

Article **A Photoelectrochemical Sensor for the Detection of Hypochlorous Acid with a Phenothiazine-Based Photosensitizer**

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Abstract: This paper presents the development of a photoelectrochemical sensor for hypochlorous acid (HOCl) detection, employing a phenothiazine-based organic photosensitizer (Dye-PZ). The designed probe, Dye-PZ, follows a D-π-A structure with phenothiazine as the electron-donating group and a cyano-substituted pyridine unit as the electron-accepting group. A specific reaction of the phenothiazine sulfur atom with HOCl enables selective recognition. The covalent immobilization of Dye-PZ onto a titanium dioxide nanorod-coated fluorine-doped tin oxide electrode (FTO/TiO²) using bromo-silane coupling agent (BrPTMS) resulted in the fabrication of the photoanode FTO/TiO₂/BrPTMS/Dye-PZ. The photoanode exhibited a significant photoresponse under visible-light irradiation, with a subsequent reduction in photocurrent upon reaction with HOCl. The oxidation of the phenothiazine sulfur atom to a sulfoxide diminished the internal charge transfer (ICT) effect. Leveraging this principle, the successful photoelectrochemical sensing of HOCl was achieved. The sensor showed high stability, excellent reproducibility, and selective sensitivity for HOCl detection. Our study provides a novel approach for the development of efficient photoelectrochemical sensors based on organic photosensitizers, with promising applications in water quality monitoring and biosensing.

Keywords: photoelectrochemical sensor; organic photosensitizer; phenothiazine; hypochlorous acid

1. Introduction

Hypochlorous acid (HOCl) is a compound renowned for its robust oxidizing capabilities, commonly utilized in household bleach and disinfectants due to its exceptional antimicrobial and stain-removing properties. With oxygen atoms embedded in its molecular structure, HOCl can serve as a potent oxidizing agent and can rapidly react with both organic and inorganic substances. In household cleaning products, HOCl can release reactive oxygen atoms to facilitate the breakdown and removal of colors, making it an indispensable component for domestic and industrial cleaning. Beyond its applications in cleaning, HOCl's potent disinfecting properties find extensive applications in the medical, food, and water treatment sectors, effectively eliminating various microorganisms to safeguard public health and ensure food safety [\[1](#page-9-0)[–4\]](#page-9-1). At the biological level, HOCl assumes a crucial role, particularly as a representative of reactive oxygen species (ROS). ROS play pivotal roles in physiological and pathological processes, and HOCl, as one of these species, participates in multiple facets of the immune system, contributing significantly to maintaining normal physiological processes [\[5](#page-9-2)[–8\]](#page-9-3). The role of HOCl in immune responses involves pathogen clearance, inflammation promotion, and facilitation of tissue repair, serving as a vital defense mechanism against external threats.

However, excessive exposure to hypochlorous acid (HOCl) can pose severe health risks due to its oxidative and damaging effects on various biological components. The intricate mechanisms underlying HOCl-induced harm involve its reactivity towards crucial

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cellular structures, leading to widespread physiological disruptions [\[9,](#page-9-4)[10\]](#page-9-5). For instance, excess HOCl reacts with cell membrane lipids, causing lipid peroxidation and compromising the integrity of the cell membrane. This may result in increased membrane permeability, disruption of cellular homeostasis, and heightened susceptibility to external threats. Additionally, excess HOCl reacts with thiol-containing amino acids, inducing protein denaturation and functional loss. HOCl also induces DNA strand breaks, base modifications, and cross-linking, impacting nucleic acid integrity and jeopardizing the cell's ability to maintain genomic stability. Due to these oxidative reactions of HOCl, elevated levels may damage cells and tissues, potentially triggering various diseases such as arthritis, cardiovascular disorders, kidney ailments, neurodegenerative conditions, and cancer [\[11](#page-9-6)[–14\]](#page-9-7). Therefore, detecting the concentration of HOCl is crucial for monitoring its presence in the environment and gaining a deeper understanding of the physiological and pathological functions of HOCl.

Scientists employ a diverse array of methods to monitor and measure HOCl levels in environmental and biological samples, ensuring that it remains within a safe and beneficial range. These techniques include electroanalytical methods [\[15](#page-9-8)[–17\]](#page-9-9), chromatography [\[18](#page-9-10)[,19\]](#page-10-0), chemiluminescence [\[20](#page-10-1)[–22\]](#page-10-2), colorimetry [\[23–](#page-10-3)[25\]](#page-10-4), and fluorescence [\[26](#page-10-5)[–30\]](#page-10-6). Each of these methods is continually evolving for HOCl detection in both scientific research and practical applications. Notably, the focus lies on detection techniques utilizing organic small molecule probes. These molecules, with precise compositions and controllable structures, selectively respond to target analytes through specific functional groups, enhancing selectivity to enable rapid and accurate detection of HOCl in diverse environmental and biological systems. Advancements in detection methods based on the specific induction of oxidation reactions in various structural units by HOCl are ongoing. These structural units include carbon–carbon double bonds, sulfur compounds, aldoxime groups, thioether/Schiff bases, *N*,*N*-dimethylthioamidoformate, phenylboronic acid/borate esters, and others [\[31](#page-10-7)[–33\]](#page-10-8). Researchers have successfully developed a range of efficient fluorescence probe molecules for HOCl detection by leveraging the specific oxidation reactions in various structural units. Notably, phenothiazine stands out as a crucial structural unit for HOCl recognition. In phenothiazine derivatives, sulfur atoms can be oxidized to sulfoxide by HOCl, achieving a highly specific response to hypochlorous acid [\[34](#page-10-9)[–40\]](#page-10-10). Moreover, phenothiazine compounds exhibit unique biological activities and optoelectronic properties, making them versatile for applications spanning medicine [\[41](#page-10-11)[,42\]](#page-10-12), photovoltaic cells [\[43](#page-10-13)[–45\]](#page-10-14), fluorescence sensing [\[46](#page-11-0)[–49\]](#page-11-1), and electrochemical detection [\[50](#page-11-2)[–52\]](#page-11-3).

In this context, we innovatively employed the phenothiazine structural unit to construct an organic photosensitive small molecule, contributing to the development of a photoelectrochemical (PEC) sensor for detecting HOCl. PEC analysis, utilizing light excitation with electrochemical signals as its output, has gained widespread attention for its improved sensitivity through the utilization of two different signal modes [\[53](#page-11-4)[–58\]](#page-11-5). Previous research has also indicated that employing organic small-molecule photosensitizers to simultaneously achieve target recognition and photoelectric conversion is an effective strategy for constructing PEC sensors [\[59](#page-11-6)[–63\]](#page-11-7). Herein, we firstly synthesized a novel D-π-A structured photoactive molecule (Dye-PZ) by coupling aldehyde-functionalized phenothiazine with cyanopyridine. Subsequently, we covalently immobilized Dye-PZ onto a TiO₂ nanoarray substrate, obtaining a specific working PEC electrode for detecting HOCl. In Dye-PZ, the phenothiazine moiety serves as the electron-donating group, while the cyanopyridine unit acts as the electron-accepting group. This D - π -A structure exhibits excellent photoelectric conversion efficiency, generating a substantial photocurrent (Scheme [1\)](#page-2-0). Upon selective reaction of Dye-PZ with hypochlorous acid, the sulfur in the molecule is oxidized to sulfoxide, disrupting the D - π -A structure and resulting in a decrease in photocurrent response. Consequently, we achieved the highly specific detection of HOCl. Furthermore, this sensor boasts several advantages, such as a fast response, high signal-to-noise ratio, good selectivity, and high sensitivity.

noise ratio, good selectivity, and high sensitivity.

Scheme 1. The proposed PEC sensor for HOCl based on the phenothiazine-structured photoactive molecule Dye-PZ. molecule Dye-PZ.

