SUPPLEMENTARY MATERIAL

LC-MS- Based Metabolomics for the Chemosystematics of Kenyan *Dodonaea viscosa* Jacq (Sapindaceae)

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Abstract.

Dodonaea viscosa Jacq (Sapindaceae) is a medicinal plant with a world-wide distribution. The species has undergone enormous taxonomic changes which causes confusion amongst plant users. In Kenya for example, two varieties are known to exist based on morphology: D. viscosa var viscosa along the coast, and D. viscosa var angustifolia in the Kenvan inland; these two taxa are recognised as distinct species in some reports. This prompted us to apply metabolomics to understand the relationship among naturally occurring populations of D. viscosa in Kenya, and to identify compounds that can assist in taxonomic delineation of the different varieties of D. viscosa from different parts of Kenya. The phytochemical variability of Kenyan D. viscosa var. angustifolia populations collected from four different geographical regions (Nanyuki, Machakos, Nairobi, Narok) and one coastal D. viscosa var viscosa (the Gazi) were analysed by LC-MS using a metabolomics-driven approach. Four known compounds, two diterpenoids (dodonic acid (1), hautriwaic acid lactone (3) and two flavonoids [5,7,4',5'-tetrahydroxy-3,6,2'-trimethoxyflavone (2) and catechin (4) were isolated and purified from the Gazi, coastal collection. The presence of these compounds and their relative abundance in other populations was determined by LC-MS analyses. The use of multivariate statistical analyses of LC-MS data allowed the visualization of the patterns of variation and identification of additional compounds. Eleven discriminant compounds responsible for separating chemometric clusters were tentatively identified. The clustering pattern of the five populations of D. viscosa suggested that the metabolite profiles were influenced by geo-environmental conditions and did not support the current classification of D. viscosa based on morphology. This study disputes the current classification of D. viscosa in Kenya and recommends revision using tools such as molecular phylogenetics. In an antimicrobial assay, hautriwaic acid lactone (3) and catechin (4) were the most active compounds followed by the extract from coastal (Gazi) population.

Keywords: African traditional medicine; antimicrobial activity; chemosystematics; Hopbush; metabolomics; natural products chemistry; phenolics; terpenoids

1.1. General characteristics of the isolated compounds 1-4

Dodonic acid (1) [1,2] white powder with molecular formula $C_{20}H_{28}O_4$ based on electrospray ionization mass spectrometry (ESIMS)([M-H] m/z 331.1910). 5,7,4',5'-Tetrahydroxy-3,6,2' –trimethoxyflavone (2) [3] yellow amorphous solid, $C_{18}H_{16}O_9$, ESIMS ([M-H] m/z 375.0723). Hautriwaic acid lactone (3) [1] $C_{20}H_{26}O_3$, ESIMS ([M-H] m/z 313.2361) white amorphous solid active under UV (254 nm). Catechin (4) [4] brown solid active under UV (254-366 nm) with ESIMS of ([M-H] m/z 289.0719), $C_{15}H_{14}O_6$.





5,7,4',5'-Tetrahydroxy-3,6,2' -trimethoxyflavone (2)



Hautriwaic acid lactone (3)

Figure S1. Chemical structures of compounds 1-4 isolated from coastal plants of D. viscosa

1.2. Spectroscopic data for compounds 1-4 isolated from *D. viscosa*

Position	δc	δ _H (m, J in Hz)	HMBC
1	17.2	1.74 (m)	
2	27.5	2.35 (m)	
3	143.9	7.25 (m)	
4	140.2		
5	44.7		
6	74.6	3.77 (dd, 4.9, 10.8)	
7	35.8	1.67 (m)	
8	33.6	1.77 (m)	C-9
9	38.7		
10	45.7	1.42 (m)	C-2, C-4, C-9
11	38.6	1.66 (m)	
12	17.2	1.77 (m)	C-9
13	125.3		
14	138.4	7.26 (br s)	C-13, C-15, C-16
15	142.6	7.38 (br s)	C-13
16	110.9	6.32 (br s)	C-13, C-14, C-15
17	15.3	0.9 (d, 6.6)	C-7, C-9
18	173.1		
19	16.3	1.26 (s)	C-3,C-6, C-10
20	17.8	0.8 (s)	C-8, C-10, C-11

