

## Supporting Information

# Antimicrobial metabolites against Methicillin-Resistant *Staphylococcus aureus* from the endophytic fungus *Neofusicoccum australe*

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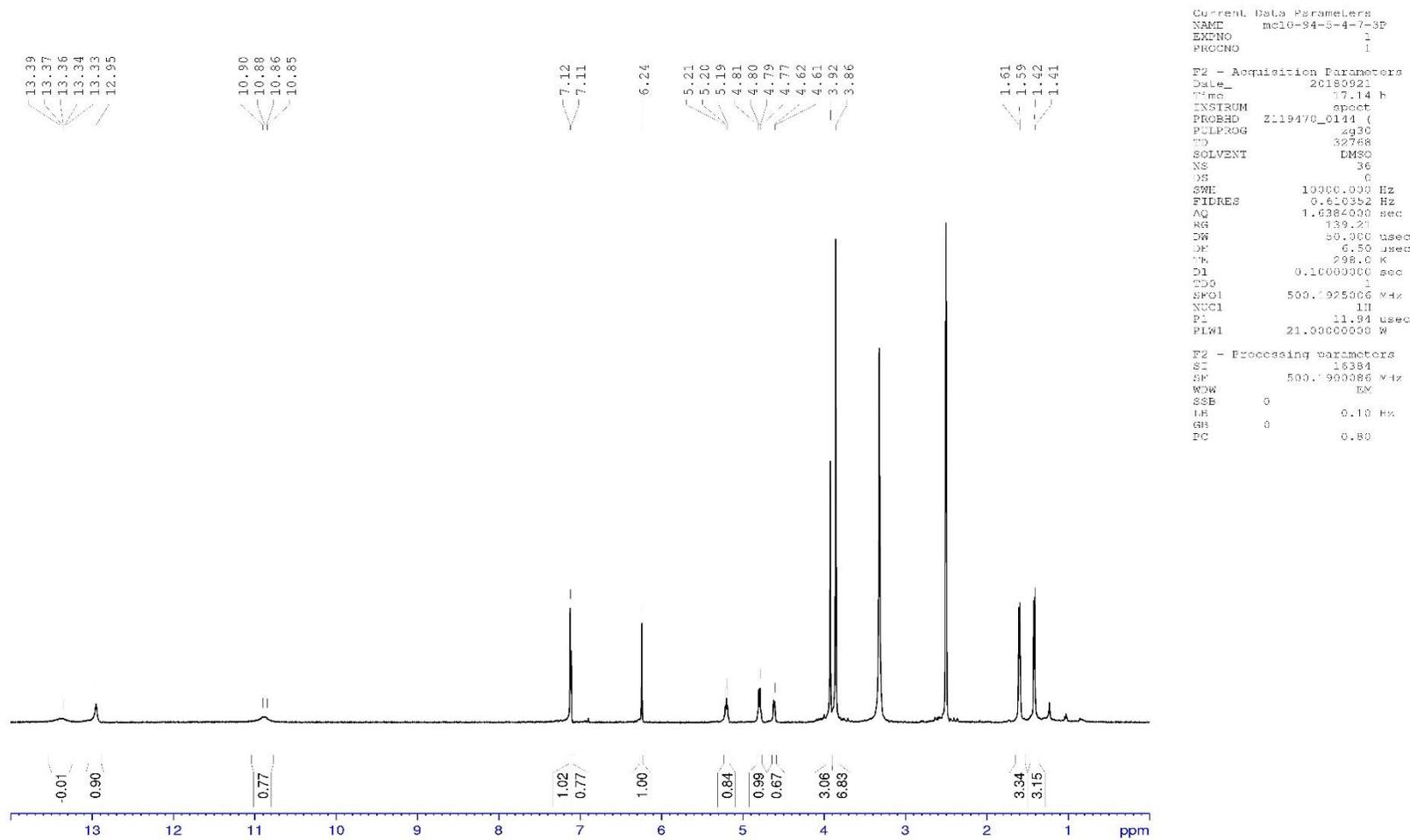


Figure S1 <sup>1</sup>H NMR spectrum (500 MHz, DMSO) of neofusnaphthoquinone B (1)

