

Review

A Review on Daphnane-Type Diterpenoids and Their Bioactive Studies

Yue-Xian Jin ^{1,2,†}, Lei-Ling Shi ^{3,†}, Da-Peng Zhang ⁴, Hong-Yan Wei ³, Yuan Si ¹, Guo-Xu Ma ^{2,3,*} and Jing Zhang ^{1,*}

¹ College of Chinese Medicine Material, Jilin Agricultural University, Changchun 130118, China; jyx0107@126.com (Y.-X.J.); siy2826405514@163.com (Y.S.)

² Institute of Medicinal Plant Development, Chinese Academy of Medical Sciences and Peking Union Medical College, Beijing 100193, China

³ Xinjiang Institute of Chinese and Ethnic Medicine, Urumqi 830002, China; shileiling@sina.com (L.-L.S.); whywlmq@sina.com (H.-Y.W.)

⁴ College of Life Science and Technology, Xinjiang University, Urumqi 830046, China; zdp88888@yeah.net

* Correspondence: mgxfl8785@163.com (G.-X.M.); zhjing0701@163.com (J.Z.)

† These authors contributed equally to this work.

Received: 16 April 2019; Accepted: 7 May 2019; Published: 13 May 2019



Abstract: Natural daphnane diterpenoids, mainly distributed in plants of the *Thymelaeaceae* and *Euphorbiaceae* families, usually include a 5/7/6-tricyclic ring system with poly-hydroxyl groups located at C-3, C-4, C-5, C-9, C-13, C-14, or C-20, while some special types have a characteristic orthoester motif triaxially connected at C-9, C-13, and C-14. The daphnane-type diterpenoids can be classified into five types: 6-epoxy daphnane diterpenoids, resiniferonoids, genkwanines, 1-alkyldaphnanes and redioides, based on the oxygen-containing functions at rings B and C, as well as the substitution pattern of ring A. Up to now, nearly 200 daphnane-type diterpenoids have been isolated and elucidated from the *Thymelaeaceae* and *Euphorbiaceae* families. In-vitro and in-vivo experiments of these compounds have shown that they possess a wide range of biological activities, including anti-HIV, anti-cancer, anti-leukemic, neurotropic, pesticidal and cytotoxic effects. A comprehensive account of the structural diversity is given in this review, along with the cytotoxic activities of daphnane-type diterpenoids, up to April 2019.

Keywords: daphnane; diterpenoid; cytotoxic activities

1. Introduction

Since the first daphnane diterpenoid characterized by a macrolactone motif was isolated from *Trigonostemon reidioides* [1], the daphnane diterpenoids have attracted the interest of many researchers because of their significant bioactive activities. Until now, nearly 200 natural products of daphnane-type diterpenoids have been isolated and identified, and they have shown good biological activities, including anti-HIV, anti-cancer, anti-leukemia, anti-hyperglycemic [2], neurotropic [3], insecticidal and cytotoxic [4] effects. Due to their rich pharmacological activities, especially strong anti-HIV activity and small cytotoxicity, daphnane-type diterpenoids have been employed in a range of clinical applications for a variety of clinical uses [5,6]. Studies have found that the natural daphnane-type diterpenoids usually embrace a 5/7/6-tricyclic ring system with poly-hydroxyl groups located at C-3, C-4, C-5, C-9, C-13, C-14, or C-20, while a special group also have a characteristic orthoester motif connected to C-9, C-13, and C-14. The daphnane-type diterpenoids can be categorized into five types (Figure 1): 6-epoxy daphnane diterpenoids, resiniferonoids, genkwanines, 1-alkyldaphnanes and redioides, based on the substitution pattern of ring A and the oxygen-containing functions at rings B and C. Besides, 6-epoxy

daphnane diterpenoids usually have a C-6 α epoxy structure in ring B; resiniferonoids usually have an α - β unsaturated ketone structure in ring A; genkwanines usually have an α - β saturated ketone structure in ring A, but without a C-6 α epoxy structure in ring B; 1-alkyldaphnanes usually have a saturated ring A, and a large ring between the end of the orthoester alkyl chain and C-1 of ring A; and redioides usually have a 12-carbon macrolide structure between C-3 and C-16, and have a special C-9, C-12, and C-14 orthoester structure. The variety of daphnane-type diterpenoid structures have continued to widen with the discovery of unusual variations with the well-established skeleton. Owing to the unique skeleton and remarkable bioactive activities, daphnane-type diterpenoids have attracted many synthetic endeavors to construct a core structure. However, few papers have reported on the total synthesis of daphnane diterpenoids—isolation from natural plants is still the only source of obtaining daphnane diterpenoids. Considering the extensive interest in daphnane-type diterpenoids, we reviewed the structural and bioactive activities of daphnane-type diterpenoids, with an emphasis on the recent progress in structure identification and bioactive evaluation.

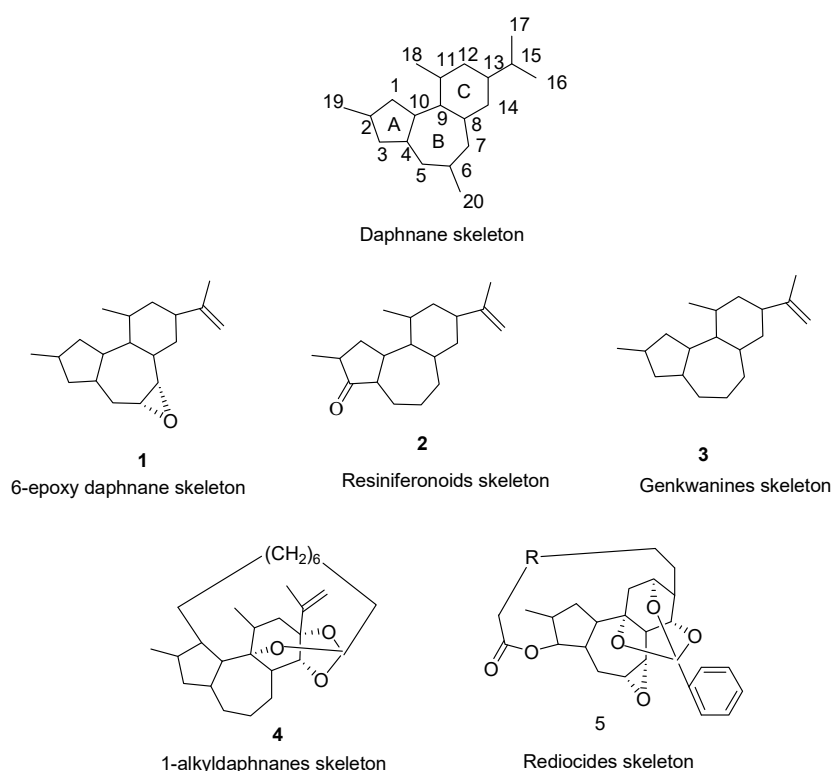


Figure 1. The kinds of daphnane-type diterpenoids skeleton.

2. Occurrence

Natural daphnane-type diterpenoids are mainly distributed in species belonging to the *Thymelaeaceae* or *Euphorbiaceae* families (Table 1). These plants grow mainly in tropical and subtropical regions of Asia [7]. Previous chemical investigations on such species have led to the isolation of a number of structurally diverse diterpenoids [8]. Various daphnane-type diterpenoids have been isolated from some parts of the following plants: The twigs and leaves of *Trigonostemonthyrsoideum*, the roots of *Trigonostemonreidioides*, the stems of *Trigonostemon lii*, the twigs and leaves of *Trigonostemonchinensis* Merr, the stem barks of *Daphne giraldii*, the air-dried roots of *Euphorbia fischeriana*, the stems of *D. acutiloba*, the roots of *Lasiosiphonkraussianus*, the flower buds of *Daphne genkwa*, and the roots of *Maprouneafricana* Muell. Arg., *Trigonostemonxyphophylloides*, *Wikstroemiaaretusa*, *Trigonostemonhowii*, and *Stellerachamaejasme* L., and so on [9].

