

Figure 1. Immunophenotyping gating strategy of Regulated Cell Death. Live (including early apoptotic) and dead cells including late apoptotic and necrotic were gated from a Zombie NIR vs. Caspase-3-BV650 dot-plot (A). Live and dead resting were defined as Caspase-3^{-ve}/RIP3^{+ve}, early or late apoptotic (Caspase-3^{+ve}/RIP3^{-ve}), RIP1-dependent apoptotic (RIP1-APO, Caspase-3^{+ve}/RIP3^{+ve}) and Double Negative (DN, Caspase-3^{-ve}/RIP3^{-ve}) (B, D). Then each of these live and dead cell populations were then gated on a H2AX vs. cleaved PARP dot-plots to show the incidence of DDR (H2AX^{+ve}/PARP^{-ve}), hyper-activation of cleaved PARP (pH2AX^{+ve}/PARP^{+ve}), apoptosis via cleaved PARP (H2AX^{-ve}/PARP^{+ve}) and DN (pH2AX^{-ve}/PARP^{-ve}) (D, E). Dead resting oncotic (F, Zombie^{+ve}/Caspase-3^{-ve}/RIP3^{+ve}) and dead oncotic DN cells (G, Zombie^{+ve}/Caspase-3^{-ve}/RIP3^{-ve}) were then analysed on pH2AX vs. cleaved PARP dot-plots (QN, quadruple negative).

