

Supplementary Information

Development of CRISPR Interference (CRISPRi) Platform for Metabolic Engineering of *Leuconostoc citreum* and Its Application for Engineering Riboflavin Biosynthesis

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Table 1. Bacterial strains and plasmids used in this study.

Plasmid	Description	Reference
Strains		
XL 1-Blue	<i>recA1 endA1 gyrA96 thi-1 hsdR17 supE44 relA1 lac</i> [<i>F' proAB lacIqZDM15 Tn10 (Tetr)</i>]	Stratagene ^a
<i>L. citreum</i> CB2567	Wild type	[38]
JW001	<i>L. citreum</i> CB2567, Δ <i>ldhD</i> ::P ₇₁₀ -sfGFP-Cm ^R	This study
Plasmids		
pCB4270	<i>L. citreum</i> - <i>E. coli</i> shuttle vector, P ₇₁₀ , Amp ^R , Cm ^R	[19]
pCB4270-Em	<i>L. citreum</i> - <i>E. coli</i> shuttle vector, P ₇₁₀ , Amp ^R , Em ^R	This study
pCB4270-sfGFP	Monocistronic, P ₇₁₀ , sfGFP, Amp ^R , Cm ^R	[19]
pCB4270V4	<i>L. citreum</i> - <i>E. coli</i> shuttle vector, P _{710V4} , Amp ^R , Cm ^R	This study
pCB4270V4-Em	<i>L. citreum</i> - <i>E. coli</i> shuttle vector, P _{710V4} , Amp ^R , Em ^R	This study
pdCas9	Plasmid carrying SpdCas9 catalytic site mutant	Addgene, USA
pgRNA-bacteria	Plasmid carrying customized tracrRNA for bacterial gene knock-down	Addgene, USA
pSOS95	Acetone operon, <i>repL</i> gene, ColE1 origin, Amp ^R , MLS ^R , Em ^R	[39]
pHldhU-sfGFP	pCB4270-sfGFP derivative containing 0.8 kb homologous region <i>Bam</i> HI upstream fragment of <i>ldhD</i>	This study
pH-InsfGFP	pCB4270ldhU-sfGFP derivative suicide vector containing 0.8 kb homologous region <i>Bam</i> HI upstream fragment of <i>ldhD</i> and 1 kb homologous region <i>Pst</i> I internal fragment of <i>ldhD</i> , Amp ^R , Cm ^R	This study
pGFP-sgR-Em	pCB4270-Em derivative carrying P _{710V4} , sfGFP targeting sgRNA, Amp ^R , Em ^R	This study
pD1-Em	pCB4270V4-Em derivative carrying bicistronic, P _{710V4} , eSD2, SpdCas9, His tag, Amp ^R , Em ^R	This study
pD2-Em	pCB4270V4-Em derivative carrying bicistronic, P _{710V4} , SD2, SpdCas9, His tag, Amp ^R , Em ^R	This study
pD3-Em	pCB4270-Em derivative carrying bicistronic, P ₇₁₀ , eSD2, SpdCas9, His tag, Amp ^R , Em ^R	This study
pD4-Em	pCB4270-Em derivative carrying bicistronic, P ₇₁₀ , SD2, SpdCas9, His tag, Amp ^R , Em ^R	This study
pD1	pCB4270V4 derivative carrying bicistronic, P _{710V4} , eSD2, SpdCas9, His tag, Amp ^R , Cm ^R	This study
pD2	pCB4270V4 derivative carrying bicistronic, P _{710V4} , SD2, SpdCas9, His tag, Amp ^R , Cm ^R	This study
pD3	pCB4270 derivative carrying bicistronic, P ₇₁₀ , eSD2, SpdCas9, His tag, Amp ^R , Cm ^R	This study
pD4	pCB4270 derivative carrying bicistronic, P ₇₁₀ , SD2, SpdCas9, His tag, Amp ^R , Cm ^R	This study
pGFP-sgR-D1-Em	pD1-Em derivative carrying bicistronic, P _{710V4} , eSD2, SpdCas9, His tag, P _{710V4} , sfGFP targeting sgRNA, Amp ^R , Em ^R	This study
pGFP-sgR-D2-Em	pD2-Em derivative carrying bicistronic, P _{710V4} , SD2, SpdCas9, His tag, P _{710V4} , sfGFP targeting sgRNA, Amp ^R , Em ^R	This study

