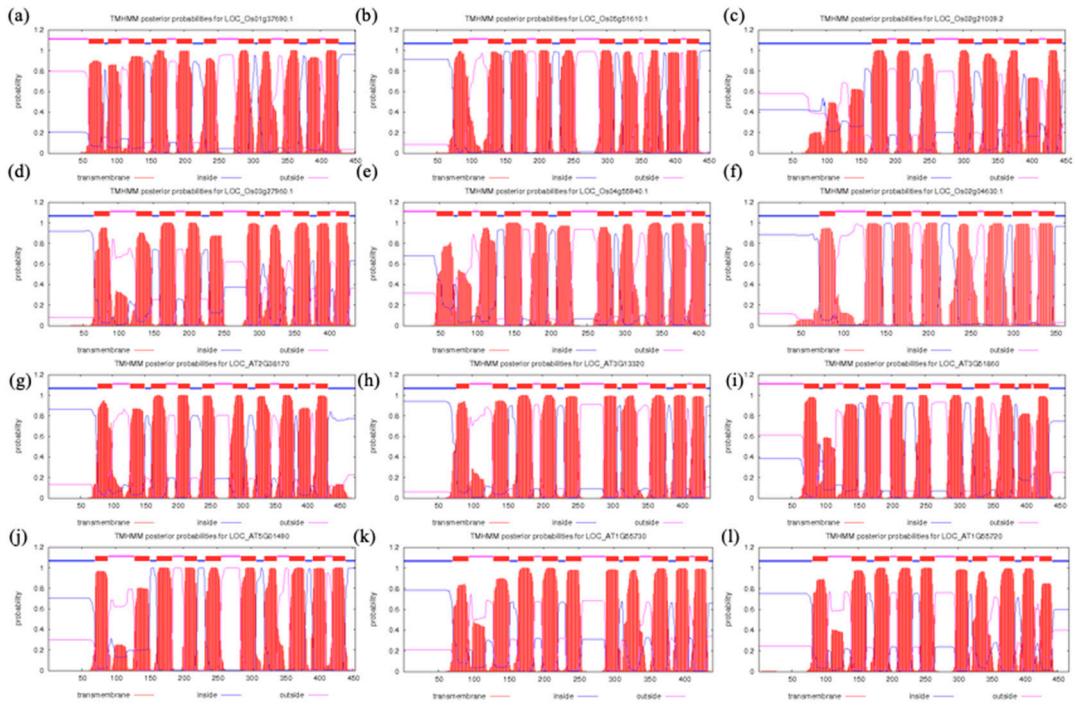
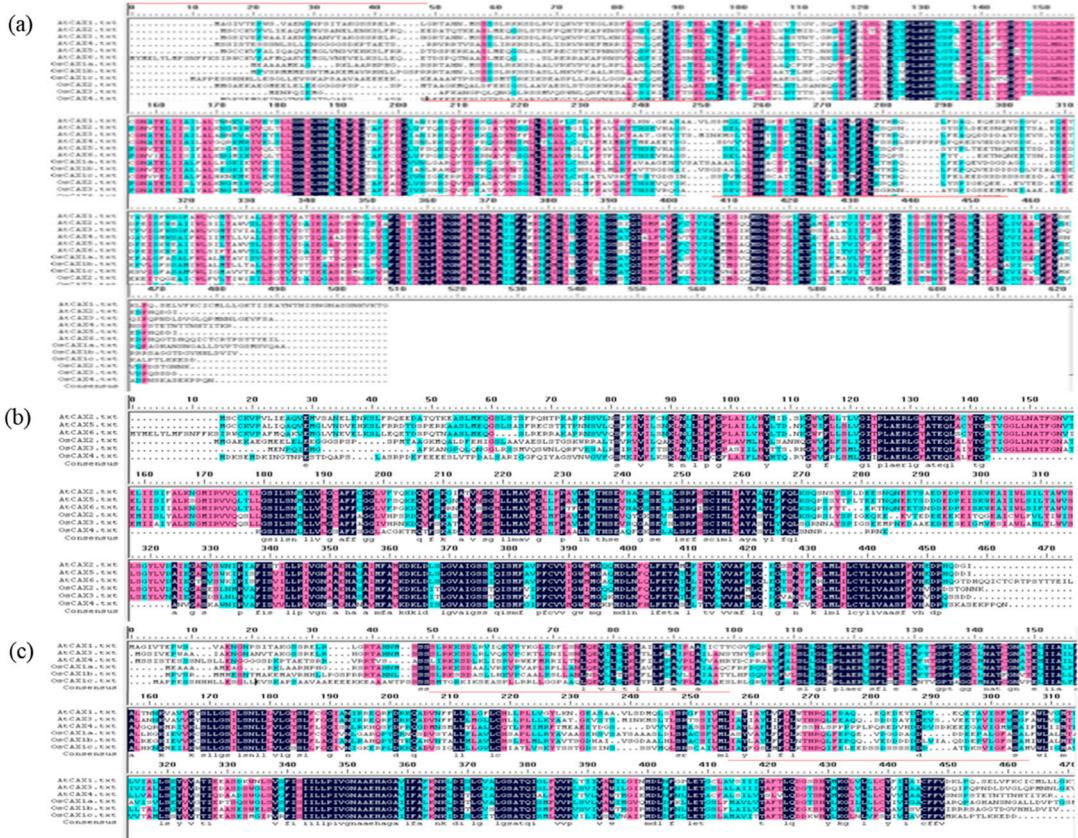


