

Article

Information Thermodynamics Derives the Entropy Current of Cell Signal Transduction as a Model of a Binary Coding System

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Abstract: The analysis of cellular signaling cascades based on information thermodynamics has recently developed considerably. A signaling cascade may be considered a binary code system consisting of two types of signaling molecules that carry biological information, phosphorylated active, and non-phosphorylated inactive forms. This study aims to evaluate the signal transduction step in cascades from the viewpoint of changes in mixing entropy. An increase in active forms may induce biological signal transduction through a mixing entropy change, which induces a chemical potential current in the signaling cascade. We applied the fluctuation theorem to calculate the chemical potential current and found that the average entropy production current is independent of the step in the whole cascade. As a result, the entropy current carrying signal transduction is defined by the entropy current mobility.

Keywords: signal transduction; fluctuation theorem; entropy production rate

1. Introduction

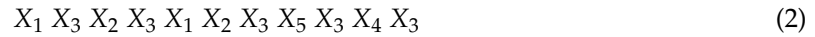
The cell is an open and non-equilibrium system, and signal transduction is one of the non-equilibrium processes characterized by a chemical potential current. In recent decades, the theoretical analysis of signal transduction has been broadly applied in various research fields, in parallel with significant development of information theory [1–5]. Informational thermodynamics for analyzing dynamic biochemical networks and systems biology have also been developed to assess the cell response to external stimuli [1–6]. In addition, in vivo analysis, a significant amount of data on signal transduction has accumulated, and the quantitative analysis of a network of signaling cascades can be performed using new technology [7–16]. In this study, a quantitative evaluation theory of a signaling cascade is described from the source coding theory of a binary code system applied for biological signal transduction. The ubiquitous signaling pathway conveys signals from the cell membrane to the nucleus shown as a form of a model scheme.

Let us consider a cell system as an open homogeneous reactor in contact with chemiostats of reactants and products, which drive the system out of equilibrium. The system is assumed to be isothermal and isovolumic.

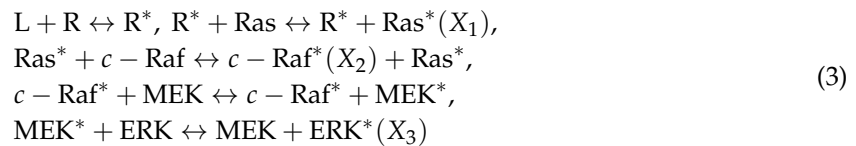
In this model, the signaling molecule at step j , denoted as X_j , induces the modification of X_{j+1} into X_{j+1}^* . In individual steps, the dephosphorylation of X_{j+1}^* into X_{j+1} occurs by phosphatase Ph_j by the release ability of inorganic phosphate Pi from X_{j+1}^* , and the pre-stimulation steady state is subsequently recovered. A signaling step in the above cascades may be described as follows:



k_j and k_{-j} are the kinetic coefficients. ATP, ADP, and Pi represent adenosine triphosphate, adenosine diphosphate, and inorganic phosphate, respectively. Consider all the possible distinct signal transduction events that correspond to all the possible combinations of the signal molecule X_j , whose transduction length is τ_j . For instance, an event is described as follows:



The cell signaling events, represented here by the symbols X_j with numbers $N_j(1 \leq j \leq n)$, will correspond to all the possible combinations of X_j . Therefore, $N_1 = 2, N_2 = 2, N_3 = 5, N_4 = 1, N_5 = 1$ and $n = 5$ in the signal event (2). In actuality, signaling cascades have been studied extensively using models of Mitogen-activated Protein Kinase (MAPK) pathways, in which the epidermal growth factor receptor, $c - Raf$, MAP kinase-extracellular signal-regulated kinase [17], and kinase-extracellular signal-regulated kinase (ERK) are phosphorylated following treatment with growth factors. The Ras- c -Raf-ERK cascade (RRE) is a ubiquitous signaling pathway that conveys mitogenic and differentiation signals from the cell membrane to the nucleus.