2. Results and Discussion 2. Results and Discussion 2. Results and Discussion

2.1. Preparation and Characterization of the FTO/TiO₂/BrPTMS/Dye-PZ Photoanode

In the design of the photoelectrochemical probe molecule Dye-PZ, the crucial structural unit phenothiazine was incorporated. Phenothiazine serves as the electron-donating moiety, contributing to the construction of a D- π -A structured photoconversion functional molecule. Simultaneously, the sulfur atom in phenothiazine acts as a reactive site for the recognition of the target analyte, hypochlorous acid. The dual functionality of phenothiazine is effectively utilized in the design of Dye-PZ. The synthetic process involves two main steps: the aldehyde functionalization of phenothiazine, followed by condensation with cyano-4-pyridylacetonitrile to yield the target molecule, Dye-PZ. Further details concerning the specific synthetic methodology and characterization are provided in the subsequent experimental section. The results were confirmed through comprehensive characterization using nuclear magnetic resonance (NMR) and mass spectrometry (MS) methods. T[he](#page-2-1) photoanode was assembled through two steps (Figure 1). In the first step, the (3-bromopropyl)trimethoxysilane (BrPTMS) was coupled with the FTO/TiO₂ electrode, resulting in a bromoalkylsilane-modified electrode, FTO/TiO₂/BrPTMS. In the second step, the alkyl bromide on the modified electrode underwent a covalent reaction with the pyridine nitrogen in Dye-PZ, forming an alkylpyridinium and resulting in photoanode FTO/TiO₂/BrPTMS/Dye-PZ. To validate the successful assembly of the photoanode, a series of characterization approaches were applied to the electrodes under different modification states.

Figure 1. The fabrication process for the photoanode FTO/TiO₂/BrPTMS/Dye-PZ.

Firstly, we examined the feasibility of covalent coupling between the bromoalkoxysilane reagent (BrPTMS) and the photoactive molecule (Dye-PZ). As shown in Figure S1, the acetonitrile solution containing only Dye-PZ had an absorption band around 420 nm, while the solution after the reaction with BrPTMS showed absorption around 510 nm. The absorption peak position underwent a redshift, and, simultaneously, the solution color changed from yellow to red. This was attributed to the successful formation of a pyridinium cation through the interaction of Dye-PZ with BrPTMS. The introduction of this pyridinium salt moiety enhances the molecule's electron-accepting capacity, thereby significantly amplifying the intramolecular photo-induced charge transfer effect (ICT). Consequently, this phenomenon leads to a noticeable redshift in the absorption spectrum. These results indicate the ability of bromoalkoxysilane to form a covalent bond with the These results indicate the ability of bromoalkoxysilane to form a covalent bond with the photosensitive molecule. photosensitive molecule.

Then, we characterized the morphology of the photoanodes. SEM results revealed Then, we characterized the morphology of the photoanodes. SEM results revealed a a well-ordered TiO₂ rod array with a radial size of approximately 100 nm grown on the FTO surface (Figure S2). Subsequently, upon modification of the electrode surface with the photoactive molecule Dye-PZ, there was no significant change observed in the surface morphology of the FTO/TiO₂/BrPTMS/Dye-PZ electrode (Figure [2a](#page-3-0)). This can be attributed to the relatively smaller size of the organic small molecule, which does not substantially alter the morphology of TiO₂ upon modification. Next, we characterized the changes in the electrode preparation process. The Fourier-transform infrared spectroscopy changes in the electrode preparation process. The Fourier-transform infrared spectros-(FTIR) outcomes revealed a pronounced absorption band within the 750–650 cm⁻¹ range for the TiO₂ sample, corresponding to the vibration of Ti-O-Ti bonds (Figure [2b](#page-3-0), green curve). In the TiO₂/BrPTMS sample, a subtle absorption band emerged at 1049 cm^{−1}, signifying the stretching vibration absorption of the Si-O bond and confirming the interaction between the silane reagent and $TiO₂$. In the $TiO₂/BrPTMS/Dye-PZ$ sample, we observed the aforementioned Ti-O-Ti and Si-O bond absorptions, along with characteristic absorptions of the photoactive [mo](#page-3-0)lecule (Figure 2b, red curve). The absorption at 1630 cm⁻¹ is due to the stretching vibration of the C=N in the pyridinium group formed after the reaction between the bromoalkylsilane reagent and the pyridine moiety of the photoactive molecule. The absorption peak observed at 1560 cm⁻¹ corresponds to the stretching vibration of the C=C bond within the pyridine ring. The absorptions at 1460 cm^{−1} and 1340 cm^{−1} correspond to the benzene ring and the C-N stretching vibration in the phenothiazine structure of the photoactive molecule, respectively. These FTIR results indicated that the photoactive the photoactive molecule, respectively. These FTIR results indicated that the photoactive
molecule Dye-PZ was covalently coupled to the TiO₂ surface through the silane reagent. Subsequently, the synthesized samples underwent X-ray photoelectron spectroscopy (XPS) elemental analysis. [As](#page-3-0) depicted in Figure 2c, the TiO₂ sample exhibited peaks at O 1s 529 eV and Ti2p1/2 458 eV. Additionally, due to the organic titanium used in TiO₂ synthesis, a peak at C 1s 284 eV was observed. In the sample resulting from the reaction of FTO/TiO₂ with bromosilane, characteristic peaks of Br 3d at 70 eV and Si 2p at 102 eV were evident. Upon covalent coupling with Dye-PZ, new peaks emerged at 163 eV and 399 eV, corresponding to S 2p and N 1s, respectively. The appearance of these peaks signified the successful integration of sulfur and nitrogen components into the sample after coupling with $\it D$ ye-PZ. These observations further validate the successful synthesis and covalent coupling of the PEC anode $FTO/TiO₂/BrPTMS/Dye-PZ.$

Figure 2. (a) SEM image of FTO/TiO₂. (b) FTIR spectra of Dye-PZ, TiO₂, TiO₂/BrPTMS, and Tio 2/BrPTMS/Dye-Partners. (c) $\frac{1}{2}$ spectra of TiO2/BrPTMS/Dye-PZ spectra of TiO2/BrPTMS/Dye-PZ TiO₂/BrPTMS/Dye-PZ samples. (**c**) XPS spectra of TiO₂, TiO₂/BrPTMS, and TiO₂/BrPTMS/Dye-PZ samples.

Moreover, the electrochemical characterization of the photoanode preparation process was conducted. The electrochemical impedance spectroscopy (EIS) results are depicted in Figure S3a. After integrating $FTO/TiO₂$ with bromosilane, there was an increase in charge transfer resistance (R_{ct}) from 3.1 × 10³ Ω to 1.6 × 10⁵ Ω . This increase suggested the effective coverage of the electrode surface by bromosilane, hindering the charge transfer process. Subsequent covalent coupling with Dye-PZ led to a reduction in R_{ct} to 9.1 \times 10⁴ Ω . This reduction could be ascribed to the electrostatic attraction between the pyridinium Ω. This reduction could be ascribed to the electrostatic attraction between the pyridinium cations and the negatively charged Fe(CN) $_6^{4-}/3-$ redox probes in the electrolyte, leading to a diminished charge transfer resistance. The cyclic voltammetry (CV) results exhibited a similar trend, as shown in Figure S3b. These findings provide additional confirmation of the covalent coupling processes, both between $FTO/TiO₂$ and BrPTMS and between TiO2/BrPTMS and Dye-PZ. This affirmed the successful fabrication of the photoanode. TiO2/BrPTMS and Dye-PZ. This affirmed the successful fabrication of the photoanode.