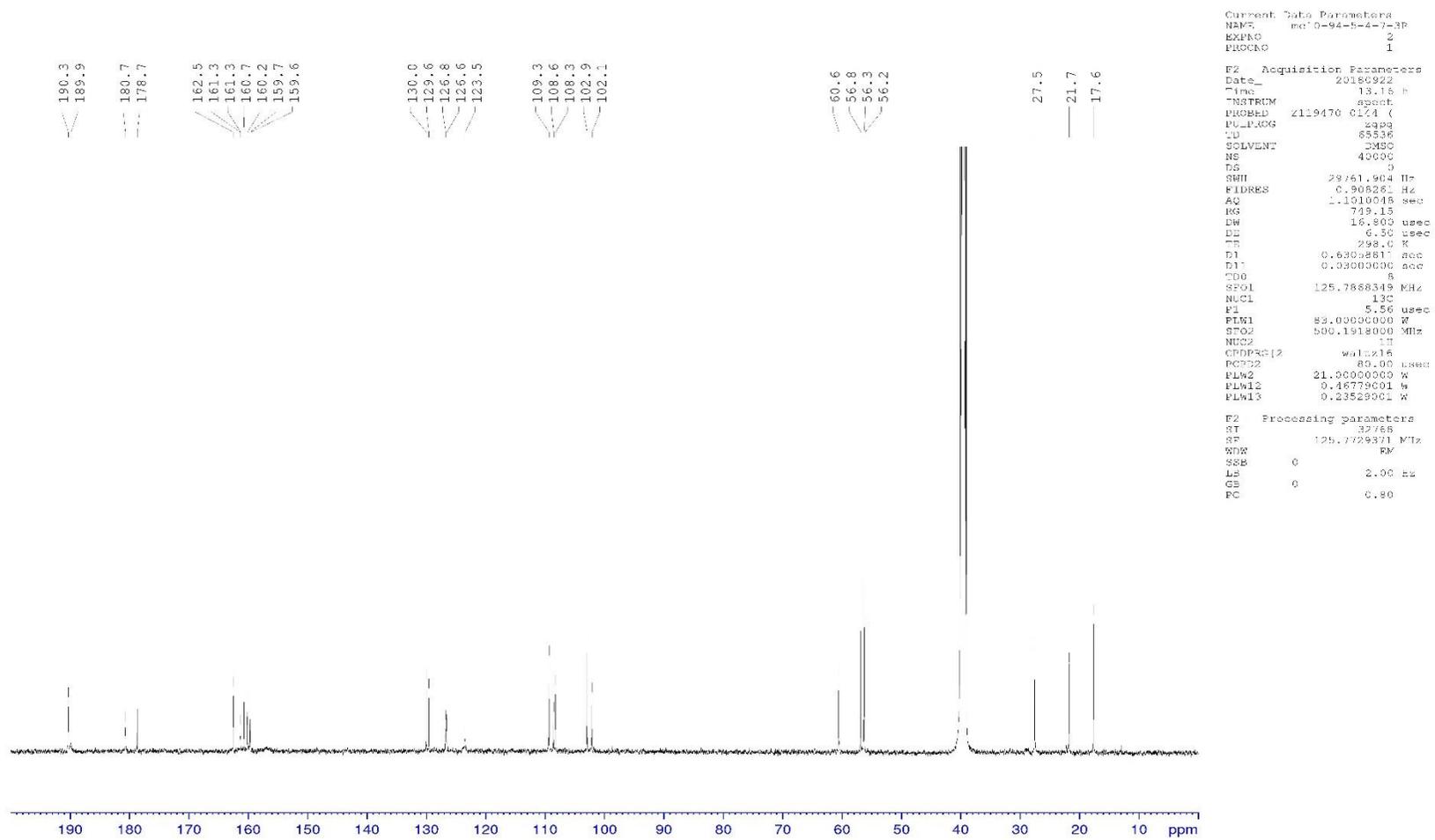


Figure S2  $^{13}\text{C}$  NMR spectrum (125 MHz, DMSO) of neofusnaphthoquinone B (1)

