

*Article*

# **Synthesis and Comparative Structure–Activity Study of Carbohydrate-Based Phenolic Compounds as** α**-Glucosidase Inhibitors and Antioxidants**

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Received: 7 November 2019; Accepted: 25 November 2019; Published: 27 November 2019



**Abstract:** Twenty-one natural and unnatural phenolic compounds containing a carbohydrate moiety were synthesized and their structure–activity relationship (SAR) was evaluated for  $\alpha$ -glucosidase inhibition and antioxidative activity. Varying the position of the galloyl unit on the 1,5-anhydro -p-glucitol (1,5-AG) core resulted in changes in the  $\alpha$ -glucosidase inhibitory activity and notably, particularly strong activity was demonstrated when the galloyl unit was present at the C-2 position. Furthermore, increasing the number of the galloyl units significantly affected the  $\alpha$ -glucosidase inhibition, and 2,3,4,6-tetra-galloyl-1,5-AG (54) and 2,3,4,6-tetra-galloyl-p-glucopyranose (61) exhibited excellent activities, which were more than 13-fold higher than the  $\alpha$ -glucosidase inhibitory activity of acertannin (**37**). Moreover, a comparative structure-activity study suggested that a hemiacetal hydroxyl functionality in the carbohydrate core and a biaryl bond of the 4,6-*O*-hexahydroxydiphenoyl (HHDP) group, which are components of ellagitannins including tellimagrandin I, are not necessary for the  $\alpha$ -glucosidase inhibitory activity. Lastly, the antioxidant activity increased proportionally with the number of galloyl units.

**Keywords:** 1,5-AG; tellimagrandin I; acertannin; maplexin; ginnalin; polyphenol; α-glucosidase; antioxidant

# **1. Introduction**

Impaired glucose tolerance increases the risk of vascular events such as atherosclerotic coronary artery disease [\[1,](#page-23-0)[2\]](#page-23-1). Particularly, postprandial hyperglycemia is a serious risk factor for cardiovascular diseases and is believed to be the cause of oxidative stress that leads to vascular events [\[3](#page-23-2)[–7\]](#page-23-3). Thus, controlling postprandial hyperglycemia is an important target to prevent diabetes as well as diabetic complications. In clinical medicine,  $\alpha$ -glucosidase inhibitors such as acarbose, miglitol, and voglibose, belong to the class of antidiabetic drugs used for improving postprandial hyperglycemia [\[8–](#page-24-0)[11\]](#page-24-1). Currently, natural products and their derivatives constitute more than half of the drugs in the clinic [\[12–](#page-24-2)[15\]](#page-24-3). Therefore, finding inspiration in nature to develop more efficient and effective medicines has attracted significant interest.

Trees belonging to the *Acer* species have been used as traditional medicinal plants for many years and are widely known for their sap, which can be concentrated to produce maple syrup [\[16\]](#page-24-4). It has been demonstrated that *Acer* extracts display various bioactivities such as anti-cancer [\[17,](#page-24-5)[18\]](#page-24-6), antioxidant [\[19](#page-24-7)[–22\]](#page-24-8), and antihyperglycemic effects [\[23,](#page-24-9)[24\]](#page-24-10). A. Honma et al. identified a compound from *Acer saccharum* extracts able to suppress hyperglycemia, namely acertannin, and revealed that its effects are a consequence of potent inhibitory activity toward  $\alpha$ -glucosidase [\[25\]](#page-24-11). The structural components of acertannin include the characteristic  $1,5$ -anhydro-p-glucitol ( $1,5$ -AG) sugar moiety,



<span id="page-1-0"></span>which lacks the hemiacetal hydroxyl group present in p-glucose, as the carbohydrate core, and two gallic acid functionalities as the phenolic units (Figure [1\)](#page-1-0) [\[26\]](#page-24-12). However, only a few plants belonging to the *Acer* genus produce the 1,5-AG core containing polyphenols [\[27,](#page-24-13)[28\]](#page-24-14). To date, maplexin A–J and ginnalin A-C have been isolated and characterized. The molecules possess varying numbers and positions of the phenol units esterified with the 1,5-AG core [29-32]. These polyphenols were shown to exhibit different bioactivities such as  $\alpha$ -glucosidase inhibition and antioxidant activity. It is noteworthy that different numbers, positions, and types of the phenol units on the 1,5-AG core display non-identical bioactivities [\[32](#page-25-0)[–36\]](#page-25-1). display non-identical bioactivities [32–36].



**Figure 1.** Structure of acertannin. **Figure 1.** Structure of acertannin.

Tellimagrandin I, which belongs to ellagitannins, has also been demonstrated to be an α-Tellimagrandin I, which belongs to ellagitannins, has also been demonstrated to be anα-glucosidase inhibitor and to show antioxidant activity  $[37,38]$ . The molecule is characterized by the presence of a hexahydroxydiphenoyl (HHDP) group, two galloyl units and the p-glucose core possesses a hemiacetal hydroxyl functionality (Figure [2\)](#page-1-1) [39]. The HHDP group provides structural diversity in polyphenols, and the macro-lactone structure is considered to be the element responsible for the pharmacological activity [\[40\]](#page-25-5). Nonetheless, to our knowledge, no reports on the evaluation of the synthesis and/or bioactivity of compounds comprising the HHDP functionality on the 1,5-AG core have been reported so far. Furthermore, the hemiacetal hydroxyl group is a fundamental moiety in the carbohydrate chemistry; however, its effects on the bioactivity remain largely unexplored. in the carbohydrate chemistry; however, its effects on the bioactivity remain largely unexplored.

<span id="page-1-1"></span>

Figure 2. The structural components of tellimagrandin I. HHDP, hexahydroxydiphenoyl.

In the present study, we report the synthesis of a series of 21 carbohydrate-based phenolic compounds to investigate the structure–activity relationship (SAR). α-glucosidase inhibition and antioxidant activity were examined by studying the effects of (1) the position and number of galloyl units, (2) the type of phenol units, (3) the existence the 4,6-*O*-HHDP group, and (4) the presence of the hemiacetal hydroxyl group. hemiacetal hydroxyl group.

# **2. Results 2. Results**

# *2.1. Syntheses of 1,5-AG-Based Polyphenols 2.1. Syntheses of 1,5-AG-Based Polyphenols*

# 2.1.1. Syntheses of Galloylated 1,5-AGs 2.1.1. Syntheses of Galloylated 1,5-AGs

Recently, A. Kamori et al. reported the synthesis of various natural and unnatural acertannin Recently, A. Kamori et al. reported the synthesis of various natural and unnatural acertannin derivatives and evaluated their SAR against ceramidase and ceramide synthase enzymes [\[35\]](#page-25-6). In addition, we have also previously reported a facile method for the preparation of 1,5-anhydroalditol via treatment of per-O-TMS-glycopyranosyl iodide with LiBH<sub>4</sub> [\[41\]](#page-25-7). In total, 1,5-AG, which can be easily synthesized from D-glucose on multi-gram scale in three days, possesses four hydroxyl groups; hence, 15 different combinations are possible for mono-, di-, tri-, and tetra-galloylation of 1,5-AG. In the current study, we attempted the synthesis of all of these galloylated compounds starting from 1,5-AG.

Firstly, 4,6-O-benzylidene-1,5-AG (1) [\[42\]](#page-25-8) was protected with benzyl (Bn) group using BnBr and NaH to afford di-benzylated compound 2 [\[42\]](#page-25-8). However, reacting 1 in a 2-phase dichloromethane (DCM)/5% NaOH system with BnBr, NaI, and tetra-n-butylammonium hydrogen sulfate provided the 3-OH analog 3 [\[35\]](#page-25-6) and the 2-OH analog 4 [35] as a mixture of products, which could be separated by the 3-OH analog **3-OH analog 2-OH analog 2**  $\sigma$  1) (Scheme 1) [43].

<span id="page-2-0"></span>

**Scheme 1.** Selective protection of 1,5-AG. **Scheme 1.** Selective protection of 1,5-AG.

corresponding hydroxyl moieties using an acid or reducing reagent (Scheme [2\)](#page-3-0). Compounds **2–4** were then deprotected with 80% acetic acid/H<sub>2</sub>O solution to give the corresponding diol analogs 5–7 [\[35](#page-25-6)[,42\]](#page-25-8). were the protected with 80% acetic acid<sub>1H2</sub>O solution to give the corresponding diol analogs **5**–1<sub>1</sub>007.11<br>advantage of 1–2, and 1 by RH (THE and trimathyloily) trifly aromathanogy flonate (TMCOTA Lt) Reduction of 1, 3, and 4 by  $BH_3/THF$  and trimethylsilyl trifluoromethanesulfonate (TMSOTf) [\[44\]](#page-25-10), and reduction of 2 by diisobutylaluminium hydride (DIBAL-H) [\[45\]](#page-25-11) resulted in the formation of the and the set of 6-OH derivatives 8–11 [\[35,](#page-25-6)[46,](#page-25-12)[47\]](#page-25-13). Moreover, compounds 1–4 were treated with triethylsilane and trifluoroacetic acid (TFA)  $[48]$  to afford the 4-OH derivatives  $12-15$   $[35,49]$  $[35,49]$ . Esterification of these -OH derivatives with Bn-protected gallic acid  $(16)$  [\[50\]](#page-25-16) using N,N-dimethyl-4-aminopyridine (DMAP) and triethylamine (TEA), as well as 2-chloro-1-methylpyridinium iodide as the condensing reagent, and triethylamine (TEA), as well as 2-chloro-1-methylpyridinium iodide as the condensing reagent, provided 1,5-AG-based galloylated derivatives **17**–**30**. Finally, hydrogenolysis using Pd(OH)2 as the provided 1,5-AG-based galloylated derivatives **17**–**30**. Finally, hydrogenolysis using Pd(OH)<sup>2</sup> as the catalyst in a MeOH/THF solvent mixture gave 14 types of the 1,5-AG core containing polyphenol catalyst in a MeOH/THF solvent mixture gave 14 types of the 1,5-AG core containing polyphenol analogs **31**–**44** (Scheme 3). analogs **31**–**44** (Scheme [3\)](#page-3-1).Subsequently, the 4,6-*O*-benzylidene groups on the 1,5-AG derivatives **1**–**4** were converted to the

<span id="page-3-1"></span><span id="page-3-0"></span>

**Scheme 3.** Synthesis of galloylated 1,5-AGs. **Scheme 3.** Synthesis of galloylated 1,5-AGs.

# 2.1.2. Synthesis of Maplexin J Analogs

Maplexin J (**54**) [\[32](#page-25-0)[,50\]](#page-25-16), which is per-galloylated 1,5-AG, could be easily synthesized from 1,5-AG. Therefore, we attempted to synthesize of maplexin J analogs, which in addition to the galloyl moiety, contained different numbers of phenolic groups at various positions intending to elucidate the effect of this functionality on the bioactivity. Phenol derivatives **45**–**48** [\[51–](#page-26-0)[53\]](#page-26-1) were prepared in 2-steps from commercially available methyl esters (Scheme [4\)](#page-4-0). Subsequently, 1,5-AG was condensed with **16** [\[50\]](#page-25-16), and **45**–**48** using DMAP, TEA, and 2-chloro-methylpyridinium iodide to obtain the corresponding Bn-protected 1,5-AG analogs **49**–**53**. Deprotection under hydrogenolysis conditions provided compounds **54**–**58** (Scheme [5\)](#page-4-1). However, esterification of benzyl glucoside **59** [\[54\]](#page-26-2) with **16** using 1-ethyl-3-(3-dimethylaminopropyl)carbodiimide (EDC) and DMAP resulted in the formation of intermediate 60. Following hydrogenolysis, tetra-galloylated p-glucopyranose 61 containing a hemiacetal moiety was formed. Notably, the structure of **61 [\[55\]](#page-26-3)** is analogous to tellimagrandin I; however, the biaryl bond of the HHDP group is missing (Scheme [6\)](#page-5-0).

<span id="page-4-0"></span>

45-48 (60-98%, 2 steps)

<span id="page-4-1"></span>

Scheme 4. Preparation of the Bn-protected phenol units 45-48.

**Scheme 5.** Synthesis of maplexin J analogs **54**–**58**. **Scheme 5.** Synthesis of maplexin J analogs **54**–**58**.

<span id="page-5-0"></span>

**Scheme 6.** Synthesis of tellimagrandin I analog **61**. **Scheme 6.** Synthesis of tellimagrandin I analog **61**.

2.1.3. Synthesis of a 1,5-AG-Based Tellimagrandin I Analog 2.1.3. Synthesis of a 1,5-AG-Based Tellimagrandin I Analog

To consider the effect of the hemiacetal group on the activity of tellimagrandin I, we also focused To consider the effect of the hemiacetal group on the activity of tellimagrandin I, we also focused on the synthesis of the 1-deoxy analog **67**. Deprotection of benzylidene acetal in **21** in a methanol/DCM solvent system using iodine, according to the method reported by Feldman et al. [\[56\]](#page-26-4), gave diol **62** in a high yield of 93% (Scheme 7). Intermediate 62 was then condensed with the gallic acid derivative **63** [57] to afford the galloylated compound **64**. The subsequent removal of the methoxymethyl (MOM) protecting groups provided 65 in 93% yield.

<span id="page-5-1"></span>

**Scheme 7.** Synthesis of the 1,5-AG-based tellimagrandin I analog **67**. **Scheme 7.** Synthesis of the 1,5-AG-based tellimagrandin I analog **67**.

The construction of the HHDP group was performed in accordance with the approach previously the biaryl derivative 66. Finally, hydrogenation of 66 in MeOH/THF with Pd(OH)<sub>2</sub> as the catalyst gave the desired 1,5-AG-based tellimagrandin I analog **67**. reported by Yamada et al. [\[57–](#page-26-5)[60\]](#page-26-6). Compound 65 was treated with  $n$  − BuNH<sub>2</sub> and CuCl<sub>2</sub> to provide

#### *2.2. Evaluation of* α*-Glucosidase Inhibition and Antioxidant Activity*

#### 2.2.1. The  $\alpha$ -Glucosidase Inhibitory Activity

The  $\alpha$ -glucosidase inhibitory activity of all samples was assayed utilizing a commercially available FUJIFILM α-glucosidase inhibitory activity assay kit. In total, 25 μL of each sample (25–2000 μg/mL in H<sub>2</sub>O) or acarbose (0.5–8  $\mu$ g/mL in H<sub>2</sub>O), 50  $\mu$ L of 18.5 mM maltose diluted in maleic anhydride buffer (100 mM,  $pH = 6.0$ ), and 25 µL of the rat  $\alpha$ -glucosidase solution were incubated in a micro-tube at 37 ◦C for 30 min. Subsequently, 400 µL of purified water was added to the solution and the reaction mixture was boiled for 3 min to deactivate α-glucosidase. The generated glucose was measured by LabAssay<sup>™</sup> glucose (mutarotase-GOD method). In total, 100 µL of the reaction solution and 150 µL of the coloring solution were incubated in 96-well plates at 37 °C for 10 min, and the absorbance was recorded at 505 nm using a microplate reader (Bio-Rad, Model 680). The inhibition percentage was calculated using the following Equation:

Inhibition percentage (%) = 
$$
1 - \frac{As - Ab}{Ac} \times 100
$$

where As is the absorbance of the analyzed sample, Ab is the absorbance of the blank (immediate deactivation), and Ac is the absorbance of the control (without α-glucosidase). Acarbose was used as the positive control.

#### 2.2.2. Evaluation of Antioxidant Activity

The antioxidant activity was assessed by employing the 2,2-diphenyl-1-picrylhydrazyl (DPPH) radical scavenging assay, according to the previously published method  $[61]$ . In total, 100  $\mu$ L of each sample (1000–10  $\mu$ g/mL in 50%EtOH/H<sub>2</sub>O) or trolox (160–0  $\mu$ M in 50% EtOH/H<sub>2</sub>O), and 50  $\mu$ L of 2-(*N*-morpholino)ethanesulfonic acid (MES)-NaOH buffer (200 mM,  $pH = 6.0$ ), and 50  $\mu$ L of an 800 µM DPPH diluted in 99.5% EtOH were mixed in 96-well plates. Following shaking the microplate for 15 min in the dark, the absorbance was recorded at 520 nm using a microplate reader (Bio-Rad, Model 680). Effective percentages (%) were calculated as follows:

Effective percentages (%) = 
$$
1 - \frac{\text{As}}{\text{Ab}} \times 100
$$

where As is the absorbance of the sample and Ab is the absorbance of the blank (without samples,  $50\%$  EtOH/H<sub>2</sub>O solution was used instead).

In addition, correlation of the absorbance on the *y*-axis and the concentrations on the *x*-axis resulted in the formation of an approximately straight-line plot. The trolox-equivalent (sample-mol/trolox-mol) was calculated as follows:

$$
Trolov-equivalent \left(\frac{mol - trolox}{mol - sample}\right) = \frac{Ss}{St}
$$

where Ss is the slope of the sample and St is the slope of trolox.

#### **3. Discussion**

The results of the biological evaluation of the 21 synthesized compounds considering the α-glucosidase inhibitory and antioxidant activities are summarized in Table [1.](#page-7-0)

<span id="page-7-0"></span>

Compounds	$\alpha$ -Glucosidase	Antioxidant	
	IC <sub>50</sub> ( $\mu$ M) <sup>a</sup>	$EC_{50}$ (µM) <sup>b</sup>	Trolox-eq <sup>b</sup>
Mono-			
2-galloyl (Ginnalin C) [31] 31	$95.1 \pm 0.22$	$19.40 \pm 1.02$	$2.04 \pm 0.08$
3-galloyl (Maplexin A) [29] 32	$137.9 \pm 1.50$	$20.50 \pm 2.05$	$2.38 \pm 0.24$
4-galloyl (Maplexin B) [29] 33	$143.9 \pm 0.52$	$20.80 \pm 1.54$	$2.09 \pm 0.19$
6-galloyl (Ginnalin B) [31] 34	$127.1 \pm 0.88$	$22.90 \pm 1.61$	$2.13 \pm 0.12$
Di-			
2,3-galloyl (Maplexin C) [29] 35	$20.50 \pm 0.50$	$11.30 \pm 0.98$	$4.29 \pm 0.23$
2,4-galloyl (Maplexin D) [29] 36	$48.30 \pm 1.25$	$14.30 \pm 0.29$	$3.36 \pm 0.13$
2,6-galloyl (Acertannin) [25] 37	$35.60 \pm 2.58$	$15.20 \pm 0.53$	$3.32 \pm 0.37$
3,4-galloyl 38 $c$	$91.40 \pm 6.56$	$14.80 \pm 0.56$	$3.55 \pm 0.42$
3,6-galloyl 39	$55.00 \pm 8.84$	$11.90 \pm 0.71$	$3.67 \pm 0.19$
4,6-galloyl 40	$26.60 \pm 2.37$	$13.00 \pm 1.12$	$3.78 \pm 0.33$
Tri-			
2,3,4-galloyl 41	$6.72 \pm 0.21$	$7.94 \pm 0.39$	$5.59 \pm 0.26$
2,3,6-galloyl (Maplexin F) [30] 42	$5.34 \pm 0.55$	$8.09 \pm 0.41$	$5.75 \pm 0.18$
2,4,6-galloyl (Maplexin E) [29] 43	$9.34 \pm 0.99$	$10.30 \pm 0.76$	$4.31 \pm 0.35$
3,4,6-galloyl 44	$12.60 \pm 0.61$	$8.48 \pm 0.21$	$5.36 \pm 0.20$
Tetra-			
2,3,4,6-galloyl (Maplexin J) [32] 54	$2.56 \pm 0.10$	$6.61 \pm 0.68$	$6.77 \pm 0.63$
$3'$ ,4'-dihydroxybenzoyl 55 $^c$	$3.28 \pm 0.16$	$6.99 \pm 0.11$	$6.54 \pm 0.15$
$3'$ ,5'-dihydroxybenzoyl 56 $^c$	$9.34 \pm 0.02$	70<	$n.d$ <sup><math>e</math></sup>
3'-hydroxybenzoyl 57 <sup>c</sup>	$\mathcal{A}$	78<	$n.d$ <sup>e</sup>
4'-hydroxybenzoyl 58 <sup>c</sup>	$-d$	78<	$n.d$ <sup><math>e</math></sup>
1-OH-2,3,4,6-galloyl 61	$1.68 \pm 0.21$	$5.60 \pm 0.28$	$8.33 \pm 0.49$
2,3-galloyl-4,6-HHDP 67 <sup>c</sup>	$3.22 \pm 0.51$	$6.79 \pm 0.21$	$6.44 \pm 0.24$
Tellimagrandin I	$3.37 \pm 0.04$	$6.31 \pm 0.27$	$7.04 \pm 0.37$
Methyl gallate	$90.50 \pm 6.97$	$19.60 \pm 0.11$	$2.42 \pm 0.27$
Methyl 3,4-dihydroxybenzoate	300 < <	$21.10 \pm 1.08$	$2.20 \pm 0.05$
Methyl 3,5-dihydroxybenzoate	300 < <	$n.d$ <sup>e</sup>	$n.d$ <sup>e</sup>
Methyl 3-hydroxybenzoate	660 < <	$n.d$ <sup><math>e</math></sup>	$n.d$ <sup><math>e</math></sup>
Methyl 4-hydroxybenzoate	660 < <	$n.d$ <sup>e</sup>	$n.d$ <sup>e</sup>
Acarbose	$0.11 \pm 0.02$		
Trolox	-	$49.40 \pm 3.87$	

**Table 1.** α-Glucosidase inhibitory and antioxidant activities.

 $^a$  IC<sub>50</sub> data represents mean  $\pm$  S.D. of n = 2. <sup>*b*</sup> EC<sub>50</sub> and trolox-eq data represents mean  $\pm$  S.D. of n = 3. <sup>*c*</sup> Novel compound. *<sup>d</sup>* Did not dissolve in water. *<sup>e</sup>* n.d.: not detected.

