



Review

# CaMKII Activity in the Inflammatory Response of Cardiac Diseases

Maria Rosaria Rusciano <sup>1,2,†</sup>, Elena Sommariva <sup>3,†</sup>, Victorine Douin-Echinard <sup>4,5</sup> ,  
Michele Ciccarelli <sup>1</sup> , Paolo Poggio <sup>6</sup> and Angela Serena Maione <sup>3,\*</sup>

<sup>1</sup> Department of Medicine, Surgery and Odontology, University of Salerno, 84081 Baronissi, Italy

<sup>2</sup> Casa di Cura Montevergine, 83013 Mercogliano, Italy

<sup>3</sup> Vascular Biology and Regenerative Medicine Unit, Centro Cardiologico Monzino IRCCS, 20138 Milan, Italy

<sup>4</sup> Institute of Cardiovascular and Metabolic Diseases, Inserm UMR 1048, 31432 Toulouse, France

<sup>5</sup> Paul Sabatier University, 31432 Toulouse, France

<sup>6</sup> Unit for the Study of Aortic, Valvular and Coronary Pathologies, Centro Cardiologico Monzino IRCCS, 20138 Milan, Italy

\* Correspondence: [angela.maione@ccfm.it](mailto:angela.maione@ccfm.it); Tel.: +39-02-5800-2753

† These authors contributed equally to this work.

Received: 7 August 2019; Accepted: 3 September 2019; Published: 6 September 2019



**Abstract:** Inflammation is a physiological process by which the body responds to external insults and stress conditions, and it is characterized by the production of pro-inflammatory mediators such as cytokines. The acute inflammatory response is solved by removing the threat. Conversely, a chronic inflammatory state is established due to a prolonged inflammatory response and may lead to tissue damage. Based on the evidence of a reciprocal regulation between inflammation process and calcium unbalance, here we described the involvement of a calcium sensor in cardiac diseases with inflammatory drift. Indeed, the Ca<sup>2+</sup>/calmodulin-dependent protein kinase II (CaMKII) is activated in several diseases with an inflammatory component, such as myocardial infarction, ischemia/reperfusion injury, pressure overload/hypertrophy, and arrhythmic syndromes, in which it actively regulates pro-inflammatory signaling, among which includes nuclear factor kappa-B (NF-κB), thus contributing to pathological cardiac remodeling. Thus, CaMKII may represent a key target to modulate the severity of the inflammatory-driven degeneration.

**Keywords:** CaMKII; inflammation; Ca<sup>2+</sup>; ROS; NF-κB; cardiac diseases

## 1. The Immune System and the Inflammatory Process in the Heart

Inflammation is a natural and necessary immune reaction that occurs when organisms experience infections, stress, or tissue damage to fight the insulting agent. Although essential for body protection against pathogens, excessive inflammation can provoke by-stander injury and cause organ dysfunction [1]. Inflammation is a complex process ensuring leukocyte infiltration at the site of tissue injury, and it is finely tuned by a large panel of molecules, tissue resident immune cells, and stromal cells [2].

Typically, the inflammatory reaction is composed of four constituents: inducers of inflammation; sensors on the cell surface that detect them; mediators, produced when prompted by the sensors; and the target tissues that respond specifically to the inflammatory mediators. Different forms exist for each constituent, and their combinations compose distinct inflammatory pathways. The type of pathway induced depends on the nature of the trigger [1].

Pathogens are recognized by several major classes of pattern recognition receptors (PRRs), expressed both in immune and non-immune sentinel cells, which are activated by pathogen-associated molecular patterns (PAMPs) [1].

Sterile inflammation takes place in the absence of pathogens. In this case the trigger is constituted by intracellular particles released by necrotic or apoptotic cells. In this case PRRs are activated by endogenous agents (danger-associated molecular patterns; DAMPs) to elicit an inflammatory response [3].

PRRs include the Toll-like receptors (TLRs), C-type lectin receptors (CLRs), retinoic acid-inducible gene (RIG)-I-like receptors (RLRs), and NOD-like receptors (NLRs) [4].

Receptor activation triggers relevant intracellular signaling pathways, among which are the mitogen-activated protein kinase (MAPK), nuclear factor kappa-B (NF- $\kappa$ B), Janus kinase, activator protein-1 (AP-1), interferon regulatory factor 3 (IRF3), and activation of transcription [5,6].

Gene transcription activation drives, in turn, the production and secretion of pro-inflammatory cytokine, such as interleukin (IL)-1, tumor necrosis factor (TNF)- $\alpha$ , IL-6, colony stimulating factor (CSF), interferons, transforming growth factor  $\beta$  (TGF- $\beta$ ), and chemokines, which contribute to the inflammatory response [7].

The acute phase of the response is characterized by a massive influx of granulocytes, then monocytes, which both play a predominant role in the clearance of the pathogen and removal of tissue debris.

The resolution of inflammation is tightly regulated [8]. The severity of disease pathogenesis may be related to effective resolution or chronicization of the inflammatory process [9,10].

In particular, a chronic inflammatory process plays a crucial role in the progression of heart diseases and exerts a deleterious role on cardiac function. Heart specific cytokines, neurohormones and pro-inflammatory molecules, which can be referred to as cardiokines, actively drive the progression of cardiac dysfunction in heart failure [11,12]. The cells composing the heart, such as cardiomyocytes, fibroblasts, vascular cells, and progenitor cells, are able to secrete several cardiokines following different environmental stimuli, realizing a specialized network that is critical for heart homeostasis. These proteins, including cytokines, such as TNF- $\alpha$  and TGF- $\beta$  or different interleukins, are able to control the balance between normal cardiac function and pathological myocardial remodeling based on their ability to influence cardiomyocyte apoptosis, fibroblast activation, and vascular cell proliferation.

Notably, low concentrations of TNF- $\alpha$  produces a cardioprotective effect, while increased levels of TNF- $\alpha$  have been associated to heart failure and diastolic dysfunction, and is positively correlated to the severity of the diseases. Transgenic mice with a cardiac-specific TNF- $\alpha$  overexpression display heart failure, cardiac dilatation, fibrosis, altered contractile function, Ca<sup>2+</sup> handling defects, and premature death [13]. Furthermore, the progression of TNF- $\alpha$ -induced cardiac remodeling is associated to the activation of cardiomyocyte apoptosis and proteasome dysfunction [14,15]. The TNF- $\alpha$  increase occurring during ischemia/reperfusion injury (I/R) is related to Ca<sup>2+</sup> overload and the resultant cardiac dysfunction [16,17]. The pharmacological modulation of TNF- $\alpha$  production is able to improve cardiac function and reduce the intracellular Ca<sup>2+</sup> overload and oxidative stress that arises following I/R stress [17–19].

In addition, TGF- $\beta$ , another cardiokine that also has a physiological cardioprotective effect, if deregulated, actively participates in the pathological cardiac remodeling mediating the tissue fibrosis that follows the tissue-injury-derived inflammation acting on fibroblast activation, differentiation, and extracellular matrix protein secretion [20,21]. TGF- $\beta$ 1 over-expression has been associated with myocardial hypertrophy, hypertensive cardiac remodeling, several cardiomyopathies, and genetic aortic syndromes [22,23]. Transgenic mice with the TGF- $\beta$  type 2 receptor conditional knockdown in cardiomyocytes display, following a sustained pressure condition, reduced interstitial fibrosis and improved heart function, and they do not exhibit cardiac dysfunction and chamber dilation [24]. The TGF- $\beta$ 1-dependent cardiac fibrosis also correlates to the regulation of intracellular Ca<sup>2+</sup> concentrations by the type 2 ryanodine receptor (RyR2). TGF- $\beta$ 1 and collagen levels are up-regulated in cardiomyocytes subjected to mechanical stress, but this event is reverted in RyR2 knockdown cardiomyocytes [25]. Pharmacological inactivation of non-canonical TGF- $\beta$  signaling by arjunolic acid treatment leads to the

up-regulation of peroxisome proliferator activated receptor alpha and results in the down-regulation of collagen gene expression in the hypertrophy-model of cardiac fibroblasts [26].

Recently, an emerging role for the IL-33/ST2 pathway in the inflammation that occurs during the cardiac stress condition has been described [26,27]. ST2, which belongs to the Toll-like receptor family, exerts an immunomodulatory effect based on its ability to regulate cytokine production [28,29]. Furthermore, the soluble ST2 form represents a predictive biomarker in patients with chronic heart failure and a severe prognosis [29]. The IL-33/ST2 interaction results in an anti-hypertrophic effect by blocking NF- $\kappa$ B activation. Mice lacking ST2 have worsened hypertrophy, cardiac dilation, ventricular fractional shortening, increase fibrosis, and reduced survival in a pressure overload condition [30].

## 2. Calcium/Calmodulin-Dependent (CaMK) II in the Heart

Calcium/calmodulin-dependent kinases are a family of serine/threonine kinases that respond to the intracellular calcium  $Ca^{2+}$  changes  $[Ca^{2+}]_i$  and consist of three members: CaMKI, CaMKII, and CaMKIV [31].  $Ca^{2+}$  transduces its functions by forming a complex with calmodulin (CaM), which acts as a ubiquitous  $Ca^{2+}$  receptor [31].

CaMKII is a multimeric enzyme consisting of 12 monomers [32]. Each monomer shares the same structure that consists of an N-terminal catalytic domain, a C-terminal association domain, and the central auto-regulatory domain where the  $Ca^{2+}$ /CaM binding site is located [33]. CaMKII is the most suitable decoder of total  $[Ca^{2+}]_i$  although it is also engaged by intracellular  $Ca^{2+}$  oscillations and transients [34,35].

