

Rational design of CYP3A4 inhibitors: A one-atom linker elongation in ritonavir-like compounds leads to a marked improvement in the binding strength

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SUPPLEMENTAL MATERIALS

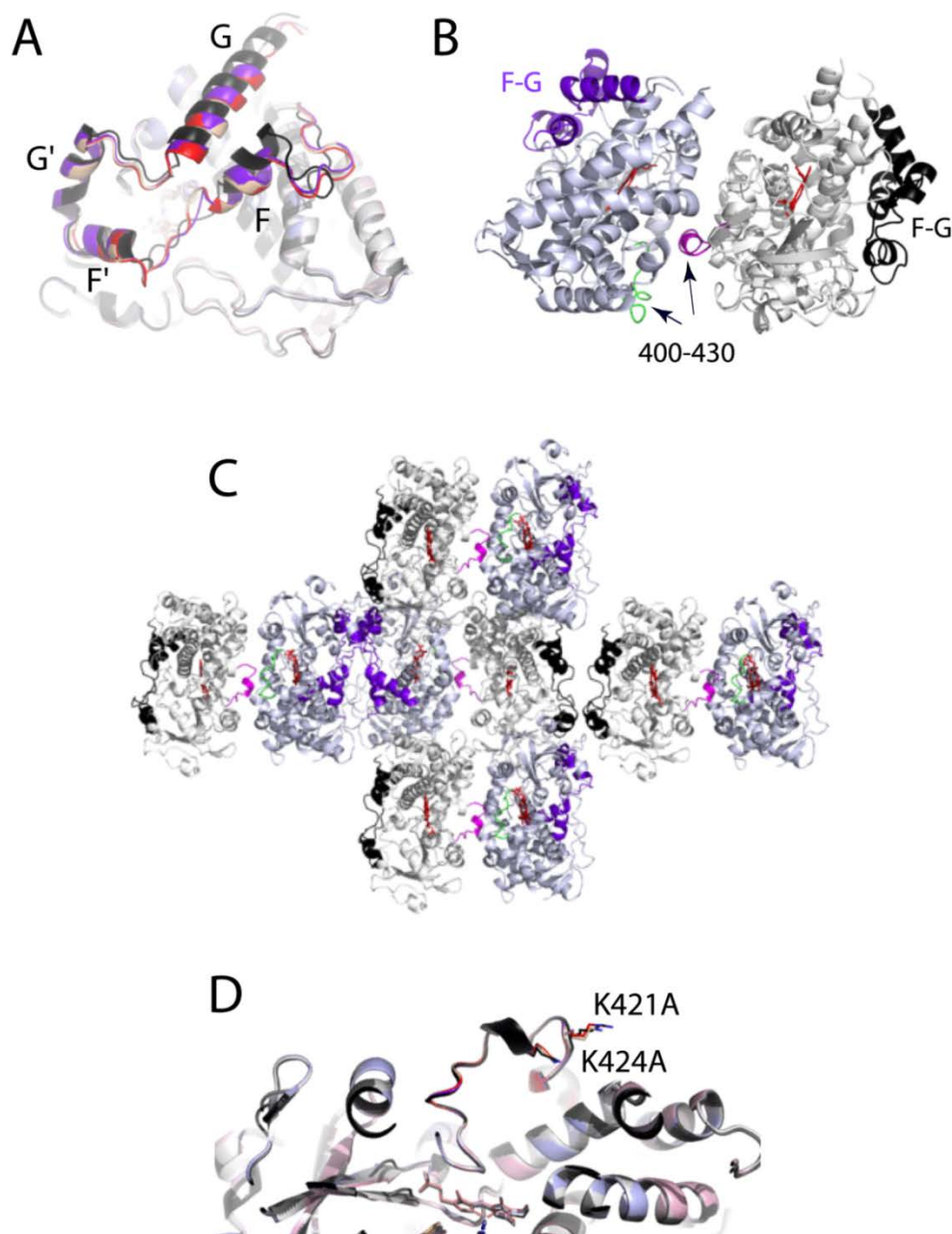


Figure S1. (A) Restructuring of the F-F' helix/loop region triggered by distinct crystal packing in the C2 space group. The F-F'-G'-G fragments of ligand-free (5VCE; I222) and **3a**-, **3e**- and **3i**-bound CYP3A4 (C2) are shown in black, red, beige and purple, respectively. (B, C) Crystallographic dimer of CYP3A4 and its spatial arrangement in C2 crystals, respectively. The 400-430 fragment (in green and magenta) is at the monomer-monomer interface, whereas the F-G fragment (in black and purple) mediates inter-dimer contacts. (D) A view at the proximal face of CYP3A4. The water-bound protein (5VCC structure) is in gray; **3a**- and **3e**-bound complexes of WT CYP3A4 are in pink and light blue, respectively; and **3i**-bound K421A/K424A CYP3A4 is in black. Structural superposition shows that substitution of surface K421 and K424 with alanine does not distort the proximal loop region. Non-mutated lysine side chains are displayed.

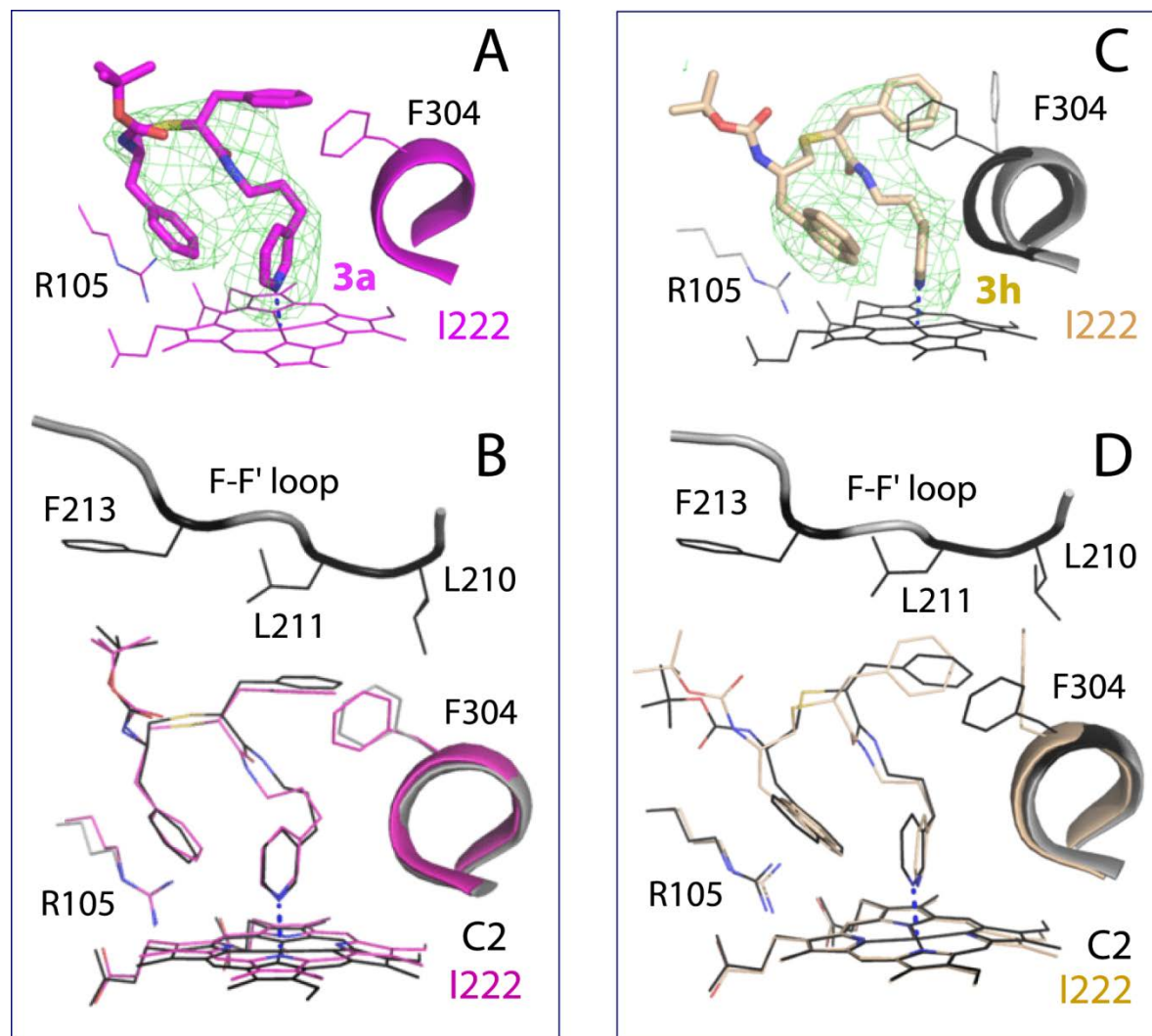


Figure S2. Binding of **3a** (**A**, **B**) and **3h** (**B**, **C**) to the active site of CYP3A4 crystallized in I222 and C2 space groups. For both **3a** and **3h**, electron density maps were not clearly defined in the I222 structures (green mesh in **A** and **C**, respectively). Therefore, **3a**- and **3h**-bound CYP3A4 was recrystallized in a more densely packed C2 space group. (**B**, **D**) Comparison of the ligand binding modes observed in the I222 and C2 structures shows that there are virtually no (for **3a**) or only minor distortions (for **3h**), primarily imposed by changes in the F-F' loop (residues 210-213).

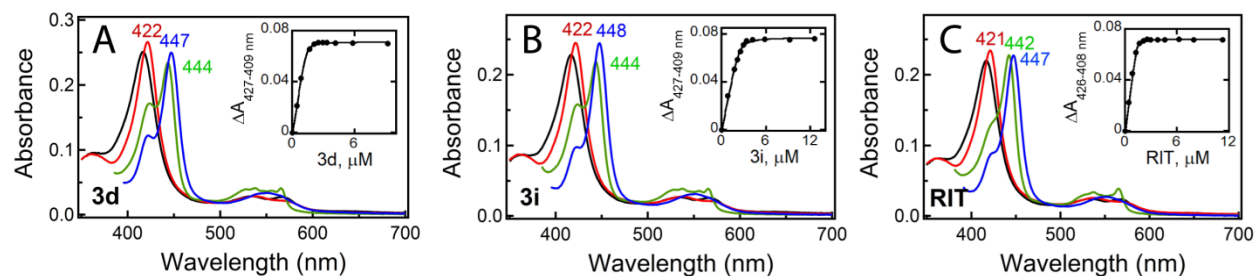


Figure S3. Spectral and ligand-binding properties of K421A/K424A CYP3A4. **(A-C)** Spectral changes induced in the mutant by **3d**, **3i** and ritonavir, respectively. Absorbance spectra of ferric ligand-free and inhibitor-bound CYP3A4 (2 μM) were recorded in 0.1 M phosphate buffer, pH 7.4, supplemented with 20% glycerol and 1 mM dithiothreitol, and displayed in black and red, respectively. Spectra of the ferrous form and its CO-adduct are in green and blue, respectively. The inhibitor concentration was 10 μM . Equilibrium titrations were conducted as described in the Materials and Methods section in the main text. Titration plots with quadratic fittings are shown in insets. The derived dissociation constants were 0,019 μM , 0.025 μM and 0.023 μM for **3d**, **3i** and ritonavir, respectively.

