



Review

Guidance of Signaling Activations by Cadherins and Integrins in Epithelial Ovarian Cancer Cells

Francesca Roggiani, Delia Mezzanzanica, Katia Rea * and Antonella Tomassetti *

Unit of Molecular Therapies, Department of Experimental Oncology and Molecular Medicine, Fondazione IRCCS Istituto Nazionale dei Tumori, Via Amadeo 42, Milan 20133, Italy; francesca.roggiani@istitutotumori.mi.it (F.R.); delia.mezzanzanica@istitutotumori.mi.it (D.M.)

* Correspondence: katia.rea@istitutotumori.mi.it (K.R.); antonella.tomassetti@istitutotumori.mi.it (A.T.); Tel.: +39-02-2390-2570 (K.R.); +39-02-2390-2568 (A.T.); Fax: +39-02-2390-3073 (K.R. & A.T.)

Academic Editor: Kwong-Kwok Wong

Received: 14 July 2016; Accepted: 13 August 2016; Published: 23 August 2016

Abstract: Epithelial ovarian cancer (EOC) is the deadliest tumor among gynecological cancer in the industrialized countries. The EOC incidence and mortality have remained unchanged over the last 30 years, despite the progress in diagnosis and treatment. In order to develop novel and more effective therapeutic approaches, the molecular mechanisms involved in EOC progression have been thoroughly investigated in the last few decades. At the late stage, peritoneal metastases originate from the attachment of small clusters of cancer cells that shed from the primary site and carried by the ascites adhere to the abdominal peritoneum or omentum. This behavior suggests that cell–cell or cell–matrix adhesion mechanisms regulate EOC growth and dissemination. Complex downstream signalings, which might be influenced by functional cross-talk between adhesion molecules and co-expressed and activated signaling proteins, can affect the proliferation/survival and the migration/invasion of EOC cells. This review aimed to define the impact of the mechanisms of cell–cell, through cadherins, and cell–extracellular matrix adhesion, through integrins, on the signaling cascades induced by membrane receptors and cytoplasmic proteins known to have a role in the proliferation, migration and invasion of EOC cells. Finally, some novel approaches using peptidomimetic ligands to cadherin and integrins are summarized.

Keywords: epithelial ovarian cancer; adhesion; cadherin; integrin; signal transduction; proliferation; migration; invasion

1. Introduction

Epithelial ovarian cancer (EOC) is a devastating disease with an overall five-year survival rate of approximately 45% [1,2]. EOCs are usually diagnosed when malignant cells have already invaded the peritoneal cavity and, although most of the patients are sensitive to the first line chemotherapy, 50% of them relapse with a chemoresistant disease. For all these reasons, EOCs are the fifth main cause of cancer-related deaths among women, and the primary cause of death from gynecological cancer [3]. Therefore, in cancer research, investigations aiming to clarify the mechanisms of EOC tumorigenesis and progression are one of the most important areas. EOC group different diseases with a common anatomical location [4] but display high molecular and etiological heterogeneity [5–8].

EOCs are divided into two large groups [9] designated types I, genetically stable and not very aggressive [10], and type II, genetically unstable and very aggressive tumors, which are usually diagnosed at the advanced-stage. Type II tumors include high-grade serous, high-grade endometrioid, malignant mixed mesodermal tumors (carcinosarcomas), and undifferentiated carcinomas being the serous high-grade ovarian carcinoma (HGSOC) the most representative tumors [11].

More than 50% of EOC patients at the late stage, in particular those with type-II tumors, present with ascites/effusions in their abdominal cavity rich of tumor cells [12]. The ascites are accumulated since implanted tumor cells give rise to the obstruction of lymphatic vessels, preventing the outflow of fluid that transpires from the tumor vessels. Hence, patients affected by HGSOC type-II tumors have the peritoneal cavity invaded by metastatic tumors, growing in the solid stromal matrices, and multicellular aggregates (MCAs) floating and growing in the malignant ascites [13]. These MCAs overcome anoikis [14] and persist as ascites [15]. Although a possible mechanism of hematogenous HGSOC metastasis formation to the omentum has also been reported [16], the general consensus is that these MCAs originate by the shedding of malignant cells into the peritoneum from the primary tumor and that disaggregation and attachment to the sub-mesothelial extracellular matrix (ECM) allow the formation of secondary lesions [17–19]. Once adhered to the peritoneum, EOC cells proliferate, migrate and invade the surrounding tissues. An elegant approach of a live image-based in vitro model determined that a myosin-generated force allows EOC MCAs to displace and remove the mesothelial monolayer. This process is now known as mesothelial cell clearance [20]. The EOC metastasis outgrowth occurs upon the remodeling of cell–cell adhesion molecules (i.e., cadherins) during spheroids dis-aggregation. In addition, the integrins expressed on the surface of EOC cells are essential to the attachment of EOC cells to the sub-mesothelial ECM [18–21].

Interestingly, a proteomic approach for MCA analysis for EOC patient stratification has been able to identify three adhesion-related subsets with potential predictive impact [22]. These data highlight the influence of the adhesion molecules in the clinical EOC behavior.

Overall, the processes of EOC progression require that cell–cell, through cadherins, and cell–ECM adhesion, through integrins, cooperate, directly or indirectly, to the activation of signaling pathways relevant to the proliferation/survival and the migration/invasion mechanisms of EOC cells.

2. Cadherin-Associated Signaling Activation

Cell–cell adhesion is mediated by cadherins (cadhs) through the calcium-dependent homophilic interaction of their extracellular domain to form, associated with cytoplasmic proteins (β - and α -catenin (ctn) together with p120ctn) intercellular structures called adherens junctions (AJs) (see, for review, [23]). The AJs constitute a physical bridge between the cadherin complex and the cortical actin filaments thus regulating cellular dynamic behaviors, such as rearrangements, movement and shape changes during embryonic development as well as during neoplastic transformation and progression. The cytoplasmic tail of cadhs has no catalytic activity; therefore, possible signaling activation must occur upon the recruitment of signaling molecules to the site of the cadh–ctn complex [24]. In epithelial tissues, the homophilic E-cadh ligation can activate the junctional GTPase signaling downstream to Rac [25–28] and Cdc42 [26,29]. E-cadh can therefore regulate the localization and function of the Rho GTPase with a mechanism described as ‘outside-in signaling’ [30,31] (Figure 1a).

Below, E- and P-cadh-associated signalings are discussed. N-cadh displays similar structural and functional properties to E-cadh (see, for review, [32]). In EOCs, N-cadh expression is associated with a more aggressive and chemoresistant phenotype [33], but its role in modulating signaling activation is still unclear.

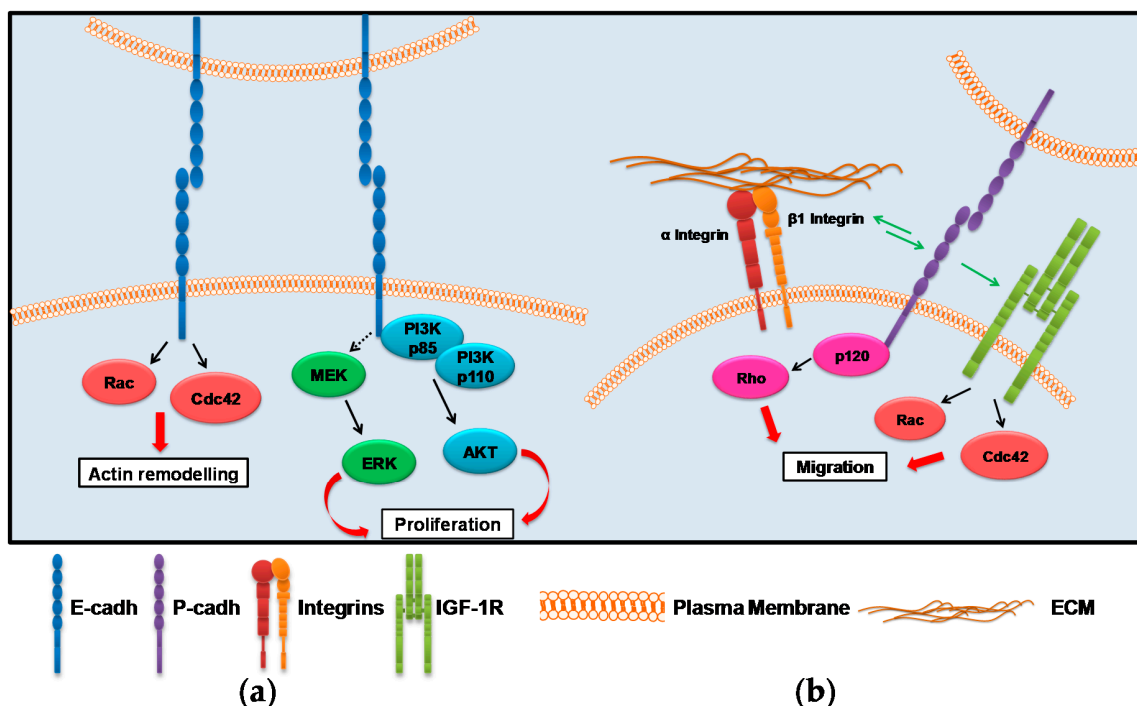


Figure 1. Schematic representation of (a) E- and (b) P-cadh-associated signaling cascades activated in EOCs (Epithelial Ovarian Cancers). The arrows indicate: green, protein-protein interaction, black, signaling cascades; discontinuous, possible signaling cascade; red, cellular effects. The abbreviation used are: cadh, cadherin; IGF-1R, Insulin Growth Factor Receptor 1; ERK, Extracellular signal-Regulated Kinase; MEK, MAP or ERK kinase; PI3K, phosphoinositide 3-kinase; p120, p120 catenin. Rho, Rac and CDC42 are Rho-GTPases.

2.1. E-cadh

In cancer, the switch in expression of E-cadh to N-cadh or the up-regulation of cadh-11 and P-cadh are important indicators of progression and are believed to facilitate the epithelial–mesenchymal transition (EMT), which leads to a more migratory and invasive phenotype [34,35]. In metastatic solid tumors, E-cadh expression is usually lost in human breast, bladder, lung and pancreatic carcinomas, which nevertheless express other cadherins, especially N-cadh [36]. For this reason, E-cadh has been proposed as a tumor suppressor since, in invasive carcinoma cells, ectopically expressed E-cadh was associated with a decreased growth potential [37]. In confluent cells, E-cadh was shown to be associated to epidermal growth factor receptor (EGFR) leading to receptor immobilization and alteration of the ligand-receptor affinity, thus inhibiting the signaling activation [38,39].

Very recent publications are beginning to revisit the general consensus that EMT is necessary for the metastasis formation occurring when the tumors become more malignant. In these reports, E-cadh expression was found maintained during the formation of lung metastasis in mouse models of breast and pancreatic cancers [40,41]. These data are not surprising since E-cadh loss was previously showed to be a rare event in both invasive ductal [42] and inflammatory breast carcinomas [43]. Mechanistically, in a breast carcinoma patient-derived 3D organoid model, E-cadh homophilic adhesion was present in a leading cell population and necessary during invasion of collagen I gel [44,45].