Under resting conditions, the CaMKII regulatory domain interacts and sterically blocks the catalytic domain, leading to its auto-inhibitory state. The activation process requires the binding of the  $Ca^{2+}$ /CaM complex, which displaces the intrastereical auto-inhibition and exposes the kinase substrate and ATP binding sites of the catalytic domain [36]. At this point, the activated monomer is able to sequentially phosphorylate at Thr286/287 (depending on CaMKII isoforms), the regulatory domains of adjacent CaMKII monomers. The auto-phosphorylation confers to CaMKII an autonomous kinase activity even after the dissociation of the  $Ca^{2+}$ /CaM complex, thus preventing the re-association of the catalytic domain with the auto-inhibitory domain [33,37]. Furthermore, the activation also induces the "CaM trapping" that leads to an increased affinity to CaM binding and to a time-sustained CaMKII activity upon low  $[Ca^{2+}]_i$  conditions [38].

An alternative route of CaMKII activation has been described involving the reactive oxygen species (ROS) produced by various sources including nicotinamide adenine dinucleotide phosphate (NADPH) oxidase and mitochondria. Specifically, ROS oxidizes CaMKII, at methionine 281/282, which remains active even in the absence of the  $Ca^{2+}$ /CaM complex [39]. Essentially, the oxidation of the methionine residues of CaMKII works as a sensor of ROS increments and correlates with a sustained kinase activity [39,40].

Another possible trigger of CaMKII autonomous activation is hyperglycemia. The extracellular glucose elevation leads to O-linked N-acetyl-glucosamine (O-GlcNAc) modification at CaMKII S279 [41]. Furthermore, CaMKII autonomous activity can be induced through a nitric oxide (NO)-dependent pathway by S-nitrosylation of Cys290 [42]. Notably, both O-GlcNAc and S-nitrosylation modifications require the initial  $Ca^{2+}$ /CaM-dependent activation and result in persistent autonomous CaMKII activation [41,42].

The CaMKII inactivation involves either phosphatase-dependent or -independent mechanisms. The dephosphorylation of Thr286 occurs through 70% of the protein phosphatase 2A (PP2A) activity; PP1 and PP2C act for the remaining activity [43]. An alternative CaMKII inactivation mechanism, which is typical of post-synaptic plasticity regulation [13], consists of the auto-phosphorylation of Thr305/306 that prevents the CaM rebinding to the regulatory domain (CaM-capping) [33] by modifying the  $Ca^{2+}$ /CaM binding site [44,45].

The CaMKII tissue distribution is variable, and the four CaMKII isoforms ( $\alpha$ ,  $\beta$ ,  $\gamma$ ,  $\delta$ ) encoded by separate genes show tissue-preferential expression [46]. CaMKII $\alpha$  and  $\beta$  are the neuronal isoforms [47] while the CaMKII $\delta$  and  $\gamma$  isoforms are predominantly expressed in cardiac tissue [48,49].

CaMKII $\delta$  is critical during the pathogenesis of cardiac hypertrophy after catecholaminergic stimulation [50]. It modulates transcription by mediating histone deacetylase (HDAC)4 phosphorylation during pressure overload [51]. CaMKII $\delta$  affects Ca<sup>2+</sup> handling by phosphorylation of RyR2 and phospholamban (PLN), thus inducing changes in sarcoplasmic reticulum (SR) Ca<sup>2+</sup> content and resulting in diastolic Ca<sup>2+</sup> leak [51], leading to diastolic dysfunction and arrhythmogenesis [52].

Several pieces of evidence correlate CaMKII activity to physiological functions such as cell proliferation and cell cycle progression. In particular, the inhibition of CaMKII reduces vascular smooth muscle [53] and endothelial cell proliferation [54] as well as S-phase progression of the cell cycle [55]. On the other hand, the over-expression of CaMKII $\gamma$  negatively regulates vascular smooth muscle proliferation [56]. Interestingly, CaMKII specific inhibitors increase proliferation of cardiomyocytes derived from induced pluripotent stem cells [57].

### 3. CaMKII and Inflammation in Cardiac Diseases

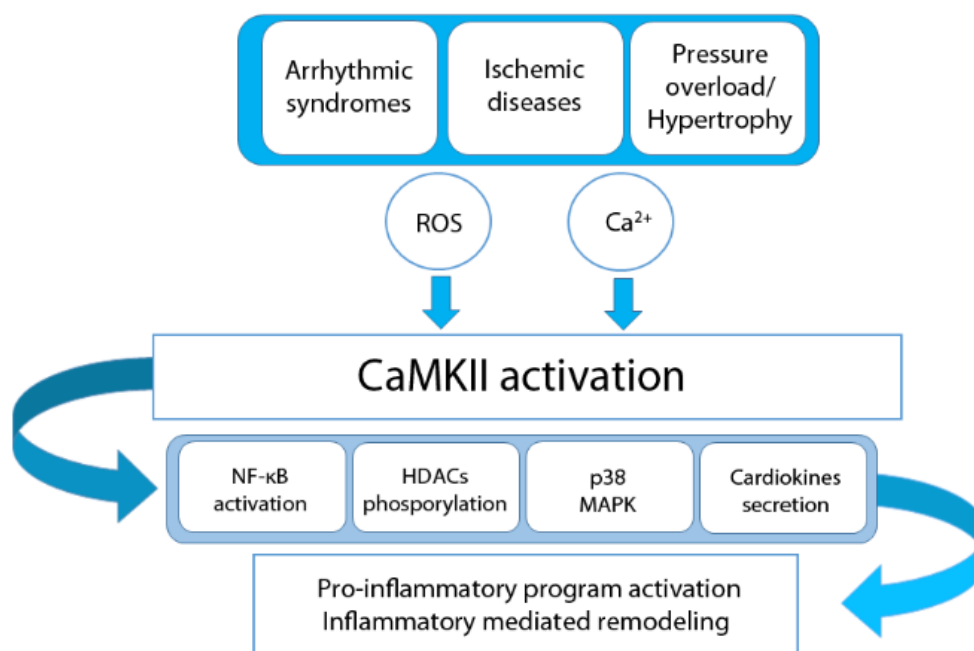
Ca<sup>2+</sup> has been associated with different events of the inflammatory response [58–61] as well as with the regulation of proliferation, anergy and cell death of T cells [62]. Based on its ability to act as an intracellular Ca<sup>2+</sup> sensor, CaMKII is recognized as a key regulator of the immune and inflammatory responses [63–65] at different levels.

CaMKII regulates the physiology of T cells. The Ca<sup>2+</sup>-independent form of CaMKII $\gamma$  enhances T cell memory formation and modulates cell death [66]. In T cells, treatment with CaMKII inhibitor KN93 modulates the NF- $\kappa$ B activation pathway by abolishing the phorbol-ester-induced phosphorylation of inhibitory  $\kappa$ B (I $\kappa$ B) proteins [67]. Moreover, CaMKII modulates IL-10, IL-2, and IL-4 production by T lymphocytes. Specifically, the overexpression of the constitutively active CaMKII form leads to increased IL-10 protein and mRNA accumulation based on its ability to directly modulate IL-10 promoter activity [68]. In addition, it is involved in the Ca<sup>2+</sup>-dependent IL-2 transcriptional arrest, causing anergy [63,64], and regulates IL-4 by direct action on its promoter [63].

Studies performed on macrophages highlighted that CaMKII boosts pro-inflammatory cytokines and type I interferon production upon TLR stimulation [69] and participates in the Wnt5A signaling-mediated inflammatory response [70].

Furthermore, CaMKII regulates dendritic cell physiology, acting at different levels on the expression and localization of MHC Class II proteins [71], cell maturation, and the antigen presentation ability following phagocytosis-induced stimulation [72].

In addition to the response to pathogens, the inflammation process in cardiac disease is mostly an adaptive response to myocardial injury [73,74]. In particular, sustained CaMKII activation is demonstrated to be involved in several cardiovascular diseases. Notably, the inhibition of CaMKII has been suggested as a novel therapeutic target to treat cardiac arrhythmias, heart failure, and hypertrophy [75–77]. The following subsections will summarize the main findings and mechanisms regarding cardiac diseases in which CaMKII and inflammation mediate pathological remodeling (Figure 1).



**Figure 1.** Schematic description of Calcium/calmodulin-dependent (CaMK) II involvement in the inflammatory response in cardiac diseases.

### 3.1. Ischemic Diseases

The cytosolic  $\text{Ca}^{2+}$  overload is one of the common events that leads to heart failure and ischemic heart disease. The resulting sustained CaMKII activation promotes L-type  $\text{Ca}^{2+}$  channel opening probability by phosphorylation of the  $\alpha$ -subunit of L-type voltage-gated  $\text{Ca}^{2+}$  channel (CaV1.2). L-type  $\text{Ca}^{2+}$  channel opening regulates cellular  $\text{Ca}^{2+}$  homeostasis, which in turn controls cardiac myocyte apoptosis [78]. In vivo, CaMKII inhibition is able to protect against myocardial apoptosis induced by myocardial infarction [79] and restores SR  $\text{Ca}^{2+}$  content. The RyR2 mutated mouse model, which lacks the CaMKII phosphorylation site, is resistant to apoptosis and displays improved cardiac function after myocardial infarction [80]. In addition, the overexpression of mutant CaV1.2, which is resistant to CaMKII binding and thus precludes CaV1.2 phosphorylation, retards cardiomyocyte death [81]. Moreover, the overexpression of CaMKII $\delta$  leads to increased cardiomyocyte apoptosis, together with elevated cytosolic  $\text{Ca}^{2+}$  and enhanced mitochondrial cytochrome C release [82].

Moreover, ischemic-induced necrotic cell death with consequent release of intracellular molecules [83] results in the activation of TLRs in cardiomyocytes, inducing pro-inflammatory transcriptional pathways [84–87]. Several pieces of evidence have established that CaMKII has a central role in regulating inflammation in myocardial infarction (MI), since it is oxidized as a consequence of increased  $\beta$ -adrenergic activation upon MI, which is followed by increased intracellular ROS [88,89]. The oxidized CaMKII is able to enhance pro-inflammatory transcriptional signaling by promoting NF- $\kappa$ B activity [67]. Gene expression profiling performed in mouse hearts of transgenic AC3-I mice, in which there is a cardiomyocyte-limited expression of a CaMKII inhibitory peptide, showed that CaMKII inhibition reduces the post-MI upregulation of pro-inflammatory genes and complement factor B [90].

