

Tannic Acid-Induced Surface-Catalyzed Secondary Nucleation during the Amyloid Fibrillation of Hen Egg White Lysozyme

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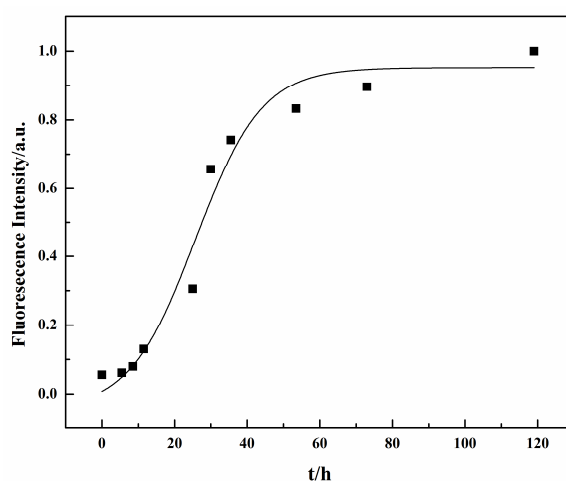


Figure S1. ThT fluorescence assay for the fibrillation kinetics of lysozyme with empirical fit using the equation of $F = F_0 + 1 / (1 + \exp[r_{\max} (\tau_{1/2} - t)])$, where $\tau_{1/2}$ is the time for half completion of aggregation, r_{\max} is the maximum growth rate, and $\tau_{1/2} - 2/r_{\max}$ is the lag phase time duration [1].

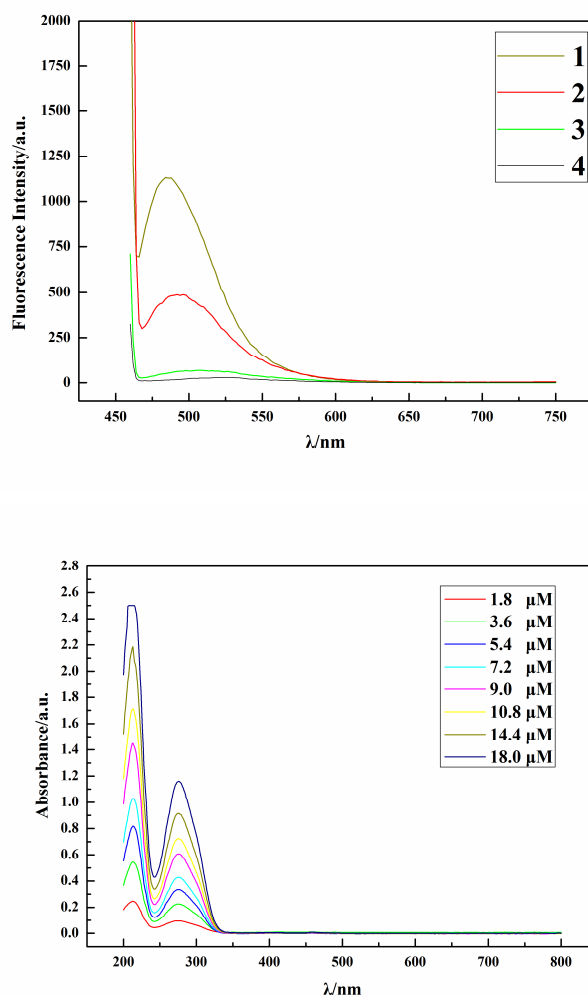


Figure S2. (Top) Fluorescence spectra of ThT bound to amyloid fibril. $C_{\text{ThT}} = 10\mu\text{M}$; Incubation condition: $C_{\text{lysozyme}} = 5\text{mg/mL}$; $t = 120\text{h}$ (1); ThT bound to non-fibrillar aggregates $C_{\text{ThT}} = 10\mu\text{M}$; Incubation condition: $C_{\text{lysozyme}} = 5\text{mg/mL}$; $C_{\text{Tannic acid}} = 1200\mu\text{M}$; $t = 0\text{h}$ (2); the control (i.e., ThT + tannic acid), $C_{\text{ThT}} = 10\mu\text{M}$ (3); and the control (tannic acid alone), $C_{\text{Tannic acid}} = 1200\mu\text{M}$ (4). (Bottom) Absorption spectra of tannic acid under different concentrations.

Note: The ThT assay was performed ex situ. For each ThT assay, only $10\mu\text{l}$ of the test solution was added in to 1mL of ThT solution. So in the test solution in ThT assay, the concentration of ThT is $10\mu\text{M}$; the concentration of lysozyme is 0.05mg/mL , and the concentration of tannic acid is only $12\mu\text{M}$.