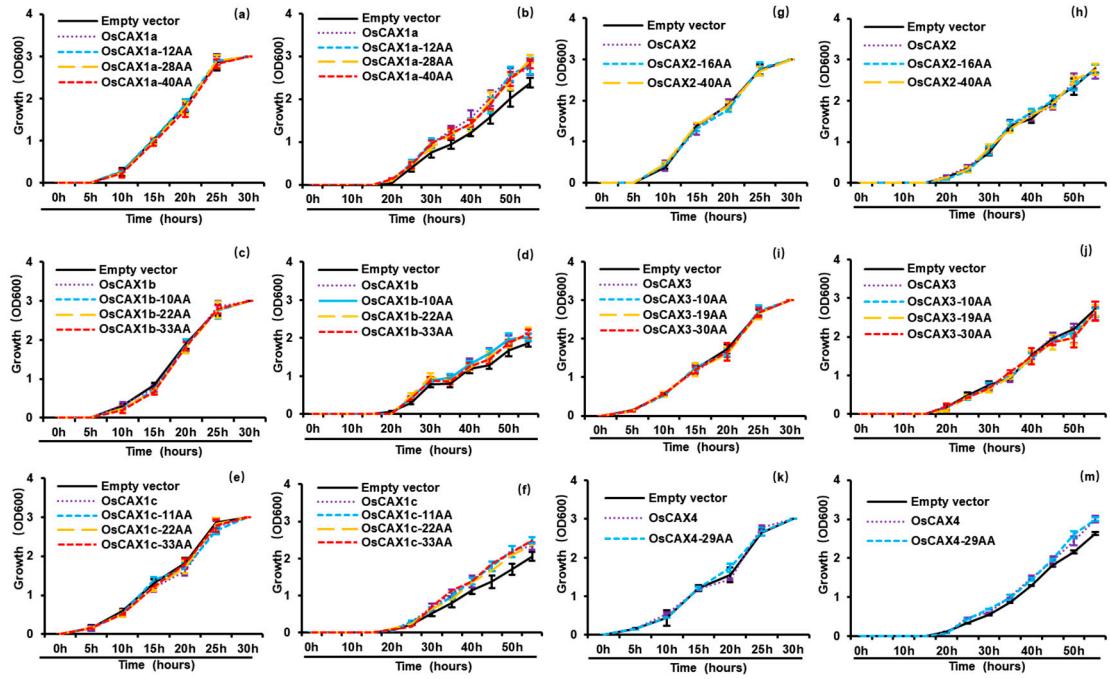
## Supplementary Material



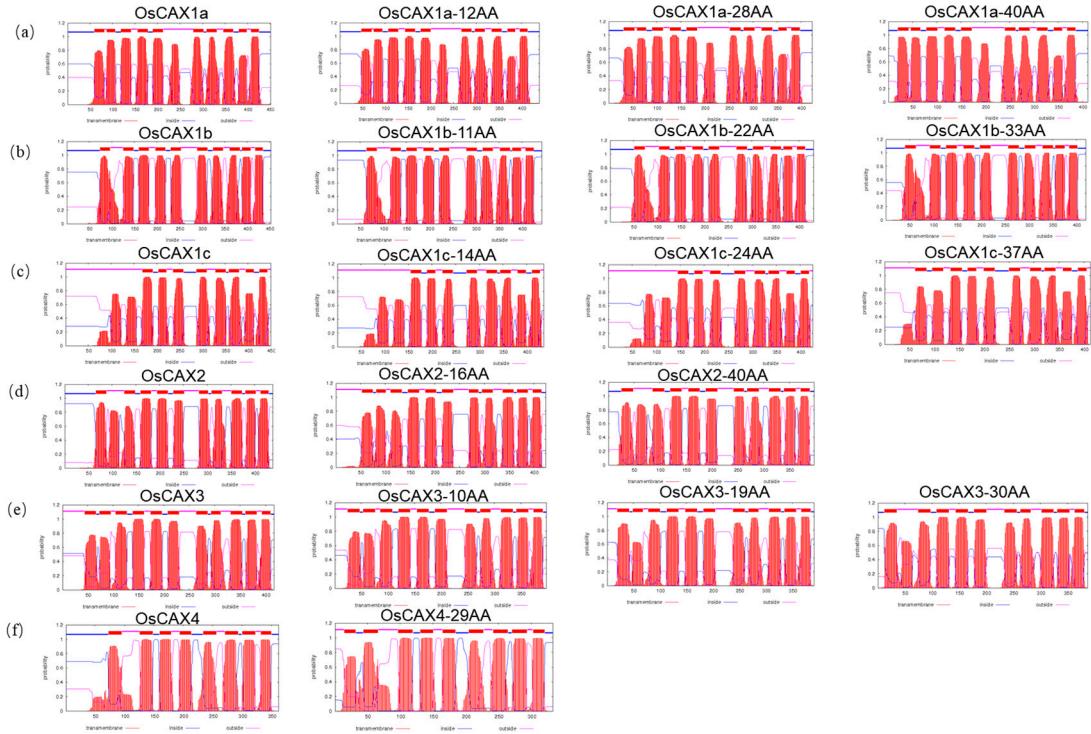
**Figure 1. Membrane topology of selected CAXs.** The topologic models were generated by the TMHMM program ([www.cbs.dtu.dk/services/TMHMM/](http://www.cbs.dtu.dk/services/TMHMM/)). TMHMM posterior probabilities for (a) OsCAX1a; (b) OsCAX1b; (c) OsCAX1c; (d) OsCAX2; (e) OsCAX3; (f) OsCAX4; (g) AtCAX1; (h) AtCAX2; (i) AtCAX3; (j) AtCAX4; (k) AtCAX5; (l) AtCAX6. The red areas indicate the predicted TM domains, and the blue and magenta lines indicate the predicted regions inside or outside of the membrane, respectively.



**Figure S2. Multiple alignment of *Oryza sativa* and *Arabidopsis thaliana* CAAX family proteins.** (a) Amino acid sequence alignments; (b) the subfamily IB; (c) the subfamily IA. Multiple alignment was performed with DNAMAN. Black, red and blue lines underneath alignments indicate reported consensus transport residues more frequently reported consensus transport residues, and less frequently reported consensus transport residues, respectively. “|” above the sequence mean every ten amino acid residues.