Table 1. The species of daphnane-type diterpenoids.

Types of Diterpenoids	Species	Medication Site
6-epoxy daphnane diterpenoids	<i>D. acutiloba</i>	Usually their effective part is roots, stems, twigs and leaves, flower buds, fresh bark.
	<i>Trigonostemonthyrsoideum</i>	
	<i>Wikstroemia retusa</i>	
	<i>Daphne genkwa</i>	
	<i>D. oleoides</i> Schreber ssp. <i>oleoides</i>	
	<i>Trigonostemon xyphophylloides</i>	
	<i>Thymelaea hirsuta</i>	
	<i>Neoboutonia glabrescens</i>	
	<i>S. kirkii</i>	
	<i>W. monticola</i>	
	<i>D. tangutica</i>	
	<i>P. elongata</i>	
	<i>T. xyphophylloides</i>	
	<i>T. thyrsoideum</i>	
<i>D. vesiculosum</i>		
<i>Stellerachamaejasme</i> L.		
<i>Trigonostemon chinensis</i> Merr		
Resiniferonoids	<i>Euphorbia fischeriana</i>	Generally, the roots and flower buds are their effective part.
	<i>Daphne genkwa</i>	
	<i>Euphorbia pilosa</i>	
Genkwanines	<i>Trigonostemon xyphophylloides</i>	Usually their effective part is roots, stems, twigs and leaves, flower buds.
	<i>Trigonostemonthyrsoideum</i>	
	<i>Trigonostemon lii</i>	
	<i>Trigonostemon chinensis</i> Merr	
	<i>Daphne genkwa</i>	
<i>Trigonostemon howii</i>		
1-alkyl daphnanes	<i>Wikstroemia chamaedaphne</i>	Usually, the flower buds and fresh bark is their effective part.
	<i>Wikstroemia retusa</i>	
	<i>Stellerachamaejasme</i> L.	
	<i>Daphne genkwa</i>	
	<i>Synaptolepis kirkii</i>	
<i>P. elongata</i>		
Redioides	<i>Trigonostemonthyrsoideum</i>	Generally, their effective part is roots, twigs and leaves.
	<i>Trigonostemon chinensis</i> Merr	
	<i>Trigonostemon reidioides</i>	

3. Species of Daphnane-Type Diterpenoids and Their Bioactive Activities

3.1. 6-Epoxy Daphnane Diterpenoids

6-epoxy daphnane diterpenoids feature a C-6 α epoxy structure in ring B and, occasionally, an α - β unsaturated ketone structure in ring A. In most cases, there is also a C-5 β hydroxyl group and a C-20 hydroxyl group in ring B (Figure 2, Table 2). Compounds acutilobins A–G (1–5, 65, 66), wikstroemia factor M₁ (74), genkwanine VIII (69), gniditrin (14), gnididin (15), gnidicin (13), daphnetoxin (6), yuanhuajine (50), kirkinine (24), excoecaria factor O₁ (8), excoecaria toxin (7), and 14'-ethyltetrahydrouratotoxin (51) have been obtained from the stems of *D. acutiloba*. Acutilobins A–G have been shown to exhibit significant anti-HIV-1 activities, with EC₅₀ below 1.5 μ M [10]. Trigoxypins A (32), B (59), and trigothysoid M (63) have been isolated from the twigs and leaves of *Trigonostemonthyrsoideum*. These compounds have been evaluated for anti-HIV activity by an assay of the inhibition of the cytopathic effects of HIV-1 and cytotoxicity against C8166 cells. However, only trigoxypin A expressed weak anti-HIV-1 activity [11]. Compounds huratotoxin (20) and wikstroelides A–D (37–40), H–J (41–42, 56), and L–N (43, 57–58) have been obtained from the fresh bark of *Wikstroemia retusa*. The orthoester compounds wikstroelides D and H, with palmitic acid at their 20-hydroxyl site, have shown the weakest cytotoxic activity [12]. Antitumor compounds genkwanin I (64) and orthobenzoate 2 (70) have been isolated from the flower buds of *Daphne genkwa*. Genkwanin I has been shown to be a potent cell growth inhibitor constituent [13]. Active ingredients genkwadane

D (9), yuanhuadine (47), yuanhuafine (45), yuanhuacine (49), yuanhuahine (44), yuanhuapine (61), genkwadaphnine (10), isoyuanhuadine (23), and genkwanine M (67) were obtained from the flower buds of *Daphne genkwa*. Among them, yuanhuadine, genkwadaphnine, yuanhuafine, yuanhuapine, and genkwanine M have exhibited the strongest cytotoxic activities against the HT-1080 cell line ($IC_{50} < 0.1 \mu M$) [14]. Maprouneacin (76) has been isolated from the roots of *Maprounea africana* Muell. Arg, and has shown potent glucose-lowering properties when administered via the oral route. [15]. The compound trigonostempene C (71) has been obtained from the twigs and leaves of *Trigonostemon thyrsoideum*, but did not show any significant activity [16]. Compounds yuanhualine (46) and yuanhuagine (48) have been isolated from *Daphne genkwa*. In the analysis of signal transduction molecules, yuanhualine and yuanhuagine appear to suppress the activation of Akt, STAT3 and Src in human lung cancer cells, and also exert potent antiproliferative activity against anticancer-drug resistant cancer cells [17]. Gnidilatidin (17), gnidilatidin-20-palmitate (18), 1, 2 α -dihydrodaphnetoxin (62), genkwadaphnin-20-palmitate (11) and gnidicin-20-palmitate (19) have successfully been obtained from the stems of *D. oleoides* Schreber ssp. *oleoides* [18]. Trigoxiphins J and K (33–34) have been isolated from the stems of *Trigonostemon xyphophylloides*, and subsequently shown to be inactive against three tumor cell lines, specifically the human chronic myelogenous leukemia cell line (K562), the human gastric carcinoma cell line (SGC-7901), and human hepatocellular carcinoma (BEL-7402) (IC_{50} value $> 10 \mu M$) [19]. Genkwanine N (68) has been obtained from the dried flower buds of *Daphne genkwa*, and the compound with esterification of the 20-hydroxyl has shown weak toxicity [20]. Trigonosin B (73) has been isolated from the roots of *Trigonostemon thyrsoideum* [21], while compounds hirseins A and B (21–22) have been isolated from *Thymelaeahirsuta*. Hirseins A and B have shown inhibition of melanogenesis in B16 murine melanoma cells [22]. Glabrescin (12) and Montanin (26) have been obtained from *Neoboutonia glabrescens* [23]. Kirkinine D (25) and synaptolepisfactor K₇ (28) have been isolated from the *S. kirkii* [24]. Wikstrotoxin C (35) has been isolated from *W. monticola*. The compound 2 α -dihydro-20-palimoyldaphnetoxin (52) has been isolated from the *D. tangutica*, while gnidiglaucin (16) has been obtained from *P. longata* [24]. Trigoxiphin C (60) has been obtained from *T. xyphophylloides*, and tested against BEL-7402 cells (human hepatocellular carcinoma), where in it has been shown to be inactive (IC_{50} value $> 10 \mu M$ was defined as inactive) [25]. Trigonosin A (72) has been isolated from *T. thyrsoideum*, and shown to exhibit significant inhibitory activity against specific tumor cells ($IC_{50} > 10 \mu M$) [21]. Isovesiculosin and vesiculosin (54–55) have been isolated from *D. vesiculosum* [26]. Genkwanine O (75) has been obtained from *D. genkwa*. Compound daphnegiraldigin (53) has been isolated from the stem barks of *Daphne giraldii* [27]. Simplexin (27) has been obtained from *Stellerachamaejasme* L. [5]. Compounds trigochinins G–I (29–31) have been isolated from the twigs and leaves of *Trigonostemon chinensis* Merr [28].

