

*Supplementary Materials*

# Persulfated Ascorbic Acid Glycoside as a Safe and Stable Derivative of Ascorbic Acid for Skin Care Application

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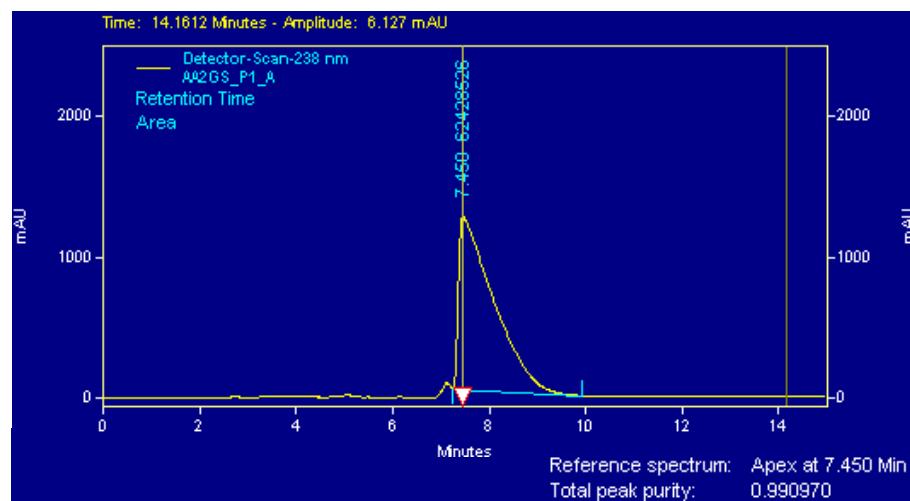
**Table S1**  $^1\text{H}$  and  $^{13}\text{C}$  NMR data of 2-O- $\alpha$ -D-glucopyranosyl-L-ascorbic acid persulfate (AAGS) and ascorbic acid persulfate (AscS) (DMSO- $d_6$ ).

$^1\text{H}^{\text{a}}$	$\delta$ (ppm)		$^{13}\text{C}^{\text{b}}$	$\delta$ (ppm)	
	AAGS	AscS*		AAGS	AscS*
<b>1</b>	-	-	1	176.5	173.1
<b>2</b>	-	-	2	114.2	95.7
<b>3</b>	-	-	3	173.6	154.2
<b>4</b>		4.29 (m)	4	114.2	92.4
<b>5</b>	3.94-4.10 (m)	4.52 (q)	5	70.4	70.2
<b>6a</b>		4.14 (q)			
<b>6b</b>	3.70-3.83 (m)	3.73 (t)	6	63.3	63.1
<b>1'</b>	5.53 (d, $J = 2.9$ Hz)	-	1'	97.3	-
<b>2'</b>	4.52-4.62 (m)	-	2'	74.0	-
<b>3'</b>	4.52-4.62 (m)	-	3'	75.4	-
<b>4'</b>	4.31 (brt)	-	4'	71.0	-
<b>5'</b>	4.13-4.18 (m)	-	5'	74.0	-
<b>6'a</b>	3.94-4.10 (m)	-			
<b>6'b</b>	3.70-3.83 (m)	-	6'	65.9	-

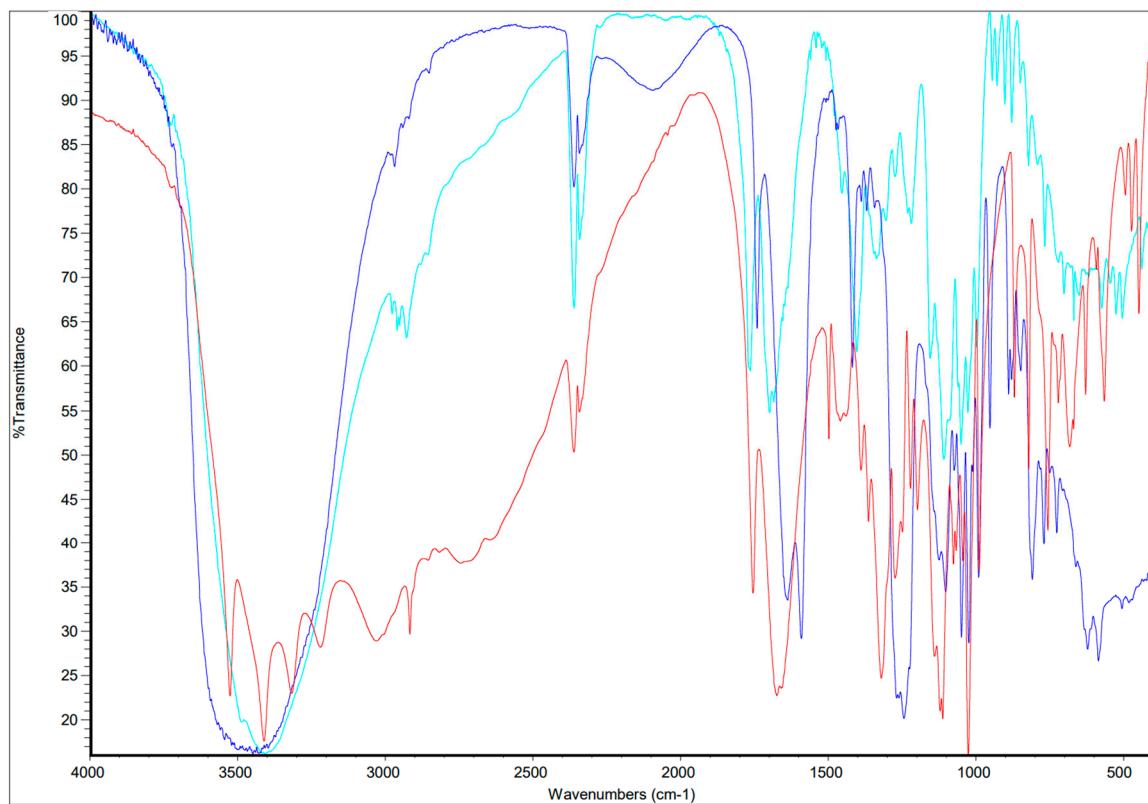
<sup>a</sup>Measured in DMSO- $d_6$  at 300.13 MHz. Values in parts per million ( $\delta_{\text{H}}$ ).

<sup>b</sup>Measured in DMSO- $d_6$  at 75.47 MHz. Values in parts per million ( $\delta_{\text{C}}$ ).