The inflammatory response also occurs during cardiac reperfusion following an acute ischemic event. In addition to the pro-inflammatory signaling-activated cardiomyocyte death described for MI, I/R injury also leads to the opening of the mitochondrial permeability transition pores, resulting in the increase of cellular  $\text{Ca}^{2+}$  and ROS [91,92]. It has been demonstrated that cardiac-specific CaMKII $\delta$  deletion protects against I/R since it decreases infarct size, attenuates apoptosis, and improves functional recovery. CaMKII $\delta$  deletion is also able to reduce I/R-induced inflammation by preventing

the reduction of I $\kappa$ B and upregulation of NF- $\kappa$ B target genes [93]. In contrast, in a similar I/R study, an effect on infarct size following I/R CaMKII $\delta$  KO, CaMKII $\gamma$  KO, and CaMKII $\gamma/\delta$  double knockout (DKO) mice has not been observed. A reduced infarct size and improved cardiac function are observed only at five weeks after I/R in CaMKII $\gamma/\delta$  DKO mice. Notably, loss of CaMKII reduces the cardiomyocyte expression and secretion of the chemokines C-C motif ligand 2 and 3, leading to decreased infiltration of CD45<sup>+</sup> leukocytes, thus attenuating inflammatory mediated post-infarct remodeling [94].

### 3.2. Pressure Overload/Hypertrophy

The recruitment of immune cells due to inflammatory responses and contribution to cardiac remodeling also occurs with pressure overload [95–97]. Angiotensin II (Ang II) infusion represents the common treatment to study the inflammatory-induced remodeling by hypertensive non-ischemic stress [98]. It has been reported that Ang II treatment induces NF- $\kappa$ B-dependent inflammatory gene expression and inflammasome activation, which were reduced in a cardiomyocyte-specific CaMKII $\delta$  KO mouse model. Therefore, CaMKII $\delta$  activation mediates inflammation-driven remodeling [99]. As an alternative mechanism, Ang II promotes ROS release, the oxidation of CaMKII, thus resulting in the activation of p38 MAPK, another major mediators of the inflammatory response [100].

An alteration of intracellular Ca<sup>2+</sup> cycling has also been observed in another experimental model of pressure overload, the transverse aortic constriction (TAC), which reflects increased afterload [101,102]. In turn, afterload is responsible for CaMKII activation based on the induced L-type calcium current increase [103,104]. The TAC model results in hypertrophy with increased fibrosis, inflammation, cardiomyocyte apoptosis, and persistent CaMKII activation [102]. As described for other models, CaMKII $\delta$  activation triggers the inflammasome through NF- $\kappa$ B and ROS signaling in cardiomyocytes, inducing chemokine production, which contributes to macrophage infiltration and the development of fibrosis [105]. Likewise, fibrosis and ventricular dilation and dysfunction can be reduced by both selective CaMKII $\delta$  deletion and by blocking CaMKII activation within the first two weeks of TAC and after the onset of inflammatory cell accumulation [105], thus confirming the CaMKII involvement in the maladaptive response during pressure overload [106].

It has been reported that CaMKII is involved in the transcriptional regulation of hypertrophic genes by regulating the phosphorylation of histone deacetylases (HDACs), which in turn affect TNF- $\alpha$  and IL-1 $\beta$  expression and cardiac fibrosis [107,108]. CaMKII is able to phosphorylate and prevent the nuclear import of HDAC4, based on the presence of two conserved CaMK phosphorylation sites in the N-terminal regions of class II HDACs, thereby inducing the repression of MEF2 and the activation of the hypertrophic program [108,109]. Mice lacking the  $\delta$  isoform of CaMKII display a reduced phosphorylation of HDAC4 and are protected against hypertrophy and fibrosis following TAC [48]. Analogously, hypertrophic genes such as ANF, brain natriuretic peptide, myosin heavy chain, and skeletal actin are overexpressed in the heart of transgenic mice with cardiomyocyte-specific expression of CaMKII $\delta$ B and CaMKII $\delta$ C due to the induced transactivation of MEF2 [51]. A common event occurring during hypertrophy is the increase of the systemic levels of Ang II, which in turn acts as an activator of cardiac fibroblast proliferation. The excessive cardiac fibroblast proliferation is associated with inflammatory cytokine secretion and promotes the progression of cardiac fibrosis, thus contributing to the heart failure. The inhibition of CaMKII is able to reduce the Ang-II-induced cardiac fibroblast proliferation as well as the secretion of TGF- $\beta$ 1 and TNF- $\alpha$ . Moreover, CaMKII inhibition also reverts the upregulation of MMP-1, 2, and 9 and collagen I and III following Ang II treatment, confirming its involvement in extracellular matrix regulation [110].

CaMKII also acts as downstream target of  $\beta$ -adrenergic receptor ( $\beta$ AR) signalling. The cytosolic Ca<sup>2+</sup> increase, following  $\beta$ AR stimulation, is related to the physiologic augment of cardiac contraction. Excessive  $\beta$ AR activation results in pathological heart remodeling and myocardial hypertrophy. A genetic mouse model of cardiac CaMKII inhibition is protected from maladaptive remodeling caused by excessive  $\beta$ AR stimulation [76].

### 3.3. Arrhythmic Syndromes

The activation of CaMKII by acting on ion channels is described as a possible trigger for some inherited cardiac arrhythmia syndromes [75]. The overactivation of CaV1.2 by CaMKII results in an enhanced peak and a slowed Ca<sup>2+</sup> inward current inactivation, causing membrane depolarization and the prolongation of the action potential duration, leading to arrhythmias [81]. CaMKII is also able to regulate both SR Ca<sup>2+</sup> uptake and release. First, CaMKII catalyzes PLN phosphorylation [111], thereby reducing its inhibitory effect on Sarco-Endoplasmic Reticulum Ca<sup>2+</sup> ATPase 2a (SERCA2a) thus causing the increase of Ca<sup>2+</sup> reuptake by the SR and myocardial relaxation. Second, CaMKII phosphorylates RyR2, leading to pro-arrhythmic abnormal diastolic Ca<sup>2+</sup> release from the SR, which results in Na<sup>+</sup>/Ca<sup>2+</sup> exchanger forward-mode activity and afterdepolarization. Moreover, CaMKII inhibition drastically reduces diastolic SR Ca<sup>2+</sup> leak in human and rodent cardiomyocytes [112], leading to decreased spontaneous Ca<sup>2+</sup>-release (arrhythmogenic event) and enhanced ability of the SR to accumulate Ca<sup>2+</sup>.

Notably, cardiokines, and in particular TNF- $\alpha$  and IL-1, can favor arrhythmias by increasing calcium currents, thus interfering with Ca<sup>2+</sup> homeostasis and triggering arrhythmic events [113,114].

CaMKII activity has also been linked to atrial fibrillation (AF) in connection with the AMP-activated protein kinase pathway, leading to apoptosis and atrial remodeling [115,116]. Indeed, several inflammatory markers such as IL-6, C-reactive protein, and complementary factors are elevated in AF [117]. The acute administration of TNF- $\alpha$  in HL1 atrial cardiomyocytes leads to a significant increase in cytosol free Ca<sup>2+</sup> levels [118]. Moreover, AF is associated with an increase of total, phosphorylated, and oxidized CaMKII [119,120], secondary to Ca<sup>2+</sup> release from the SR [121].

TNF- $\alpha$  also promotes mitochondrial ROS production, which in turn leads to an enhanced oxidation of CaMKII [122]. Once active, CaMKII acts on downstream targets, such as ion channels (promoting arrhythmias) and pro-fibrotic pathways (promoting atrial remodeling), and mitochondria (promoting ROS-induced cell death). Moreover, animal studies showed that CaMKII inhibition is protective against AF [123]. The central role of CaMKII in the pathophysiology of AF makes it an attractive therapeutic target.

### 3.4. Influence of Cardiac Therapies on CaMKII Activation

CaMKII is a downstream target of multiple agonists for which effective antagonists are available and already used as routine clinical practice for cardiac diseases. These include ranolazine, ivabradine, beta-blockers, angiotensin-converting enzyme inhibitors (ACEI), and aldosterone antagonists.

Late Na<sup>+</sup> current dysregulation in hypertrophic cardiomyopathy is responsible for the intracellular Ca<sup>2+</sup> accumulation and activation of CaMKII [124]. Acute ranolazine administration reduces both the intracellular Na<sup>+</sup> and Ca<sup>2+</sup> levels and CaMKII activity, thus contributing to the reduction of hypertrophic cardiomyopathy-related cardiac remodeling myocardial dysfunction [125].

The pharmacological treatment of cardiac hypertrophy also includes the  $\beta$ -blockers, the renin inhibitors, and ACEI such as carvedilol, aliskiren and enalapril. Carvedilol exerts a beneficial effect based on its antioxidant, anti-inflammatory, and anti-fibrotic properties and significantly reduces CaMKII levels in isoproterenol-hypertrophied rats (also in combination with aliskiren treatment) [126,127]. Treatment with enalapril, in spontaneously hypertensive rats, is able to prevent hypertrophy, apoptosis, and CaMKII activity [128].

Furthermore, treatment with the mineralocorticoid receptor antagonist spironolactone reduces both ROS and ox-CaMKII levels in cultured neonatal myocytes stimulated with aldosterone, thus confirming CaMKII activity contribution to aldosterone-induced mortality during myocardial infarction [129].