2.2. PEC Response of FTO/TiO2/BrPTMS/Dye-PZ to HOCl 2.2. PEC Response of FTO/TiO2/BrPTMS/Dye-PZ to HOCl

To validate the feasibility of the prepared photoanode in responding to HOCl, we To validate the feasibility of the prepared photoanode in responding to HOCl, we initially tested the spectral response of the synthesized photoactive probe Dye-PZ to HOCl initially tested the spectral response of the synthesized photoactive probe Dye-PZ to HOCl in solution. HPLC-MS experiments were conducted using different solution systems in solution. HPLC-MS experiments were conducted using different solution systems to to verify the occurrence of the reaction. Subsequently, we employed a three-electrode system to assess the PEC response of the photoanode to HOCl. In a mixture of DMSO and phosphate buffer (*v*/*v*, 5/5, 5 mM, pH 7.4), the spectral response of Dye-PZ to HOCl was investigated. Dye-PZ (10 μ M) exhibited a blue shift in the UV absorption spectrum from 422 nm to 400 nm upon reacting with HOCl. Simultaneously, a significant enhancement in fluorescence emission at 470 nm was observed, and, under UV light, the solution's fluorescence emission at 470 nm was observed, and, under UV light, the solution's fluo-fluorescence changed from red to blue (Figure 3). [The](#page-4-0) spectral response of Dye-PZ to HOCl can likely be attributed to the oxidation of the sulfur in the phenothiazine moiety induced by HOCl, leading to the attenuation of the intra-molecular charge transfer (ICT) effect, resulting in spectral shifts. Subsequently, the reaction process was monitored using HPLC-MS (Figure S4). A chromatographic peak appeared at 5.14 min for Dye-PZ alone HPLC-MS (Figure S4). A chromatographic peak appeared at 5.14 min for Dye-PZ alone (Figure S4a), while after reacting with HOCl, two peaks emerged at 5.14 min and 5.83 min (Figure S4a), while after reacting with HOCl, two peaks emerged at 5.14 min and 5.83 min (Figure S4b), corresponding to Dye-PZ (*m*/*z* [M + H⁺] + : theoretical value 342.1, measured (Figure S4b), corresponding to Dye-PZ (*m/z* [M + H+]+: theoretical value 342.1, measured value m/z : 342.1) and the product $(m/z [M + H⁺]$ ⁺: theoretical value 358.1, measured value *m*/*z*: 358.1), respectively. These results indicated the oxidation of sulfur in Dye-PZ to sub sulfoxide after reacting with H OCl.

Figure 3. (**a**) Schematic representation of the reaction between Dye-PZ and HOCl. (**b**) UV–visible absorption and (**c**) fluorescence emission spectra of Dye-PZ (10 µM) solution and the reaction solution of Dye-PZ (10 µM) with HOCl (50 µM), with corresponding photographs under 365 nm UV light.

Following that, the PEC response of the photoanode to HOCl was tested. As depicted in Scheme [1,](#page-2-0) the phenothiazine moiety within the molecule functioned as the electrondonating group, while the pyridine unit served as the electron-accepting group, inducing the generation of the ICT effect and absorption in the visible region. Upon reacting with

HOCl, the sulfur in the molecule underwent oxidation to a sulfoxide, leading to the elimination of the ICT effect. The color of the photoanode $\text{FTO/TiO}_2/\text{BrPTMS}/\text{Dye-PZ}$ also changed from red to colorless (Figure 4 in[set](#page-5-0)). Photocurrent tests demonstrated stable photocurrent responses under 500 nm excitation light. The process of photocurrent generation begins with the excitation of Dye-PZ under illumination, where electrons transition from the HOMO state to the LUMO excited state. Subsequently, the injected electrons from the LUMO into the covalently coupled TiO_2 conduction band are collected by the electrode, generating a photocurrent signal. The oxidized state of Dye-PZ, after losing electrons, is then reduced to its initial state by an electron-donating species in the solution, such as ascorbic acid, thereby continuing the photocurrent generation. However, upon reacting with HOCl, the molecular structure of the D-π-A system is disrupted, preventing the effective generation of photocurrent. This phenomenon can be observed as a significant decrease in photocurrent in Figure [4.](#page-5-0) The results confirm the effectiveness of the photoelectrode as a PEC sensing platform for HOCl.

Figure 4. Photocurrent response and color change of the photoanode FTO/TiO₂/BrPTMS/Dye-PZ HOCl. to HOCl.

2.3. Analytical Performance 2.3. Analytical Performance

During PEC testing, factors such as bios voltage, the excitation light, and the electron During PEC testing, factors such as bios voltage, the excitation light, and the electron donor can influence the photocurrent response. Therefore, we explored and optimized the donor can influence the photocurrent response. Therefore, we explored and optimized the conditions of this method to obtain the best experimental results. Firstly, we investigated conditions of this method to obtain the best experimental results. Firstly, we investigated the impact of bias voltage (−0.6 V to −0.2 V) and excitation wavelength (440–600 nm) on the impact of bias voltage (−0.6 V to −0.2 V) and excitation wavelength (440–600 nm) on the PEC response. As shown in Figure S5a,b, the photoanode exhibited a stable response the PEC response. As shown in Figure S5a,b, the photoanode exhibited a stable response and significant current changes at -0.4 V (vs. Hg/Hg₂SO₄) and 500 nm excitation wavelength. Additionally, three electrolytes containing ascorbic acid (AA), sodium oxalate, and length. Additionally, three electrolytes containing ascorbic acid (AA), sodium oxalate, and triethanolamine (TEOA) as electron donors were chosen for comparative testing to explore triethanolamine (TEOA) as electron donors were chosen for comparative testing to explore the influence of different electron donors on PEC testing. The results in Figure S5c indicate the influence of different electron donors on PEC testing. The results in Figure S5c indicate that the photocurrent response showed the largest relative reduction when ascorbic acid that the photocurrent response showed the largest relative reduction when ascorbic acid was used as the electron donor. Therefore, we selected a −0.4 V bias voltage, a 500 nm was used as the electron donor. Therefore, we selected a −0.4 V bias voltage, a 500 nm excitation light, and an electrolyte-containing ascorbic acid for subsequent testing. Under excitation light, and an electrolyte-containing ascorbic acid for subsequent testing. Under the optimal conditions, the photoanode was tested for its photocurrent response to different concentrations of HOCl (Figure [5a](#page-6-0)). We observed that within the concentration range of 5.0–40.0 µM HOCl, the photocurrent exhibited a decrease as the concentration increased, showing a robust linear correlation with the logarithm of HOCl concentration, as illustrated in Figure [5b](#page-6-0), represented by the fitting equation I = $-0.348 \times \text{lg[HOCl]} + 0.861 \text{ (R}^2 = 0.981)$. The detection limit was established as $2.58 \mu M (3\sigma/k)$.

Figure 5. (a) The photocurrent response of FTO/TiO₂/BrPTMS/Dye-PZ to varying concentrations of HOCl (0, 5, 10, 15, 20, 30, and 40 μ M). (b) The fitted curve depicting the linear relationship between photocurrent values and the logarithm of HOCl concentration. photocurrent values and the logarithm of HOCl concentration. photocurrent values and the logarithm of HOCl concentration.