**Figure S3. Functional assay of rice CAX genes by heterologous expression in yeast.** Yeast strains were grown in liquid media with 0  $\mu\text{mol/L}$   $\text{CdCl}_2$  for 30 h (a), (c), (e), (g), (i) and (k); or 10  $\mu\text{mol/L}$   $\text{CdCl}_2$  for 55 h (b), (d), (f), (h), (j) and (m). The absorbance at 600 nm (OD600) of cell cultures was measured every 5 h. Values are mean  $\pm$  SE ( $n = 3$ ).



**Figure S4. Transmembrane structure of the truncated rice CAX proteins.** The topologic models were generated by the TMHMM program. TMHMM posterior probabilities for (a) OsCAX1a, OsCAX1a-12A, OsCAX1a-28AA, OsCAX1a-40AA; (b) OsCAX1b, OsCAX1b-11AA, OsCAX1b-22AA, OsCAX1b-33AA; (c) OsCAX1c-14AA, OsCAX1c-24AA, OsCAX1c-37AA; (d) OsCAX2, OsCAX2-16AA, OsCAX2-40AA; (e) OsCAX3, OsCAX3-10AA, OsCAX3-19AA, OsCAX3-40AA; (f) OsCAX4, OsCAX4-29AA. The red, blue and magenta areas indicate the predicted TM domains, the regions inside or outside of the membrane, respectively.