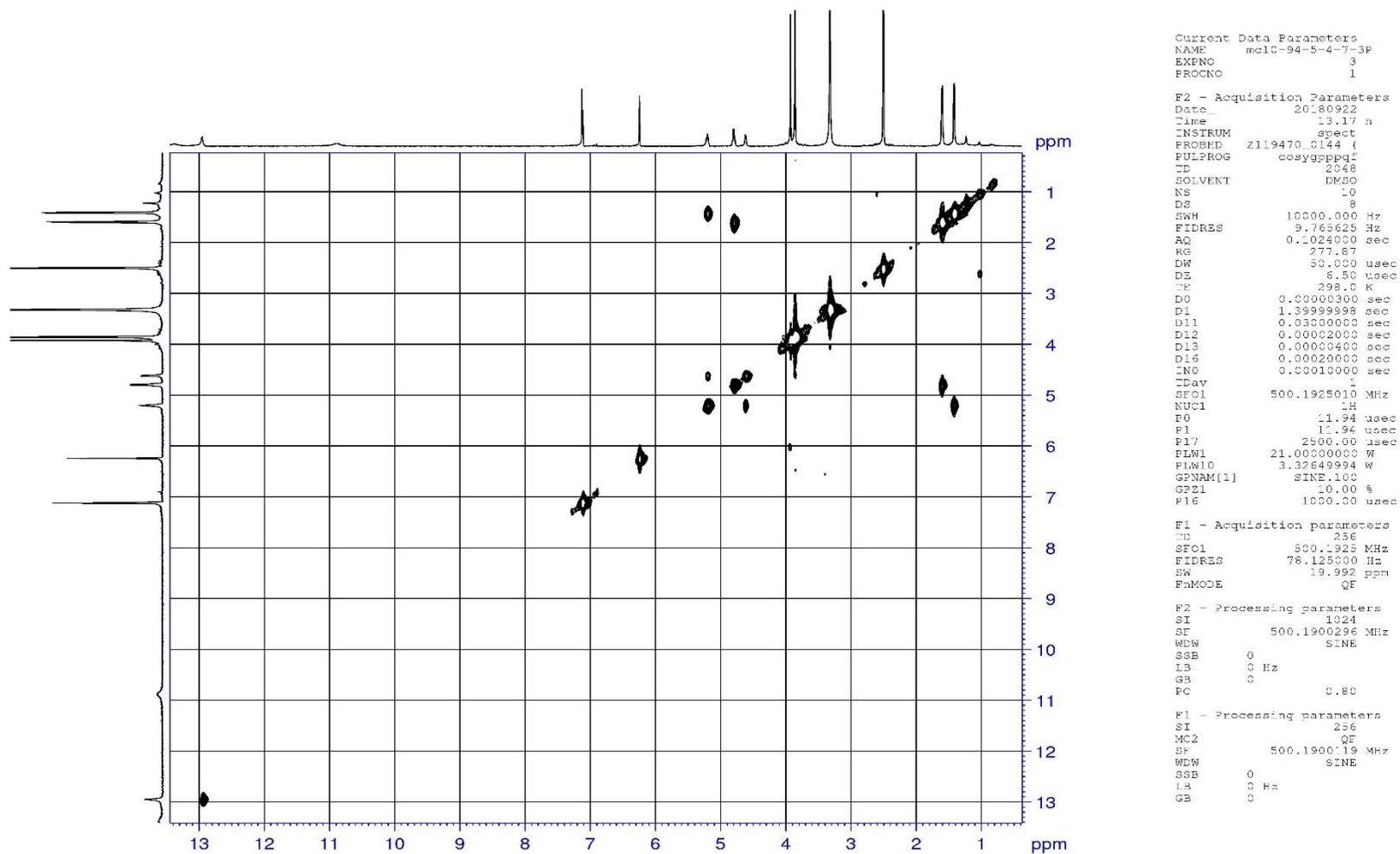


Figure S3 COSY spectrum of neofusnaphthoquinone B (1)