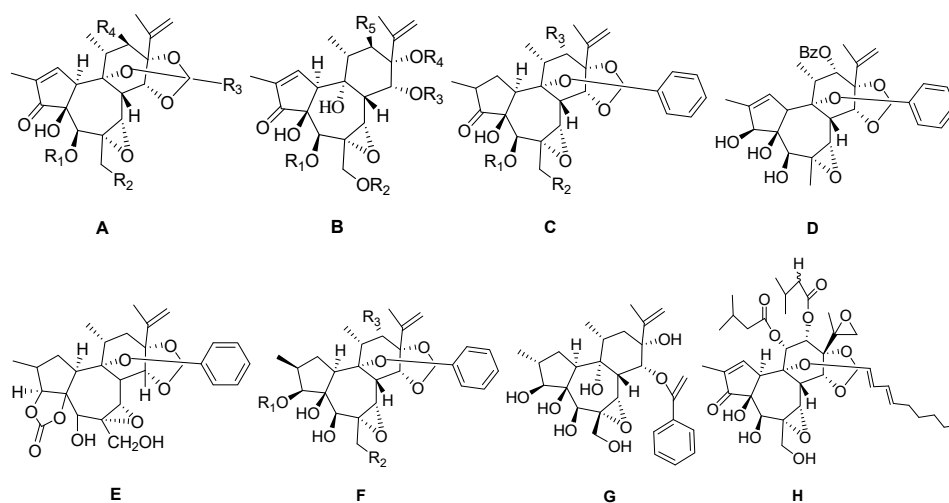


Figure 2. Eight types (A–H) of 6-epoxy daphnane skeletons.

Table 2. Reported structures of 6-epoxy daphnane skeletons.


No.	Name	R ₁	R ₂	R ₃	R ₄	R ₅	Type
1	Acutilobin A	H	OH	Ph	OCO(CH=CH) ₂ COC(CH ₂) ₂ CH ₃	–	A
2	Acutilobin B	H	OH	Ph	OCO(CH=CH) ₃ CHCH ₂ CH ₃ OH	–	A
3	Acutilobin C	H	OH	(CH=CH) ₃ (CH ₂) ₂ CH ₃	OCOCH=CHPhCH ₃ OH	–	A
4	Acutilobin D	H	OH	(CH=CH) ₂ (CH ₂) ₄ CH ₃	OCOCH=CHPhCH ₃ OH	–	A
5	Acutilobin E	H	OH	Ph	OCOCH=CHPhCH ₃ OH	–	A
6	Daphnetoxin	H	OH	Ph	H	–	A
7	Excoecaria toxin	H	OH	(CH=CH) ₂ (CH ₂) ₄ CH ₃	H	–	A
8	Excoecaria factor O ₁	H	OH	(CH=CH) ₃ (CH ₂) ₂ CH ₃	H	–	A
9	Genkwadane D	H	OH	(CH=CH) ₂ (CH ₂) ₄ CH ₃	OCOCH(CH ₃) ₂	–	A
10	Genkwadaphnine	H	OH	Ph	OBz	–	A
11	Genkwadaphnin-20-palmitate	H	OCO(CH ₂) ₁₄ CH ₃	Ph	OCOPh	–	A
12	Glabrescin	H	OCOCH ₂ (CH ₂) ₁₃ CH ₃	(CH ₂) ₁₀ CH ₃	H	–	A
13	Gnidicin	H	OH	Ph	OCOCH=CHPh	–	A
14	Gniditrin	H	OH	Ph	OCO(CH=CH) ₃ (CH ₂) ₂ CH ₃	–	A
15	Gnididin	H	OH	Ph	OCO(CH=CH) ₂ (CH ₂) ₄ CH ₃	–	A
16	Gnidiglaucin	H	OH	(CH ₂) ₈ CH ₃	OAc	–	A
17	Gnidilatidin	H	OH	(CH=CH) ₂ (CH ₂) ₄ CH ₃	OCOPh	–	A
18	Gnidilatidin-20-palmitate	H	OCO(CH ₂) ₁₄ CH ₃	(CH=CH) ₂ (CH ₂) ₄ CH ₃	OCOPh	–	A
19	Gnidicin-20-palmitate	H	OCO(CH ₂) ₁₄ CH ₃	Ph	OCOCH=CHPh	–	A
20	Huratoxin	H	OH	(CH=CH) ₂ (CH ₂) ₈ CH ₃	H	–	A
21	Hirsein A	H	OH	CH=CH(CH ₂) ₄ CH ₃	OCOCH=CHPh	–	A
22	Hirsein B	H	OH	CH=CH(CH ₂) ₄ CH ₃	OCOCH=CHPhOH	–	A
23	Isoyuanhuadine	H	OH	(CH=CH) ₂ (CH ₂) ₄ CH ₃	OAc	–	A
24	Kirkinine	H	OH	CH=CH(CH ₂) ₁₂ CH ₃	OAc	–	A
25	Kirkinine D	H	OH	(CH=CH) ₃ (CH ₂) ₂ CH ₃	OAc	–	A
26	Montanin	H	OH	(CH ₂) ₁₀ CH ₃	H	–	A
27	Simplexin	H	OH	(CH ₂) ₈ CH ₃	H	–	A
28	Synaptolepisfactor K ₇	H	OH	CH=CH(CH ₂) ₁₂ CH ₃	H	–	A
29	Trigochinin G	H	H	Ph	OCOCH ₂ CH(CH ₃) ₂	–	A
30	Trigochinin H	H	H	Ph	OCOC ₆ H ₄ (4-OH)	–	A
31	Trigochinin I	H	H	Ph	OCOC ₆ H ₃ (3-OMe)(4-OH)	–	A
32	Trigoxyphin A	H	H	Ph	OBz	–	A
33	Trigoxyphin J	H	OH	CH ₃	OCO(CH ₂) ₁₄ CH ₃	–	A
34	Trigoxyphin K	H	H	Ph	OBz	–	A
35	Wikstrotoxin C		OH	(CH=CH) ₂ (CH ₂) ₄ CH ₃	OAc	–	A
36	Wikstrotoxin D	H	OH	n-C ₉ H ₁₉	H	–	A

Table 2. Cont.