Next, the selectivity and anti-interference performance of the PEC sensor were investigated. Interfering ions included common ions such as Al^{3+} , Fe²⁺, Cu²⁺, Zn²⁺, Mg²⁺, Mn²⁺, $NO₃⁻$, Cl⁻, BrO₃⁻, SO₄²⁻, H₂O₂, C₂H₅OH, and alanine (Ala). None of the mentioned interfering substances led to a substantial reduction in the PEC sensor's photocurrent. Although there are studies reporting that thiol-containing compounds (such as dimethyl sulfide, DMS) can be oxidized to sulfoxides by hydrogen peroxide [\[64,](#page-11-8)[65\]](#page-11-9), in the case of this Dye-PZ probe, the sulfur (S) in phenothiazine is linked to two phenyl rings, with its electron cloud distributed in a larger conjugated system. Therefore, the reactivity of sulfur in phenothiazine is expected to be lower than that of alkyl thioethers, making it resistant to oxidation by hydrogen peroxide and selectively oxidizable by the stronger oxidizing agent, hypochlorous acid, thus achieving the interference-free detection of hydrogen peroxide. Phenothiazine is also widely utilized as a high-specificity recognition moiety in constructing probes for hypochlorous acid, and these probe molecules have also demon-strated interference-free characteristics against hydrogen peroxide [\[66](#page-11-10)[–70\]](#page-11-11). Furthermore, competitive experiments revealed that the presence of other interferences did not impact
 \mathcal{L} the detection of HOCl (Figure [6a](#page-6-1)). The selected competitive results indicated that the PEC sensor exhibited outstanding anti-interference capability. Finally, the stability of the sensor
expansion that testing its continuous PEC substances of final the redes of testing was examined by testing its continuous PEC pulse response. After eight cycles of testing, the stating, the stating, the photocurrent remained highly stable, with a relative standard deviation (RSD) of less than 1.1% (Figure [6b](#page-6-1)), demonstrating the sensor's high photostability. $\frac{1}{100}$ (Figure $\frac{1}{10}$), demonstrating the sensor $\frac{1}{100}$ photostability.

Figure 6. (**a**) The black column represents the photocurrent signal of the PEC sensor incubated with an interfering substance (40 μ M), and the red column represents the photocurrent signal of the sensor incubated with both the interfering substance (40 µM) and HOCl (40 µM). (**b**) Stability of the sensor under a continuous cycle of periodic "off-on-off" light.

3. Materials and Methods

3.1. Materials and Apparatus

The materials for synthesizing the photoactive molecule Dye-PZ, including 10-methyl-10H-phenothiazine, phosphorus oxychloride, cyanopyridine, and piperidine, were purchased from Shanghai Titan Technology Co., Ltd. (Shanghai, China). Titanium butoxide and acetonitrile were procured from Tianjin Chemreagent Company (Tianjin, China). Solvents used in synthesis, such as dichloromethane and ethyl acetate, were obtained from Tianjin Damao Chemical Reagent Factory. Analytical-grade small molecules and salts were sourced from China National Pharmaceutical Group Corporation. All reagents were of analytical purity. Deionized water used for preparing testing solutions was purified through a secondary deionization process.

UV–visible spectrophotometry signals were recorded using a Lambda-850 instrument from PerkinElmer Inc. (Waltham, MA, USA). Fluorescence measurements were conducted with an RF-5301PC fluorometer from Shimadzu Corporation (Kyoto, Japan). Mass spectrometry analysis employed a Q-Tof Premier mass spectrometer from Waters Corporation (Milford, MA, USA). Nuclear magnetic resonance (NMR) spectra were acquired using a Bruker ARX-400 spectrometer (Bruker Limited, Fällanden, Switzerland). Infrared spectra were recorded with the Bruker Vector 22 Fourier-transform infrared spectrometer (Bruker Limited, Switzerland). X-ray photoelectron spectroscopy (XPS) measurements were conducted using a Thermo Scientific K-Alpha instrument (Thermo Fisher Scientific Inc., Waltham, MA, USA). Photoelectrochemical tests were carried out using a DY200B electrochemical workstation from Digi-Ivy, Inc. (Austin, TX, USA).

3.2. Synthesis of Dye-PZ

The synthetic route for Dye-PZ is depicted in Figure S6. Under an ice bath $(0 \degree C)$ and a nitrogen atmosphere, dimethylformamide (0.17 mL, 2.4 mmol) was combined with phosphorus oxychloride (0.2 mL, 2.4 mmol), followed by stirring for 15 min. Subsequently, compound **1** (426 mg, 2 mmol in 2 mL anhydrous DMF) was introduced into the mixture. The reaction mixture was heated to 60 \degree C and stirred for 4 h. After completing the reaction, the mixture was transferred into 100 mL of ice water and neutralized with a 10% NaHCO₃ solution. The resulting viscous material was subjected to three extractions with 100 mL of dichloromethane. The organic layer was separated, washed sequentially with saturated sodium chloride solution and water, dried over anhydrous $Na₂SO₄$, and the solvent was evaporated under reduced pressure. The residue was further purified through silica gel column chromatography, using dichloromethane as the eluent. This process resulted in the isolation of purified compound **2** (0.322 g, 67% yield). Compound **2** (241 mg, 1.0 mmol), cyanopyridine (118 mg, 1.0 mmol), and 4 mL anhydrous ethanol were combined and refluxed for 5 h. After the completion of the reaction, the solvent was removed under reduced pressure, and further purification was carried out using silica gel column chromatography. A mixture of dichloromethane and ethyl acetate (1:1, *v*/*v*) was used as the eluent for purification. The resulting product, Dye-PZ, was obtained as a deep red solid (0.143 g, 43% yield). Characterization of the synthesized photosensitizer Dye-PZ was conducted using several techniques, including nuclear magnetic resonance (NMR) spectroscopy and mass spectrometry. ¹HNMR (400 MHz, DMSO) δ 8.67 (d, J = 6.1 Hz, 2H), 8.19 (s, 1H), 7.91 (dd, J = 8.6, 1.9 Hz, 1H), 7.81 (d, J = 1.9 Hz, 1H), 7.69 (d, J = 6.1 Hz, 2H), 7.26 (t, J = 7.7 Hz, 1H), 7.20 (d, J = 7.4 Hz, 1H), 7.13 (d, J = 8.7 Hz, 1H), 7.03 (t, J = 7.3 Hz, 2H), 3.39 (s, 3H) (Figure S7). ¹³CNMR (101 MHz, DMSO) δ 150.85, 148.41, 144.87, 144.29, 141.84, 130.84, 128.55, 128.09, 127.66, 127.40, 123.85, 122.68, 121.48, 119.95, 117.96, 115.71, 115.18, 104.96, 35.91 (Figure S8). MS [ESI]: *m*/*z*, theoretical molecular weight [M⁺] ⁺ 341.0981; mass spectrum data 341.0983 (Figure S9).

3.3. Preparation of FTO/TiO2/BrPTMS/Dye-PZ

The FTO/TiO² nanorod-based electrode was prepared according to a reported method [\[60\]](#page-11-12). The assembly of the photoanode is illustrated in Figure [1,](#page-2-1) and the spe-

cific assembly steps were as follows: Firstly, 3-(bromopropyl)trimethoxysilane (150 μ L) and isopropanol (10 mL) were added to a beaker. After thorough mixing, the prepared FTO/TiO₂ was placed into the beaker and the reaction mixture was heated at 70 °C for 30 min. Subsequently, the electrode was removed and washed with isopropanol, and dried with nitrogen gas. Then, the electrode was placed in a vacuum drying oven at 60 \degree C to obtain the bromosilane-modified electrode $FTO/TiO₂/BrPTMS$. Next, $FTO/TiO₂/BrPTMS$ was immersed in a solution containing Dye-PZ (1 mM) in acetonitrile (20 mL) at 80 ◦C for 5 h oil bath reflux. Finally, the electrode was thoroughly rinsed with dimethyl sulfoxide to remove non-covalently bound dye, resulting in the desired working electrode FTO/TiO2/BrPTMS/Dye-PZ.

3.4. Characterization and Photoelectrochemical Testing of FTO/TiO2/BrPTMS/Dye-PZ

The photoanode and its preparation process were characterized using a three-electrode system with $K_3Fe(CN)_6$ (10 mM) and KCl (0.1 M) solution as the electrolyte. The three electrodes consisted of differently modified FTO as the working electrode (WE), Ag/AgCl (saturated KCl at 25 $^{\circ}$ C) as the reference electrode (RE), and platinum wire as the counter electrode (CE). The photoanode was subjected to electrochemical impedance spectroscopy (EIS) and cyclic voltammetry (CV) using a CHI 660E electrochemical analyzer. Infrared spectroscopy (IR) was employed for functional group analysis, and X-ray photoelectron spectroscopy (XPS) was used for elemental analysis of the photoanode surface.