It has been demonstrated that resveratrol has a cardioprotective effect based on its anti-inflammatory and antioxidant properties [130]. Resveratrol significantly prevents the diastolic intracellular Ca<sup>2+</sup> increase, ROS production, and activation of CaMKII induced by H<sub>2</sub>O<sub>2</sub> treatment in ventricular myocytes, which are overall responsible for stress-induced arrhythmogenic events [131]. The beneficial effect of resveratrol has also been demonstrated in a pressure overload model in

which it exerts an anti-hypertrophic effect, increases cardiac systolic function, reduces interstitial and perivascular fibrosis, and prevents CaMKII activation [131].

#### 4. Conclusions

Inflammation comprises a wide range of processes that affect many aspects of normal physiology and pathology. Inflammation switches from physiological to pathological mechanisms when it is not a time-limited event but a chronic process causing tissue damage or even death. The identification of possible modulators of the inflammatory response could be beneficial not only for chronic inflammatory diseases but also for the diseases in which the inflammatory component represents a limit to the resolution of the pathology, such as, in the cardiac scenario, myocardial infarction, pressure overload, I/R injury, and arrhythmic diseases. Increasing evidence suggests a pivotal role of CaMKII as a versatile kinase in many cardiac pathophysiological conditions involving inflammation. This is both a consequence of its activation properties in the presence of inflammatory states dysregulating Ca<sup>2+</sup> balance, and to its ability to enhance the pro-inflammatory transcriptional signaling leading to inflammatory state amplification and persistence. Consequently, achieving a deeper knowledge of the mechanism by which CaMKII, the immune system, and inflammation are reciprocally modulated will be of potential therapeutic importance to mitigate the severity of many cardiac diseases.

**Funding:** A.S.M., E.S., and V.D.-E. acknowledge financial support from Transnational Research Projects on Cardiovascular Diseases (ACM-HF JTC2016\_FP-40-021).

**Conflicts of Interest:** The authors declare no conflict of interest.

#### References

1. Medzhitov, R. Inflammation 2010: New adventures of an old flame. *Cell* **2010**, *140*, 771–776. [[CrossRef](#)] [[PubMed](#)]
2. McGettrick, H.M.; Butler, L.M.; Buckley, C.D.; Rainger, G.E.; Nash, G.B. Tissue stroma as a regulator of leukocyte recruitment in inflammation. *J. Leukoc. Biol.* **2012**, *91*, 385–400. [[CrossRef](#)] [[PubMed](#)]
3. Chen, G.Y.; Nunez, G. Sterile inflammation: Sensing and reacting to damage. *Nat. Rev. Immunol.* **2010**, *10*, 826–837. [[CrossRef](#)] [[PubMed](#)]
4. Takeuchi, O.; Akira, S. Pattern recognition receptors and inflammation. *Cell* **2010**, *140*, 805–820. [[CrossRef](#)] [[PubMed](#)]
5. Kyriakis, J.M.; Avruch, J. Mammalian mitogen-activated protein kinase signal transduction pathways activated by stress and inflammation. *Physiol. Rev.* **2001**, *81*, 807–869. [[CrossRef](#)] [[PubMed](#)]
6. Chen, L.; Deng, H.; Cui, H.; Fang, J.; Zuo, Z.; Deng, J.; Li, Y.; Wang, X.; Zhao, L. Inflammatory responses and inflammation-associated diseases in organs. *Oncotarget* **2018**, *9*, 7204–7218. [[CrossRef](#)] [[PubMed](#)]
7. Iwasaki, A.; Medzhitov, R. Toll-like receptor control of the adaptive immune responses. *Nat. Immunol.* **2004**, *5*, 987–995. [[CrossRef](#)] [[PubMed](#)]
8. Buckley, C.D.; Gilroy, D.W.; Serhan, C.N. Proresolving lipid mediators and mechanisms in the resolution of acute inflammation. *Immunity* **2014**, *40*, 315–327. [[CrossRef](#)] [[PubMed](#)]
9. Newson, J.; Stables, M.; Karra, E.; Arce-Vargas, F.; Quezada, S.; Motwani, M.; Mack, M.; Yona, S.; Audzevich, T.; Gilroy, D.W. Resolution of acute inflammation bridges the gap between innate and adaptive immunity. *Blood* **2014**, *124*, 1748–1764. [[CrossRef](#)]
10. Netea, M.G.; Quintin, J.; van der Meer, J.W. Trained immunity: A memory for innate host defense. *Cell Host Microbe* **2011**, *9*, 355–361. [[CrossRef](#)]
11. Doroudgar, S.; Glembotski, C.C. The cardiokine story unfolds: Ischemic stress-induced protein secretion in the heart. *Trends Mol. Med.* **2011**, *17*, 207–214. [[CrossRef](#)] [[PubMed](#)]
12. Wu, Y.S.; Zhu, B.; Luo, A.L.; Yang, L.; Yang, C. The Role of Cardiokines in Heart Diseases: Beneficial or Detrimental? *Biomed Res. Int.* **2018**, *2018*, 8207058. [[CrossRef](#)] [[PubMed](#)]
13. Giese, K.P.; Fedorov, N.B.; Filipkowski, R.K.; Silva, A.J. Autophosphorylation at Thr286 of the alpha calcium-calmodulin kinase II in LTP and learning. *Science* **1998**, *279*, 870–873. [[CrossRef](#)] [[PubMed](#)]



14. Haudek, S.B.; Taffet, G.E.; Schneider, M.D.; Mann, D.L. TNF provokes cardiomyocyte apoptosis and cardiac remodeling through activation of multiple cell death pathways. *J. Clin. Investig.* **2007**, *117*, 2692–2701. [[CrossRef](#)] [[PubMed](#)]
15. Hartupee, J.; Szalai, G.D.; Wang, W.; Ma, X.; Diwan, A.; Mann, D.L. Impaired Protein Quality Control During Left Ventricular Remodeling in Mice With Cardiac Restricted Overexpression of Tumor Necrosis Factor. *Circ. Heart Fail.* **2017**, *10*. [[CrossRef](#)] [[PubMed](#)]
16. Rathi, S.S.; Xu, Y.J.; Dhalla, N.S. Mechanism of cardioprotective action of TNF-alpha in the isolated rat heart. *Exp. Clin. Cardiol.* **2002**, *7*, 146–150.
17. Zhang, M.; Xu, Y.J.; Saini, H.K.; Turan, B.; Liu, P.P.; Dhalla, N.S. TNF-alpha as a potential mediator of cardiac dysfunction due to intracellular Ca<sup>2+</sup>-overload. *Biochem. Biophys. Res. Commun.* **2005**, *327*, 57–63. [[CrossRef](#)]
18. Turan, B.; Saini, H.K.; Zhang, M.; Prajapati, D.; Elimban, V.; Dhalla, N.S. Selenium improves cardiac function by attenuating the activation of NF-kappaB due to ischemia-Reperfus. injury. *Antioxid. Redox Signal.* **2005**, *7*, 1388–1397. [[CrossRef](#)]
19. Zhang, M.; Xu, Y.J.; Saini, H.K.; Turan, B.; Liu, P.P.; Dhalla, N.S. Pentoxifylline attenuates cardiac dysfunction and reduces TNF-alpha level in ischemic-reperfused heart. *Am. J. Physiol.* **2005**, *289*, H832–H839. [[CrossRef](#)]
20. Perrucci, G.L.; Barbagallo, V.A.; Corliano, M.; Tosi, D.; Santoro, R.; Nigro, P.; Poggio, P.; Bulfamante, G.; Lombardi, F.; Pompilio, G. Integrin alphanubeta5 in vitro inhibition limits pro-fibrotic response in cardiac fibroblasts of spontaneously hypertensive rats. *J. Transl. Med.* **2018**, *16*, 352. [[CrossRef](#)]
21. Gambini, E.; Perrucci, G.L.; Bassetti, B.; Spaltro, G.; Campostrini, G.; Lionetti, M.C.; Pillozzi, A.; Martinelli, F.; Farruggia, A.; DiFrancesco, D.; et al. Preferential myofibroblast differentiation of cardiac mesenchymal progenitor cells in the presence of atrial fibrillation. *Transl. Res.* **2018**, *192*, 54–67. [[CrossRef](#)] [[PubMed](#)]
22. Almendral, J.L.; Shick, V.; Rosendorff, C.; Atlas, S.A. Association between transforming growth factor-β(1) and left ventricular mass and diameter in hypertensive patients. *J. Am. Soc. Hypertens.* **2010**, *4*, 135–141. [[CrossRef](#)] [[PubMed](#)]
23. Ayca, B.; Sahin, I.; Kucuk, S.H.; Akin, F.; Kafadar, D.; Avsar, M.; Avci, I.I.; Gungor, B.; Okuyan, E.; Dinckal, M.H. Increased Transforming Growth Factor-beta Levels Associated With Cardiac Adverse Events in Hypertrophic Cardiomyopathy. *Clin. Cardiol.* **2015**, *38*, 371–377. [[CrossRef](#)] [[PubMed](#)]
24. Koitabashi, N.; Danner, T.; Zaiman, A.L.; Pinto, Y.M.; Rowell, J.; Mankowski, J.; Zhang, D.; Nakamura, T.; Takimoto, E.; Kass, D.A. Pivotal role of cardiomyocyte TGF-beta signaling in the murine pathological response to sustained pressure overload. *J. Clin. Investig.* **2011**, *121*, 2301–2312. [[CrossRef](#)] [[PubMed](#)]
25. Ding, Z.; Yuan, J.; Liang, Y.; Wu, J.; Gong, H.; Ye, Y.; Jiang, G.; Yin, P.; Li, Y.; Zhang, G.; et al. Ryanodine Receptor Type 2 Plays a Role in the Development of Cardiac Fibrosis under Mechanical Stretch Through TGFbeta-1. *Int. Heart J.* **2017**, *58*, 957–961. [[CrossRef](#)]
26. Bansal, T.; Chatterjee, E.; Singh, J.; Ray, A.; Kundu, B.; Thankamani, V.; Sengupta, S.; Sarkar, S. Arjunolic acid, a peroxisome proliferator-activated receptor alpha agonist, regresses cardiac fibrosis by inhibiting non-canonical TGF-beta signaling. *J. Biol. Chem.* **2017**, *292*, 16440–16462. [[CrossRef](#)] [[PubMed](#)]
27. Ciccone, M.M.; Cortese, F.; Gesualdo, M.; Riccardi, R.; Di Nunzio, D.; Moncelli, M.; Iacoviello, M.; Scicchitano, P. A novel cardiac bio-marker: ST2: A review. *Molecules* **2013**, *18*, 15314–15328. [[CrossRef](#)]
28. Sweet, M.J.; Leung, B.P.; Kang, D.; Sogaard, M.; Schulz, K.; Trajkovic, V.; Campbell, C.C.; Xu, D.; Liew, F.Y. A novel pathway regulating lipopolysaccharide-induced shock by ST2/T1 via inhibition of Toll-like receptor 4 expression. *J. Immunol.* **2001**, *166*, 6633–6639. [[CrossRef](#)]
29. Schmitz, J.; Owyang, A.; Oldham, E.; Song, Y.; Murphy, E.; McClanahan, T.K.; Zurawski, G.; Moshrefi, M.; Qin, J.; Li, X.; et al. IL-33, an interleukin-1-like cytokine that signals via the IL-1 receptor-related protein ST2 and induces T helper type 2-associated cytokines. *Immunity* **2005**, *23*, 479–490. [[CrossRef](#)]
30. Sanada, S.; Hakuno, D.; Higgins, L.J.; Schreiter, E.R.; McKenzie, A.N.; Lee, R.T. IL-33 and ST2 comprise a critical biomechanically induced and cardioprotective signaling system. *J. Clin. Investig.* **2007**, *117*, 1538–1549. [[CrossRef](#)]
31. Racioppi, L.; Means, A.R. Calcium/calmodulin-dependent protein kinase kinase 2: Roles in signaling and pathophysiology. *J. Biol. Chem.* **2012**, *287*, 31658–31665. [[CrossRef](#)] [[PubMed](#)]
32. Chao, L.H.; Stratton, M.M.; Lee, I.H.; Rosenberg, O.S.; Levitz, J.; Mandell, D.J.; Kortemme, T.; Groves, J.T.; Schulman, H.; Kuriyan, J. A mechanism for tunable autoinhibition in the structure of a human Ca<sup>2+</sup>/calmodulin- dependent kinase II holoenzyme. *Cell* **2011**, *146*, 732–745. [[CrossRef](#)] [[PubMed](#)]