No.	Name	R ₁	R ₂	R ₃	R ₄	R ₅	Type
37	Wikstroelide A	H	OH	(CH=CH) ₂ (CH ₂) ₈ CH ₃	OAc	–	A
38	Wikstroelide B	H	OH	(CH=CH) ₂ (CH ₂) ₉ CH ₃	OAc	–	A
39	Wikstroelide C	H	O-trans-5-pentadecenoic acid	(CH=CH) ₂ (CH ₂) ₈ CH ₃	OAc	–	A
40	Wikstroelide D	H	O-palmitic acid	(CH=CH) ₂ (CH ₂) ₈ CH ₃	OAc	–	A
41	Wikstroelide H	H	OH	(CH=CH) ₂ (CH ₂) ₆ CH ₃	OAc	–	A
42	Wikstroelide I	H	O-palmitic acid	(CH=CH) ₂ (CH ₂) ₉ CH ₃	OAc	–	A
43	Wikstroelide L	H	OH	(CH=CH) ₂ (CH ₂) ₈ CH ₃	OAc	–	A
44	Yuanhuahine	H	OH	(CH=CH) ₂ (CH ₂) ₄ CH ₃	OCOCH ₂ CH ₃	–	A
45	Yuanhuafine	H	H	Ph	OAc	–	A
46	Yuanhualine	H	OH	(CH=CH) ₂ (CH ₂) ₄ CH ₃	OCO(CH ₂) ₂ CH ₃	–	A
47	Yuanhuadine	H	OH	(CH=CH) ₂ (CH ₂) ₄ CH ₃	OAc	–	A
48	Yuanhuagine	H	OH	(CH=CH)(CH ₂) ₂ CH ₃	OCOCH ₃	–	A
49	Yuanhuacine	H	OH	(CH=CH) ₂ (CH ₂) ₄ CH ₃	OBz	–	A
50	Yuanhuajine	H	OH	(CH=CH) ₃ (CH ₂) ₂ CH ₃	OBz	–	A
51	14'-ethyltetrahydrohuratoxin	H	OH	(CH ₂) ₁₄ CH ₃	H	–	A
52	2α-dihydro-20-palimoyldaphnetoxin	H	OH	CH=CH(CH ₂) ₆ CH ₃	OAc	–	A
53	Daphnegiraldigin	H	OH	COPh	H	H	B
54	Isovesiculosin	Ac	Ac	Ac	CO(CH=CH) ₂ (CH ₂) ₄ CH ₃	H	B
55	Vesiculosin	H	H	CO(CH=CH) ₂ (CH ₂) ₄ CH ₃	H	H	B
56	Wikstroelide J	H	H	CO(CH=CH) ₂ (CH ₂) ₈ CH ₃	H	OAc	B
57	Wikstroelide M	H	H	CO(CH=CH) ₂ (CH ₂) ₈ CH ₃	H	H	B
58	Wikstroelide N	H	H	CO(CH=CH) ₂ (CH ₂) ₉ CH ₃	H	H	B
59	Trigoxypin B	H	H	OBz	–	–	C
60	Trigoxypin C	Ac	H	OBz	–	–	C
61	Yuanhuapine	H	OH	OAc	–	–	C
62	1,2α-dihydrodaphnetoxin	H	OH	H	–	–	C
63	Trigothyoid M	–	–	–	–	–	D
64	Genkwanin I	–	–	–	–	–	E
65	Acutilobin F	CO(CH=CH) ₃ (CH ₂) ₂ CH ₃	OH	H	–	–	F
66	Acutilobin G	COCH=CHPh	OH	H	–	–	F
67	Genkwanine M	H	OBz	H	–	–	F
68	Genkwanine N	Bz	OH	H	–	–	F
69	Genkwanine VIII	COPh	OH	H	–	–	F
70	Orthobenzoate 2	H	OH	H	–	–	F
71	Trigonostempene C	H	H	OH	–	–	F
72	Trigonosin A	H	H	OBz	–	–	F
73	Trigonosin B	H	OH	OBz	–	–	F
74	Wikstroemia factor M ₁	CO(CH=CH) ₂ (CH ₂) ₄ CH ₃	OH	H	–	–	F
75	Genkuanine O	–	–	–	–	–	G
76	Maprouneacin	–	–	–	–	–	H

3.2. Resiniferonoids

Relative to 6-epoxy daphnane diterpenoids, there is no C-6 α epoxy structure in ring B for resiniferonoids. However, resiniferonoids do possess an α - β unsaturated ketone structure in ring A (Figure 3, Table 3). Compounds 4 β , 9 α , 20- trihydroxy- 13, 15- secotiglia- 1,6- diene- 3,13- dione 20-O- β -D- [6-galloyl] glu-copyranoside (**86**) and euphopiloside A (**84**) have been isolated from the air-dried roots of *Euphorbia fischeriana*, and display moderate inhibitory effects against α -glucosidase in in-vitro bioassays [29]. Yuanhuatine (**78**) has been isolated from the flower buds of *Daphne genkwa* [14]. Compounds daphneresiniferins A and B (**80–81**) have been obtained from the flower buds of *Daphne genkwa*. A study found that daphneresiniferin A was able to dependently inhibit melanin production [30]. Genkwanine L (**77**) has been isolated from the bud of *Daphne genkwa* [31]. Euphopiloside B (**83**), langduin A (**85**) and phorbol (**87**) have been obtained from the *Euphorbia Pilosa* [32], while compounds genkwadane A (**79**) and yuanhuaate B (**82**) have been isolated from the flower buds of *Daphne genkwa* [14].

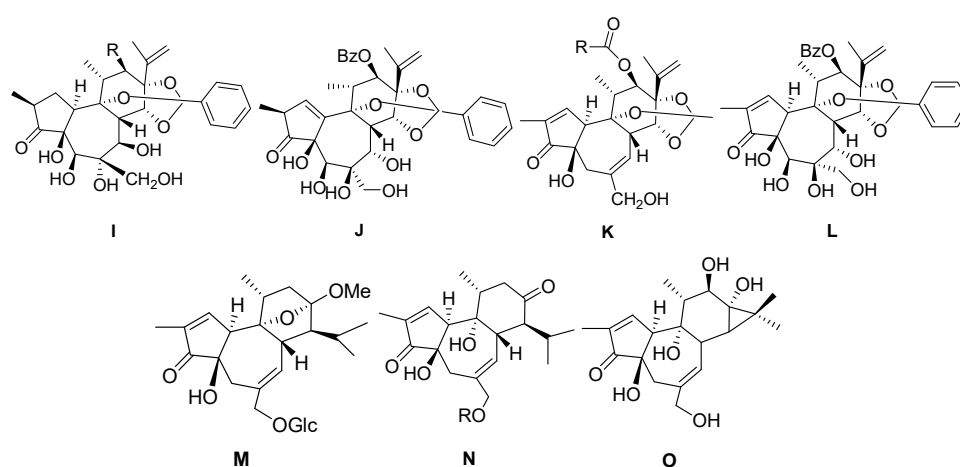
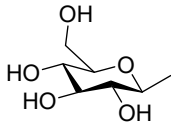
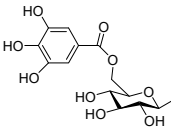


Figure 3. Seven types (I–O) of resiniferonoids skeletons.

Table 3. Reported structures of resiniferonoids skeletons.

No.	Name	R	Type
77	Genkwanine L	OAc	I
78	Yuanhuatine	OBz	I
79	Genkwadane A	–	J
80	Daphneresiniferin A	Me	K
81	Daphneresiniferin B	Ph	K
82	Yuanhuaate B	–	L
83	Euphopiloside B	–	M
84	Euphopiloside A		N
85	Langduin A	H	N
86	4 β ,9 α ,20-trihydroxy-13,15-secotiglia-1,6-diene-3,13-dione 20-O- β -D-[6-galloyl]glu-copyranoside		N
87	Phorbol	–	O

3.3. Genkwamines

Relative to 6-epoxy daphnane diterpenoids and resiniferonoids, genkwamines have an α - β saturated ketone structure in ring A, but do not possess a C-6 α epoxy structure in ring B (Figure 4, Table 4). Compound trigoxyphin H (**100**) has been isolated from the twigs of *Trigonostemonxyphophylloides* [33]. The active ingredients trigothysoids A–L (**122–124**, **96–99**, **139–141**, **131,128**), trigochinins A–E (**145–146**, **130**,