In the above equation, R and L represent the receptor in the cell membrane and the ligand that is substance stimulating receptor, respectively. External stimulation on the cell induces a concentration fluctuation in the phosphorylation of the signaling molecules. More specifically, a fluctuation in the signaling molecules' concentration tentatively increases, followed by a decrease over a long duration, τ_j , of several hours [7–24] (Figure 1). Here, we defined the occurrence probability, p_j and p_j^* , which represents the selection probability of j -th step using X_j or X_j^* , respectively:

$$p_j = X_j / X \tag{4}$$

$$p_j^* = X_j^* / X \tag{5}$$

with

$$\sum_{j=1}^n p_j + p_j^* = 1 \tag{6}$$

Here, X represents the total concentration of signaling molecules.

$$X = \sum_{j=1}^n X_j + X_j^* \tag{7}$$

Because the sum of the concentrations of active j molecules, X_j^* , and non-active j molecules, X_j , participating in signaling cascades is regarded as constant, the protein production process is relatively slower than the signal transduction step:

$$p_j + p_j^* = p_j^0 \tag{8}$$

The entire duration, τ , which signifies the sum of cascades, is determined using the total concentration of signaling molecules, X , and the probabilities, p_j and p_j^* .

Here, τ_j signifies the signal step duration of the j -step of the cascade. Subsequently, the total number of signal events, ψ , is introduced for the whole cascade, as follows:

$$\psi = \frac{X!}{\prod_{j=1}^n X_j! \prod_{j=1}^n X_j^*!} \tag{9}$$

The logarithm of ψ , which is Shannon’s entropy S , is given using Stirling’s approximation, as follows:

$$S = \log \psi \simeq -k_B X \left(\sum_{j=1}^n p_j \log p_j + \sum_{j=1}^n p_j^* \log p_j^* \right) \tag{10}$$

In Equation (10), k_B represents the Boltzmann constant.