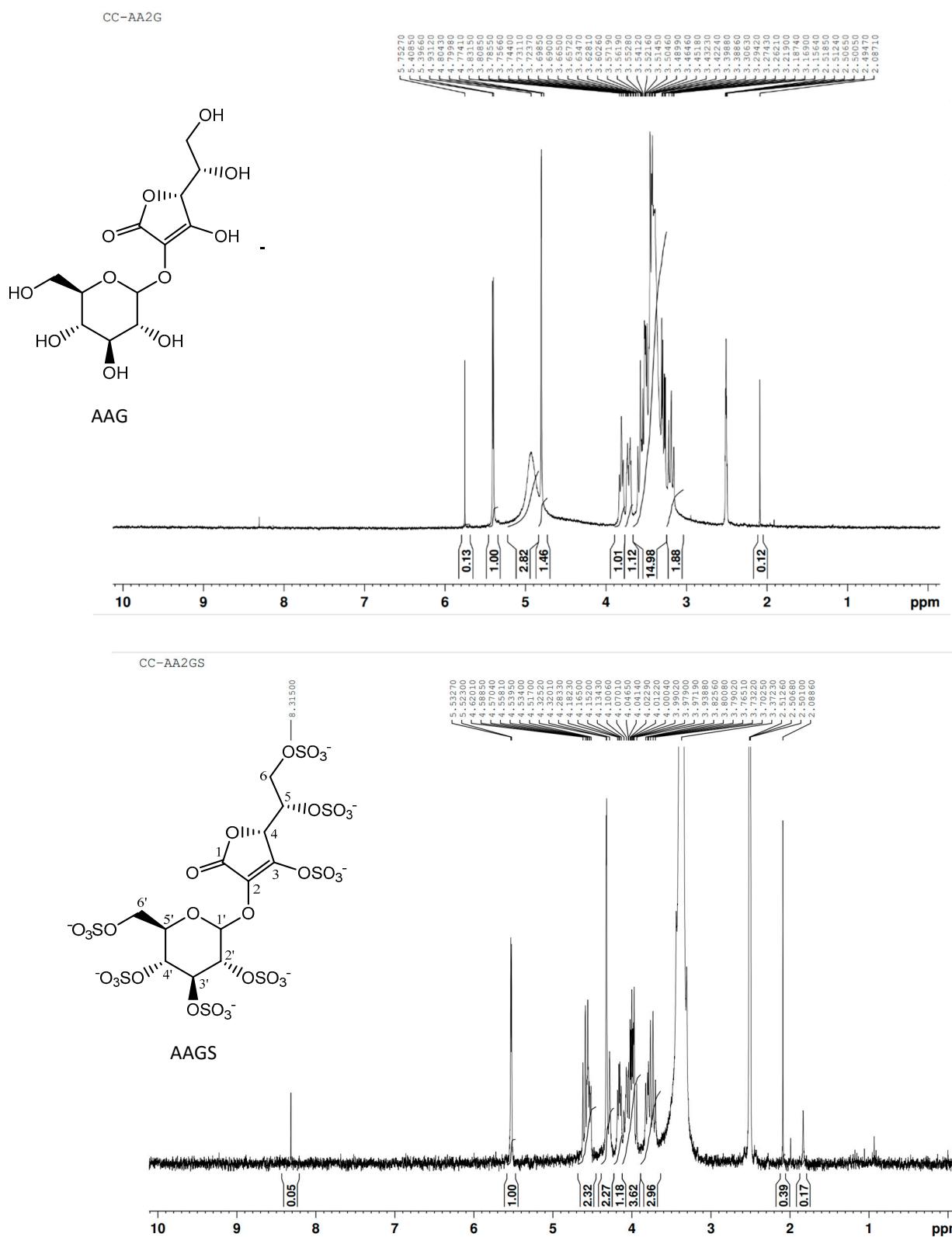
\*Literature assignments (1)



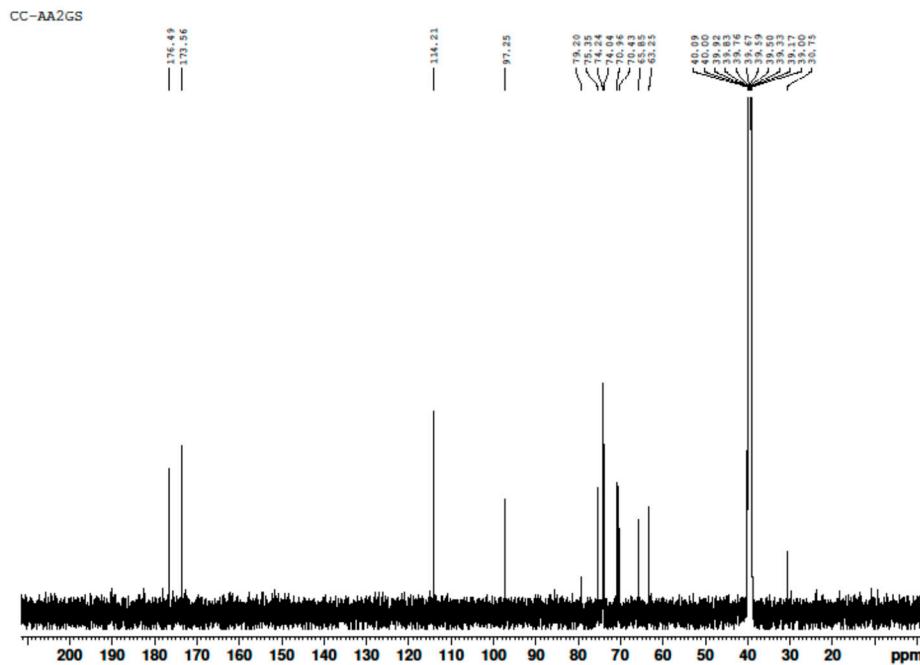
**Figure S1** Obtained chromatogram for AAGS by HPLC-DAD analysis (column C18; mobile phase: 25 mM of tetrabutylammonium and acetonitrile (38:62)).



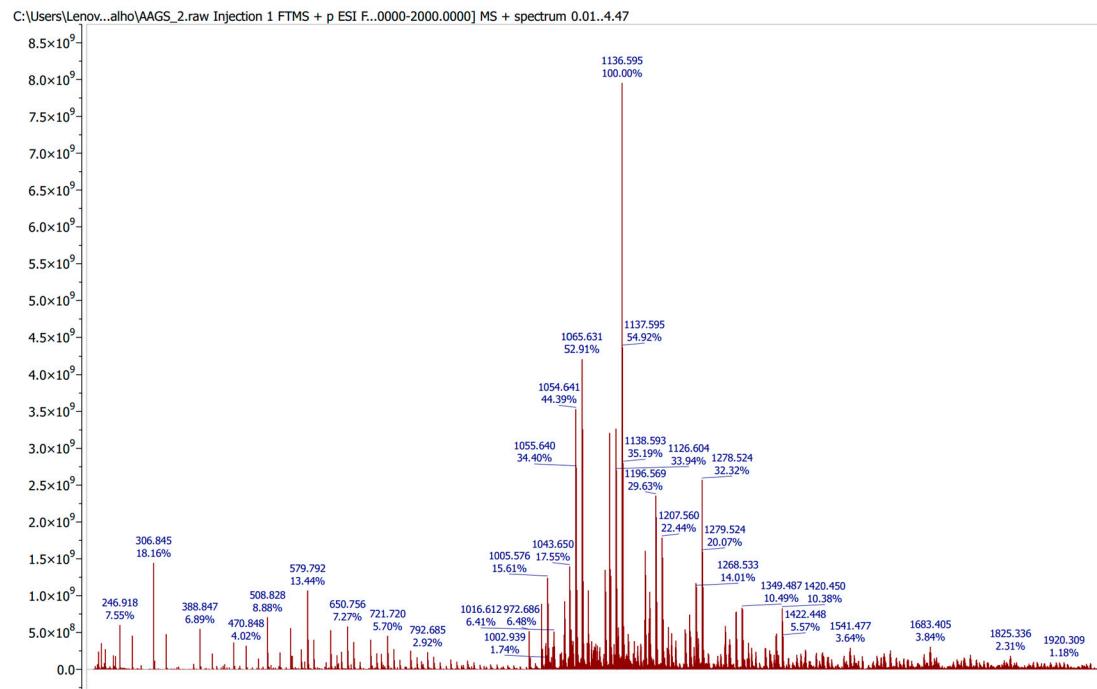
**Figure S2** IR (KBr) spectra: dark blue= 2-O- $\alpha$ -D-glucopyranosyl-L-ascorbic acid persulfate (AAGS), light blue= ascorbic acid 2-glucoside, and red= ascorbic acid.