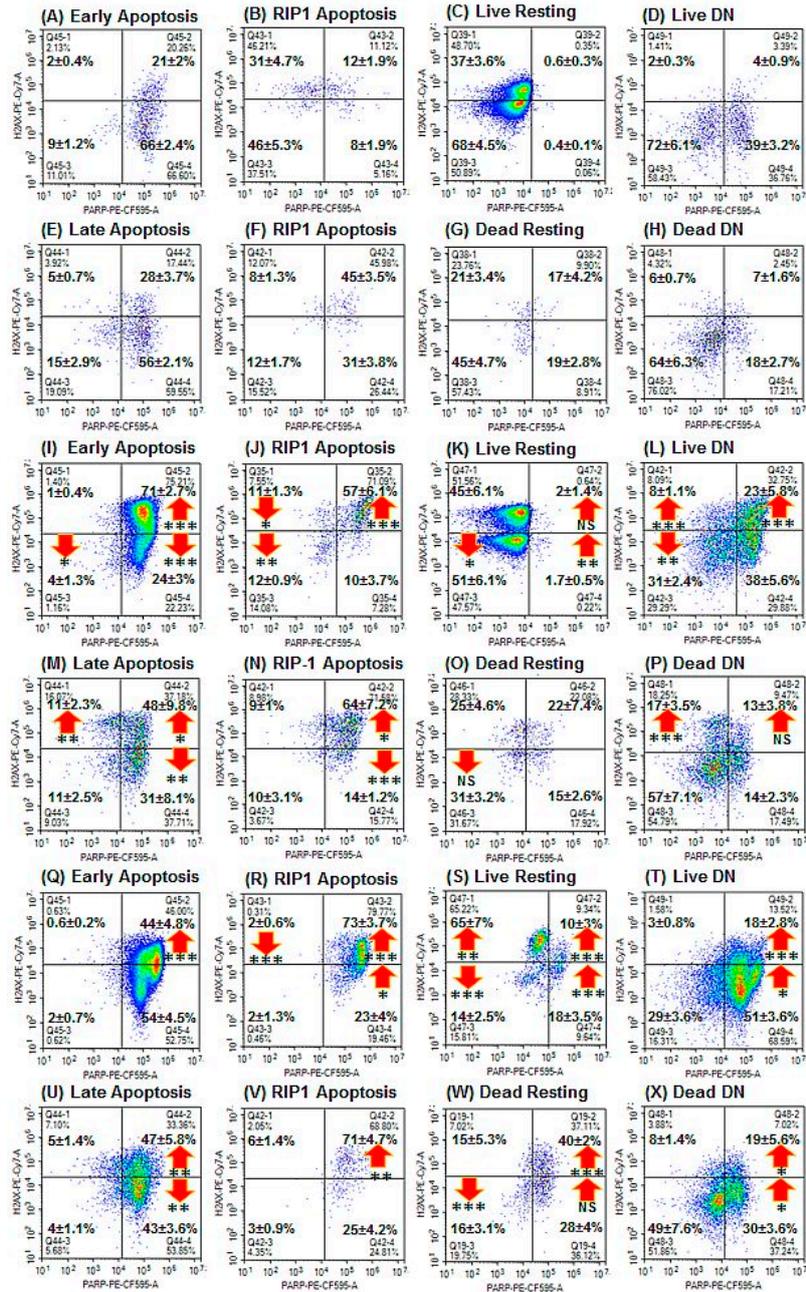


Figure 2. pH2AX hyper-activation of cleaved PARP/parthanatos, apoptosis via cleaved PARP and DNA Damage assay. Jurkat cells were incubated with live-fix dye, Zombie NIR, fixed and permeabilised and incubated with anti-active caspase-3-BV650, RIP3-PE, cleaved PARP-PE-CF595, pH2AX-PE-Cy7 according to the Materials and Methods and 100,000 cells analysed flow cytometrically. After gating on live and dead cells from a Zombie NIR vs Caspase-3-BV650 dot-plot untreated (A-H), 0.25 % NaN_3 (I-P) or 1 μM Etop (Q-X) treated live and dead Jurkat cells were analysed on a RIP3-PE vs Caspase-3-BV650 dot-plot. The early and late apoptosis phenotypes indicated by RIP3^{ve}/Caspase-3^{ve} (A, E, I, M, Q, U), RIP1-dependent apoptosis phenotype indicated by RIP3^{ve}/Caspase-3^{ve} (B, F, J, N, R, V), resting phenotype indicated by RIP3^{ve}/Caspase-3^{ve} (C, G, K, O, S, W), or double negative indicated by RIP3^{ve}/Caspase-3^{ve} (D, H, L, P, T, X), respectively. Such live and dead populations were then analysed for cleaved PARP and pH2AX. n=3, % mean \pm % SEM, student t test NS (not significant), $P < 0.05^*$, $P < 0.01^{**}$, $P < 0.001^{***}$, with arrows indicating change compared to untreated cells.