In EOCs, initial immunohistochemical analysis revealed low E-cadh expression in advanced-stage disease [46,47]. Data made on a higher number of EOC biopsies assessed that E-cadh is still expressed at cell–cell contacts in late stage tumors [48–51]. Furthermore, in EOC MCAs, the predominant cadherin expressed at cell–cell contact is E-cadh (our unpublished results and [52]) and contributes to aggregate formation [53]. E-cadh was first assessed to upregulate ligand-independent EGFR trans-phosphorylation leading to AKT and MAPK activations [54] (Figure 1a). Furthermore,

E-cadh-mediated AJ formation recruits PI3K-p85 to the cell membrane contributing to activation of the PI3K/AKT pathway and to the up-modulation of EOC growth [55]. In the EOC cell line SKOV3, E-cadh functionality can also increase mitogen-activated protein kinase/extracellular-signal regulated kinase (MEK/ERK) activation, although no physical association with MEK or even EGFR was demonstrated [56] (Figure 1b).

While in normal epithelial cells E-cadh is maintained on the membrane by p120 catenin and its levels are controlled through the endocytic pathway [57], in transformed cells, E-cadh loss is also due to the proteolytic cleavage of the extra-cellular domain resulting in a 80-KDa fragment. This soluble (s) E-cadh might be generated through the action of metalloproteases (MMTs), such as ADAM10 and MMPs [58,59] usually upregulated in cancers, or γ -secretases, like presenilin 1 [60]. The 80 KDa soluble form of E-cadh has been found in the serum of cancer patients with poor prognosis [61]. In EOCs, the levels of sE-cadh in cystic fluid from patients presenting with complex ovarian masses was higher than in patients presenting with benign tumors [60]. sE-cadh was found in the EOC ascites, and in vitro experiments demonstrated that the integrin clustering occurring during the adhesion to the sub-mesothelial collagen I stimulated EGFR-dependent MMP9 expression [62], thus inducing sE-cadh production [63]. These data suggest that EGFR activation and production of sE-cadh constitute a feedback loop able to promote proliferation and/or migration/invasion. More recently, the 85-KDa and 23-KDa E-cadh, produced by calpain, were found in solid metastatic peritoneal masses of advanced EOC patients [64]. The presence of the 85-KDa E-cadh fragment correlated with a worse patient outcome, and the authors hypothesized that the 85-KDa fragment might enable tumor cluster formation and peritoneal dissemination [64].

2.2. P-cadh

Among cadherins, P-cadh has a proven role during the progression of several carcinomas, as cervix [65], esophagous [66], breast [67] and colon [68], including EOCs [69]. In EOC, an increase in P-cadh expression in the tumor masses, concomitant with a decrease of E-cadh expression, is associated with a progression from stage I to stage II tumors [69]. The pro-metastatic role of P-cadh is mainly due to a cross-talk with the axis constituted by the cytoplasmic p120ctn and the activation of the Rho GTPase signaling. In this context, once Gonadotropin-releasing hormone (GnRH), whose physiologic role is the control of pituitary gonadotropin secretion, induces the E- to P-cadh switching, p120ctn moves from the membrane to the cytoplasm, thus leading to activation of mechanisms of migration and invasion [70]. The role of p120ctn as determinant of migration upon GnRH stimulation is due to a trans-activation of insulin-like growth factor-1 receptor (IGF-1R) by P-cadh, which induces Rac1 and Cdc42 activations [71].

P-cadh also contributes to the adhesion of EOC cells to the peritoneum and in the metastasis formation and has been considered the predominant cadherin subtype expressed in the MCAs of the peritoneal effusion [69]. Furthermore, its inhibition reduced tumor growth, ascites formation and metastasis in in vivo pre-clinical models [72]. Besides its role in the MCA formation, P-cadh can also contribute to inhibition of anoikis. Accordingly, in vitro P-cadh knockdown increases cell death of EOC cells growth in suspension and in vivo decreases the dissemination of EOC cells injected in the peritoneum of immunocompromised mice [73], once again through a cross-talk with p120ctn and Rho GTPase activation. Another report has recently showed a P-cadh role in the MCA formation through a cross-talk with β 1 integrin. In particular, P-cadh induces the Golgi glycosyltransferase (ST6Gal-1), which mediate β 1 integrin hypersialylation, a post-translational modification that results in maturation of this integrin and activation of the downstream signaling [74] (Figure 1b).

3. Integrin-Associated Signaling Activation

Integrins are $\alpha\beta$ heterodimeric transmembrane proteins implicated in numerous physiological processes including adhesion to the extracellular matrix, proliferation, survival, migration and differentiation [75]. The molecular and physical interactions among the cells and with ECM can

strongly affect cell behavior, thus inducing cancer cell invasion and dissemination. A group of integrins recognizes and binds to the Arg-Gly-Asp (RGD) motif present on the ECM proteins, and the specificity of integrin binding to different ECM proteins is determined, in part, by other amino acids surrounding the RGD sequence [76]. As already said for the cadherins, integrin cytoplasmic tails do not exert a kinase activity but instead are able to activate specific intracellular kinases, such as FAK (Focal Adhesion Kinase), which, in turn, recruit the src kinase [77]. Src phosphorylates a number of FAK-associated proteins including paxilin, tensin and the adaptor p130CAS (Crk-Associated Substrate). FAK activation also leads to the recruitment of other SH2-containing proteins, including the PI3K, PLC- γ and the adapter proteins Grb7 [77] and Grb2 to partially mediate ERK activation [78]. Finally, the FAK/Src complex modulates the activity of small GTPases leading to the actin cytoskeleton remodeling necessary for cell adhesion and migration [79].

In EOC, the expression of ECM components and integrins has been found to be involved not only in the cell detachment from the primary tumor and from the peritoneal metastasis [80–82], but also in the resistance to anoikis of the EOC spheroids [83,84]. Dysregulation and aberrant deposition of ECM components, due to the activation of different MMPs [85], contribute to tumor progression. The expression of vitronectin [80] and fibronectin [86] at the periphery of mesothelial cells actively participates in EOC cell motility and metastasis. In addition, fibronectin has been also shown to mediate EOC cell migration and invasion through the upregulation of the FAK/PI3K/Akt pathway [87] and is an indicator of poor prognosis in invasive EOC [88].

3.1. Integrins and Receptor Tyrosine Kinase (RTKs)

Numerous studies suggest that the cooperation between integrins and RTKs exists and exerts a central role in cancer progression regulating invasion, proliferation and survival (see, for review, [89]). Considering that neither α or β subunit possess catalytic activity, it is possible that multiple mechanisms may regulate the cross-talk between integrins and RTKs. Three main types of integrins/RTKs interaction have been identified [90] (Figure 2): (1) integrins can physically bind to RTKs; (2) integrins clustering upon the binding to ECM can enhance signaling pathways that are activated following ligand-dependent RTKs activation; and (3) integrins and RTKs reciprocally control their surface expression.

The RTK represents the largest family of oncogenes, and the inhibitors targeting these receptors have already shown clinical efficacy as anti-cancer agents in solid tumors other than EOCs [91]. RTKs have been found to be over-expressed in EOCs as compared to normal counterparts [92].

3.1.1. EGFR

The EGFR is a transmembrane glycoprotein receptor that belongs to the ErbB family of RTKs. The ligands for the EGFR are the EGF, the betacellulin (BTC), TGF- α , amphiregulin (AR), and epiregulin (EPR) [93]. EGF ligands can induce the homodimerization of EGFR as well as the heterodimerization with other members of the family: HER2, HER3 and HER4 [93].

In solid tumors, EGFR can interact with many integrins such as $\beta 1$ [94], $\alpha 6\beta 4$ [95] and $\alpha v\beta 3$ [96,97], likely by forming a multimeric complex that also includes Src and the adaptor protein p130Cas [96]. This complex is necessary for the ligand-independent activation of the EGFR leading to signaling involved in cell survival and proliferation in response to ECM [94]. EGFR is expressed in up to 70% of EOCs [98], and its activation contributes to cell proliferation, invasion, angiogenesis and resistance to apoptosis [99]. EGFR activation can also induce a pro-inflammatory program with the co-expression of IL-6 and PAI-1 in a subset of EGFR-expressing EOCs with shorter progression-free survival after chemotherapy [100]. A direct correlation between EGFR and integrin expression has been reported. Indeed, reduced expression of EGFR in NIH:OVCA8 cells was associated with reduced integrin $\alpha 6$, a laminin-1 receptor, and cell adhesion with the down-modulation of MMP-9 activity, thus reducing the aggressiveness of EOC cells [101]. Conversely, the expression of a constitutively active EGFR has

been proposed to contribute to a more aggressive disease throughout the down-modulation of the integrin $\alpha 2$ expression, which leads to changes in cell shape and focal adhesion formation [102].

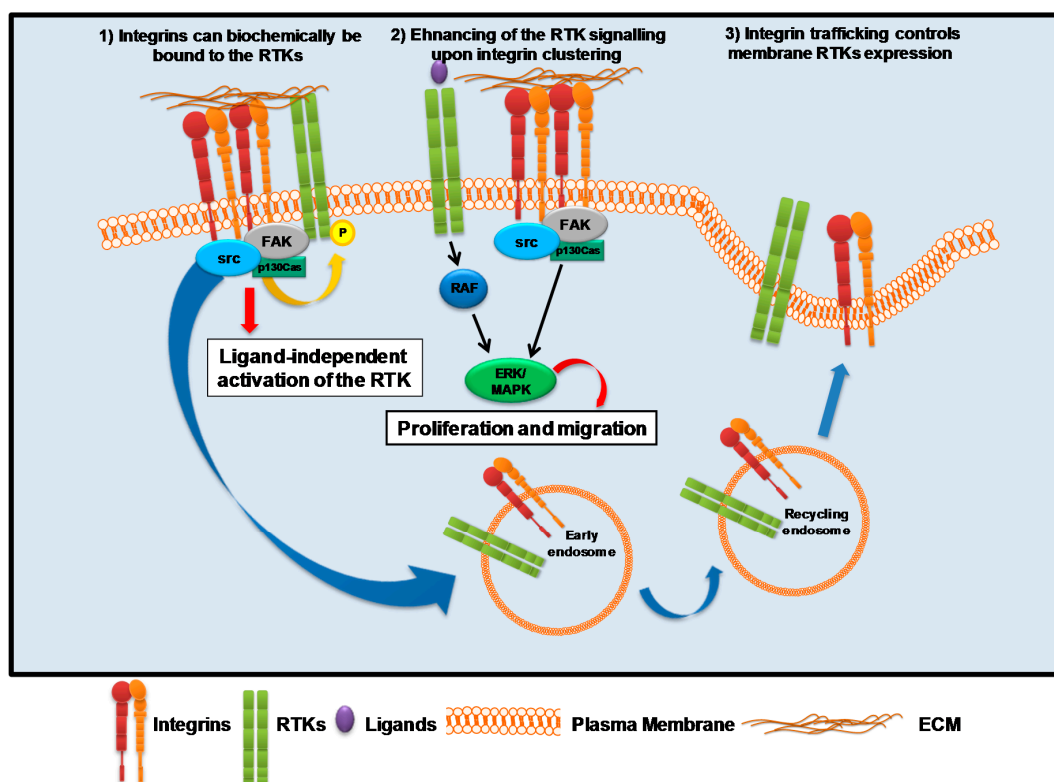


Figure 2. Schematic representation of the the signaling cascades activated in cancer cells by the cross-talk between integrins and RTKs. The arrays indicate: black, signaling cascades; yellow, phosphorylation; discontinuous, possible signaling cascades; blue, trafficking; red, effect. The abbreviations are: RTK, Receptor Tyrosine Kinase; ECM, Extra Cellular Matrix, MAPK, Mitogen Activated Protein Kinase, FAK, Focal Adhesion Kinase; src, Sarcoma viral oncogene; ERK, Extracellular signal-Regulated Kinase.