The comparison of the 1,5-AG-based polyphenol analogs **31**–**44** and **54** revealed that the α-glucosidase inhibitory activity significantly increased with the number of the galloyl units in the compounds and the highest inhibition was observed for tetra-O-galloyl-1,5-AG (maplexin J) **54** (IC<sub>50</sub> = 2.56  $\mu$ M). This result is in accordance with the previous reports [\[32\]](#page-25-0). In addition, different the position of the galloyl unit on the 1,5-AG core appeared to influence the α-glucosidase inhibitory activity, even for the compounds with the same number of these moieties. Analogous outcomes were noted for 2-galloyl-1,5-AG (**31**) and methyl gallate (IC<sub>50</sub> = 95.1, 90.5 µM, respectively). Conversely, differing  $\alpha$ -glucosidase inhibitory activities were obtained for mono-galloyl analogs **32**–**34**, which are weaker inhibitors than methyl gallate (IC<sub>50</sub> = 127.1–143.9 µM). Furthermore, compound **33** (galloyl unit at the C-4 position) exhibited lowest activity (IC<sub>50</sub> = 143.9 µM). Higher inhibitory activity was detected for di-galloylated analogs **35–37** (IC<sub>50</sub> = 20.5–48.3  $\mu$ M), which possessed the galloyl unit at the C-2 position than for analogs **38**–**40** (IC<sup>50</sup> = 26.6–91.4 µM). In particular, compound **35** that contains esterified galloyl units at the C-2 and C-3 positions exhibited three-times higher inhibitory activity than gallic acid. Moreover, analog 38, galloylated at the C-3 and C-4 positions, displayed significantly lower activity ( $IC_{50} = 91.4$ ). Among the tri-galloyllated analogs, compound **42** without a galloyl unit at the C-4 position showed stronger inhibitory activity (IC<sub>50</sub> = 5.34  $\mu$ M) than analogs 41, 43, and 44 (IC<sub>50</sub> = 6.72, 9.34, and 12.6  $\mu$ M,

respectively). In addition, analog **44** without a galloyl moiety at the C-2 position displayed weak activity. Thus, our results suggested that a galloyl unit at the C-2 position considerably increases the α-glucosidase inhibitory activity, while the presence of this group at the C-4 position causes a decrease in the activity.

Subsequently, we compared maplexin J (**54**) and its analogs (**55** and **56**) to elucidate the effect of the phenolic hydroxyl group. The 3',4'-di-hydroxybenzoyl analog 55 exhibited good inhibitory activity, whereas the 3',5'-di-hydroxybenzoyl analog 56 was a weaker inhibitor of the α-glucosidase enzyme (IC<sub>50</sub> = 3.28, 9.34  $\mu$ M, respectively). Consequently, these results implied that the presence of two adjacent phenolic hydroxyl groups is essential for the desired activity. We then focused on the evaluation of the influence of the hemiacetal hydroxyl and the HHDP functionalities against the α-glucosidase inhibitory activity. The effect of the hemiacetal hydroxyl moiety can be observed by the comparison of the activity of maplexin J (54) and its analog 61 ( $IC_{50} = 1.68$ , 2.56  $\mu$ M, respectively). Furthermore, tellimagrandin I and 67 showed analogous inhibitory activity ( $IC_{50} = 3.37, 3.22 \mu M$ , respectively). Therefore, the obtained outcomes suggested that the presence of a hemiacetal hydroxyl group did not have a significant effect on the α-glucosidase inhibitory activity. Lastly, to examine the influence of the HHDP group, the results for maplexin J (**54**) and the 4,6-*O*-HHDP analog **67** were compared and it transpired that maplexin J (**54**) displayed marginally higher activity than its analog  $67$  (IC<sub>50</sub> = 2.56, 3.22  $\mu$ M, respectively). Likewise, the assessment of the tellimagrandin I activity and the activity of analog **61** without the HHDP group revealed that **61** was a stronger inhibitor than tellimagrandin I (IC<sub>50</sub> = 3.37, 1.68  $\mu$ M, respectively). Intriguingly, our results suggested that the 4,6-*O*-HHDP group has a weakening effect on the α-glucosidase inhibitory activity [\[38,](#page-25-3)[62\]](#page-26-8).

Meanwhile, we also investigated the antioxidant activity of these polyphenols and their analogs. Firstly, the mono-galloylated analogs **31**–**34** showed nearly equivalent antioxidative activity with methyl gallate ( $EC_{50} = 19.4$ –22.9 µM, TE = 2.04–2.42). Moreover, the activity improved as the number of galloyl units increased, with di-galloylated analogs ( $EC_{50} = 11.9-15.2 \mu M$ , TE = 3.32-4.29) exhibiting lower activity than the tri-galloylated analogs ( $EC_{50} = 7.94-10.3 \mu M$ , TE = 4.31–5.75) and the tetra-galloylated analogs **54**, **61** displaying the highest activity ( $EC_{50} = 5.60$ , 6.61  $\mu$ M, TE = 8.33, 6.77, respectively). Unlike the significant increase observed for the  $\alpha$ -glucosidase inhibitory activity, the antioxidant activity increased proportionally to the number of galloyl units. Moreover, the antioxidant activity was not affected by the position of the galloyl units on the carbohydrate core. In addition, the comparison of compounds **54**, **61**, and **67** with tellimagrandin I revealed similar activity ( $EC_{50} = 6.61$ , 5.60, 6.79, and 6.31  $\mu$ M, TE = 6.77, 8.33, 6.44, and 7.04, respectively). This implied that the antioxidative activity was not influenced by the presence of a hemiacetal hydroxyl and the 4,6-*O*-HHDP groups. The presence of the phenolic hydroxyl groups in 3,4-dihydroxybenzoyl analogs **55** and methyl 3,4-dihydroxybenzoate appeared to result in improved antioxidative activity (EC<sub>50</sub> = 6.99  $\mu$ M, TE = 6.54); however, no antioxidant activity was observed for 3,5-dihydroxybenzoyl analog **56** and the monohydroxybenzoyl analogs (**57** and **58)**. Likewise, the activity of methyl 3,4-dihydroxybenzoate was comparable with methyl gallate ( $EC_{50} = 21.1 \mu M$ , TE = 2.20), whereas methyl 3,5-dihydroxybenzoate, methyl 3- and 4-hydroxybenzoate did not exhibit any notable activity. This data therefore suggested that the presence of two adjacent phenolic hydroxyl groups is necessary for the antioxidant activity, as was the case with  $\alpha$ -glucosidase inhibitory activity.

## **4. Conclusions**

We synthesized 21 carbohydrate-based phenolic analogs including a series of compounds containing all possible combinations of galloylation on 1,5-AG. The  $\alpha$ -glucosidase inhibition and antioxidant activities of these compounds were further studied to evaluate the SAR. Our results suggested that the  $\alpha$ -glucosidase inhibitory activity; 1) is significantly enhanced with the increasing number of galloyl units, and changing the position of the galloyl moiety substitution on the 1,5-AG unit tends to affect the activity; particularly, the presence of this functionality at the C-2 position improves the  $\alpha$ -glucosidase inhibition, whereas substitution at the C-4 position reduces it, 2) requires

two adjacent phenolic hydroxyl groups, 3) is not affected by the presence of the biaryl bond on the 4,6-*O*-HHDP group, 4) is not influenced by the hemiacetal hydroxyl functionality on the carbohydrate unit. Moreover, the following trends were determined for the antioxidant activity; 1) the activity is dependent on the number of galloyl units; however, it is not affected by their position, 2) the presence of two adjacent phenolic hydroxyl groups is significant, 3) the activity is not affected by the HHDP group or the hemiacetal hydroxyl group. The  $\alpha$ -glucosidase inhibitory activity is undoubtedly affected by the position of the galloyl group on 1,5-AG. This outcome indicates that the carbohydrate core is not only a store unit for the galloyl moiety but can also act as a carrier to the biological targets. Thus, derivatives modified at the C-2 or C-4 positions on the carbohydrate unit, which is not  $p$ -glucose, have the potential to exhibit stronger antidiabetic activity. The synthesis and profiling of further analogs will be reported in due course.

#### **5. Materials and Methods**

#### *5.1. General Information*

Proton nuclear magnetic resonance  $(^1H$  NMR) spectra were recorded on JEOL JNM-ECX600 spectrometers. Chemical shifts are reported relative to internal standard (tetramethylsilane;  $\delta_H$  0.00, CDCl<sub>3</sub>;  $\delta$ <sub>H</sub> 7.26). Data are presented as follows: chemical shift ( $\delta$ , ppm), multiplicity (s = singlet,  $d =$  doublet,  $t =$  triplet,  $q =$  quartet,  $m =$  multiplet), coupling constant and integration. Carbon nuclear magnetic resonance (<sup>13</sup>C NMR) spectra were recorded on JEOL JNM-ECX600 (150 MHz) spectrometers. The following internal reference was used: (tetramethylsilane: δ 0.00, CDCl<sub>3</sub> δ 77.0, Acetone –  $d_6$ ; δ 29.8, CD3OD; δ 49.0). Optical rotations were measured on a JASCO P-1030 digital polarimeter at the sodium D line (589 nm). Electron impact (EI) mass analyses and fast atom bombardment (FAB) mass analyses were carried out with a JEOL JMS-GCMATE. C.C. was carried out on Kanto Silica gel 60N spherical (63–210 mesh). Analytical thin-layer chromatography (TLC) was carried out using Merck Kieselgel 60 F254 plates with visualization by ultraviolet light or stained by  $8\%$  H<sub>2</sub>SO<sub>4</sub>/EtOH solution on hot-plate. Tellimagrandin I was purchased from Nagara Science (Gifu, Japan). Methyl gallate, methyl 3,4-dihydroxybenzoate, methyl 3,5-dihydroxybenzoate, methyl 3-hydroxybenzoate, methyl 4-hydroxybenzoate, Trolox and MES were purchased from TOKYO CHEMICAL INDUSTRY CO., LTD (Tokyo, Japan). DPPH was purchased from Sigma-Aldrich Japan (Tokyo, Japan).

# *5.2. Chemical Synthesis*

*2-O-Benzyl-4,6-O-benzylidene-1,5-anhydro-*d-*glucitol* (**3**) [\[35\]](#page-25-6) and *3-O-Benzyl-4,6-O-benzylidene-1,5 -anhydro-*b-*glucitol* (4) [\[35\]](#page-25-6); Compound **1** (2.5 g, 10 mmol) and TBAHSO<sub>4</sub> (0.68 g, 2.0 mmol) in 160 mL of DCM and 14 mL of 5% NaOH was stirred at rt. Then, BnBr (2.1 mL, 17 mmol) was slowly added, and the mixture was refluxed for 30 h. After the addition 50 mL of water, the mixture was extracted with DCM ( $3 \times 100$ ). The combined organic layers were washed with brine, dried over Na<sub>2</sub>SO<sub>4</sub>, filtered and concentrated under reduced pressure. The crude product was separated by C.C (Hex/EtOAc =  $4/1$ ) to obtain 2-*O*-Bn compound **3** (2.0 g, 59%) as colorless needles and 3-*O*-Bn compound **4** (1.0 g, 30% yield) as a white solid. Compound 3: m.p. 163 °C;  $[\alpha]_D^{20} = -3.16$  (c 1.0, CHCl<sub>3</sub>); <sup>1</sup>H NMR (CDCl<sub>3</sub>, 600 MHz) δ 7.49–7.47 (m, 2H), 7.38–7.29 (m, 8H), 5.50 (s, 1H), 4.76, 4.67 (ABq, *J* = 11.7 Hz, 2H), 4.30 (dd, *J* = 10.5, 5.0 Hz, 1H), 4.01 (dd, *J* = 11.3, 5.5 Hz, 1H), 3.84 (m, 1H), 3.65 (t, *J* = 10.3 Hz, 1H), 3.59–3.55 (m, 1H), 3.45 (t, *J* = 9.3 Hz, 1H), 3.35 (m, 1H), 3.30 (t, *J* = 11.0 Hz, 1H); <sup>13</sup>C NMR (CDCl3, 150 MHz) δ 138.02, 137.05, 129.23, 128.56, 128.33, 128.03, 127.89, 126.29, 101.88, 81.05, 77.74, 74.81, 73.43, 70.91, 68.79, 68.45. Compound 4: m.p. 137 °C; [α] $^{20}_{\text{D}}$  = +5.3 (c 1.0, CHCl<sub>3</sub>); <sup>1</sup>H NMR (CDCl<sub>3</sub>, 600 MHz) δ 7.50–7.48 (m, 2H), 7.41–7.29 (m, 8H), 5.58 (s, 1H), 5.03, 4.72 (ABq, *J* = 11.3 Hz, 2H), 4.34 (dd, *J* = 10.5, 5.0 Hz, 1H), 4.06 (dd, *J* = 11.2, 5.7 Hz, 1H), 3.80–3.76 (m, 1H), 3.72 (t, *J* = 10.3 Hz, 1H), 3.66 (t, *J* = 9.1 Hz, 1H), 3.58 (t, *J* = 8.8 Hz, 1H), 3.44–3.40 (m, 1H), 3.34 (t, *J* = 10.8 Hz, 1H); <sup>13</sup>C NMR (CDCl3, 150 MHz) δ 138.30, 137.32, 129.00, 128.61, 128.30, 128.13, 128.01, 125.98, 101.21, 82.66, 82.13, 74.71, 71.53, 69.91, 69.79, 68.88.

*2,3-Di-O-benzyl-1,5-anhydro-*d-*glucitol* (**5**) [\[42\]](#page-25-8); Compound **2** (0.86g, 2.0 mmol) in 10 mL of 80% AcOH/H<sub>2</sub>O solution was stirred at 80 °C for 5 h. After the addition 5 mL of saturated aq. NaHCO<sub>3</sub>, the reaction solution was extracted by EtOAc  $(3 \times 30)$ . The organic layer was washed with brine, dried over  $Na<sub>2</sub>SO<sub>4</sub>$ , filtered and concentrated under reduced pressure. The crude was purified by recrystallization (EtOAc/Hex) to obtain **5** (0.65 g, 95%) as colorless needles. m.p. 129 °C;  $[\alpha]_D^{20} = -10.4$ (c 1.00, CHCl3); <sup>1</sup>H NMR (CDCl3, 600 MHz) δ 7.43–7.29 (m, 10H), 5.03, 4.70 (ABq, *J* = 11.7 Hz, 2H), 4.64, 4.03 (ABq, *J* = 11.3, 2H), 3.86–3.82 (m, 1H), 3.72–3.68 (m, 1H), 3.63–3.59 (m, 1H), 3.50–3.44 (m, 2H), 3.28–3.23 (m, 2H); <sup>13</sup>C NMR (CDCl3, 150 MHz) δ 138.54, 137.97, 128.67, 128.53, 127.98, 127.93, 127.83, 85.30, 79.53, 78.29, 75.10, 73.07, 70.45, 67.94, 62.89.

*1,5-Anhydro-2-O-benzyl-*d-*glucitol* (**6**) [\[35\]](#page-25-6); Compound **3** (860 mg, 2.5 mmol) in 10 mL of 80% AcOH/H<sub>2</sub>O solution was stirred at 80 °C for 5 h. By the same procedure previously described for the preparation of compound **5**, compound **6** (503 mg, 76%) was obtained as colorless needles. m.p. 131 ◦C;  $[\alpha]_D^{20}$  = +8.4 (c 1.00, MeOH); <sup>1</sup>H NMR (CD<sub>3</sub>OD, 600 MHz); δ 7.40–7.36 (m, 2H), 7.33–7.31 (m, 2H), 7.28–7.25 (m, 1H), 4.75, 4.64 (ABq, *J* = 11.7 Hz, 2H), 3.96 (dd, *J* = 11.0, 5.2 Hz, 1H), 3.81 (dd, *J* = 11.9, 2.2 Hz, 1H), 3.59 (dd, *J* = 11.9, 6.0 Hz, 1H), 3.43 (t, *J* = 8.9 Hz, 1H), 3.39–3.35 (m, 1H), 3.26–3.22 (m, 1H), 3.15–3.11 (m, 2H); <sup>13</sup>C NMR (CD3OD, 150 MHz); δ 140.09, 129.37, 129.05, 128.76, 82.42, 79.31, 79.27, 74.14, 71.92, 68.95, 63.07.

*1,5-Anhydro-3-O-benzyl-*d-*glucitol* (**7**); Compound **4** (680 mg, 2.0 mmol) in 10 mL of 80% AcOH/H2O solution was stirred at 80 °C for 5 h. By the same procedure previously described for the preparation of compound **5**, compound **7** (410 mg, 80%) was obtained as colorless needles. m.p. 153 °C;  $[\alpha]_D^{20} = +28.6$ (c 1.00, MeOH); <sup>1</sup>H NMR (CD<sub>3</sub>OD, 600 MHz);  $\delta$  7.44 (m, 2H), 7.32–7.30 (m, 2H), 7.26–7.23 (m, 1H), 4.90 (overlap, 2H), 3.89 (dd, *J* = 11.3, 5.5 Hz, *1*H), 3.83 (dd, *J* = 11.9, 2.2 Hz, 1H), 3.62–3.57 (m, 2H), 3.37  $(dd, J = 18.0, 8.71 \text{ Hz}, 1\text{H}$ ), 3.31–3.28 (overlap, 1H), 3.22–3.16 (m, 2H); <sup>13</sup>C NMR (CD<sub>3</sub>OD, 150 MHz);  $\delta$  140.53, 129.20, 129.09, 128.49, 88.20, 82.68, 76.10, 71.67, 71.54, 71.09, 63.05. HRMS (ESI,  $m/z$ ):  $[M + Na]$ <sup>+</sup>, calcd for  $[C_{13}H_{18}O_5\,Na]^+$ : 277.1052; found 277.1052.

1,5-Anhydro-4-O-benzyl-p-glucitol (8) [\[35\]](#page-25-6); Compound 1 (0.76 g, 3.0 mmol) in 15 mL of DCM was stirred at 0 °C. Then, 15 mL of borane-THF (ca. 1M THF solution) and TMSOTf (0.11 mL, 0.60 mmol) were successively added to the mixture. The mixture was allowed to stir for 4 h and added MeOH carefully. After the addition 1 mL of saturated aq. NaHCO<sub>3</sub>, the reaction solution was extracted with DCM (5  $\times$  40 mL). The combined organic layer was dried over Na<sub>2</sub>SO<sub>4</sub>, filtered and concentrated under reduced pressure. The crude product was purified by C.C. (CHCl<sub>3</sub>/MeOH = 8/1) to obtain 8 (0.64 g, 74%) as a white solid.  $[\alpha]_D^{20}$  = +27.7 (c 0.45, CHCl<sub>3</sub>); <sup>1</sup>H NMR (CD<sub>3</sub>OD, 600 MHz) δ 7.38–7.25 (m, 5H), 4.94, 4.64 (ABq, *J* = 11.0 Hz, 2H), 3.89 (dd, *J* = 13.3, 5.4 Hz, 1H), 3.78 (dd, *J* = 12.0, 2.1 Hz, 1H), 3.60 (dd, 1H, *J* = 11.4, 5.73 Hz,), 3.49–3.46 (m, 2H), 3.33–3.30 (overlap, 1H), 3.20 (ddd, *J* = 9.7, 5.2, 2.1 Hz, 1H), 3.14 (t, *J* = 10.7, 1H); <sup>13</sup>C NMR (CD<sub>3</sub>OD, 150 MHz); δ 140.09, 129.33, 129.14, 128.70, 81.75, 80.43, 79.50, 75.90, 71.77, 70.93, 62.72.

*2,4-Di-O-benzyl-1,5-anhydro-*d-*glucitol* (**9**) [\[46\]](#page-25-12); Compound **3** (1.0 g, 3.0 mmol) in 15 mL of DCM was successively added 15 mL of borane-THF (ca. 1M THF solution) and TMSOTf (0.11 mL, 0.6 mmol) at 0 °C. The mixture was allowed to stir at rt for 10 h and added 1 mL of MeOH carefully. After the addition 1 mL of saturated aq. NaHCO<sub>3</sub>, the mixture was extracted with DCM ( $3 \times 40$  mL). The organic layer was washed with brine, dried over Na<sub>2</sub>SO<sub>4</sub>, filtered and concentrated under reduced pressure. The crude was purified by recrystallization (Hex/EtOAc) to obtain **9** (820 mg, 82%) as colorless needles. m.p. 112 °C;  $[\alpha]_D^{20}$  = +34.6 (c 1.00, CHCl<sub>3</sub>); <sup>1</sup>H NMR (CDCl<sub>3</sub>, 600 MHz)  $\delta$  7.37–7.28 (m, 10H), 4.86, 4.70 (ABq, *J* = 11.3 Hz, 2H), 4.65 (s, 2H), 3.99 (dd, *J* = 11.3, 5.2 Hz, 1H), 3.84 (ddd, *J* = 11.8, 5.8, 2.7 Hz, 1H), 3.74 (td, *J* = 8.9, 2.1 Hz, 1H), 3.68–3.64 (m, 1H), 3.45–3.41 (m, 1H), 3.41 (t, *J* = 9.2 Hz, 1H), 3.26 (ddd, *J* = 9.6, 4.5, 2.7 Hz, 1H), 3.19 (t, *J* = 10.8 Hz, 1H); <sup>13</sup>C NMR (CDCl3, 150 MHz) δ 138.18, 138.00, 128.60, 128.56, 128.09, 127.98, 127.85, 79.34, 78.18, 77.97, 77.39, 74.74, 73.05, 67.44, 62.27.

*3,4-Di-O-benzyl-1,5-anhydro-*d-*glucitol* (**10**) [\[47\]](#page-25-13); Compound **4** (850 g, 2.5 mmol) in 15 mL of DCM was successively added 12.5 mL of borane-THF (ca. 1M THF solution) and TMSOTf (90  $\mu$ L, 0.5 mmol) at 0 ◦C. The mixture was allowed to stir at rt for 7 h and added 5 mL of MeOH carefully. After the

addition 1 mL of saturated aq. NaHCO<sub>3</sub>, the mixture was extracted with DCM ( $3 \times 40$  mL). The organic layer was washed with brine, dried over Na<sub>2</sub>SO<sub>4</sub>, filtered and concentrated under reduced pressure. The crude was purified by recrystallization (Hex/EtOAc) to obtain **10** (523 mg, 61%) as colorless needles. m.p. 99 °C; [α] $_{\text{D}}^{20}$  = +48.3 (c 1.40, CHCl<sub>3</sub>); <sup>1</sup>H NMR (CDCl<sub>3</sub>, 600 MHz) δ 7.38–7.30 (m, 10H), 4.97, 4.77 (ABq, *J* = 11.3 Hz, 2H), 4.86, 4.68 (ABq, *J* = 11.0 Hz, 2H), 3.98 (dd, *J* = 11.2, 5.3 Hz, 1H), 3.84 (ddd, *J* = 11.9, 5.5, 2.6 Hz, 1H), 3.71–3.65 (m, 2H), 3.52 (t, *J* = 9.1 Hz, 1H), 3.47 (t, *J* = 8.8 Hz, 1H), 3.32–3.29 (m, 1H), 3.23 (t, *J* = 10.8 Hz, 1H); <sup>13</sup>C NMR (CDCl3, 150 MHz) δ 138.45, 137.84, 128.71, 128.56, 128.02, 127.86, 86.73, 79.92, 77.79, 75.25, 74.95, 70.16, 69.30, 61.99.