Table S1: ¹H (500 MHz), ¹³C (125 MHz) NMR data along with HMBC correlations (in CD_2Cl_2) for dodonic acid (1)

TableS2: ¹H (500 MHz), ¹³C (125 MHz) NMR data along with HMBC correlations (in CD_2Cl_2) for 5,7,4',5'-tetrahydroxy-3,6,2'-trimethoxyflavone (**2**)

Position	δς	δ _H (m)	НМВС
2	155.9		
3	136.4		
4	177.8		
5	151.2		
6	130.2		
7	155.3		
8	93.3	6.60 (s)	C-6, C-7, C-9, C-10
9	152.8		
10	106.0		
1'	109.8		
2'	151.8		
3'	102.2	6.66	C-1', C-4', C-5'
4'	150.2		
5'	139.9		

6'	113.3	7.26 (s)	C-2, C-5'
2 '-OMe	56.15	3.98	C-2'
3-OMe	61.90	3.88	C-3
6-OMe	60.73	4.05	C-6
OH-4'		7.84 (s)	
OH-7		6.67 (s)	
OH-5		12.74 (s)	_

Table S3: ¹H (500 MHz), ¹³C (125 MHz) NMR data along with HMBC correlations (in CDCl₃) for hautriwaic acid lactone (**3**)

Position	δc	δн	HMBC
1	17.9	1.65 (m)	
		1.47 (m)	
2	27.6	2.19 (m)	
3	135.9	6.68 (m)	C-4, C-5, C-18
4	141.5		
5	42		
6	34.5	1.42 (m)	
7	26.6	1.68 (m)	
		1.38 (m)	
8	16.7		
9	19.5		
10	45.7	1.14 (m)	
11	38.6	1.48 (m)	
12	17.2	2.43 (m)	
13	125.3		
14	138.5	7.26 (s)	C-13, C-16, C-15
15	142.7	7.37	
16	110.9	6.31 (br s)	
17	15.3	0.88 (d, 6.6)	
18	169.6		
19	65.1	4.12 (d, 4.2)	C-4, C-5, C-6, C-10, C-18
		3.72 (d, 3.8)	C-4, C-5, C-6, C-10
20	17.3	0.78 (s)	C-17, C-19

Position	δς	δ н (m , <i>J</i> in Hz)	HMBC
2	82.9	4.57 (d, 7.5)	C-3, C-1', C-2', 6 '
3	68.8	3.98 (td, 5.4, 7.8)	
4	28.5	2.51 (dd, 8.1, 16.1)	C-2, C-3, C-5, C-9, C-10
5	157.6		
6	96.4	4.93 (d, 2.3)	C-8, C-10
7	157.9		
8	95.6	5.86 (d, 2.3)	C-6, C-10
9	156.9		
10	100.9		
1'	132.3		
2'	115.3	6.84 (d, 2.0)	C-2, C-4' C-6'
3'	146.2		
4'	146.3		
5'	116.1	6.76 (d, 8.1)	C-2, C-1' C-3'
6'	120.0	6.72 (dd, 2.0, 8.2)	C-2, C-2', C-6'

Table S4: 1 H (500 MHz), 13 C (125 MHz) NMR data along with HMBC correlations (in CD₃OD) for catechin (4)

1.2.NMR and MS Spectra of the isolated compounds 1-4

1.2.1. Dodonic acid (1)



Figure S2. The ¹H NMR spectrum of dodonic acid (1) observed at 500 MHz in CD₂Cl₂ solution at 25 °C. Assignment is provided in Table S1.



Figure S3. The ¹³C NMR spectrum of dodonic acid (1) observed at 500 MHz in CD_2Cl_2 solution at 25 °C. Assignment is provided in Table S1.



Figure S4. The ${}^{1}H{}^{-1}H$ COSY NMR spectrum of dodonic acid (1) observed at 500 MHz in CD₂Cl₂ solution at 25 °C.