Table S1. Data collection and refinement statistics

Ligand	3a	3b	3c	3d	8
CYP3A4	WT	WT	WT	K421A/K424A	WT
PDB ID	7KNH	7KVI	7KVJ	7KVK	7KVM
<i>Data statistics</i>					
Space group	C2	I222	I222	C2	I222
Unit cell	$a, b, c = 93 \times 95 \times 153 \text{ \AA};$ $\alpha, \beta, \gamma = 90 \times 90 \times 124^\circ$	$a, b, c = 77 \times 103 \times 126 \text{ \AA};$ $\alpha, \beta, \gamma = 90^\circ$	$a, b, c = 76 \times 100 \times 124 \text{ \AA};$ $\alpha, \beta, \gamma = 90^\circ$	$a, b, c = 76 \times 101 \times 125 \text{ \AA};$ $\alpha, \beta, \gamma = 90^\circ$	$a, b, c = 78 \times 101 \times 129 \text{ \AA};$ $\alpha, \beta, \gamma = 90^\circ$
Resolution range (Å)	39.55-2.79 (2.79-2.94) ^a	79.64-2.55 (2.55-2.69)	78.17-2.65 (2.79-2.65)	77.80-2.55 (2.69-2.55)	79.64-2.70 (2.85-2.70)
Total reflections	167,933	81,756	51,716	159,292	56,723
Unique reflections	27,176	16,474	13,659	37,311	13,668
Redundancy	6.2 (6.0)	5.0 (4.8)	3.8 (4.0)	4.3 (4.4)	4.2 (4.3)
Completeness	98.3 (96.9)	99.2 (99.5)	97.1 (98.3)	99.8 (99.8)	95.9 (97.4)
Average I/σ	5.8 (1.1)	8.3 (1.0)	7.4 (1.0)	8.4 (1.1)	8.5 (0.9)
R_{merge}	0.163 (2.561)	0.077 (1.163)	0.070 (2.029)	0.080 (1.443)	0.047 (2.926)
R_{pim}	0.107 (1.155)	0.038 (0.592)	0.042 (1.127)	0.043 (0.768)	0.038 (1.358)
CC ½	0.993 (0.489)	0.998 (0.532)	0.998 (0.320)	0.999 (0.398)	0.996 (0.313)
<i>Refinement statistics</i>					
R/R_{free} ^b	23.8/27.8	22.5/26.2	22.5/28.0	23.7/27.7	22.7/27.6
Number of atoms:					
Protein	3657/3555 ^c	3654	3341	3716/3581	3720
Solvent	24	21	4	14	0
R.m.s. deviations					
Bond lengths, Å	0.002	0.002	0.002	0.002	0.003
Bond angles, °	0.490	0.531	0.490	0.467	0.661
Wilson B-factor, Å ²	83	79	96	73	112
Average B-factor, Å ² :					
Protein	91/110 ^c	112	136	89/131	141
Ligand	85/104 ^c	140	162	102/126	149
Ramachandran plot ^d (residues; %)					
Preferred	816 (93.0%)	427 (96.0%)	388 (95.0%)	848 (94.9%)	435 (95.0%)
Allowed	60 (6.8%)	17 (3.8%)	20 (5.0%)	44 (4.9%)	25 (5.0%)
Outliers	2 (0.2%)	1 (0.2%)	none	2 (0.2%)	none

^aValues in brackets are for the highest resolution shell.^b R_{free} was calculated from a subset of 5% of the data that were excluded during refinement.^cValues for two molecules in the asymmetric unit.^dAnalyzed with PROCHECK.

Table S2. Data collection and refinement statistics

Ligand	3e	3f	3g	3h	3i
CYP3A4	WT	WT	WT	WT	K421A/K424A
PDB ID	7KVN	7KVO	7KVP	7KVQ	7KVS
<i>Data statistics</i>					
Space group	C2	I2	I222	C2	C2
Unit cell	$a, b, c = 93 \times 96 \times 153 \text{ \AA};$ $\alpha, \beta, \gamma = 90 \times 90 \times 124^\circ$	$a, b, c = 93 \times 98 \times 128 \text{ \AA};$ $\alpha, \beta, \gamma = 90 \times 90 \times 93^\circ$	$a, b, c = 78 \times 102 \times 128 \text{ \AA};$ $\alpha, \beta, \gamma = 90^\circ$	$a, b, c = 93 \times 97 \times 154 \text{ \AA};$ $\alpha, \beta, \gamma = 90 \times 90 \times 124^\circ$	$a, b, c = 93 \times 98 \times 155 \text{ \AA};$ $\alpha, \beta, \gamma = 90 \times 90 \times 124^\circ$
Resolution range (Å)	76.93-2.70 (2.85-2.70) ^a	49.04-2.65 (2.79-2.65)	80.00-2.75 (2.907-2.75)	77.28-2.70 (2.85-2.70)	77.82-2.50 (2.64-2.50)
Total reflections	136,528	223,503	43,178	177,490	246,329
Unique reflections	28,152	33,059	13,250	30,116	38,953
Redundancy	4.8 (4.3)	6.8 (6.7)	3.3 (3.4)	5.9 (6.0)	6.3 (6.3)
Completeness	91.9 (84.0)	99.9 (99.9)	97.3 (98.6)	94.4 (97.2)	97.6 (97.4)
Average I/σ	4.0 (1.0)	12.3 (1.1)	5.9 (0.7)	8.5 (1.1)	11.0 (1.0)
R_{merge}	0.233 (1.598)	0.070 (2.278)	0.072 (2.466)	0.093 (1.641)	0.075 (1.922)
R_{pim}	0.115 (0.841)	0.029 (0.944)	0.045 (1.566)	0.040 (0.723)	0.032 (0.830)
CC $\frac{1}{2}$	0.970 (0.571)	0.999 (0.390)	0.998 (0.481)	0.999 (0.372)	0.999 (0.345)
<i>Refinement statistics</i>					
R/R_{free}^b	24.8/29.0	24.1/29.3	22.7/27.3	25.0/28.0	23.5/27.5
Number of atoms:					
Protein	3653/3646 ^c	3721/3631	3653	3669/3605	3689/3538
Solvent	88	0	2	0	47
R.m.s. deviations					
Bond lengths, Å	0.003	0.003	0.004	0.002	0.002
Bond angles, °	0.563	0.570	0.731	0.497	0.500
Wilson B-factor, Å ²	69	91	113	89	80
Average B-factor, Å ² :					
Protein	75/93 ^c	107/160	96	100/167	99/151
Ligand	65/88 ^c	164/177	163	128/177	132/182
Ramachandran plot ^d (residues; %)					
Preferred	826 (92.9%)	842 (94.1%)	435 (97.0%)	837 (93.9%)	836 (94.1%)
Allowed	62 (9.0%)	52 (5.8%)	14 (3.0%)	53 (6.0%)	51 (5.8%)
Outliers	1 (0.1%)	1 (0.1%)	none	1 (0.1%)	1 (0.1%)

^aValues in brackets are for the highest resolution shell.^b R_{free} was calculated from a subset of 5% of the data that were excluded during refinement.^cValues for two molecules in the asymmetric unit.^dAnalyzed with PROCHECK

Table S3. Local correlation coefficients (CC) and CC_{peak} between three polder maps ($F_{\text{obs}} = |F_{\text{model}}|$)

Map 1 (m1), calculated F_{obs} assuming that the omitted ligand atoms are present. Map 2 (m2), calculated F_{obs} assuming that the omitted ligand atoms are not present. Map 3 (m3), polder map using experimental data.

Ligand/PDB ID	m1-m2		m1-m3		m2-m3	
	CC	CC_{peak}	CC	CC_{peak}	CC	CC_{peak}
3a 7KVH	0.4949	0.4797	0.8790	0.8579	0.4998	0.4709
3b 7KVI	0.6654	0.6106	0.8934	0.8439	0.6464	0.6431
3c 7KNJ	0.6304	0.6167	0.8333	0.7082	0.4517	0.3954
3d 7KVK	0.5390	0.4609	0.9078	0.8797	0.5149	0.4463
8 7KVM	0.6183	0.5721	0.8442	0.7824	0.6879	0.6878
3e 7KVN	0.5025	0.5048	0.8715	0.8431	0.4884	0.4780
3f 7KVO	0.7812	0.7598	0.8015	0.7899	0.6965	0.6741
3g 7KVP	0.6169	0.6394	0.8884	0.8461	0.5352	0.5618
3h 7KVQ	0.6701	0.6616	0.8005	0.7598	0.6414	0.6466
3i 7KVS	0.6324	0.6383	0.8551	0.8222	0.7047	0.6985

Correlation coefficients for polder maps were calculated using *phenix.polder* in the PHENIX software package.¹

For all ligands, CC(1,3) was > 0.8 and higher than CC(1,2) and CC(2,3), meaning that the polder maps displayed in Figures 3 and 6 of the main text show the omitted ligand atoms.²

¹Adams, P.D.; Afonine, P.V.; Bunkoczi, G.; Chen, V.B.; Davis, I.W.; Echols, N.; Headd, J.J.; Hung, L.W.; Kapral, G.J.; Grosse-Kunstleve, R.W., et al. PHENIX: a comprehensive Python-based system for macromolecular structure solution. *Acta Crystallogr. Section D* **2010**, *66*, 213-321

²Liebschner D., Afonine P.V.; Moriarty N.W.; Poon B.K.; Sobolev O.V.; Terwilliger T.C.; Adams P.D. Polder maps: improving OMIT maps by excluding bulk solvent. *Acta Crystallogr. Section D* **2016**, *73*, 148-157.

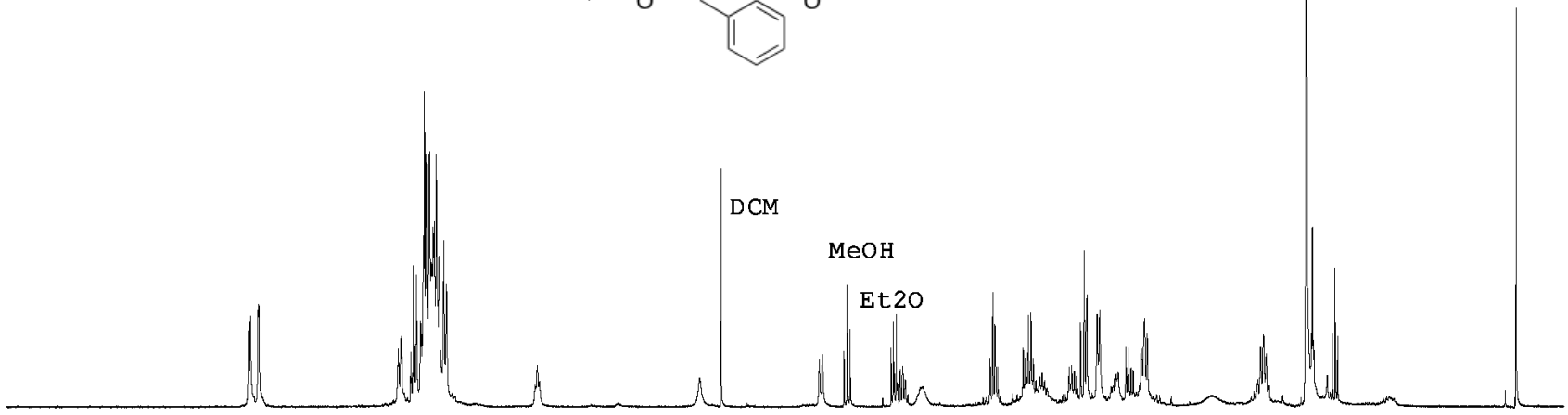
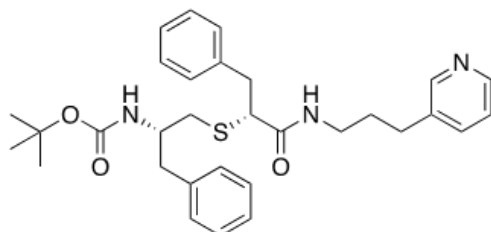
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¹H spectrum