33. Hudmon, A.; Schulman, H. Structure-function of the multifunctional Ca<sup>2+</sup>/calmodulin-dependent protein kinase II. *Biochem. J.* **2002**, *364*, 593–611. [[CrossRef](#)] [[PubMed](#)]
34. De Koninck, P.; Schulman, H. Sensitivity of CaM kinase II to the frequency of Ca<sup>2+</sup> oscillations. *Science* **1998**, *279*, 227–230. [[CrossRef](#)] [[PubMed](#)]
35. Erickson, J.R. Mechanisms of CaMKII Activation in the Heart. *Front. Pharmacol.* **2014**, *5*, 59. [[CrossRef](#)] [[PubMed](#)]
36. Rosenberg, O.S.; Deindl, S.; Sung, R.J.; Nairn, A.C.; Kuriyan, J. Structure of the autoinhibited kinase domain of CaMKII and SAXS analysis of the holoenzyme. *Cell* **2005**, *123*, 849–860. [[CrossRef](#)]
37. Lai, Y.; Nairn, A.C.; Gorelick, F.; Greengard, P. Ca<sup>2+</sup>/calmodulin-dependent protein kinase II: Identification of autophosphorylation sites responsible for generation of Ca<sup>2+</sup>/calmodulin-independence. *Proc. Natl. Acad. Sci. USA* **1987**, *84*, 5710–5714. [[CrossRef](#)]
38. Meyer, T.; Hanson, P.I.; Stryer, L.; Schulman, H. Calmodulin trapping by calcium-calmodulin-dependent protein kinase. *Science* **1992**, *256*, 1199–1202. [[CrossRef](#)]
39. Erickson, J.R.; Joiner, M.L.; Guan, X.; Kutschke, W.; Yang, J.; Oddis, C.V.; Bartlett, R.K.; Lowe, J.S.; O'Donnell, S.E.; Aykin-Burns, N.; et al. A dynamic pathway for calcium-independent activation of CaMKII by methionine oxidation. *Cell* **2008**, *133*, 462–474. [[CrossRef](#)]
40. Luo, M.; Guan, X.; Luczak, E.D.; Lang, D.; Kutschke, W.; Gao, Z.; Yang, J.; Glynn, P.; Sossalla, S.; Swaminathan, P.D.; et al. Diabetes increases mortality after myocardial infarction by oxidizing CaMKII. *J. Clin. Investig.* **2013**, *123*, 1262–1274. [[CrossRef](#)]
41. Erickson, J.R.; Pereira, L.; Wang, L.; Han, G.; Ferguson, A.; Dao, K.; Copeland, R.J.; Despa, F.; Hart, G.W.; Ripplinger, C.M.; et al. Diabetic hyperglycaemia activates CaMKII and arrhythmias by O-linked glycosylation. *Nature* **2013**, *502*, 372–376. [[CrossRef](#)] [[PubMed](#)]
42. Erickson, J.R.; Nichols, C.B.; Uchinoumi, H.; Stein, M.L.; Bossuyt, J.; Bers, D.M. S-Nitrosylation Induces Both Autonomous Activation and Inhibition of Calcium/Calmodulin-dependent Protein Kinase II delta. *J. Biol. Chem.* **2015**, *290*, 25646–25656. [[CrossRef](#)] [[PubMed](#)]
43. Strack, S.; Barban, M.A.; Wadzinski, B.E.; Colbran, R.J. Differential inactivation of postsynaptic density-associated and soluble Ca<sup>2+</sup>/calmodulin-dependent protein kinase II by protein phosphatases 1 and 2A. *J. Neurochem.* **1997**, *68*, 2119–2128. [[CrossRef](#)] [[PubMed](#)]
44. Colbran, R.J.; Smith, M.K.; Schworer, C.M.; Fong, Y.L.; Soderling, T.R. Regulatory domain of calcium/calmodulin-dependent protein kinase II. Mechanism of inhibition and regulation by phosphorylation. *J. Biol. Chem.* **1989**, *264*, 4800–4804. [[PubMed](#)]
45. Rellos, P.; Pike, A.C.; Niesen, F.H.; Salah, E.; Lee, W.H.; von Delft, F.; Knapp, S. Structure of the CaMKIIdelta/calmodulin complex reveals the molecular mechanism of CaMKII kinase activation. *PLoS Biol.* **2010**, *8*, e1000426. [[CrossRef](#)] [[PubMed](#)]
46. Tombes, R.M.; Faison, M.O.; Turbeville, J.M. Organization and evolution of multifunctional Ca(2+)/CaM-dependent protein kinase genes. *Gene* **2003**, *322*, 17–31. [[CrossRef](#)] [[PubMed](#)]
47. Hudmon, A.; Schulman, H. Neuronal CA<sup>2+</sup>/calmodulin-dependent protein kinase II: The role of structure and autoregulation in cellular function. *Annu. Rev. Biochem.* **2002**, *71*, 473–510. [[CrossRef](#)] [[PubMed](#)]
48. Backs, J.; Backs, T.; Neef, S.; Kreusser, M.M.; Lehmann, L.H.; Patrick, D.M.; Grueter, C.E.; Qi, X.; Richardson, J.A.; Hill, J.A.; et al. The delta isoform of CaM kinase II is required for pathological cardiac hypertrophy and remodeling after pressure overload. *Proc. Natl. Acad. Sci. USA* **2009**, *106*, 2342–2347. [[CrossRef](#)]
49. Edman, C.F.; Schulman, H. Identification and characterization of delta B-CaM kinase and delta C-CaM kinase from rat heart, two new multifunctional Ca<sup>2+</sup>/calmodulin-dependent protein kinase isoforms. *Biochim. Biophys. Acta* **1994**, *1221*, 89–101. [[CrossRef](#)]
50. Li, C.; Cai, X.; Sun, H.; Bai, T.; Zheng, X.; Zhou, X.W.; Chen, X.; Gill, D.L.; Li, J.; Tang, X.D. The deltaA isoform of calmodulin kinase II mediates pathological cardiac hypertrophy by interfering with the HDAC4-MEF2 signaling pathway. *Biochem. Biophys. Res. Commun.* **2011**, *409*, 125–130. [[CrossRef](#)]
51. Zhang, T.; Kohlhaas, M.; Backs, J.; Mishra, S.; Phillips, W.; Dybkova, N.; Chang, S.; Ling, H.; Bers, D.M.; Maier, L.S.; et al. CaMKIIdelta isoforms differentially affect calcium handling but similarly regulate HDAC/MEF2 transcriptional responses. *J. Biol. Chem.* **2007**, *282*, 35078–35087. [[CrossRef](#)] [[PubMed](#)]