3.1.2. c-MET

c-MET oncogene is a membrane receptor that is essential for embryonic development and wound healing [103]. The natural ligand for this receptor is the hepatocyte growth factor, HGF, whose binding to the c-MET receptor leads to receptor phosphorylation and activation [103].

As in other carcinomas, also in EOCs, Sawada et al. found that the over-expression of c-MET was associated with the worst prognosis [104]. The knockdown of c-Met expression by small interfering RNA (siRNA) induced a decreased activation of the MEK/ERK and PI3K/AKT signaling pathways and reduced adhesion, invasion, peritoneal dissemination, and tumor growth through the inhibition of the expression of $\alpha 5\beta 1$ integrin [104]. Furthermore, $\alpha 5\beta 1$ integrin binding to the fibronectin also leads to a ligand-independent activation of c-MET signaling to FAK and Src. This mechanism is due to a biochemical association between $\alpha 5$ integrin and c-MET [105]. Since it has been reported that cancer-associated mesothelial cells that colonize the peritoneum produce fibronectin [86], able to trap HGF produced by the fibroblasts of the tumor microenvironment [106], the inhibition of c-MET activation seems to be a promising target to block cancer metastasis. Recently, Moran-Jones et al. investigated the effect of a novel ATP competitive inhibitor, INC280 (Novartis, Basel, Switzerland), which selectively inhibits c-MET activation in a number of EOC in vitro models [107], also decreasing the migration of EOC cells adherent to ex vivo peritoneal tissue.

3.1.3. VEGFR Family

For EOC patients, targeting angiogenesis is a novel therapeutic option, which includes the use of bevacizumab, an antibody that binds to VEGF, thus inhibiting its association with the receptor [108]. The VEGF and the VEGFR are expressed by EOC cells, and increased VEGF expression has been associated with the tumor progression and poor survival [109,110].

The interaction between the VEGFR and integrins is very important for angiogenesis. In particular, ligand-dependent VEGFR-2 phosphorylation activates c-src which then phosphorylates the cytosolic tail of $\beta 3$ integrin, promoting a feedback loop with the formation of the VEGFR-2/ $\alpha v\beta 3$ complex and increased integrin and VEGFR-2 signaling [111].

In in vivo EOC models, the combination of anti-angiogenic and anti-integrin combination therapy using Bevacizumab and etaracizumab (an antibody that binds to the integrin $\alpha v\beta 3$ inhibiting its activation) was more effective in inhibiting tumor growth and microvessel density than individual drugs [112]. In particular, the etaracizumab was reported to be particularly efficient in reducing the volumes of VEGFR-dependent SKOV3ip tumors in nude mice [113]. These findings might open the possibility that a dual blockade of VEGF and the $\alpha v\beta 3$ integrin in combination with chemotherapy may impair EOC growth in those patients whose tumors display high vessel density.

3.1.4. Axl

Deregulation of the receptor tyrosine kinase Axl, a member of the TAM RTK family, which also includes Mer and Tyro3, has been implicated in aggressive phenotype, tumor metastases and the progression of several human cancers [114]. The ligand of TAM RTKs, Gas6, displays the highest activity for Axl, which has been proposed as a therapeutic target since its genetic and pharmacologic inhibition could prevent or overcome acquired resistance to EGFR inhibitors [115]. Indeed, an EMT signature including Axl was identified and associated with the resistance to EGFR and PI3K inhibitors [116]. Multiple downstream signaling pathways, including PI3K/Akt and MAPK, can be elicited by Axl/Gas6 in different cell types [117,118].

Axl expression increased in type II EOC primary tumors and metastases compared to the ovarian surface epithelium [119] and Gas6 has been identified as prognostic marker in HGEOCs [120]. Therapeutic blockade of Axl activation reduced the migratory and invasive capability of HGSOC cells in vitro and in xenograft models [121]. Recently, the signaling elicited by Axl activation was dissected showing a cross-talk with the integrin/ECM pathway through the adaptor protein p130Cas leading to Rac activation [122]. Furthermore, an Axl-driven 61-gene signature, which included several collagens and ECM-associated proteins, was associated to the most aggressive HGSOC subtype [122].

3.2. Integrins and RNASET2

A novel factor that may play a role in the complex integrin signaling is the protein RNASET2, a ribonuclease that belongs to the T2 family. This protein, known to be downregulated in EOC and EOC cell lines [123], has been found to display tumorigenic and metastatic suppressor properties, independent from its ribonuclease activity, in vivo [123–126]. Recently, a new role of this protein was discovered that led to the hypothesis that RNASET2 may affect the integrin signaling pathway. RNASET2 proved to have the capability to reorganize the actin cytoskeleton and consequently influence cells motility, invasion capability and cell adhesion to ECM. In fact, silencing of the protein in the EOC cell line NIH-OVCAR3 triggered a disruption of the network of actin filaments and stress fibers, inducing a pattern of peripheral actin filaments [126], thus leading to increased migration and adhesion to laminin, collagen I and IV. Of note, the adhesion of these cells on collagen I was accompanied to an increase of paxillin activation and, accordingly, to an increase of mature focal adhesions (FAs) [126]. Although RNASET2 and tr-T2-50, a recombinant truncated form of the human RNASET2 deprived of its RNase activity [127,128], showed the capability to bind actin in vitro, Acquati et al. did not find any co-localization between the actin cytoskeleton and RNASET2 in NIH-OVCAR3 cells [126],

supporting the hypothesis of a cross-talk with the integrin activation and consequent re-organization of the actin cytoskeleton. Conversely, RNASET2 resulted in being significantly down-regulated in drug-resistant EOC biopsies and EOC cell lines, suggesting an involvement of this protein in the response to chemotherapy [129]. Since integrin signaling pathway is also associated with therapy resistance in cancer (see, for reviews, [130,131]) it can be assumed that RNASET2 may act on integrin signaling pathway, not only influencing FA dynamics, cell motility and adhesion on ECM, but also influencing downstream PI3K/AKT pathways and, in turn, drug resistance.

4. Targeting Cell Adhesion Using Peptidomimetic Ligands

Besides the use of specific antibodies, in the last few years, the possibility of designing small molecules acting as antagonists or agonists of the interactions between cell/ECM or cell/cell and the consequent intracellular signalings has been developed. Peptidomimetic integrin ligands have proved to be valuable for tumor directed delivery of diagnostics or therapeutics [132–135]. A subset of integrins (8 out of 24) recognizes RGD sequence and several RGD peptide mimetics entered in clinical trials as integrin-targeted agents in EOC [136]. Integrin ligands tested in tumor cells are summarized in Table 1. The cilengitide, designed against the $\alpha v\beta 3$, has been considered the most promising compound; however, the phase III clinical trials failed likely due to a low specificity, metabolic instability and low biodistribution [137,138]. To accomplish an adequate biodistribution and uptake in cancer cells, new compounds were obtained by the conjugation between the peptidomimetic ligands and a drug or a nanoparticle made of new materials (see, for review, [139]).

As far as the cadherins are concerned, some short peptide sequences, corresponding to the binding domain of N-cadh, have been shown to disrupt cell adhesion-induced apoptosis in an antagonist manner [140] (Table 1). In cancer cells in vitro and in xenotransplanted animal models, one of these small peptides, the ADH-1, enhanced antitumor activity of melphalan-treated melanomas [141] by altering both homotypic (between cancer cells) and heterotypic (between cancer cells and surrounding endothelial cells) cadherin interactions. Although ADH-1 has been well tolerated in phase I and II clinical trials, its antitumor efficacy as a single agent has been moderate [142,143]. This is probably due to its low affinity for N-cadh and the lack of stability in biological fluids. Very recently, Turley et al. assessed that ADH-1 may have a dichotomous effect: it can increase tumor growth rate and sensitivity to some chemotherapy agents by increasing AKT activation and, on the other side, ADH-1 facilitates drug delivery, enhancing the vascular permeability [144]. More recently, some peptidomimetics able to bind at the sites of the homophilic N- and E-cadh binding have been shown to inhibit cell–cell adhesion [145] and E-cadherin-mediated AKT phosphorylation of EOC cells [146].

Table 1. Integrins or cadherins expressed on EOC cells and their suitable ligands.

Integrins	Ligands	Chemical Scaffold	Tumor Cell Model ¹	Reference
$\alpha v\beta 3$	cRGDFV ²	Cyclopentapeptide	GBM	[147]
	RGD4C	Cyclopentapeptide	BC	[148]
	cAbaRGD	Azabicycloalkane	EOC	[149]
	(DKP)-RGD	Dichetopiperazine	EOC	[135]
	CisoDGR	CDAK 22-mer peptide	BC	[150]
	Cyclo[DKP-isoDGR]	Dichetopiperazine/CDAK	GBM	[151]
$\alpha 5\beta 3$	H2009.1	20-mer peptide	NSCL-C	[152]
Cadherins				
N-cadh	N-Ac-CHAVC-NH ₂ ³	Disulphate-linked cyclic peptide	PC	[153]
N- and E-cadh	Compound 3	Benzyl ring	EOC	[145]

¹ Only ligands tested on cancer cells are reported. The abbreviations are: GBM, Glioblastoma; BC, Breast Cancer; NSCL-C Non-Small Cell Lung cancer; PC, Pancreatic cancer; ² Cilengitide (Merck, Darmstadt, Germany). It also binds to $\alpha 5\beta 3$ integrin; ³ ADH-1 or Exherin.

5. Conclusions

The present review has reported the more recent knowledge on the cooperation of cell–cell and cell–ECM adhesion molecules, such as cadherins and integrins, as well as their elicited mechanisms, together with membrane or cytoplasmic signaling molecules that are required for the growth, migration and invasion of EOC cells, especially taking into account those mechanisms necessary for EOC intra-peritoneal dissemination. The reported information suggests that different cadherins and integrins could affect several aspects of EOC progression. For example, E-cadh is important in spheroid formation, but, at the late stage, the switch in cadherins expression could enhance metastasis through a different pattern of integrins at the plasma membrane.