pGFP-sgR-D3-Em	pD3-Em derivative carrying bicistronic, P ₇₁₀ , eSD2, SpdCas9, His tag, P _{710V4} , sfGFP targeting sgRNA, Amp ^R , Em ^R	This study
pGFP-sgR-D4-Em	pD4-Em derivative carrying bicistronic, P ₇₁₀ , SD2, SpdCas9, His tag, P _{710V4} , sfGFP targeting sgRNA, Amp ^R , Em ^R	This study
pFolE-sgR-D4	pD4 derivative carrying bicistronic, P ₇₁₀ , SD2, SpdCas9, His tag, P _{710V4} , <i>folE</i> targeting sgRNA, Amp ^R , Cm ^R	This study
pRibF-sgR-D4	pD4 derivative carrying bicistronic, P ₇₁₀ , SD2, SpdCas9, His tag, P _{710V4} , <i>ribF</i> targeting sgRNA, Amp ^R , Cm ^R	This study
pFRdual-D4	pFolE-sgR-D4 derivative carrying bicistronic, P ₇₁₀ , SD2, SpdCas9, His tag, P _{710V4} , <i>folE</i> targeting sgRNA, P _{710V4} , <i>ribF</i> targeting sgRNA, Amp ^R , Cm ^R	This study
pH-rib	pCB4270 derivative carrying monocistronic, P _{710V4} , <i>rib</i> operon, Amp ^R , Cm ^R	This study
pFRdual-Rib-D4	pFRdual-D4 derivative carrying bicistronic, P ₇₁₀ , SD2, SpdCas9, His tag, P _{710V4} , <i>folE</i> targeting sgRNA, P _{710V4} , <i>ribF</i> targeting sgRNA, monocistronic, P _{710V4} , <i>rib</i> operon, Amp ^R , Cm ^R	This study

Table S2: Oligonucleotides primers used in this study

Primer	Sequence (5' to 3')
F-LDHU	GGATCCAATGTTATGTTGCGAATATATTTTAAATAA
R-LDHU	GGATCCAAGATCCTCCAAAATTTATATTACCA
F-LDHD	CTGCAGGACTTATTGACTAAGCTCGGG
R-LDHD	CTGCAGTTTCATGGCACCAAGTGC
IF-stCAT	GGACTTCATTTACTGGGTTT
OR-ldh	ATATTGTGCAATGAAACCAGC
F1-v4sfgsgR	TGAGTATAACCAGAACATCTAATTCAACAAGAATTGTTTTAGAGCTAGAAATAG CAAGTTAAAATAAGG
F2-v4sfgsgR	AAGTTAATAGGTGTTTTAGCCTGAAGTGTATAATGAGTATAACCAGAACATCT AATTCAACAA
F3-v4sfgsgR	CCGCGCCGAGAACAGTTTAAAGTTAATAGGTGTTTTAGCCTGAAGT
R-v4sfgsgR	GGCGCCAGTTCACCGACAAACAACAGAT
F1-v4folEsgR	CAGAAATATTTGACGCCCTGCACGAGTTTTAGAGCTAGAAATAGCAAGTTAAA ATAAG
F2-v4folEsgR	AGTTAATAGGTGTTTTAGCCTGAAGTGTATAATGAGTATAACCAGAAATATTT GACGCCCTGC
F3-v4folEsgR	AATCTAGAGAGAACAGTTTAAAGTTAATAGGTGTTTTAGCCTGAAGT
F1-v4ribsgR	AATGAGTATAACCAGAAGGCTGTTTGAAATTTTGAATGTTTTAGAGCTAGAAAT AGCAAGTTAAAATAAG
F2-v4ribsgR	GAGAACAGTTTAAAGTTAATAGGTGTTTTAGCCTGAAGTGTATAATGAGTATA ACCAGAAGGCTGTTTG
F1-eSD2dCas9	ATCTTAATCATGGAAGGGAGGGTTTTTAATGGATAAGAAATACTCAATAGGCTT AGCTA
F2-eSD2dCas9	GGCTCGAGATGAAAGCAATTTTCGTA CTGAAACATCTTAATCATGGAAGGGAG GGT
F1-SD2dCas9	ACATCTTAATCATGCAAAGGAGGTGTTTAAATGGATAAGAAATACTCAATAGGC TTAGC
F2-SD2dCas9	AGCTCGAGATGAAAGCAATTTTCGTA CTGAAACATCTTAATCATGCAAAGGAG GTG
R-Not-dCas9	GCGCGGCCGCTTATTAGTGG
F-RT-sfGFP	TGCTACAAACGGAAAACCTCA
R-RT-sfGFP	GCGTTCCTGTACATAACCTT
F-RT-foIE	ACCTCGCTTGGTTGATTGGG
R-RT-foIE	CGTGTGTTGTTTTCGCTCCCTG
F-RT-ribF	TCCACATCCAAGTGTCGCAT
R-RT-ribF	CCTGTGGTGATAAACCCGCT
F-RT-16srRNA	TATGTCCCGAGCGTTATCCG
R-RT-16srRNA	TCTACGCATTCCACCGCTAC