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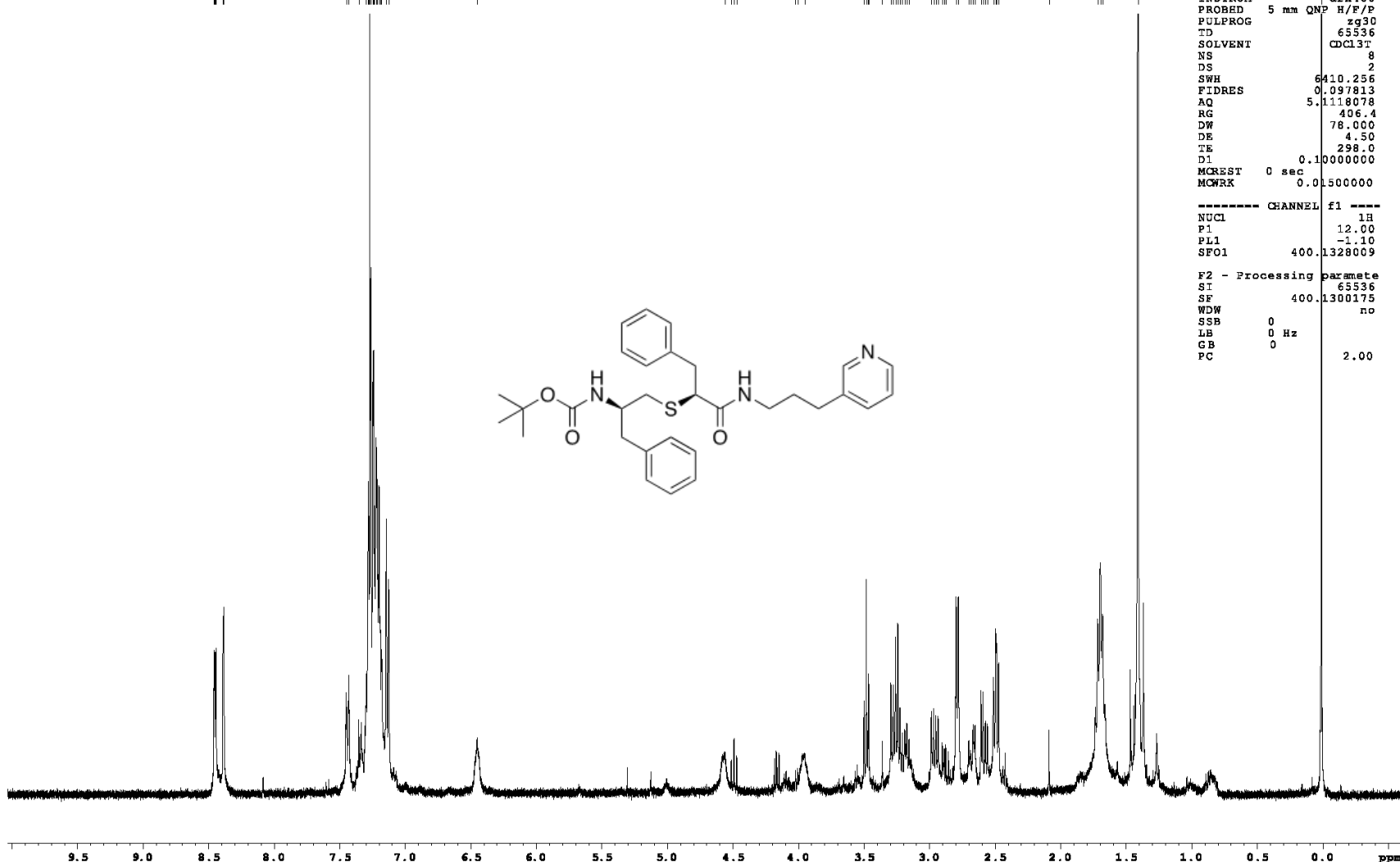
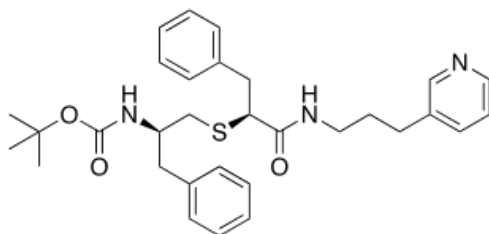
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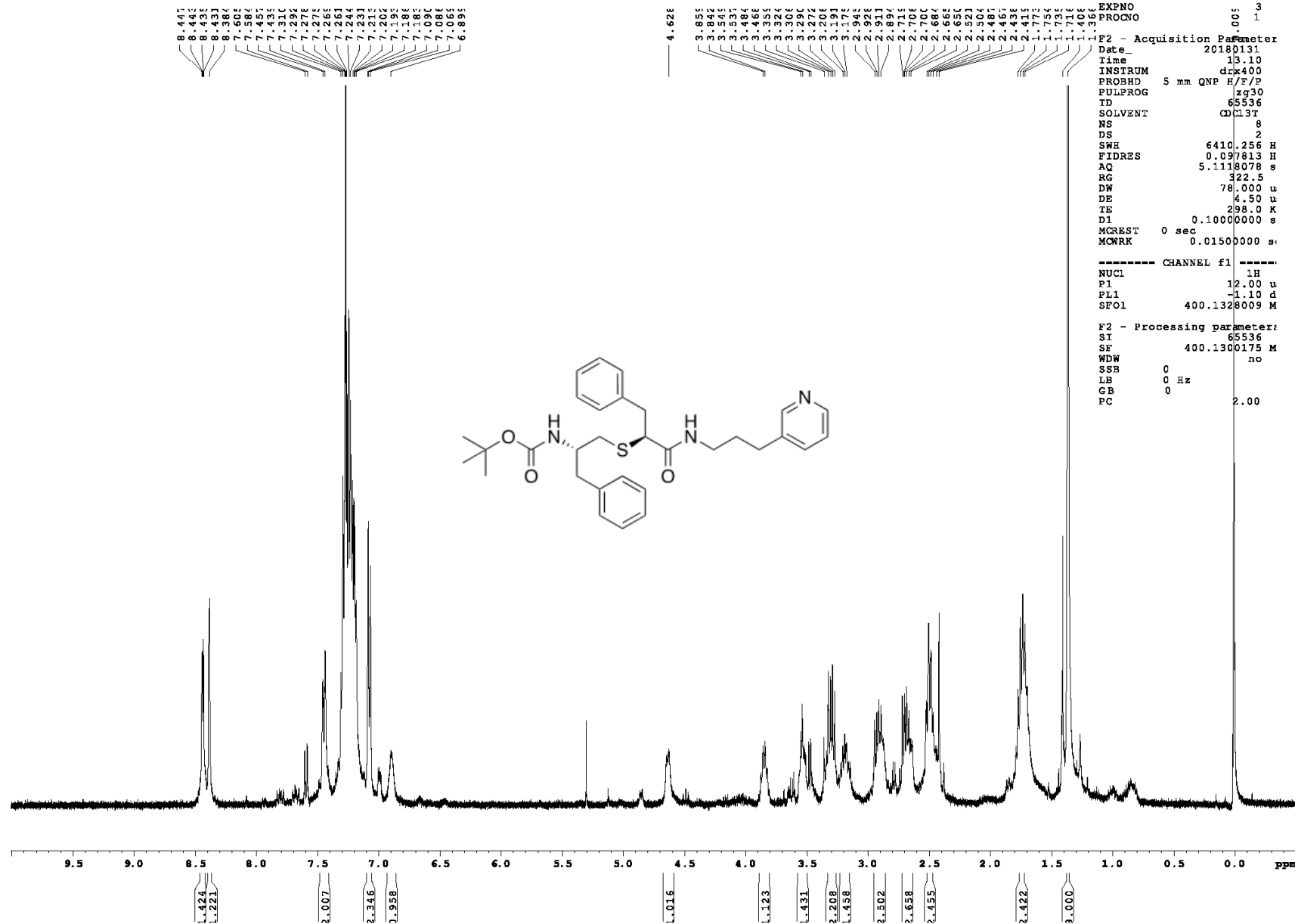
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¹H spectrum

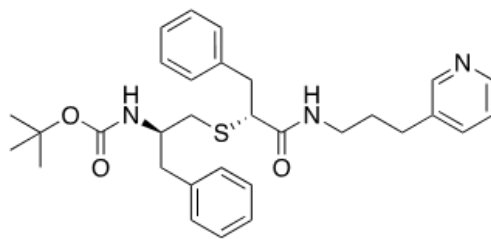


1H spectrum

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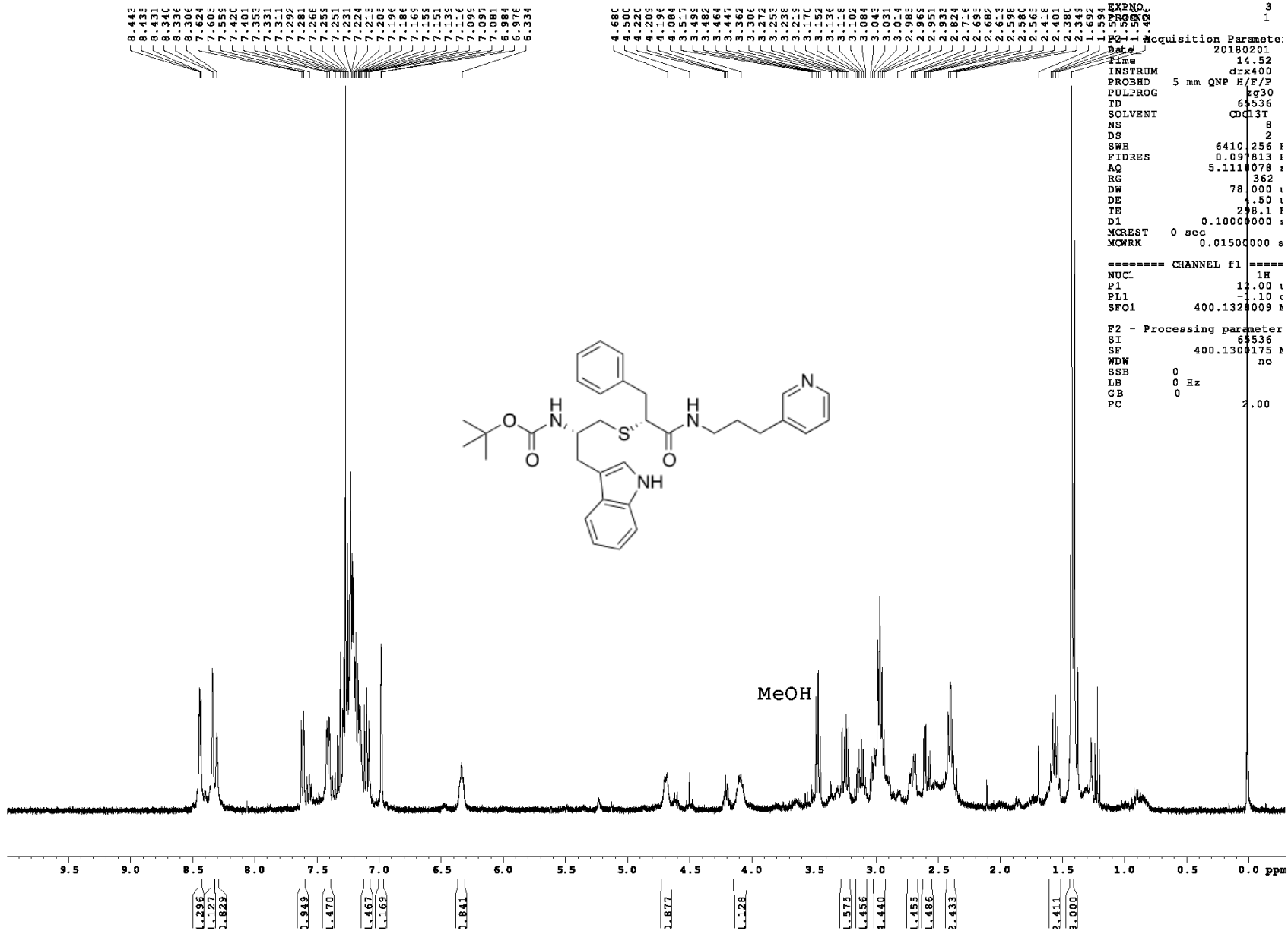
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¹H spectrum



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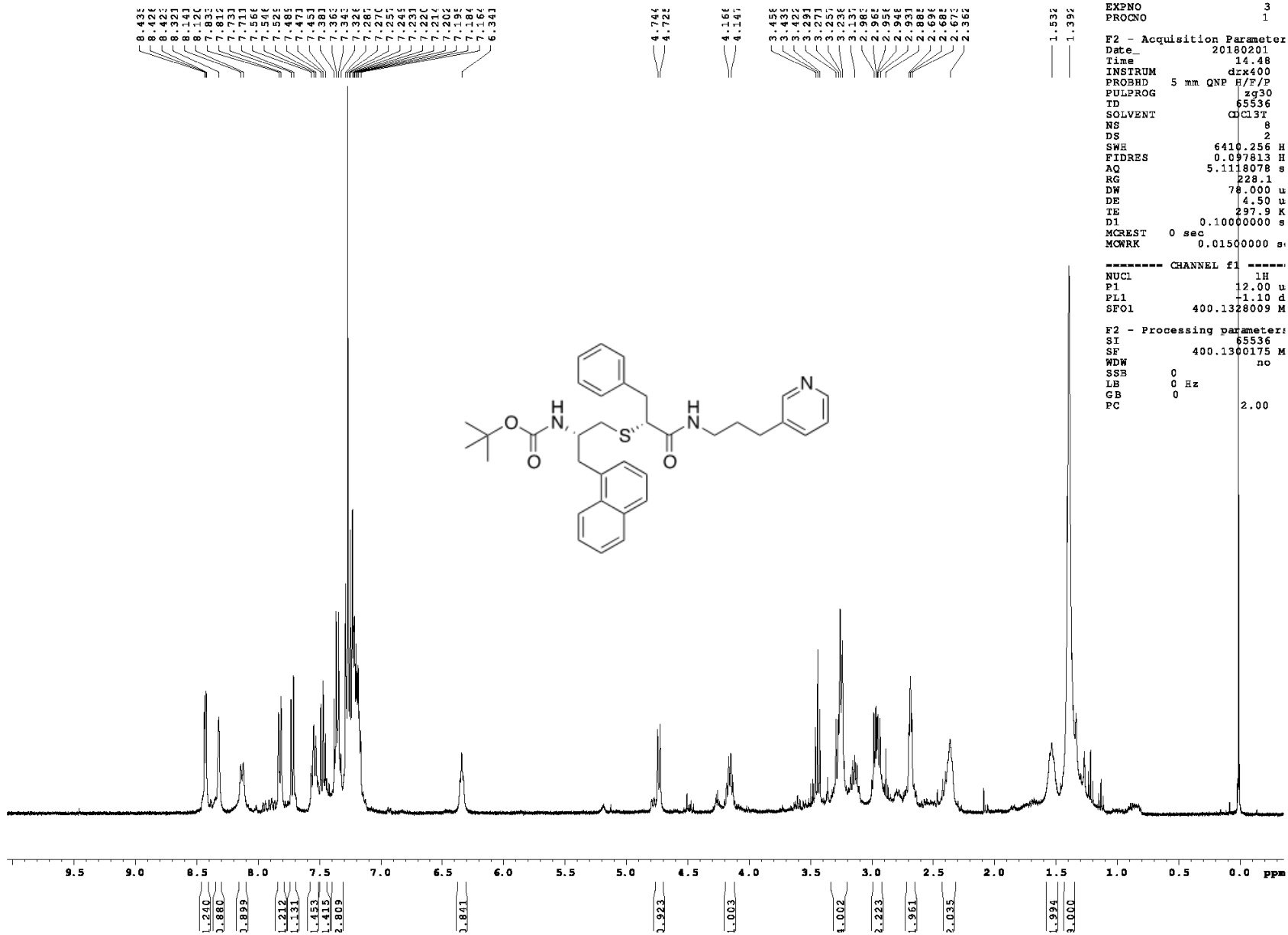
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¹H spectrum