147–148), andtrigonothyryns D, E (143–144) and G (121) have been obtained from the twigs and leaves of *Trigonostemonthyrsoideum*. These compounds have been evaluated for their anti-HIV activity using an assay to determine their inhibition of the cytopathic effects of HIV-1 and their cytotoxicity against C8166 cells. Amongst them, trigothysoid A and L exhibited moderate anti-HIV-1 activity; andtrigothysoid C and K andtrigochinins A, B and D expressed weak anti-HIV-1 activity [11]. Trigolins A–G (132–138) and trigonothyryrin F (107) have been isolated from the stems of *Trigonostemon lii*. Trigolins A, G, H, and K have been shown to exhibit modest anti-HIV-1 activity with EC₅₀ values of 2.04, 9.17, 11.42, and 9.051 µg/mL, respectively [34]. Compound trigochinin F (149) has been obtained from the twigs and leaves of *Trigonostemon chinensis* Merr, and has shown strong inhibition of HL-60 tumor cell lines [28]. Trigonothyryns A–C (125–127) have been isolated from the stems of *Trigonostemonthyrsoideum* [6]. Among them, trigonothyryrin C has shown significant activity to prevent the cytopathic effects of HIV-1 in C8166 cells, with an EC₅₀ value of 2.19 µg/mL [35]. Compounds genkwaniens F, I, and J (93, 113, 114) have been isolated from the flower buds of *Daphne genkwa* [14]. Genkwanine H (95) has been obtained from the flower buds of *Daphne genkwa*, and the compound has been shown to dependently inhibit melanin production [30]. Compounds trigonostempenes A (150) and B (129) have been isolated from the twigs and leaves of *Trigonostemonthyrsoideum*. Studies have shown that the discovery of these NO inhibitory daphnane diterpenoids—including compound trigonostempene A—which possess IC₅₀ values comparable to positive controls may have the potential to be developed as anti-neuroinflammatory agents for Alzheimer disease (AD) and other related neurological disorders [16]. Most inhibitors of acetylcholinesterase (AChE) are alkaloids that often possess several side effects, whereas these daphnane-type diterpenoids do not belong to the class of alkaloids, and therefore they may constitute novel active AChE inhibitors with fewer side effects. It is important to search for new AChE inhibitors not belonging to this structural class [36,37]. Genkwaniens A–E (88–92), G (94), I (113), and K (115) have been obtained from the bud of *Daphne genkwa*. Among these compounds, genkwanine D has been shown to exhibit strong activity to inhibit the endothelium cell HMEC at IC₅₀ levels of 2.90–15.0 µM [31]. Compounds trigoxyphins U and W (105–116) have been isolated from the twigs of *Trigonostemon xyphophylloides*. Trigoxyphin W has shown modest cytotoxicity against BEL-7402, SPCA-1 and SGC-7901, with IC₅₀ values of 5.62, 16.79 and 17.19 µM, respectively [33]. Trigonosins C–D (106, 142) have been obtained from the roots of *Trigonostemonthyrsoideum* [21]. Trigoxyphin I (104) has been isolated from the *Trigonostemon xyphophylloides* [38]. Compounds trigohownins D and E (101–102), and trigohownins A–C (108–110) and F–I (117–120) have been obtained from the *Trigonostemon howii*. Among them, trigohownins A and D have been shown to exhibit moderate cytotoxic activity against the HL-60 tumor cell line, with IC₅₀ values of 17.0 and 9.3 µM, respectively [39]. Trigoxyphins D–F (111–112, 103) have been isolated from *Trigonostemon xyphophylloides*, with all three compounds found to be inactive against BEL-7402 cells (IC₅₀ value > 10 µM) [25].

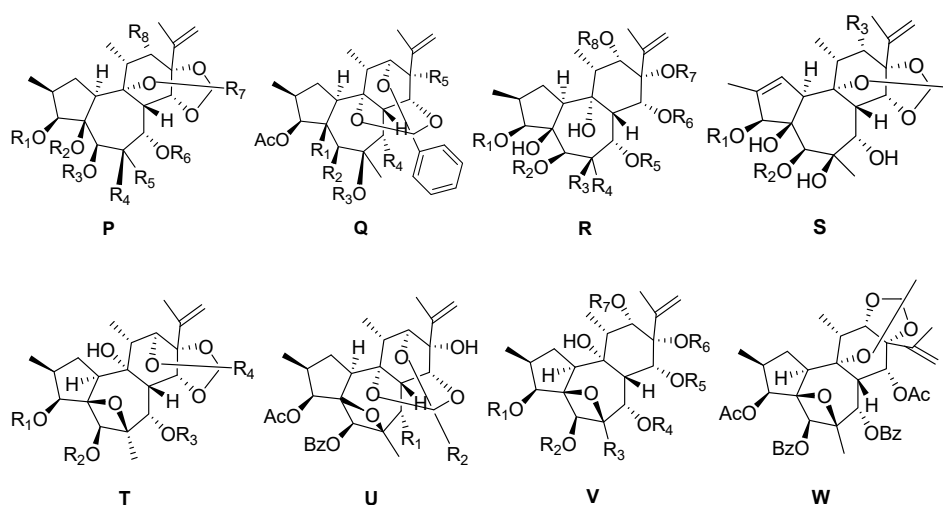


Figure 4. Eight types (P–W) of genkwaniens skeletons.

3.4. 1-Alkyldaphnanes

1-alkyldaphnanes have a large ring between the end of the orthoester alkyl chain and C-1 of ring A (Figure 5, Table 5). Pimelea factors S_6 (**168**) and S_7 (**169**) have been isolated from the flower buds of *Wikstroemiachamaedaphne* and have shown moderate cytotoxic activities against human myeloid leukemia HL-60, hepatocellular carcinoma SMMC-7721, lung cancer A549, breast cancer MCF-7, and colon cancer SW480 [1]. Compound pimelea factor P_2 (**155**) has been obtained from the fresh bark of *Wikstroemiaretusa*, and has been shown to exhibit cytotoxicity in 10 cell lines (including HeLa, HepG2, HT-1080, HCT116, A375-S2, MCF-7, A549, U-937, K562 and HL60 cell lines) [14]. Wikstroelides E–G, K and O (**163–167**) have been isolated from the fresh bark of *Wikstroemiaretusa*. Among them, compound wikstroelide E has been shown to exhibit the highest activity against cell lines PC-6 (human lung cancer cell line) and P388 (mouse leukaemia cell line), followed by wikstroelides A and J, which have the orthoester group without a fatty acid at the 20-hydroxyl [12]. Compounds stellaralides A–C (**151–152**, **174**) and gnidimacrin (**153**) have been isolated from the *Stellerachamaejasme* L. [5]. Genkwadane B (**154**), pimelotides A and C (**170**, **172**), and genkwadane C (**156**) have been isolated from the flower buds of *Daphne genkwa* [14]. Compounds wikstroelides R–T (**157–159**) have been obtained from the flower buds of *Wikstroemiachamaedaphne*. Wikstroelide R has been shown to have moderate cytotoxic activities against human cancer cell lines [1]. Compounds kirkinines B, C, and E (**160–162**) were isolated from *Synaptolepiskirkii*. Pimelotides B and D (**171**, **173**) have been obtained from *Pelongata* [40].

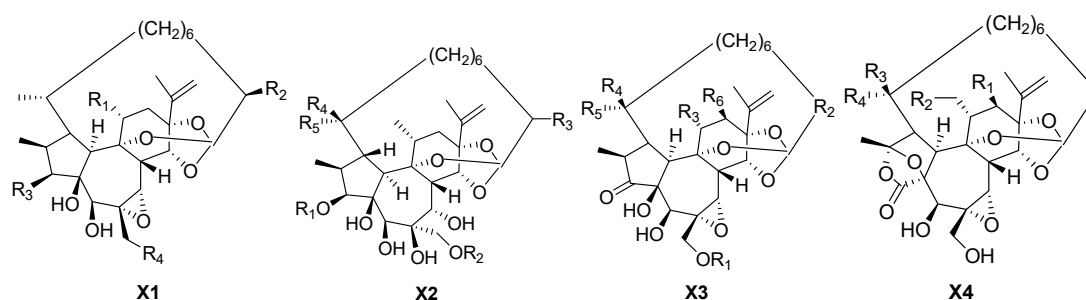


Figure 5. Four types (X1–X4) of 1-alkyldaphnanes skeletons.