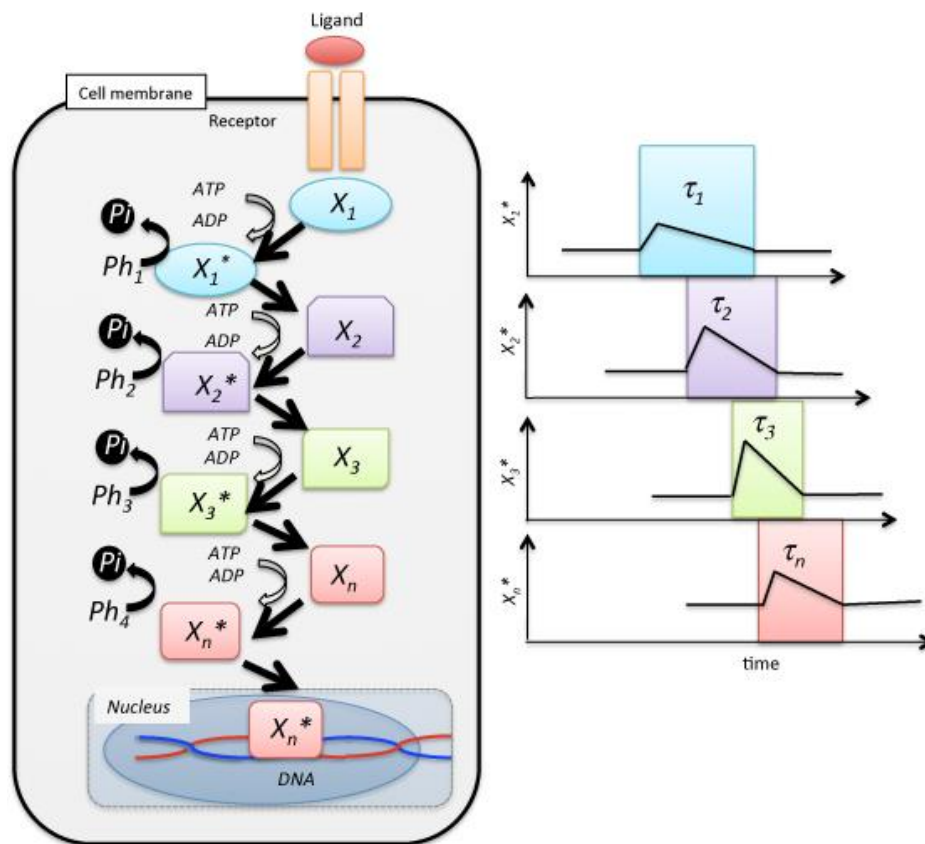


Figure 1. Schematic of a reaction cascade in cell signal transduction. The receptor mediates the cellular response to the presence of the ligand in the extracellular medium. A is a messenger, ATP, of signal transduction. Individual signaling molecules $X_j(1 \leq j \leq n)$ relay the modification of individual steps, and the last species X_n is translocated to the nucleus, where it controls gene expression by the transcription of mRNA. Ph denotes a phosphatase.

2. Mixing Entropy in Signal Transduction

Here, we noticed that the right side of Equation (10) is identical to mixing entropy in the system in which X_j^* and X_j are mixed. Here, our aim was to estimate the entropy change at individual steps in the cascade. Because the signaling molecules X_j are macromolecules, they are localized and the individual steps are compartmentalized in the cytoplasm. In the steady state, the signal transduction system stands at steady state. Here, let us consider that the ligand molecule stimulates the given system, and this stimulation produces a fluctuation in the transduction system. Considering

the entropy current from the j -th to $(j + 1)$ -th step by the mixing entropy consisting of active molecule X_j^* and X_j difference between the steps, the mixing entropy change of the j -th step, dS_j^{mix} , with a minimal concentration difference in X_j^* , dp_j^* , and in X_j , $dp_j = -dp_j^*$, is described:

$$dS_j^{\text{mix}} \triangleq -k_B X [(p_j + dp_j) \log(p_j + dp_j) + (p_j^* + dp_j^*) \log(p_j^* + dp_j^*)] \quad (11)$$

In Equation (11), dp_j^* and dp_j denote the fluctuations in the transduction system. On the other hand, because the increase and the decrease are not observed in the $(j + 1)$ -th step in the initial phase of the signal transduction from the j -th to $(j + 1)$ -th step:

$$dS_{j+1}^{\text{mix}} \triangleq -k_B X [p_j \log p_j + p_j^* \log p_j^*] \quad (12)$$

Here, T represents the temperature of the given system. Then, we have the entropy signal current C_j arising from chemical potential difference on the left side of Equations (11) and (12) using differential coefficient of missing entropy for p_j^* , which is the probability of selection of an active molecule that transmits the signal transduction:

$$C_j = T \frac{\partial dS_j^{\text{mix}}}{\partial p_j^*} \Delta p_j^* \approx k_B T X \log \frac{p_j}{p_j^*} \Delta p_j^* = k_B T \log \frac{p_j}{p_j^*} \Delta X_j^* \quad (13)$$

Therefore, the entropy signal current density c_j from the entropy change is given:

$$c_j = \frac{C_j}{\Delta X_j} = k_B T \log \frac{p_j}{p_j^*} \quad (14)$$

In general, such a nonequilibrium steady system is given by the occurrence probability p during signal step duration τ_j for the current c_j from the left reservoir at temperature β_L^{-1} and chemical potential μ_L to the right reservoir at β_R^{-1} and μ_R satisfies the steady fluctuation theorem [25]:

$$\lim_{\tau_j \rightarrow \infty} \frac{1}{\tau_j} \log \left\{ \frac{p(j+1|j; q_j^*)}{p(j|j+1; -q_j^*)} \right\} = \frac{\beta_L \mu_L - \beta_R \mu_R}{\tau_j} q_j^* \quad (15)$$

Here, q_j represents the flow of the signal current.