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¹H spectrum

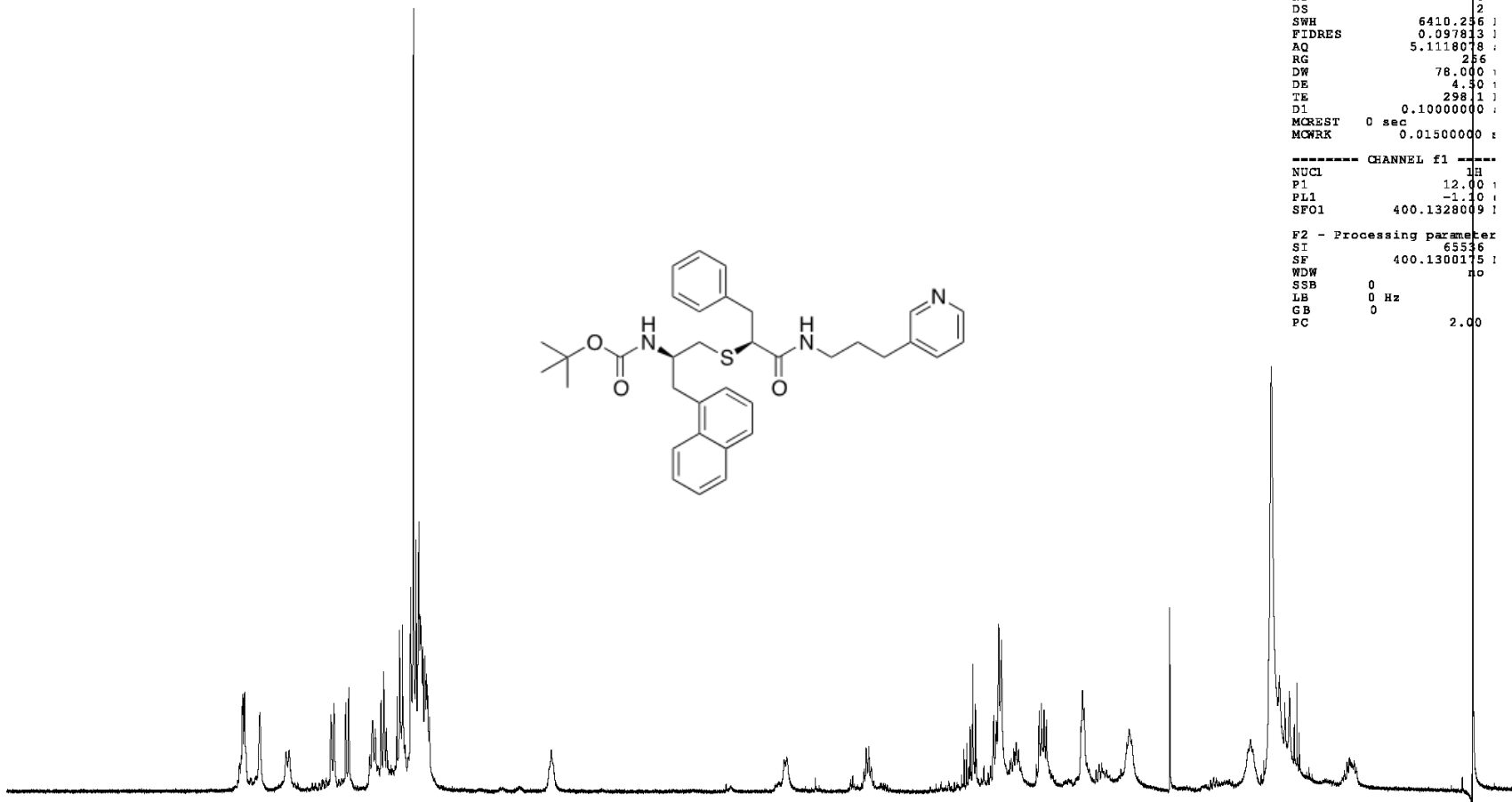
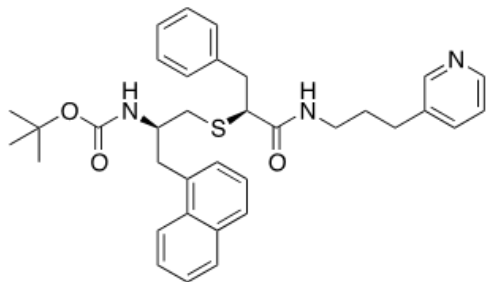
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7.175
7.171

6.324

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4.141
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Time 14.42
INSTRUM dx400
PROBHD 5 mm QNP H/R/P
PULPROG zgpg30
TD 65536
SOLVENT CDCl3
NS 8
DS 2
SWH 6410.236 |
FIDRES 0.097813 |
AQ 5.1118078 |
RG 256
DW 78.000 |
DE 4.30 |
TE 298.11 |
D1 0.1000000 |
MCREST 0 sec
MCWRK 0.0150000 |
----- CHANNEL f1 -----
NUC1 ¹H
P1 12.00 |
PL1 -1.10 |
SFO1 400.1328009 |
F2 - Processing parameter
SI 65536
SF 400.1300175 |
WDW no
SSB 0
LB 0 Hz
GB 0
PC 2.00



1.594
1.948
1.038
1.341
1.251
1.804
1.756
3.081

1.987

1.950

1.084

1.308

2.276

2.040

2.140

2.180

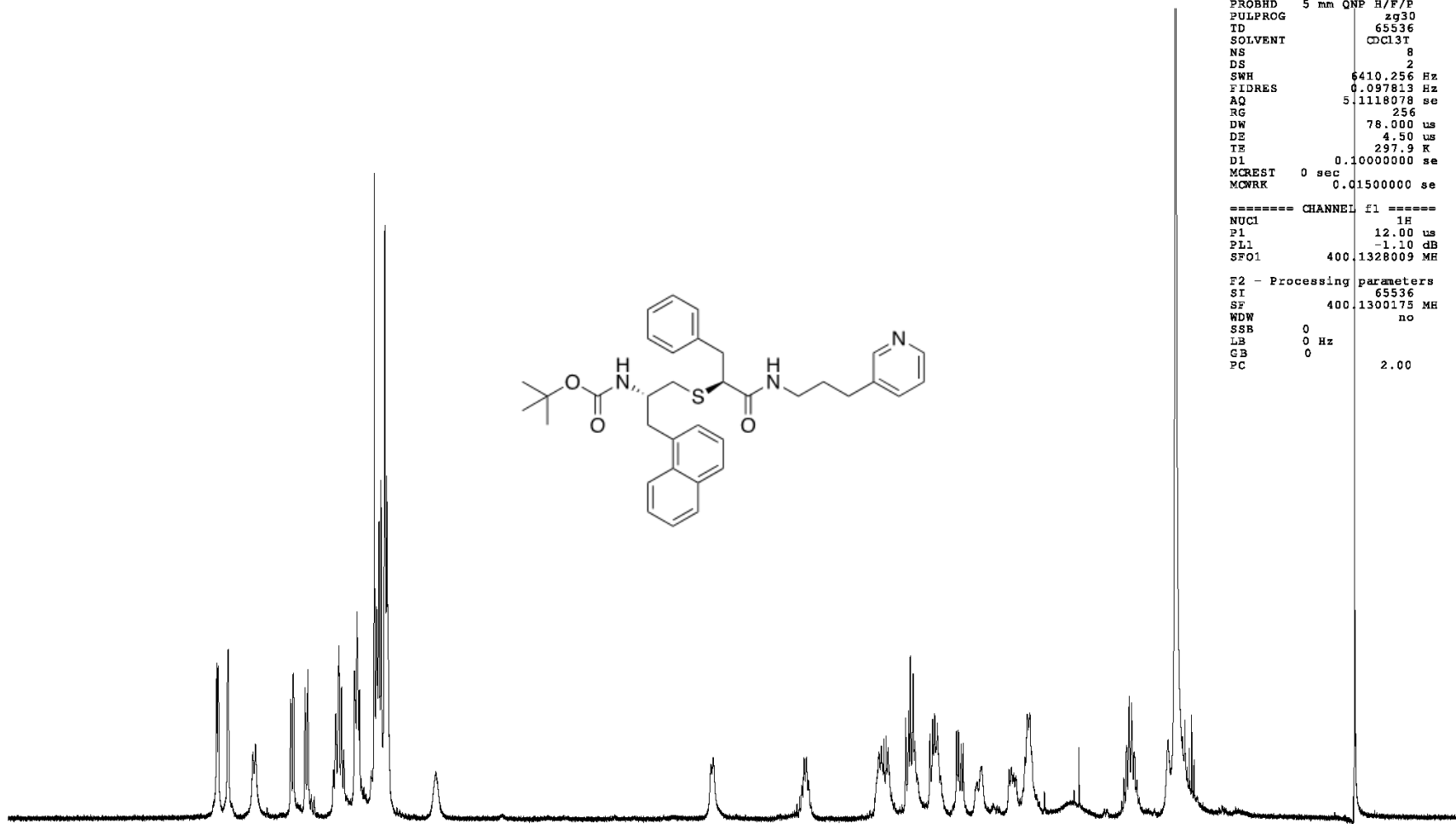
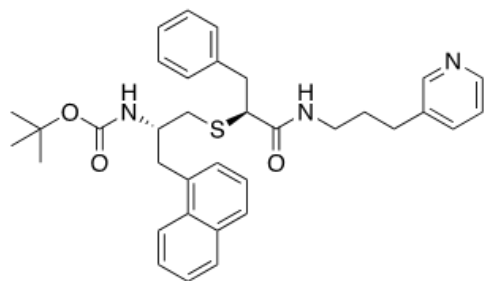
3.000

¹H spectrum

8.437
8.433
8.423
8.421
8.350
8.167
8.146
7.886
7.865
7.781
7.760
7.570
7.534
7.512
7.494
7.415
7.396
7.378
7.365
7.265
7.250
7.234
7.215
7.197
7.176
6.814

4.758

4.103
4.086
4.065
3.530
3.514
3.497
3.475
3.462
3.334
3.310
3.296
3.266
3.151
3.131
3.115
3.095
2.958
2.942
2.924
2.905
2.770
2.568
2.532
2.437
2.414
2.051
1.715
1.697
1.680
1.662
1.635
1.622
1.336