Table 5. Reported structures of 1-alkyldaphnanes skeletons.

No.	Name	R ₁	R ₂	R ₃	R ₄	R ₅	R ₆	Type
151	Stellaralide A	CH ₂ OAc	OH	OBz	OH	–	–	X1
152	Stellaralide B	CH ₂ OBz	H	OBz	OH	–	–	X1
153	Gnidimacrin	CH ₂ OBz	OH	OBz	OH	–	–	X1
154	Genkwadane B	Me	H	OH	OBz	–	–	X1
155	Pimelea factor P ₂	CH ₂ OH	H	OBz	OH	–	–	X1
156	Genkwadane C	H	benzoyl	H	H	Me	–	X2
157	Wikstroelide R	H	benzoyl	OH	H	Me	–	X2
158	Wikstroelide S	benzoyl	H	H	Me	H	–	X2
159	Wikstroelide T	H	trans-cinnamoyl	H	H	Me	–	X2
160	Kirkinine B	H	CH=CH(CH ₂) ₅	Me	H	Me	H	X3
161	Kirkinine C	H	CH=CH(CH ₂) ₅	Me	H	Me	OAc	X3
162	Kirkinine E	H	CH=CH(CH ₂) ₅	Me	OH	Me	H	X3
163	Wikstroelide E	H	CH ₂	Me	H	Me	H	X3
164	Wikstroelide F	H	CH ₂	CH ₂ OBz	H	Me	H	X3
165	Wikstroelide G	palmitic acid	CH ₂	CH ₂ OBz	H	Me	H	X3
166	Wikstroelide K	CO(CH ₂) ₁₄ CH ₃	CH ₂	CH ₂ OBz	Me	H	H	X3
167	Wikstroelide O	H	CH ₂	CH ₂ OBz	Me	H	H	X3
168	Pimelea factor S ₆	OH	CH ₂	Me	H	Me	H	X3
169	Pimelea factor S ₇	OH	CH ₂	Me	Me	H	H	X3
170	Pimelotide A	H	H	Me	H	–	–	X4
171	Pimelotide B	OAc	H	H	Me	–	–	X4
172	Pimelotide C	H	H	H	Me	–	–	X4
173	Pimelotide D	OAc	H	Me	H	–	–	X4
174	Stellaralide C	H	OBz	Me	H	–	–	X4

3.5. Redioides

Redioides usually have a 12-carbon macrolide structure between C-3 and C-16, and have a special C-9, C-12, and C-14 orthoester structure (Figure 6, Table 6). The active compounds trigothysoids N–P (182–184), redioides A, C, and F (176–177, 179), and trigonosin F (181) have been obtained from the twigs and leaves of *Trigonostemonthyrsoideum*. Amongst them, compounds trigothysoid N, redioides A, C, and F, and trigonosins F have shown potent anti-HIV-1 activity, with EC_{50} values ranging from 0.001 to 0.015 nM. Additionally, trigothysoid O has been shown to exhibit moderate anti-HIV-1 activity [11], while redioid A has shown potent activities against mosquito larvae in an in-vitro assay study and against fleas (*Ctenocephalides felis*) in an artificial membrane feeding system, exhibiting LD_{90} values of 1 and 0.25 ppm, respectively [39]. Trigochilides A and B (175, 186) have been isolated from the twigs and leaves of *Trigonostemonchinensis* Merr. Trigochilide A has shown modest cytotoxicity against HL-60 (human leukemia) and BEL-7402 (human hepatoma), with demonstrated IC_{50} values of 3.68 and 8.22 μ M, respectively, whereas compound trigochilide B has only been shown to exhibit weak cytotoxicity against two tumor cell lines, with IC_{50} values of 33.35 and 54.85 μ M [1]. Compound redioid E (178) has been obtained from the roots of *Trigonostemonreidioides*, and has shown significant acaricidal activity on *D. pteronyssinus* [40]. Trigonosin E (180) and trigonostempene D (185) have been isolated from the twigs and leaves of *Trigonostemonthyrsoideum* [16,21]. Redioides B, G, and D (187–189) have been isolated from the *Trigonostemonreidioides*, and have been evaluated for their insecticidal properties in an anti-flea artificial membrane feeding assay (as detailed earlier). In this assay, redioides B and D exhibited LD_{90} values of 0.25 and 0.5 ppm, respectively, and thus were equipotent with redioid A (LD_{90} 0.25 ppm) [41].

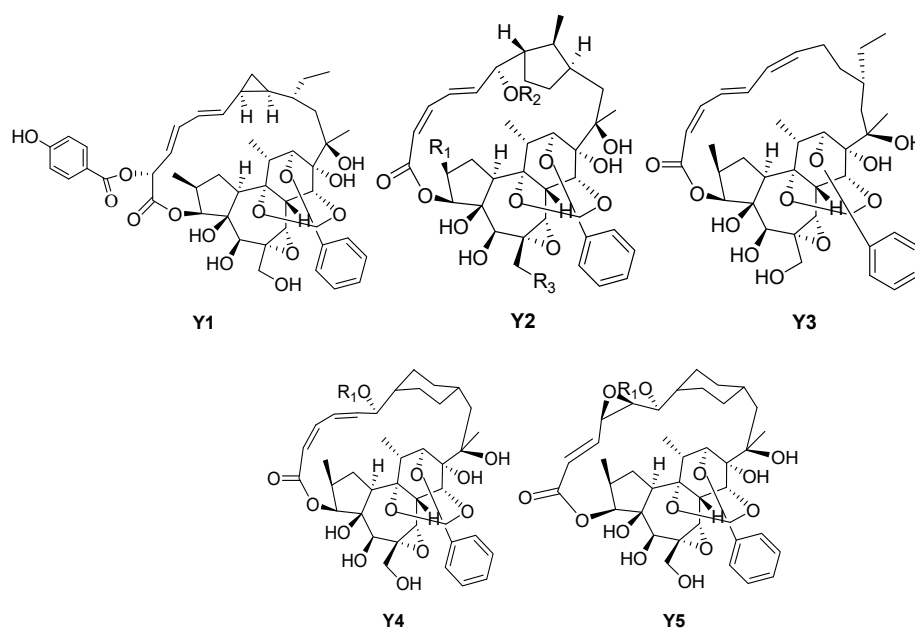


Figure 6. Five types (Y1–Y5) of redioides skeletons.

Table 6. Reported structures of redioides skeletons.

No.	Name	R ₁	R ₂	R ₃	Type
175	Trigochilide A	–	–	–	Y1
176	Redioid A	Me	COCH ₂ CH(CH ₃) ₂	OH	Y2
177	Redioid C	Me	Bz	OH	Y2
178	Redioid E	H	COCH ₂ CH(CH ₃) ₂	OH	Y2
179	Redioid F	H	Bz	OH	Y2
180	Trigonosin E	Me	COPh	OH	Y2
181	Trigonosin F	Me	COPh	OH	Y2

Table 6. Cont.