The photoanode functioned as the working electrode, accompanied by a platinum wire as the counter electrode and Hg/Hg_2SO_4 (saturated K_2SO_4 at 25 °C) as the reference electrode. Incubation took place in a phosphate buffer solution (5 mM, pH 7.4) containing a specific concentration of HOCl. The electrolyte utilized was a phosphate buffer solution (0.1 M, pH 7.4) comprising 0.1 M sodium sulfate and 0.1 M ascorbic acid (AA). The photoelectrochemical (PEC) characteristics were monitored using an electrochemical workstation and a fluorescence spectrophotometer (model: RF-5301PC, Shimadzu, Japan) equipped with a xenon lamp for excitation.

4. Conclusions

A PEC sensor for HOCl detection was successfully developed using a phenothiazinebased photoactive probe. The designed probe, Dye-PZ, featuring a D - π -A structure with phenothiazine as the electron-donating group and a cyano-pyridine unit as the electronwithdrawing group, demonstrated the specific reactivity with HOCl. Covalent attachment of Dye-PZ onto a titanium dioxide nanorod substrate using BrPTMS yielded the photoanode FTO/TiO₂/BrPTMS/Dye-PZ. The photoanode exhibited a pronounced PEC response under visible light, and, upon reaction with HOCl, the oxidation of the sulfur atom in the phenothiazine resulted in a weakened internal charge transfer effect and a subsequent decrease in photocurrent. Exploiting this mechanism, the PEC sensor demonstrated effective detection of HOCl. The sensor displayed high stability and excellent reproducibility, allowing for the selective and sensitive detection of HOCl.

Supplementary Materials: The following supporting information can be downloaded at: [https:](https://www.mdpi.com/article/10.3390/molecules29030614/s1) [//www.mdpi.com/article/10.3390/molecules29030614/s1,](https://www.mdpi.com/article/10.3390/molecules29030614/s1) Figure S1: Absorption spectra of Dye-PZ and Dye-PZ with BrPTMS in acetonitrile; Figure S2: SEM images of TiO₂ nanorod grown on FTO substrate before (a,b), SEM images of photoanode FTO/TiO₂/BrPTMS/Dye-PZ (c,d); Figure S3: Nyquist plots (**a**) and cyclic voltammetry (**b**) of photoanodes with different modifications in a phosphate buffer solution (10 mM, pH 7.4) containing 0.1 M $\rm Na_2SO_4$ and 1 mM $\rm K_3[Fe(CN)_6].$ The reference electrode used was Ag/AgCl; Figure S4: HPLC-MS analysis of the reaction process between Dye-PZ and HOCl. (**a**) Results for Dye-PZ alone. (**b**) Results for the reaction between Dye-PZ and HOCl; Figure S5: (**a**) Photocurrent response of the photoanode at voltages ranging from −0.6 V to −0.2 V and its response after reacting with HOCl; (**b**) photocurrent response of the photoanode before and after reacting with HOCl under different excitation lights from 440 nm to 600 nm; (**c**) photocurrent response of the photoanode before and after reacting with HOCl in solutions with three different electron donors; Figure S6: Synthetic route to Dye-PZ; Figure S7: ¹H NMR spectrum

of Dye-PZ in DMSO-*d⁶* ; Figure S8: ¹³C NMR spectrum of Dye-PZ in DMSO-*d⁶* ; Figure S9: Mass spectrum of Dye-PZ.