*2,3,4-Tri-O-benzyl-1,5-anhydro-*d-*glucitol* (**11**) [\[46\]](#page-25-12); Compound **2** (860 mg, 2.0 mmol) in 10 mL of toluene was stirred at rt. The reaction solution was added DIBAL-H (ca. 1M toluene solution, 6 mL) and stirred at rt for 20 h. The reaction solution was slowly added 4.2 mL of MeOH and 7.2 mL of 30% Rochelle salt solution and stirred for 1 h. After the addition 20 mL of EtOAc, the mixture was extracted with 30% Rochelle salt solution  $(3 \times 15 \text{ mL})$ . The organic layer was washed with brine, dried over Na<sub>2</sub>SO<sub>4</sub>, filtered and concentrated under reduced pressure. The crude was purified by C.C.  $(Hex/EtOAc = 4/1–2/1)$  to obtain **11** (760 mg, 90% yield) as colorless needles. m.p. 83 °C;  $[\alpha]_D^{20} = +8.9$ (c 0.90, CHCl3); <sup>1</sup>H NMR (CDCl3, 600 MHz) δ 7.37–7.28 (m, 15H), 4.98–4.64 (m, 6H), 3.99 (dd, *J* = 11.3, 5.2 Hz, 1H), 3.84–3.81 (m, 1H), 3.67–3.58 (m, 3H), 3.48 (t, *J* = 9.3 Hz, 1H), 3.29–3.25 (m, 1H), 3.23 (t, *J* = 10.8 Hz, 1H); <sup>13</sup>C NMR (CDCl3, 150 MHz) δ 138.61, 138.09, 138.01, 128.48, 128.41, 128.05, 127.89, 127.83, 127.64, 86.17, 79.69, 78.55, 77.57, 75.55, 75.13, 73.34, 67.96, 62.25.

*1,5-Anhydro-6-O-benzyl-*d-*glucitol* (**12**) [\[35\]](#page-25-6); Compound **1** (760 mg, 3.0 mmol) in 15 mL of DCM was added triethylsilane (2.4 mL, 15 mmol) and trifluoracetic acid (1.2 mL, 15 mmol) at 0 ◦C. The reaction solution was stirred at rt for 8 h. After the addition 10 mL of saturated aq. NaHCO<sub>3</sub>, the mixture was extracted with DCM ( $5 \times 30$  mL). The combined organic layer was dried over Na<sub>2</sub>SO<sub>4</sub>, filtered and concentrated under reduced pressure. The crude was purified by C.C (CHCl<sub>3</sub>/MeOH =  $100/1-10/1$ ) to obtain **12** (471 mg, 62%) as a colorless oil.  $[\alpha]_D^{20} = +8.9$  (c 0.30, CHCl<sub>3</sub>); <sup>1</sup>H NMR (CDCl<sub>3</sub>, 600 MHz) δ 7.35–7.25 (m, 5H), 4.55 (d, *J* = 2.1 Hz, 2H), 3.88 (dd, *J* = 11.0, 5.5 Hz, 1H), 3.77 (dd, *J* = 10.8, 1.9 Hz, 1H), 3.60 (dd, *J* = 10.8, 5.7 Hz, 1H), 3.48–3.42 (m, 1H), 3.32–3.24 (overlap, 3H), 3.15 (t, *J* = 10.8 Hz, 1H); <sup>13</sup>C NMR (CDCl<sub>3</sub>, 150 MHz) δ 139.57, 129.34, 128.90, 128.67, 81.38, 79.95, 74.48, 71.85, 71.31, 71.19, 70.95.

*2,3,6-Tri-O-benzyl-1,5-anhydro-*d-*glucitol* (**13**) [\[49\]](#page-25-15); Compound **2** (860 mg, 2.0 mmol) in 10 mL of DCM was added triethylsilane (1.6 mL, 10 mmol) and tifluoroacetic acid (0.8 mL, 10 mmol) at 0 ◦C. The reaction solution was stirred at rt for 15 h. After the addition 5 mL of saturated aq. NaHCO<sub>3</sub>, the mixture was extracted with DCM  $(3 \times 30 \text{ mL})$ . The organic layer was washed with brine, dried over Na2SO4, filtered and concentrated under reduced pressure. The crude product was purified by C.C. (Hex/EtOAc = 4/1) to obtain **13** (505 mg, 59%) as a colorless oil.  $[\alpha]_D^{20} = -7.0$  (c 1.00, CHCl<sub>3</sub>); <sup>1</sup>H NMR (CDCl3, 600 MHz) δ 7.37–7.25 (m, 15H), 5.00, 4.76 (ABq, *J* = 11.5 Hz, 2H), 4.69, 4.63 (ABq, *J* = 11.7 Hz, 2H), 4.58, 4.54 (ABq, *J* = 12.2 Hz, 2H), 4.04 (dd, *J* = 11.3, 5.2 Hz, 1H), 3.70 (dd, *J* = 10.5, 3.3 Hz, 1H), 3.64–3.60 (m, 2H), 3.54 (td, *J* = 9.2, 2.1 Hz, 1H), 3.44 (t, *J* = 8.9 Hz, 1H), 3.37–3.34 (m, 1H), 3.23 (t, *J* = 11.0 Hz, 1H); <sup>13</sup>C NMR (CDCl3, 150 MHz) δ 138.64, 138.05, 137.84, 128.58, 128.48, 128.39, 127.95, 127.90, 127.87, 127.83, 127.77, 127.70, 85.40, 78.75, 78.04, 75.10, 73.65, 73.05, 70.87, 69.98, 68.06.

*2,6-Di-O-benzyl-1,5-anhydro-*d-*glucitol* (**14**); Compound **3** (860 mg, 2.5 mmol) in 15 mL of DCM was added triethylsilane (2.0 mL, 12.5 mmol) and tifluoroacetic acid (1.0 mL, 12.5 mmol) at 0 ◦C. The reaction solution was stirred at rt for 8 h. After the addition 10 mL of saturated aq. NaHCO<sub>3</sub>, the mixture was extracted with DCM  $(3 \times 40 \text{ mL})$ . The organic layer was washed with brine, dried over  $Na<sub>2</sub>SO<sub>4</sub>$ , filtered and concentrated under reduced pressure. The crude product was purified by C.C. (CHCl<sub>3</sub>/MeOH = 8/1) to obtain **14** (500 mg, 58%) as a white solid.  $[\alpha]_D^{20} = +17.4$  (c 1.00, CHCl<sub>3</sub>); <sup>1</sup>H NMR (CDCl3, 600 MHz) δ 7.36–7.27 (m, 10H), 4.64 (s, 2H), 4.59, 4.54 (ABq, *J* = 12.0 Hz, 2H), 4.02 (dd, *J* = 11.2, 5.0 Hz, 1H), 3.69 (dd, *J* = 10.5, 3.6 Hz, 1H), 3.65 (dd, *J* = 10.5, 5.0 Hz, 1H), 3.55 (td, *J* = 8.8, 2.1 Hz, 1H), 3.50 (td, *J* = 9.0, 2.6 Hz, 1H), 3.47–3.43 (m, 1H), 3.37–3.34 (m, 1H), 3.19 (t, *J* = 10.8 Hz, 1H); <sup>13</sup>C NMR (CDCl3, 150 MHz) δ 137.96, 137.71, 128.59, 128.46, 128.07, 127.90, 127.81, 78.31, 77.74, 77.41,

73.68, 72.95, 71.34, 70.01, 67.70. HRMS (ESI, *m/z*):  $[M + Na]^+$ , calcd for  $[C_{20}H_{24}O_5Na]^+$ : 367.1521; found 367.1521.

*1,5-Anhydro-3,6-di-O-benzyl-*d-*glucitol* (**15**); Compound **4** (680 mg, 2.0 mmol) in 15 mL of DCM was added triethylsilane (1.6 mL, 10 mmol) and trifluoroacetic acid (0.80 mL, 10 mmol) at 0 ◦C. The reaction solution was stirred at rt for 4 h. After the addition 10 mL of saturated aq. NaHCO<sub>3</sub>, the mixture was extracted with DCM ( $3 \times 40$  mL). The combined organic layer was washed with brine, dried over Na2SO4, filtered and concentrated under reduced pressure. The crude product was purified by C.C. (CHCl<sub>3</sub>/MeOH = 8/1) to obtain **15** (420 mg, 62%) as a white solid.  $[\alpha]_D^{20} = +22.7$  (c 1.00, CHCl3); <sup>1</sup>H NMR (CDCl3, 600 MHz) δ 7.39–7.27 (m, 10H), 4.91, 4.83 (ABq, *J* = 11.7 Hz, 2H), 4.59, 4.54 (ABq, *J* = 12.2 Hz, 2H), 3.96 (dd, *J* = 11.2, 5.3 Hz, 1H), 3.72–3.66 (m, 3H), 3.61 (dt, *J* = 9.1, 2.6 Hz, 1H), 3.39–3.36 (m, 1H), 3.31 (t, *J* = 8.8 Hz, 1H), 3.21 (t, *J* = 10.8 Hz, 1H); <sup>13</sup>C NMR (CDCl3, 150 MHz) δ 138.60, 137.61, 128.70, 128.47, 128.02, 127.93, 127.86, 127.82, 86.47, 78.47, 74.87, 73.74, 72.11, 70.38, 69.79, 69.57. HRMS (ESI, *m*/z):  $[M + Na]$ <sup>+</sup>, calcd for  $[C_{20}H_{24}O_5Na]$ <sup>+</sup>: 367.1521; found 367.1522.

*1,5-Anhydro-2-O-(3*<sup>0</sup> *,4*0 *,5*0 *-tribenzyloxybenzoyl)-3-O-benzyl-4,6-O-benzylidene-*d-*glucitol* (**17**); Compound **3** (340 mg, 1.0 mmol), compound **16** [\[39\]](#page-25-4) (650 mg, 1.5 mmol), 2-chloro-1-methylpyridinium iodide (380 mg, 1.5 mmol), DMAP (37 mg, 0.30 mmol), TEA (416 µL, 3.0 mmol) in 15 mL of DCM was stirred at rt for 20 h. After the addition 100 mL of saturated aq. NH4Cl, the reaction solution was extracted with DCM ( $3 \times 60$  mL). The combined organic layer was washed with brine, dried over Na<sub>2</sub>SO<sub>4</sub>, filtered and concentrated under reduced pressure. The crude was purified by C.C. (DCM/MeOH = 500/1 − 200/1) to obtain **17** (350 mg, 41%) as a colorless amorphous oil.  $[\alpha]_D^{20} = +8.4$  (c 1.04, CHCl<sub>3</sub>); <sup>1</sup>H NMR (CDCl<sub>3</sub>, 600 MHz) δ 7.53–7.11 (m, 29H), 5.61 (s, 1H), 5.27–5.20 (m, 1H), 5.15 (s, 2H), 5.09 (s, 4H), 4.86, 4.71 (ABq, *J* = 12.0 Hz, 2H), 4.36 (dd, *J* = 10.3, 4.8 Hz, 1H), 4.19 (dd, *J* = 11.0, 5.8 Hz, 1H), 3.87 (t, *J* = 9.1 Hz, 1H), 3.79–3.74 (m, 2H), 3.45 (td, *J* = 9.7, 4.9 Hz, 1H), 3.36 (t, *J* = 10.8 Hz, 1H); <sup>13</sup>C NMR (CDCl3, 150 MHz) δ 165.06, 152.55, 142.74, 138.10, 137.31, 137.28, 136.56, 129.03, 128.57, 128.30, 128.25, 128.22, 128.07, 128.02, 127.89, 127.58, 127.47, 126.02, 124.50, 109.41, 101.32, 81.96, 79.12, 75.14, 74.33, 71.50, 71.30, 68.76, 67.68; HRMS (ESI, *m*/z):  $[M + Na]$ <sup>+</sup>, calcd for  $[C_{48}H_{44}O_9Na]$ <sup>+</sup>: 787.2883; found 787.2881.

*1,5-Anhydro-2-O-benzyl-3-O-(3*<sup>0</sup> *,4*0 *,5*0 *-tribenzyloxybenzoyl)-4,6-O-benzylidene-*d-*glucitol* (**18**); Compound **2** (0.51 g, 1.5 mmol), compound **16** (1.0 g, 2.3 mmol), 2-chloro-1-methylpyridinium iodide (0.59 g, 2.3 mmol), DMAP (28 mg, 0.23 mmol), TEA (0.62 mL, 4.5 mmol) in 20 mL of DCM was stirred at rt for 20 h. By the same procedure previously described for the preparation of compound **17**, compound **18** (0.87 g, 68%) was obtained as a colorless amorphous oil.  $[\alpha]_D^{20} = -45.6$  (c 0.95, CHCl<sub>3</sub>); <sup>1</sup>H NMR (CDCl3, 600 MHz) δ 7.24–7.44 (m, 24H), 7.20–7.14 (m, 5H), 5.53 (t, *J* = 9.3 Hz, 1H), 5.46 (s, 1H), 5.15–5.10 (m, 6H), 4.55, 4.46 (ABq, *J* = 12.4 Hz, 2H), 4.35 (dd, *J* = 10.5, 5.0 Hz, 1H), 4.13 (dd, *J* = 11.3, 5.5 Hz, 1H), 3.76–3.72 (m, 1H), 3.70 (t, *J* = 10.3 Hz, 1H), 3.65 (t, *J* = 9.5 Hz, 1H), 3.51 (td, *J* = 9.7, 4.8 Hz, 1H), 3.48 (t, *J* = 11.0 Hz, 1H); <sup>13</sup>C NMR (CDCl3, 150 MHz) δ 165.16, 152.47, 142.58, 137.53, 137.39, 136.69, 128.95, 128.54, 128.38, 128.21, 128.16, 128.02, 127.98, 127.87, 127.53, 126.14, 125.05, 109.55, 101.35, 79.18, 75.62, 75.15, 74.94, 72.97, 71.48, 71.33, 68.80, 68.74; HRMS (ESI, *m/z*):  $\left[\text{M} + \text{Na}\right]^+$ , calcd for  $\left[\text{C}_{48}\text{H}_{44}\text{O}_9\text{Na}\right]^+$ : 787.2883; found 787.2882.

*1,5-Anhydro-2,3,6-tris-O-benzyl-4-O-(3*<sup>0</sup> *,4*0 *,5*0 *-tribenzyloxybenzoyl)-*d-*glucitol* (**19**); Compound **13** (340 mg, 0.78 mmol), compound **16** (530 mg, 1.2 mmol), 2-chloro-1-methylpyridinium iodide (307 mg, 1.2 mmol), DMAP (95 g, 0.78 mmol), TEA (315  $\mu$ L, 2.3 mmol) in 15 mL of DCM was stirred at rt for 2 d. By the same procedure previously described for the preparation of compound **17**, compound **19** (607 mg, 91%) was obtained as a colorless amorphous oil.  $[\alpha]_D^{20} = -32.0$  (c 0.95, CHCl<sub>3</sub>); <sup>1</sup>H NMR (CDCl3, 600 MHz) δ 7.44–7.26 (m, 20H), 7.23–7.15 (m, 7H), 7.11–7.03 (m, 5H), 5.18–5.15 (m, 3H), 5.13–5.07 (m, 4H), 4.76, 4.54 (ABq, *J* = 11.3 Hz, 2H), 4.74, 4.65 (ABq, *J* = 11.7 Hz, 1H), 4.45 (s, 2H), 4.08 (dd, *J* = 11.3, 5.2 Hz, 1H), 3.75–3.71 (m, 1H), 3.65 (t, *J* = 9.1 Hz, 1H), 3.59–3.56 (m, 1H), 3.50 (dd, *J* = 10.8, 2.6 Hz, 1H), 3.45 (dd, *J* = 10.7, 5.8 Hz, 1H), 3.29 (t, *J* = 11.0 Hz, 1H); <sup>13</sup>C NMR (CDCl3, 150 MHz) δ 164.83, 152.41, 142.58, 138.03, 137.58, 137.37, 136.62, 128.57, 128.50, 128.23, 128.14, 128.06, 128.01, 127.94, 127.86, 127.83, 127.58, 127.49, 127.45, 124.67, 109.34, 83.08, 78.18, 78.08, 75.13, 74.94, 73.67, 73.40, 71.24, 69.47, 68.23; HRMS (ESI, *m/z*):  $[M + Na]$ <sup>+</sup>, calcd for  $[C_{55}H_{52}O_9Na]$ <sup>+</sup>: 879.3509; found 879.3511.

*1,5-Anhydro-2,3,4-tris-O-benzyl-6-O-(3*<sup>0</sup> *,4*0 *,5*0 *-tribenzyloxybenzoyl)-*d-*glucitol* (**20**); Compound **11** (0.43 g, 1.0 mmol), compound **16** (0.66 g, 1.5 mmol), 2-chloro-1-methylpyridinium iodide (0.38 g, 1.5 mmol), DMAP (0.18 g, 1.5 mmol), TEA (0.42 mL, 3.0 mmol) in 20 mL of DCM was stirred at rt for 2 d. By the same procedure previously described for the preparation of compound **17**, compound **20** (0.50 g, 58%) was obtained as a colorless amorphous oil.  $[\alpha]_D^{20} = +27.9$  (c 1.00, CHCl<sub>3</sub>); <sup>1</sup>H NMR (CDCl3, 600 MHz) δ 7.43–7.18 (m, 32H), 5.14–5.10 (m, 6H), 5.03, 4.88 (ABq, *J* = 10.7 Hz, 2H), 4,85, 4.53 (ABq, *J* = 11.8 Hz, 2H), 4.74, 4.67 (ABq, *J* = 11.7 Hz, 2H), 4.52 (dd, *J* = 12.0, 2.1 Hz, 1H), 4.37 (dd, *J* = 12.0, 4.5 Hz, 1H), 4.03 (dd, *J* = 11.3, 5.2 Hz, 1H), 3.69 (t, *J* = 8.8 Hz, 1H), 3.66–3.62 (m, 1H), 3.52–3.49 (m, 1H), 3.46 (t, *J* = 8.9 Hz, 1H), 3.23 (t, *J* = 10.8 Hz, 1H); <sup>13</sup>C NMR (CDCl3, 150 MHz) δ 165.82, 152.41, 142.46, 138.55, 138.04, 137.69, 137.37, 136.65, 128.59, 128.53, 128.48, 128.46, 128.20, 128.08, 128.01, 127.96, 127.93, 127.85, 127.76, 127.45, 124.91, 109.30, 86.33, 78.48, 77.83, 77.57, 75.73, 75.23, 75.09, 73.29, 71.17, 68.12, 63.86; HRMS (ESI, *m*/z):  $[M + Na]$ <sup>+</sup>, calcd for  $[C_{55}H_{52}O_9Na]$ <sup>+</sup>: 879.3509; found 879.3509.

*1,5-Anhydro-2,3-bis-O-(3*<sup>0</sup> *,4*0 *,5*0 *-tribenzyloxybenzoyl)-4,6-O-benzylidene-*d-*glucitol* (**21**); Compound **1** (380 mg, 1.5 mmol), compound **16** (2.0 g, 4.5 mmol), 2-chloro-1-methylpyridinium iodide (1.2 g, 4.5 mmol), DMAP (0.55 g, 4.5 mmol), TEA (1.3 mL, 9.0 mmol) in 20 mL of DCM was stirred at rt for 2 d. By the same procedure previously described for the preparation of compound **17**, compound **21** (1.6 g, 88%) was obtained as a colorless amorphous oil.  $[\alpha]_D^{20}$  = +46.4 (c 1.00, CHCl<sub>3</sub>); <sup>1</sup>H NMR ( CDCl<sub>3</sub>, 600 MHz) δ 7.44–7.42 (m, 6H), 7.34–7.31 (m, 26H), 7.25–7.22 (m, 4H), 5.80 (t, *J* = 9.5 Hz, 1H), 5.56 (s, 1H), 5.27–5.23 (m, 1H), 5.11–4.93 (m, 12H), 4.45–4.41 (m, 2H), 3.87 (t, *J* = 16.0 Hz, 1H), 3.82 (t, *J* = 17.0 Hz, 1H), 3.65–3.62 (m, 1H), 3.55 (t, *J* = 10.5 Hz, 1H); <sup>13</sup>C NMR (CDCl3, 150 MHz) δ 165.6, 165.4, 152.6, 142.9, 142.8, 137.4, 136.8, 136.5, 129.1, 128.5, 128.4, 128.3, 128.2, 128.1, 128.0, 127.5, 126.2, 109.3, 109.1, 101.6, 78.8, 75.1, 73.0, 71.9, 71.2, 71.2, 71.1, 68.7, 67.7; HRMS (ESI, *m/*z):  $[M + Na]^+$ , calcd for  $[C_{48}H_{44}O_9\,Na]^+$ : 1119.3932; found 1119.3901.

*1,5-Anhydro-2,4-bis-O-(3*<sup>0</sup> *,4*0 *,5*0 *-tribenzyloxybenzoyl)-3,6-bis-O-benzyl-*d-*glucitol* (**22**); Compound **15** (0.30 mg, 0.87 mmol), compound **16** (1.3 g, 3.0 mmol), 2-chloro-1-methylpyridinium iodide (0.77 g, 3.0 mmol), DMAP (0.37 g, 3.0 mmol), TEA (0.83 mL, 6.0 mmol) in 20 mL of DCM was stirred at rt for 2 d. By the same procedure previously described for the preparation of compound **17**, compound **22** (0.93 g, 77%) was obtained as a colorless amorphous oil.  $[\alpha]_D^{20} = +7.87$  (c 1.00, CHCl<sub>3</sub>); <sup>1</sup>H NMR <sup>1</sup>H NMR (CDCl<sub>3</sub>, 600 MHz) δ 7.58–7.14 (m, 45H), 7.08–6.95 (m, 5H), 5.32–5.25 (m, 2H), 5.19–5.08 (m, 13H), 4.61–4.45 (m, 4H), 4.29 (dd, *J* = 11.3, 5.5 Hz, 1H), 3.94 (t, *J* = 9.1 Hz, 1H), 3.72–3.68 (m, 1H), 3.58–3.53 (m, 2H), 3.39 (t, *J* = 10.8 Hz, 1H); <sup>13</sup>C NMR (CDCl3, 150 MHz) δ 164.87, 164.71, 152.61, 152.51, 142.88, 142.79, 137.54, 137.52, 137.32, 137.27, 136.56, 136.49, 128.59, 128.57, 128.55, 128.28, 128.24, 128.22, 128.16, 128.10, 128.05, 128.02, 127.85, 127.76, 127.65, 127.53, 127.49, 124.51, 109.43, 80.72, 78.29, 75.16, 74.16, 73.72, 72.12, 71.35, 71.32, 69.44, 67.03; HRMS (ESI,  $m/z$ ):  $[M+Na]^+$ , calcd for  $[C_{76}H_{68}O_{13}Na]^+$ : 1211.4558; found 1211.4554.