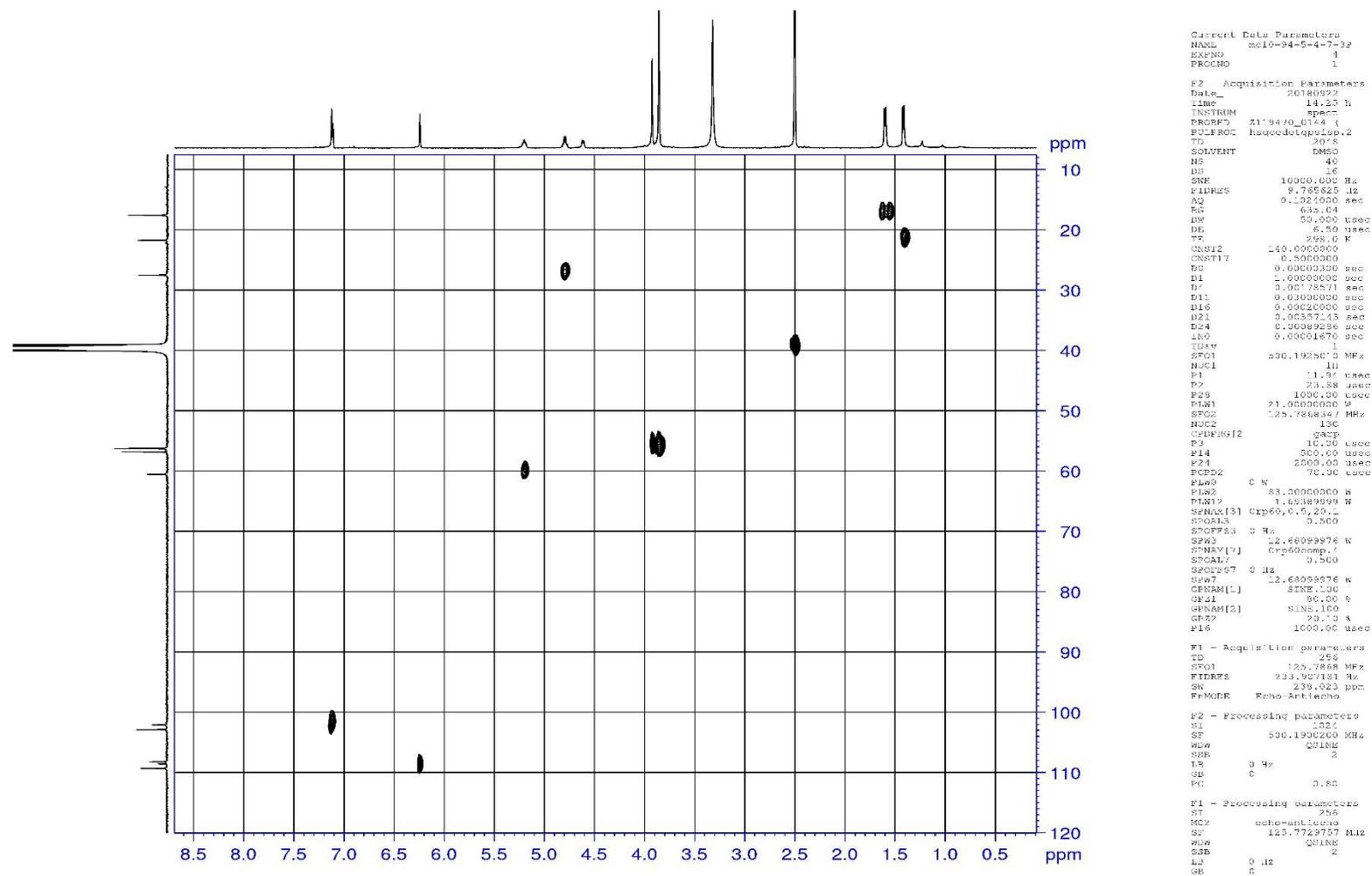


Figure S4 HSQC spectrum of neofusnaphthoquinone B (1)