Recent evidence shows that mono-targeting, particularly of RTKs, inefficiently impacts tumor growth control, suggesting that a multi-target treatment might be more efficient. As we have shown here, RTKs might require specific integrins for full activation, and for their contribution to tumorigenesis and metastasis formation, it may be plausible to tailor the use of integrin antagonists in combination with RTK inhibitors. For instance, inhibitors to adhesion molecules together with target-specific drugs could be effective as adjuvant therapy in advanced-stage EOC patients. Peptidomimetics to E-cadh could be relevant for EOC relapsing patients presenting E-cadh-expressing MCAs, and labelled E-cadh binding compound/s could be exploited for the detection of minimal residual disease after first debulking or during chemotherapeutic treatment. Overall, these new approaches, aimed to inhibit adhesion-dependent mechanisms, represent a challenge in the field of cancer research likely to counteract the processes of mesothelial cell clearance and MCA formation occurring during the intra-peritoneal dissemination of EOC cells. Investigations through large scale/high-throughput technologies could help to identify further relevant cross-talks that involve adhesion molecules.

Further efforts are necessary for a better understanding of the different roles exerted by the numerous players participating in these complex mechanisms for a more rational development of better treatments in the aim to improve EOC patient outcome.

Acknowledgments: The authors acknowledge Anna Maria Invernizzi for her collaboration in the development of the projects in the EOC field. This work was supported by the Associazione Italiana per La Ricerca sul Cancro (AIRC, IG13055 to Antonella Tomassetti and IG17475 to Delia Mezzanzanica), by the Italian Ministry of Health ('Characterization of the molecular basis of tumor progression: a focus on ovarian cancer' Ricerca Corrente 2013–2016 to Antonella Tomassetti) and the Cariplo Foundation (2013-0865 to Delia Mezzanzanica).

Author Contributions: All authors contributed equally to the preparation of the manuscript.

Conflicts of Interest: The authors declare no conflict of interest.

References

1. Jelovac, D.; Armstrong, D.K. Recent progress in the diagnosis and treatment of ovarian cancer. *CA Cancer J. Clin.* **2011**, *61*, 183–203. [[CrossRef](#)] [[PubMed](#)]
2. Baldwin, L.A.; Huang, B.; Miller, R.W.; Tucker, T.; Goodrich, S.T.; Podzielinski, I.; DeSimone, C.P.; Ueland, F.R.; van Nagell, J.R.; Seamon, L.G. Ten-year relative survival for epithelial ovarian cancer. *Obstet. Gynecol.* **2012**, *120*, 612–618. [[CrossRef](#)] [[PubMed](#)]
3. Jayson, G.C.; Kohn, E.C.; Kitchener, H.C.; Ledermann, J.A. Ovarian cancer. *Lancet* **2014**, *384*, 1376–1388. [[CrossRef](#)]
4. Vaughan, S.; Coward, J.I.; Bast, R.C., Jr.; Berchuck, A.; Berek, J.S.; Brenton, J.D.; Coukos, G.; Crum, C.C.; Drapkin, R.; Etemadmoghadam, D.; et al. Rethinking ovarian cancer: Recommendations for improving outcomes. *Nat. Rev. Cancer* **2011**, *11*, 719–725. [[CrossRef](#)] [[PubMed](#)]
5. Tothill, R.W.; Tinker, A.V.; George, J.; Brown, R.; Fox, S.B.; Lade, S.; Johnon, D.S.; Trivett, M.K.; Etemadmoghadam, D.; Locandro, B.; et al. Novel molecular subtypes of serous and endometrioid ovarian cancer linked to clinical outcome. *Clin. Cancer Res.* **2008**, *14*, 5198–5208. [[CrossRef](#)] [[PubMed](#)]
6. The Cancer Genome Atlas Database. 2012. Available online: <https://tcga-data.nci.nih.gov/> (accessed on 30 June 2011).

7. Sieh, W.; Salvador, S.; McGuire, V.; Weber, R.P.; Terry, K.L.; Rossing, M.A.; Risch, H.; Wu, A.H.; Webb, P.M.; Moysich, K.; et al. Tubal ligation and risk of ovarian cancer subtypes: A pooled analysis of case-control studies. *Int. J. Epidemiol.* **2013**, *42*, 579–589. [[CrossRef](#)] [[PubMed](#)]
8. Munksgaard, P.S.; Blaakaer, J. The association between endometriosis and ovarian cancer: A review of histological, genetic and molecular alterations. *Gynecol. Oncol.* **2012**, *124*, 164–169. [[CrossRef](#)] [[PubMed](#)]
9. Kurman, R.J.; Shih, I.M. Pathogenesis of ovarian cancer: Lessons from morphology and molecular biology and their clinical implications. *Int. J. Gynecol. Pathol.* **2008**, *27*, 151–160. [[CrossRef](#)] [[PubMed](#)]
10. Zorn, K.K.; Bonome, T.; Gangi, L.; Chandramouli, G.V.; Awtrey, C.S.; Gardner, G.J.; Barrett, J.C.; Boyd, J.; Birrer, M.J. Gene expression profiles of serous, endometrioid, and clear cell subtypes of ovarian and endometrial cancer. *Clin. Cancer Res.* **2005**, *11*, 6422–6430. [[CrossRef](#)] [[PubMed](#)]
11. Bowtell, D.D.; Bohm, S.; Ahmed, A.A.; Aspuria, P.J.; Bast, R.C., Jr.; Beral, V.; Berek, J.S.; Birrer, M.J.; Blagden, S.; Bookman, M.A.; et al. Rethinking ovarian cancer II: Reducing mortality from high-grade serous ovarian cancer. *Nat. Rev. Cancer* **2015**, *15*, 668–679. [[CrossRef](#)] [[PubMed](#)]
12. Kipps, E.; Tan, D.S.; Kaye, S.B. Meeting the challenge of ascites in ovarian cancer: New avenues for therapy and research. *Nat. Rev. Cancer* **2013**, *13*, 273–282. [[CrossRef](#)] [[PubMed](#)]
13. Ahmed, N.; Stenvers, K.L. Getting to know ovarian cancer ascites: Opportunities for targeted therapy-based translational research. *Front. Oncol.* **2013**, *3*, 256. [[CrossRef](#)] [[PubMed](#)]
14. Tan, D.S.; Agarwal, R.; Kaye, S.B. Mechanisms of transcoelomic metastasis in ovarian cancer. *Lancet Oncol.* **2006**, *7*, 925–934. [[CrossRef](#)]
15. Freedman, R.S.; Deavers, M.; Liu, J.; Wang, E. Peritoneal inflammation—A microenvironment for Epithelial Ovarian Cancer (EOC). *J. Transl. Med.* **2004**, *2*, 23. [[CrossRef](#)] [[PubMed](#)]
16. Pradeep, S.; Kim, S.W.; Wu, S.Y.; Nishimura, M.; Chaluvally-Raghavan, P.; Miyake, T.; Pecot, C.V.; Kim, S.J.; Choi, H.J.; Bischoff, F.Z.; et al. Hematogenous metastasis of ovarian cancer: Rethinking mode of spread. *Cancer Cell* **2014**, *26*, 77–91. [[CrossRef](#)] [[PubMed](#)]
17. Birbeck, M.S.; Wheatley, D.N. An electron microscopy study of the invasion of ascites tumor cells into the abdominal wall. *Cancer Res.* **1965**, *25*, 490–497. [[PubMed](#)]
18. Shield, K.; Riley, C.; Quinn, M.A.; Rice, G.E.; Ackland, M.L.; Ahmed, N. $\alpha 2\beta 1$ Integrin affects metastatic potential of ovarian carcinoma spheroids by supporting disaggregation and proteolysis. *J. Carcinog.* **2007**, *6*, 11. [[CrossRef](#)] [[PubMed](#)]
19. Burleson, K.M.; Hansen, L.K.; Skubitz, A.P. Ovarian carcinoma spheroids disaggregate on type I collagen and invade live human mesothelial cell monolayers. *Clin. Exp. Metastasis* **2004**, *21*, 685–697. [[CrossRef](#)] [[PubMed](#)]
20. Iwanicki, M.P.; Davidowitz, R.A.; Ng, M.R.; Besser, A.; Muranen, T.; Merritt, M.; Danuser, G.; Ince, T.A.; Brugge, J.S. Ovarian cancer spheroids use myosin-generated force to clear the mesothelium. *Cancer Discov.* **2011**, *1*, 144–157. [[CrossRef](#)] [[PubMed](#)]
21. Barbolina, M.V.; Moss, N.M.; Westfall, S.D.; Liu, Y.; Burkhalter, R.J.; Marga, F.; Forgacs, G.; Hudson, L.G.; Stack, M.S. Microenvironmental regulation of ovarian cancer metastasis. *Cancer Treat. Res.* **2009**, *149*, 319–334. [[PubMed](#)]
22. Kim, G.; Davidson, B.; Henning, R.; Wang, J.; Yu, M.; Annunziata, C.; Hetland, T.; Kohn, E.C. Adhesion molecule protein signature in ovarian cancer effusions is prognostic of patient outcome. *Cancer* **2012**, *118*, 1543–1553. [[CrossRef](#)] [[PubMed](#)]
23. Takeichi, M. Dynamic contacts: Rearranging adherens junctions to drive epithelial remodelling. *Nat. Rev. Mol. Cell Biol.* **2014**, *15*, 397–410. [[CrossRef](#)] [[PubMed](#)]
24. Shapiro, L.; Fannon, A.M.; Kwong, P.D.; Thompson, A.; Lehmann, M.S.; Grübel, G.; Legrand, J.-F.; Als-Nielsen, J.; Colman, D.R.; Hendrickson, W.A. Structural basis of cell-cell adhesion by cadherins. *Nature* **1995**, *374*, 327–337. [[CrossRef](#)] [[PubMed](#)]
25. Kovacs, E.M.; Ali, R.G.; McCormack, A.J.; Yap, A.S. E-cadherin homophilic ligation directly signals through Rac and phosphatidylinositol 3-kinase to regulate adhesive contacts. *J. Biol. Chem.* **2002**, *277*, 6708–6718. [[CrossRef](#)] [[PubMed](#)]
26. Kraemer, A.; Goodwin, M.; Verma, S.; Yap, A.S.; Ali, R.G. Rac is a dominant regulator of cadherin-directed actin assembly that is activated by adhesive ligation independently of Tiam1. *Am. J. Physiol. Cell Physiol.* **2007**, *292*, C1061–C1069. [[CrossRef](#)] [[PubMed](#)]