**Figure S3**  $^1\text{H}$  NMR spectra of ascorbic acid 2-glucoside (AAG) and 2-O- $\alpha$ -D-glucopyranosyl-L-ascorbic acid persulfate (AAGS). \*Measured in DMSO- $d_6$  at 300.13 MHz.



**Figure S4** <sup>13</sup>C NMR spectrum of 2-O- $\alpha$ -D-glucopyranosyl-L-ascorbic acid persulfate (AAGS). \*Measured in DMSO-*d*<sub>6</sub> at 75.47 MHz.



**Figure S5.** ESI-HRMS of 2-O- $\alpha$ -D-glucopyranosyl-L-ascorbic acid persulfate (AAGS). **Experimental conditions.** Analysis was done on an Orbitrap Exploris 120 mass spectrometer (Thermo Fischer Scientific, Bremen, Germany) controlled by *Orbitrap Exploris Tune Application 2.0.185.35* and *Xcalibur 4.4.16.14*. The capillary voltage of the electrospray ionization source (ESI) was set to 3.4 kV and 2 kV for positive and negative mode. The capillary temperature was 320°C. The sheath gas and auxiliary gas flow rate were at 5 (arbitrary unit as provided by the software settings). The resolution of MS scan was 60 000. Data dependent MS/MS was performed on HCD using nitrogen as gas with collision energy settings of 30 V. The m/z range were 200–2000 Da. The resolution of SIM MS scan was 60,000. MS data handling software (Xcalibur QualBrowser software, Thermo Fischer Scientific) was used to search for predicted metabolites by their m/z value and MS/MS value.

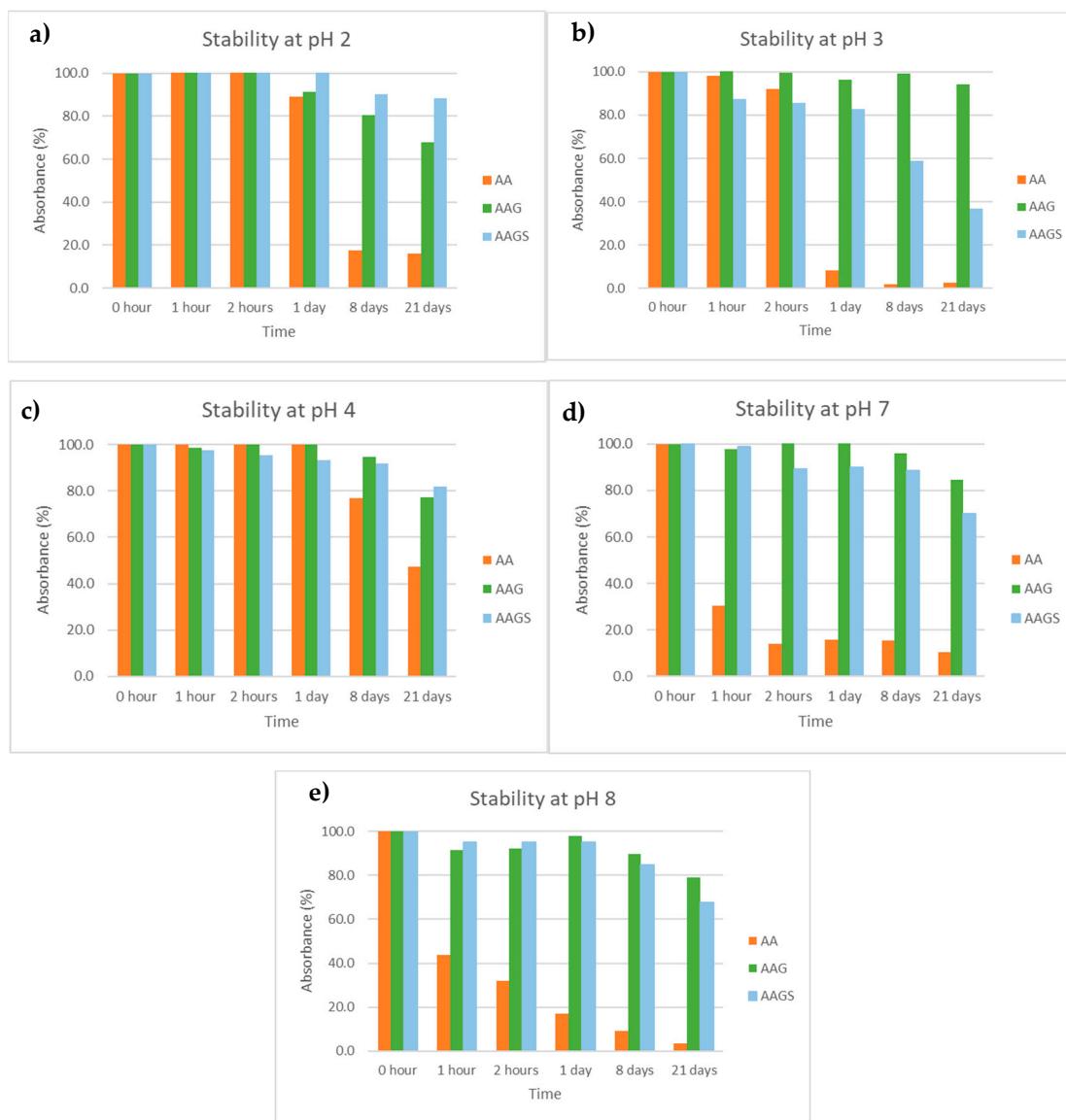
**Table S2** - DPPH scavenging activity of AA, AAG and AAGS given by EC<sub>50</sub> values (μM).