1,5-Anhydro-2,6-bis-O-(3',4',5'-tribenzyloxybenzoyl)-3,4-bis-O-benzyl-D-glucitol (23); Compound 10 (0.45 g, 1.3 mmol), compound **16** (1.8 g, 4.0 mmol), 2-chloro-1-methylpyridinium iodide (1.0 g, 4.0 mmol), DMAP (0.49  $g$ , 4.0 mmol), TEA (1.1 mL, 8.0 mmol) in 30 mL of DCM was stirred at rt for 2 d. By the same procedure previously described for the preparation of compound **17**, compound **23** (1.4 g, 92%) was obtained as a colorless amorphous oil.  $\alpha|_{D}^{20}$  = +51.2 (c 1.00, CHCl<sub>3</sub>); <sup>1</sup>H NMR (CDCl<sub>3</sub>, 600 MHz) δ 7.43–7.18 (m, 44H), 5.27–5.21 (m, 1H), 5.17–5.07 (m, 12H), 4.82, 4.52 (ABq, *J* = 11.0 Hz, 2H), 4.77, 4.70 (ABq, *J* = 11.3 Hz, 2H), 4.56 (m, 1H), 4.42–4.39 (m, 1H), 4.19 (dd, *J* = 11.2, 5.7 Hz, 1H), 3.85–3.82 (m, 1H), 3.60–3.56 (m, 2H), 3.31 (t, *J* = 10.7 Hz, 1H); <sup>13</sup>C NMR (CDCl3, 150 MHz) δ 165.80, 165.05, 152.56, 152.43, 142.76, 142.47, 137.88, 137.49, 137.37, 137.28, 136.66, 136.53, 128.62, 128.60, 128.57, 128.51, 128.39, 128.21, 128.12, 128.06, 128.01, 127.88, 127.80, 127.40, 127.37, 124.80, 124.49, 109.32, 109.25, 84.32, 77.91, 77.42, 75.45, 75.26, 75.13, 75.09, 72.22, 71.26, 71.15, 67.11, 63.52; HRMS (ESI, *m*/*z*): [M + Na] + , calcd for  $[C_{76}H_{68}O_{13}Na]^+$ : 1211.4558; found 1211.4550.

*1,5-Anhydro-2,6-bis-O-benzyl-3,4-bis-O-(3*<sup>0</sup> *,4*0 *,5*0 *-tribenzyloxybenzoyl)-*d-*glucitol* (**24**); Compound **14** (380 mg, 1.1 mmol), compound **16** (1.5 g, 3.3 mmol), 2-chloro-1-methylpyridinium iodide (0.84 g, 3.3 mmol), DMAP (0.91 g, 3.3 mmol), TEA (0.91 mL, 6.6 mmol) in 30 mL of DCM was stirred at rt for

2 d. By the same procedure previously described for the preparation of compound **17**, compound **24** (1.2 g, 92%) was obtained as a colorless amorphous oil.  $[\alpha]_D^{20}$  = -80.8 (c 1.00, CHCl<sub>3</sub>); <sup>1</sup>H NMR (CDCl<sub>3</sub>, 600 MHz) δ 7.41–7.14 (m, 44H), 5.58 (t, *J* = 9.5 Hz, 1H), 5.31 (t, *J* = 9.8 Hz, 1H), 5.09–5.00 (m, 12H), 4.54, 4.48 (ABq, *J* = 12.2 Hz, 2H), 4.52, 4.47 (ABq, *J* = 12.0 Hz, 2H), 4.19 (dd, *J* = 11.5, 5.3 Hz, 1H), 3.79 (td, *J* = 9.9, 5.2 Hz, 1H), 3.73 (m, 1H), 3.59 (dd, *J* = 10.7, 2.4 Hz, 1H), 3.51 (dd, *J* = 10.8, 5.3 Hz, 1H), 3.46 (t, *J* = 11.0 Hz, 1H); <sup>13</sup>C NMR (CDCl3, 150 MHz) δ 165.62, 165.18, 152.43, 142.67, 142.53, 137.56, 137.48, 137.43, 137.39, 136.57, 136.56, 128.49, 128.41, 128.38, 128.36, 128.27, 128.16, 127.98, 127.89, 127.87, 127.84, 127.65, 127.55, 124.67, 124.20, 109.15, 77.99, 76.35, 75.22, 75.10, 75.08, 73.70, 72.96, 71.12, 70.07, 69.06, 68.30; HRMS (ESI,  $m/z$ ):  $[M + Na]$ <sup>+</sup>, calcd for  $[C_{76}H_{68}O_{13}Na]$ <sup>+</sup>: 1211.4558; found 1211.4558.

*1,5-Anhydro-2,4-bis-O-benzyl-3,6-bis-O-(3*<sup>0</sup> *,4*0 *,5*0 *-tribenzyloxybenzoyl)-*d-*glucitol* (**25**); Compound **9** (0.52 mg, 1.5 mmol), compound **16** (2.0 g, 4.5 mmol), 2-chloro-1-methylpyridinium iodide (1.2 g, 4.5 mmol), DMAP (0.55 g, 4.5 mmol), TEA (1.2 mL, 9.0 mmol) in 20 mL of DCM was stirred at rt for 2 d. By the same procedure previously described for the preparation of compound **17**, compound **25** (1.6 g, 91%) was obtained as a colorless amorphous oil.  $[\alpha]_D^{20} = +16.1$  (c 1.00, CHCl<sub>3</sub>); <sup>1</sup>H NMR (CDCl3, 600 MHz) δ 7.43–7.24 (m, 34H), 7.18–7.05 (m, 10H), 5.50 (t, *J* = 9.3 Hz, 1H), 5.19–5.08 (m, 12H), 4.56, 4.43 (ABq, *J* = 12.4 Hz, 2H), 4.55 (dd, *J* = 12.0 Hz, 2.0 Hz, 1H), 4.45 (dd, *J* = 12.0, 5.5 Hz, 1H) 4.41 (s, 2H), 4.11 (dd, *J* = 11.3, 5.2 Hz, 1H), 3.67–3.62 (m, 2H), 3.54 (t, *J* = 9.5 Hz, 1H), 3.36 (t, *J* = 11.0 Hz, 1H); <sup>13</sup>C NMR (CDCl3, 150 MHz) δ 165.84, 165.22, 152.49, 142.66, 137.65, 137.31, 136.99, 136.59, 128.56, 128.36, 128.22, 128.16, 128.05, 127.94, 127.83, 127.52, 127.47, 124.96, 124.88, 109.54, 109.51, 78.19, 77.87, 76.42, 75.26, 75.13, 74.68, 72.63, 71.32, 67.97, 63.91; HRMS (ESI,  $m/z$ ):  $[M+Na]^+$ , calcd for  $[C_{76}H_{68}O_{13}Na]^+$ : 1211.4558; found 1211.4557.

*1,5-Anhydro-2,3-bis-O-benzyl-4,6-bis-O-(3*<sup>0</sup> *,4*0 *,5*0 *-tribenzyloxybenzoyl)-*d-*glucitol* (**26**); Compound **5** (0.34 g, 1.0 mmol), compound **16** (1.3 g, 3.0 mmol), 2-chloro-1-methylpyridinium iodide (0.77 g, 3.0 mmol), DMAP (0.37 g, 3.0 mmol), TEA (0.83 mL, 6.0 mmol) in 20 mL of DCM was stirred at rt for 2 d. By the same procedure previously described for the preparation of compound **17**, compound **26** (1.0 g, 87%) was obtained as a colorless amorphous oil.  $[\alpha]_D^{20}$  = +13.7 (c 1.00, CHCl<sub>3</sub>); <sup>1</sup>H NMR (CDCl<sub>3</sub>, 600 MHz) δ 7.45–7.21 (m, 39H), 7.12–7.03 (m, 5H), 5.32 (t, *J* = 9.5 Hz, 1H), 5.15–5.08 (m, 8H), 5.03–5.01 (m, 4H), 4.79, 4.58 (ABq, *J* = 11.5, 2H), 4.76, 4.66 (ABq, *J* = 11.5 Hz, 2H), 4.63 (dd, *J* = 12.0, 2.8 Hz, 1H), 4.12 (dd, *J* = 12.2, 5.3 Hz, 1H), 4.07 (dd, *J* = 11.5, 5.0 Hz, 1H), 3.70–3.68 (m, 2H), 3.70 (t, *J* = 8.9 Hz, 1H), 3.30 (t, *J* = 10.8 Hz, 1H); <sup>13</sup>C NMR (CDCl3, 150 MHz) δ 165.80, 164.76, 152.48, 152.44, 142.77, 142.42, 137.98, 137.91, 137.47, 137.37, 136.73, 136.56, 128.53, 128.50, 128.47, 128.20, 128.17, 128.07, 128.03, 128.01, 127.94, 127.84, 127.60, 127.53, 124.75, 124.54, 109.40, 109.14, 82.95, 78.22, 76.56, 75.15, 75.08, 73.47, 71.24, 71.06, 70.96, 68.32, 63.44; HRMS (ESI, *m/z*):  $[M+Na]^+$ , calcd for  $[C_{76}H_{68}O_{13}Na]^+$ : 1211.4558; found 1211.4553.

*1,5-Anhydro-2,3,4-tris-O-(3*<sup>0</sup> *,4*0 *,5*0 *-tribenzyloxybenzoyl)-6-O-benzyl-*d-*glucitol* (**27**); Compound **12** (180 mg, 0.70 mmol), compound **16** (1.4 g, 3.2 mmol), 2-chloro-1-methylpyridinium iodide (0.82 g, 3.2 mmol), DMAP (0.39 g, 3.2 mmol), TEA (0.89 mL, 6.4 mmol) in 20 mL of DCM was stirred at rt for 2 d. By the same procedure previously described for the preparation of compound **17**, compound **27** (0.94 g, 91%) was obtained as a colorless amorphous oil.  $\left[\alpha\right]_D^{20} = -4.9$  (c 0.65, CHCl<sub>3</sub>); <sup>1</sup>H NMR (CDCl3, 600 MHz) δ 7.43–7.16 (m, 56H), 5.82 (t, *J* = 9.6 Hz, 1H), 5.55 (t, *J* = 10.0 Hz, 1H), 5.29 (td, *J* = 10.0, 5.5 Hz, 1H), 5.13–4.96 (m, 14H), 4.90 (s, 4H), 4.58, 4.53 (ABq, *J* = 12.0, 2H), (dd, *J* = 10.2, 5.6 Hz, 1H), 3.88–3.84 (m, 1H), 3.67 (dd, *J* = 11.0, 2.4 Hz, 1H), 3.61 (dd, *J* = 10.8, 5.3 Hz, 1H), 3.57 (t, *J* = 10.8 Hz, 1H) <sup>13</sup>C NMR (CDCl3, 150 MHz) δ 165.92, 165.17, 165.02, 152.54, 152.50, 142.88, 142.82, 142.71, 137.44, 137.34, 136.49, 136.45, 136.36, 128.55, 128.51, 128.39, 128.32, 128.27, 128.17, 128.15, 128.10, 128.06, 128.02, 127.95, 127.92, 127.89, 127.81, 127.72, 127.56, 127.52, 124.08, 124.03, 109.20, 109.11, 109.02, 78.36, 75.12, 75.09, 75.07, 74.65, 73.77, 71.18, 71.10, 71.02, 70.75, 69.71, 69.00, 67.26; HRMS (ESI, *m*/*z*): [M + Na] + , calcd for  $[C_{97}H_{84}O_{17}Na]^+$ : 1543.5606; found 1543.5601.

*1,5-Anhydro-2,3,6-tris-O-(3*<sup>0</sup> *,4*0 *,5*0 *-tribenzyloxybenzoyl)-4-O-benzyl-*d-*glucitol* (**28**); Compound **8** (0.38 mg, 1.5 mmol), compound **16** (3.0 g, mmol), 2-chloro-1-methylpyridinium iodide (1.8 g, 7.0 mmol), DMAP  $(0.12 g, 1.0 mmol)$ , TEA  $(1.9 mL, 14 mmol)$  in 30 mL of DCM was stirred at rt for 1 d. By the same procedure

previously described for the preparation of compound **17**, compound **28** (1.0 g, 45% yield) was obtained as a colorless amorphous oil.  $[α]_D^{20} = +47.2$  (c 0.40, CHCl<sub>3</sub>); <sup>1</sup>H NMR (CDCl<sub>3</sub>, 600 MHz) δ 7.47–7.19 (m, 51H), 7.11–7.05 (m, 5H), 5.75 (t, *J* = 9.1 Hz, 1H), 5.19–5.03 (m, 15H), 5.00, 4.92 (ABq, *J* = 11.7 Hz, 4H), 4.60–4.53 (m, 2H), 4.45 (s, 2H), 4.41 (dd, *J* = 11.3, 5.5 Hz, 1H), 3.75–3.70 (m, 2H), 3.46 (t, *J* = 10.8 Hz, 1H); <sup>13</sup>C NMR (CDCl3, 150 MHz) δ 165.84, 165.51, 165.41, 152.59, 152.54, 142.94, 142.75, 142.65, 137.35, 137.32, 137.27, 136.86, 136.62, 136.51, 136.39, 128.60, 128.54, 128.46, 128.44, 128.38, 128.22, 128.18, 128.14, 128.08, 128.06, 128.01, 127.97, 127.90, 127.61, 127.50, 127.43, 124.81, 124.40, 124.05, 109.54, 109.29, 109.03, 78.11, 76.54, 76.04, 75.14, 75.11, 75.09, 74.96, 71.34, 71.20, 71.06, 70.81, 67.07, 63.58; HRMS (ESI, *m*/*z*):  $[M+Na]^+$ , calcd for  $[C_{97}H_{84}O_{17}Na]^+$ : 1543.5606; found 1543.5604.

*1,5-Anhydro-2,4,6-tris-O-(3*<sup>0</sup> *,4*0 *,5*0 *-tribenzyloxybenzoyl)-3-O-benzyl-*d-*glucitol* (**29**); Compound **7** (0.33 mg, 1.3 mmol), compound **16** (2.6 g, 6.0 mmol), 2-chloro-1-methylpyridinium iodide (1.5 g, 6.0 mmol), DMAP  $(0.73 \text{ g}, 6.0 \text{ mmol})$ , TEA  $(1.7 \text{ mL}, 12 \text{ mmol})$  in 20 mL of DCM was stirred at rt for 2 d. By the same procedure previously described for the preparation of compound **17**, compound **29** (1.3 g, 65% yield) was obtained as a colorless amorphous oil.  $[\alpha]_D^{20}$  = +26.7 (c 0.95, CHCl<sub>3</sub>); <sup>1</sup>H NMR (CDCl<sub>3</sub>, 600 MHz) δ 7.46–7.21 (m, 49H), 7.07–6.96 (m, 5H), 5.48 (t, *J* = 9.5 Hz, 1H), 5.34–5.29 (m, 1H), 5.17–5.07 (m, 14H), 5.04 (s, 4H), 4.68 (dd, *J* = 12.0, 3.1 Hz, 1H), 4.62, 4.54 (ABq, *J* = 11.7 Hz, 2H), 4.30 (dd, *J* = 11.2, 5.7 Hz, 1H), 4.22 (dd, *J* = 12.2, 5.0 Hz, 1H), 3.99 (t, *J* = 9.1 Hz, 1H), 3.90–3.87 (m, 1H), 3.42 (t, *J* = 10.8 Hz, 1H); <sup>13</sup>C NMR (CDCl3, 150 MHz) δ 165.81, 164.83, 164.61, 152.63, 152.55, 152.46, 142.94, 142.49, 137.46, 137.41, 137.33, 137.24, 136.67, 136.50, 136.46, 128.57, 128.53, 128.51, 128.46, 128.21, 128.19, 128.14, 128.10, 128.06, 128.02, 127.95, 127.85, 127.80, 127.62, 127.56, 127.47, 124.64, 124.41, 124.38, 109.47, 109.44, 109.13, 80.62, 76.53, 75.16, 75.08, 74.34, 72.02, 71.35, 71.30, 71.05, 70.98, 67.05, 63.33; HRMS (ESI, *m*/*z*): [M + Na] + , calcd for  $[C_{97}H_{84}O_{17}Na]^+$ : 1543.5606; found 1543.5599.

*1,5-Anhydro-2-O-benzyl-3,4,6-tris-O-(3*<sup>0</sup> *,4*0 *,5*0 *-tribenzyloxybenzoyl)-*d-*glucitol* (**30**); Compound **6** (0.38 g, 1.5 mmol), compound **16** (3.0 g, 6.8 mmol), 2-chloro-1-methylpyridinium iodide (1.7 g, 6.8 mmol), DMAP (0.83 g, 6.8 mmol), TEA (2.0 mL, 14 mmol) in 30 mL of DCM was stirred at rt for 2 days. By the same procedure previously described for the preparation of compound **17**, compound **30** (2.0 g, 89% yield) was obtained as a colorless amorphous oil.  $[\alpha]_D^{20}$  = -36.2 (c 1.00, CHCl<sub>3</sub>); <sup>1</sup>H NMR (CDCl<sub>3</sub>, 600 MHz) δ 7.42–7.14 (m, 54H), 5.67 (t, *J* = 9.6 Hz, 1H), 5.41 (t, *J* = 9.8 Hz, 1H), 5.13–4.94 (m, 18H), 4.69 (dd, *J* = 12.2, 2.9 Hz, 1H), 4.59, 4.51 (ABq, *J* = 12.4 Hz, 2H), 4.23 (dd, *J* = 12.4, 5.5 Hz, 1H), 4.19 (dd, *J* = 11.5, 5.3 Hz, 1H) 3.92 (m, 1H), 3.83 (td, *J* = 9.8, 5.3 Hz, 1H), 3.48 (t, *J* = 11.0 Hz, 1H); <sup>13</sup>C NMR (CDCl<sub>3</sub>, 150 MHz) δ 165.68, 165.65, 165.29, 152.51, 152.48, 152.40, 142.87, 142.66, 142.53, 137.54, 137.43, 137.38, 136.63, 136.46, 128.51, 128.46, 128.43, 128.38, 128.29, 128.15, 128.11, 127.98, 127.92, 127.89, 127.87, 127.80, 127.78, 127.57, 127.52, 124.65, 124.53, 123.97, 109.18, 76.69, 76.16, 75.17, 75.08, 72.95, 71.13, 71.03, 70.20, 68.38, 63.54; HRMS (ESI, *m*/z):  $[M + Na]^+$ , calcd for  $[C_{97}H_{84}O_{17}Na]^+$ : 1543.5606; found 1543.5607.

1,5-Anhydro-2-O-(3<sup>,</sup> ,4' ,5'-trihydroxybenzoyl)- $p$ -glucitol (**31**) [\[31\]](#page-25-17); Pd(OH)<sub>2</sub> on C (20 wt.%, 20 mg) was added to a solution of **17** (270 mg, 0.35 mmol) in 10 mL of MeOH and 10 mL of THF under the argon. After the replaced argon atmosphere to hydrogen gas, the suspension was stirred at rt for 6 h. The reaction mixture was filtered and concentrated under reduced pressure to obtain purple amorphous oil. The purple amorphous oil was dissolved by 2 mL acetone and filtered through a whatman™ puradisc 0.1  $\mu$ M TF and concentrated under reduced pressure. In addition, the purple amorphous oil dissolved with 2 mL of MeOH and added acidic resin until becoming a clear solution. After the filtered through the whatman™ puradisc 0.1 µm TF, the solution was concentrated under reduced pressure to give 31 (106 mg, 96%) as pale yellow amorphous oil.  $[\alpha]_{D}^{20} = +58.5$  (c 0.70, MeOH); <sup>1</sup>H NMR (CD3OD, 600 MHz) δ 7.08 (d, *J* = 5.5 Hz, 2H), 4.87–4.83 (overlap, 1H), 4.08 (dd, *J* = 10.5, 5.5 Hz, 1H), 3.86 (dd, *J* = 11.9, 2.2 Hz, 1H), 3.66–3.63 (m, 2H), 3.36 (t, *J* = 9.5 Hz, 1H), 3.29 (t, *J* = 10.7 Hz, 1H), 3.24 (ddd, *J* = 9.7, 5.9, 2.3 Hz, 1H). <sup>13</sup>C NMR (CD3OD, 150 MHz) δ 167.80, 146.42, 139.92, 121.14, 110.25, 82.56, 77.06, 73.28, 71.97, 67.84, 62.93; HRMS (ES<sup>−</sup>, *m*/*z*): [M − H]<sup>−</sup>, calcd for [C<sub>13</sub>H<sub>15</sub>O<sub>9</sub>]<sup>−</sup>: 315.0716; found 315.0718. Please find <sup>1</sup>H NMR, <sup>13</sup>C NMR spectra of compounds **31**–**44**, **54**–**58**, **61** and **67** in the Supplementary Materials.