27. Nakagawa, M.; Fukata, M.; Yamaga, M.; Itoh, N.; Kaibuchi, K. Recruitment and activation of Rac1 by the formation of E-cadherin-mediated cell-cell adhesion sites. *J. Cell Sci.* **2001**, *114*, 1829–1838. [[PubMed](#)]
28. Yamada, S.; Nelson, W.J. Localized zones of Rho and Rac activities drive initiation and expansion of epithelial cell-cell adhesion. *J. Cell Biol.* **2007**, *178*, 517–527. [[CrossRef](#)] [[PubMed](#)]
29. Kim, S.H.; Li, Z.; Sacks, D.B. E-cadherin-mediated cell-cell attachment activates Cdc42. *J. Biol. Chem.* **2000**, *275*, 36999–37005. [[CrossRef](#)] [[PubMed](#)]
30. Fukata, M.; Kaibuchi, K. Rho-family GTPases in cadherin-mediated cell-cell adhesion. *Nat. Rev. Mol. Cell Biol.* **2001**, *2*, 887–897. [[CrossRef](#)] [[PubMed](#)]
31. Wheelock, M.J.; Johnson, K.R. Cadherin-mediated cellular signaling. *Curr. Opin. Cell Biol.* **2003**, *15*, 509–514. [[CrossRef](#)]
32. Oda, H.; Takeichi, M. Evolution: Structural and functional diversity of cadherin at the adherens junction. *J. Cell Biol.* **2011**, *193*, 1137–1146. [[CrossRef](#)] [[PubMed](#)]
33. Miow, Q.H.; Tan, T.Z.; Ye, J.; Lau, J.A.; Yokomizo, T.; Thiery, J.P.; Mori, S. Epithelial-mesenchymal status renders differential responses to cisplatin in ovarian cancer. *Oncogene* **2014**, *34*, 1899–1907. [[CrossRef](#)] [[PubMed](#)]
34. Hanahan, D.; Weinberg, R.A. Hallmarks of cancer: The next generation. *Cell* **2011**, *144*, 646–674. [[CrossRef](#)] [[PubMed](#)]
35. Thiery, J.P.; Lim, C.T. Tumor dissemination: An EMT affair. *Cancer Cell* **2013**, *23*, 272–273. [[CrossRef](#)] [[PubMed](#)]
36. Gheldof, A.; Berx, G. Cadherins and epithelial-to-mesenchymal transition. *Prog. Mol. Biol. Transl. Sci.* **2013**, *116*, 317–336. [[PubMed](#)]
37. Wong, A.S.; Gumbiner, B.M. Adhesion-independent mechanism for suppression of tumor cell invasion by E-cadherin. *J. Cell Biol.* **2003**, *161*, 1191–1203. [[CrossRef](#)] [[PubMed](#)]
38. Qian, X.; Karpova, T.; Sheppard, A.M.; McNally, J.; Lowy, D.R. E-cadherin-mediated adhesion inhibits ligand-dependent activation of diverse receptor tyrosine kinases. *EMBO J.* **2004**, *23*, 1739–1748. [[CrossRef](#)] [[PubMed](#)]
39. Andl, C.D.; Rustgi, A.K. No one-way street: Cross-talk between e-cadherin and receptor tyrosine kinase (RTK) signaling: A mechanism to regulate RTK activity. *Cancer Biol. Ther.* **2005**, *4*, 28–31. [[CrossRef](#)] [[PubMed](#)]
40. Fischer, K.R.; Durrans, A.; Lee, S.; Sheng, J.; Li, F.; Wong, S.T.; Choi, H.; El, R.T.; Ryu, S.; Troeger, J.; et al. Epithelial-to-mesenchymal transition is not required for lung metastasis but contributes to chemoresistance. *Nature* **2015**, *527*, 472–476. [[CrossRef](#)] [[PubMed](#)]
41. Zheng, X.; Carstens, J.L.; Kim, J.; Scheible, M.; Kaye, J.; Sugimoto, H.; Wu, C.C.; LeBleu, V.S.; Kalluri, R. Epithelial-to-mesenchymal transition is dispensable for metastasis but induces chemoresistance in pancreatic cancer. *Nature* **2015**, *527*, 525–530. [[CrossRef](#)] [[PubMed](#)]
42. Bertos, N.R.; Park, M. Breast cancer—One term, many entities? *J. Clin. Investig.* **2011**, *121*, 3789–3796. [[CrossRef](#)] [[PubMed](#)]
43. Colpaert, C.G.; Vermeulen, P.B.; Benoy, I.; Soubry, A.; Van, R.F.; van, B.P.; Goovaerts, G.; Dirix, L.Y.; van, D.P.; Fox, S.B.; et al. Inflammatory breast cancer shows angiogenesis with high endothelial proliferation rate and strong E-cadherin expression. *Br. J. Cancer* **2003**, *88*, 718–725. [[CrossRef](#)] [[PubMed](#)]
44. Cheung, K.J.; Gabrielson, E.; Werb, Z.; Ewald, A.J. Collective invasion in breast cancer requires a conserved basal epithelial program. *Cell* **2013**, *155*, 1639–1651. [[CrossRef](#)] [[PubMed](#)]
45. Nguyen-Ngoc, K.V.; Cheung, K.J.; Brenot, A.; Shamir, E.R.; Gray, R.S.; Hines, W.C.; Yaswen, P.; Werb, Z.; Ewald, A.J. ECM microenvironment regulates collective migration and local dissemination in normal and malignant mammary epithelium. *Proc. Natl. Acad. Sci. USA* **2012**, *109*, E2595–E2604. [[CrossRef](#)] [[PubMed](#)]
46. Darai, E.; Scoazec, J.Y.; Walker-Combrouze, F.; Mlika-Cabanne, N.; Feldmann, G.; Madelenat, P.; Potet, F. Expression of cadherins in benign, borderline, and malignant ovarian epithelial tumors: A clinicopathologic study of 60 cases. *Hum. Pathol.* **1997**, *28*, 922–928. [[CrossRef](#)]
47. Davies, B.R.; Worsley, S.D.; Ponder, B.A. Expression of E-cadherin, α -catenin and β -catenin in normal ovarian surface epithelium and epithelial ovarian cancers. *Histopathology* **1998**, *32*, 69–80. [[CrossRef](#)] [[PubMed](#)]
48. Sundfeldt, K.; Piontkewitz, Y.; Ivarsson, K.; Nilsson, O.; Hellberg, P.; Brannstrom, M.; Janson, P.O.; Enerback, S.; Hedin, L. E-cadherin expression in human epithelial ovarian cancer and normal ovary. *Int. J. Cancer* **1997**, *74*, 275–280. [[CrossRef](#)]

49. Imai, T.; Horiuchi, A.; Shiozawa, T.; Osada, R.; Kikuchi, N.; Ohira, S.; Oka, K.; Konishi, I. Elevated expression of E-cadherin and α -, β -, and γ -catenins in metastatic lesions compared with primary epithelial ovarian carcinomas. *Hum. Pathol.* **2004**, *35*, 1469–1476. [[CrossRef](#)] [[PubMed](#)]
50. Hsia, D.A.; Mitra, S.K.; Hauck, C.R.; Strebblow, D.N.; Nelson, J.A.; Ilic, D.; Huang, S.; Li, E.; Nemerow, G.R.; Leng, J.; et al. Differential regulation of cell motility and invasion by FAK. *J. Cell Biol.* **2003**, *160*, 753–767. [[CrossRef](#)] [[PubMed](#)]
51. Tomassetti, A.; de Santis, G.; Castellano, G.; Miotti, S.; Mazzi, M.; Tomasoni, D.; van Roy, F.; Carcangiu, M.L.; Canevari, S. Variant HNF1 modulates epithelial plasticity of normal and transformed ovary cells. *Neoplasia* **2008**, *10*, 1481–1492. [[CrossRef](#)] [[PubMed](#)]
52. Strauss, R.; Li, Z.Y.; Liu, Y.; Beyers, I.; Persson, J.; Sova, P.; Moller, T.; Pesonen, S.; Hemminki, A.; Hamerlik, P.; et al. Analysis of epithelial and mesenchymal markers in ovarian cancer reveals phenotypic heterogeneity and plasticity. *PLoS ONE* **2011**, *6*, e16186. [[CrossRef](#)]
53. Xu, S.; Yang, Y.; Dong, L.; Qiu, W.; Yang, L.; Wang, X.; Liu, L. Construction and characteristics of an E-cadherin-related three-dimensional suspension growth model of ovarian cancer. *Sci. Rep.* **2014**, *4*, 5646. [[CrossRef](#)] [[PubMed](#)]
54. Reddy, P.; Liu, L.; Ren, C.; Lindgren, P.; Boman, K.; Shen, Y.; Lundin, E.; Ottander, U.; Rytinki, M.; Liu, K. Formation of E-cadherin-mediated cell-cell adhesion activates AKT and mitogen activated protein kinase via phosphatidylinositol 3 kinase and ligand-independent activation of epidermal growth factor receptor in ovarian cancer cells. *Mol. Endocrinol.* **2005**, *19*, 2564–2578. [[CrossRef](#)] [[PubMed](#)]
55. Huber, M.A.; Kraut, N.; Beug, H. Molecular requirements for epithelial-mesenchymal transition during tumor progression. *Curr. Opin. Cell Biol.* **2005**, *17*, 548–558. [[CrossRef](#)] [[PubMed](#)]
56. Dong, L.L.; Liu, L.; Ma, C.H.; Li, J.S.; Du, C.; Xu, S.; Han, L.H.; Li, L.; Wang, X.W. E-cadherin promotes proliferation of human ovarian cancer cells *in vitro* via activating MEK/ERK pathway. *Acta Pharmacol. Sin.* **2012**, *33*, 817–822. [[CrossRef](#)] [[PubMed](#)]
57. Davis, M.A.; Ireton, R.C.; Reynolds, A.B. A core function for p120-catenin in cadherin turnover. *J. Cell Biol.* **2003**, *163*, 525–534. [[CrossRef](#)] [[PubMed](#)]
58. Maretzky, T.; Reiss, K.; Ludwig, A.; Buchholz, J.; Scholz, F.; Proksch, E.; de Strooper, B.; Hartmann, D.; Saftig, P. ADAM10 mediates E-cadherin shedding and regulates epithelial cell-cell adhesion, migration, and β -catenin translocation. *Proc. Natl. Acad. Sci. USA* **2005**, *102*, 9182–9187. [[CrossRef](#)] [[PubMed](#)]
59. Noe, V.; Fingleton, B.; Jacobs, K.; Crawford, H.C.; Vermeulen, S.; Steelant, W.; Bruyneel, E.; Matrisian, L.M.; Mareel, M. Release of an invasion promoter E-cadherin fragment by matrilysin and stromelysin-1. *J. Cell Sci.* **2001**, *114*, 111–118. [[PubMed](#)]
60. Marambaud, P.; Shioi, J.; Serban, G.; Georgakopoulos, A.; Sarner, S.; Nagy, V.; Baki, L.; Wen, P.; Efthimiopoulos, S.; Shao, Z.; et al. A presenilin-1/ γ -secretase cleavage releases the E-cadherin intracellular domain and regulates disassembly of adherens junctions. *EMBO J.* **2002**, *21*, 1948–1956. [[CrossRef](#)] [[PubMed](#)]
61. Gogali, A.; Charalabopoulos, K.; Zampira, I.; Konstantinidis, A.K.; Tachmazoglou, F.; Daskalopoulos, G.; Constantopoulos, S.H.; Dalavanga, Y. Soluble adhesion molecules E-cadherin, intercellular adhesion molecule-1, and E-selectin as lung cancer biomarkers. *Chest* **2010**, *138*, 1173–1179. [[CrossRef](#)] [[PubMed](#)]
62. Cowden Dahl, K.D.; Symowicz, J.; Ning, Y.; Gutierrez, E.; Fishman, D.A.; Adley, B.P.; Stack, M.S.; Hudson, L.G. Matrix metalloproteinase 9 is a mediator of epidermal growth factor-dependent E-cadherin loss in ovarian carcinoma cells. *Cancer Res.* **2008**, *68*, 4606–4613. [[CrossRef](#)] [[PubMed](#)]
63. Symowicz, J.; Adley, B.P.; Gleason, K.J.; Johnson, J.J.; Ghosh, S.; Fishman, D.A.; Hudson, L.G.; Stack, M.S. Engagement of collagen-binding integrins promotes matrix metalloproteinase-9-dependent E-cadherin ectodomain shedding in ovarian carcinoma cells. *Cancer Res.* **2007**, *67*, 2030–2039. [[CrossRef](#)] [[PubMed](#)]
64. Trillsch, F.; Kuerti, S.; Eulenburg, C.; Burandt, E.; Woelber, L.; Prieske, K.; Eylmann, K.; Oliveira-Ferrer, L.; Milde-Langosch, K.; Mahner, S. E-Cadherin fragments as potential mediators for peritoneal metastasis in advanced epithelial ovarian cancer. *Br. J. Cancer* **2016**, *114*, 213–220. [[CrossRef](#)] [[PubMed](#)]
65. De Boer, C.J.; van, D.E.; van, K.H.; van Jansen Rhijn, C.M.; Warnaar, S.O.; Fleuren, G.J.; Litvinov, S.V. Changing roles of cadherins and catenins during progression of squamous intraepithelial lesions in the uterine cervix. *Am. J. Pathol.* **1999**, *155*, 505–515. [[CrossRef](#)]
66. Bailey, T.; Biddlestone, L.; Shepherd, N.; Barr, H.; Warner, P.; Jankowski, J. Altered cadherin and catenin complexes in the Barrett's esophagus-dysplasia-adenocarcinoma sequence: Correlation with disease progression and dedifferentiation. *Am. J. Pathol.* **1998**, *152*, 135–144. [[PubMed](#)]