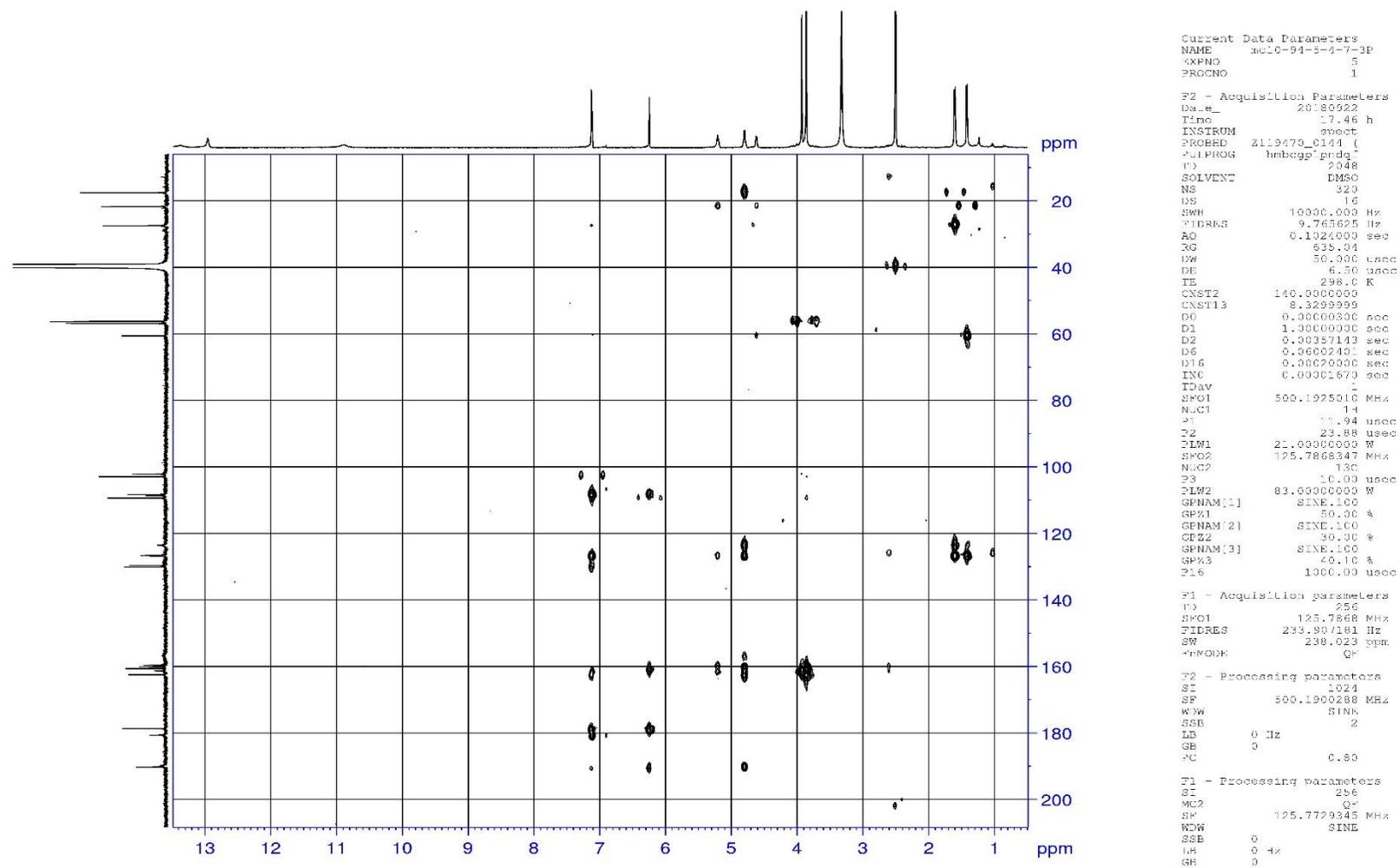
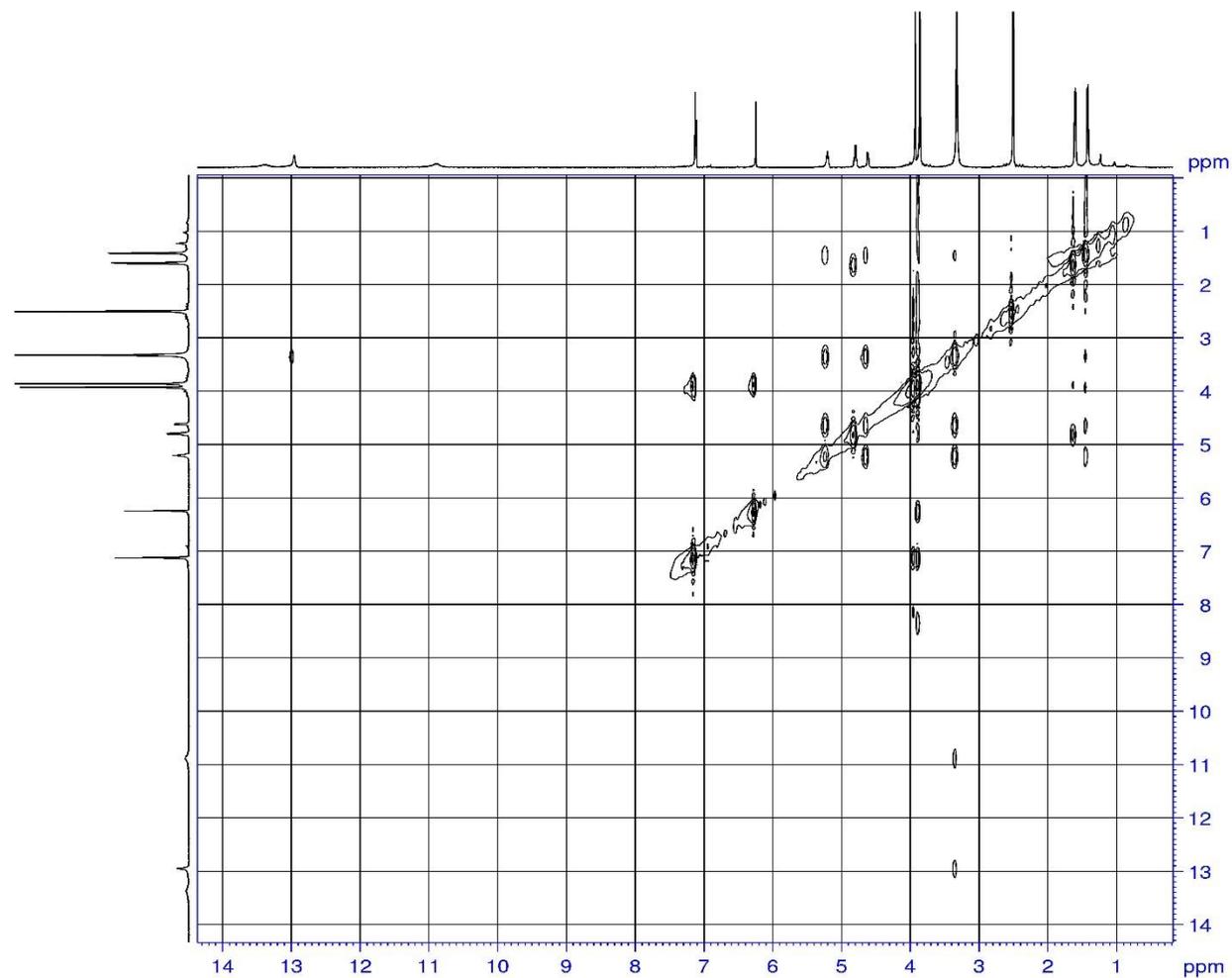