This fluctuation theorem leads to various nonequilibrium relations among cumulates of the current. Because in the biological systems, $\beta_L^{-1} = \beta_R^{-1} = \beta^{-1}$ and using $\beta^{-1} = k_B T$, we have using signal current density:

$$\lim_{\tau_j \rightarrow \infty} \frac{1}{\tau_j} \log \frac{p(j+1|j)}{p(j|j+1)} = \frac{c_j}{k_B T \tau_j} \Delta X_j^* \quad (16)$$

On the left side of (16), $p(j+1|j)$, the transitional probability of step $j+1$ given step j , is defined in the forward direction of the signal is also defined. From Equations (15) and (16) we have an important result:

$$\lim_{\tau_j \rightarrow \infty} \frac{1}{\tau_j} \log \frac{p(j+1|j)}{p(j|j+1)} = \frac{1}{\tau_j} \log \frac{p_j}{p_j^*} \quad (17)$$

Equation (17) shows the relation between the transitional probability and the occurrence probability. In most cases of biological signal transduction, the signal duration is sufficiently long, and therefore Equation (17) can be further described simply as follows:

$$\log \frac{p(j+1|j)}{p(j|j+1)} = \log \frac{p_j}{p_j^*} \quad (18)$$

In our previous reports [26,27], when the number of the events or messages per a given duration is maximized, the occurrence probability can be described using an arbitrary parameter ζ independent of the step number j :

$$-\log p_j = \zeta \tau_j \quad (19)$$

Further, as shown in Appendix A, Equation (18) can be rewritten as follows:

$$\frac{1}{\tau_j} \log \frac{p(j+1|j)}{p(j|j+1)} = -\zeta \quad (20)$$

As a result, we have an important result from Equations (16) and (20):

$$\zeta = -\frac{c_j \Delta X_j^*}{k_B T \tau_j} = -\frac{C_j}{k_B T \tau_j} = -\frac{J}{k_B T} \quad (21)$$

with

$$J \triangleq \frac{C_j}{\tau_j} = k_B T \zeta \quad (22)$$

Here, J represents the average entropy production current along the cascade, and the suffix j representing the step number is omitted because the average current is independent of the step number. The above equation represents that ζ has the dimension of the average entropy production rate.

3. Entropy Current and Signal Transduction

Subsequently, we aimed to formulate the signal current from the perspectives of the entropy current. The entropy current depends on the spatial gradient of entropy and is given using the signal current intensity from (13) and (22) using an intracellular spatial coordinate parameter r :

$$-T \nabla_r S_j \approx -k_B T \log \frac{p_j}{p_j^*} \nabla_r X_j^* = -c_j \nabla_r X_j^* = -\nabla_r C_j \quad (23)$$

Further, using the diffusion coefficient of an active signaling molecule, the average entropy current per signal duration is given using the diffusion coefficient of the signal, D_j and from (22):

$$\nabla_r J = -\frac{D_j \nabla_r X_j^*}{\tau_j} = -\frac{c_j \nabla_r X_j^*}{\tau_j} \quad (24)$$

Here, the diffusion coefficient is obtained from (23):

$$D_j \triangleq c_j \quad (25)$$

From the Stokes–Einstein equation, the diffusion coefficients can be described using the signal mobility, ω_j :

$$D_j \triangleq k_B T \omega_j \quad (26)$$

Therefore, we have:

$$c_j = k_B T \omega_j \quad (27)$$

4. Conclusions

In the current study, we hypothesized that the signaling cascade is a binary code system in which an increase in the concentration of the active signal molecule at each step (although accompanied by a decrease in the inactive form of the signal molecule) transmits a signal transduction between steps. This simple binary-code hypothesis enabled us to have several important equities about the quantification of signal transformation. This hypothesis includes a theoretical basis that can introduce

a duration parameter to analyze the development of signal transduction over time. We postulate that quantitative estimation of signal transduction is possible based on the amount of ATP consumed. In an actual experiment, preventing other biochemical reactions, apart from signal transduction, is challenging, but we have a plan for establishing a model for measurement. Therefore, if data with adequate comprehensive signal events is accumulated, quantification of the signal events might be possible by measurement of ATP concentration changes.