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References

- 1. Block, M.S.; Rowan, B.G. Hypochlorous Acid: A Review. *J. Oral Maxillofac. Surg.* **2020**, *78*, 1461–1466. [\[CrossRef\]](https://doi.org/10.1016/j.joms.2020.06.029) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/32653307)
- 2. Boyce, J.M. Modern technologies for improving cleaning and disinfection of environmental surfaces in hospitals. *Antimicrob. Resist. Infect. Control* **2016**, *5*, 10. [\[CrossRef\]](https://doi.org/10.1186/s13756-016-0111-x) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/27069623)
- 3. Robins, L.I.; Clark, A.; Gafken, P.R.; Alam, S.; Milici, J.; Hassan, R.; Wang, C.Y.; Williams, J.; Meyers, C. Hypochlorous acid as a disinfectant for high-risk HPV: Insight into the mechanism of action. *J. Med. Virol.* **2022**, *94*, 3386–3393. [\[CrossRef\]](https://doi.org/10.1002/jmv.27716) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/35277984)
- 4. Stubbs, A.D.; Lao, M.; Wang, C.; Abbatt, J.P.D.; Hoffnagle, J.; VandenBoer, T.C.; Kahan, T.F. Near-source hypochlorous acid emissions from indoor bleach cleaning. *Environ. Sci.-Process. Impacts* **2023**, *25*, 56–65. [\[CrossRef\]](https://doi.org/10.1039/D2EM00405D)
- 5. Duclos, F.; Abell, L.M.; Harden, D.G.; Pike, K.; Nowak, K.; Locke, G.A.; Duke, G.J.; Liu, X.Q.; Fernando, G.; Shaw, S.A.; et al. Triazolopyrimidines identified as reversible myeloperoxidase inhibitors. *Medchemcomm* **2017**, *8*, 2093–2099. [\[CrossRef\]](https://doi.org/10.1039/C7MD00268H)
- 6. Flemmig, J.; Remmler, J.; Zschaler, J.; Arnhold, J. Detection of the halogenating activity of heme peroxidases in leukocytes by aminophenyl fluorescein. *Free Radic. Res.* **2015**, *49*, 768–776. [\[CrossRef\]](https://doi.org/10.3109/10715762.2014.999676)
- 7. Liu, Q.C.; Liu, C.; He, S.; Zhao, L.C.; Zeng, X.S.; Zhou, J.; Gong, J. A New Phenylazo-Based Fluorescent Probe for Sensitive Detection of Hypochlorous Acid in Aqueous Solution. *Molecules* **2022**, *27*, 2978. [\[CrossRef\]](https://doi.org/10.3390/molecules27092978)
- 8. Schieven, G.L.; De Fex, H.; Stephenson, L. Hypochlorous acid activates tyrosine phosphorylation signal pathways leading to calcium signaling and TNFα production. *Antioxid. Redox Signal.* **2002**, *4*, 501–507. [\[CrossRef\]](https://doi.org/10.1089/15230860260196308) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/12215218)
- 9. Spickett, C.M.; Jerlich, A.; Panasenko, O.M.; Arnhold, J.; Pitt, A.R.; Stelmaszynska, T.; Schaur, R.J. The reactions of hypochlorous acid, the reactive oxygen species produced by myeloperoxidase, with lipids. *Acta Biochim. Pol.* **2000**, *47*, 889–899. [\[CrossRef\]](https://doi.org/10.18388/abp.2000_3944)
- 10. Tang, V.; Fu, S.L.; Rayner, B.S.; Hawkins, C.L. 8-Chloroadenosine induces apoptosis in human coronary artery endothelial cells through the activation of the unfolded protein response. *Redox Biol.* **2019**, *26*, 101274. [\[CrossRef\]](https://doi.org/10.1016/j.redox.2019.101274) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/31307008)
- 11. Andrés, C.M.C.; de la Lastra, J.M.P.; Juan, C.A.; Plou, F.J.; Pérez-Lebeña, E. Hypochlorous Acid Chemistry in Mammalian Cells-Influence on Infection and Role in Various Pathologies. *Int. J. Mol. Sci.* **2022**, *23*, 10735. [\[CrossRef\]](https://doi.org/10.3390/ijms231810735)
- 12. Aratani, Y. Myeloperoxidase: Its role for host defense, inflammation, and neutrophil function. *Arch. Biochem. Biophys.* **2018**, *640*, 47–52. [\[CrossRef\]](https://doi.org/10.1016/j.abb.2018.01.004)
- 13. Koyani, C.N.; Scheruebel, S.; Jin, G.; Kolesnik, E.; Zorn-Pauly, K.; Mächler, H.; Hoefler, G.; von Lewinski, D.; Heinzel, F.R.; Pelzmann, B.; et al. Hypochlorite-Modified LDL Induces Arrhythmia and Contractile Dysfunction in Cardiomyocytes. *Antioxidants* **2022**, *11*, 25. [\[CrossRef\]](https://doi.org/10.3390/antiox11010025)
- 14. Pravalika, K.; Sarmah, D.; Kaur, H.; Wanve, M.; Saraf, J.; Kalia, K.; Borah, A.; Yavagal, D.R.; Dave, K.R.; Bhattacharya, P. Myeloperoxidase and Neurological Disorder: A Crosstalk. *ACS Chem. Neurosci.* **2018**, *9*, 421–430. [\[CrossRef\]](https://doi.org/10.1021/acschemneuro.7b00462)
- 15. Dong, H.; Zhou, Y.L.; Zhao, L.; Hao, Y.Q.; Zhang, Y.T.; Ye, B.X.; Xu, M.T. Dual-Response Ratiometric Electrochemical Microsensor for Effective Simultaneous Monitoring of Hypochlorous Acid and Ascorbic Acid in Human Body Fluids. *Anal. Chem.* **2020**, *92*, 15079–15086. [\[CrossRef\]](https://doi.org/10.1021/acs.analchem.0c03089)
- 16. Hajnsek, M.; Schiffer, D.; Harrich, D.; Koller, D.; Verient, V.; van der Palen, J.; Heinzle, A.; Binder, B.; Sigl, E.; Sinner, F.; et al. An electrochemical sensor for fast detection of wound infection based on myeloperoxidase activity. *Sens. Actuators B-Chem.* **2015**, *209*, 265–274. [\[CrossRef\]](https://doi.org/10.1016/j.snb.2014.11.125)
- 17. Zhang, R.; Song, B.; Yuan, J.L. Bioanalytical methods for hypochlorous acid detection: Recent advances and challenges. *Trac-Trends Anal. Chem.* **2018**, *99*, 1–33. [\[CrossRef\]](https://doi.org/10.1016/j.trac.2017.11.015)
- 18. Okamoto, N.; Bito, T.; Hiura, N.; Yamamoto, A.; Iida, M.; Baba, Y.; Fujita, T.; Ishihara, A.; Yabuta, Y.; Watanabe, F. Food Additives (Hypochlorous Acid Water, Sodium Metabisulfite, and Sodium Sulfite) Strongly Affect the Chemical and Biological Properties of Vitamin B₁₂ in Aqueous Solution. *ACS Omega* **2020**, *5*, 6207–6214. [\[CrossRef\]](https://doi.org/10.1021/acsomega.0c00425)
- 19. Wang, Y.; Hong, F.; Li, D.P.; Qi, J.; Liu, X.M. A novel strategy for evaluation of natural products acting on the myeloperoxidase/hypochlorous acid system by combining high-performance liquid chromatography-photodiode array detectionchemiluminescence and ultrafiltration-mass spectrometry techniques. *J. Sep. Sci.* **2018**, *41*, 194–1201. [\[CrossRef\]](https://doi.org/10.1002/jssc.201800658) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/30194713)
- 20. Dai, Y.C.; Zhang, K.X.; Yuan, X.H.; Xie, X.B.; Zhan, Z.X.; Lv, Y. Novel Near-Infrared Iridium(III) Complex for Chemiluminescence Imaging of Hypochlorous Acid. *Anal. Chem.* **2023**, *95*, 8310–8317. [\[CrossRef\]](https://doi.org/10.1021/acs.analchem.3c00738) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/37200201)
- 21. Magalhaes, L.M.; Segundo, M.A.; Reis, S.; Lima, J.; Estela, J.M.; Cerdà, V. Automatic in vitro determination of hypochlorous acid scavenging capacity exploiting multisyringe flow injection analysis and chemiluminescence. *Anal. Chem.* **2007**, *79*, 3933–3939. [\[CrossRef\]](https://doi.org/10.1021/ac0700068) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/17425285)
- 22. Wang, C.A.F.; Wang, T.; Zhang, M.Y.; Li, N.; Fan, M.T.; Cui, X.Y. Chemiluminescence molecular sensor for endogenous HOCl in vivo. *Sens. Actuators B-Chem.* **2021**, *339*, 129927. [\[CrossRef\]](https://doi.org/10.1016/j.snb.2021.129927)