Compounds	EC <sub>50</sub> * ± SD (μM)
AA <sup>#</sup>	1.54 ± 0.21
AAG	9.15 ± 0.13
AAGS	5.28 ± 0.36

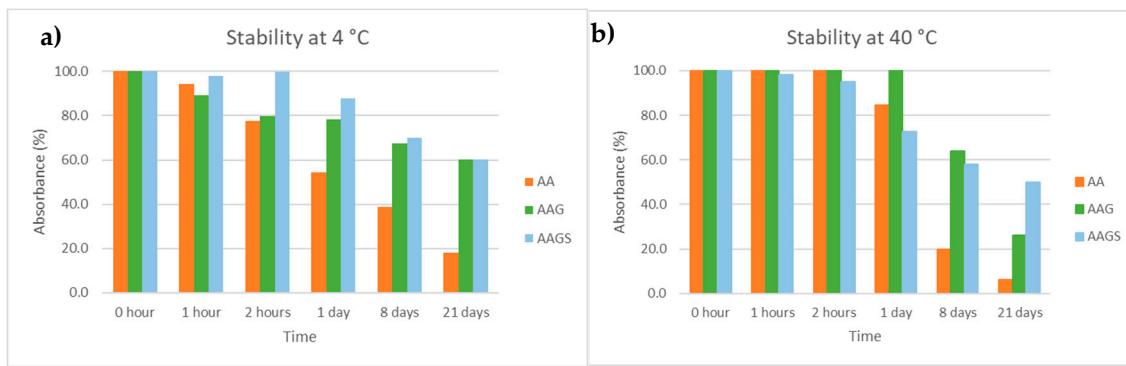
\* EC50: Concentration of a compound required to scavenge 50% of the initial DPPH radical.

SD: Standard deviation

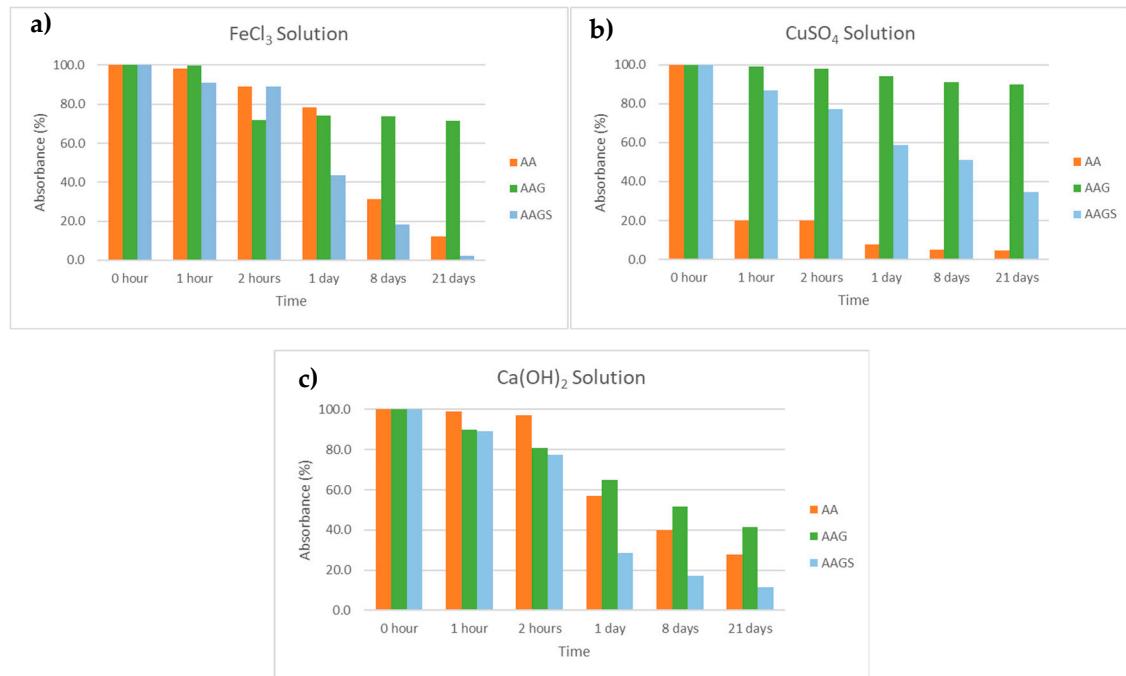
<sup>#</sup> EC<sub>50</sub> of AA in DPPH assay (literature value) = 16.3 ± 0.569 μM (2).



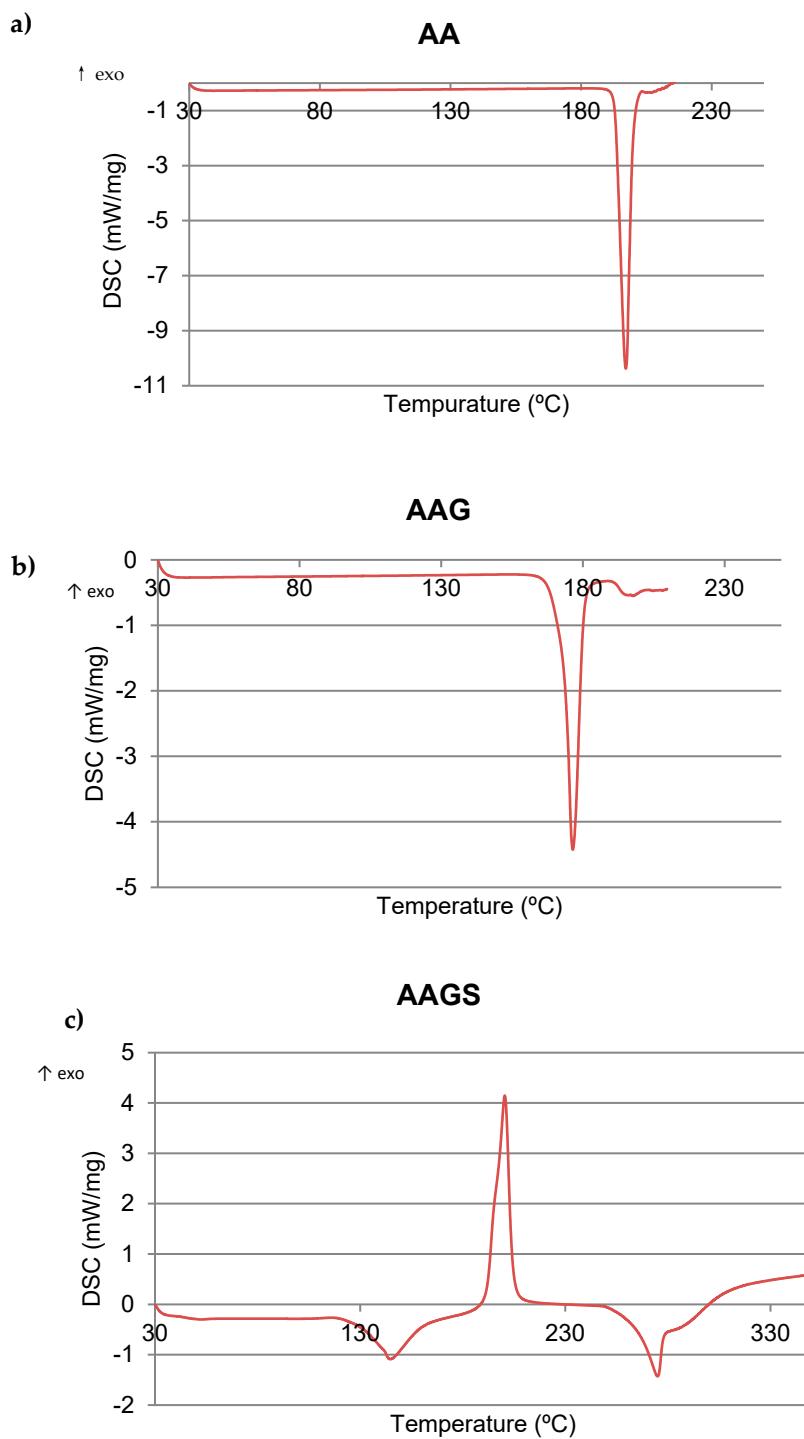
**Figure S6** Stability studies at different pH for compounds AA; AAG and AAGS. a) pH 2, b) pH 3, c) pH 4, d) pH 7 and e) pH 8. Results are expressed as % of absorbance normalized to the absorbance at the time t=0, in triplicate.