1,5-Anhydro-3-O-(3',4',5'-trihydroxybenzoyl)-<mark>p-glucitol **(32)** [\[29\]](#page-24-15); Pd(OH)<sub>2</sub> on C (20 wt.%, 20 mg)</mark> was added to a solution of compound **18** (270 mg, 0.35 mmol) in 10 mL of MeOH and 10 mL of THF under the argon. By the same procedure previously described for the preparation of compound **31**, desired compound **32** (110 mg, quant.) was obtained as a colorless amorphous oil.  $[\alpha]_D^{20} = +24.8$ (c 1.0 MeOH); <sup>1</sup>H NMR CD3OD, 600 MHz) δ 7.13 (s, 2H), 5.04 (t, *J* = 9.3 Hz, 1H), 3.97 (dd, *J* = 11.3, 5.5 Hz, 1H), 3.85 (dd, *J* = 11.9, 2.2 Hz, 1H), 3.73–3.69 (m, 1H), 3.66 (dd, *J* = 12.0, 5.8 Hz, 1H), 3.51 (t, *J* = 9.5 Hz, 1H), 3.31–3.27 (overlap, 2H). <sup>13</sup>C NMR (CD<sub>3</sub>OD, 150 MHz) δ 168.50, 146.39, 139.64, 121.88, 110.34, 82.51, 81.20, 70.95, 69.98, 69.89, 62.75; HRMS (ES<sup>−</sup>, *m*/z): [M − H]<sup>−</sup>, calcd for  $[C_{13}H_{15}O_9]$ <sup>−</sup>: 315.0716; found 315.0721.

1,5-Anhydro-4-O-(3',4',5'-trihydroxybenzoyl)-<mark>p-glucitol **(33**) [</mark>29]; Pd(OH)<sub>2</sub> on C (20 wt.%, 20 mg) was added to a solution of compound **19** (280 mg, 0.33 mmol) in 10 mL of MeOH and 10 mL of THF under the argon. By the same procedure previously described for the preparation of compound **31**, desired compound **33** (100 mg, 96%) was obtained as a colorless amorphous oil.  $[\alpha]_D^{20} = -5.7$  (c 0.90, MeOH); <sup>1</sup>H NMR (Acetone − *d*<sub>6</sub>, 600 MHz) δ 7.14 (s, 2H), 4.88 (t, *J* = 9.3 Hz, 1H), 3.94 (dd, *J* = 11.2, 5.3 Hz, 1H), 3.68 (t, *J* = 8.9 Hz, 1H), 3.64–3.60 (m, 1H), 3.57 (dd, *J* = 12.0, 2.1 Hz, 1H), 3.51–3.44 (m, 2H), 3.25 (t, *J* = 10.8 Hz, 1H); <sup>13</sup>C NMR (Acetone − *d*<sub>6</sub>, 150 MHz) δ 166.59, 145.90, 138.86, 121.46, 110.07, 80.56, 77.44, 72.70, 71.38, 70.45, 62.54; HRMS (ES<sup>−</sup>, *m*/*z*): [M − H]<sup>−</sup>, calcd for [C<sub>13</sub>H<sub>15</sub>O<sub>9</sub>]<sup>−</sup>: 315.0716; found 315.0718.

1,5-Anhydro-6-O-(3',4',5'-trihydroxybenzoyl)-<mark>p-glucitol (**34**) [\[31\]](#page-25-17); Pd(OH)<sub>2</sub> on C (20 wt.%, 20 mg)</mark> was added to a solution of compound **20** (210 mg, 0.25 mmol) in 10 mL of MeOH and 10 mL of THF under the argon. By the same procedure previously described for the preparation of compound **31**, desired compound **55** (74 mg, 96%) was obtained as a colorless amorphous oil.  $[\alpha]_D^{20} = +30.3$  (c 1.00, MeOH); <sup>1</sup>H NMR (CD3OD, 600 MHz) δ 7.09 (s, 2H), 4.54 (dd, *J* = 11.9, 1.9 Hz, 1H), 4.35 (dd, *J* = 12.0, 5.5 Hz, 1H), 3.93 (dd, *J* = 11.2, 5.3 Hz, 1H), 3.51–3.55 (m, 1H), 3.48–3.45 (m, 1H), 3.40 (t, *J* = 8.6 Hz, 1H), 3.37 (t, *J* = 8.6 Hz, 1H), 3.23 (t, *J* = 10.8 Hz, 1H). <sup>13</sup>C NMR (CD<sub>3</sub>OD, 150 MHz) δ 168.32, 146.36, 139.71, 121.26, 110.09, 79.91, 79.63, 71.59, 71.20, 70.86, 65.02; HRMS (ES<sup>−</sup> , *m*/*z*): [M − H] − , calcd for  $[C_{13}H_{15}O_9]$  : 315.0716; found 315.0718.

1,5-Anhydro-2,3-bis-O-(3',4',5'-trihydroxybenzoyl)-<mark>p-glucitol **(35**) [</mark>29]; Pd $(OH)_2$  on C  $(20 \text{ wt.}\%$ , 50 mg) was added to a solution of compound **21** (210 mg, 0.25 mmol) in 10 mL of MeOH and 10 mL of THF under the argon. By the same procedure previously described for the preparation of compound **31**, desired compound **35** (313 mg, quant.) was obtained as a colorless amorphous oil.  $[\alpha]_D^{20} = +139.7$ (c 1.00, MeOH); <sup>1</sup>H NMR (CD3OD, 600 MHz) δ 7.05 (s, 2H), 6.96 (s, 2H), 5.41 (t, *J* = 9.5 Hz, 1H), 5.08 (td, *J* = 10.1, 5.3 Hz, 1H), 4.20 (dd, *J* = 11.0, 5.5 Hz, 1H), 3.90 (dd, *J* = 11.9, 2.2 Hz, 1H), 3.74–3.69 (m, 2H), 3.45 (t, *J* = 10.8 Hz, 1H), 3.40 (ddd, *J* = 9.6, 5.5, 2.1 Hz, 1H); <sup>13</sup>C NMR (CD3OD, 150 MHz) δ 168.18, 167.38, 146.38, 146.34, 140.08, 139.85, 121.29, 120.55, 110.30, 110.25, 82.65, 77.91, 71.39, 69.83, 67.81, 62.57; HRMS (ES<sup>-</sup>, *m*/*z*):  $[M - H]$ <sup>-</sup>, calcd for  $[C_{20}H_{19}O_{13}]$ <sup>-</sup>: 467.0826; found 467.0831.

1,5-Anhydro-2,4-bis-O-(3',4',5'-trihydroxybenzoyl)-D-glucitol (36) [\[29\]](#page-24-15); Pd(OH)<sub>2</sub> on C (20 wt.%, 50 mg) was added to a solution of compound **22** (680 mg, 0.57 mmol) in 20 mL of MeOH and 20 mL of THF under the argon. By the same procedure previously described for the preparation of compound **31**, desired compound **36** (267 mg, quant.) was obtained as a colorless amorphous oil.  $[\alpha]_D^{20}$  = +11.3 (c 0.70, MeOH); <sup>1</sup>H NMR (Acetone – *d*<sub>6</sub>, 600 MHz) δ 7.17 (s, 2H), 7.14 (dd, *J* = 9.8, 4.0 Hz, 2H), 5.04 (t, *J* = 9.5 Hz, 1H), 5.01–4.97 (m, 1H), 4.15–4.11 (m, 2H), 3.63–3.54 (m, 3H), 3.41 (t, *J* = 10.8 Hz, 1H); <sup>13</sup>C NMR (Acetone − *d*<sub>6</sub>, 150 MHz) δ 166.39, 166.21, 146.00, 145.98, 138.99, 138.94, 121.43, 121.38, 110.15, 110.09, 80.72, 74.30, 73.19, 72.73, 67.38, 62.49; HRMS (ES<sup>−</sup> , *m*/*z*): [M − H] − , calcd for  $[C_{20}H_{19}O_{13}]^-$ : 467.0826; found 467.0827.

*1,5-Anhydro-2,6-bis-O-(3*<sup>0</sup> *,4*0 *,5*0 *-trihydroxybenzoyl)-*d-*glucitol* (**37**) [\[26\]](#page-24-12); Pd(OH)<sup>2</sup> on C (20 wt.%, 50 mg) was added to a solution of compound **23** (440 mg, 0.37 mmol) in 20 mL of MeOH and 20 mL of THF under the argon. By the same procedure previously described for the preparation of compound **31**, desired compound **37** (170 mg, quant.) was obtained as a yellow amorphous oil.  $[\alpha]_D^{20} = +19.4$ (c 1.00 in MeOH); <sup>1</sup>H NMR (Acetone − *d*<sub>6</sub>, 600 MHz) δ 7.16 (s, 2H), 7.14 (s, 2H), 4.92–4.88 (m, 1H), 4.57 (d, *J* = 10.7 Hz, 1H), 4.41–4.35 (m, 1H), 4.07 (dd, *J* = 10.8, 5.3 Hz, 1H), 3.83–3.78 (m, 1H), 3.61–3.57 (m, 2H), 3.37 (t, *J* = 10.7 Hz, 1H); <sup>13</sup>C NMR (Acetone − *d*<sub>6</sub>, 150 MHz) δ 166.63, 166.30, 146.02, 145.96, 138.89, 138.77, 121.74, 121.46, 110.07, 109.85, 79.47, 76.57, 72.94, 71.60, 67.45, 64.54; HRMS (ES<sup>−</sup> , *m*/*z*): [M – H]<sup>-</sup>, calcd for  $[C_{20}H_{19}O_{13}]$ <sup>-</sup>: 467.0826; found 467.0831.

1,5-Anhydro-3,4-bis-O-(3',4',5'-trihydroxybenzoyl)- $p$ -glucitol (38); Pd(OH)<sub>2</sub> on C (20 wt.%, 50 mg) was added to a solution of compound **24** (710 mg, 0.60 mmol) in 20 mL of MeOH and 20 mL of THF under the argon. By the same procedure previously described for the preparation of compound **31**, desired compound **38** (280 mg, quant.) was obtained as a yellow amorphous oil.  $[\alpha]_D^{20} = -78.2$  (c 1.00, MeOH); <sup>1</sup>H NMR (Acetone − *d*<sub>6</sub>, 600 MHz) δ 7.06 (s, 2H), 7.04 (s, 2H), 5.40 (t, *J* = 9.5 Hz, 1H), 5.16 (t, *J* = 9.6 Hz, 1H), 4.06 (dd, *J* = 11.3, 5.5 Hz, 1H), 3.98–3.93 (m, 1H), 3.71–3.64 (m, 2H), 3.58 (dd, *J* = 12.5, 5.7 Hz, 1H), 3.45 (t, *J* = 10.8 Hz, 1H); <sup>13</sup>C NMR (Acetone − *d*<sub>6</sub>, 150 MHz) δ 166.41, 166.12, 145.83, 145.72, 139.04, 138.66, 121.52, 120.78, 110.05, 110.01, 80.29, 78.04, 70.50, 70.17, 69.50, 62.25; HRMS (ES<sup>−</sup> , *m*/*z*): [M − H]<sup>-</sup>, calcd for [C<sub>20</sub>H<sub>19</sub>O<sub>13</sub>]<sup>-</sup>: 467.0826; found 467.0829.

1,5-Anhydro-3,6-bis-O-(3<sup>7</sup>,4<sup>1</sup>,5'-trihydroxybenzoyl)- $p$ -glucitol (**39**) [\[35\]](#page-25-6); Pd(OH)<sub>2</sub> on C (20 wt.%, 50 mg) was added to a solution of compound **25** (800 mg, 0.67 mmol) in 20 mL of MeOH and 20 mL of THF under the argon. By the same procedure previously described for the preparation of compound **31**, desired compound **39** (310 mg, quant.) was obtained as a yellow amorphous oil.  $[\alpha]_D^{20} = +28.7$  (c 0.60, MeOH); <sup>1</sup>H NMR (Acetone − *d*<sub>6</sub>, 600 MHz) δ 7.17 (s, 2H), 7.16 (s, 2H), 5.11 (t, *J* = 9.1 Hz, 1H), 4.57 (dd, *J* = 11.9, 1.9 Hz, 1H), 4.40 (dd, *J* = 11.9, 5.3 Hz, 1H), 3.99 (dd, *J* = 11.0, 5.5 Hz, 1H), 3.84–3.79 (m, 1H), 3.72 (t, *J* = 9.5 Hz, 1H), 3.65–3.63 (m, 1H), 3.38 (t, *J* = 10.8 Hz, 1H); <sup>13</sup>C NMR (Acetone − *d*<sub>6</sub>, 150 MHz) δ 166.90, 166.63, 145.97, 145.82, 138.71, 138.52, 122.16, 121.76, 110.10, 109.85, 81.10, 79.61, 70.73, 69.66, 69.46, 64.54; HRMS (ES<sup>-</sup>, *m*/*z*): [M – H]<sup>-</sup>, calcd for  $[C_{20}H_{19}O_{13}]$ <sup>-</sup>: 467.0826; found 467.0828.

1,5-Anhydro-4,6-bis-O-(3',4',5'-trihydroxybenzoyl)- $_{p}$ -glucitol (**40**) [\[35\]](#page-25-6); Pd(OH)<sub>2</sub> on C (20 wt.%, 50 mg) was added to a solution of compound **26** (430 mg, 0.36 mmol) in 20 mL of MeOH and 20 mL of THF under the argon. By the same procedure previously described for the preparation of compound **31**, desired compound **40** (160 mg, 95%) was obtained as a yellow amorphous oil.  $[\alpha]_D^{20} = +41.7$  (c 0.60, MeOH); <sup>1</sup>H NMR (Acetone − *d*<sub>6</sub>, 600 MHz) δ 7.15 (s, 2H), 7.14 (s, 2H), 5.09 (t, *J* = 9.5 Hz, 1H), 4.41 (dd, *J* = 12.0, 2.1 Hz, 1H), 4.13 (dd, *J* = 12.0, 5.8 Hz, 1H), 3.97 (dd, *J* = 11.2, 5.3 Hz, 1H), 3.82–3.79 (m, 1H), 3.75 (t, *J* = 9.1 Hz, 1H), 3.71–3.66 (m, 1H), 3.33 (t, *J* = 10.8 Hz, 1H); <sup>13</sup>C NMR (Acetone − *d*<sub>6</sub>, 150 MHz) δ 166.47, 166.13, 145.92, 138.90, 138.77, 121.53, 121.38, 110.12, 109.91, 77.56, 77.42, 72.24, 71.28, 70.51, 69.79, 63.98, 55.32; HRMS (ES<sup>−</sup>, *m*/z): [M − H]<sup>−</sup>, calcd for  $[C_{20}H_{19}O_{13}]$ <sup>−</sup>: 467.0826; found 467.0829.

1,5-Anhydro-2,3,4-tris-O-(3',4',5'-trihydroxybenzoyl)-D-glucitol (41) [\[35\]](#page-25-6); Pd(OH)<sub>2</sub> on C (20 wt.%, 50 mg) was added to a solution of compound **27** (520 mg, 0.34 mmol) in 20 mL of MeOH and 20 mL of THF under the argon. By the same procedure previously described for the preparation of compound **31**, desired compound **41** (210 mg, quant.) was obtained as a yellow amorphous oil.  $[\alpha]_D^{20} = -4.8$  (c 1.00, MeOH); <sup>1</sup>H NMR (Acetone − *d*<sub>6</sub>, 600 MHz) δ 7.07 (d, *J* = 3.8 Hz, 4H), 7.01 (d, *J* = 3.8 Hz, 2H), 5.77 (t, *J* = 9.6 Hz, 1H), 5.36 (t, *J* = 9.8 Hz, 1H), 5.23 (td, *J* = 10.1, 5.4 Hz, 1H), 4.28 (dd, *J* = 11.2, 5.7 Hz, 1H), 3.83–3.80 (m, 1H), 3.72 (dd, *J* = 12.4, 2.1 Hz, 1H), 3.62–3.65 (m, 2H); <sup>13</sup>C NMR (Acetone − *d*<sub>6</sub>, 150 MHz) δ 166.12, 165.91, 165.89, 145.80, 145.65, 139.00, 138.76, 120.93, 120.70, 120.62, 110.03, 109.96, 109.91, 80.37, 74.46, 70.83, 69.96, 67.27, 61.99; HRMS (ES<sup>−</sup>, *m*/*z*): [M − H]<sup>−</sup>, calcd for  $[C_{27}H_{23}O_{17}]$ <sup>−</sup>: 619.0935; found 619.0934.

1,5-Anhydro-2,3,6-tris-O-(3',4',5'-trihydroxybenzoyl)-D-glucitol (**42**) [\[30\]](#page-24-16); Pd(OH)<sub>2</sub> on C (20 wt.%, 50 mg) was added to a solution of compound **28** (410 mg, 0.27 mmol) in 20 mL of MeOH and 20 mL of THF under the argon. By the same procedure previously described for the preparation of compound **31**, desired compound **42** (170 mg, quant.) was obtained as a yellow amorphous oil.  $[\alpha]_D^{20}$  = +80.9 (c 0.70, MeOH); <sup>1</sup>H NMR (Acetone – *d*<sub>6</sub>, 600 MHz) δ 7.19 (s, 2H), 7.10 (s, 2H), 7.04 (s, 2H), 5.50 (t, *J* = 9.5 Hz, 1H), 5.13–5.09 (m, 1H), 4.60 (dd, *J* = 11.9, 1.9 Hz, 1H), 4.47 (dd, *J* = 12.0, 4.8 Hz, 1H), 4.20 (dd, *J* = 11.0, 5.5 Hz, 1H), 3.93 (t, *J* = 9.5 Hz, 1H), 3.79–3.77 (m, 1H), 3.56 (t, *J* = 10.8 Hz, 1H) <sup>13</sup>C NMR (Acetone − *d*<sub>6</sub>, 150 MHz) δ 166.60, 166.39, 166.03, 146.05, 145.96, 145.87, 139.15, 138.86,

138.81, 121.64, 121.56, 120.75, 110.06, 110.00, 109.89, 79.67, 77.07, 70.84, 69.60, 67.44, 64.23; HRMS (ES<sup>−</sup> , *m*/*z*): [M − H]<sup>-</sup>, calcd for [C<sub>27</sub>H<sub>23</sub>O<sub>17</sub>]<sup>-</sup>: 619.0935; found 619.0931.

1,5-Anhydro-2,4,6-tris-O-(3',4',5'-trihydroxybenzoyl)-D-glucitol (43) [\[29\]](#page-24-15); Pd(OH)<sub>2</sub> on C (20 wt.%, 50 mg) was added to a solution of compound **29** (480 mg, 0.32 mmol) in 20 mL of MeOH and 20 mL of THF under the argon. B By the same procedure previously described for the preparation of compound **31**, desired compound **37** (200 mg, quant.) was obtained as a yellow amorphous oil.  $[\alpha]_D^{20}$  = +36.3 (c 1.00, MeOH); <sup>1</sup>H NMR (Acetone – *d*<sub>6</sub>, 600 MHz) δ 7.17 (s, 2H), 7.16 (s, 2H), 7.15 (s, 2H), 5.25 (dd, *J* = 10.0, 9.3 Hz, 1H), 5.07–5.03 (m, 1H), 4.46 (dd, *J* = 12.2, 1.9 Hz, 1H), 4.20–4.16 (m, 3H), 3.95–3.92 (m, 1H), 3.50 (t, *J* = 10.8 Hz, 1H); <sup>13</sup>C NMR (Acetone − *d*<sub>6</sub>, 150 MHz) δ 166.44, 166.19, 165.95, 145.98, 138.98, 138.83, 121.56, 121.35, 121.31, 110.20, 110.11, 109.96, 77.63, 74.27, 72.96, 72.22, 67.45, 63.76; HRMS (ES<sup>-</sup>, *m*/*z*):  $[M - H]$ <sup>-</sup>, calcd for  $[C_{27}H_{23}O_{17}]$ <sup>-</sup>: 619.0935; found 619.0934.

1,5-Anhydro-3,4,6-tris-O-(3',4',5'-trihydroxybenzoyl)-D-glucitol (44) [\[35\]](#page-25-6); Pd(OH)<sub>2</sub> on C (20 wt.%, 50 mg) was added to a solution of compound **30** (765 mg, 0.50 mmol) in 20 mL of MeOH and 20 mL of THF under the argon. By the same procedure previously described for the preparation of compound **31**, desired compound **37** (310 mg, quant.) was obtained as a colorless amorphous oil.  $[\alpha]_D^{20} = -6.12$  (c 0.80, MeOH); <sup>1</sup>H NMR (Acetone − *d*6, 600 MHz) δ 7.20 (s, 2H), 7.07 (s, 2H), 7.06 (s, 2H), 5.46 (t, *J* = 9.5 Hz, 1H), 5.34 (t, *J* = 9.8 Hz, 1H), 4.47 (dd, *J* = 12.4, 2.1 Hz, 1H), 4.27 (dd, *J* = 12.2, 5.3 Hz, 1H), 4.12 (dd, *J* = 11.3, 5.5 Hz, 1H), 4.04–4.00 (m, 2H), 3.55 (t, *J* = 10.8 Hz, 1H) <sup>13</sup>C NMR (Acetone − *d*<sub>6</sub>, 150 MHz) δ 166.39, 166.35, 165.72, 145.83, 145.73, 145.66, 138.98, 138.74, 138.64, 121.44, 121.38, 120.67, 110.05, 109.97, 109.94, 77.86, 77.41, 70.54, 69.73, 69.41, 63.52; HRMS (ES<sup>−</sup>, *m*/z): [M − H]<sup>−</sup>, calcd for  $[C_{27}H_{23}O_{17}]$ <sup>−</sup>: 619.0935; found 619.0941.

*3,4-bis(Benzyloxy)benzoic acid* (**45**) [\[51\]](#page-26-0); Methyl 3,4-dihydroxybenzoate (2.5 g, 15 mmol) and K2CO<sup>3</sup>  $(8.2 g, 60 mmol)$  and KI  $(2.0 g, 12 mmol)$  in 100 mL of acetone was stirred at rt. The reaction suspension was slowly added BnCl (4.2 mL, 36 mmol) and refluxed for 7 h. TLC indicated full conversion of the start material, added MeOH and stirred 1 h. The reaction mixture was filtered by celite and filtrate was evaporated under reduced pressure. The residue was purified by recrystallization with hexane, and the mother liquid was purified by C.C (Hex/EtOAc = 100/1–4/1) to afford methyl ester of **45** (total 5.0 g, 97%) as a white solid. Further on, methyl ester of **45** (3.0 g, 9.0 mmol), KOH (5.0 g, 90 mmol) in 90 mL of 1,4-dioxane and 30 mL of MeOH was stirred at 85 ◦C for 2 h. TLC indicated full conversion of the start material, the reaction mixture was cooled to  $0 °C$  and slowly added 6 M HCl until pH = 1. The precipitating muddy suspension was filtrated, and the white residue was washed by water and MeOH until pH = 7. The white solid was dried *in vacuo*, purified by recrystallizing with MeOH to obtain desired compound **45** (2.4 g, 79%) as colorless needles. m.p. 211 ◦C; <sup>1</sup>H NMR (DMSO-*d*6, 600 MHz) δ 7.56–7.15 (m, 13H), 5.22 (s, 2H), 5.18 (s, 2H); <sup>13</sup>C NMR (DMSO − *d*6, 150 MHz) δ 166.88, 151.94, 147.50, 136.93, 136.62, 128.36, 128.30, 127.82, 127.71, 127.46, 127.34, 123.36, 123.22, 114.43, 112.97, 69.87, 69.73.