**Table S1. Primers for qRT-PCR analysis of rice CAX family genes.**

<b>Genes</b>	<b>Forward primer (5'-3')</b>	<b>Reverse primer (5'-3')</b>
<i>Osactin</i>	GGAAGTGGTATGGTCAAGGC	AGTCTCATGGATAACCGCAG
<i>OsCAX1a</i>	TGTGGGTGTTGCTCTTAGT	GTGCAATCTGCTCTGTGAGG
<i>OsCAX1b</i>	ATCTCAACGACGTCCTCCTC	GCTAAGCACAAACACCCACA
<i>OsCAX1c</i>	TGAGCTTCCTGAGCGAACAT	TTATGCAGAGCGAACAGTGC
<i>OsCAX2</i>	CGTCATGCTCCACTACCTCT	TCCCAATCTCTCAGCCAAGG
<i>OsCAX3</i>	CTGGCGGTATTGTTCATCGG	AGTCCCAGACAGCCATCAA
<i>OsCAX4</i>	TTCCCTTGGCTGAGCGATTG	ACTCGGTTGCATTGCCAAAT

**Table S2. Primers used to amplify the open reading frame of the candidate genes.**

Genes	Primer (5'-3')
OsCAX1a-F	ATGGAGGCAGG CGGCAGCGAT
OsCAX1a-14AA-F	ATGGCGCGCGGCACCCGCACGG
OsCAX1a-28AA-F	ATGTCGTCGTCGCTGCAGGAAGAA
OsCAX1a-40AA-F	ATGGTGCAGAAGGTCCCCGTGGC
OsCAX1a-R	TCATGCAGCTGAACACTCAT
OsCAX1b-F	ATGCCAGTGT CGCGGATGAT GAT
OsCAX1b-10AA-F	ATGACGATGGCCAAGGAGATGGC
OsCAX1b-22A-F	ATGCTTCCGGGTCTCCTCGCCG
OsCAX1b-33A-F	ATGAACCTCTCATCTTCCTCCCT
OsCAX1b-R	CTACACAATC ACATCCAGATGG
OsCAX1c-F	ATGGCGCCGCCGGAGAGCA
OsCAX1c-14AA-F	ATGCTGCTGAAGTCAGCAAGGC
OsCAX1c-24AA-F	ATGGCGGTGGCTGCCGAGGAAGA
OsCAX1c-37AA-F	ATGGCGTGGACACCGTCGTCGTC
OsCAX1c-R	TTAGTCATCTTCCTTTAGCGTTGGG
OsOsCAX2-F	ATGATGGGGCGGGAGAAGG
OsCAX2-14AA-F	ATGGAGGAGGGCGGGGGCT
OsCAX2-40AA-F	ATGGGGTCGCTGCCGCGGT
OsCAX2-R	TCACTTGTGTTACCTGT
OsCAX3-F	ATGGAGAACCTCAGATTG
OsCAX3-10AA-F	ATGGGTGGGCTACGTTCCAGCAT
OsCAX3-19AA-F	ATGGTTCAGTCCTGGAACCTGCA
OsCAX3-30AA-F	ATGAACCTGCAGAGATTGTTGA
OsCAX3-R	TCAATCATC CTGGATTGTG G
OsCAX4-F	ATGAGCCAACAGCTTCAA
OsCAX4-29AA-F	ATGCAGGCCCTTCATTGGCTAG
OsCAX4-R	TTAGTTTGTGGAGGCTCT