According to this idea and the fluctuation theorem [28–30], we obtained an important result: the diffusion coefficient of the signal event is equal to the entropy current. In this way, the signal transduction in the cell system is definitely formulated in the entropy production and the current. Significantly, the average entropy production rate current is independent of the step number; this implies that the whole cascade of the signal transduction is integrated well under stable entropy production. Based on this finding, the quantification of the signal events is possible by measuring the chemical potential change during individual signal event in the cell system. For example, the consumption of ATP, which mediates signal transduction, is anticipated to provide data regarding the entropy production during the events. In conclusion, the current signaling cascade model provides a basis for informational thermodynamics, and the relationship between the chemical potential and information or entropy was established.

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Conflicts of Interest: The author declares no conflict of interest.

Appendix A

To obtain Equation (20) from Equations (8), (18) and (19), we can calculate as follows:

$$\begin{aligned} \frac{1}{\tau_j} \log \frac{p_j}{p_j^*} &= \frac{1}{\tau_j} \log \frac{\exp(-\zeta\tau_j)}{p_j^0 - \exp(-\zeta\tau_j)} = \frac{1}{\tau_j} \log \frac{\exp(-\zeta\tau_j)}{p_j^0 [1 - \exp(-\zeta\tau_j)/p_j^0]} \\ &= -\zeta - \frac{1}{\tau_j} \log p_j^0 - \frac{1}{\tau_j} \log [1 - \exp(-\zeta\tau_j)/p_j^0] \\ &= -\zeta - \frac{1}{\tau_j} \log p_j^0 + \frac{1}{\tau_j} \exp(-\zeta\tau_j)/p_j^0 \\ &\approx -\zeta (\tau_j \rightarrow \infty) \end{aligned}$$

References

1. Selimkhanov, J.; Taylor, B.; Yao, J.; Pilko, A.; Albeck, J.; Hoffmann, A.; Tsimring, L.; Wollman, R. Systems biology. Accurate information transmission through dynamic biochemical signaling networks. *Science* **2014**, *346*, 1370–1373. [[CrossRef](#)] [[PubMed](#)]
2. Sagawa, T.; Kikuchi, Y.; Inoue, Y.; Takahashi, H.; Muraoka, T.; Kinbara, K.; Ishijima, A.; Fukuoka, H. Single-cell *E. coli* response to an instantaneously applied chemotactic signal. *Biophys. J.* **2014**, *107*, 730–739. [[CrossRef](#)] [[PubMed](#)]
3. Cheong, R.; Rhee, A.; Wang, C.J.; Nemenman, I.; Levchenko, A. Information transduction capacity of noisy biochemical signaling networks. *Science* **2011**, *334*, 354–358. [[CrossRef](#)] [[PubMed](#)]
4. Watters, N.; Reeke, G.N. Neuronal spike train entropy estimation by history clustering. *Neural Comput.* **2014**, *26*, 1840–1872. [[CrossRef](#)] [[PubMed](#)]
5. Hanel, R.; Thurner, S.; Gell-Mann, M. How multiplicity determines entropy and the derivation of the maximum entropy principle for complex systems. *Proc. Natl. Acad. Sci. USA* **2014**, *111*, 6905–6910. [[CrossRef](#)] [[PubMed](#)]
6. Teschendorff, A.E. Increased entropy of signal transduction in the cancer metastasis phenotype. *BMC Syst. Biol.* **2010**, *4*, e104. [[CrossRef](#)] [[PubMed](#)]