*3,5-bis(Benzyloxy)benzoic acid* (**46**) [\[53\]](#page-26-1); Methyl 3,5-dihydroxybenzoate (2.5 g, 15 mmol) and K2CO<sup>3</sup>  $(8.2 g, 60 mmol)$  and KI  $(2.0 g, 12 mmol)$  in 100 mL of acetone was stirred at rt. The reaction suspension was slowly added BnCl (4.2 mL, 36 mmol) and refluxed for 9 h. By the same procedure previously described for the preparation of compound **45**, methyl ester of **46** (5.2 g, quant.) was obtained as a white solid. Further on, methyl ester of **46** (3.0 g, 9.0 mmol), KOH (5.0 g, 90 mmol) in 90 mL of 1,4-dioxane and 30 mL of MeOH was stirred at 85 ◦C for 2 h. By the same procedure previously described for the preparation of compound **45**, desire compound **46** (2.5 g, 81%) was obtained as colorless needles. m.p. 185 ◦C; <sup>1</sup>H NMR (DMSO − *d*6, 600 MHz) δ 7.46–7.30 (m, 11H), 7.19–7.16 (m, 2H), 6.93–6.93 (m, 1H), 5.15 (s, 4H); <sup>13</sup>C NMR (DMSO − *d*6, 150 MHz) δ 166.80, 159.30, 136.64, 132.80, 128.36, 127.79, 127.58, 107.87, 106.43, 69.38.

*3-(Benzyloxy)benzoic acid* (**47**) [\[52\]](#page-26-9); Methyl 3-hydroxybenzoate (2.3 g, 15 mmol) and K2CO<sup>3</sup> (4.2 g, 30 mmol) and KI (1.0 g, 6.0 mmol) in 100 mL of acetone was stirred at rt. The reaction suspension was slowly added BnCl (2.1 mL, 18 mmol) and refluxed for 10 h. By the same procedure previously described for the preparation of compound **45**, methyl ester of **47** (3.7 g, 95%) was obtained as a white

solid. Further on, methyl ester of **47** (1.9 g, 7.4 mmol), KOH (4.2 g, 74 mmol) in 90 mL of 1,4-dioxane and 30 mL of MeOH was stirred at 85 ◦C for 2 h. By the same procedure previously described for the preparation of compound procedure described for previously **45** preparation, desire compound **47** (1.4 g, 85%) was obtained as colorless needles. m.p. 136 ◦C; <sup>1</sup>H NMR (600 MHz, DMSO-*d*6) δ 7.57–7.53 (m, 2H), 7.48–7.39 (m, 5H), 7.35–7.32 (m, 1H), 7.28–7.26 (m, 1H), 5.17 (s, 2H); <sup>13</sup>C-NMR (150 MHz, DMSO-*d*6) δ 167.12, 158.34, 136.84, 132.23, 129.79, 128.50, 127.92, 127.70, 121.82, 119.77, 114.89, 69.35.

4-(*Benzyloxy*)benzoic acid (48) [\[51\]](#page-26-0); Methyl 4-hydroxybenzoate (2.3 g, 15 mmol) and K<sub>2</sub>CO<sub>3</sub> (4.2 g, 30 mmol) and KI (1.0 g, 6.0 mmol) in 100 mL of acetone was stirred at rt. The reaction suspension was slowly added BnCl (2.1 mL, 18 mmol) and refluxed for 9 h. By the same procedure previously described for the preparation of compound **45**, methyl ester of **48** (3.6 g, quant.) was obtained as a white solid. Further on, methyl ester of **48** (2.3 g, 9.0 mmol), KOH (5.0 g, 90 mmol) in 90 mL of 1,4-dioxane and 30 mL of MeOH was stirred at 85  $\degree$ C for 2 h. By the same procedure previously described for the preparation of compound procedure described for previously **45** preparation, desired compound **48** (1.4 g, 85%) was obtained as colorless needles. m.p. 192 ◦C; <sup>1</sup>H NMR (600 MHz, DMSO-*d*6) δ 7.95–7.90 (m, 2H), 7.48–7.46 (m, 2H), 7.42–7.40 (m, 2H), 7.36–7.34 (m, 1H), 7.12–7.09 (m, 2H), 5.19 (s, 2H); <sup>13</sup>C-NMR (150 MHz, DMSO-*d*6) δ 166.90, 161.83, 136.43, 131.27, 128.40, 127.92, 127.73, 123.09, 114.50, 69.35.

*1,5-Anhydro-2,3,4,6-tetrakis-O-(3*<sup>0</sup> *,4*0 *,5*0 *-tribenzyloxybenzoyl)-*d-*glucitol* (**49**) [\[50\]](#page-25-16); 1,5-AG (82 mg, 0.50 mmol), compound **16** (1.1 g, 2.4 mmol), 2-chloro-1-methylpyridinium iodide (0.61 g, 2.4 mmol), DMAP (0.29 g, 2.4 mmol), TEA (0.67 mL, 4.8 mmol) in 20 mL of DCM was stirred at rt for 2 d. By the same procedure previously described for the preparation of compound **17**, desire compound **49** (0.86 g, 92%) was obtained as a colorless amorphous oil.  $\lbrack \alpha \rbrack_{D}^{20} = +7.0$  (c 1.00, CHCl<sub>3</sub>); <sup>1</sup>H NMR (600 MHz, CHLOROFORM-D) δ 7.56–7.07 (m, 68H), 5.89 (t, *J* = 9.8 Hz, 1H), 5.63 (t, *J* = 9.8 Hz, 1H), 5.35–5.31 (m, 1H), 5.18–4.94 (m, 22H), 4.88–4.84 (m, 4H), 4.76 (dd, *J* = 12.2, 2.9 Hz, 1H), 4.52 (dd, *J* = 11.3, 5.5 Hz, 1H), 4.30 (dd, *J* = 12.4, 5.2 Hz, 1H), 4.06–4.03 (m, 1H), 3.60 (t, *J* = 10.8 Hz, 1H); <sup>13</sup>C NMR (CDCl3, 150 MHz) δ 165.91, 165.71, 165.14, 165.10, 152.54, 152.43, 143.06, 142.93, 142.79, 142.62, 137.41, 137.31, 136.59, 136.44, 136.35, 136.25, 128.54, 128.45, 128.44, 128.37, 128.27, 128.23, 128.14, 128.11, 128.08, 128.02, 128.00, 127.95, 127.90, 127.87, 127.81, 127.55, 127.52, 124.56, 123.95, 123.79, 109.22, 109.14, 109.03, 75.10, 75.06, 74.53, 71.18, 71.11, 71.05, 70.51, 69.78, 67.31, 63.36 HRMS (ESI, *m*/*z*): [M + Na] + , calcd for  $[C_{118}H_{100}O_{21}Na]^+$ : 1875.6644; found 1875.6653.

*1,5-Anhydro-2,3,4,6-tetrakis-O-(3*<sup>0</sup> *,4*0 *-dibenzyloxybenzoyl)-*d-*glucitol* (**50**); 1,5-AG (0.12 g, 0.7 mmol), compound **45** (1.4 g, 4.2 mmol), 2-chloro-1-methylpyridinium iodide (1.1 g, 4.2 mmol), DMAP (0.52 g, 4.2 mmol), TEA (1.1 mL, 8.0 mmol) in 20 mL of DCM was stirred at rt for 2 d. By the same procedure previously described for the preparation of compound **17**, desired compound **50** (0.61 g, 72%) was obtained as a colorless amorphous oil.  $[\alpha]_D^{20}$  = +18.8 (c 0.90, CHCl<sub>3</sub>); <sup>1</sup>H NMR (CDCl<sub>3</sub>, 600 MHz) δ 7.65–7.23 (m, 48H), 6.89–6.86 (m, 2H), 6.82–6.80 (m, 1H), 6.77–6.75 (m, 1H), 5.81 (t, *J* = 9.8 Hz, 1H), 5.56 (t, *J* = 9.6 Hz, 1H), 5.29 (td, *J* = 10.1, 5.5 Hz, 1H), 5.22–5.03 (m, 14H), 4.99 (s, 2H), 4.63 (dd, *J* = 12.4, 2.7 Hz, 1H), 4.43 (dd, *J* = 11.3, 5.5 Hz, 1H), 4.29 (dd, *J* = 12.4, 5.2 Hz, 1H), 3.97–3.94 (m, 1H), 3.53 (t, *J* = 10.8 Hz, 1H); <sup>13</sup>C NMR (CDCl3, 150 MHz) δ 165.83, 165.69, 165.17, 164.91, 153.26, 153.14, 152.96, 148.27, 148.17, 136.87, 136.68, 136.51, 136.38, 128.57, 128.53, 128.46, 128.40, 127.94, 127.86, 127.46, 127.41, 127.09, 127.01, 126.98, 124.36, 124.25, 121.79, 115.37, 115.20, 113.01, 76.91, 73.90, 71.07, 71.04, 70.95, 70.70, 70.64, 70.16, 69.38, 67.29, 63.16; HRMS (ESI, *m/*z):  $[M+{\rm Na}]^+$ , calcd for  $[{\rm C}_{62}{\rm H}_{56}{\rm O}_{13}{\rm Na}]^+$ : 1451.4980; found 1451.4977.

*1,5-Anhydro-2,3,4,6-tetrakis-O-(3*<sup>0</sup> *,5*0 *-dibenzyloxybenzoyl)-*d-*glucitol* (**51**); 1,5-AG (0.15 g, 0.9 mmol), compound **46** (1.8 g, 5.4 mmol), 2-chloro-1-methylpyridinium iodide (1.3 g, 5.4 mmol), DMAP (0.66 g, 5.4 mmol), TEA (1.5 mL, 10.8 mmol) in 20 mL of DCM was stirred at rt for 2 d. By the same procedure previously described for the preparation of compound **17**, desired compound **51** (0.69 g, 63%) was obtained as a colorless amorphous oil.  $[\alpha]_D^{20} = +8.3$  (c 0.90, CHCl<sub>3</sub>); <sup>1</sup>H NMR (CDCl<sub>3</sub>, 600 MHz) δ 7.45–7.18 (m, 48H), 6.79–6.76 (m, 2H), 6.72–6.69 (m, 2H), 5.89 (t, *J* = 9.6 Hz, 1H), 5.62 (t, *J* = 9.8 Hz, 1H), 5.37 (td, *J* = 10.1, 5.6 Hz, 1H), 5.06–4.98 (m, 8H), 4.95–4.84 (m, 8H), 4.66 (dd, *J* = 12.2, 2.9 Hz, 1H), 4.49 (dd, *J* = 11.3, 5.5 Hz, 1H), 4.40 (dd, *J* = 12.2, 5.3 Hz, 1H), 4.04–4.01 (m, 1H), 3.58 (t, *J* = 10.8 Hz, 1H); <sup>13</sup>C NMR (CDCl<sub>3</sub>, 150 MHz) δ 165.84, 165.80, 165.20, 165.06, 159.76, 159.75, 159.70, 136.44, 136.31, 136.26, 136.22, 131.45, 130.91, 130.87, 130.71, 128.60, 128.57, 128.51, 128.09, 128.06, 127.59, 127.57, 108.53, 108.49, 108.41, 108.11, 107.74, 107.70, 107.51, 74.28, 70.41, 70.23, 70.19, 69.73, 67.21, 63.51; HRMS (ESI, *m*/*z*):  $[M+Na]^+$ , calcd for  $[C_{62}H_{56}O_{13}Na]^+$ : 1451.4980; found 1451.4980.

*1,5-Anhydro-2,3,4,6-tetrakis-O-(3*<sup>0</sup> *-benzyloxybenzoyl)-*d-*glucitol* (**52**); 1,5-AG (0.10 g, 0.6 mmol), compound **47** (0.82 g, 3.6 mmol), 2-chloro-1-methylpyridinium iodide (0.81 g, 3.6 mmol), DMAP (0.44 g, 3.6 mmol), TEA (1.0 mL, 7.2 mmol) in 20 mL of DCM was stirred at rt for 2 d. By the same procedure previously described for the preparation of compound **17**, desire compound **52** (0.56 g, 92%) was obtained as a colorless amorphous oil.  $[\alpha]_D^{20}$  = +22.7 (c 1.00, CHCl<sub>3</sub>); <sup>1</sup>H NMR (CDCl<sub>3</sub>, 600 MHz) δ 7.68–7.05 (m, 36H), 5.90 (t, *J* = 9.6 Hz, 1H), 5.65 (t, *J* = 9.8 Hz, 1H), 5.40 (td, *J* = 10.1, 5.6 Hz, 1H), 5.02–5.13 (m, 4H), 5.00 (s, 2H), 4.96 (s, 2H), 4.65 (dd, *J* = 12.0, 2.7 Hz, 1H), 4.48 (dd, *J* = 11.2, 5.7 Hz, 1H), 4.42 (dd, *J* = 12.2, 5.3 Hz, 1H), 4.02–4.05 (m, 1H), 3.59 (t, *J* = 10.8 Hz, 1H); <sup>13</sup>C NMR (CDCl3, 150 MHz) δ 166.03, 165.85, 165.34, 165.11, 158.66, 136.57, 136.37, 130.91, 130.30, 130.12, 129.56, 129.49, 128.61, 128.56, 128.13, 127.62, 127.58, 122.52, 121.12, 120.85, 120.71, 120.53, 115.16, 115.12, 115.00, 114.84, 76.89, 74.07, 70.27, 70.12, 69.50, 67.21, 63.31; HRMS (ESI, *m/*z):  $[M+{\rm Na}]^+$ , calcd for  $[{\rm C}_{62}{\rm H}_{56}{\rm O}_{13}{\rm Na}]^+$ : 1027.3306; found 1027.3301.

*1,5-Anhydro-2,3,4,6-tetrakis-O-(4*<sup>0</sup> *-benzyloxybenzoyl)-*d-*glucitol* (**53**); 1,5-AG (0.16 g, 1.0 mmol), compound **48** (1.4 g, 6.1 mmol), 2-chloro-1-methylpyridinium iodide (1.4 g, 6.0 mmol), DMAP (0.73 g, 6.0 mmol), TEA (1.7 mL, 12 mmol) in 30 mL of DCM was stirred at rt for 2 d. By the same procedure previously described for the preparation of compound **17**, desire compound **53** (0.99 g, 98%) was obtained as a colorless amorphous oil.  $[\alpha]_D^{20}$  = +38.2 (c 0.90, CHCl<sub>3</sub>); <sup>1</sup>H NMR (CDCl<sub>3</sub>, 600 MHz) δ 8.06–7.96 (m, 2H), 7.92–7.86 (m, 6H), 7.44–7.17 (m, 20H), 6.93–6.78 (m, 8H), 5.89 (t, *J* = 9.5 Hz, 1H), 5.63 (t, *J* = 9.8 Hz, 1H), 5.40 (td, *J* = 10.0, 5.7 Hz, 1H), 5.02–4.85 (m, 8H), 4.61–4.59 (m, 1H), 4.41 (td, *J* = 12.5, 5.4 Hz, 2H), 3.99–3.96 (m, 1H), 3.54 (t, *J* = 10.8 Hz, 1H); <sup>13</sup>C NMR (CDCl3, 150 MHz) δ 165.83, 165.55, 165.15, 164.86, 162.78, 162.62, 162.57, 136.22, 136.07, 131.93, 131.86, 128.63, 128.61, 128.17, 127.46, 127.41, 127.19, 122.30, 121.65, 121.59, 121.46, 114.52, 114.47, 114.43, 114.38, 73.69, 69.96, 69.90, 69.24, 67.23, 63.05; HRMS (ESI, *m*/z):  $[M + Na]^+$ , calcd for  $[C_{62}H_{56}O_{13}Na]^+$ : 1027.3306; found 1027.3304.

2,3,4,6-tetrakis-O-(3',4',5'-Trihydroxybenzoyl)- $p$ -glucitol (54) [\[50\]](#page-25-16); Pd(OH)<sub>2</sub> on C (20 wt.%, 50 mg) was added to a solution of compound **50** (860 mg, 0.46 mmol) in 20 mL of MeOH and 20 mL of THF under the argon. By the same procedure previously described for the preparation of compound **31**, desired compound **54** (321 mg, 90%) was obtained as a yellow amorphous oil.  $[\alpha]_D^{20} = +58.0$  (c 1.04, MeOH); <sup>1</sup>H NMR (Acetone − *d*<sub>6</sub>, 600 MHz) δ 7.19 (s, 2H), 7.06 (s, 2H) × 2, 6.99 (s, 2H), 5.81 (t, *J* = 9.6 Hz, 1H), 5.50 (t, *J* = 9.8 Hz, 1H), 5.28–5.24 (m, 1H), 4.49 (dd, *J* = 12.4, 2.1 Hz, 1H), 4.33–4.29 (m, 2H), 4.17−4.15 (m, 1H), 3.72 (t, *J* = 10.8 Hz, 1H); <sup>13</sup>C NMR (Acetone − *d*<sub>6</sub>, 150 MHz) δ 166.36, 166.05, 165.90, 165.63, 146.00, 145.97, 145.92, 145.80, 139.22, 138.97, 138.90, 121.50, 120.90, 120.63, 110.17, 110.08, 110.01, 109.98, 77.57, 74.31, 70.74, 69.61, 67.44, 63.32; HRMS-ESI (*m*/z):  $\rm{[M+Na]}^{+}$ , calcd for  $\rm{[C_{34}H_{28}O_{21}Na]}^{+}$ : 795.1021; found 795.1019.

1,5-Anhydro-2,3,4,6-tetrakis-O-(3',4'-dihydroxybenzoyl)-D-glucitol (55); Pd(OH)<sub>2</sub> on C (20 wt.%, 50 mg) was added to a solution of compound **50** (365 mg, 0.30 mmol) in 10 mL of MeOH and 10 mL of THF under the argon. By the same procedure previously described for the preparation of compound **31**, desired compound 55 (196 mg, 92%) was obtained as a colorless amorphous oil.  $[\alpha]_D^{20} = +48.3$ (c 0.65, MeOH); <sup>1</sup>H NMR (Acetone − *d*6, 600 MHz); δ 7.56 (d, *J* = 2.1 Hz, 1H), 7.49 (dd, *J* = 8.4, 1.9 Hz, 1H), 7.43–7.33 (m, 6H), 6.90 (d, *J* = 8.2 Hz, 1H), 6.84 (dd, *J* = 11.3, 8.2 Hz, 2H), 6.76 (d, *J* = 8.2 Hz, 1H), 5.85 (t, *J* = 9.6 Hz, 1H), 5.55 (t, *J* = 9.8 Hz, 1H), 5.30 (td, *J* = 10.1, 5.4 Hz, 1H), 4.52 (dd, *J* = 12.0, 2.4 Hz, 1H), 4.37–4.32 (m, 2H), 4.20–4.17 (m, 1H), 3.75 (t, *J* = 10.8 Hz, 1H); <sup>13</sup>C NMR (Acetone − *d*<sub>6</sub>, 150 MHz) δ 166.22, 166.00, 165.76, 165.56, 151.21, 151.01, 145.55, 145.39, 123.78, 123.71, 123.62, 122.53, 121.96, 121.74, 117.41, 117.29, 117.19, 117.09, 115.74, 115.68, 77.53, 74.40, 70.78, 69.87, 67.48, 63.52; HRMS (ES<sup>−</sup> , *m*/*z*): [M − H]<sup>-</sup>, calcd for [C<sub>34</sub>H<sub>27</sub>O<sub>17</sub>]<sup>-</sup>: 707.1248; found 707.1255.

 $1,5$ -Anhydro-2,3,4,6-tetrakis-O-(3,5-dihydroxybenzoyl)- $p$ -glucitol (**56**);  $Pd(OH)_2$  on  $C$  (20 wt.%, 50 mg) was added to a solution of compound **51** (365 mg, 0.30 mmol) in 20 mL of MeOH and 20 mL of THF

under the argon. By the same procedure previously described for the preparation of compound **31**, desired compound **56** (210 mg, quant.) was obtained as a colorless amorphous oil.  $[\alpha]_D^{20} = +39.1$  (c 0.50, MeOH); <sup>1</sup>H NMR (Acetone − *d*<sub>6</sub>, 600 MHz); δ 7.09 (s, 2H), 6.95 (d, *J* = 1.4 Hz, 4H), 6.89 (d, *J* = 1.7 Hz, 2H), 6.63–6.49 (m, 4H), 5.90 (t, *J* = 9.6 Hz, 1H), 5.59 (t, *J* = 9.6 Hz, 1H), 5.37 (td, *J* = 10.0, 5.6 Hz, 1H), 4.55 (d, *J* = 12.4 Hz, 1H), 4.43 (dd, *J* = 12.2, 4.6 Hz, 1H), 4.37 (dd, *J* = 11.3, 5.5 Hz, 1H), 4.25 (dd, *J* = 9.8, 4.3 Hz, 1H), 3.80 (t, *J* = 10.8 Hz, 1H) <sup>13</sup>C NMR (Acetone − *d*<sub>6</sub>, 150 MHz) δ 166.33, 166.13, 165.82, 165.67, 159.36, 159.31, 159.24, 132.73, 132.10, 131.91, 129.67, 128.96, 108.81, 108.74, 108.65, 108.39, 108.24, 108.08, 77.25, 74.65, 70.85, 69.79, 67.25, 63.45; HRMS (ES<sup>−</sup>, *m*/*z*): [M − H]<sup>−</sup>, calcd for  $[C_{34}H_{27}O_{17}]$ <sup>−</sup>: 707.1248; found 707.1255.