67. Peralta, S.A.; Knudsen, K.A.; Salazar, H.; Han, A.C.; Keshgegian, A.A. P-cadherin expression in breast carcinoma indicates poor survival. *Cancer* **1999**, *86*, 1263–1272.
68. Hardy, R.G.; Tselepis, C.; Hoyland, J.; Wallis, Y.; Pretlow, T.P.; Talbot, I.; Sanders, D.S.; Matthews, G.; Morton, D.; Jankowski, J.A. Aberrant P-cadherin expression is an early event in hyperplastic and dysplastic transformation in the colon. *Gut* **2002**, *50*, 513–519. [[CrossRef](#)] [[PubMed](#)]
69. Patel, I.S.; Madan, P.; Getsios, S.; Bertrand, M.A.; MacCalman, C.D. Cadherin switching in ovarian cancer progression. *Int. J. Cancer* **2003**, *106*, 172–177. [[CrossRef](#)] [[PubMed](#)]
70. Cheung, L.W.; Leung, P.C.; Wong, A.S. Cadherin switching and activation of p120 catenin signaling are mediators of gonadotropin-releasing hormone to promote tumor cell migration and invasion in ovarian cancer. *Oncogene* **2010**, *29*, 2427–2440. [[CrossRef](#)] [[PubMed](#)]
71. Cheung, L.W.; Mak, A.S.; Cheung, A.N.; Ngan, H.Y.; Leung, P.C.; Wong, A.S. P-cadherin cooperates with insulin-like growth factor-1 receptor to promote metastatic signaling of gonadotropin-releasing hormone in ovarian cancer via p120 catenin. *Oncogene* **2011**, *30*, 2964–2974. [[CrossRef](#)] [[PubMed](#)]
72. Cheung, L.W.; Yung, S.; Chan, T.M.; Leung, P.C.; Wong, A.S. Targeting gonadotropin-releasing hormone receptor inhibits the early step of ovarian cancer metastasis by modulating tumor-mesothelial adhesion. *Mol. Ther.* **2013**, *21*, 78–90. [[CrossRef](#)] [[PubMed](#)]
73. Usui, A.; Ko, S.Y.; Barengo, N.; Naora, H. P-cadherin promotes ovarian cancer dissemination through tumor cell aggregation and tumor-peritoneum interactions. *Mol. Cancer Res.* **2014**, *12*, 504–513. [[CrossRef](#)] [[PubMed](#)]
74. Ip, C.K.; Yung, S.; Chan, T.M.; Tsao, S.W.; Wong, A.S. p70 S6 kinase drives ovarian cancer metastasis through multicellular spheroid-peritoneum interaction and P-cadherin/ β 1 integrin signaling activation. *Oncotarget* **2014**, *5*, 9133–9149. [[PubMed](#)]
75. Barczyk, M.; Carracedo, S.; Gullberg, D. Integrins. *Cell Tissue Res.* **2010**, *339*, 269–280. [[CrossRef](#)]
76. Ruoslahti, E. RGD and other recognition sequences for integrins. *Annu. Rev. Cell Biol.* **1996**, *12*, 697–715. [[CrossRef](#)] [[PubMed](#)]
77. Zhao, X.; Guan, J.L. Focal adhesion kinase and its signaling pathways in cell migration and angiogenesis. *Adv. Drug Deliv. Rev.* **2011**, *63*, 610–615. [[CrossRef](#)] [[PubMed](#)]
78. Schlaepfer, D.D.; Hanks, S.K.; Hunter, T.; van der, G.P. Integrin-mediated signal transduction linked to Ras pathway by GRB2 binding to focal adhesion kinase. *Nature* **1994**, *372*, 786–791. [[CrossRef](#)] [[PubMed](#)]
79. Mitra, S.K.; Schlaepfer, D.D. Integrin-regulated FAK-Src signaling in normal and cancer cells. *Curr. Opin. Cell Biol.* **2006**, *18*, 516–523. [[CrossRef](#)] [[PubMed](#)]
80. Heyman, L.; Leroy-Dudal, J.; Fernandes, J.; Seyer, D.; Dutoit, S.; Carreiras, F. Mesothelial vitronectin stimulates migration of ovarian cancer cells. *Cell Biol. Int.* **2010**, *34*, 493–502. [[CrossRef](#)] [[PubMed](#)]
81. Sawada, K.; Mitra, A.K.; Radjabi, A.R.; Bhaskar, V.; Kistner, E.O.; Tretiakova, M.; Jagadeeswaran, S.; Montag, A.; Becker, A.; Kenny, H.A.; et al. Loss of E-cadherin promotes ovarian cancer metastasis via α 5-integrin, which is a therapeutic target. *Cancer Res.* **2008**, *68*, 2329–2339. [[CrossRef](#)] [[PubMed](#)]
82. Ahmed, N.; Pansino, F.; Clyde, R.; Murthi, P.; Quinn, M.A.; Rice, G.E.; Agrez, M.V.; Mok, S.; Baker, M.S. Overexpression of α v β 6 integrin in serous epithelial ovarian cancer regulates extracellular matrix degradation via the plasminogen activation cascade. *Carcinogenesis* **2002**, *23*, 237–244. [[CrossRef](#)] [[PubMed](#)]
83. Carduner, L.; Picot, C.R.; Leroy-Dudal, J.; Blay, L.; Kellouche, S.; Carreiras, F. Cell cycle arrest or survival signaling through α v integrins, activation of PKC and ERK1/2 lead to anoikis resistance of ovarian cancer spheroids. *Exp. Cell Res.* **2014**, *320*, 329–342. [[CrossRef](#)] [[PubMed](#)]
84. Carduner, L.; Leroy-Dudal, J.; Picot, C.R.; Gallet, O.; Carreiras, F.; Kellouche, S. Ascites-induced shift along epithelial-mesenchymal spectrum in ovarian cancer cells: Enhancement of their invasive behavior partly dependant on α v integrins. *Clin. Exp. Metastasis* **2014**, *31*, 675–688. [[CrossRef](#)] [[PubMed](#)]
85. Cho, A.; Howell, V.M.; Colvin, E.K. The Extracellular matrix in epithelial ovarian cancer—A piece of a Puzzle. *Front. Oncol.* **2015**, *5*, 245. [[PubMed](#)]
86. Kenny, H.A.; Chiang, C.Y.; White, E.A.; Schryver, E.M.; Habis, M.; Romero, I.L.; Ladanyi, A.; Penicka, C.V.; George, J.; Matlin, K.; et al. Mesothelial cells promote early ovarian cancer metastasis through fibronectin secretion. *J. Clin. Investig.* **2014**, *124*, 4614–4628. [[CrossRef](#)] [[PubMed](#)]
87. Yousif, N.G. Fibronectin promotes migration and invasion of ovarian cancer cells through up-regulation of FAK-PI3K/Akt pathway. *Cell Biol. Int.* **2014**, *38*, 85–91. [[CrossRef](#)] [[PubMed](#)]