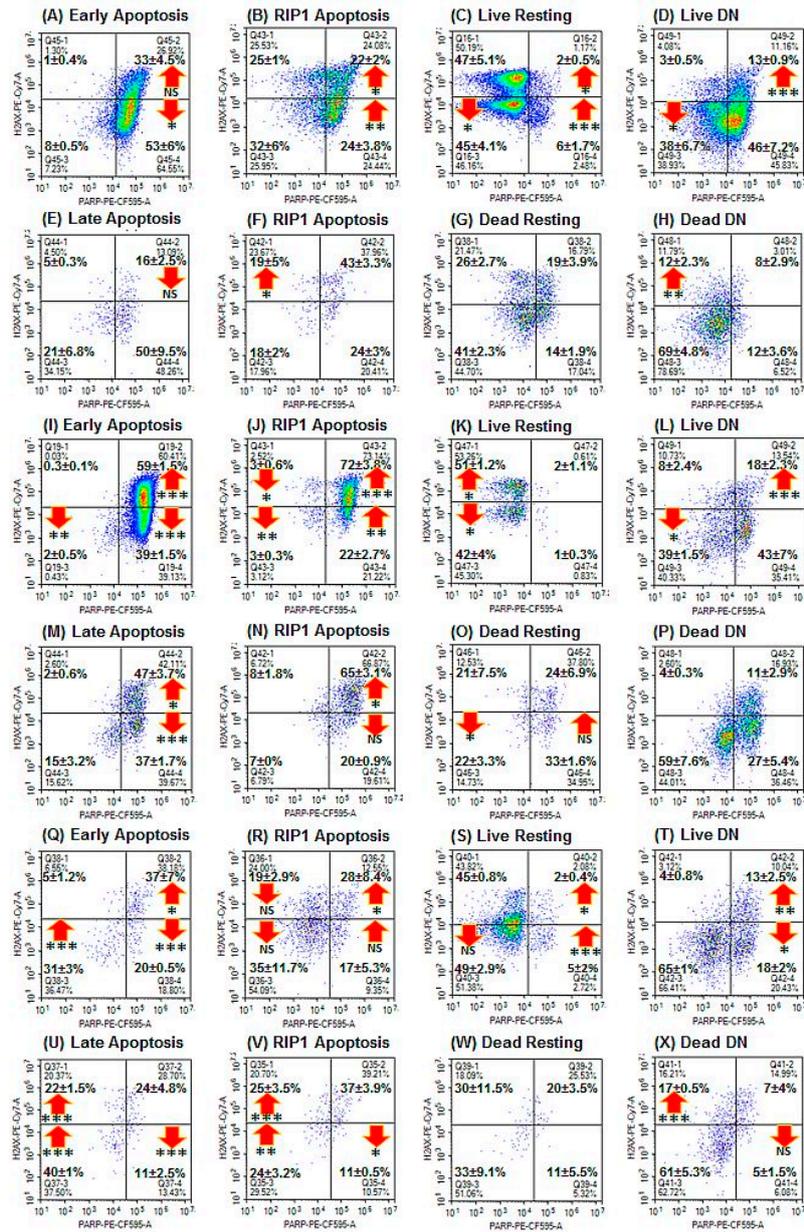


Figure 3. pH2AX hyper-activation of cleaved PARP/parthanatos, apoptosis via cleaved PARP and DNA Damage of blocked oncosis. Jurkat cells were pre-treated zVAD (20 μ M) and or Nec-1 (60 μ M) for 2 h then incubated with 0.25 % NaN₃ for 24 h. Cells were then loaded with live-fix dye, Zombie NIR, fixed and permeabilised and incubated with anti-active caspase-3-BV650, RIP3-PE, PARP-PE-CF595, and pH2AX-PE-Cy7, according to the Materials and Methods and 100,00 cells analysed flow cytometrically. After gating on live and dead cells from a Zombie NIR v Caspase-3-BV650 dot-plot treated live and dead Jurkat cells with zVAD with 0.25 % NaN₃ (A-H), Nec-1 with 0.25 % NaN₃ (I-P) or zVAD with Nec-1 and 0.25 % NaN₃ (Q-X) were analysed on a RIP3-PE vs. Caspase-3-BV650 dot-plot. The early and late apoptosis phenotype indicated by RIP3^{ve}/Caspase-3^{ve} (A, E, I, M, Q, U), RIP1-dependent apoptosis phenotype indicated by RIP3^{ve}/Caspase-3^{ve} (B, F, J, N, R, V), resting phenotype indicated by RIP3^{ve}/Caspase-3^{ve} (C, G, K, O, S, W), or double negative indicated by RIP3^{ve}/Caspase-3^{ve} (D, H, L, P, T, X), respectively and such live and dead populations were then analysed for PARP and pH2AX. n=3, % mean \pm % SEM, student t test NS (not significant), $P < 0.05^*$, $P < 0.01^{**}$, $P < 0.001^{***}$, with arrows indicating change compared to untreated cells.