52. Sossalla, S.; Maurer, U.; Schotola, H.; Hartmann, N.; Didie, M.; Zimmermann, W.H.; Jacobshagen, C.; Wagner, S.; Maier, L.S. Diastolic dysfunction and arrhythmias caused by overexpression of CaMKII $\delta$ (C) can be reversed by inhibition of late Na<sup>+</sup> current. *Basic Res. Cardiol.* **2011**, *106*, 263–272. [[CrossRef](#)] [[PubMed](#)]
53. Maione, A.S.; Cipolletta, E.; Sorriento, D.; Borriello, F.; Soprano, M.; Rusciano, M.R.; D'Esposito, V.; Markabaoui, A.K.; De Palma, G.D.; Martino, G.; et al. Cellular subtype expression and activation of CaMKII regulate the fate of atherosclerotic plaque. *Atherosclerosis* **2017**, *256*, 53–61. [[CrossRef](#)] [[PubMed](#)]
54. Cipolletta, E.; Monaco, S.; Maione, A.S.; Vitiello, L.; Campiglia, P.; Pastore, L.; Franchini, C.; Novellino, E.; Limongelli, V.; Bayer, K.U.; et al. Calmodulin-dependent kinase II mediates vascular smooth muscle cell proliferation and is potentiated by extracellular signal regulated kinase. *Endocrinology* **2010**, *151*, 2747–2759. [[CrossRef](#)] [[PubMed](#)]
55. Monaco, S.; Rusciano, M.R.; Maione, A.S.; Soprano, M.; Gomathinayagam, R.; Todd, L.R.; Campiglia, P.; Salzano, S.; Pastore, L.; Leggiero, E.; et al. A novel crosstalk between calcium/calmodulin kinases II and IV regulates cell proliferation in myeloid leukemia cells. *Cell. Signal.* **2015**, *27*, 204–214. [[CrossRef](#)] [[PubMed](#)]
56. Saddouk, F.Z.; Sun, L.Y.; Liu, Y.F.; Jiang, M.; Singer, D.V.; Backs, J.; Van Riper, D.; Ginnan, R.; Schwarz, J.J.; Singer, H.A. Ca<sup>2+</sup>/calmodulin-dependent protein kinase II-gamma (CaMKII $\gamma$ ) negatively regulates vascular smooth muscle cell proliferation and vascular remodeling. *FASEB J.* **2016**, *30*, 1051–1064. [[CrossRef](#)] [[PubMed](#)]
57. Uosaki, H.; Magadam, A.; Seo, K.; Fukushima, H.; Takeuchi, A.; Nakagawa, Y.; Moyes, K.W.; Narazaki, G.; Kuwahara, K.; Laflamme, M.; et al. Identification of chemicals inducing cardiomyocyte proliferation in developmental stage-specific manner with pluripotent stem cells. *Circ. Cardiovasc. Genet.* **2013**, *6*, 624–633. [[CrossRef](#)]
58. Sloan-Lancaster, J.; Allen, P.M. Altered peptide ligand-induced partial T cell activation: Molecular mechanisms and role in T cell biology. *Annu. Rev. Immunol.* **1996**, *14*, 1–27. [[CrossRef](#)]
59. Schwartz, R.H. Models of T cell anergy: Is there a common molecular mechanism? *J. Exp. Med.* **1996**, *184*, 1–8. [[CrossRef](#)]
60. Bullens, D.M.; Rafiq, K.; Charitidou, L.; Peng, X.; Kasran, A.; Warmerdam, P.A.; Van Gool, S.W.; Ceuppens, J.L. Effects of co-stimulation by CD58 on human T cell cytokine production: A selective cytokine pattern with induction of high IL-10 production. *Int. Immunol.* **2001**, *13*, 181–191. [[CrossRef](#)]
61. Rafiq, K.; Charitidou, L.; Bullens, D.M.; Kasran, A.; Lorre, K.; Ceuppens, J.; van Gool, S.W. Regulation of the IL-10 production by human T cells. *Scand. J. Immunol.* **2001**, *53*, 139–147. [[CrossRef](#)] [[PubMed](#)]
62. McConkey, D.J.; Orrenius, S. The role of calcium in the regulation of apoptosis. *Biochem. Biophys. Res. Commun.* **1997**, *239*, 357–366. [[CrossRef](#)] [[PubMed](#)]
63. Hama, N.; Paliogianni, F.; Fessler, B.J.; Boumpas, D.T. Calcium/calmodulin-dependent protein kinase II downregulates both calcineurin and protein kinase C-mediated pathways for cytokine gene transcription in human T cells. *J. Exp. Med.* **1995**, *181*, 1217–1222. [[CrossRef](#)] [[PubMed](#)]
64. Nghiem, P.; Ollick, T.; Gardner, P.; Schulman, H. Interleukin-2 transcriptional block by multifunctional Ca<sup>2+</sup>/calmodulin kinase. *Nature* **1994**, *371*, 347–350. [[CrossRef](#)] [[PubMed](#)]
65. Lin, M.Y.; Zal, T.; Ch'en, I.L.; Gascoigne, N.R.; Hedrick, S.M. A pivotal role for the multifunctional calcium/calmodulin-dependent protein kinase II in T cells: From activation to unresponsiveness. *J. Immunol.* **2005**, *174*, 5583–5592. [[CrossRef](#)] [[PubMed](#)]
66. Bui, J.D.; Calbo, S.; Hayden-Martinez, K.; Kane, L.P.; Gardner, P.; Hedrick, S.M. A role for CaMKII in T cell memory. *Cell* **2000**, *100*, 457–467. [[CrossRef](#)]
67. Singh, M.V.; Swaminathan, P.D.; Luczak, E.D.; Kutschke, W.; Weiss, R.M.; Anderson, M.E. MyD88 mediated inflammatory signaling leads to CaMKII oxidation, cardiac hypertrophy and death after myocardial infarction. *J. Mol. Cell. Cardiol.* **2012**, *52*, 1135–1144. [[CrossRef](#)]
68. Boubali, S.; Liopeta, K.; Virgilio, L.; Thyphronitis, G.; Mavrothalassitis, G.; Dimitracopoulos, G.; Paliogianni, F. Calcium/calmodulin-dependent protein kinase II regulates IL-10 production by human T lymphocytes: A distinct target in the calcium dependent pathway. *Mol. Immunol.* **2012**, *52*, 51–60. [[CrossRef](#)]
69. Liu, X.; Yao, M.; Li, N.; Wang, C.; Zheng, Y.; Cao, X. CaMKII promotes TLR-triggered proinflammatory cytokine and type I interferon production by directly binding and activating TAK1 and IRF3 in macrophages. *Blood* **2008**, *112*, 4961–4970. [[CrossRef](#)]

70. Pereira, C.; Schaer, D.J.; Bachli, E.B.; Kurrer, M.O.; Schoedon, G. Wnt5A/CaMKII signaling contributes to the inflammatory response of macrophages and is a target for the antiinflammatory action of activated protein C and interleukin-10. *Arterioscler. Thromb. Vasc. Biol.* **2008**, *28*, 504–510. [[CrossRef](#)]
71. Herrmann, T.L.; Agrawal, R.S.; Connolly, S.F.; McCaffrey, R.L.; Schlomann, J.; Kusner, D.J. MHC Class II levels and intracellular localization in human dendritic cells are regulated by calmodulin kinase II. *J. Leukoc. Biol.* **2007**, *82*, 686–699. [[CrossRef](#)] [[PubMed](#)]
72. Herrmann, T.L.; Morita, C.T.; Lee, K.; Kusner, D.J. Calmodulin kinase II regulates the maturation and antigen presentation of human dendritic cells. *J. Leukoc. Biol.* **2005**, *78*, 1397–1407. [[CrossRef](#)] [[PubMed](#)]
73. Ren, G.; Dewald, O.; Frangogiannis, N.G. Inflammatory mechanisms in myocardial infarction. *Curr. Drug Targets-Inflamm. Allergy* **2003**, *2*, 242–256. [[CrossRef](#)] [[PubMed](#)]
74. Frangogiannis, N.G.; Smith, C.W.; Entman, M.L. The inflammatory response in myocardial infarction. *Cardiovasc. Res.* **2002**, *53*, 31–47. [[CrossRef](#)]
75. Rokita, A.G.; Anderson, M.E. New therapeutic targets in cardiology: Arrhythmias and Ca<sup>2+</sup>/calmodulin-dependent kinase II (CaMKII). *Circulation* **2012**, *126*, 2125–2139. [[CrossRef](#)] [[PubMed](#)]
76. Zhang, R.; Khoo, M.S.; Wu, Y.; Yang, Y.; Grueter, C.E.; Ni, G.; Price, E.E., Jr.; Thiel, W.; Guatimosim, S.; Song, L.S.; et al. Calmodulin kinase II inhibition protects against structural heart disease. *Nat. Med.* **2005**, *11*, 409–417. [[CrossRef](#)]
77. Cipolletta, E.; Rusciano, M.R.; Maione, A.S.; Santulli, G.; Sorriento, D.; Del Giudice, C.; Ciccarelli, M.; Franco, A.; Crola, C.; Campiglia, P.; et al. Targeting the CaMKII/ERK Interaction in the Heart Prevents Cardiac Hypertrophy. *PLoS ONE* **2015**, *10*, e0130477. [[CrossRef](#)]
78. Zhu, W.Z.; Wang, S.Q.; Chakir, K.; Yang, D.; Zhang, T.; Brown, J.H.; Devic, E.; Kobilka, B.K.; Cheng, H.; Xiao, R.P. Linkage of beta1-adrenergic stimulation to apoptotic heart cell death through protein kinase A-independent activation of Ca<sup>2+</sup>/calmodulin kinase II. *J. Clin. Investig.* **2003**, *111*, 617–625. [[CrossRef](#)]
79. Yang, Y.; Zhu, W.Z.; Joiner, M.L.; Zhang, R.; Oddis, C.V.; Hou, Y.; Yang, J.; Price, E.E.; Gleaves, L.; Eren, M.; et al. Calmodulin kinase II inhibition protects against myocardial cell apoptosis in vivo. *Am. J. Physiol.* **2006**, *291*, H3065–H3075. [[CrossRef](#)]
80. Di Carlo, M.N.; Said, M.; Ling, H.; Valverde, C.A.; De Giusti, V.C.; Sommese, L.; Palomeque, J.; Aiello, E.A.; Skapura, D.G.; Rinaldi, G.; et al. CaMKII-dependent phosphorylation of cardiac ryanodine receptors regulates cell death in cardiac ischemia/reperfusion injury. *J. Mol. Cell. Cardiol.* **2014**, *74*, 274–283. [[CrossRef](#)]
81. Koval, O.M.; Guan, X.; Wu, Y.; Joiner, M.L.; Gao, Z.; Chen, B.; Grumbach, I.M.; Luczak, E.D.; Colbran, R.J.; Song, L.S.; et al. CaV1.2 beta-subunit coordinates CaMKII-triggered cardiomyocyte death and afterdepolarizations. *Proc. Natl. Acad. Sci. USA* **2010**, *107*, 4996–5000. [[CrossRef](#)] [[PubMed](#)]
82. Zhu, W.; Woo, A.Y.; Yang, D.; Cheng, H.; Crow, M.T.; Xiao, R.P. Activation of CaMKIIdeltaC is a common intermediate of diverse death stimuli-induced heart muscle cell apoptosis. *J. Biol. Chem.* **2007**, *282*, 10833–10839. [[CrossRef](#)] [[PubMed](#)]
83. Schaefer, L. Complexity of danger: The diverse nature of damage-associated molecular patterns. *J. Biol. Chem.* **2014**, *289*, 35237–35245. [[CrossRef](#)] [[PubMed](#)]
84. Arslan, F.; de Kleijn, D.P.; Pasterkamp, G. Innate immune signaling in cardiac ischemia. *Nat. Rev. Cardiol.* **2011**, *8*, 292–300. [[CrossRef](#)] [[PubMed](#)]
85. Ma, Y.; Yabluchanskiy, A.; Iyer, R.P.; Cannon, P.L.; Flynn, E.R.; Jung, M.; Henry, J.; Cates, C.A.; DeLeon-Pennell, K.Y.; Lindsey, M.L. Temporal neutrophil polarization following myocardial infarction. *Cardiovasc. Res.* **2016**, *110*, 51–61. [[CrossRef](#)]
86. Zhang, X.; Mosser, D.M. Macrophage activation by endogenous danger signals. *J. Pathol.* **2008**, *214*, 161–178. [[CrossRef](#)] [[PubMed](#)]
87. Liu, L.; Wang, Y.; Cao, Z.Y.; Wang, M.M.; Liu, X.M.; Gao, T.; Hu, Q.K.; Yuan, W.J.; Lin, L. Up-regulated TLR4 in cardiomyocytes exacerbates heart failure after long-term myocardial infarction. *J. Cell. Mol. Med.* **2015**, *19*, 2728–2740. [[CrossRef](#)]
88. Yoo, B.; Lemaire, A.; Mangmool, S.; Wolf, M.J.; Curcio, A.; Mao, L.; Rockman, H.A. Beta1-adrenergic receptors stimulate cardiac contractility and CaMKII activation in vivo and enhance cardiac dysfunction following myocardial infarction. *Am. J. Physiol. Heart Circ. Physiol.* **2009**, *297*, H1377–H1386. [[CrossRef](#)]
89. Christensen, M.D.; Dun, W.; Boyden, P.A.; Anderson, M.E.; Mohler, P.J.; Hund, T.J. Oxidized calmodulin kinase II regulates conduction following myocardial infarction: A computational analysis. *PLoS Comput. Biol.* **2009**, *5*, e1000583. [[CrossRef](#)]