Figure S5 HMBC spectrum of neofusnaphthoquinone B (1)



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Current Data Parameters
NAME      mcl0-94-5-4-7-3P
EXPNO    6
PROCNO   1

F2 - Acquisition Parameters
Date_    20180923
Time     20.32 h
INSTRUM  spect
PROBHD   zll9470.0144.4
PULPROG  roesyphpp
TD        2048
SOLVENT  DMSO
NS        200
DS        16
SWH       10000.000 Hz
FIDRES    9.765625 Hz
AQ        0.2024000 sec
RG         635.04
DW         50.000 usec
DE         6.50 usec
TE         298.0 K
D0         0.00003840 sec
D1         1.0000000 sec
D11        0.0000000 sec
D12        0.00002000 sec
IN0        0.00010000 sec
TDav       1
SF01      500.1925010 MHz
NUC1       1H
F1         11.94 usec
F15        23000.00 usec
P17        2500.00 usec
PLW1       21.0000000 W
PLW10      3.32649994 W
PLW11      0.24742000 W

F1 - Acquisition parameters
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SF01      500.1925 MHz
FIDRES    123.456787 Hz
SW         19.992 ppm
FmMODE    TPFI

F2 - Processing parameters
SI         1024
SF         500.1900000 MHz
WDW        QSINE
SSB        2
LB         0 Hz
GB         0
PC         0.80