88. Franke, F.E.; Von, G.R.; Zygmunt, M.; Munstedt, K. Association between fibronectin expression and prognosis in ovarian carcinoma. *Anticancer Res.* **2003**, *23*, 4261–4267. [[PubMed](#)]
89. Desgrosellier, J.S.; Cheresh, D.A. Integrins in cancer: Biological implications and therapeutic opportunities. *Nat. Rev. Cancer* **2010**, *10*, 9–22. [[CrossRef](#)] [[PubMed](#)]
90. Ivaska, J.; Heino, J. Cooperation between integrins and growth factor receptors in signaling and endocytosis. *Annu. Rev. Cell Dev. Biol.* **2011**, *27*, 291–320. [[CrossRef](#)] [[PubMed](#)]
91. Regad, T. Targeting RTK signaling pathways in cancer. *Cancers* **2015**, *7*, 1758–1784. [[CrossRef](#)] [[PubMed](#)]
92. Jiao, Y.; Ou, W.; Meng, F.; Zhou, H.; Wang, A. Targeting HSP90 in ovarian cancers with multiple receptor tyrosine kinase coactivation. *Mol. Cancer* **2011**, *10*, 125. [[CrossRef](#)] [[PubMed](#)]
93. Citri, A.; Yarden, Y. EGF-ERBB signalling: Towards the systems level. *Nat. Rev. Mol. Cell Biol.* **2006**, *7*, 505–516. [[CrossRef](#)] [[PubMed](#)]
94. Moro, L.; Venturino, M.; Bozzo, C.; Silengo, L.; Altruda, F.; Beguinot, L.; Tarone, G.; Defilippi, P. Integrins induce activation of EGF receptor: Role in MAP kinase induction and adhesion-dependent cell survival. *EMBO J.* **1998**, *17*, 6622–6632. [[CrossRef](#)] [[PubMed](#)]
95. Mariotti, A.; Kedeshian, P.A.; Dans, M.; Curatola, A.M.; Gagnoux-Palacios, L.; Giancotti, F.G. EGF-R signaling through Fyn kinase disrupts the function of integrin $\alpha 6 \beta 4$ at hemidesmosomes: Role in epithelial cell migration and carcinoma invasion. *J. Cell Biol.* **2001**, *155*, 447–458. [[CrossRef](#)] [[PubMed](#)]
96. Moro, L.; Dolce, L.; Cabodi, S.; Bergatto, E.; Boeri, E.E.; Smeriglio, M.; Turco, E.; Retta, S.F.; Giuffrida, M.G.; Venturino, M.; et al. Integrin-induced epidermal growth factor (EGF) receptor activation requires c-Src and p130Cas and leads to phosphorylation of specific EGF receptor tyrosines. *J. Biol. Chem.* **2002**, *277*, 9405–9414. [[CrossRef](#)] [[PubMed](#)]
97. Cabodi, S.; Moro, L.; Bergatto, E.; Boeri, E.E.; Di, S.P.; Turco, E.; Tarone, G.; Defilippi, P. Integrin regulation of epidermal growth factor (EGF) receptor and of EGF-dependent responses. *Biochem. Soc. Trans.* **2004**, *32*, 438–442. [[CrossRef](#)] [[PubMed](#)]
98. Gui, T.; Shen, K. The epidermal growth factor receptor as a therapeutic target in epithelial ovarian cancer. *Cancer Epidemiol.* **2012**, *36*, 490–496. [[CrossRef](#)] [[PubMed](#)]
99. Lafky, J.M.; Wilken, J.A.; Baron, A.T.; Maihle, N.J. Clinical implications of the ErbB/epidermal growth factor (EGF) receptor family and its ligands in ovarian cancer. *Biochim. Biophys. Acta* **2008**, *1785*, 232–265. [[CrossRef](#)] [[PubMed](#)]
100. Alberti, C.; Pinciroli, P.; Valeri, B.; Ferri, R.; Ditto, A.; Umezawa, K.; Sensi, M.; Canevari, S.; Tomassetti, A. Ligand-dependent EGFR activation induces the co-expression of IL-6 and PAI-1 via the NF κ B pathway in advanced-stage epithelial ovarian cancer. *Oncogene* **2012**, *31*, 4139–4149. [[CrossRef](#)] [[PubMed](#)]
101. Alper, O.; Bergmann-Leitner, E.S.; Bennett, T.A.; Hacker, N.F.; Stromberg, K.; Stetler-Stevenson, W.G. Epidermal growth factor receptor signaling and the invasive phenotype of ovarian carcinoma cells. *J. Natl. Cancer Inst.* **2001**, *93*, 1375–1384. [[CrossRef](#)] [[PubMed](#)]
102. Ning, Y.; Zeineldin, R.; Liu, Y.; Rosenberg, M.; Stack, M.S.; Hudson, L.G. Down-regulation of integrin $\alpha 2$ surface expression by mutant epidermal growth factor receptor (EGFRvIII) induces aberrant cell spreading and focal adhesion formation. *Cancer Res.* **2005**, *65*, 9280–9286. [[CrossRef](#)] [[PubMed](#)]
103. Comoglio, P.M. Pathway specificity for Met signalling. *Nat. Cell Biol.* **2001**, *3*, E161–E162. [[CrossRef](#)] [[PubMed](#)]
104. Sawada, K.; Radjabi, A.R.; Shinomiya, N.; Kistner, E.; Kenny, H.; Becker, A.R.; Turkyilmaz, M.A.; Salgia, R.; Yamada, S.D.; Vande Woude, G.F.; et al. c-Met overexpression is a prognostic factor in ovarian cancer and an effective target for inhibition of peritoneal dissemination and invasion. *Cancer Res.* **2007**, *67*, 1670–1679. [[CrossRef](#)] [[PubMed](#)]
105. Mitra, A.K.; Sawada, K.; Tiwari, P.; Mui, K.; Gwin, K.; Lengyel, E. Ligand-independent activation of c-Met by fibronectin and $\alpha 5 \beta 1$ -integrin regulates ovarian cancer invasion and metastasis. *Oncogene* **2011**, *30*, 1566–1576. [[CrossRef](#)] [[PubMed](#)]
106. Kwon, Y.; Smith, B.D.; Zhou, Y.; Kaufman, M.D.; Godwin, A.K. Effective inhibition of c-MET-mediated signaling, growth and migration of ovarian cancer cells is influenced by the ovarian tissue microenvironment. *Oncogene* **2015**, *34*, 144–153. [[CrossRef](#)] [[PubMed](#)]

107. Moran-Jones, K.; Brown, L.M.; Samimi, G. INC280, an orally available small molecule inhibitor of c-MET, reduces migration and adhesion in ovarian cancer cell models. *Sci. Rep.* **2015**, *5*, 11749. [[CrossRef](#)] [[PubMed](#)]
108. Ferrara, N.; Hillan, K.J.; Gerber, H.P.; Novotny, W. Discovery and development of bevacizumab, an anti-VEGF antibody for treating cancer. *Nat. Rev. Drug Discov.* **2004**, *3*, 391–400. [[CrossRef](#)] [[PubMed](#)]
109. Mu, J.; Abe, Y.; Tsutsui, T.; Yamamoto, N.; Tai, X.G.; Niwa, O.; Tsujimura, T.; Sato, B.; Terano, H.; Fujiwara, H.; et al. Inhibition of growth and metastasis of ovarian carcinoma by administering a drug capable of interfering with vascular endothelial growth factor activity. *Jpn. J. Cancer Res.* **1996**, *87*, 963–971. [[CrossRef](#)] [[PubMed](#)]
110. Yamamoto, S.; Konishi, I.; Mandai, M.; Kuroda, H.; Komatsu, T.; Nanbu, K.; Sakahara, H.; Mori, T. Expression of vascular endothelial growth factor (VEGF) in epithelial ovarian neoplasms: Correlation with clinicopathology and patient survival, and analysis of serum VEGF levels. *Br. J. Cancer* **1997**, *76*, 1221–1227. [[CrossRef](#)] [[PubMed](#)]
111. Serini, G.; Napione, L.; Arese, M.; Bussolino, F. Besides adhesion: New perspectives of integrin functions in angiogenesis. *Cardiovasc. Res.* **2008**, *78*, 213–222. [[CrossRef](#)] [[PubMed](#)]
112. Kim, T.J.; Landen, C.N.; Lin, Y.G.; Mangala, L.S.; Lu, C.; Nick, A.M.; Stone, R.L.; Merritt, W.M.; rmaiz-Pena, G.; Jennings, N.B.; et al. Combined anti-angiogenic therapy against VEGF and integrin $\alpha v \beta 3$ in an orthotopic model of ovarian cancer. *Cancer Biol. Ther.* **2009**, *8*, 2263–2272. [[CrossRef](#)] [[PubMed](#)]
113. Landen, C.N.; Kim, T.J.; Lin, Y.G.; Merritt, W.M.; Kamat, A.A.; Han, L.Y.; Spannuth, W.A.; Nick, A.M.; Jennings, N.B.; Kinch, M.S.; et al. Tumor-selective response to antibody-mediated targeting of $\alpha v \beta 3$ integrin in ovarian cancer. *Neoplasia* **2008**, *10*, 1259–1267. [[CrossRef](#)] [[PubMed](#)]
114. Brown, M.; Black, J.R.; Sharma, R.; Stebbing, J.; Pinato, D.J. Gene of the month: *Axl*. *J. Clin. Pathol.* **2016**, *69*, 391–397. [[CrossRef](#)] [[PubMed](#)]
115. Zhang, Z.; Lee, J.C.; Lin, L.; Olivás, V.; Au, V.; LaFramboise, T.; bdel-Rahman, M.; Wang, X.; Levine, A.D.; Rho, J.K.; et al. Activation of the AXL kinase causes resistance to EGFR-targeted therapy in lung cancer. *Nat. Genet.* **2012**, *44*, 852–860. [[CrossRef](#)] [[PubMed](#)]
116. Byers, L.A.; Diao, L.; Wang, J.; Saintigny, P.; Girard, L.; Peyton, M.; Shen, L.; Fan, Y.; Giri, U.; Tumula, P.K.; et al. An epithelial-mesenchymal transition gene signature predicts resistance to EGFR and PI3K inhibitors and identifies *Axl* as a therapeutic target for overcoming EGFR inhibitor resistance. *Clin. Cancer Res.* **2013**, *19*, 279–290. [[CrossRef](#)] [[PubMed](#)]
117. Linger, R.M.; Keating, A.K.; Earp, H.S.; Graham, D.K. TAM receptor tyrosine kinases: Biologic functions, signaling, and potential therapeutic targeting in human cancer. *Adv. Cancer Res.* **2008**, *100*, 35–83. [[PubMed](#)]
118. Korshunov, V.A. *Axl*-dependent signalling: A clinical update. *Clin. Sci.* **2012**, *122*, 361–368. [[CrossRef](#)] [[PubMed](#)]
119. Sun, W.; Fujimoto, J.; Tamaya, T. Coexpression of *Gas6/Axl* in human ovarian cancers. *Oncology* **2004**, *66*, 450–457. [[CrossRef](#)] [[PubMed](#)]
120. Buehler, M.; Tse, B.; Leboucq, A.; Jacob, F.; Caduff, R.; Fink, D.; Goldstein, D.R.; Heinzelmann-Schwarz, V. Meta-analysis of microarray data identifies *GAS6* expression as an independent predictor of poor survival in ovarian cancer. *BioMed Res. Int.* **2013**, *2013*, 238284. [[CrossRef](#)] [[PubMed](#)]
121. tRankin, E.B.; Fuh, K.C.; Taylor, T.E.; Krieg, A.J.; Musser, M.; Yuan, J.; Wei, K.; Kuo, C.J.; Longacre, T.A.; Giaccia, A.J. *Axl* is an essential factor and therapeutic target for metastatic ovarian cancer. *Cancer Res.* **2010**, *70*, 7570–7579. [[CrossRef](#)] [[PubMed](#)]
122. Rea, K.; Pinciroli, P.; Sensi, M.; Alciato, F.; Bisaro, B.; Lozneau, L.; Raspagliesi, F.; Centritto, F.; Cabodi, S.; Defilippi, P.; et al. Novel *Axl*-driven signaling pathway and molecular signature characterize high-grade ovarian cancer patients with poor clinical outcome. *Oncotarget* **2015**, *6*, 30859–30875. [[PubMed](#)]
123. Acquati, F.; Morelli, C.; Cinquetti, R.; Bianchi, M.G.; Porrini, D.; Varesco, L.; Gismondi, V.; Rocchetti, R.; Talevi, S.; Possati, L.; et al. Cloning and characterization of a senescence inducing and class II tumor suppressor gene in ovarian carcinoma at chromosome region 6q27. *Oncogene* **2001**, *20*, 980–988. [[CrossRef](#)] [[PubMed](#)]
124. Acquati, F.; Possati, L.; Ferrante, L.; Campomenosi, P.; Talevi, S.; Bardelli, S.; Margiotta, C.; Russo, A.; Bortoletto, E.; Rocchetti, R.; et al. Tumor and metastasis suppression by the human *RNASET2* gene. *Int. J. Oncol.* **2005**, *26*, 1159–1168. [[CrossRef](#)] [[PubMed](#)]