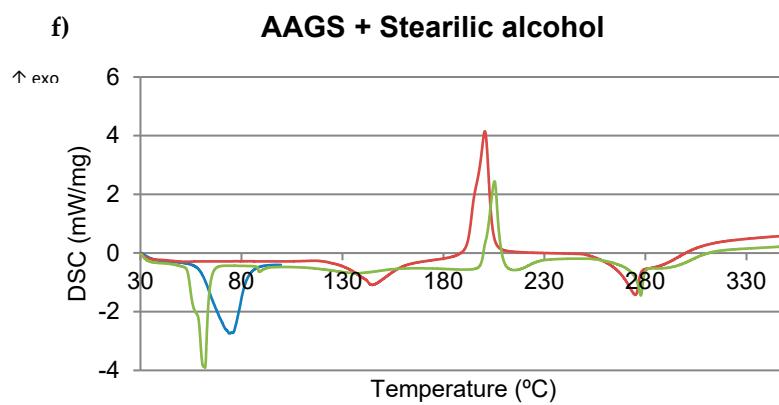
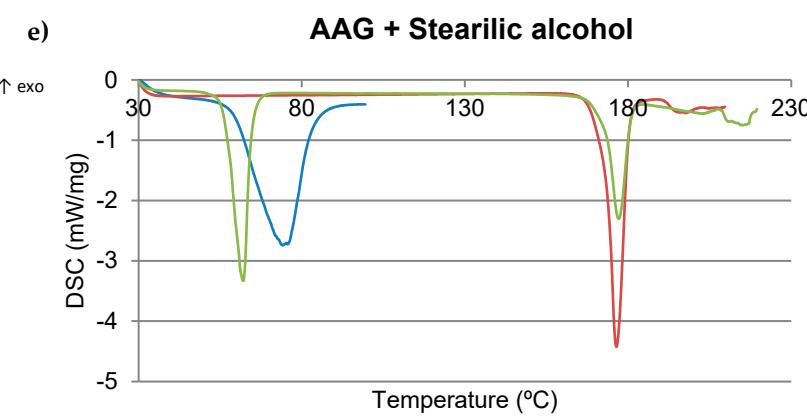
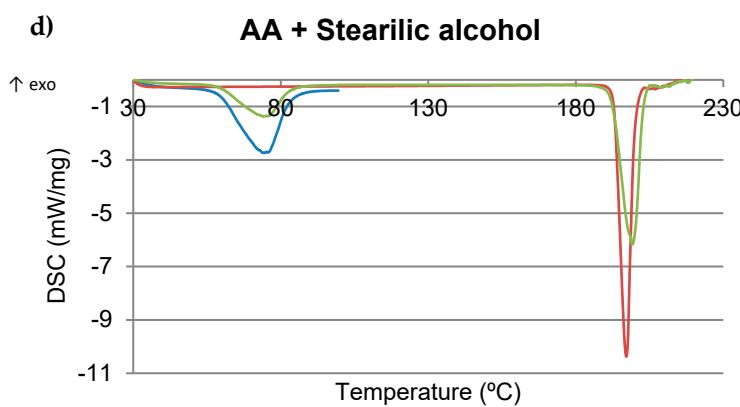


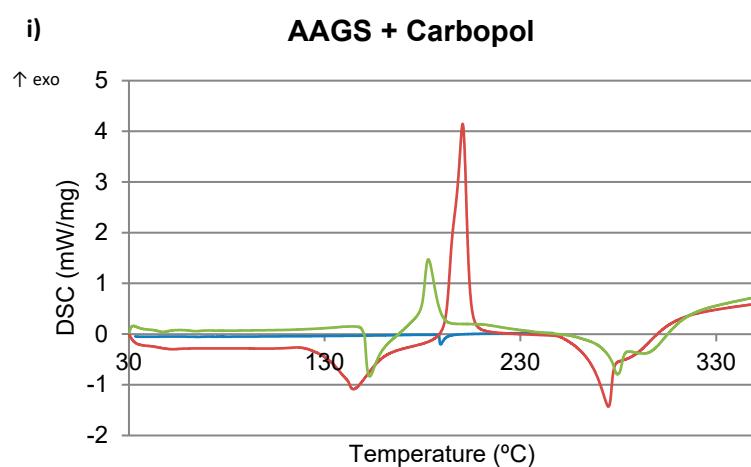
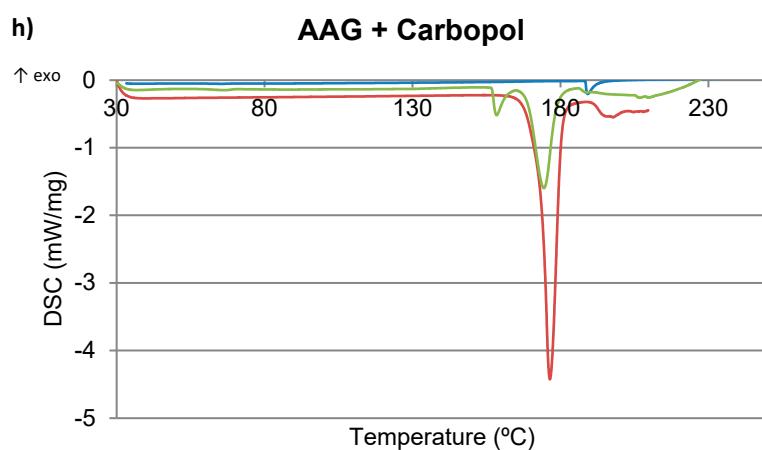
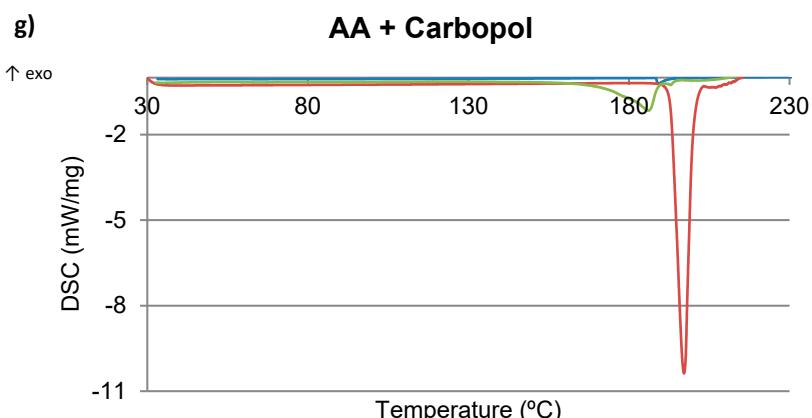
**Figure S7** Stability studies for compounds AA; AAG and AAGS at pH 5, at a) 4 °C, and b) 40 °C. Results are expressed as % of absorbance normalized to the absorbance at the time t=0, in triplicate.