1,5-Anhydro-2,3,4,6-tetrakis-O-(3-hydroxybenzoyl)- $_{\text{D}}$ -glucitol (57); Pd(OH)<sub>2</sub> on C (20 wt.%, 50 mg) was added to a solution of compound **52** (300 mg, 0.30 mmol) in 20 mL of MeOH and 20 mL of THF under the argon. By the same procedure previously described for the preparation of compound **31**, desired compound 57 (181 mg, 94%) was obtained as a colorless amorphous oil.  $[\alpha]_D^{20}$  = +36.4 (c 0.70, MeOH); <sup>1</sup>H NMR (Acetone − *d*<sub>6</sub>, 600 MHz); δ 7.55–6.97 (m, 20H), 5.94 (t, *J* = 9.5 Hz, 1H), 5.65 (t, *J* = 9.6 Hz, 1H), 5.41 (td, *J* = 10.1, 5.4 Hz, 1H), 4.57 (dd, *J* = 12.2, 2.6 Hz, 1H), 4.47 (dd, *J* = 12.4, 4.8 Hz, 1H), 4.39 (dd, *J* = 11.2, 5.7 Hz, 1H), 4.29–4.26 (m, 1H), 3.83 (t, *J* = 10.8 Hz, 1H); <sup>13</sup>C NMR (Acetone − *d*<sub>6</sub>, 150 MHz) δ 166.38, 166.23, 165.90, 165.82, 158.46, 158.25, 132.14, 131.57, 131.43, 130.49, 130.44, 130.40, 129.72, 129.00, 126.08, 121.43, 121.39, 121.21, 121.08, 117.04, 116.93, 116.88, 116.73, 77.23, 74.83, 70.96, 70.23, 67.33, 63.83; HRMS (ES<sup>-</sup>, *m*/*z*): [M – H]<sup>-</sup>, calcd for  $[C_{34}H_{27}O_{13}]$ <sup>-</sup>: 643.1452; found 643.1459.

1,5-Anhydro-2,3,4,6-tetrakis-O-(4-hydroxybenzoyl)- $p$ -glucitol (58); Pd $(OH)_2$  on C (20 wt.%, 50 mg) was added to a solution of compound **54** (288 mg, 0.29 mmol) in 15 mL of MeOH and 15 mL of THF under the argon. By the same procedure previously described for the preparation of compound **31**, desired compound **58** (190 mg, quant.) was obtained as a colorless amorphous oil.  $[\alpha]_D^{20} = +45.3$  (c 0.50, MeOH); <sup>1</sup>H NMR (Acetone − *d*6, 600 MHz); δ 7.93–7.76 (m, 8H), 6.92–6.77 (m, 8H), 5.89 (t, *J* = 9.6 Hz, 1H), 5.61 (t, *J* = 9.8 Hz, 1H), 5.34 (td, *J* = 10.0, 5.3 Hz, 1H), 4.55 (dd, *J* = 12.4, 2.7 Hz, 1H), 4.41–4.35 (m, 2H), 4.23–4.20 (m, 1H), 3.77 (t, *J* = 10.8 Hz, 1H); <sup>13</sup>C NMR (Acetone − *d*<sub>6</sub>, 150 MHz) δ 166.17, 165.99, 165.69, 165.54, 163.17, 163.13, 162.88, 162.82, 132.75, 132.68, 132.61, 132.58, 121.99, 121.44, 121.21, 121.19, 116.09, 116.05, 116.00, 115.94, 77.45, 74.47, 70.76, 70.05, 67.50, 63.65; HRMS (ES<sup>−</sup> , *m*/*z*): [M − H] − , calcd for  $[C_{34}H_{27}O_{13}]^-$ : 643.1452; found 643.1459.

*Benzyl 2,3,4,6-tetrakis-O-(3',4',5'-tribenzyloxybenzoyl)-α,β-D-glucopyranoside* (60); Compound 59 [\[43\]](#page-25-9) (211 mg, 0.78 mmol), EDC·HCl (1.2 g, 6.3 mmol), compound **16** (2.06 g, 4.7 mmol), DMAP (47.7 mg, 0.39 mmol) in 30 mL of DCM was stirred at rt for overnight. After the addition 20 mL of water, the reaction mixture was extracted with DCM (20 mL). The combined organic layer was washed with water (30 mL  $\times$  4) and brine, dried over Na<sub>2</sub>SO<sub>4</sub>, filtered and concentrated under reduced pressure. The crude was purified by C.C (CHCl3/MeOH = 200/1) to obtain desired compound **60** (791 mg, 52%) as a colorless amorphous oil.  $[\alpha]_D^{20}$  = +18.1 (c 1.00, CHCl<sub>3</sub>); <sup>1</sup>H NMR (CDCl<sub>3</sub>, 600 MHz) (assigned for the major anomer; α) δ 7.42–7.16 (m, 80H), 6.21 (t, *J* = 10.0, 1H), 5.69 (t, *J* = 10.0 Hz, 1H), 5.45 (d, *J* = 3.8 Hz, 1H), 5.12–4.95 (m, 16H), 4.73 (dd, *J* = 3.1, 12.4 Hz, 1H), 4.54–4.51 (m, 1H), 4.73 (dd, *J* = 3.1, 12.4 Hz, 1H); <sup>13</sup>C NMR (CDCl3, 150 MHz) (assigned for the major anomer; α) δ 165.68, 165.16, 152.55, 152.52, 152.49, 143.00, 142.86, 142.69, 142.64, 137.41, 137.36, 137.32, 136.56, 136.53, 136.48, 136.40, 136.29, 128.54, 128.46, 128.38, 128.29, 128.24, 128.16, 128.12, 128.09, 128.05, 127.98, 127.94, 127.92, 127.88, 127.81, 127.58, 127.54, 127.52, 124.64, 124.15, 124.02, 123.85, 109.23, 109.16, 109.01, 95.27, 75.11, 75.09, 75.06, 72.17, 71.18, 71.15, 71.05, 70.16, 69.96, 67.96, 63.17. HRMS (ESI, *m/z*):  $\left[ M + Na \right]^{+}$ , calcd for  $\left[ C_{125}H_{106}O_{22}Na \right]^{+}$ : 1981.7073; found 1981.7067.

*2,3,4,6-tetrakis-O-(3*<sup>0</sup> *,4*0 *,5*0 *-Trihydroxybenzoyl)-* <sup>α</sup>*,*β*-*d-*glucopyranose* (**61**) [\[55\]](#page-26-3); Pd(OH)<sup>2</sup> on C (20 wt.%, 20 mg) was added to a solution of compound **54** (100 mg, 0.51 mmol) in 10 mL of MeOH and 10 mL of THF under the argon. By the same procedure previously described for the preparation of compound **31**, desire compound **58** (34 mg, 85%) was obtained as a colorless amorphous oil.  $[\alpha]_D^{20} = +54.4$  (c 0.30, MeOH); <sup>1</sup>H NMR (CDCl3, 600 MHz) δ [α-form] 7.20 (s, 2H), 7.09 (s, 2H), 7.07 (s, 2H), 7.00 (s, 2H), 6.11 (t, *J* = 9.6 Hz, 1H), 5.62 (d, *J* = 3.4 Hz, 1H), 5.56 (t, *J* = 10.2 Hz, 1H), 5.16 (dd, *J* = 10.3, 3.5 Hz, 1H), 4.47

(dd, *J* = 2.4, 11.4 Hz, 1H), 4.35 (dd, *J* = 12.6, 4.8 Hz, 1H), 4.32–4.29 (m, 1H); [β-form] 7.19 (s, 2H), 7.08 (s, 2H), 7.05 (s, 2H), 6.95 (s, 2H), 5.80 (t, *J* = 9.6 Hz, 1H), 5.51 (t, *J* = 9.6 Hz, 1H), 5.29 (t, *J* = 9.0 Hz, 1H), 5.24 (d, *J* = 9.0 Hz, 1H), 4.50 (d, *J* = 1.7, 11.4 Hz, 1H), 4.31 (m, 2H); <sup>13</sup>C NMR (Acetone − *d*<sub>6</sub>, 150 MHz) (assigned for the major anomer; α) δ 166.48, 166.22, 165.99, 165.73, 146.04, 145.99, 145.84, 139.27, 139.24, 139.00, 138.93, 121.61, 121.02, 120.78, 120.67, 110.21, 110.17, 110.06, 110.01, 91.04, 73.02, 70.70, 69.88, 68.55, 63.27; HRMS (ESI,  $m/z$ ):  $[M + Na]^+$ , calcd for  $[C_{62}H_{56}O_{13}Na]^+$ : 811.0970; found 811.0970.

1,5-Anhydro-2,3-bis-O-(3',4',5'-tribenzyloxybenzoyl)-D-glucitol (62); Compound 21 (3.5 g, 3.2 mmol), iodine (0.71 g, 5.6 mmol) in 100 mL of DCM and 50 mL of MeOH was stirred at 70 ◦C for 7 days. The reaction mixture was washed with sodium thiosulfate solution and brine. The crude product was purified by C.C (DCM/MeOH = 4/1) to obtain desired compound **62** (3.0 g, 93%) as a white solid.  $[\alpha]_D^{20}$  = +80.3 (c 1.06, CHCl<sub>3</sub>); <sup>1</sup>H NMR (CDCl<sub>3</sub>, 600 MHz) δ 7.38–7.21 (m, 32H), 5.34 (t, *J* = 9.0 Hz, 1H), 5.28–5.26 (m, 1H), 5.07–4.97 (m, 12H), 4.35–4.32 (m, 1H), 4.00–3.99 (m, 1H), 3.91–3.88 (m, 2H), 3.51–3.48 (m, 2H), 3.07 (d, *J* = 4.0 Hz, 1H), 2.01 (t, *J* = 6.0 Hz, 1H); <sup>13</sup>C NMR (CDCl3, 150 MHz) δ 167.2, 165.3, 152.6, 143.1, 142.9, 137.3, 136.5, 136.4, 128.5, 128.4, 128.2, 128.1, 128.0, 127.7, 127.5, 109.3, 109.1, 80.4, 78.5, 75.1, 71.1, 67.0, 69.8, 66.9, 62.4; HRMS-ESI (*m*/z):  $[M + Na]^{+}$ , calcd for  $[C_{62}H_{56}O_{13}Na]^{+}$ : 1031.3612; found 1031.3619.

*1,5-Anhydro-2,3-bis-O-(3*<sup>0</sup> *,4*0 *,5*0 *-tribenzyloxybenzoyl)-4,6-bis-O-(3*<sup>0</sup> *,5*0 *-dimetoxymetoxy-4*<sup>0</sup> *-benzyloxybenzoyl)* -d-*glucitol* (**64**); Compound **62** (2.5 g, 2.5 mmol), **63** [\[46\]](#page-25-12) (2.5 g, 6.2 mmol), EDC·HCl (1.4 g, 7.2 mmol), DMAP (0.15 g, 1.2 mmol) in 15 mL of DCM was stirred at rt for 18 h. After the addition 30 mL of water, the reaction mixture was extracted with EtOAc  $(3 \times 50 \text{ mL})$ . The combined organic layer was washed with brine, dried over  $Na<sub>2</sub>SO<sub>4</sub>$ , filtered and concentrated under reduced pressure. The crude was purified by C.C (CHCl3/acetone = 100/1) to obtain **64** (2.3 g, 55% yield) as a colorless amorphous oil.  $[\alpha]_D^{20}$  = +24.7 (c 2.15 in CHCl<sub>3</sub>); <sup>1</sup>H NMR (CDCl<sub>3</sub>, 600 MHz)  $\delta$  7.58 (s, 2H), 7.46–7.25 (m, 50H), 5.85 (t, *J* = 10Hz, 1H), 5.61 (t, *J* = 10.0Hz, 1H), 5.28–5.26 (m, 1H), 5.22–4.93 (m, 24H), 4.73–4.71 (m, 1H), 4.50–4.47 (m, 1H), 4.35–4.32 (m, 1H), 4.06–4.03 (m, 1H), 3.56 (t, *J* = 11.0 Hz, 1H), 3.48 (s, 6H), 3.42(s, 6H); <sup>13</sup>C NMR (CDCl<sub>3</sub>, 150 MHz) δ 165.7, 165.5, 165.1, 164.8, 152.5, 150.9, 150.8, 143.7, 143.4, 142.8, 142.7, 137.4, 137.3, 137.2, 136.5, 136.4, 132.4, 130.8, 128.7, 128.5, 128.4, 128.3, 128.2, 128.1, 128.0, 127.9, 127.8, 127.6, 127.5, 125.1, 124.2, 124.1, 124.0, 112.4, 112.3, 109.1, 109.0, 95.4, 75.2, 75.1, 75.0, 74.4, 71.1, 71.0, 70.6, 69.3, 68.1, 67.1, 63.3, 56.4; HRMS-ESI ( $m/z$ ):  $[M + Na]^+$ , calcd for  $[C_{98}H_{92}O_{25}Na]^+$ : 1691.5824; found 1691.5825.

1,5-Anhydro-2,3-bis-O-(3',4',5'-tribenzyloxybenzoyl)-4,6-bis-O-(3',5'-dihydroxy-4'-benzyloxybenzoyl) *-*d-*glucitol* (**65**); Compound **64** (2.0 g, 1.2 mmol) in 8 mL of THF solution was added 8.8 mL of 2-propanol and 0.2 mL of conc. HCl solution, and the mixture was stirred at 60 ◦C for 6 h. After the addition 3 mL of NaHCO<sub>3</sub>, the mixture was extracted with EtOAc ( $3 \times 100$  mL). The combined organic layer was washed with brine, dried over  $Na<sub>2</sub>SO<sub>4</sub>$ , filtered and concentrated under reduced pressure. The crude was purified by C.C (CHCl3/MeOH = 1000/1–500/1) to obtain **65** (1.7 g, 93% yield) as a white amorphous oil.  $[\alpha]_D^{20}$  = +26.8 (c 2.36, CHCl<sub>3</sub>); <sup>1</sup>H NMR (CDCl<sub>3</sub>, 600 MHz) δ 7.43–7.14 (m, 48H), 6.00 (s, 4H), 5.82 (t, *J* = 9.5 Hz, 1H), 5.59 (t, *J* = 9.5 Hz, 1H), 5.37–5.30 (m, 1H), 5.10–4.82 (m, 16H), 4.70–4.68 (m, 1H), 4.54–4.52 (m, 1H), 4.45–4.42 (m, 1H), 3.97–3.95 (m, 1H), 3.57 (t, *J* = 10.5 Hz, 1H); <sup>13</sup>C NMR (CDCl3, 150 MHz) δ 166.2, 165.8, 165.5, 165.2, 152.5, 149.0, 148.9, 142.8, 142.7, 138.2, 137.8, 137.3, 137.2, 136.6, 136.4, 136.2,128.8, 128.7, 128.5, 128.4, 128.3, 128.1, 128.0, 127.9, 127.7, 127.5, 124.5, 123.9, 123.8, 109.9, 109.8, 109.1, 108.9, 77.6, 75.2, 75.1, 75.0, 74.3, 71.1, 71.0, 70.7, 69.7, 68.6, 67.3, 62.6; HRMS-ESI (*m*/*z*):  $[M+Na]^+$ , calcd for  $[C_{90}H_{76}O_{21}Na]^+$ : 1515.4775; found 1515.4777.

*Benzyl protected 1,5-AG-tellimagrandin analog* (66); To a solution of CuCl<sub>2</sub> (135 mg, 1.0 mmol) in 10 mL of MeOH was added *n*-butylamine (400 µL, 4.0 mmol). After the stirred at rt for 1.5 h, the mixture was added to solution of compound **65** (500 mg, 0.34 mmol) in 20 mL of 1,2-dichloroethane (DCE) and stirred at rt for 30 min. The reaction mixture was diluted with 50 mL of diethyl ether, and 50 mL of 5M aq. HCl and 50 mL of diethyl ether were added. The separating organic layer washed with water, NaHCO<sub>3</sub> and brine. The organic layer was dried over  $Na<sub>2</sub>SO<sub>4</sub>$ , filtered and concentrated under reduced pressure. The crude was purified by C.C (DCM/MeOH = 250/1) and HPLC (column,

FNED01048, 250 × 20 mm, SG80–5 µm, eluant DCM/MeOH = 200/1) to afford **66** (237 mg, 48%) as a peal yellow amorphous oil.  $[\alpha]_D^{20}$  = +69.5 (c 1.43, CHCl<sub>3</sub>); <sup>1</sup>H NMR (CDCl<sub>3</sub>, 600 MHz) δ 7.43–7.23 (m, 50H), 6.75 (s, 1H), 6.67 (s, 1H), 5.67 (t, *J* = 10.0 Hz, 1H), 5.37–5.34 (m, 1H), 5.29 (t, *J* = 10.0 Hz, 1H), 5.25–5.24 (m, 1H), 5.13–4.85 (m, 16H), 4.47–4.45 (m, 1H), 3.98 (d, *J* = 12.5 Hz, 1H), 3.96–3.94 (m. 1H), 3.46 (t, *J* = 11.0 Hz, 1H); <sup>13</sup>C NMR (CDCl<sub>3</sub>, 150 MHz) δ 167.1, 166.6, 165.8, 165.2, 152.5, 149.0, 147.0, 142.8, 137.4, 137.3, 136.5, 136.4, 136.3, 136.2, 135.7, 135.5, 130.3, 129.6, 129.0, 128.9, 128.6, 128.5, 128.4, 128.3, 128.1, 128.0, 127.9, 127.8, 127.6, 127.5, 123.9, 113.7, 113.2, 109.2, 108.4, 107.9, 75.6, 75.1, 75.0, 74.4, 71.1, 71.0, 70.8, 70.1, 67.7, 63.6; HRMS-ESI (*m*/z):  $[M + Na]^+$ , calcd for  $[C_{90}H_{74}O_{21}Na]^+$ : 1513.4619; found 1513.4620.

1,5-Anhydro-2,3-bis-O-(3<sup>,</sup> ,4' ,5'-trihydroxybenzoyl)-4,6-O-hexahydroxydiphenyl- $p$ -glucitol (**67**); Pd $(OH)_2$ on C (20 wt.%, 20 mg) was added to a solution of compound **66** (60 mg, 0.40 mmol) in 2 mL of MeOH and 2 mL of THF under the argon. By the same procedure previously described for the preparation of compound procedure described for previously **31** preparation, desired compound **67** (32 mg, quant.) was obtained as a peal yellow amorphous oil.  $[\alpha]_D^{20} = +113.3$  (c 0.29, MeOH); <sup>1</sup>H NMR (CD3OD, 600 MHz) δ 7.00 (s, 2H), 6.95 (s, 2H), 6.60 (s, 1H), 6.40 (s, 1H), 5.61 (t, *J* = 9.5 Hz, 1H), 5.27–5.19 (m, 2H), 5.03 (t, *J* = 10.0 Hz, 1H), 4.25–4.22 (m, 1H), 4.08–4.07 (m, 1H), 3.80 (t, *J* = 13.0 Hz, 1H), 3.54 (t, *J* = 11.0 Hz, 1H); <sup>13</sup>C NMR (CD<sub>3</sub>OD, 150 MHz); 171.1, 170.7, 168.8, 168.3, 150.6, 147.6, 147.1, 144.6, 143.5, 141.8, 139.7, 126.8, 126.7, 126.3, 120.7, 120.3, 120.1, 119.4, 111.1, 110.8, 108.3, 107.7, 78.7, 78.5, 76.2, 72.1, 72.0, 71.8, 71.4, 71.3, 69.4, 68.7, 64.7; HRMS (ESI, *m/z*):  $[M + Na]^+$ , calcd for  $[C_{34}H_{26}O_{21}Na]^+$ : 793.0864; found 793.0865.

**Supplementary Materials:** The following are available online <sup>1</sup>H NMR, <sup>13</sup>C NMR spectra of compounds **31**–**44**, **54**–**58**, **61** and **67**.

**Author Contributions:** Experiment: S.M. (Shota Machida), S.M. (Saki Mukai), R.K. and M.F.; Writing—Original Draft preparation: S.M. (Shota Machida); Writing—Review and Editing: S.M. (Shota Machida), H.S. and T.U.; project administration: T.U.

**Funding:** This study was partly supported by Japan Society for the Promotion of Science (JSPS), 18K06725 (T.U.) and Nihon University Mutidisciplinary Research Grant for 2019 (19-016).

**Acknowledgments:** We thank Koichi Metori (Chemical Analysis Center, School of Pharmacy, Nihon University) for performing mass measurements.