7. Lee, C.S.; Park, M.; Han, J.; Lee, J.H.; Bae, I.H.; Choi, H.; Son, E.D.; Park, Y.H.; Lim, K.M. Liver X receptor activation inhibits melanogenesis through the acceleration of ERK-mediated MITF degradation. *J. Investig. Dermatol.* **2013**, *133*, 1063–1071. [[CrossRef](#)] [[PubMed](#)]
8. Mackeigan, J.P.; Murphy, L.O.; Dimitri, C.A.; Blenis, J. Graded mitogen-activated protein kinase activity precedes switch-like c-Fos induction in mammalian cells. *Mol. Cell. Biol.* **2005**, *25*, 4676–4682. [[CrossRef](#)] [[PubMed](#)]
9. Newman, D.R.; Li, C.M.; Simmons, R.; Khosla, J.; Sannes, P.L. Heparin affects signaling pathways stimulated by fibroblast growth factor-1 and-2 in type II cells. *Am. J. Physiol. Lung Cell. Mol. Physiol.* **2004**, *287*, L191–L200. [[CrossRef](#)] [[PubMed](#)]
10. Petropavlovskaja, M.; Daoud, J.; Zhu, J.; Moosavi, M.; Ding, J.; Makhlin, J.; Assouline-Thomas, B.; Rosenberg, L. Mechanisms of action of islet neogenesis-associated protein, comparison of the full-length recombinant protein and a bioactive peptide. *Am. J. Physiol. Endocrinol. Metab.* **2012**, *303*, E917–E927. [[CrossRef](#)] [[PubMed](#)]
11. Tao, R.; Hoover, H.E.; Honbo, N.; Kalinowski, M.; Alano, C.C.; Karliner, J.S.; Raffai, R. High-density lipoprotein determines adult mouse cardiomyocyte fate after hypoxia-reoxygenation through lipoprotein-associated sphingosine 1-phosphate. *Am. J. Physiol. Heart Circ. Physiol.* **2010**, *298*, H1022–H1028. [[CrossRef](#)] [[PubMed](#)]
12. Mina, M.; Magi, S.; Jurman, G.; Itoh, M.; Kawaji, H.; Lassmann, T.; Arner, E.; Forrest, A.R.R.; Carninci, P.; Hayashizaki, Y. Promoter-level expression clustering identifies time development of transcriptional regulatory cascades initiated by ErbB receptors in breast cancer cells. *Sci. Rep.* **2015**, *5*, 11999. [[CrossRef](#)] [[PubMed](#)]
13. Wang, H.; Ubl, J.J.; Stricker, R.; Reiser, G. Thrombin (PAR-1)-induced proliferation in astrocytes via MAPK involves multiple signaling pathways. *Am. J. Physiol. Cell Physiol.* **2002**, *283*, C1351–C1364. [[CrossRef](#)] [[PubMed](#)]
14. Wang, Y.Y.; Liu, Y.; Ni, X.Y.; Bai, Z.H.; Chen, Q.Y.; Zhang, Y.; Gao, F.G. Nicotine promotes cell proliferation and induces resistance to cisplatin by alpha7 nicotinic acetylcholine receptor-mediated activation in Raw264.7 and EL4 cells. *Oncol. Rep.* **2014**, *31*, 1480–1488. [[CrossRef](#)] [[PubMed](#)]
15. Yeung, K.; Seitz, T.; Li, S.; Janosch, P.; McFerran, B.; Kaiser, C.; Fee, F.; Katsanakis, K.D.; Rose, D.W.; Mischak, H.; et al. Suppression of Raf-1 kinase activity and MAP kinase signalling by RKIP. *Nature* **1999**, *401*, 173–177. [[CrossRef](#)] [[PubMed](#)]
16. Zhang, W.Z.; Yano, N.; Deng, M.; Mao, Q.; Shaw, S.K.; Tseng, Y.-T. β -Adrenergic Receptor-PI3K Signaling Crosstalk in Mouse Heart, Elucidation of Immediate Downstream Signaling Cascades. *PLoS ONE* **2011**, *6*, e26581. [[CrossRef](#)] [[PubMed](#)]