Current Data Parameters
NAME 04ERS129
EXPNO 3
PROCNO 1

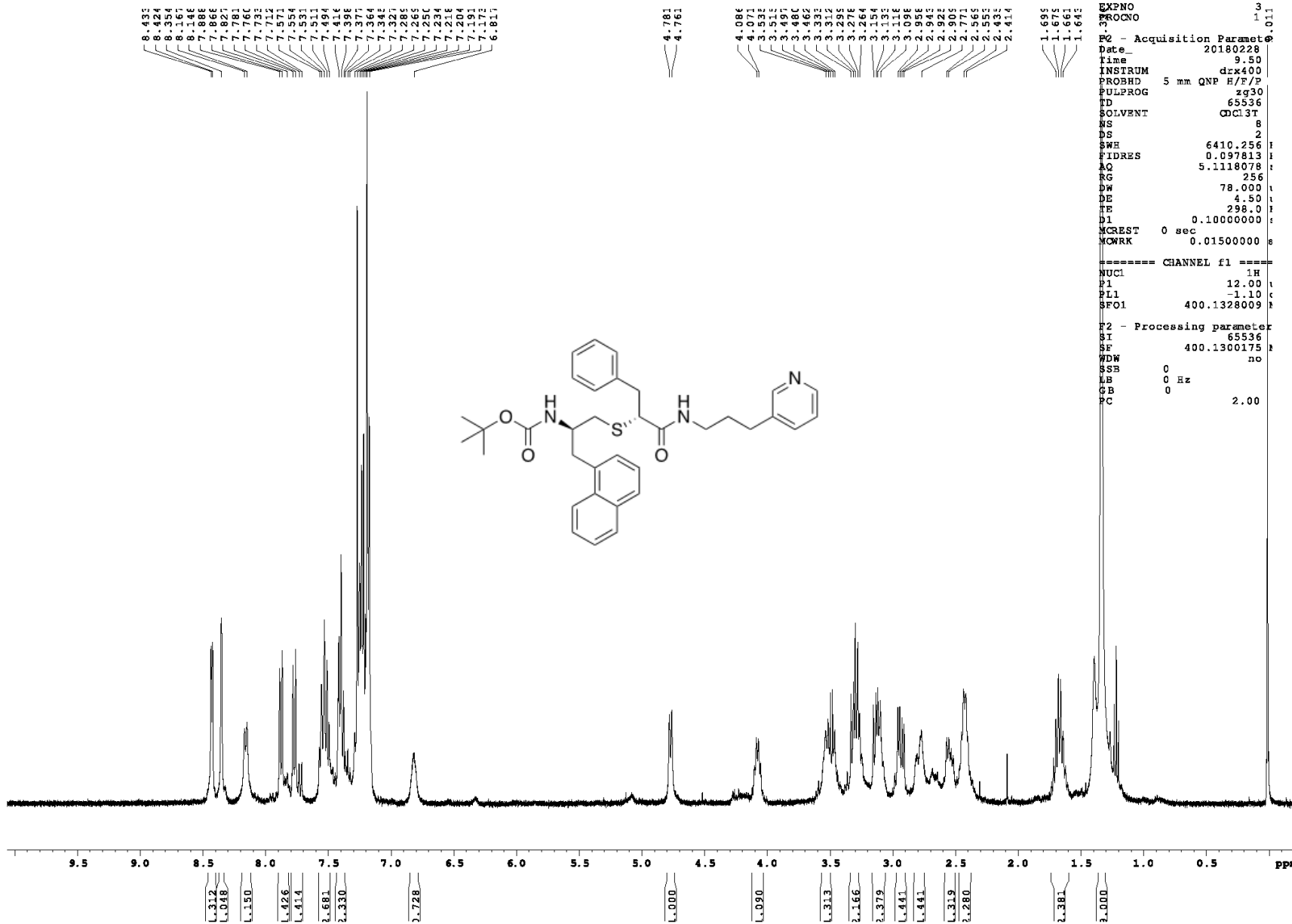
F2 - Acquisition Parameters
Date_ 20180202
Time 13.58
INSTRUM drx400
PROBHD 5 mm QNP H/F/P
PULPROG zg30
TD 65536
SOLVENT CDCl3T
NS 8
DS 2
SWH 6410.256 Hz
FIDRES 0.097813 Hz
AQ 5.1118078 se
RG 256
DW 78.000 us
DE 4.50 us
TE 297.9 K
D1 0.10000000 se
MCREST 0 sec
MCWRK 0.01500000 se

----- CHANNEL f1 -----
NUC1 1H
P1 12.00 us
PL1 -1.10 dB
SFO1 400.1328009 MHz

F2 - Processing parameters
SI 65536
SF 400.1300175 MHz
WDW no
SSB 0
LB 0 Hz
GB 0
PC 2.00

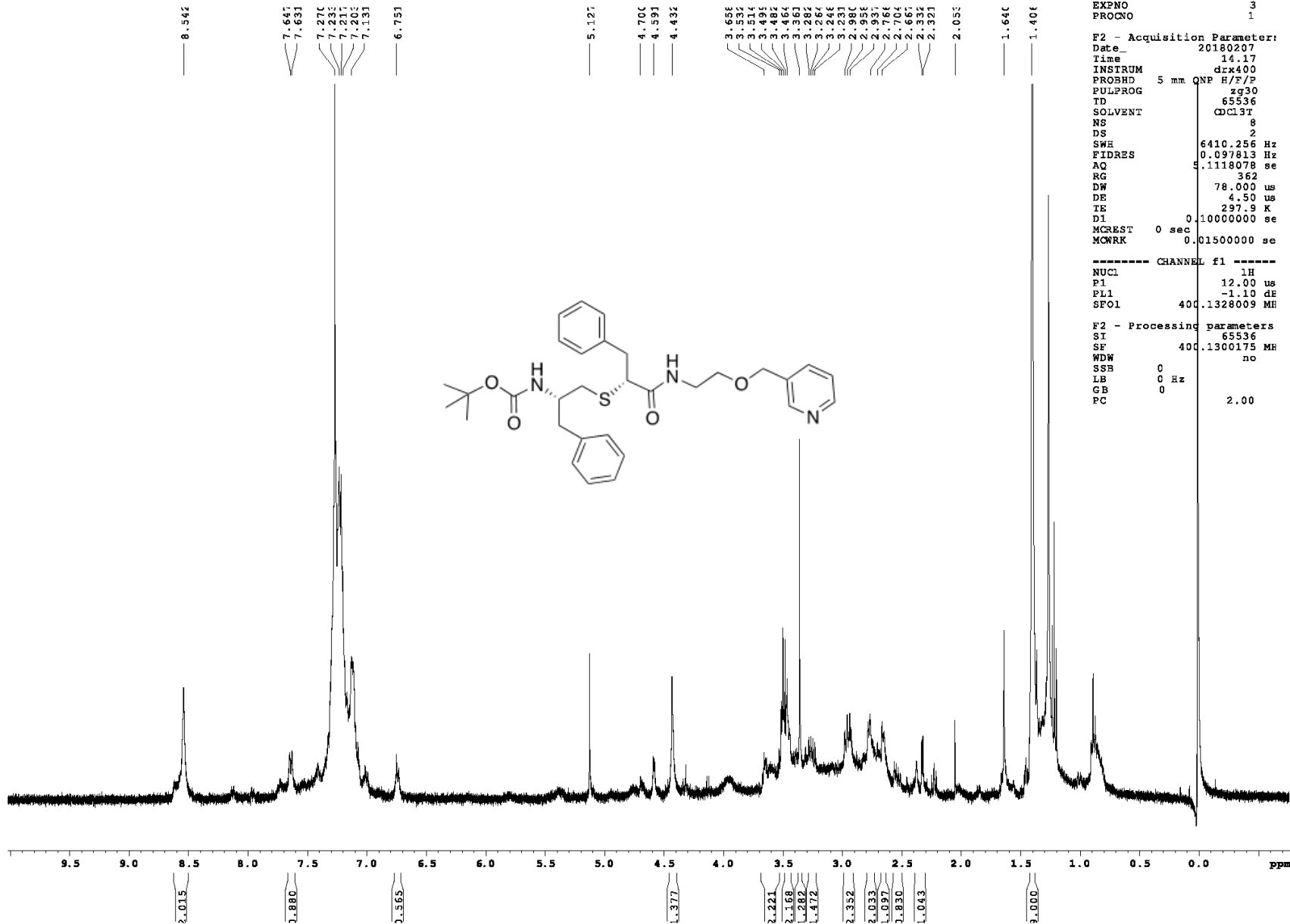
1.105
1.922
1.948
1.977
1.010
1.292
1.215
1.719
1.852
1.984
1.030
1.153
1.169
1.178
1.971
1.033
1.117
1.119
1.000

¹H spectrum



Current Data Parameters
NAME 04ERS145
EXPNO 3
PROCNO 1
F2 - Acquisition Parameters
Date_ 20180228
Time 9.50
INSTRUM dx400
PROBHD 5 mm QNP H₂/P
PULPROG zg30
TD 65536
SOLVENT CDCl₃
NS 8
DS 2
SWH 6410.256 MHz
FIDRES 0.097813 Hz
AQ 5.1118078 sec
RG 256
DW 78.000 nsec
DE 4.50 nsec
TE 298.0 K
D1 0.10000000 sec
MCREST 0 sec
MCWRK 0.01500000 sec
----- CHANNEL f1 -----
NUC1 ¹H
P1 12.00 nsec
PL1 -1.10 dB
SFO1 400.1328009 MHz
F2 - Processing parameters
SI 65536
SF 400.1300175 MHz
WDW no
SSB 0
LB 0 Hz
GB 0
PC 2.00

¹H spectrum



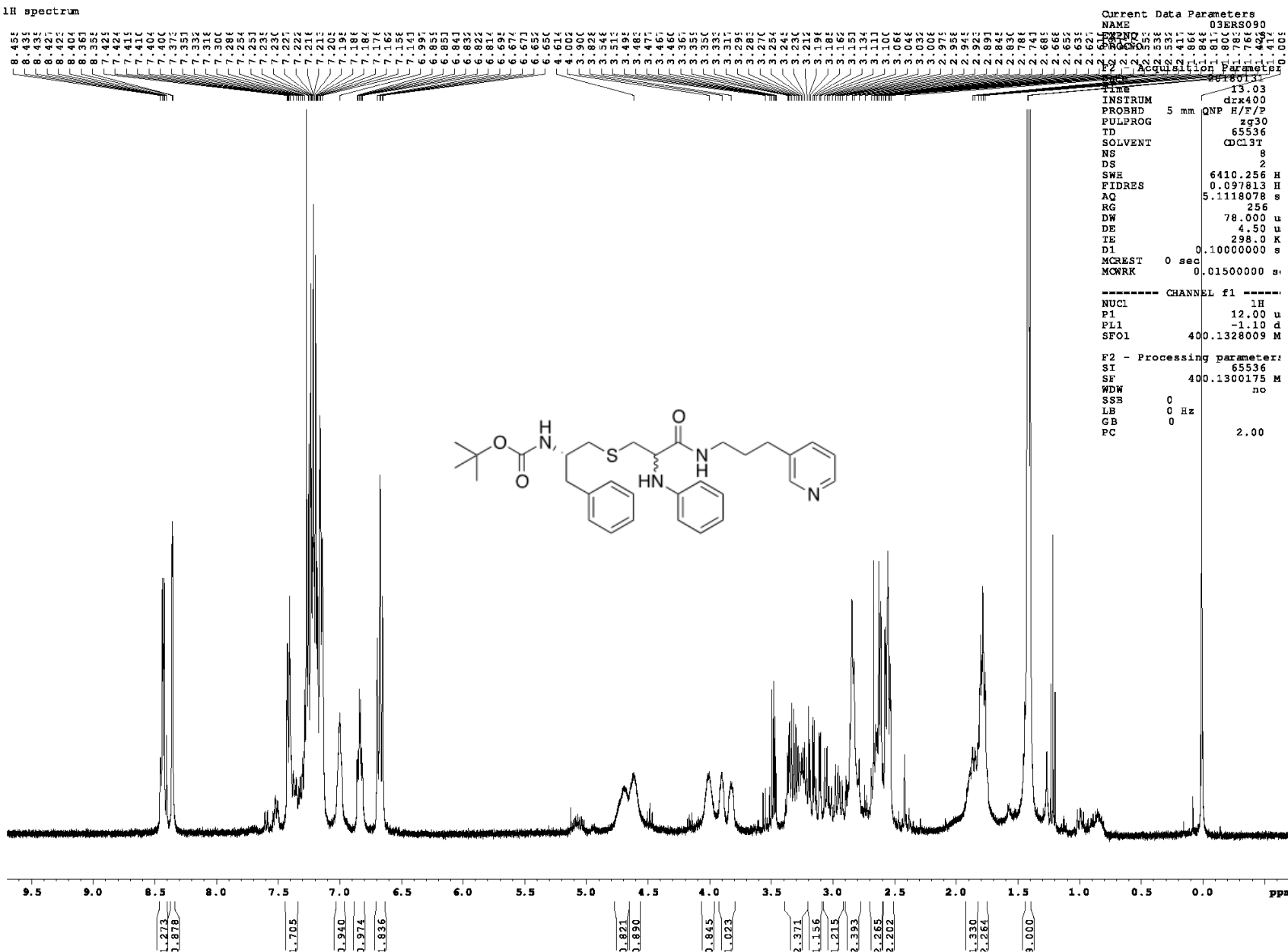
Current Data Parameters
NAME 04ERS102
EXPNO 3
PROCNO 1

F2 - Acquisition Parameters:
Date_ 20180207
Time 14.17
INSTRUM drx400
PROBHD 5 mm QNP H/P/P
PULPROG zg30
TD 65536
SOLVENT CDCl3
NS 8
DS 2
SWE 6410.256 Hz
FIDRES 0.097813 Hz
AQ 5.1118078 se
RG 362
DW 78.000 us
DE 4.50 us
TE 297.9 K
D1 0.10000000 se
MCREST 0 sec
MCWRK 0.01500000 se