**Table S3. The CAX family genes involve in metal stress tolerance.**

Genes	Species	Subcellular localization	Functions	References
<i>OsCAX1a</i>	Yeast	Vacuolar	Ca transport and Ca homeostasis	[1, 2]
<i>OsCAX1b</i>	Yeast	-	Ca tolerance	[1]
<i>OsCAX1c</i>	Yeast	-	Ca tolerance	[1]
<i>OsCAX3</i>	Yeast	Plasma membrane	Ca tolerance	[1]
<i>OsCAX4</i>	Yeast	-	Ca, Cu and Mn transport	[3]
	Yeast	-	Ca, Mn and Ba tolerance; Ca, Zn and Cd transport	[4-6]
	Arabidopsis	Vacuolar	Ca, Mn, Mg, salt and freezing tolerance; Ca transport	[7-9]
<i>AtCAX1</i>	Arabidopsis halleri	-	Cd tolerance	[10, 11]
	Tobacco	-	Ca homeostasis	[12]
	Petunia	-	Cd tolerance and accumulation	[13]
	Tomato	-	Ca accumulation	[14]
	Yeast	Vacuolar	Ca tolerance and translocation	[15-17]
<i>AtCAX2</i>	Tobacco	-	Cd, Mn, Zn tolerance; Cd and Mn transport	[18-21]
	Potato	-	Ca transport	[22]
	Tomato	-	Ca accumulation	[23]
<i>AtCAX3</i>	Yeast	Vacuolar	Ca tolerance and translocation	[24]
	Arabidopsis	-	salt tolerance	[25]
	yeast	Vacuolar	Ca, Ba and salt tolerance	[26]
	Arabidopsis	-	Cd, Ca and salt tolerance	[9, 27]
<i>AtCAX4</i>	Tobacco	-	Cd, Mn and Zn tolerance; Cd transport and accumulation	[19-21]
	tomato	-	Ca accumulation	[28]
<i>AtCAX5</i>	Yeast	Vacuolar	Ca and Mn transport; ion homeostasis	[16, 29]
<i>PutCAX1</i>	Yeast	Vacuolar	Ca and Ba tolerance	[6]
<i>PutCAX2</i>	Yeast	Golgi apparatus	Ca and Ba tolerance	[26]
<i>TuCAX1a</i>	Arabidopsis	-	Ca and Zn tolerance and translocation	[30]
<i>TuCAX1b</i>	Arabidopsis	-	Ca and Zn tolerance; Cd, Ca and Zn translocation	[30]
<i>LeCAX2</i>	Yeast	-	Ca and Mn transport	[16]
<i>GhCAX3</i>	Yeast	-	Ca Transport and stress tolerance	[31]
<i>GhCAX3</i>	Arabidopsis	-	cross-talk of ABA and cold signal transduction	[31]
<i>SeCAX3</i>	Yeast	-	Ca transport	[32]
<i>VvCAX3</i>	Yeast	-	Na, Li and Cu tolerance; Ca transport	[33]