90. Singh, M.V.; Kapoun, A.; Higgins, L.; Kutschke, W.; Thurman, J.M.; Zhang, R.; Singh, M.; Yang, J.; Guan, X.; Lowe, J.S.; et al.  $\text{Ca}^{2+}$ /calmodulin-dependent kinase II triggers cell membrane injury by inducing complement factor B gene expression in the mouse heart. *J. Clin. Investig.* **2009**, *119*, 986–996. [[CrossRef](#)]
91. Murphy, E.; Steenbergen, C. Mechanisms underlying acute protection from cardiac ischemia-reperfusion injury. *Physiol. Rev.* **2008**, *88*, 581–609. [[CrossRef](#)] [[PubMed](#)]
92. Weiss, J.N.; Korge, P.; Honda, H.M.; Ping, P. Role of the mitochondrial permeability transition in myocardial disease. *Circ. Res.* **2003**, *93*, 292–301. [[CrossRef](#)] [[PubMed](#)]
93. Ling, H.; Gray, C.B.; Zambon, A.C.; Grimm, M.; Gu, Y.; Dalton, N.; Purcell, N.H.; Peterson, K.; Brown, J.H.  $\text{Ca}^{2+}$ /Calmodulin-dependent protein kinase II delta mediates myocardial ischemia/reperfusion injury through nuclear factor-kappaB. *Circ. Res.* **2013**, *112*, 935–944. [[CrossRef](#)] [[PubMed](#)]
94. Weinreuter, M.; Kreuzer, M.M.; Beckendorf, J.; Schreiter, F.C.; Leuschner, F.; Lehmann, L.H.; Hofmann, K.P.; Rostovsky, J.S.; Diemert, N.; Xu, C.; et al. CaM Kinase II mediates maladaptive post-infarct remodeling and pro-inflammatory chemoattractant signaling but not acute myocardial ischemia/reperfusion injury. *EMBO Mol. Med.* **2014**, *6*, 1231–1245. [[CrossRef](#)] [[PubMed](#)]
95. Nevers, T.; Salvador, A.M.; Grodecki-Pena, A.; Knapp, A.; Velazquez, F.; Aronovitz, M.; Kapur, N.K.; Karas, R.H.; Blanton, R.M.; Alcaide, P. Left Ventricular T-Cell Recruitment Contributes to the Pathogenesis of Heart Failure. *Circ. Heart Fail.* **2015**, *8*, 776–787. [[CrossRef](#)] [[PubMed](#)]
96. Patel, B.; Ismahil, M.A.; Hamid, T.; Bansal, S.S.; Prabhu, S.D. Mononuclear Phagocytes Are Dispensable for Cardiac Remodeling in Established Pressure-Overload Heart Failure. *PLoS ONE* **2017**, *12*, e0170781. [[CrossRef](#)] [[PubMed](#)]
97. Kai, H.; Mori, T.; Tokuda, K.; Takayama, N.; Tahara, N.; Takemiya, K.; Kudo, H.; Sugi, Y.; Fukui, D.; Yasukawa, H.; et al. Pressure overload-induced transient oxidative stress mediates perivascular inflammation and cardiac fibrosis through angiotensin II. *Hypertens. Res.* **2006**, *29*, 711–718. [[CrossRef](#)] [[PubMed](#)]
98. Duerschmid, C.; Trial, J.; Wang, Y.; Entman, M.L.; Haudek, S.B. Tumor necrosis factor: A mechanistic link between angiotensin-II-induced cardiac inflammation and fibrosis. *Circ. Heart Fail.* **2015**, *8*, 352–361. [[CrossRef](#)] [[PubMed](#)]
99. Willeford, A.; Suetomi, T.; Nickle, A.; Hoffman, H.M.; Miyamoto, S.; Heller Brown, J. CaMKII $\delta$ -mediated inflammatory gene expression and inflammasome activation in cardiomyocytes initiate inflammation and induce fibrosis. *JCI Insight* **2018**, *3*. [[CrossRef](#)] [[PubMed](#)]
100. Palomeque, J.; Rueda, O.V.; Sapia, L.; Valverde, C.A.; Salas, M.; Petroff, M.V.; Mattiazzi, A. Angiotensin II-induced oxidative stress resets the  $\text{Ca}^{2+}$  dependence of  $\text{Ca}^{2+}$ -calmodulin protein kinase II and promotes a death pathway conserved across different species. *Circ. Res.* **2009**, *105*, 1204–1212. [[CrossRef](#)]
101. Vangheluwe, P.; Sipido, K.R.; Raeymaekers, L.; Wuytack, F. New perspectives on the role of SERCA2's  $\text{Ca}^{2+}$  affinity in cardiac function. *Biochim. Biophys. Acta* **2006**, *1763*, 1216–1228. [[CrossRef](#)] [[PubMed](#)]
102. Toischer, K.; Rokita, A.G.; Unsold, B.; Zhu, W.; Kararigas, G.; Sossalla, S.; Reuter, S.P.; Becker, A.; Teucher, N.; Seidler, T.; et al. Differential cardiac remodeling in preload versus afterload. *Circulation* **2010**, *122*, 993–1003. [[CrossRef](#)] [[PubMed](#)]
103. Wang, Y.; Tandan, S.; Cheng, J.; Yang, C.; Nguyen, L.; Sugianto, J.; Johnstone, J.L.; Sun, Y.; Hill, J.A.  $\text{Ca}^{2+}$ /calmodulin-dependent protein kinase II-dependent remodeling of  $\text{Ca}^{2+}$  current in pressure overload heart failure. *J. Biol. Chem.* **2008**, *283*, 25524–25532. [[CrossRef](#)] [[PubMed](#)]
104. Wang, Z.; Kutschke, W.; Richardson, K.E.; Karimi, M.; Hill, J.A. Electrical remodeling in pressure-overload cardiac hypertrophy: Role of calcineurin. *Circulation* **2001**, *104*, 1657–1663. [[CrossRef](#)] [[PubMed](#)]
105. Suetomi, T.; Willeford, A.; Brand, C.S.; Cho, Y.; Ross, R.S.; Miyamoto, S.; Brown, J.H. Inflammation and NLRP3 Inflammasome Activation Initiated in Response to Pressure Overload by  $\text{Ca}^{2+}$ /Calmodulin-Dependent Protein Kinase II  $\delta$  Signaling in Cardiomyocytes Are Essential for Adverse Cardiac Remodeling. *Circulation* **2018**, *138*, 2530–2544. [[CrossRef](#)] [[PubMed](#)]
106. Ling, H.; Zhang, T.; Pereira, L.; Means, C.K.; Cheng, H.; Gu, Y.; Dalton, N.D.; Peterson, K.L.; Chen, J.; Bers, D.; et al. Requirement for  $\text{Ca}^{2+}$ /calmodulin-dependent kinase II in the transition from pressure overload-induced cardiac hypertrophy to heart failure in mice. *J. Clin. Investig.* **2009**, *119*, 1230–1240. [[CrossRef](#)] [[PubMed](#)]
107. Cardinale, J.P.; Sriramula, S.; Pariaut, R.; Guggilam, A.; Mariappan, N.; Elks, C.M.; Francis, J. HDAC inhibition attenuates inflammatory, hypertrophic, and hypertensive responses in spontaneously hypertensive rats. *Hypertension* **2010**, *56*, 437–444. [[CrossRef](#)] [[PubMed](#)]