Figure S5. The ¹H-¹³C HSQC NMR spectrum of dodonic acid (1) observed at 500 MHz in CD_2Cl_2 solution at 25 °C.



Figure S6. The ${}^{1}\text{H}{}^{-13}\text{C}$ HMBC NMR spectrum of dodonic acid (1) observed at 500 MHz in CD₂Cl₂ solution at 25 °C. Assignment is provided in Table S1.



Figure S7. ESIMS spectrum for dodonic acid (1)





Figure S8. The ¹H NMR spectrum of 5, 7, 4', 5' -tetrahydroxy-3, 6, 2' -trimethoxyflavone (2) observed at 500 MHz in CD_2Cl_2 solution at 25 °C. Assignment is provided in Table S2.



Figure S9. ¹³C NMR spectrum of 5,7,4',5'-tetrahydroxy-3,6,2'-trimethoxyflavone (2) observed at 500 MHz in CD₂Cl₂ solution at 25 °C. Assignment is provided in Table S2.



Figure S10. ¹H- ¹³C HSQC spectrum of 5,7,4',5'-tetrahydroxy-3,6,2'-trimethoxyflavone (**2**) observed at 500 MHz in CD₂Cl₂ solution at 25 °C.



Figure S11. ¹H- ¹³C HMBC spectrum of 5,7,4',5'-tetrahydroxy-3,6,2'-trimethoxyflavone (2) observed at 500 MHz in CD_2Cl_2 solution at 25 °C. Assignment is provided in Table S2.





Figure S12. ESIMS spectrum for 5,7,4',5' -tetrahydroxy-3,6,2'-trimethoxyflavone (2)

1.3.3. Hautriwaic acid lactone (3)



Figure S13. The ¹H- NMR spectrum of hautriwaic acid lactone (**3**) observed at 500 MHz in CDCl₃ solution at 25 °C. Assignment is provided in Table S3.



Figure S14. ¹³C- NMR spectrum of hautriwaic acid lactone (**3**) observed at 500 MHz. in CDCl₃ solution at 25 °C. Assignment is provided in Table S3.



Figure S15. The ¹H- ¹H COSY- NMR spectrum of hautriwaic acid lactone (**3**) observed at 500 MHz in CDCl₃ solution at 25 °C.



Figure S16. The ¹H- ¹³C- HSQS NMR spectrum of hautriwaic acid lactone (**3**) observed at 500 MHz in CDCl₃ solution at 25 °C.



Figure S17. The ¹H- ¹³C- HMBC NMR spectrum of hautriwaic acid lactone (**3**) observed at 500 MHz in CDCl₃ solution at 25 °C. Assignment is provided in Table S3.

MK_Bot_190627_11.raw#1152 @8.98 MS1 c -, base peak: 313.2361 m/z (1.3E3)



Figure S18. ESIMS for hautriwaic acid lactone (3)



1.3.4. Catechin (4)

Figure S19. The ¹H NMR spectrum of catechin (**4**) observed at 500 MHz in MeOD solution at 25 °C. Assignment is provided in Table S4.



Figure S20. The 13 C NMR spectrum of catechin (4) observed at 500 MHz in MeOD solution at 25 °C. Assignment is provided in Table S4.



Figure S21. The ¹H-¹H COSY spectrum of catechin (4) observed at 500 MHz in MeOD solution at 25 °C.



Figure S22. The 1 H- 13 C HSQC NMR spectrum of catechin (4) observed at 500 MHz in MeOD solution at 25 °C.



Figure S23. The ¹H-¹³C HMBC NMR spectrum of catechin (**4**) observed at 500 and 125 MHz in MeOD solution at 25 °C. Assignment is given in Table S4

D4.mzML#403 @3.16 MS1 c -, base peak: 289.0714 m/z (9.4E3)





Figure S24. ESIMS for catechin (4).

1.3.Random permutation and k-fold cross-validation of discriminant chemicals for Kenyan *D. viscosa*



Figure S25. Random permutation of discriminant chemicals for the five populations of *D*. *viscosa* in Kenya

Cluster 1, n=45



Figure S26. k-fold cross-validation of discriminant chemicals for the five populations of *D. viscosa* in Kenya

References

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