- 23. Han, Q.; Zhou, F.; Wang, Y.; Feng, H.; Meng, Q.T.; Zhang, Z.Q.; Zhang, R. A Redox-Switchable Colorimetric Probe for "Naked-Eye" Detection of Hypochlorous Acid and Glutathione. *Molecules* **2019**, *24*, 2455. [\[CrossRef\]](https://doi.org/10.3390/molecules24132455) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/31277409)
- 24. Yu, Q.; Chen, X.L.; Liu, H.; Zhang, Q.L. Recent Progress in Colorimetric and Fluorimetric Probes for the Detection of Hypochlorous Acid. *Chin. J. Org. Chem.* **2020**, *40*, 1206–1231. [\[CrossRef\]](https://doi.org/10.6023/cjoc201911004)
- 25. He, S.; Dong, H.; Hao, Y.Q.; Zhang, Y.T.; Zhou, Y.L.; Zhang, F.Y.; Li, J.; Jia, Y.Y.; Xiao, G.Q.; Xu, M.T. Quantifying Hypochlorous Acid Concentration in Environmental Water Using Smartphone Colorimetry. *J. Anal. Test.* **2021**, *5*, 360–369. [\[CrossRef\]](https://doi.org/10.1007/s41664-021-00156-1)
- 26. Hao, Y.Q.; Chen, S.; Zhou, Y.L.; Zhang, Y.T.; Xu, M.T. Recent Progress in Metal-Organic Framework (MOF) Based Luminescent Chemodosimeters. *Nanomaterials* **2019**, *9*, 974. [\[CrossRef\]](https://doi.org/10.3390/nano9070974) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/31277318)
- 27. Wu, D.; Chen, L.Y.; Xu, Q.L.; Chen, X.Q.; Yoon, J.Y. Design Principles, Sensing Mechanisms, and Applications of Highly Specific Fluorescent Probes for HOCl/OCl−. *Acc. Chem. Res.* **2019**, *52*, 2158–2168. [\[CrossRef\]](https://doi.org/10.1021/acs.accounts.9b00307)
- 28. Fang, Y.Y.; Dehaen, W. Fluorescent Probes for Selective Recognition of Hypobromous Acid: Achievements and Future Perspectives. *Molecules* **2021**, *26*, 363. [\[CrossRef\]](https://doi.org/10.3390/molecules26020363)
- 29. Dai, M.C.; Yang, Y.J.; Sarkar, S.; Ahn, K.H. Strategies to convert organic fluorophores into red/near-infrared emitting analogues and their utilization in bioimaging probes. *Chem. Soc. Rev.* **2023**, *52*, 6344–6358. [\[CrossRef\]](https://doi.org/10.1039/D3CS00475A)
- 30. Hou, J.T.; Kwon, N.; Wang, S.; Wang, B.Y.; He, X.J.; Yoon, J.; Shen, J.L. Sulfur-based fluorescent probes for HOCl: Mechanisms, design, and applications. *Coord. Chem. Rev.* **2022**, *450*, 214232. [\[CrossRef\]](https://doi.org/10.1016/j.ccr.2021.214232)
- 31. Zhang, Y.-R.; Liu, Y.; Feng, X.; Zhao, B.-X. Recent progress in the development of fluorescent probes for the detection of hypochlorous acid. *Sens. Actuators B Chem.* **2017**, *240*, 18–36. [\[CrossRef\]](https://doi.org/10.1016/j.snb.2016.08.066)
- 32. Kwon, N.; Chen, Y.; Chen, X.; Kim, M.H.; Yoon, J. Recent progress on small molecule-based fluorescent imaging probes for hypochlorous acid (HOCl)/hypochlorite (OCl−). *Dye. Pigment.* **2022**, *200*, 110132. [\[CrossRef\]](https://doi.org/10.1016/j.dyepig.2022.110132)
- 33. Ren, C.P.; Nie, W.; Leng, J.Q.; Liu, Z.B. Reactive Fluorescent Probe for Hypochlorite. *Prog. Chem.* **2021**, *33*, 942–957. [\[CrossRef\]](https://doi.org/10.7536/pc200869)
- 34. Liu, D.; Yue, X.X.; Zhang, H.K.; Li, K.; Yang, Z.H.; Wang, B.H.; Song, X.Z. Probing drug-mediated fluctuations of HClO levels in the endoplasmic reticulum by a ratiometric fluorescent probe with a large emission shift. *Dye. Pigment.* **2023**, *215*, 111257. [\[CrossRef\]](https://doi.org/10.1016/j.dyepig.2023.111257)
- 35. Wang, B.Y.; Guo, X.M.; Liu, Z.D.; Wu, Y.Q.; Hou, J.T. A Long-wavelength Emissive Phenothiazine Derived Fluorescent Probe for Detecting HOCl Upregulation in 5-FU Stimulated Living Cells. *Chem. Res. Chin. Univ.* **2022**, *38*, 609–615. [\[CrossRef\]](https://doi.org/10.1007/s40242-021-1173-8)
- 36. Hao, Y.Q.; Zhang, Y.T.; Sun, Q.L.; Chen, S.; Tang, Z.L.; Zeng, R.J.; Xu, M.T. Phenothiazine-coumarin-pyridine hybrid as an efficient fluorescent probe for ratiometric sensing hypochlorous acid. *Microchem. J.* **2021**, *171*, 106851. [\[CrossRef\]](https://doi.org/10.1016/j.microc.2021.106851)
- 37. Jiao, X.J.; Huang, K.; He, S.; Liu, C.; Zhao, L.C.; Zeng, X.S. A mitochondria-targeted near-infrared fluorescent probe with a large Stokes shift for real-time detection of hypochlorous acid. *Org. Biomol. Chem.* **2019**, *17*, 108–114. [\[CrossRef\]](https://doi.org/10.1039/C8OB02583E) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/30520928)
- 38. Zheng, D.S.; Qiu, X.Y.; Liu, C.; Jiao, X.J.; He, S.; Zhao, L.C.; Zeng, X.S. Synthesis and bioapplication of a highly selective and sensitive fluorescent probe for HOCl based on a phenothiazine-dicyanoisophorone conjugate with large Stokes shift. *New J. Chem.* **2018**, *42*, 5135–5141. [\[CrossRef\]](https://doi.org/10.1039/C8NJ00279G)
- 39. Zhi, X.; Qian, Y. A novel red-emission phenothiazine fluorescent protein chromophore based on oxygen–chlorine bond (O-Cl) formation for real-time detection of hypochlorous acid in cells. *Talanta* **2021**, *222*, 121503. [\[CrossRef\]](https://doi.org/10.1016/j.talanta.2020.121503)
- 40. Han, J.; Yang, S.; Wang, B.; Song, X. Tackling the Selectivity Dilemma of Benzopyrylium–Coumarin Dyes in Fluorescence Sensing of HClO and SO2. *Anal. Chem.* **2021**, *93*, 5194–5200. [\[CrossRef\]](https://doi.org/10.1021/acs.analchem.0c05266)
- 41. Onoabedje, E.A.; Egu, S.A.; Ezeokonkwo, M.A.; Okoro, U.C. Highlights of molecular structures and applications of phenothiazine & phenoxazine polycycles. *J. Mol. Struct.* **2019**, *1175*, 956–962. [\[CrossRef\]](https://doi.org/10.1016/j.molstruc.2018.08.064)
- 42. Simon, A.; Alexandru, T.; Boni, M.; Damian, V.; Stoicu, A.; Dutschk, V.; Pascu, M.L. Interaction of solutions containing phenothiazines exposed to laser radiation with materials surfaces, in view of biomedical applications. *Int. J. Pharm.* **2014**, *475*, 270–281. [\[CrossRef\]](https://doi.org/10.1016/j.ijpharm.2014.08.031)
- 43. Devadiga, D.; Selvakumar, M.; Shetty, P.; Santosh, M.S.; Chandrabose, R.S.; Karazhanov, S. Recent developments in metal-free organic sensitizers derived from carbazole, triphenylamine, and phenothiazine for dye-sensitized solar cells. *Int. J. Energy Res.* **2021**, *45*, 6584–6643. [\[CrossRef\]](https://doi.org/10.1002/er.6348)
- 44. Buene, A.F.; Almenningen, D.M. Phenothiazine and phenoxazine sensitizers for dye-sensitized solar cells—an investigative review of two complete dye classes. *J. Mater. Chem. C* **2021**, *9*, 11974–11994. [\[CrossRef\]](https://doi.org/10.1039/D1TC03207K)
- 45. Luo, J.S.; Wan, Z.Q.; Jia, C.Y. Recent advances in phenothiazine-based dyes for dye-sensitized solar cells. *Chin. Chem. Lett.* **2016**, *27*, 1304–1318. [\[CrossRef\]](https://doi.org/10.1016/j.cclet.2016.07.002)
- 46. Lu, X.L.; Zhan, Y.; He, W. Recent development of small-molecule fluorescent probes based on phenothiazine and its derivates. *J. Photochem. Photobiol. B-Biol.* **2022**, *234*, 112528. [\[CrossRef\]](https://doi.org/10.1016/j.jphotobiol.2022.112528) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/35907277)