No.	Name	R ₁	R ₂	R ₃	Type
182	Trigothyoid N	Me	COCH ₂ CH(CH ₃) ₂	OH	Y2
183	Trigothyoid O	Me	COPh	H	Y2
184	Trigothyoid P	Me	COCH ₂ CH(CH ₃) ₂	H	Y2
185	Trigonostempene D	Me	Val	H	Y2
186	Trigochilide B	–	–	–	Y3
187	Rediocide B	COCH ₂ CH(CH ₃) ₂	–	–	Y4
188	Rediocide G	Bz	–	–	Y4
189	Rediocide D	COCH ₂ CH(CH ₃) ₂	–	–	Y5

4. Conclusions

It can be concluded that the bioactive activities of daphnane-type diterpenoids is obviously related to structure types. The most important points of them are the following: (1) The orthoester groups at C-9, C-13 and C-14 are essential to the cytotoxic activity. Daphnane-type diterpenoids with orthoester groups at C-9, C-13, and C-14 usually have stronger activity than daphnane-type diterpenoids with orthoester groups at C-9, C-12, and C-14 or C-12, C-13 and C-14. The absence of the orthoester group is unhelpful to the cytotoxic activity. (2) Specific to the 6-epoxyl groups, free 20-hydroxyl and 3-carbonyl are important for their activities. (3) Side chains at C-10 are crucial for cytotoxic activities. Generally speaking, long C-10 alkyl chains are more important than phenyl at C-10. Interestingly, the structure with macro-lactones exhibited much stronger activity than the others. Due to the rich activities of daphnane-type diterpenoids, researchers have not stopped exploring and researching such compounds and their bioactive activities from plants.

Funding: This research was funded by the National Natural Science Foundation of China [grant number 81860759].

Conflicts of Interest: There is no conflict of interest associated with the authors of this paper.

References

- Jayasuriya, H.; Zink, D.L.; Singh, S.B.; Borris, R.P.; Nanakorn, W.; Beck, H.T.; Balick, M.J.; Goetz, M.A.; Slayton, L.; Gregory, L.; et al. Structure and Stereochemistry of Rediocide A, a Highly Modified Daphnane from *Trigonostemon reidioides* Exhibiting Potent Insecticidal Activity. *ChemInform* **2000**, *31*, 4998–4999.
- Carney, J.R.; Krenisky, J.M.; Williamson, R.T.; Luo, J.; Carlson, T.J.; Hsu, V.L.; Moswa, J.L. Maprouneacin, a new daphnane diterpenoid with potent antihyperglycemic activity from *Maprounea africana*. *J. Nat. Prod.* **1999**, *62*, 345–347. [[CrossRef](#)] [[PubMed](#)]
- He, W.; Cik, M.; Lesage, A. Kirkinine, a new daphnane orthoester with potent neurotrophic activity from *Synaptolepis kirkii*. *J. Nat. Prod.* **2000**, *63*, 1185–1187. [[CrossRef](#)] [[PubMed](#)]
- He, W.; Cik, M.; Appendino, G. Daphnane-type diterpene orthoesters and their biological activities. *Mini-Rev. Med. Chem.* **2002**, *2*, 185–200. [[CrossRef](#)] [[PubMed](#)]
- Asada, Y.; Sukemori, A.; Watanabe, T.; Malla, K.J.; Yoshikawa, T.; Li, W.; Koike, K.; Chen, C.H.; Akiyama, T.; Qian, K.; et al. Stelleralides A-C, Novel Potent Anti-HIV Daphnane-Type Diterpenoids from *Stellera chamaejasme* L. *Org. Lett.* **2011**, *13*, 2904–2907. [[CrossRef](#)] [[PubMed](#)]
- Zhang, L.; Luo, R.H.; Wang, F.; Dong, Z.J.; Yang, L.M.; Zheng, Y.T.; Liu, J.K. Daphnane diterpenoids isolated from *Trigonostemon thyrsoides* as HIV-1 antivirals. *Phytochemistry* **2010**, *71*, 1879–1883. [[CrossRef](#)] [[PubMed](#)]
- Chen, S.K.; Chen, B.Y.; Li, H. *Flora of China (Zhongguo Zhiwu Zhi)*; Science: Beijing, China, 1997; Volume 44, pp. 162–170.
- Zhang, L.; Luo, R.H.; Wang, F.; Jiang, M.Y.; Dong, Z.J.; Yang, L.M.; Zheng, Y.T.; Liu, J.K. Highly functionalized daphnane diterpenoids from *Trigonostemon thyrsoides*. *Org. Lett.* **2010**, *12*, 152–155. [[CrossRef](#)]
- Wang, H.B.; Liu, L.P.; Wang, X.Y. ¹³C-NMR data of daphnane diterpenoids. *Magn. Reson. Chem.* **2013**, *51*, 580–592. [[CrossRef](#)]