F1 - Processing parameters
SI         256
MC2        TPFI
SF         500.1900000 MHz
WDW        QSINE
SSB        2
LB         0 Hz
GB         0

```

Figure S6 ROESY spectrum of neofusnaphthoquinone B (1)

### Acquisition Parameter

Source Type	ESI	Ion Polarity	Positive	Set Nebulizer	0.4 Bar
Focus	Not active	Set Capillary	4500 V	Set Dry Heater	180 °C
Scan Begin	50 m/z	Set End Plate Offset	-500 V	Set Dry Gas	4.0 l/min
Scan End	1000 m/z	Set Collision Cell RF	150.0 Vpp	Set Divert Valve	Waste

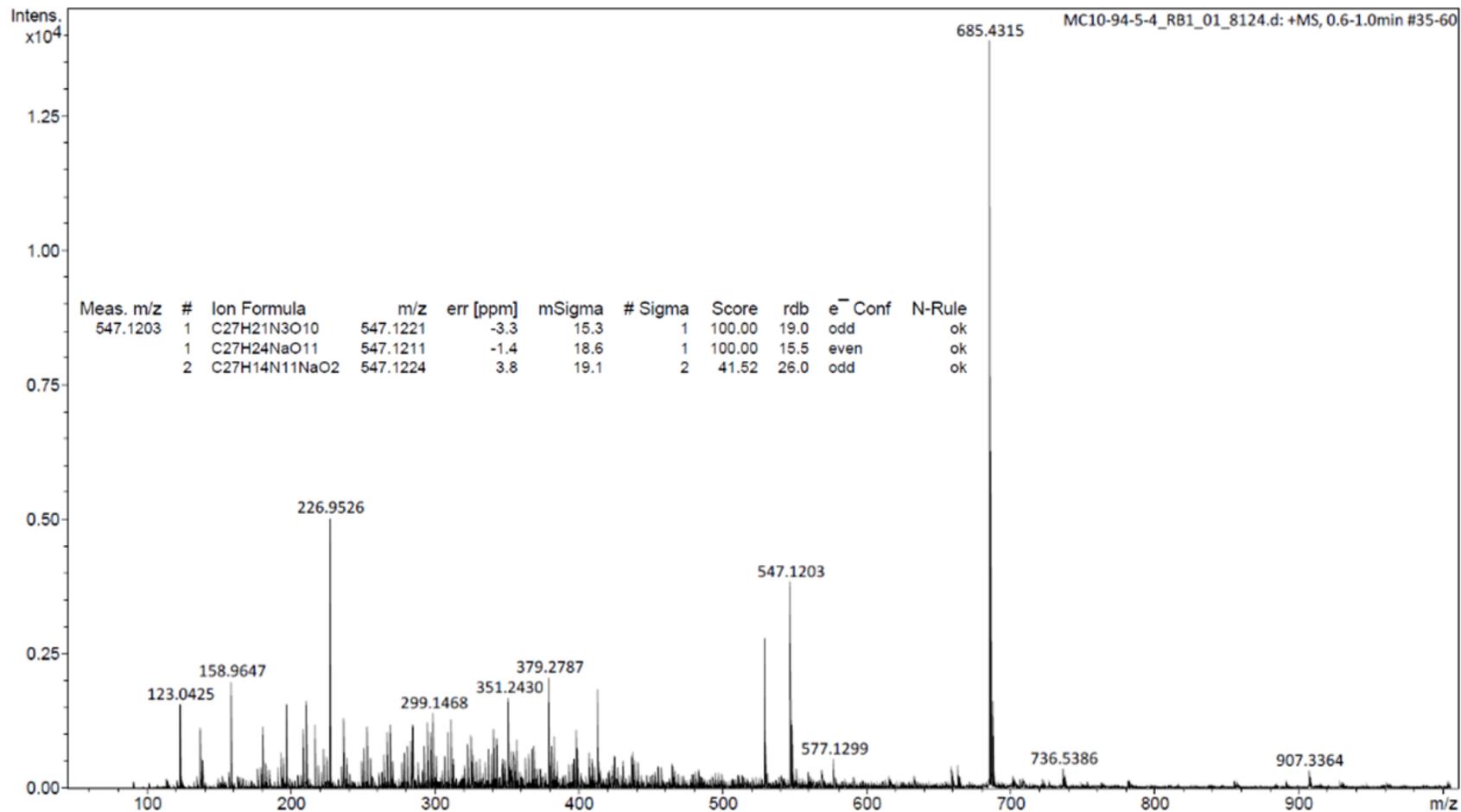
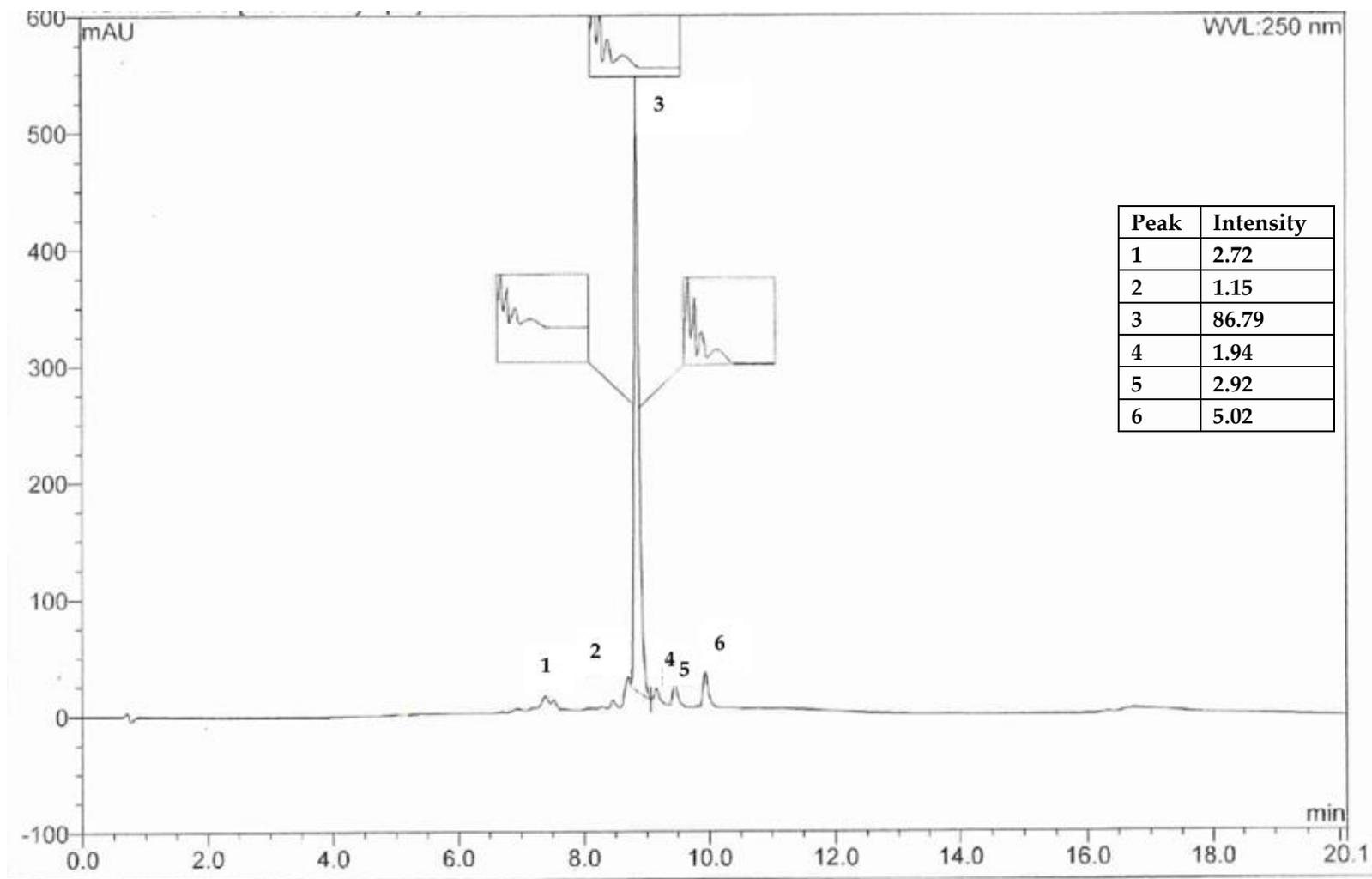


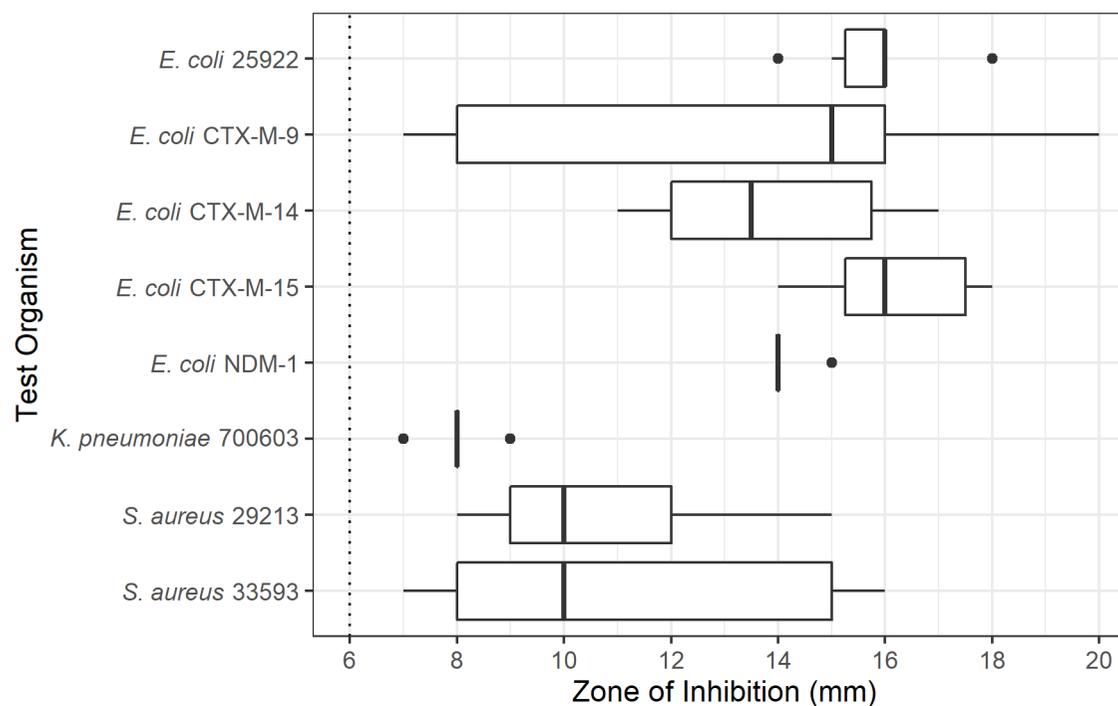
Figure S7 HRESIMS of neofusnaphthoquinone B (1)



**Figure S8** HPLC trace of neofusnaphthoquinone B (1) (Analytical reversed-phase HPLC on a Dionex UltiMate 3000RS using a C<sub>8</sub> column (3 μm Platinum, 33 × 7 mm) and eluting with a linear gradient of H<sub>2</sub>O (0.05% TFA) to MeCN over 20 mins at 2 mL/min and monitoring at 216 nm).