**Conflicts of Interest:** The authors declare no conflict of interest.

## **References**

- <span id="page-23-0"></span>1. Oizumi, T.; Daimon, M.; Jimbu, Y.; Wada, K.; Kameda, W.; Susa, S.; Yamaguchi, H.; Ohnuma, H.; Tominaga, M.; Kato, T. Impaired Glucose Tolerance is a Risk Factor for Stroke in a Japanese Sample-the Funagata Study. *Metabolism* **2008**, *57*, 333–338. [\[CrossRef\]](http://dx.doi.org/10.1016/j.metabol.2007.10.007) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/18249204)
- <span id="page-23-1"></span>2. Bornfeldt, K.E.; Tabas, I. Insulin Resistance, Hyperglycemia, and Atherosclerosis. *Cell Metab.* **2011**, *14*, 575–585. [\[CrossRef\]](http://dx.doi.org/10.1016/j.cmet.2011.07.015) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/22055501)
- <span id="page-23-2"></span>3. Ceriello, A.; Bortolotti, N.; Motz, E.; Pieri, C.; Marra, M.; Tonutti, L.; Lizzio, S.; Feletto, F.; Catone, B.; Taboga, C. Meal-Induced Oxidative Stress and Low-Density Lipoprotein Oxidation in Diabetes: The Possible Role of Hyperglycemia. *Metabolism* **1999**, *48*, 1503–1508. [\[CrossRef\]](http://dx.doi.org/10.1016/S0026-0495(99)90237-8)
- 4. Ten Bruggencate, S.J.; Frederiksen, P.D.; Pedersen, S.M.; Floris-Vollenbroek, E.G.; Lucas-van de Bos, E.; van Hoffen, E.; Wejses, P.L. Dietary Milk-Fat-Globule Membrane Affects Resistance to Diarrheagenic Escherichia Coli in Healthy Adults in a Randomized, Placebo-Controlled, Double-Blind Study. *J. Nutr.* **2016**, *146*, 249–255. [\[CrossRef\]](http://dx.doi.org/10.3945/jn.115.214098) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/26701793)
- 5. Ceriello, A.; Genovese, S. Atherogenicity of Postprandial Hyperglycemia and Lipotoxicity. *Rev. Endocr. Metab. Dis.* **2016**, *17*, 111–116. [\[CrossRef\]](http://dx.doi.org/10.1007/s11154-016-9341-8) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/26880302)
- 6. Mah, E.; Bruno, R.S. Postprandial Hyperglycemia on Vascular Endothelial Function: Mechanisms and Consequences. *Nutr. Res.* **2012**, *32*, 727–740. [\[CrossRef\]](http://dx.doi.org/10.1016/j.nutres.2012.08.002) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/23146769)
- <span id="page-23-3"></span>7. Kitasato, L.; Tojo, T.; Hatakeyama, Y.; Kameda, R.; Hashikata, T.; Yamaoka-Tojo, M. Postprandial Hyperglycemia and Endothelial Function in Type 2 Diabetes: Focus on Mitiglinide. *Cardiovasc. Diabetol.* **2012**, *11*, 79. [\[CrossRef\]](http://dx.doi.org/10.1186/1475-2840-11-79)
- <span id="page-24-0"></span>8. Derosa, G.; Maffioli, P. α-Glucosidase Inhibitors and their Use in Clinical Practice. *Arch. Med. Sci.* **2012**, *8*, 899–906. [\[CrossRef\]](http://dx.doi.org/10.5114/aoms.2012.31621)
- 9. Tuccori, M.; Convertino, I.; Galiulo Maria, T.; Marino, A.; Capogrosso-Sansone, A.; Blandizzi, C. Diabetes Drugs and the Incidence of Solid Cancers: A Survey of the Current Evidence. *Expert Opin. Drug Saf.* **2017**, *16*, 1133–1148. [\[CrossRef\]](http://dx.doi.org/10.1080/14740338.2017.1361401)
- 10. Liu, Z.; Ma, S. Recent Advances in Synthetic α-Glucosidase Inhibitors. *Chem. Med. Chem.* **2017**, *12*, 819–829. [\[CrossRef\]](http://dx.doi.org/10.1002/cmdc.201700216)
- <span id="page-24-1"></span>11. Singla, R.K.; Singh, R.; Dubey, A.K. Important Aspects of Post-Prandial Antidiabetic Drug, Acarbose. *Curr. Top Med. Chem.* **2016**, *16*, 2625–2633. [\[CrossRef\]](http://dx.doi.org/10.2174/1568026616666160414123500) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/27086787)
- <span id="page-24-2"></span>12. Newman, D.J.; Cragg, G.M. Natural Products as Sources of New Drugs from 1981 to 2014. *J. Nat. Prod.* **2016**, *79*, 629–661. [\[CrossRef\]](http://dx.doi.org/10.1021/acs.jnatprod.5b01055) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/26852623)
- 13. Che, C.-T.; Zhang, H. Plant natural products for human health. *Int. J. Mol. Sci.* **2019**, *20*, 830. [\[CrossRef\]](http://dx.doi.org/10.3390/ijms20040830) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/30769917)
- 14. Wright, G.D. Opportunities for Natural Products in 21(st) Century Antibiotic Discovery. *Nat. Prod. Rep.* **2017**, *34*, 694–701. [\[CrossRef\]](http://dx.doi.org/10.1039/C7NP00019G)
- <span id="page-24-3"></span>15. Zhao, C.; Yang, C.F.; Wai, S.T.C.; Zhang, Y.B.; Portillo, M.P.; Paoli, P.; Wu, Y.; Cheang, W.S.; Liu, B.; Carpéné, C.; et al. Regulation of Glucose Metabolism by Bioactive Phytochemicals for the Management of Type 2 Diabetes Mellitus. *Crit. Rev. Food Sci.* **2019**, *59*, 830–847. [\[CrossRef\]](http://dx.doi.org/10.1080/10408398.2018.1501658)
- <span id="page-24-4"></span>16. Bi, W.; Gao, Y.; Shen, J.; He, C.; Liu, H.; Peng, Y.; Zhang, C.; Xiao, P. Traditional Uses, Phytochemistry, and Pharmacology of the Genus Acer (Maple): A review. *J. Ethnopharmacol.* **2016**, *189*, 31–60. [\[CrossRef\]](http://dx.doi.org/10.1016/j.jep.2016.04.021)
- <span id="page-24-5"></span>17. González-Sarrías, A.; Li, L.; Seeram, N.P. Effects of Maple (Acer) Plant Part Extracts on Proliferation, Apoptosis and Cell Cycle Arrest of Human Tumorigenic and Non-Tumorigenic Colon Cells. *Phyther. Res.* **2012**, *26*, 995–1002. [\[CrossRef\]](http://dx.doi.org/10.1002/ptr.3677)
- <span id="page-24-6"></span>18. Bi, W.; Liu, H.; Shen, J.; Zhang, L.H.; Li, P.; Peng, B.; Cao, L.; Zhang, P.; He, C.; Xiao, P. Chemopreventive Effects of Ku-Jin Tea Against AOM-Induced Precancerous Colorectal Lesions in Rats and Metabolomic Analysis. *Sci. Rep.* **2017**, *7*, 15893. [\[CrossRef\]](http://dx.doi.org/10.1038/s41598-017-16237-0)
- <span id="page-24-7"></span>19. Han, S.S.; Lo, S.C.; Choi, Y.W.; Kim, J.H.; Baek, S.H. Antioxidant Activity of Crude Extract and Pure Compounds of Acer Ginnala Max. *Bull. Korean Chem. Soc.* **2004**, *25*, 389–391.
- 20. Royer, M.; Diouf, P.N.; Stevanovic, T. Polyphenol Contents and Radical Scavenging Capacities of Red Maple (Acer Rubrum L.) Extracts. *Food Chem. Toxicol.* **2011**, *49*, 2180–2188. [\[CrossRef\]](http://dx.doi.org/10.1016/j.fct.2011.06.003)
- 21. Watanabe, M.; Prasad, D.H. Antioxidant Phenolic Constituents from the Leaves of Acer Ginnala var Aidzuense. *J. Nat. Remedies* **2017**, *17*, 9–12. [\[CrossRef\]](http://dx.doi.org/10.18311/jnr/2017/15632)
- <span id="page-24-8"></span>22. Bi, W.; Shen, J.; Gao, Y.; He, C.; Peng, Y.; Xiao, P. Ku-Jin Tea (Acer Tataricum Subsp. Ginnala or A. Tataricum Subsp. Theiferum), an Underestimated Functional Beverage Rich in Antioxidant Phenolics. *J. Funct. Foods* **2016**, *24*, 75–84. [\[CrossRef\]](http://dx.doi.org/10.1016/j.jff.2016.04.002)
- <span id="page-24-9"></span>23. Apostolidis, E.; Li, L.; Lee, C.; Seeram, N.P. In Vitro Evaluation of Phenolic-Enriched Maple Syrup Extracts for Inhibition of Carbohydrate Hydrolyzing Enzymes Relevant to Type 2 Diabetes Management. *J. Funct. Foods* **2011**, *3*, 100–106. [\[CrossRef\]](http://dx.doi.org/10.1016/j.jff.2011.03.003)
- <span id="page-24-10"></span>24. Honma, A.; Koyama, T.; Yazawa, K. Antihyperglycemic effects of Japanese maple Acer amoenum leaf extract and its constituent corilagin. *J. Wood Sci.* **2010**, *56*, 507–512. [\[CrossRef\]](http://dx.doi.org/10.1007/s10086-010-1130-5)
- <span id="page-24-11"></span>25. Honma, A.; Koyama, T.; Yazawa, K. Anti-Hyperglycemic Effects of Sugar Maple Acer Saccharum and its Constituent Acertannin. *Food Chem.* **2010**, *123*, 390–394. [\[CrossRef\]](http://dx.doi.org/10.1016/j.foodchem.2010.04.052)
- <span id="page-24-12"></span>26. Bock, K.; LaCour, N.F.; Jensen, S.R.; Nielsen, B.J. The structure of acertannin. *Phytochemistry* **1980**, *19*, 2033. [\[CrossRef\]](http://dx.doi.org/10.1016/0031-9422(80)83034-2)
- <span id="page-24-13"></span>27. Bate-Smith, E.C. Astringent Tannins of Acer Species. *Phytochemistry* **1977**, *16*, 1421–1426. [\[CrossRef\]](http://dx.doi.org/10.1016/S0031-9422(00)88795-6)
- <span id="page-24-14"></span>28. Hatano, T.; Hattori, S.; Ikeda, Y.; Shingu, T.; Okuda, T. Tannins of Aceraceous Plants. Part II. Gallotannins Having a 1,5-Anhydro-D-Glucitol Core and Some Ellagitannins from Acer Species. *Chem. Pharm. Bull.* **1990**, *38*, 1902–1905. [\[CrossRef\]](http://dx.doi.org/10.1248/cpb.38.1902)
- <span id="page-24-15"></span>29. Wan, C.; Yuan, T.; Li, L.; Kandhi, V.; Cech, N.B.; Xie, M.; Seeram, N.P. Maplexins, New α-Glucosidase Inhibitors from Red Maple (Acer Rubrum) Stems. *Bioorganic Med. Chem. Lett.* **2012**, *22*, 597–600. [\[CrossRef\]](http://dx.doi.org/10.1016/j.bmcl.2011.10.073)
- <span id="page-24-16"></span>30. Yuan, T.; Wan, C.; Liu, K.; Seeram, N.P. New Maplexins F-I and Phenolic Glycosides from Red Maple (Acer Rubrum) Bark. *Tetrahedron* **2012**, *68*, 959–964. [\[CrossRef\]](http://dx.doi.org/10.1016/j.tet.2011.11.062)
- <span id="page-25-17"></span>31. Honma, A.; Koyama, T.; Yazawa, K. Anti-Hyperglycaemic Effects of the Japanese Red Maple Acer Pycnanthum and its Constituents the Ginnalins B and C. J. Enzyme Inhib. *Med. Chem.* **2011**, *26*, 176–180.
- <span id="page-25-0"></span>32. Ma, H.; Wang, L.; Niesen, D.B.; Cai, A.; Cho, B.P.; Tan, W.; Gu, Q.; Xu, J.; Seeram, N.P. Structure Activity Related, Mechanistic, and Modeling Studies of Gallotannins Containing a Glucitol-Core and α-Glucosidase. *RSC Adv.* **2015**, *5*, 107904–107915. [\[CrossRef\]](http://dx.doi.org/10.1039/C5RA19014B) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/26989482)
- 33. González-Sarrías, A.; Yuan, T.; Seeram, N.P. Cytotoxicity and Structure Activity Relationship Studies of Maplexins A-I, Gallotannins from Red Maple (Acer Rubrum). *Food Chem. Toxicol.* **2012**, *50*, 1369–1376. [\[CrossRef\]](http://dx.doi.org/10.1016/j.fct.2012.02.031) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/22387705)
- 34. González-Sarrías, A.; Ma, H.; Edmonds, M.E.; Seeram, N.P. Maple Polyphenols, Ginnalins A-C, Induce Sand G2/M-Cell Cycle Arrest in Colon and Breast Cancer Cells Mediated by Decreasing Cyclins A and D1 Levels. *Food Chem.* **2013**, *136*, 636–642. [\[CrossRef\]](http://dx.doi.org/10.1016/j.foodchem.2012.08.023) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/23122108)
- <span id="page-25-6"></span>35. Kamori, A.; Kato, A.; Miyawaki, S.; Koyama, J.; Nash, R.J.; Fleet, G.W.J.; Miura, D.; Ishikawa, F.; Adachi, I. Dual Action of Acertannins as Potential Regulators of Intracellular Ceramide Levels. *Tetrahedron: Asymmetry* **2016**, *27*, 1177–1185. [\[CrossRef\]](http://dx.doi.org/10.1016/j.tetasy.2016.09.006)
- <span id="page-25-1"></span>36. Kato, A.; Koyama, J.; Shinzawa, K.; Imaeda, S.; Adachi, I.; Nash, R.J.; Fleet, G.W.J.; Shintani, M.; Takeuchi, C.; Ishikawa, F. Ginnalin B Induces Differentiation Markers and Modulates the Proliferation/Differentiation Balance via the Upregulation of NOTCH1 in Human Epidermal Keratinocytes. *Bioorganic Med. Chem.* **2019**, *27*, 2172–2180. [\[CrossRef\]](http://dx.doi.org/10.1016/j.bmc.2019.04.008)
- <span id="page-25-2"></span>37. Yokozawa, T.; Chen, C.P.; Dong, E.; Tanaka, T.; Nonaka, G.; Nishioka, I. Study on the Inhibitory Effect of Tannins and Flavonoids Against the 1,1-Diphenyl-2-Picrylhydrazyl Radical. *Biochem. Pharmacol.* **1998**, *56*, 213–222. [\[CrossRef\]](http://dx.doi.org/10.1016/S0006-2952(98)00128-2)
- <span id="page-25-3"></span>38. Lee, D.Y.; Kim, H.W.; Yang, H.; Sung, S.H. Hydrolyzable Tannins from the Fruits of Terminalia Chebula Retz and their α-Glucosidase Inhibitory Activities. *Phytochemistry* **2017**, *137*, 109–116. [\[CrossRef\]](http://dx.doi.org/10.1016/j.phytochem.2017.02.006)
- <span id="page-25-4"></span>39. Wilkins, C.K.; Bohm, B.A. Ellagitannins from Tellima Grandiflora. *Phytochemistry* **1976**, *15*, 211–214. [\[CrossRef\]](http://dx.doi.org/10.1016/S0031-9422(00)89087-1)
- <span id="page-25-5"></span>40. Zheng, S.; Laraia, L.; O'Connor, C.J.; Sorrell, D.; Tan, Y.S.; Xu, Z.; Venkitaraman, A.R.; Wu, W.; Spring, D.R. Synthesis and Biological Profiling of Tellimagrandin I and Analogues Reveals that the Medium Ring Can Significantly Modulate Biological Activity. *Org. Biomol. Chem.* **2012**, *10*, 2590–2593. [\[CrossRef\]](http://dx.doi.org/10.1039/c2ob25065a)
- <span id="page-25-7"></span>41. Uchiyama, T.; Shishikura, K.; Ogawa, K.; Ohshima, Y.; Miyairi, S. An Efficient Method for the Preparation of 1,5-Anhydroalditol from Unprotected Carbohydrates via Glycopyranosyl Iodide. *Tetrahedron Lett.* **2016**, *57*, 5294–5296. [\[CrossRef\]](http://dx.doi.org/10.1016/j.tetlet.2016.10.063)
- <span id="page-25-8"></span>42. Nie, X.; Wang, G. Synthesis of a Ring-Oxygenated Variant of the 2-Carboxy-6-Hydroxyoctahydroindole Core of Aeruginosin 298-A from Glucose. *J. Org. Chem.* **2005**, *70*, 8687–8692. [\[CrossRef\]](http://dx.doi.org/10.1021/jo0507901) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/16238296)
- <span id="page-25-9"></span>43. Lonnecker, A.T.; Lim, Y.H.; Felder, S.E.; Besset, C.J.; Wooley, K.L. Four Different Regioisomeric Polycarbonates Derived from One Natural Product, D-Glucose. *Macromolecules* **2016**, *49*, 7857–7867. [\[CrossRef\]](http://dx.doi.org/10.1021/acs.macromol.6b00591)
- <span id="page-25-10"></span>44. Tanaka, N.; Ogawa, I.; Yoshigase, S.; Nokami, J. Regioselective Ring Opening of Benzylidene Acetal Protecting Group(s) of Hexopyranoside Derivatives by DIBAL-H. *Carbohydr. Res.* **2008**, *343*, 2675–2679. [\[CrossRef\]](http://dx.doi.org/10.1016/j.carres.2008.07.017) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/18718576)
- <span id="page-25-11"></span>45. Daragics, K.; Fügedi, P. Regio- and Chemoselective Reductive Cleavage of 4,6-*O*-Benzylidene-Type Acetals of Hexopyranosides Using BH<sup>3</sup> ·THF-TMSOTf. *Tetrahedron Lett.* **2009**, *50*, 2914–2916. [\[CrossRef\]](http://dx.doi.org/10.1016/j.tetlet.2009.03.194)
- <span id="page-25-12"></span>46. Haslam, E.; Radford, T. Synthesis of 1,3-Anhydro-D-Glucitol and Some Derivatives of 1,5-Anhydro-D-Glucitol. *Carbohydr. Res.* **1966**, *2*, 301–314. [\[CrossRef\]](http://dx.doi.org/10.1016/S0008-6215(00)82565-3)
- <span id="page-25-13"></span>47. Qian, P.; Yao, W.; Huang, L.; Meng, X.; Li, Z. A Mild and Efficient Method for the Selective Cleavage of Primary p-Methoxybenzyl Protecting Group of Saccharides by Co<sub>2</sub>(CO)<sub>8</sub>–Me<sub>2</sub>PhSiH–CO System. *Tetrahedron Lett.* **2015**, *56*, 5238–5241. [\[CrossRef\]](http://dx.doi.org/10.1016/j.tetlet.2015.07.051)
- <span id="page-25-14"></span>48. DeNinno, M.P.; Etienne, J.B.; Duplantier, K.C. A Method for the Selective Reduction of Carbohydrate 4,6-*O*-Benzylidene Acetals. *Tetrahedron Lett.* **1995**, *36*, 669–672. [\[CrossRef\]](http://dx.doi.org/10.1016/0040-4039(94)02348-F)
- <span id="page-25-15"></span>49. Cocinero, E.J.; Gamblin, D.P.; Davis, B.G.; Simons, J.P. The Building Blocks of Cellulose: The Intrinsic Conformational Structures of Cellobiose, its Epimer, Lactose, and their Singly Hydrated Complexes. *J. Am. Chem. Soc.* **2009**, *131*, 11117–11123. [\[CrossRef\]](http://dx.doi.org/10.1021/ja903322w)
- <span id="page-25-16"></span>50. Ren, Y.; Himmeldirk, K.; Chen, X. Synthesis and Structure–Activity Relationship Study of Antidiabetic Penta-O-galloyl-p-glucopyranose and its Analogues. *J. Med. Chem.* 2006, 49, 2829–2837. [\[CrossRef\]](http://dx.doi.org/10.1021/jm060087k)
- <span id="page-26-0"></span>51. Tranchimand, S.; Tron, T.; Gaudin, C.; Iacazio, G. First Chemical Synthesis of Three Natural Depsides Involved in Flavonol Catabolism and Related to Quercetinase Catalysis. *Synth. Commun.* **2006**, *36*, 587–597. [\[CrossRef\]](http://dx.doi.org/10.1080/00397910500406534)
- <span id="page-26-9"></span>52. Fergusson, K.M.; Hird, M. The Dramatic Influence of the Location of Bend and of Lateral Fluoro Substitution on the Mesomorphic Properties of Angular Chiral Esters Based on a 1,3-Disubstituted Benzene Ring. *J. Mater. Chem.* **2010**, *20*, 3069–3078. [\[CrossRef\]](http://dx.doi.org/10.1039/b923267b)
- <span id="page-26-1"></span>53. Hawker, C.J.; Lee, R.; Frechet, J.M.J. One-Step Synthesis of Hyperbranched Dendritic Polyesters. *J. Am. Chem. Soc.* **1991**, *113*, 4583–4588. [\[CrossRef\]](http://dx.doi.org/10.1021/ja00012a030)
- <span id="page-26-2"></span>54. Furukawa, M.; Kamo, S.; Makino, M.; Kurita, M.; Tabata, K.; Matsuzaki, K.; Suzuki, T.; Uchiyama, T. Triterpenoid Glycosides from Ladenbergia Hexandra Klotzsch. *Phytochemistry* **2017**, *136*, 147–155. [\[CrossRef\]](http://dx.doi.org/10.1016/j.phytochem.2017.01.014) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/28153444)
- <span id="page-26-3"></span>55. Gramshaw, J.W.; Haslam, E.; Haworth, R.D.; Searle, T. 571. Gallotannins. Part, V. The Structure of Penta- and Tetra-*O*-Galloylglucoses, and Some Observations on the Molecular Weights of the Gallotannins. *J. Chem. Soc.* **1962**, 2944. [\[CrossRef\]](http://dx.doi.org/10.1039/jr9620002944)
- <span id="page-26-4"></span>56. Feldman, K.S.; Ensel, S.M.; Minard, R.D. Ellagitannin Chemistry. The First Total Chemical Synthesis of an Ellagitannin Natural Product, Tellimagrandin I. *J. Am. Chem. Soc.* **1994**, *116*, 1742–1745. [\[CrossRef\]](http://dx.doi.org/10.1021/ja00084a015)
- <span id="page-26-5"></span>57. Michihata, N.; Kaneko, Y.; Kasai, Y.; Tanigawa, K.; Hirokane, T.; Higasa, S.; Yamada, H. High-Yield Total Synthesis of (-)-Strictinin through Intramolecular Coupling of Gallates. *J. Org. Chem.* **2013**, *78*, 4319–4328. [\[CrossRef\]](http://dx.doi.org/10.1021/jo4003135)
- 58. Yamada, H.; Nagao, K.; Dokei, K.; Kasai, Y.; Michihata, N. Total Synthesis of (-)-Corilagin. *J. Am. Chem. Soc.* **2008**, *130*, 7566–7567. [\[CrossRef\]](http://dx.doi.org/10.1021/ja803111z)
- 59. Kasai, Y.; Michihata, N.; Nishimura, H.; Hirokane, T.; Yamada, H. Total Synthesis of (+)-Davidiin. *Angew. Chemie. Int. Ed.* **2012**, *51*, 8026–8029. [\[CrossRef\]](http://dx.doi.org/10.1002/anie.201203305)
- <span id="page-26-6"></span>60. Asakura, N.; Fujimoto, S.; Michihata, N.; Nishii, K.; Imagawa, H.; Yamada, H. Synthesis of Chiral and Modifiable Hexahydroxydiphenoyl Compounds. *J. Org. Chem.* **2011**, *76*, 9711–9719. [\[CrossRef\]](http://dx.doi.org/10.1021/jo201750d)
- <span id="page-26-7"></span>61. Oki, T.; Masuda, M.; Furuta, S.; Nishiba, Y.; Suda, I. Radical Scavenging Activity of Fried Chips Made from Purple-Fleshed Sweet Potato. *J. Jpn. Soc. Food Sci.* **2011**, *48*, 926–932. [\[CrossRef\]](http://dx.doi.org/10.3136/nskkk.48.926)
- <span id="page-26-8"></span>62. Toda, M.; Kawabata, J.; Kasai, T. α-Glucosidase Inhibitors from Clove (Syzgium aromaticum). *Biosci. Biotechnol. Biochem.* **2000**, *64*, 294–298. [\[CrossRef\]](http://dx.doi.org/10.1271/bbb.64.294) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/10737184)

**Sample Availability:** Samples of the compounds are not available from the authors.



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