10. Huang, S.Z.; Zhang, X.J.; Li, X.Y.; Kong, L.M.; Jiang, H.J.; Ma, Q.Y.; Liu, Y.Q.; Hu, J.M.; Zheng, Y.T.; Li, Y. Daphnane-type diterpene esters with cytotoxic and anti-HIV-1 activities from *Daphne acutiloba* Rehd. *Phytochemistry* **2012**, *75*, 99–107. [[CrossRef](#)]
11. Cheng, Y.Y.; Chen, H.; He, H.P.; Zhang, Y.; Li, S.F.; Tang, G.H.; Guo, L.L.; Yang, W.; Zhu, F.; Zheng, Y.T. Anti-HIV active daphnane diterpenoids from *Trigonostemon thyrsoideum*. *Phytochemistry* **2013**, *96*, 360–369. [[CrossRef](#)]
12. Abe, F.; Iwase, Y.; Yamauchi, T.; Kinjo, K.; Yaga, S.; Ishii, M.; Iwahana, M. Minor daphnane-type diterpenoids from *Wikstroemia retusa*. *Phytochemistry* **1998**, *47*, 833–837. [[CrossRef](#)]
13. Li, L.Z.; Gao, P.Y.; Peng, Y.; Wang, L.H.; Song, S.J. A novel daphnane-type diterpene from the flower bud of *Daphne genkwa*. *Chem. Nat. Compd.* **2010**, *46*, 380–382. [[CrossRef](#)]
14. Li, F.; Sun, Q.; Hong, L.; Li, L.; Wu, Y.; Xia, M.; Ikejima, T.; Peng, Y.; Song, S. Daphnane-type diterpenes with inhibitory activities against human cancer cell lines from *Daphne genkwa*. *Bioorg. Med. Chem. Lett.* **2013**, *23*, 2500–2504. [[CrossRef](#)] [[PubMed](#)]
15. Liu, F.; Yang, X.; Ma, J.; Yang, Y.; Xie, C.; Tuerhong, M.; Jin, D.Q.; Xu, J.; Lee, D.; Ohizumi, Y.; et al. Nitric oxide inhibitory daphnane diterpenoids as potential anti-neuroinflammatory agents for AD from the twigs of *Trigonostemon thyrsoideus*. *Bioorg. Chem.* **2017**, *75*, 149–156. [[CrossRef](#)] [[PubMed](#)]
16. Jo, S.K.; Hong, J.Y.; Park, H.J.; Lee, S.K. Anticancer Activity of Novel Daphnane Diterpenoids from *Daphne genkwa* through Cell-Cycle Arrest and Suppression of Akt/STAT/Src Signalings in Human Lung Cancer Cells. *Biomol. Ther. (Seoul)*. **2012**, *20*, 513–519. [[CrossRef](#)]
17. Taninaka, H.; Takaishi, Y.; Honda, G.; Imakura, Y.; Sezik, E.; Yesilada, E. Terpenoids and aromatic compounds from *Daphne oleoides* ssp. *oleoides*. *Phytochemistry (Oxford)* **1999**, *52*, 1525–1529. [[CrossRef](#)]
18. Yu, L.; Zuo, W.J.; Mei, W.L.; Guo, Z.K.; Li, X.N.; Dai, H.F. Three new terpenoids from *Trigonostemon xyphophylloides* (Croiz.) L.K. Dai and T.L. Wu. *Phytochem. Lett.* **2013**, *6*, 472–475. [[CrossRef](#)]
19. Chen, Y.Y.; Guo, J.M.; Qian, Y.F.; Guo, S.; Ma, C.H.; Duan, J.A. Toxicity of daphnane-type diterpenoids from *Genkwa Flos* and their pharmacokinetic profile in rat. *Phytomedicine* **2013**, *21*, 82–89. [[CrossRef](#)]
20. Li, S.F.; Di, Y.T.; Li, S.L.; Zhang, Y.; Yang, F.M.; Sun, Q.Y.; Simo, J.M.; He, H.P.; Hao, X.J. Trigonosins A–F, Daphnane Diterpenoids from *Trigonostemon thyrsoideum*. *J. Nat. Prod.* **2011**, *74*, 464–469. [[CrossRef](#)]
21. Miyamae, Y.; Villareal, M.O.; Abdrabbah, M.B.; Isoda, H.; Shigemori, H. Hirseins A and B, Daphnane Diterpenoids from *Thymelaea hirsuta* That Inhibit Melanogenesis in B16 Melanoma Cells. *J. Nat. Prod.* **2009**, *72*, 938–941. [[CrossRef](#)]
22. Tchinda, A.Y.; Tsopmo, A.; Tene, M.; Kamnaing, P.; Ngnokam, D.; Tane, P.; Ayafor, J.F.; Connolly, J.D.; Farrugia, L.J. Diterpenoids from *Neoboutonia glabrescens* (Euphorbiaceae). *Phytochemistry* **2003**, *64*, 549–553. [[CrossRef](#)]
23. Hayes, P.Y.; Chow, S.; Somerville, M.J.; Fletcher, M.T. Daphnane- and Tiglane-Type Diterpenoid Esters and Orthoesters from *Pimelea elongata*. *J. Nat. Prod.* **2010**, *73*, 1907–1913. [[CrossRef](#)] [[PubMed](#)]
24. Lin, B.D.; Han, M.L.; Ji, Y.C.; Chen, H.D.; Yang, S.P.; Zhang, S.; Geng, M.Y.; Yue, J.M. Trigoxypins A–G: diterpenes from *Trigonostemon xyphophylloides*. *J. Nat. Prod.* **2010**, *73*, 1301–1305. [[CrossRef](#)] [[PubMed](#)]
25. Powell, R.G.; Weisleder, D.; Smith, C.R. Daphnane Diterpenes from *Diarthron vesiculosum*: Vesiculosin and Isovesiculosin. *J. Nat. Prod.* **1985**, *48*, 102–107. [[CrossRef](#)]
26. Su, J.; Wu, Z. A New Daphnane-Type Diterpenoid from *Daphne giraldii*. *Chem. Nat. Compd.* **2014**, *50*, 285–287. [[CrossRef](#)]
27. Chen, H.D.; Yang, S.P.; He, X.F.; Liu, H.B.; Ding, J.; Yue, J.M. Trigochinins D–I: six new daphnane-type diterpenoids from *Trigonostemon chinensis*. *Tetrahedron* **2010**, *66*, 5065–5070. [[CrossRef](#)]
28. Wei, Y.L.; Yu, Z.L.; Huo, X.K.; Tian, X.G.; Feng, L.; Huang, S.S.; Deng, S.; Ma, X.C.; Jia, J.M.; Wang, C. Diterpenoids from the roots of *Euphorbia fischeriana* and their inhibitory effects on alpha-glucosidase. *J. Asian Nat. Prod. Res.* **2018**, *20*, 977–984. [[CrossRef](#)]
29. Bang, K.K.; Yun, C.Y.; Lee, C.; Jin, Q.; Lee, J.W.; Jung, S.H.; Lee, D.; Lee, M.K.; Hong, J.T.; Kim, Y.; et al. Melanogenesis inhibitory daphnane diterpenoids from the flower buds of *Daphne genkwa*. *Bioorg. Med. Chem. Lett.* **2013**, *23*, 3334. [[CrossRef](#)]
30. Zhan, Z.J.; Fan, C.Q.; Ding, J.; Yue, J.M. Novel diterpenoids with potent inhibitory activity against endothelium cell HMEC and cytotoxic activities from a well-known TCM plant *Daphne genkwa*. *Bioorg. Med. Chem.* **2005**, *13*, 645–655. [[CrossRef](#)]

31. Zhang, X.D.; Ni, W.; Yan, H.; Li, G.T.; Zhong, H.M.; Li, Y.; Liu, H.Y. Daphnane-Type Diterpenoid Glucosides and Further Constituents of *Euphorbia pilosa*. *Chem. Biodivers.* **2014**, *11*, 760–766. [[CrossRef](#)]
32. Yang, B.; Meng, Z.; Li, Z.; Sun, L.; Hu, Y.; Wang, Z.; Ding, G.; Xiao, W.; Han, C. Three daphnane diterpenoids from *Trigonostemon xyphophylloides*. *Phytochem. Lett.* **2015**, *11*, 270–274. [[CrossRef](#)]
33. Li, S.F.; Zhang, Y.; Huang, N.; Zheng, Y.T.; Di, Y.T.; Li, S.L.; Cheng, Y.Y.; He, H.P.; Hao, X.J. Daphnane diterpenoids from the stems of *Trigonostemon lii* and their anti-HIV-1 activity. *Phytochemistry* **2013**, *93*, 216–221. [[CrossRef](#)]
34. Pejin, B.; Iodice, C.; Tommonaro, G. Synthesis and biological activities of thio-avarol derivatives. *J. Nat. Prod.* **2008**, *71*, 1850–1853. [[CrossRef](#)]
35. Tommonaro, G.; Vitale, R.M.; Pejin, B.; Iodice, C.; Canadas, S. Avarol derivatives as competitive AChE inhibitors, non hepatotoxic and neuroprotective agents for Alzheimer's disease. *Eur. J. Med. Chem.* **2016**, *122*, 326–338. [[CrossRef](#)]
36. Yang, B.; Chen, G.Y.; Song, X.P.; Yang, L.Q.; Han, C.R.; Wu, X.Y.; Li, X.M.; Zou, B.Y. Trigoxylins H and I: Two new daphnane diterpenoids from *Trigonostemon xyphophylloides*. *ChemInform* **2012**, *22*, 3828–3830. [[CrossRef](#)]
37. Dong, S.H.; Zhang, C.R.; Xu, C.H.; Ding, J.; Yue, J.M. Daphnane-Type Diterpenoids from *Trigonostemon howii*. *J. Nat. Prod.* **2011**, *74*, 1255–1261. [[CrossRef](#)]
38. Guo, J.; Tian, J.; Yao, G.; Zhu, H.; Xue, Y.; Luo, Z.; Zhang, J.; Zhang, Y. Three new 1 α -alkyldaphnane-type diterpenoids from the flower buds of *Wikstroemia chamaedaphne*. *Fitoterapia* **2015**, *106*, 242–246. [[CrossRef](#)]
39. Hayes, P.Y.; Chow, S.; Somerville, M.J.; Fletcher, M.T. Pimelotides A and B, Diterpenoid Ketal-Lactone Orthoesters with an Unprecedented Skeleton from *Pimelea elongata*. *J. Nat. Prod.* **2009**, *72*, 2081–2083. [[CrossRef](#)]
40. Chen, H.D.; He, X.F.; Ai, J.; Geng, M.Y.; Yue, J.M. Trigochilides A and B, Two Highly Modified Daphnane-Type Diterpenoids from *Trigonostemon chinensis*. *Org. Lett.* **2009**, *11*, 4080–4083. [[CrossRef](#)]
41. Jayasuriya, H.; Zink, D.L.; Borris, R.P.; Nanakorn, W.; Beck, H.T.; Balick, M.J.; Goetz, M.A.; Gregory, L.; Shoop, W.L.; Singh, S.B. Rediocides B-E, Potent Insecticides from *Trigonostemon reidioides*. *J. Nat. Prod.* **2004**, *67*, 228–231. [[CrossRef](#)]

Sample Availability: Samples of the compounds are available from the authors.



© 2019 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (<http://creativecommons.org/licenses/by/4.0/>).