- 47. Huang, T.H.; Yan, S.R.; Yu, Y.B.; Xue, Y.S.; Yu, Y.Y.; Han, C.P. Dual-Responsive Ratiometric Fluorescent Probe for Hypochlorite and Peroxynitrite Detection and Imaging In Vitro and In Vivo. *Anal. Chem.* **2022**, *94*, 1415–1424. [\[CrossRef\]](https://doi.org/10.1021/acs.analchem.1c04729)
- 48. Qiu, X.Y.; Liu, S.J.; Hao, Y.Q.; Sun, J.W.; Chen, S. Phenothiazine-based fluorescence probe for ratiometric imaging of hydrazine in living cells with remarkable Stokes shift. *Spectrochim. Acta Part A-Mol. Biomol. Spectrosc.* **2020**, *227*, 117675. [\[CrossRef\]](https://doi.org/10.1016/j.saa.2019.117675)
- 49. Rout, Y.; Montanari, C.; Pasciucco, E.; Misra, R.; Carlotti, B. Tuning the Fluorescence and the Intramolecular Charge Transfer of Phenothiazine Dipolar and Quadrupolar Derivatives by Oxygen Functionalization. *J. Am. Chem. Soc.* **2021**, *143*, 9933–9943. [\[CrossRef\]](https://doi.org/10.1021/jacs.1c04173)
- 50. Manusha, P.; Yadav, S.; Satija, J.; Senthilkumar, S. Designing electrochemical NADH sensor using silver nanoparticles/phenothiazine nanohybrid and investigation on the shape dependent sensing behavior. *Sens. Actuators B-Chem.* **2021**, *347*, 130649. [\[CrossRef\]](https://doi.org/10.1016/j.snb.2021.130649)
- 51. Manusha, P.; Senthilkumar, S. Design and synthesis of phenothiazine based imidazolium ionic liquid for electrochemical nonenzymatic detection of sulfite in food samples. *J. Mol. Liq.* **2020**, *301*, 112412. [\[CrossRef\]](https://doi.org/10.1016/j.molliq.2019.112412)
- 52. Lawrence, N.S.; Wang, J. Chemical adsorption of phenothiazine dyes onto carbon nanotubes: Toward the low potential detection of NADH. *Electrochem. Commun.* **2006**, *8*, 71–76. [\[CrossRef\]](https://doi.org/10.1016/j.elecom.2005.10.026)
- 53. Abdi, G.; Alluhaibi, L.; Kowalewska, E.; Mazur, T.; Mech, K.; Podborska, A.; Sławek, A.; Tanaka, H.; Szaciłowski, K. Reservoir computing and photoelectrochemical sensors: A marriage of convenience. *Coord. Chem. Rev.* **2023**, *487*, 215155. [\[CrossRef\]](https://doi.org/10.1016/j.ccr.2023.215155)
- 54. Liu, Q.; Zhang, H.; Jiang, H.; Yang, P.; Luo, L.; Niu, Q.; You, T. Photoactivities regulating of inorganic semiconductors and their applications in photoelectrochemical sensors for antibiotics analysis: A systematic review. *Biosens. Bioelectron.* **2022**, *216*, 114634. [\[CrossRef\]](https://doi.org/10.1016/j.bios.2022.114634) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/36049349)
- 55. Li, T.; Dong, H.; Hao, Y.; Zhang, Y.; Chen, S.; Xu, M.; Zhou, Y. Near-infrared Responsive Photoelectrochemical Biosensors. *Electroanalysis* **2022**, *34*, 956–965. [\[CrossRef\]](https://doi.org/10.1002/elan.202100355)
- 56. Shu, J.; Tang, D. Recent Advances in Photoelectrochemical Sensing: From Engineered Photoactive Materials to Sensing Devices and Detection Modes. *Anal. Chem.* **2020**, *92*, 363–377. [\[CrossRef\]](https://doi.org/10.1021/acs.analchem.9b04199)
- 57. Zhao, W.-W.; Xu, J.-J.; Chen, H.-Y. Photoelectrochemical bioanalysis: The state of the art. *Chem. Soc. Rev.* **2015**, *44*, 729–741. [\[CrossRef\]](https://doi.org/10.1039/C4CS00228H)
- 58. Zhou, Y.; Yin, H.; Ai, S. Applications of two-dimensional layered nanomaterials in photoelectrochemical sensors: A comprehensive review. *Coord. Chem. Rev.* **2021**, *447*, 214156. [\[CrossRef\]](https://doi.org/10.1016/j.ccr.2021.214156)
- 59. Cheng, J.; Luo, Y.; Hao, Y.; Han, H.; Hu, X.; Yang, Y.; Long, X.; He, J.; Zhang, P.; Zeng, R.; et al. A responsive organic probe based photoelectrochemical sensor for hydrazine detection. *Spectrochim. Acta Part A Mol. Biomol. Spectrosc.* **2024**, *305*, 123463. [\[CrossRef\]](https://doi.org/10.1016/j.saa.2023.123463)
- 60. Li, T.; Hao, Y.; Dong, H.; Li, C.; Liu, J.; Zhang, Y.; Tang, Z.; Zeng, R.; Xu, M.; Chen, S. Target-Induced In Situ Formation of Organic Photosensitizer: A New Strategy for Photoelectrochemical Sensing. *ACS Sens.* **2022**, *7*, 415–422. [\[CrossRef\]](https://doi.org/10.1021/acssensors.1c02595)
- 61. Hao, Y.; Li, T.; Luo, L.; Fan, S.; Chen, S.; Zhang, Y.; Tang, Z.; Xu, M.; Zeng, R.; Chen, S. A reaction based dual-modal probe for fluorescent and photoelectrochemical determination of thiophenol. *Sens. Actuators B Chem.* **2022**, *369*, 132405. [\[CrossRef\]](https://doi.org/10.1016/j.snb.2022.132405)
- 62. Xiang, Y.; Kong, Y.; Feng, W.; Ye, X.; Liu, Z. A ratiometric photoelectrochemical microsensor based on a small-molecule organic semiconductor for reliable in vivo analysis. *Chem. Sci.* **2021**, *12*, 12977–12984. [\[CrossRef\]](https://doi.org/10.1039/D1SC03069H) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/34745528)
- 63. Wu, S.; Tu, W.; Zhao, Y.; Wang, X.; Song, J.; Yang, X. Phosphonate-Substituted Ruthenium(II) Bipyridyl Derivative as a Photoelectrochemical Probe for Sensitive and Selective Detection of Mercury(II) in Biofluids. *Anal. Chem.* **2018**, *90*, 14423–14432. [\[CrossRef\]](https://doi.org/10.1021/acs.analchem.8b03985)
- 64. Chu, J.-W.; Trout, B.L. On the Mechanisms of Oxidation of Organic Sulfides by H2O2 in Aqueous Solutions. *J. Am. Chem. Soc.* **2004**, *126*, 900–908. [\[CrossRef\]](https://doi.org/10.1021/ja036762m) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/14733566)
- 65. Zeida, A.; Trujillo, M.; Ferrer-Sueta, G.; Denicola, A.; Estrin, D.A.; Radi, R. Catalysis of Peroxide Reduction by Fast Reacting Protein Thiols. *Chem. Rev.* **2019**, *119*, 10829–10855. [\[CrossRef\]](https://doi.org/10.1021/acs.chemrev.9b00371) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/31498605)
- 66. Yin, C.; Zhu, H.; Xie, C.; Zhang, L.; Chen, P.; Fan, Q.; Huang, W.; Pu, K. Organic Nanoprobe Cocktails for Multilocal and Multicolor Fluorescence Imaging of Reactive Oxygen Species. *Adv. Funct. Mater.* **2017**, *27*, 1700493. [\[CrossRef\]](https://doi.org/10.1002/adfm.201700493)
- 67. Liu, Y.; Li, Y.; Yu, L.; Yang, Z.; Ding, J.; Wang, K.-N.; Zhang, Y. Monocomponent Nanodots with Dichromatic Output Regulated by Synergistic Dual-Stimuli for Cervical Cancer Tissue Imaging and Photodynamic Tumor Therapy. *Anal. Chem.* **2022**, *94*, 811–819. [\[CrossRef\]](https://doi.org/10.1021/acs.analchem.1c03488)
- 68. Wang, S.; Zhang, B.; Wang, W.; Feng, G.; Yuan, D.; Zhang, X. Elucidating the Structure-Reactivity Correlations of Phenothiazine-Based Fluorescent Probes toward ClO−. *Chem.-A Eur. J.* **2018**, *24*, 8157–8166. [\[CrossRef\]](https://doi.org/10.1002/chem.201800356)
- 69. Xiao, H.; Xin, K.; Dou, H.; Yin, G.; Quan, Y.; Wang, R. A fast-responsive mitochondria-targeted fluorescent probe detecting endogenous hypochlorite in living RAW 264.7 cells and nude mouse. *Chem. Commun.* **2015**, *51*, 1442–1445. [\[CrossRef\]](https://doi.org/10.1039/C4CC07411D)
- 70. Liang, L.; Liu, C.; Jiao, X.; Zhao, L.; Zeng, X. A highly selective and sensitive photoinduced electron transfer (PET) based HOCl fluorescent probe in water and its endogenous imaging in living cells. *Chem. Commun.* **2016**, *52*, 7982–7985. [\[CrossRef\]](https://doi.org/10.1039/C6CC02603F)

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