## References

1. Kamiya, T.; Akahori, T.; Maeshima, M. Expression profile of the genes for rice cation/H<sup>+</sup> exchanger family and functional analysis in yeast. *Plant Cell Physiol.* **2005**, *46*, 1735–1740.
2. Kamiya, T.; Akahori, T.; Ashikari, M.; Maeshima, M. Expression of the vacuolar Ca<sup>2+</sup>/H<sup>+</sup> exchanger, OsCAX1a, in rice: cell and age specificity of expression, and enhancement by Ca<sup>2+</sup>. *Plant Cell Physiol.* **2006**, *47*, 96–106.
3. Yamada, N.; Theerawitaya, C.; Cha-um, S.; Kirdmanee, C.; Takabe, T. Expression and functional analysis of putative vacuolar Ca<sup>2+</sup>-transporters (CAXs and ACAs) in roots of salt tolerant and sensitive rice cultivars. *Protoplasma* **2014**, *251*, 1067–1075.
4. Shigaki, T.; Barkla, B.J.; Miranda-Vergara, M.C.; Zhao, J.; Pantoja, O.; Hirschi, K.D. Identification of a crucial histidine involved in metal transport activity in the Arabidopsis cation/H<sup>+</sup> exchanger CAX1. *J. Biol. Chem.* **2005**, *280*, 30136–30142.
5. Shigaki, T.; Mei, H.; Marshall, J.; Li, X.; Manohar, M.; Hirschi, K.D. The expression of the open reading frame of Arabidopsis CAX1, but not its cDNA, confers metal tolerance in yeast. *Plant Biol. (Stuttg.)* **2010**, *12*, 935–939.
6. Guo, H.; Tsugama, D.; Takano, T.; Bu, Y.; Liu, S. AtCAX1 N-terminal can improve Ca<sup>2+</sup> and Ba<sup>2+</sup> tolerance in yeast. *Cell Biol. Biophys.* **2015**, *4*, 1–4.
7. Catala, R.; Santos, E.; Alonso, J.M.; Ecker, J.R.; Martinez-Zapater, J.M.; Salinas, J. Mutations in the Ca<sup>2+</sup>/H<sup>+</sup> transporter CAX1 increase CBF/DREB1 expression and the cold-acclimation response in Arabidopsis. *Plant Cell.* **2003**, *15*, 2940–2951.
8. Cheng, N.H.; Pittman, J.K.; Barkla, B.J.; Shigaki, T.; Hirschi, K.D. The Arabidopsis *cax1* mutant exhibits impaired ion homeostasis, development, and hormonal responses and reveals interplay among vacuolar transporters. *Plant Cell.* **2003**, *15*, 347–364.
9. Bu, Y.; Fu, W.; Chen, J.; Takano, T.; Liu, S. Description of AtCAX4 in Response to Abiotic Stress in Arabidopsis. *Int. J. Mol. Sci.* **2021**, *22*, 856.
10. Baliardini, C.; Meyer, C.L.; Salis, P.; Saumitou-Laprade, P.; Verbruggen, N. CATION EXCHANGER1 Cosegregates with Cadmium Tolerance in the Metal Hyperaccumulator Arabidopsis halleri and Plays a Role in Limiting Oxidative Stress in Arabidopsis Spp. *Plant Physiol.* **2015**, *169*, 549–559.
11. Ahmadi, H.; Corso, M.; Weber, M.; Verbruggen, N.; Clemens, S. CAX1 suppresses Cd-induced generation of reactive oxygen species in *Arabidopsis halleri*. *Plant Cell Environ.* **2018**, *41*, 2435–2448.
12. Hirschi, K. Expression of Arabidopsis CAX1 in tobacco: altered calcium homeostasis and increased stress sensitivity. *Plant Cell.* **1999**, *11*, 2113–2122.
13. Wu, Q.; Shigaki, T.; Williams, K.A.; Han, J.S.; Kim, C.K.; Hirschi, K.D.; Park, S. Expression of an Arabidopsis Ca<sup>2+</sup>/H<sup>+</sup> antiporter CAX1 variant in petunia enhances cadmium tolerance and accumulation. *J. Plant Physiol.* **2011**, *168*, 167–173.
14. Zorrilla, C.; Schabow, J.E.; Chernov, V.; Plata, J.P. CAX1 Vacuolar Antiporter Overexpression in Potato Results in Calcium Deficiency in Leaves and Tubers by Sequestering Calcium as Calcium Oxalate. *Crop Sci.* **2018**, *59*, 1.
15. Hirschi, K.D.; Zhen, R.G.; Cunningham, K.W.; Rea, P.A.; Fink, G.R. CAX1, an H<sup>+</sup>/Ca<sup>2+</sup> antiporter from Arabidopsis. *Proc. Natl. Acad. Sci. USA.* **1996**, *93*, 8782–8786.
16. Edmond, C.; Shigaki, T.; Ewert, S.; Nelson, M.D.; Connerton, J.M.; Chalova, V.; Noordally, Z.; Pittman, J.K. Comparative analysis of CAX2-like cation transporters indicates functional and regulatory diversity. *Biochem. J.* **2009**, *418*, 145–154.
17. Pittman, J.K.; Shigaki, T.; Hirschi, K.D. Evidence of differential pH regulation of the Arabidopsis vacuolar Ca<sup>2+</sup>/H<sup>+</sup> antiporters CAX1 and CAX2. *FEBS Lett.* **2005**, *579*, 2648–2656.
18. Hirschi, K.D.; Korenkov, V.D.; Wilganowski, N.L.; Wagner, G.J. Expression of arabidopsis CAX2 in tobacco. Altered metal accumulation and increased manganese tolerance. *Plant Physiol.* **2000**, *124*, 125–133.
19. Koren'kov, V.; Park, S.; Cheng, N.H.; Sreevidya, C.; Lachmansingh, J.; Morris, J.; Hirschi, K.; Wagner, G.J. Enhanced Cd<sup>2+</sup>-selective root-tonoplast-transport in tobacco expressing Arabidopsis cation exchangers. *Planta* **2007**, *225*, 403–411.
20. Korenkov, V.; Hirschi, K.; Crutchfield, J.D.; Wagner, G.J. Enhancing tonoplast Cd/H antiport activity increases Cd, Zn, and Mn tolerance, and impacts root/shoot Cd partitioning in *Nicotiana tabacum* L. *Planta* **2007**, *226*, 1379–1387.
21. Korenkov, V.; King, B.; Hirschi, K.; Wagner, G.J. Root-selective expression of AtCAX4 and AtCAX2 results in reduced lamina cadmium in field-grown *Nicotiana tabacum* L. *Plant Biotechnol. J.* **2009**, *7*,