----- CHANNEL f1 -----
NUC1 1H
P1 12.00 us
PL1 -1.10 dB
SFO1 400.1328009 MHz

F2 - Processing parameters
SI 65536
SF 400.1300175 MHz
WDW no
SSB 0
LB 0 Hz
GB 0
PC 2.00

¹H spectrum

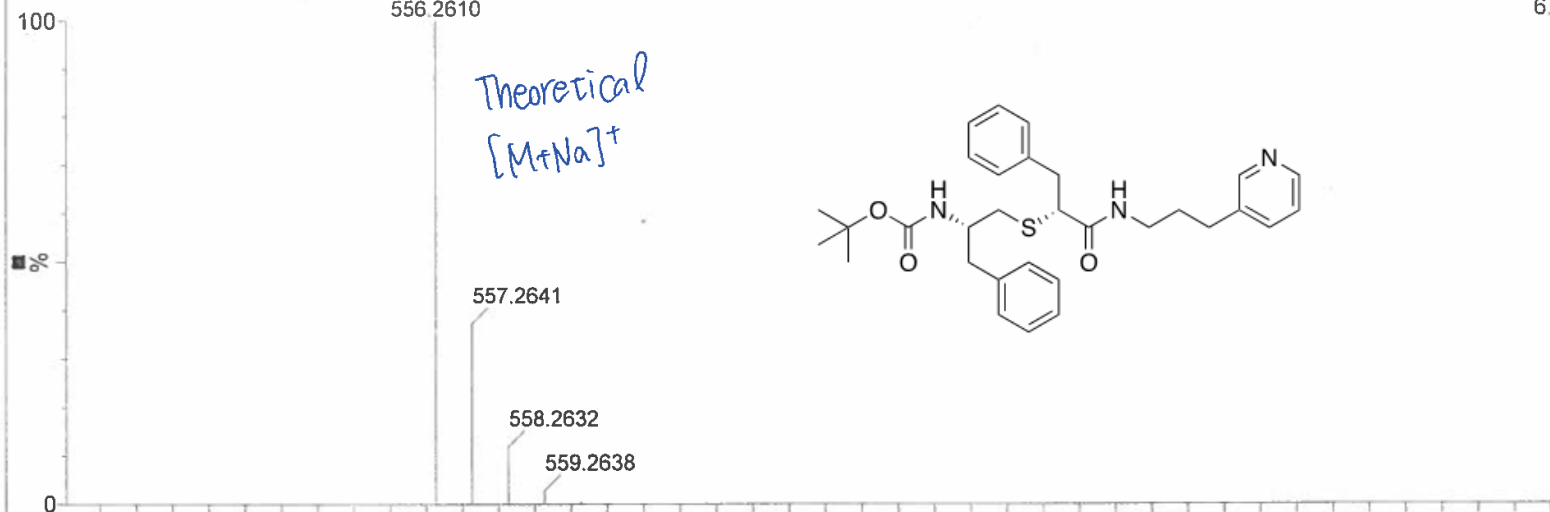


8.452
8.438
8.435
8.425
8.422
8.402
8.361
8.358
7.425
7.424
7.415
7.410
7.404
7.400
7.375
7.350
7.332
7.302
7.288
7.254
7.251
7.235
7.230
7.221
7.222
7.216
7.213
7.205
7.198
7.186
7.176
7.174
7.162
7.158
7.141
6.997
6.855
6.853
6.841
6.832
6.822
6.816
6.692
6.675
6.657
6.650
4.614
4.002
3.900
3.828
3.548
3.511
3.492
3.482
3.471
3.461
3.460
3.350
3.335
3.333
3.317
3.295
3.283
3.270
3.254
3.245
3.230
3.230
3.212
3.198
3.182
3.152
3.139
3.133
3.111
3.100
3.068
3.048
3.032
3.008
3.006
2.975
2.958
2.941
2.925
2.899
2.883
2.843
2.781
2.741
2.681
2.668
2.652
2.635
2.621
2.607

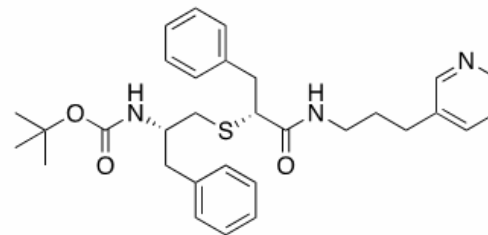
Current Data Parameters
 NAME 03ERS090
 EXPNO 2
 PROCNO 2
 F2 Acquisition Parameters
 Time 13.03
 INSTRUM dirx400
 PROBHD 5 mm QNP H/F/P
 PULPROG zg30
 TD 65536
 SOLVENT CDCl3T
 NS 8
 DS 2
 SWH 6410.256 H
 FIDRES 0.097813 H
 AQ 5.1118078 s
 RG 256
 DW 78.000 u
 DE 4.50 u
 TE 298.0 K
 D1 0.10000000 s
 MCREST 0 sec
 MCWRK 0.01500000 s
 ----- CHANNEL f1 -----
 NUCL ¹H
 P1 12.00 u
 PL1 -1.10 d
 SF01 400.1328009 M
 F2 - Processing parameters:
 SI 65536
 SF 400.1300175 M
 WDW no
 SSB 0
 LB 0 Hz
 GB 0
 PC 2.00

ES-377-a (0.019) Is (1.00,0.01) C₃₁H₃₉N₃O₃Na

TOF MS ES+
6.58e12

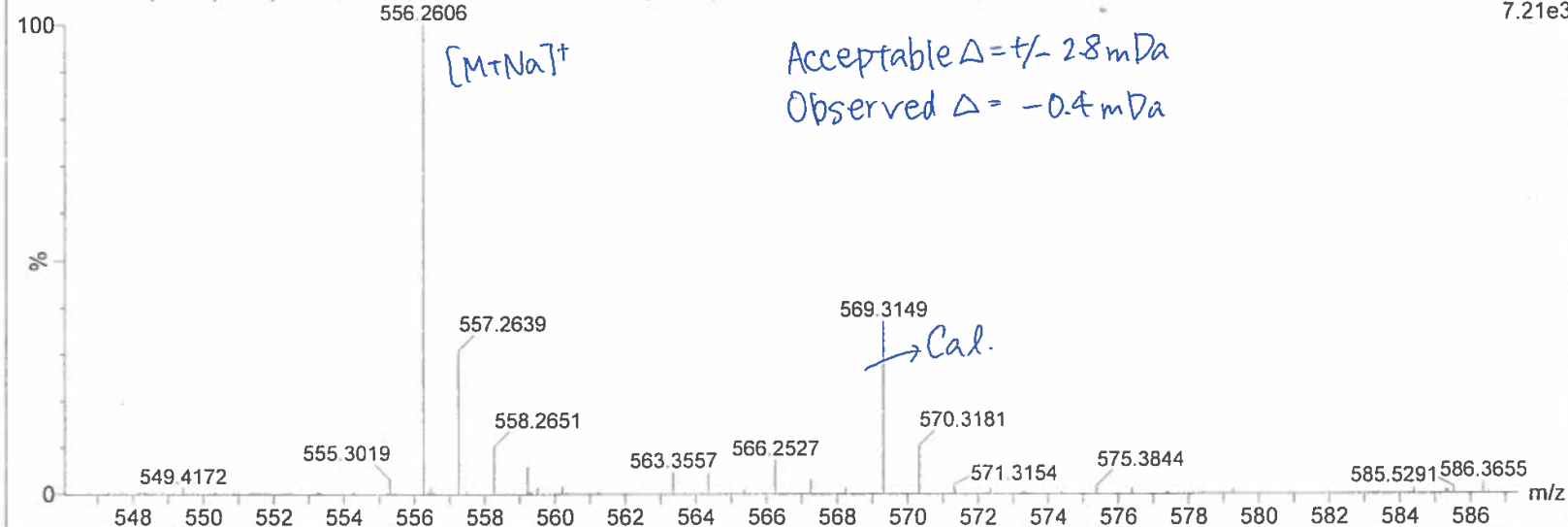


Theoretical
[M+Na]⁺



ES-377-a 18 (0.330) AM (Cen,5, 80.00, Ar,8000.0,569.31,0.70); Sm (SG, 2x3.00); Sb (1,40.00)

TOF MS ES+
7.21e3

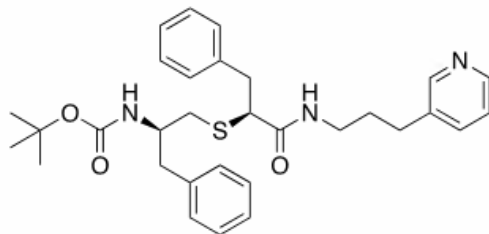


[M+Na]⁺

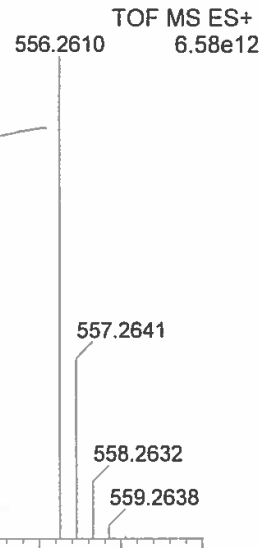
Acceptable $\Delta = \pm 2.8$ mDa
Observed $\Delta = -0.4$ mDa

Cal.

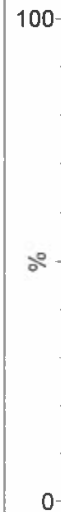
4109_a (0.019) Is (1.00,0.01) C₃₁H₃₉N₃O₃Na



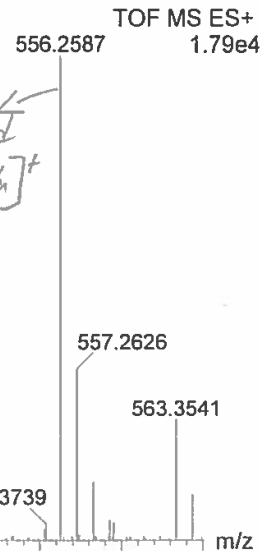
Theoretical $[M+Na]^+$



4109_a 20 (0.367) AM (Cen,5, 80.00, Ar,8000.0,481.26,0.70); Sm (SG, 2x3.00); Sb (1,40.00); Cm (17:31)