108. Zhang, C.L.; McKinsey, T.A.; Chang, S.; Antos, C.L.; Hill, J.A.; Olson, E.N. Class II histone deacetylases act as signal-responsive repressors of cardiac hypertrophy. *Cell* **2002**, *110*, 479–488. [[CrossRef](#)]
109. Backs, J.; Song, K.; Bezprozvannaya, S.; Chang, S.; Olson, E.N. CaM kinase II selectively signals to histone deacetylase 4 during cardiomyocyte hypertrophy. *J. Clin. Investig.* **2006**, *116*, 1853–1864. [[CrossRef](#)] [[PubMed](#)]
110. Zhang, W.; Chen, D.Q.; Qi, F.; Wang, J.; Xiao, W.Y.; Zhu, W.Z. Inhibition of calcium-calmodulin-dependent kinase II suppresses cardiac fibroblast proliferation and extracellular matrix secretion. *J. Cardiovasc. Pharmacol.* **2010**, *55*, 96–105. [[CrossRef](#)]
111. Valverde, C.A.; Mundina-Weilenmann, C.; Reyes, M.; Kranias, E.G.; Escobar, A.L.; Mattiazzi, A. Phospholamban phosphorylation sites enhance the recovery of intracellular Ca<sup>2+</sup> after perfusion arrest in isolated, perfused mouse heart. *Cardiovasc. Res.* **2006**, *70*, 335–345. [[CrossRef](#)] [[PubMed](#)]
112. Neef, S.; Mann, C.; Zwenger, A.; Dybkova, N.; Maier, L.S. Reduction of SR Ca<sup>2+</sup> leak and arrhythmogenic cellular correlates by SMP-114, a novel CaMKII inhibitor with oral bioavailability. *Basic Res. Cardiol.* **2017**, *112*, 45. [[CrossRef](#)] [[PubMed](#)]
113. Greensmith, D.J.; Nirmalan, M. The effects of tumor necrosis factor-alpha on systolic and diastolic function in rat ventricular myocytes. *Physiol. Rep.* **2013**, *1*, e00093. [[CrossRef](#)] [[PubMed](#)]
114. Duncan, D.J.; Yang, Z.; Hopkins, P.M.; Steele, D.S.; Harrison, S.M. TNF- $\alpha$  and IL-1 $\beta$  increase Ca<sup>2+</sup> leak from the sarcoplasmic reticulum and susceptibility to arrhythmia in rat ventricular myocytes. *Cell Calcium* **2010**, *47*, 378–386. [[CrossRef](#)]
115. Lenski, M.; Schleider, G.; Kohlhaas, M.; Adrian, L.; Adam, O.; Tian, Q.; Kaestner, L.; Lipp, P.; Lehrke, M.; Maack, C.; et al. Arrhythmia causes lipid accumulation and reduced glucose uptake. *Basic Res. Cardiol.* **2015**, *110*, 40. [[CrossRef](#)]
116. Oakes, R.S.; Badger, T.J.; Kholmovski, E.G.; Akoum, N.; Burgon, N.S.; Fish, E.N.; Blauer, J.J.; Rao, S.N.; DiBella, E.V.; Segerson, N.M.; et al. Detection and quantification of left atrial structural remodeling with delayed-enhancement magnetic resonance imaging in patients with atrial fibrillation. *Circulation* **2009**, *119*, 1758–1767. [[CrossRef](#)] [[PubMed](#)]
117. Issac, T.T.; Dokainish, H.; Lakkis, N.M. Role of inflammation in initiation and perpetuation of atrial fibrillation: A systematic review of the published data. *J. Am. Coll. Cardiol.* **2007**, *50*, 2021–2028. [[CrossRef](#)]
118. Kao, Y.H.; Chen, Y.C.; Cheng, C.C.; Lee, T.I.; Chen, Y.J.; Chen, S.A. Tumor necrosis factor-alpha decreases sarcoplasmic reticulum Ca<sup>2+</sup>-ATPase expressions via the promoter methylation in cardiomyocytes. *Crit. Care Med.* **2010**, *38*, 217–222. [[CrossRef](#)]
119. Voigt, N.; Li, N.; Wang, Q.; Wang, W.; Trafford, A.W.; Abu-Taha, I.; Sun, Q.; Wieland, T.; Ravens, U.; Nattel, S.; et al. Enhanced sarcoplasmic reticulum Ca<sup>2+</sup> leak and increased Na<sup>+</sup>-Ca<sup>2+</sup> exchanger function underlie delayed afterdepolarizations in patients with chronic atrial fibrillation. *Circulation* **2012**, *125*, 2059–2070. [[CrossRef](#)]
120. Purohit, A.; Rokita, A.G.; Guan, X.; Chen, B.; Koval, O.M.; Voigt, N.; Neef, S.; Sowa, T.; Gao, Z.; Luczak, E.D.; et al. Oxidized Ca<sup>2+</sup>/calmodulin-dependent protein kinase II triggers atrial fibrillation. *Circulation* **2013**, *128*, 1748–1757. [[CrossRef](#)]
121. Hove-Madsen, L.; Llach, A.; Bayes-Genis, A.; Roura, S.; Rodriguez Font, E.; Aris, A.; Cinca, J. Atrial fibrillation is associated with increased spontaneous calcium release from the sarcoplasmic reticulum in human atrial myocytes. *Circulation* **2004**, *110*, 1358–1363. [[CrossRef](#)] [[PubMed](#)]
122. Zuo, S.; Li, L.L.; Ruan, Y.F.; Jiang, L.; Li, X.; Li, S.N.; Wen, S.N.; Bai, R.; Liu, N.; Du, X.; et al. Acute administration of tumour necrosis factor-alpha induces spontaneous calcium release via the reactive oxygen species pathway in atrial myocytes. *EP Eur.* **2018**, *20*, 1367–1374. [[CrossRef](#)]
123. Mesubi, O.O.; Anderson, M.E. Atrial remodelling in atrial fibrillation: CaMKII as a nodal proarrhythmic signal. *Cardiovasc. Res.* **2016**, *109*, 542–557. [[CrossRef](#)] [[PubMed](#)]
124. Fischer, T.H.; Herting, J.; Mason, F.E.; Hartmann, N.; Watanabe, S.; Nikolaev, V.O.; Sprenger, J.U.; Fan, P.; Yao, L.; Popov, A.F.; et al. Late INa increases diastolic SR-Ca<sup>2+</sup>-leak in atrial myocardium by activating PKA and CaMKII. *Cardiovasc. Res.* **2015**, *107*, 184–196. [[CrossRef](#)] [[PubMed](#)]
125. Coppini, R.; Mazzoni, L.; Ferrantini, C.; Gentile, F.; Pioner, J.M.; Laurino, A.; Santini, L.; Bargelli, V.; Rotellini, M.; Bartolucci, G.; et al. Ranolazine Prevents Phenotype Development in a Mouse Model of Hypertrophic Cardiomyopathy. *Circ. Heart Fail.* **2017**, *10*. [[CrossRef](#)] [[PubMed](#)]

126. Packer, M.; Bristow, M.R.; Cohn, J.N.; Colucci, W.S.; Fowler, M.B.; Gilbert, E.M.; Shusterman, N.H. The effect of carvedilol on morbidity and mortality in patients with chronic heart failure. U.S. Carvedilol Heart Failure Study Group. *New Engl. J. Med.* **1996**, *334*, 1349–1355. [[CrossRef](#)] [[PubMed](#)]
127. Bin-Dayel, A.F.; Abdel Baky, N.A.; Fadda, L.M.; Mohammad, R.A.; Al-Mohanna, F. Effect of aliskiren and carvedilol on expression of Ca<sup>2+</sup>/calmodulin-dependent protein kinase II delta-subunit isoforms in cardiac hypertrophy rat model. *Toxicol. Mech. Methods* **2016**, *26*, 122–131. [[CrossRef](#)]
128. Velez Rueda, J.O.; Palomeque, J.; Mattiazzi, A. Early apoptosis in different models of cardiac hypertrophy induced by high renin-angiotensin system activity involves CaMKII. *J. Appl. Physiol.* **2012**, *112*, 2110–2120. [[CrossRef](#)]
129. He, B.J.; Joiner, M.L.; Singh, M.V.; Luczak, E.D.; Swaminathan, P.D.; Koval, O.M.; Kutschke, W.; Allamargot, C.; Yang, J.; Guan, X.; et al. Oxidation of CaMKII determines the cardiotoxic effects of aldosterone. *Nat. Med.* **2011**, *17*, 1610–1618. [[CrossRef](#)]
130. Huang, F.C.; Kuo, H.C.; Huang, Y.H.; Yu, H.R.; Li, S.C. Anti-inflammatory effect of resveratrol in human coronary arterial endothelial cells via induction of autophagy: Implication for the treatment of Kawasaki disease. *BMC Pharmacol. Toxicol.* **2017**, *18*, 3. [[CrossRef](#)]
131. Li, W.; Wang, Y.P.; Gao, L.; Zhang, P.P.; Zhou, Q.; Xu, Q.F.; Zhou, Z.W.; Guo, K.; Chen, R.H.; Yang, H.T.; et al. Resveratrol protects rabbit ventricular myocytes against oxidative stress-induced arrhythmogenic activity and Ca<sup>2+</sup> overload. *Acta Pharmacol. Sin.* **2013**, *34*, 1164–1173. [[CrossRef](#)] [[PubMed](#)]



© 2019 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (<http://creativecommons.org/licenses/by/4.0/>).