17. Jung, P.; Menssen, A.; Mayr, D.; Hermeking, H. AP4 encodes a c-MYC-inducible repressor of p21. *Proc. Natl. Acad. Sci. USA* **2008**, *105*, 15046–15051. [[CrossRef](#)] [[PubMed](#)]
18. Blosser, R.; Bodart, J.F.; Devys, A.; Goudon, T.; Lafitte, P. Signal propagation of the MAPK cascade in *Xenopus* oocytes, role of bistability and ultrasensitivity for a mixed problem. *J. Math. Biol.* **2012**, *64*, 1–39. [[CrossRef](#)] [[PubMed](#)]
19. Hollenberg, M.D. PARs in the stars, proteinase-activated receptors and astrocyte function. Focus on “Thrombin (PAR-1)-induced proliferation in astrocytes via MAPK involves multiple signaling pathways”. *Am. J. Physiol. Cell Physiol.* **2002**, *283*, C1347–C1350. [[CrossRef](#)] [[PubMed](#)]
20. Purutçuoğlu, V.; Wit, E. Estimating Network Kinetics of the MAPK/ERK Pathway Using Biochemical Data. *Math. Probl. Eng.* **2012**, *2012*, 1–34. [[CrossRef](#)]
21. Qiao, L.; Nachbar, R.B.; Kevrekidis, I.G.; Shvartsman, S.Y. Bistability and oscillations in the Huang-Ferrell model of MAPK signaling. *PLoS Comput. Biol.* **2007**, *3*, 1819–1826. [[CrossRef](#)] [[PubMed](#)]
22. Xin, X.; Zhou, L.; Reyes, C.M.; Liu, F.; Dong, L.Q. APPL1 mediates adiponectin-stimulated p38 MAPK activation by scaffolding the TAK1-MKK3-p38 MAPK pathway. *Am. J. Physiol. Endocrinol. Metab.* **2011**, *300*, E103–E110. [[CrossRef](#)] [[PubMed](#)]
23. Yoon, J.; Deisboeck, T.S. Investigating differential dynamics of the MAPK signaling cascade using a multi-parametric global sensitivity analysis. *PLoS ONE* **2009**, *4*, e4560. [[CrossRef](#)] [[PubMed](#)]
24. Zumsande, M.; Gross, T. Bifurcations and chaos in the MAPK signaling cascade. *J. Theor. Biol.* **2010**, *265*, 481–491. [[CrossRef](#)] [[PubMed](#)]

25. Watanabe, K.L.; Hayakawa, H. Geometric fluctuation theorem for a spin-boson system. *Phys. Rev. E* **2017**, *96*, 022118. [[CrossRef](#)] [[PubMed](#)]
26. Tsuruyama, T. Channel Capacity of Coding System on Tsallis Entropy and q-Statistics. *Entropy* **2017**, *19*, 682. [[CrossRef](#)]
27. Brillouin, L. *Science and Information Theory*, 2nd ed.; Dover Publication Inc.: New York, NY, USA, 2013.
28. Andrieux, D.; Gaspard, P. Fluctuation theorem and Onsager reciprocity relations. *J. Chem. Phys.* **2004**, *121*, 6167–6174, Erratum in **2006**, *125*, 219902. [[CrossRef](#)] [[PubMed](#)]
29. Andrieux, D.; Gaspard, P. Fluctuation theorem and mesoscopic chemical clocks. *J. Chem. Phys.* **2008**, *128*, 154506. [[CrossRef](#)] [[PubMed](#)]
30. Gaspard, P. Fluctuation theorem for nonequilibrium reactions. *J. Chem. Phys.* **2004**, *120*, 8898–8905. [[CrossRef](#)] [[PubMed](#)]



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