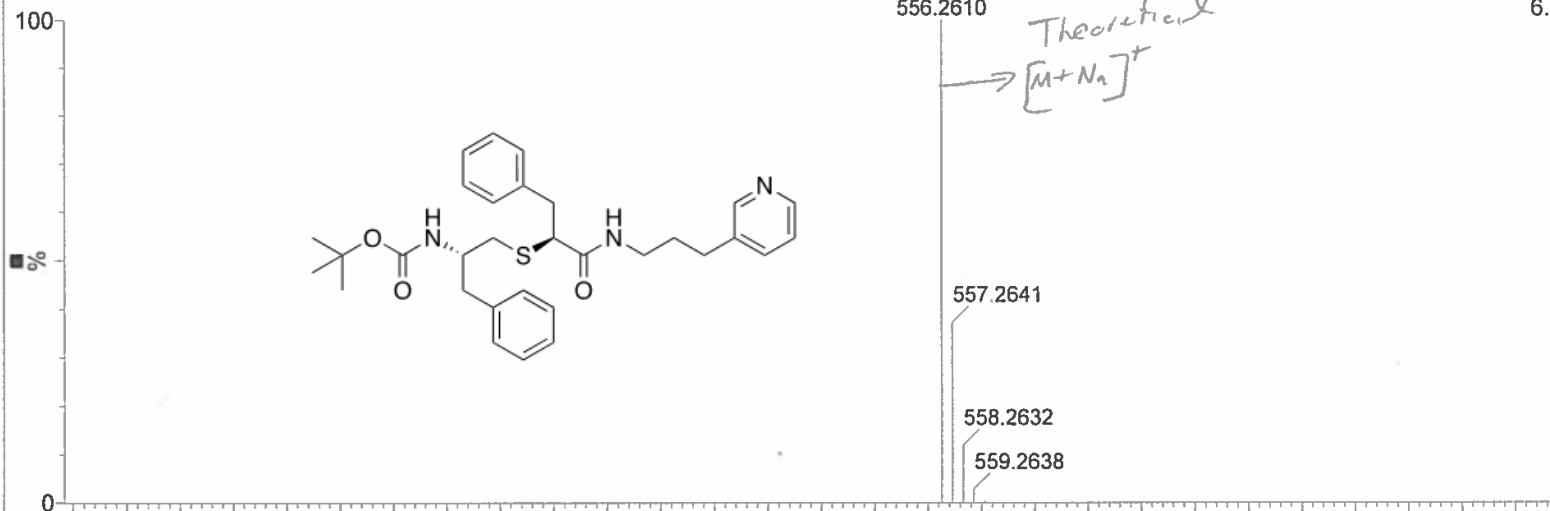


Acceptable = ± 3 mDa
Observed = -2.3 mDa
Observed $[M+Na]^+$



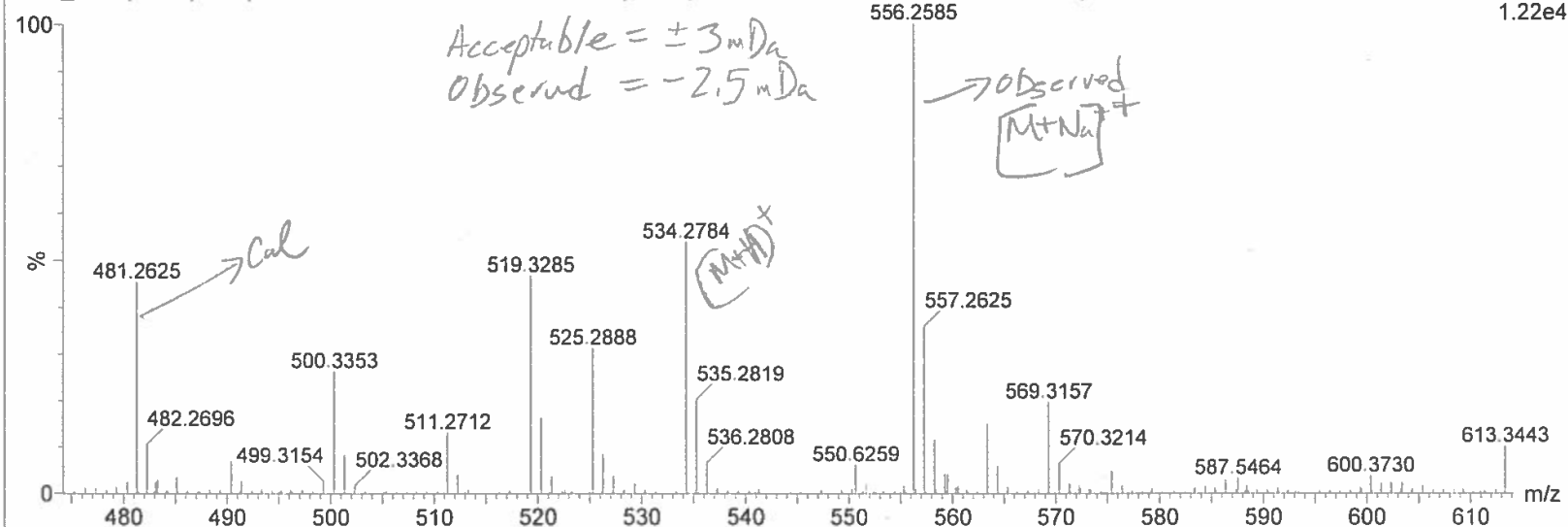
4111_a (0.019) Is (1.00,0.01) C₃₁H₃₉N₃O₃Na

TOF MS ES+
6.58e12



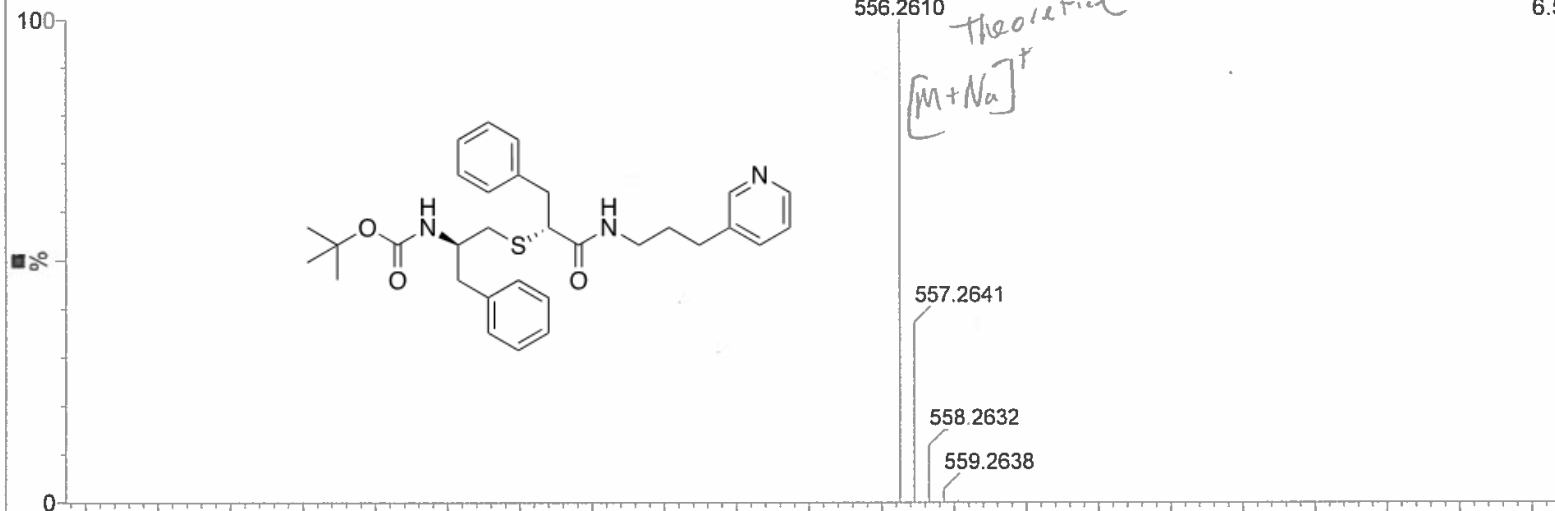
4111_a 18 (0.330) AM (Cen,5, 80.00, Ar,8000.0,481.26,0.70); Sm (SG, 2x3.00); Sb (1,40.00); Cm (18:23)

TOF MS ES+
1.22e4



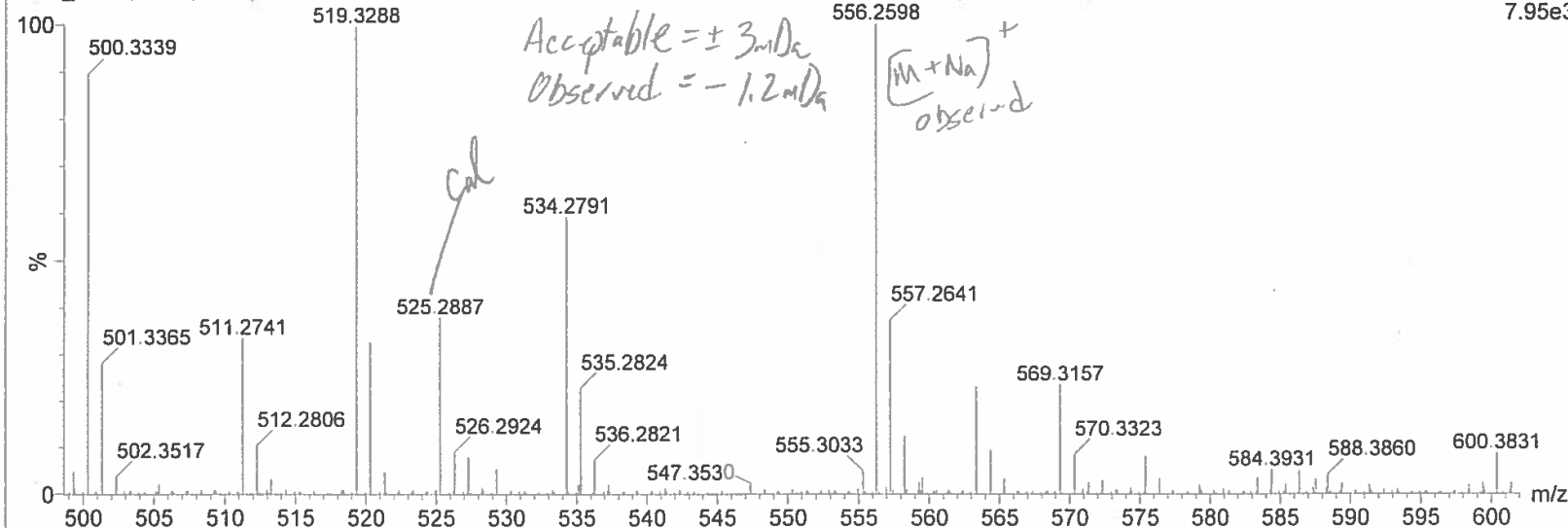
4115_a (0.019) Is (1.00,0.01) C₃₁H₃₉N₃O₃Na

TOF MS ES+
6.58e12



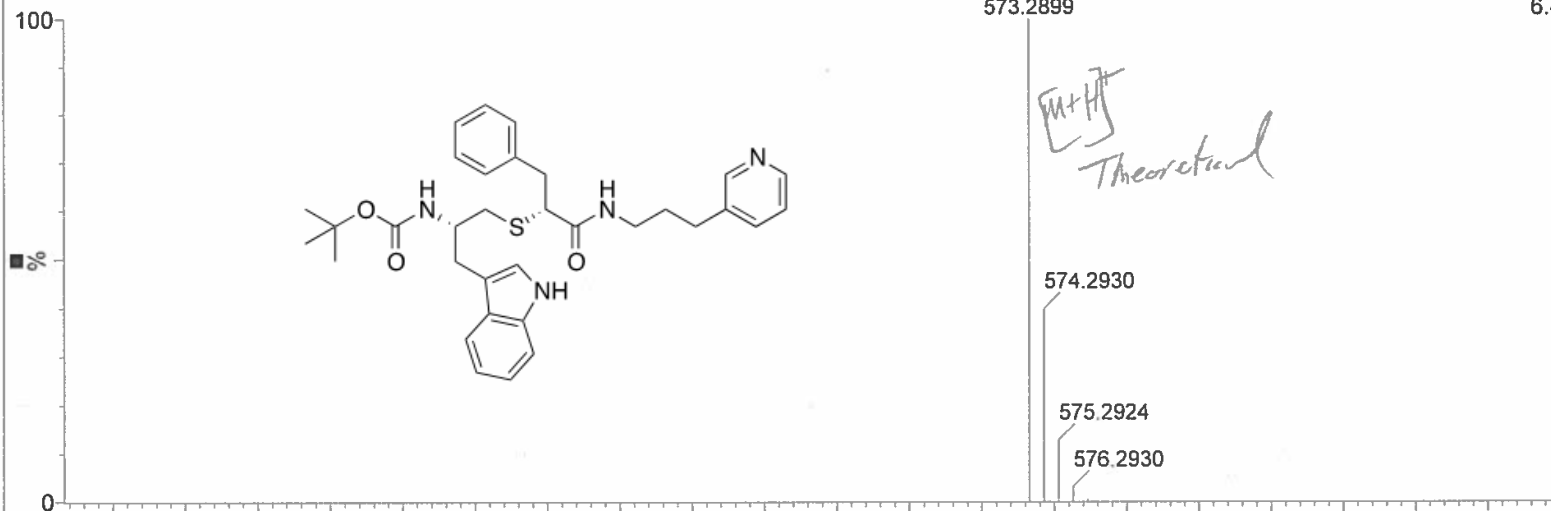
4115_a 29 (0.532) AM (Cen,5, 80.00, Ar,8000.0,525.29,0.70); Sm (SG, 2x3.00); Sb (1,40.00); Cm (29:39)

TOF MS ES+
7.95e3



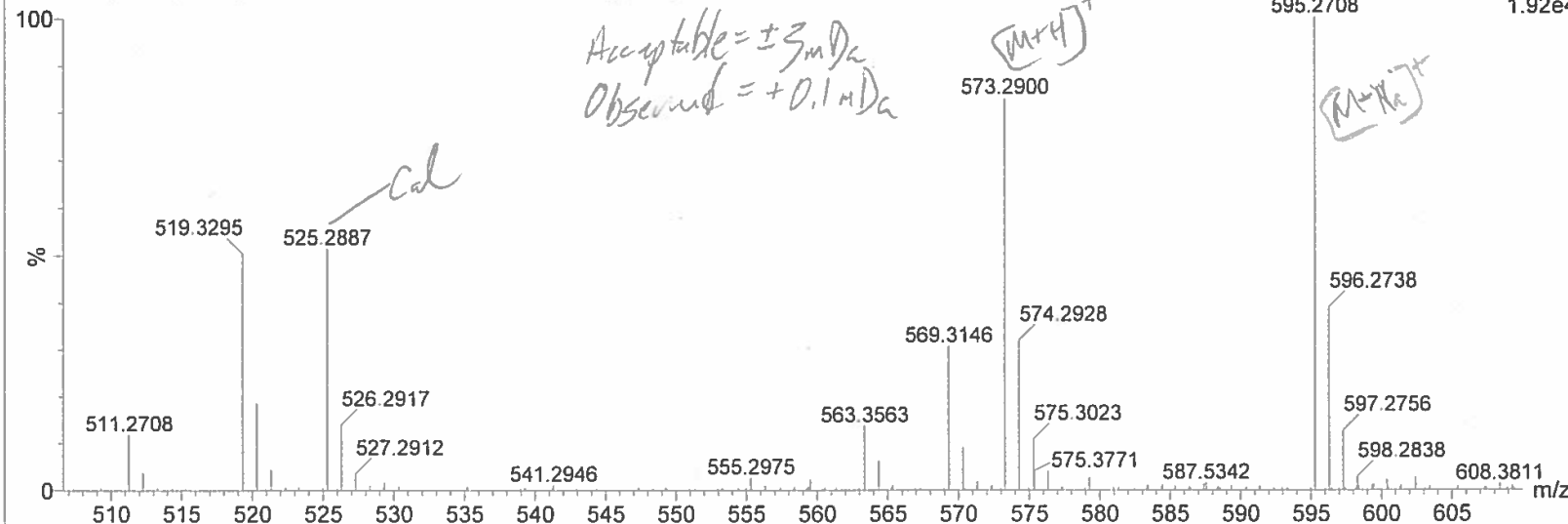
397_a (0.019) Is (1.00,0.01) C33H41N4O3S