**Figure S9** Antibacterial activity testing using the zone of inhibition (ZOI) assay

To perform zone of inhibition (ZOI) testing, potato dextrose agar (PDA) plates were inoculated with a lawn of either antibiotic-sensitive *Escherichia coli* (ATCC 25922) or resistant clinical isolates (CTX-M-9, CTX-M-14, CTX-M-15, NDM-1) or antibiotic-resistant *Klebsiella pneumoniae* (ATCC 700603). Similarly, Mueller-Hinton agar plates were inoculated with a lawn of either antibiotic-sensitive *Staphylococcus aureus* (ATCC 29213) or antibiotic-resistant *S. aureus* (ATCC 33593). *Neofusicoccum australe* was grown on PDA plates and fungal plugs removed using a 6 mm punch biopsy tool (Catalogue number: SH241, Amtech Medical, New Zealand). Fungal plugs were placed onto the bacterial lawns, alongside PDA plugs containing no fungus. Plates were incubated inverted at 37°C for 24 h before measuring any zones of inhibition (in mm) produced.



**Antibacterial activity of *Neofusicoccum australe* (ICMP 21498) against *E. coli*, *K. pneumoniae*, and *S. aureus*.**

The dotted line represents the diameter of the fungal plugs with no activity. Data is presented as box-whisker plots. Raw data is available at DOI:

10.17608/k6.auckland.11888184.