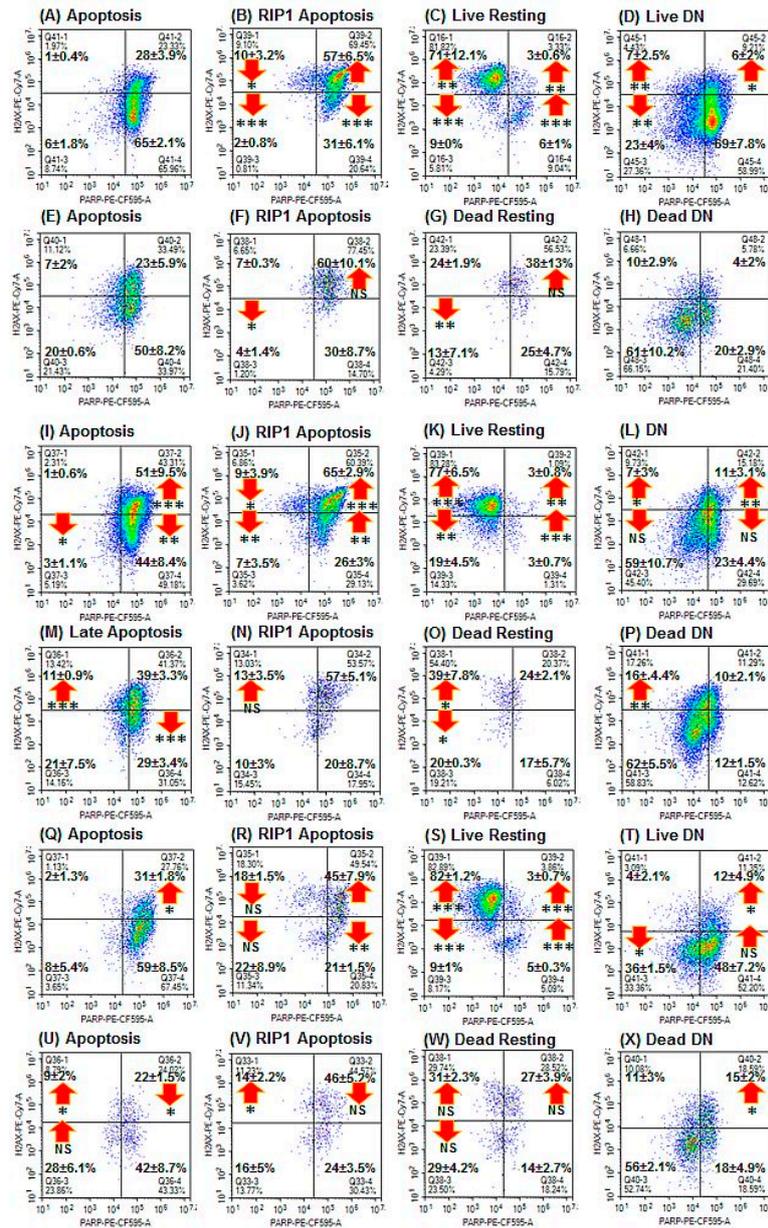


Figure 4. pH2AX Hyper-activation of cleaved PARP/parthanatos, apoptosis via cleaved PARP and DNA Damage blocked apoptosis. Jurkat cells were pre-treated zVAD (20 μ M) and or Nec-1 (60 μ M) for 2 h then incubated with 1 μ M Etop for 24 h. Cells were then loaded with live-fix dye, Zombie NIR, fixed and permeabilised and incubated with anti-active caspase-3-BV650, RIP3-PE, PARP-PE-CF595, and pH2AX-PE-Cy7, according to the Materials and Methods and 100,000 cells analysed flow cytometrically. After gating on live and dead cells from a Zombie NIR v Caspase-3-BV650 dot-plot treated live and dead Jurkat cells with zVAD with 0.5 μ M Etop (A-H), Nec-1 with 1 μ M Etop (I-P) or zVAD with Nec-1 and 1 μ M Etop (Q-X) were analysed on a RIP3-PE vs Caspase-3-BV650 dot-plot. The early and late apoptosis phenotype indicated by RIP3^{ve}/Caspase-3^{ve} (A, E, I, M, Q, U), RIP1-dependent apoptosis phenotype indicated by RIP1^{ve}/Caspase-3^{ve} (B, F, J, N, R, V), resting phenotype indicated by RIP3^{ve}/Caspase-3^{ve} (C, G, K, O, S, W), or double negative indicated by RIP3^{ve}/Caspase-3^{ve} (D, H, L, P, T, X), respectively and such live and dead populations were then analysed for cleaved PARP and pH2AX. n=3, % mean \pm % SEM, student t test NS (not significant), $P < 0.05^*$, $P < 0.01^{**}$, $P < 0.001^{***}$, with arrows indicating change compared to untreated cells.