Table S3. sgRNA used in this study

sgRNA name	Target location	Spacer sequence (5' to 3')	PAM sequence (5' to 3')
GFP-sgRNA	CDS of sfGFP	AATTCTTGTTGAATTAGATG	CCC
folE-sgRNA	CDS of <i>folE</i>	ATATTTGACGCCCTGCACGA	CCC
ribF-sgRNA	CDS of <i>ribF</i>	GGCTGTTTGAAATTTGAAT	CCA

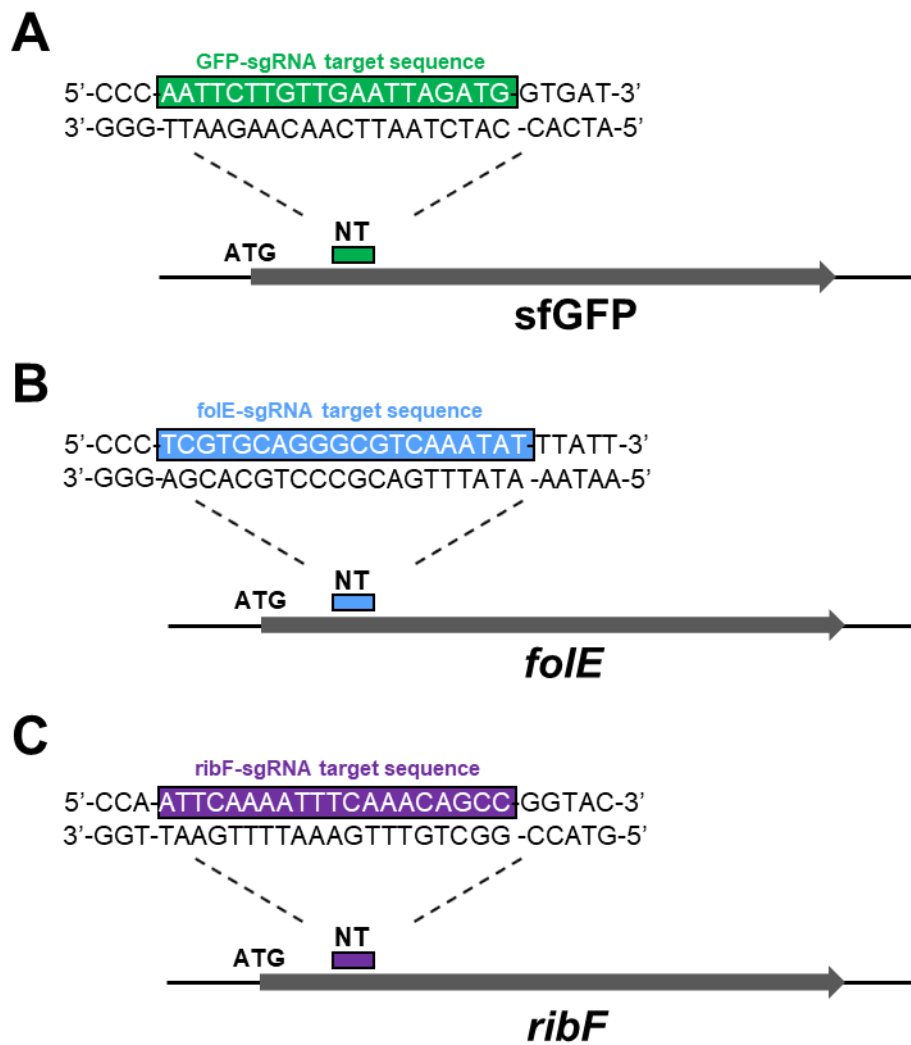


Figure S1. Schematic of synthetic guide RNAs target sequence used in this study. NT represents non-template strand of target gene. **(A)** Target sequence of GFP-sgRNA for sfGFP gene. **(B)** Target sequence of folE-sgRNA for *folE* gene. **(C)** Target sequence of ribF-sgRNA for *ribF* gene.