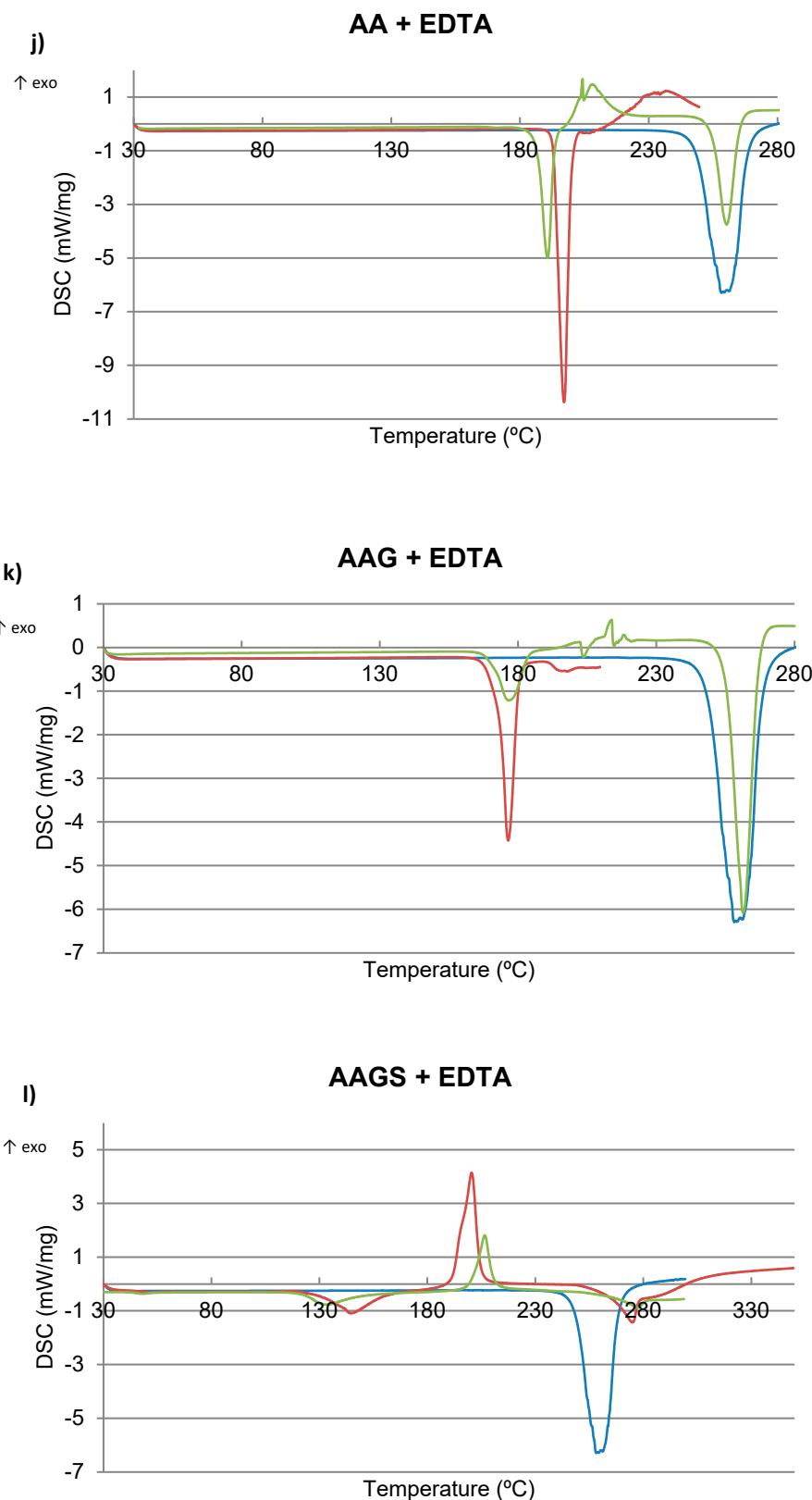


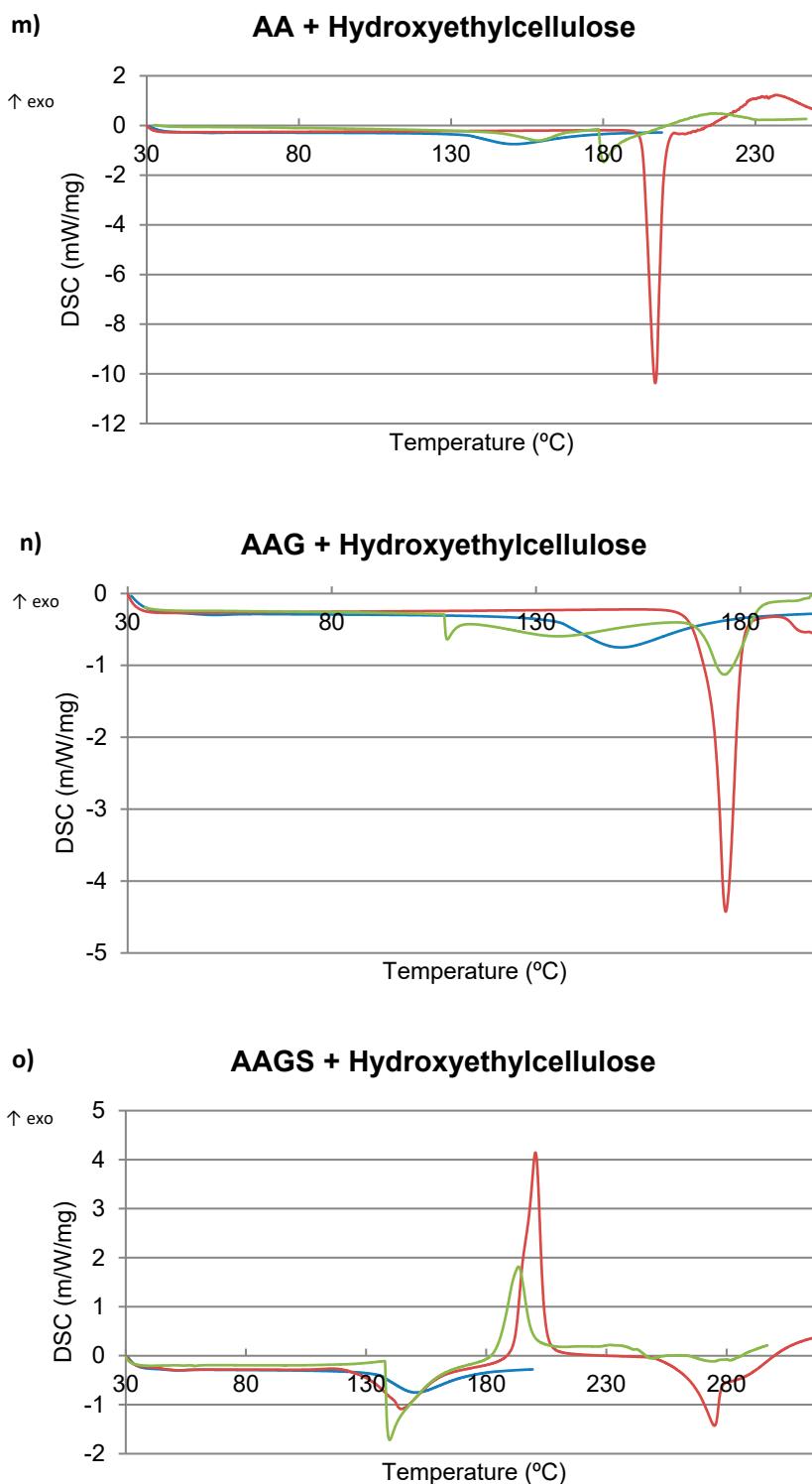
**Figure S8** Stability studies for compounds AA; AAG and AAGS at pH 5, for different metals, a) Fe(III), b) Cu(II) and c) Ca(II). Results are expressed as % of absorbance normalized to the absorbance at the time t=0, in triplicate.