125. Acquati, F.; Lualdi, M.; Bertilaccio, S.; Monti, L.; Turconi, G.; Fabbri, M.; Grimaldi, A.; Anselmo, A.; Inforzato, A.; Collotta, A.; et al. Loss of function of Ribonuclease T2, an ancient and phylogenetically conserved RNase, plays a crucial role in ovarian tumorigenesis. *Proc. Natl. Acad. Sci. USA* **2013**, *110*, 8140–8145. [[CrossRef](#)] [[PubMed](#)]
126. Lualdi, M.; Pedrini, E.; Rea, K.; Monti, L.; Scaldaferrì, D.; Gariboldi, M.; Camporeale, A.; Ghia, P.; Monti, E.; Tomassetti, A.; et al. Pleiotropic modes of action in tumor cells of RNASET2, an evolutionary highly conserved extracellular RNase. *Oncotarget* **2015**, *6*, 7851–7865. [[CrossRef](#)] [[PubMed](#)]
127. Smirnoff, P.; Roiz, L.; Angelkovitch, B.; Schwartz, B.; Shoseyov, O. A recombinant human RNASET2 glycoprotein with antitumorigenic and antiangiogenic characteristics: Expression, purification, and characterization. *Cancer* **2006**, *107*, 2760–2769. [[CrossRef](#)] [[PubMed](#)]
128. Nesiël-Nuttman, L.; Schwartz, B.; Shoseyov, O. Human recombinant truncated RNASET2, devoid of RNase activity; A potential cancer therapeutic agent. *Oncotarget* **2014**, *5*, 11464–11478. [[CrossRef](#)] [[PubMed](#)]
129. Yin, F.; Liu, L.; Liu, X.; Li, G.; Zheng, L.; Li, D.; Wang, Q.; Zhang, W.; Li, L. Downregulation of tumor suppressor gene ribonuclease T2 and gametogenetin binding protein 2 is associated with drug resistance in ovarian cancer. *Oncol. Rep.* **2014**, *32*, 362–372. [[CrossRef](#)] [[PubMed](#)]
130. Eke, I.; Cordes, N. Focal adhesion signaling and therapy resistance in cancer. *Semin. Cancer Biol.* **2015**, *31*, 65–75. [[CrossRef](#)] [[PubMed](#)]
131. Seguin, L.; Desgrosellier, J.S.; Weis, S.M.; Cheresch, D.A. Integrins and cancer: Regulators of cancer stemness, metastasis, and drug resistance. *Trends Cell Biol.* **2015**, *25*, 234–240. [[CrossRef](#)] [[PubMed](#)]
132. Lanzardo, S.; Conti, L.; Brioschi, C.; Bartolomeo, M.P.; Arosio, D.; Belvisi, L.; Manzoni, L.; Maiocchi, A.; Maisano, F.; Forni, G. A new optical imaging probe targeting $\alpha\beta 3$ integrin in glioblastoma xenografts. *Contrast Media Mol. Imaging* **2011**, *6*, 449–458. [[CrossRef](#)] [[PubMed](#)]
133. Manzoni, L.; Belvisi, L.; Arosio, D.; Bartolomeo, M.P.; Bianchi, A.; Brioschi, C.; Buonsanti, F.; Cabella, C.; Casagrande, C.; Civera, M.; et al. Synthesis of Gd and ^{68}Ga complexes in conjugation with a conformationally optimized RGD sequence as potential MRI and PET tumor-imaging probes. *ChemMedChem* **2012**, *7*, 1084–1093. [[CrossRef](#)] [[PubMed](#)]
134. Pilkington-Miksa, M.; Arosio, D.; Battistini, L.; Belvisi, L.; De, M.M.; Vasile, F.; Burreddu, P.; Carta, P.; Rassa, G.; Perego, P.; et al. Design, synthesis, and biological evaluation of novel cRGD-paclitaxel conjugates for integrin-assisted drug delivery. *Bioconjug. Chem.* **2012**, *23*, 1610–1622. [[CrossRef](#)] [[PubMed](#)]
135. Colombo, R.; Mingozzi, M.; Belvisi, L.; Arosio, D.; Piarulli, U.; Carenni, N.; Perego, P.; Zaffaroni, N.; de, C.M.; Castiglioni, V.; et al. Synthesis and biological evaluation (in vitro and in vivo) of cyclic arginine-glycine-aspartate (RGD) peptidomimetic-paclitaxel conjugates targeting integrin $\alpha\beta 3$. *J. Med. Chem.* **2012**, *55*, 10460–10474. [[CrossRef](#)] [[PubMed](#)]
136. Sawada, K.; Ohyagi-Hara, C.; Kimura, T.; Morishige, K. Integrin inhibitors as a therapeutic agent for ovarian cancer. *J. Oncol.* **2012**, *2012*, 915140. [[CrossRef](#)] [[PubMed](#)]
137. Stupp, R.; Hegi, M.E.; Gorlia, T.; Erridge, S.C.; Perry, J.; Hong, Y.K.; Aldape, K.D.; Lhermitte, B.; Pietsch, T.; Grujicic, D.; et al. Cilengitide combined with standard treatment for patients with newly diagnosed glioblastoma with methylated MGMT promoter (CENTRIC EORTC 26071–22072 study): A multicentre, randomised, open-label, phase 3 trial. *Lancet Oncol.* **2014**, *15*, 1100–1108. [[CrossRef](#)]
138. Merck Press Release on Cilengitide Studies. Available online: <http://www.merck.de/de/press/extNewsDetail.html?newsId=C47977D13865FCB9C1257B1D001EF9CA&newsType=1> (accessed on 30 June 2016).
139. Arosio, D.; Casagrande, C. Advancement in integrin facilitated drug delivery. *Adv. Drug Deliv. Rev.* **2016**, *97*, 111–143. [[CrossRef](#)] [[PubMed](#)]
140. Blaschuk, O.W. Discovery and development of N-cadherin antagonists. *Cell Tissue Res.* **2012**, *348*, 309–313. [[CrossRef](#)] [[PubMed](#)]
141. Augustine, C.K.; Yoshimoto, Y.; Gupta, M.; Zipfel, P.A.; Selim, M.A.; Febbo, P.; Pendergast, A.M.; Peters, W.P.; Tyler, D.S. Targeting N-cadherin enhances antitumor activity of cytotoxic therapies in melanoma treatment. *Cancer Res.* **2008**, *68*, 3777–3784. [[CrossRef](#)] [[PubMed](#)]
142. Beasley, G.M.; Riboh, J.C.; Augustine, C.K.; Zager, J.S.; Hochwald, S.N.; Grobmyer, S.R.; Peterson, B.; Royal, R.; Ross, M.I.; Tyler, D.S. Prospective Multicenter Phase II trial of systemic ADH-1 in combination with melphalan via isolated limb infusion in patients with advanced extremity melanoma. *J. Clin. Oncol.* **2011**, *29*, 1210–1215. [[CrossRef](#)] [[PubMed](#)]

143. Perotti, A.; Sessa, C.; Mancuso, A.; Noberasco, C.; Cresta, S.; Locatelli, A.; Carcangiu, M.L.; Passera, K.; Braghetti, A.; Scaramuzza, D.; et al. Clinical and pharmacological phase I evaluation of Exherin (ADH-1), a selective anti-N-cadherin peptide in patients with N-cadherin-expressing solid tumours. *Ann. Oncol.* **2009**, *20*, 741–745. [[CrossRef](#)] [[PubMed](#)]
144. Turley, R.S.; Tokuhisa, Y.; Toshimitsu, H.; Lidsky, M.E.; Padussis, J.C.; Fontanella, A.; Deng, W.; Augustine, C.K.; Beasley, G.M.; Davies, M.A.; et al. Targeting N-cadherin increases vascular permeability and differentially activates AKT in melanoma. *Ann. Surg.* **2015**, *261*, 368–377. [[CrossRef](#)] [[PubMed](#)]
145. Doro, F.; Colombo, C.; Alberti, C.; Arosio, D.; Belvisi, L.; Casagrande, C.; Fanelli, R.; Manzoni, L.; Parisini, E.; Piarulli, U.; et al. Computational design of novel peptidomimetic inhibitors of cadherin homophilic interactions. *Org. Biomol. Chem.* **2015**, *13*, 2570–2573. [[CrossRef](#)] [[PubMed](#)]
146. Nardone, V.; Lucarelli, A.P.; Dalle, V.A.; Fanelli, R.; Tomassetti, A.; Belvisi, L.; Civera, M.; Parisini, E. Crystal structure of human E-Cadherin-EC1EC2 in complex with a peptidomimetic competitive inhibitor of cadherin homophilic interaction. *J. Med. Chem.* **2016**, *59*, 5089–5094. [[CrossRef](#)] [[PubMed](#)]
147. Dechantsreiter, M.A.; Planker, E.; Matha, B.; Lohof, E.; Holzemann, G.; Jonczyk, A.; Goodman, S.L.; Kessler, H. N-Methylated cyclic RGD peptides as highly active and selective $\alpha V\beta 3$ integrin antagonists. *J. Med. Chem.* **1999**, *42*, 3033–3040. [[CrossRef](#)] [[PubMed](#)]
148. Arap, W.; Pasqualini, R.; Ruoslahti, E. Cancer treatment by targeted drug delivery to tumor vasculature in a mouse model. *Science* **1998**, *279*, 377–380. [[CrossRef](#)] [[PubMed](#)]
149. Manzoni, L.; Belvisi, L.; Arosio, D.; Civera, M.; Pilkington-Miksa, M.; Potenza, D.; Caprini, A.; Araldi, E.M.; Monferini, E.; Mancino, M.; et al. Cyclic RGD-containing functionalized azabicycloalkane peptides as potent integrin antagonists for tumor targeting. *ChemMedChem* **2009**, *4*, 615–632. [[CrossRef](#)] [[PubMed](#)]
150. Hou, L.; Zhao, X.; Wang, P.; Ning, Q.; Meng, M.; Liu, C. Antitumor activity of antimicrobial peptides containing CisoDGRC in CD13 negative breast cancer cells. *PLoS ONE* **2013**, *8*, e53491. [[CrossRef](#)] [[PubMed](#)]
151. Panzeri, S.; Zanella, S.; Arosio, D.; Vahdati, L.; Dal, C.A.; Pignataro, L.; Paolillo, M.; Schinelli, S.; Belvisi, L.; Gennari, C.; et al. Cyclic isoDGR and RGD peptidomimetics containing bifunctional diketopiperazine scaffolds are integrin antagonists. *Chemistry* **2015**, *21*, 6265–6271. [[CrossRef](#)] [[PubMed](#)]
152. Li, S.; Gray, B.P.; McGuire, M.J.; Brown, K.C. Synthesis and biological evaluation of a peptide-paclitaxel conjugate which targets the integrin $\alpha v\beta 6$. *Bioorg. Med. Chem.* **2011**, *19*, 5480–5489. [[CrossRef](#)] [[PubMed](#)]
153. Shintani, Y.; Fukumoto, Y.; Chaika, N.; Grandgenett, P.M.; Hollingsworth, M.A.; Wheelock, M.J.; Johnson, K.R. ADH-1 suppresses N-cadherin-dependent pancreatic cancer progression. *Int. J. Cancer* **2008**, *122*, 71–77. [[CrossRef](#)] [[PubMed](#)]



© 2016 by the authors; licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC-BY) license (<http://creativecommons.org/licenses/by/4.0/>).