- 219–226.
- 22. Kim, C.K.; Han, J.S.; Lee, H.S.; Oh, J.Y.; Shigaki, T.; Park, S.H.; Hirschi, K. Expression of an *Arabidopsis* CAX2 variant in potato tubers increases calcium levels with no accumulation of manganese. *Plant Cell Rep.* **2006**, *25*, 1226–1232.
  - 23. Mi, Y.C.; Han, J.S.; Giovannoni, J.; Yang, L.; Chang, K.K.; Lim, K.B.; Chung, J.D. Modest calcium increase in tomatoes expressing a variant of *Arabidopsis* cation/H<sup>+</sup> antiporter. *Plant Biotechnol. Rep.* **2010**, *4*, 15–21.
  - 24. Shigaki, T.; Cheng, N.H.; Pittman, J.K.; Hirschi, K. Structural determinants of Ca<sup>2+</sup> transport in the *Arabidopsis* H<sup>+</sup>/Ca<sup>2+</sup> antiporter CA1. *J. Biol. Chem.* **2001**, *276*, 43152–43159.
  - 25. Zhao, J.; Barkla, B.J.; Marshall, J.; Pittman, J.K.; Hirschi, K.D. The *Arabidopsis* *cax3* mutants display altered salt tolerance, pH sensitivity and reduced plasma membrane H<sup>+</sup>-ATPase activity. *Planta* **2008**, *227*, 659–669.
  - 26. Chen, J.; Takano, T.; Liu, S.; Bu, Y. Identification and characterization of a cation<sup>2+</sup>/H<sup>+</sup> antiporter Atcax4 gene from *Arabidopsis thaliana*. *Mol. Soil. Biol.* **2016**, *7*, 1–5.
  - 27. Liao, Q.; Jian, S.F.; Song, H.X.; Guan, C.Y.; Lepo, J.; Ismail, A.M.; Zhang, Z.H. Balance between nitrogen use efficiency and cadmium tolerance in *Brassica napus* and *Arabidopsis thaliana*. *Plant Sci.* **2019**, *284*, 57–66.
  - 28. Jeong, S.W.; Han, J.S.; Kim, K.M.; Oh, J.Y.; Chung, J.D. Expression of *Arabidopsis* CAX4 in tomato fruits increases calcium level with no accumulation of other metallic cations. *J. Plant Biotechnol.* **2008**, *35*, 337–343.
  - 29. Choe, M.; Choe, W.; Cha, S.; Lee, I. Changes of cationic transport in AtCAX5 transformant yeast by electromagnetic field environments. *J. Biol. Phys.* **2018**, *44*, 433–448.
  - 30. Qiao, K.; Wang, F.; Liang, S.; Hu, Z.; Chai, T. Heterologous expression of *TuCAX1a* and *TuCAX1b* enhances Ca<sup>2+</sup> and Zn<sup>2+</sup> translocation in *Arabidopsis*. *Plant Cell Rep.* **2019**, *38*, 597–607.
  - 31. Xu, L.; Zahid, K.R.; He, L.; Zhang, W.; He, X.; Zhang, X.; Yang, X.; Zhu, L. GhCAX3 gene, a novel Ca<sup>2+</sup>/H<sup>+</sup> exchanger from cotton, confers regulation of cold response and ABA induced signal transduction. *PLoS ONE* **2013**, *8*, e66303.
  - 32. Zhang, L.; Hao, J.; Bao, M.; Hasi, A.; Niu, Y. Cloning and characterization of a Ca<sup>2+</sup>/H<sup>+</sup> exchanger from the halophyte *Salicornia europaea* L. *Plant Physiol. Biochem.* **2015**, *96*, 321–328.
  - 33. Martins, V.; Carneiro, F.; Conde, C.; Sottomayor, M.; Geros, H. The grapevine VvCAX3 is a cation/H(+) exchanger involved in vacuolar Ca<sup>(2+)</sup> homeostasis. *Planta* **2017**, *246*, 1083–1096.