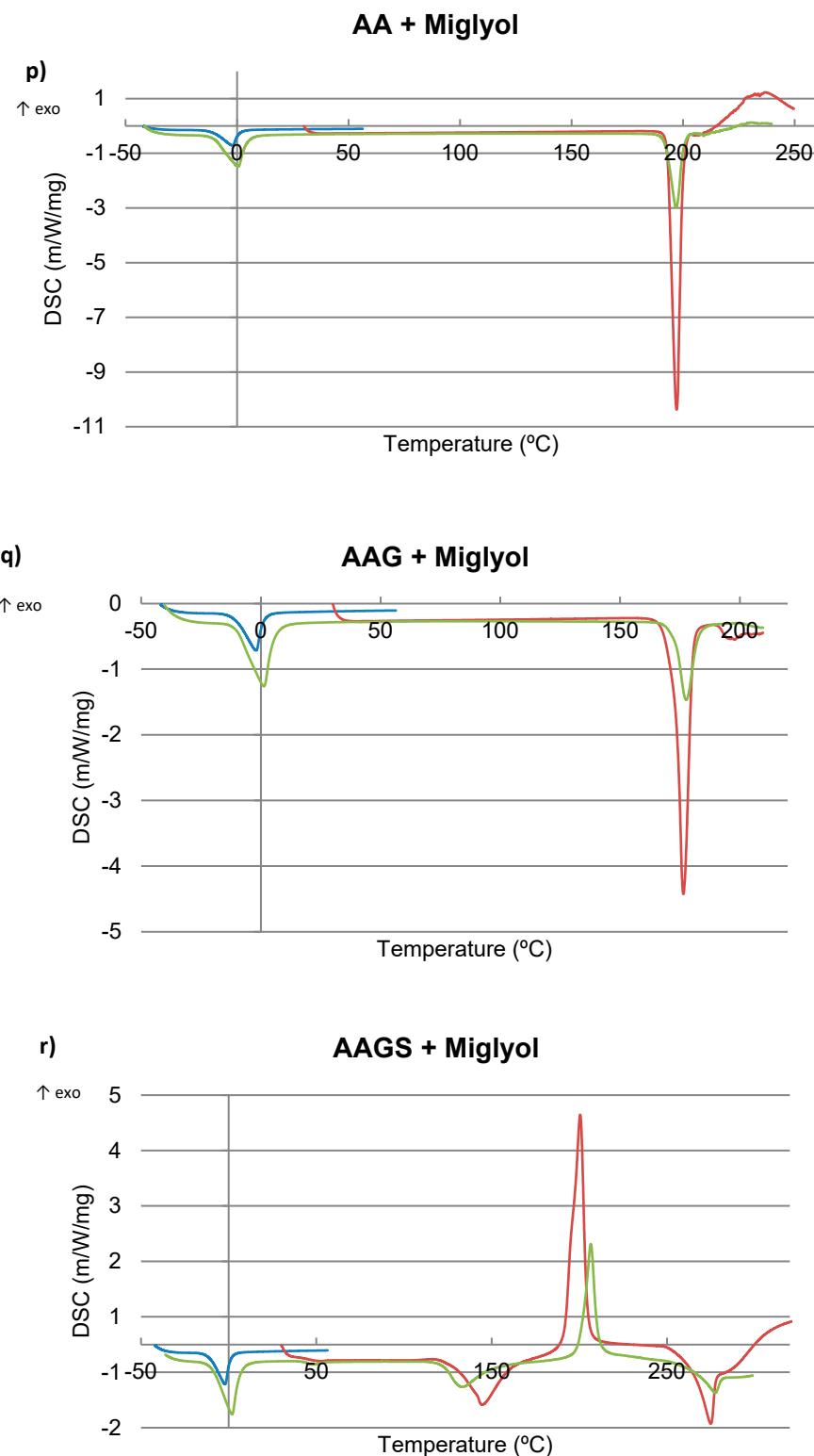


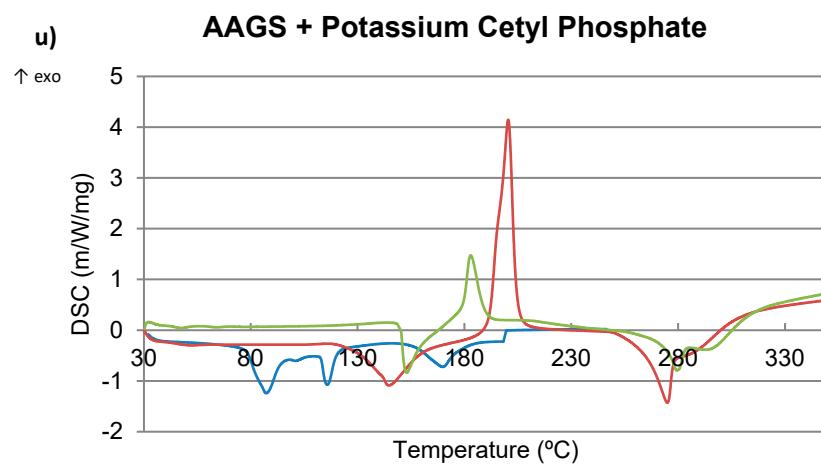
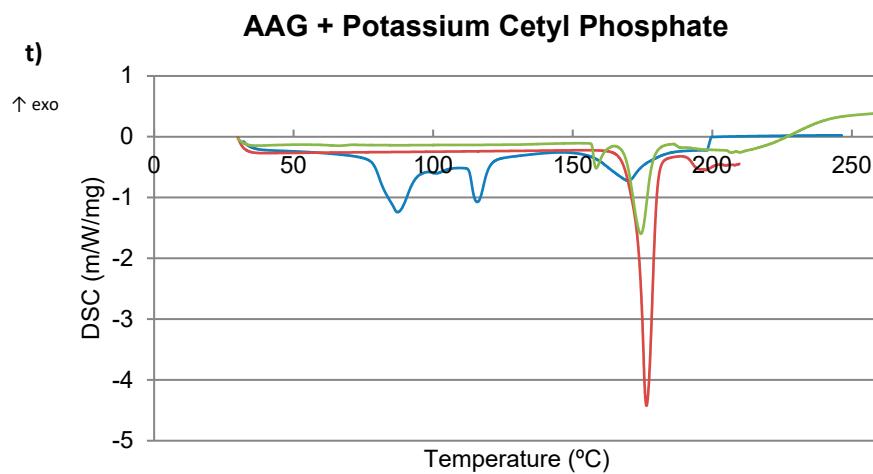
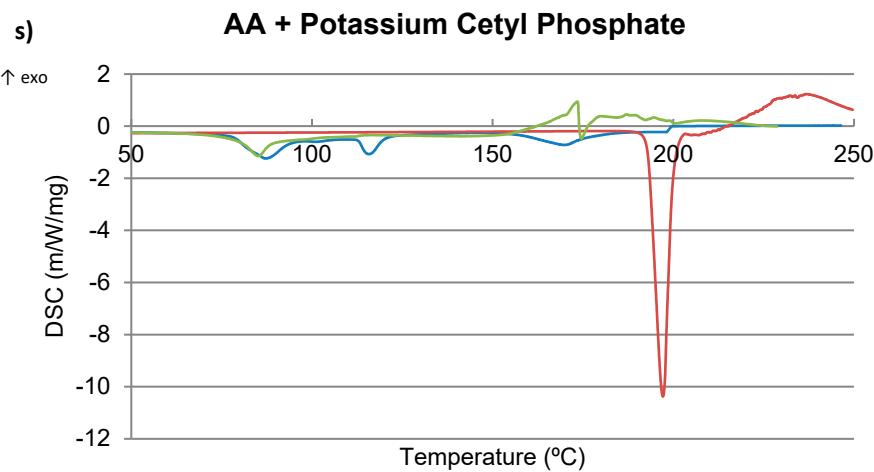