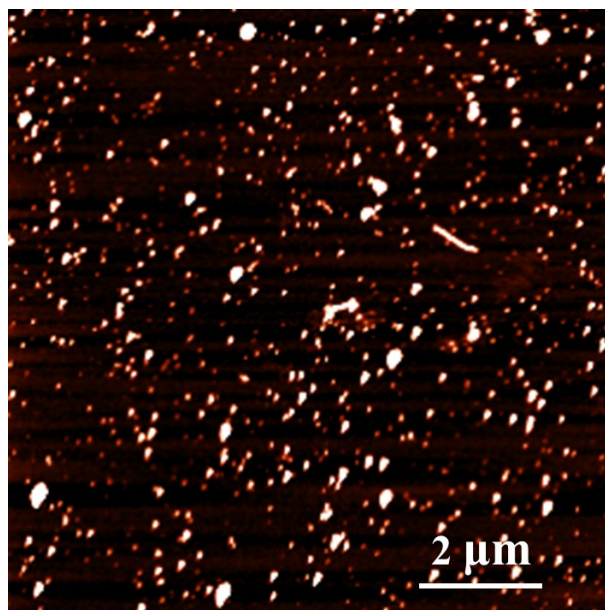


Figure S3. AFM evidence for the presence of the non-fibrillar aggregates at the end of incubation in the presence of tannic acid (i.e., $t=137\text{h}$). $C_{\text{lysozyme}}=5\text{mg/mL}$; $C_{\text{Tannic acid}}=1200\mu\text{M}$. This image was from the same mica surface for Figure 4F.

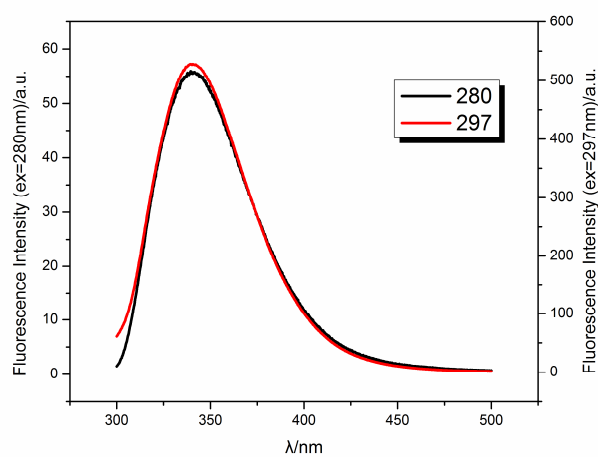


Figure S4. Fluorescence spectra of lysozyme at 5mg/mL excited with 280 nm and 297 nm.

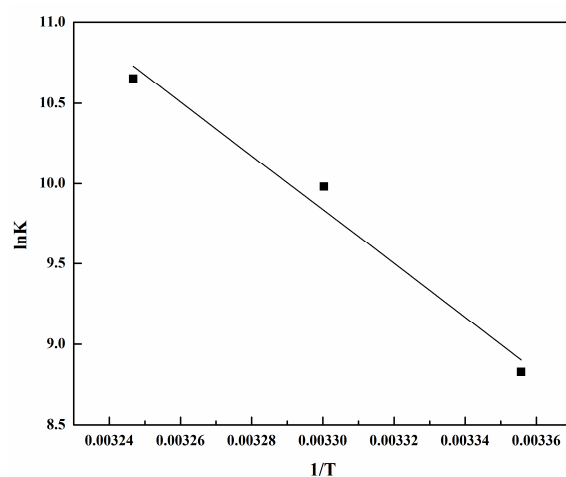


Figure S5. Van 't Hoff plot for lysozyme quenched by tannic acid at 323 K, 328 K and 333 K.

$C_{\text{lysozyme}}=3.6 \times 10^{-6} \text{ mol/L}$; $C_{\text{Tannic acid (1-9)}} = (0, 1.8, 3.6, 5.4, 7.2, 9.0, 10.8, 14.4, 18.0) \times 10^{-6} \text{ mol/L}$

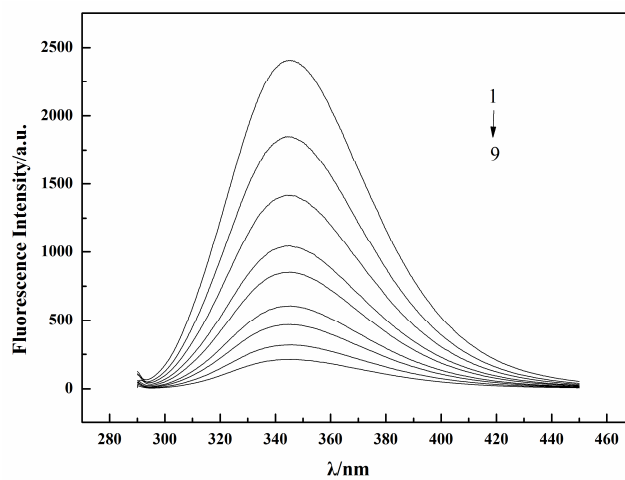


Figure S6. Fluorescence spectra of lysozyme quenched by tannic acid at 298 K.