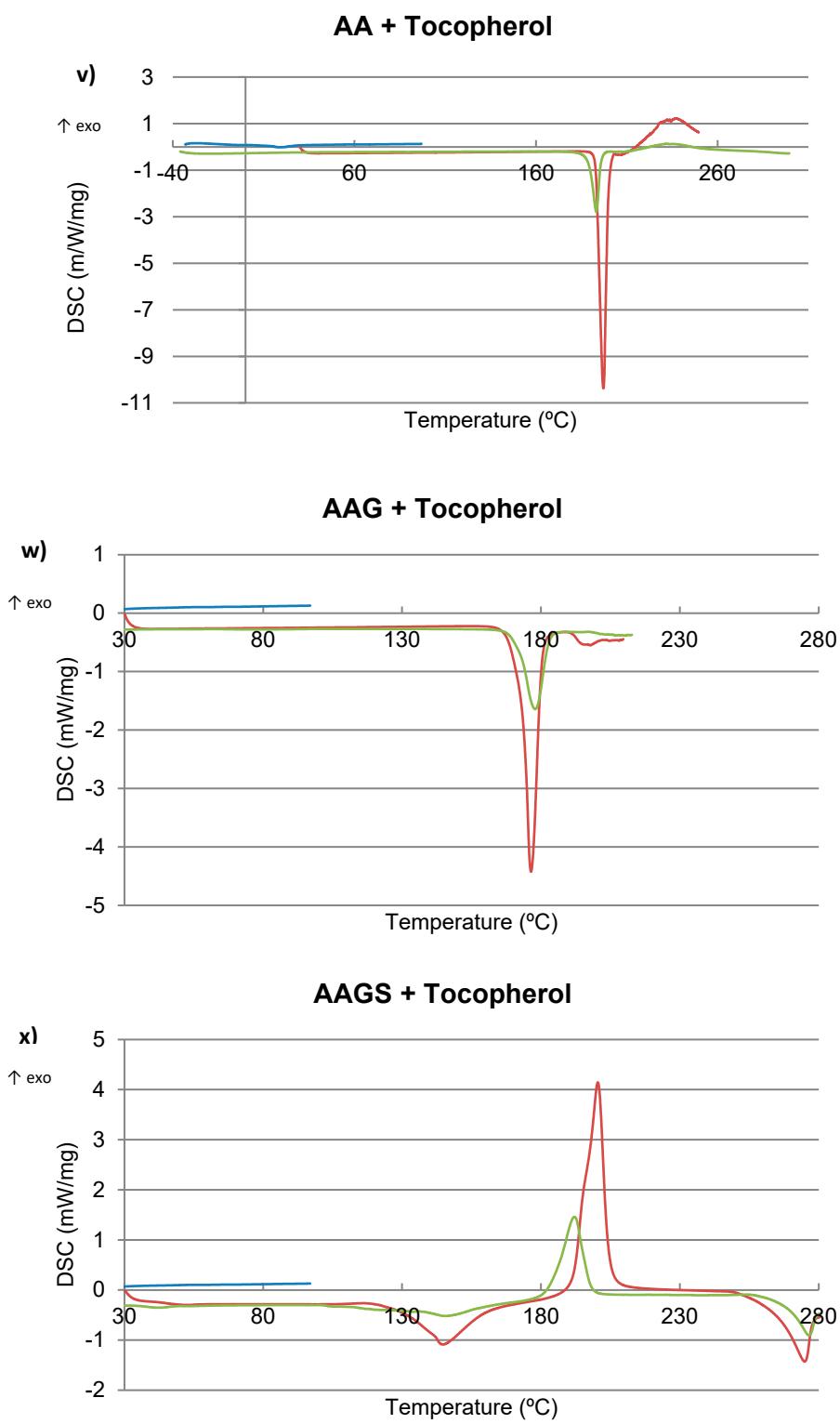












**Figure S9** Thermograms of individual compounds and the respective equivalent (1:1) mixtures for the compounds a) AA, b) AAG, c) AAGS, d) AA + stearilic acid, e) AAG + stearilic acid, f) AAGS + stearilic acid, g) AA + carbopol, h) AAG + carbopol, i) AAGS + carbopol, j) AA + EDTA, k) AAG + EDTA, l) AAGS + EDTA, m) AA + hydroxyethylcellulose, n) AAG + hydroxyethylcellulose, o) AAGS + hydroxyethylcellulose, p) AA + miglyol, q) AAG + miglyol, r) AAGS + miglyol, s) AA + potassium cetyl phosphate, t) AAG + potassium cetyl phosphate, u) AAGS + potassium cetyl phosphate, v) AA + tocopherol w) AAG + tocopherol x) AAGS + tocopherol. Red line: antioxidant (AA, AAG or AAGS), blue line (excipients), and green line: 1:1 (w/w) mixtures.

**References:**

1. Correia-da-Silva M, Sousa E, Duarte B, Marques F, Cunha-Ribeiro LM, Pinto MM. Dual anticoagulant/antiplatelet persulfated small molecules. *Eur J Med Chem.* 2011;46(6):2347-58.
2. Afsar T, Razak S, Shabbir M, Khan MR. Antioxidant activity of polyphenolic compounds isolated from ethyl-acetate fraction of *Acacia hydaspica* R. Parker. (1752-153X (Print)).