$C_{\text{lysozyme}}=3.6 \times 10^{-6} \text{ mol/L}$; $C_{\text{Tannic acid (1-9)}} = (0, 1.8, 3.6, 5.4, 7.2, 9.0, 10.8, 14.4, 18.0) \times 10^{-6} \text{ mol/L}$

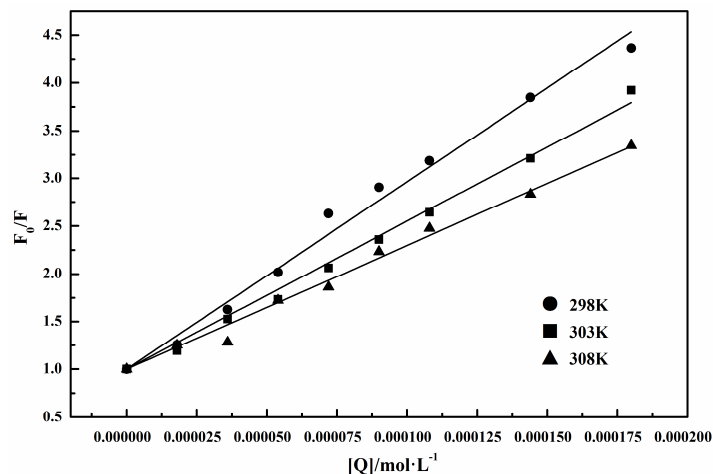


Figure S7. Stern-Volmer plots for lysozyme quenched by tannic acid at 298 K, 303 K and 308 K.

Table S1. The Stern-Volmer parameters of lysozyme-tannic acid system at 298 K, 303 K and 308 K.

T/K	$K_{sv}/(\text{L}\cdot\text{mol}^{-1})$	$K_q/(\text{L}\cdot\text{mol}^{-1}\cdot\text{s}^{-1})$	r^*
298	$(1.65\pm 0.28)\times 10^4$	$(1.83\pm 0.31)\times 10^{12}$	0.9897 ± 0.0081
303	$(1.47\pm 0.08)\times 10^4$	$(1.63\pm 0.09)\times 10^{12}$	0.9944 ± 0.0030
308	$(1.34\pm 0.13)\times 10^4$	$(1.49\pm 0.14)\times 10^{12}$	0.9917 ± 0.0039

* r is correlation coefficient.

Table S2. Binding parameters of lysozyme-tannic acid system at 298 K, 303 K and 308 K.

T/K	$K_A/(\text{L}\cdot\text{mol}^{-1})$	n	r
298	$(1.17\pm 0.86)\times 10^5$	1.20 ± 0.08	0.9931 ± 0.0051
303	$(6.20\pm 4.54)\times 10^4$	1.13 ± 0.10	0.9936 ± 0.0036
308	$(3.27\pm 1.03)\times 10^4$	1.10 ± 0.03	0.9866 ± 0.0114

Table S3. Parameters of E , J , R_0 , r of the lysozyme-tannic acid system at 298 K, 303 K and 308 K.

T/K	$E/\%$	$J/(\text{cm}^3\cdot\text{L}\cdot\text{mol}^{-1})$	R_0/nm	r/nm
298	32.43 ± 8.52	$(5.96\pm 0.01)\times 10^{-15}$	$2.34\pm 3.27\times 10^{-4}$	2.65 ± 0.17
303	31.48 ± 3.02	$(5.99\pm 0.04)\times 10^{-15}$	$2.34\pm 2.34\times 10^{-3}$	2.67 ± 0.06
308	26.61 ± 4.28	$(5.96\pm 0.06)\times 10^{-15}$	$2.34\pm 4.18\times 10^{-3}$	2.78 ± 0.11

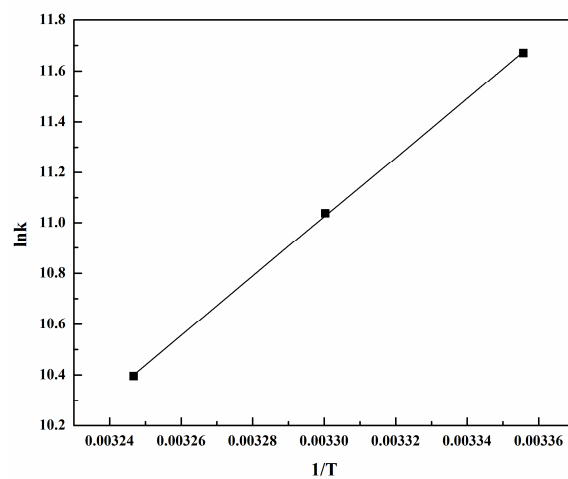


Figure S8. Van 't Hoff plot for lysozyme quenched by tannic acid at 298 K, 303 K and 308 K.

$C_{\text{lysozyme}}=3.6 \times 10^{-6} \text{ mol/L}$; $C_{\text{Tannic acid (1-9)}} = (0, 1.8, 3.6, 5.4, 7.2, 9.0, 10.8, 14.4, 18.0) \times 10^{-6} \text{ mol/L}$

1. Abelein, A.; Jarvet, J.; Barth, A.; Graslund, A.; Danielsson, J. Ionic strength modulation of the free energy landscape of a beta(40) peptide fibril formation. *J. Am. Chem. Soc.* **2016**, *138*, 6893-6902, 10.1021/jacs.6b04511