TOF MS ES+
6.41e12



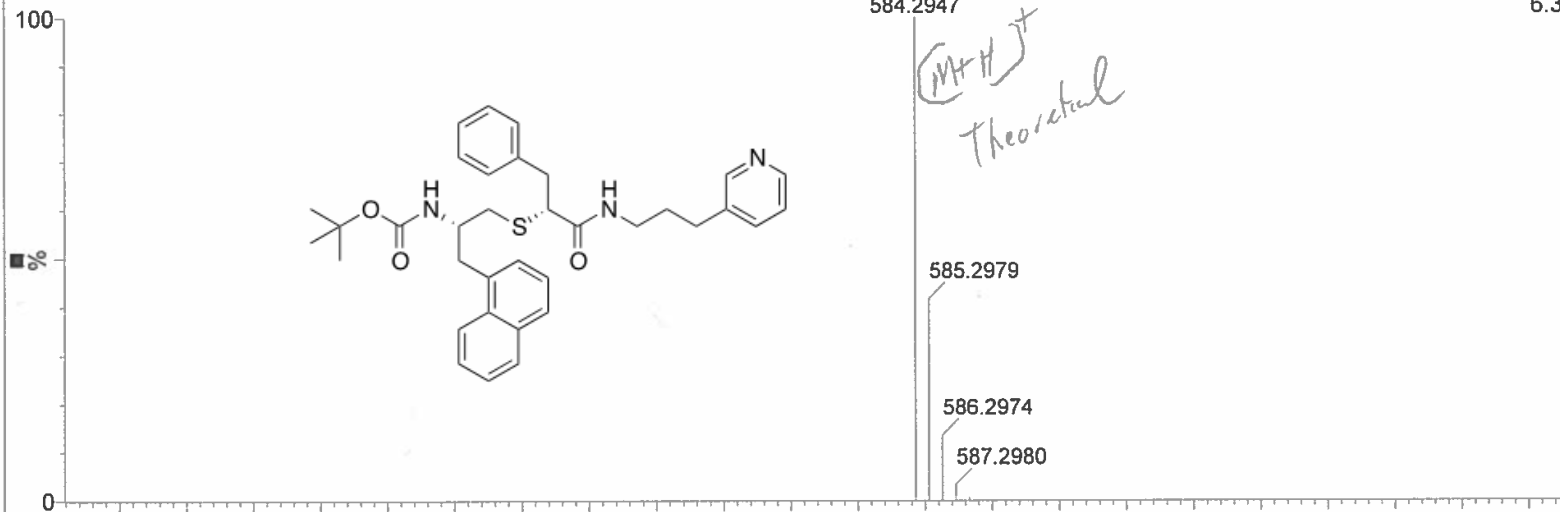
397_a 19 (0.349) AM (Cen,5, 80.00, Ar,8000.0,525.29,0.70); Sm (SG, 2x3.00); Sb (1,40.00); Cm (19:30)

TOF MS ES+
1.92e4



383_a (0.019) Is (1.00,0.01) C35H42N3O3S

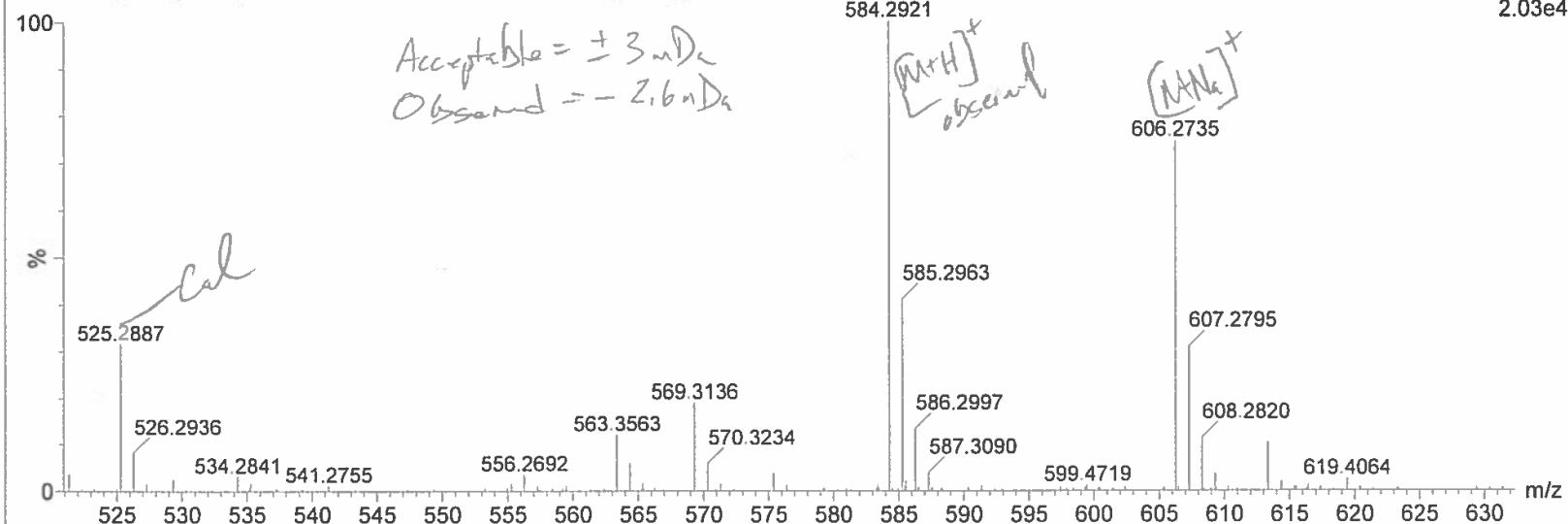
TOF MS ES+
6.30e12



$[M+H]^+$
Theoretical

383_a 22 (0.404) AM (Cen,5, 80.00, Ar,8000.0,525.29,0.70); Sm (SG, 2x3.00); Sb (1,40.00); Cm (22:34)

TOF MS ES+
2.03e4

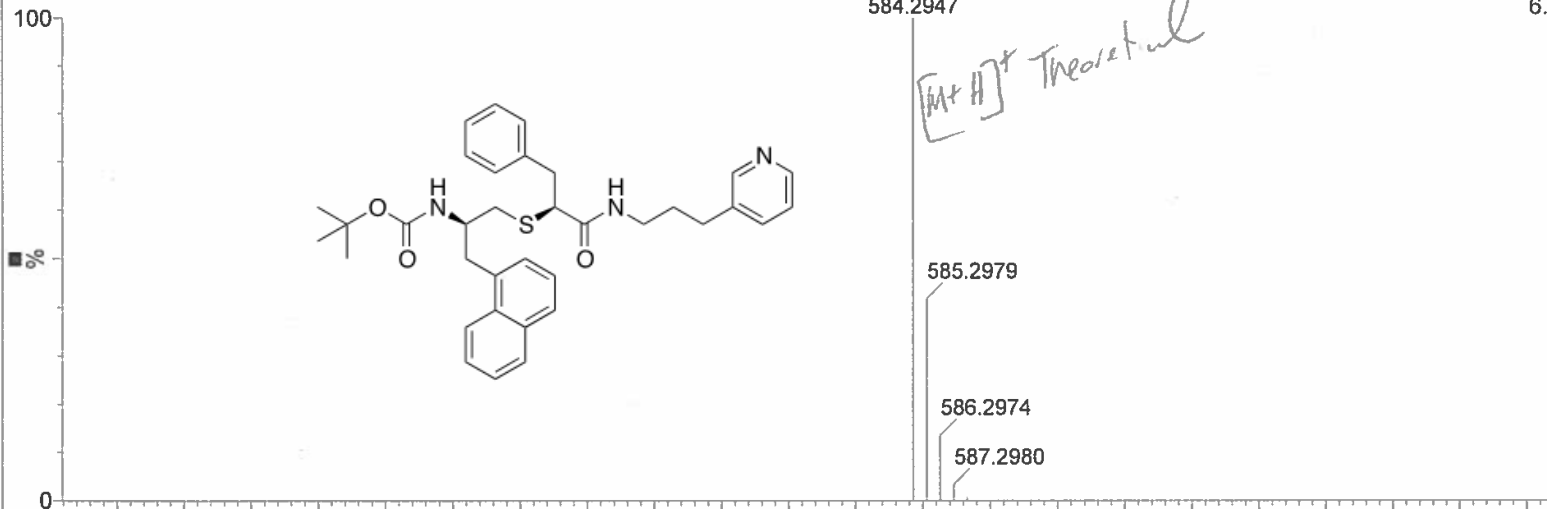


$[M+H]^+$
observed

$[M+Na]^+$

381_a (0.019) Is (1.00,0.01) C35H42N3O3S

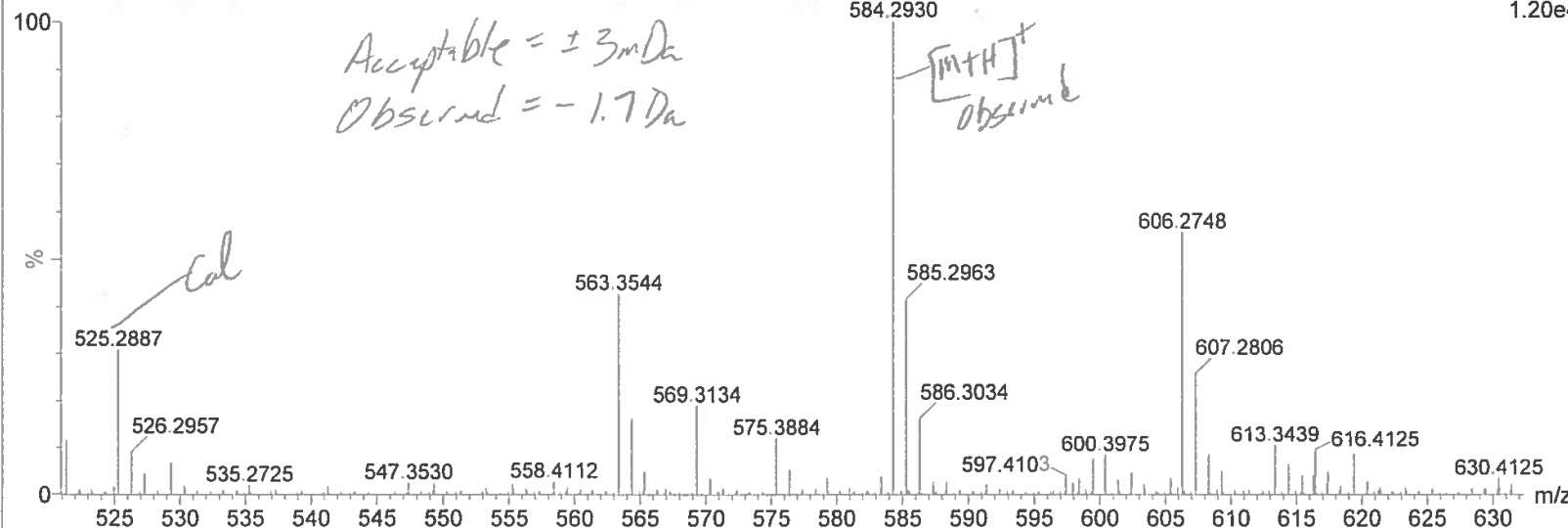
TOF MS ES+
6.30e12



$[M+H]^+$ theoretical

381_a 25 (0.458) AM (Cen,5, 80.00, Ar,8000.0,525.29,0.70); Sm (SG, 2x3.00); Sb (1,40.00); Cm (24:43)

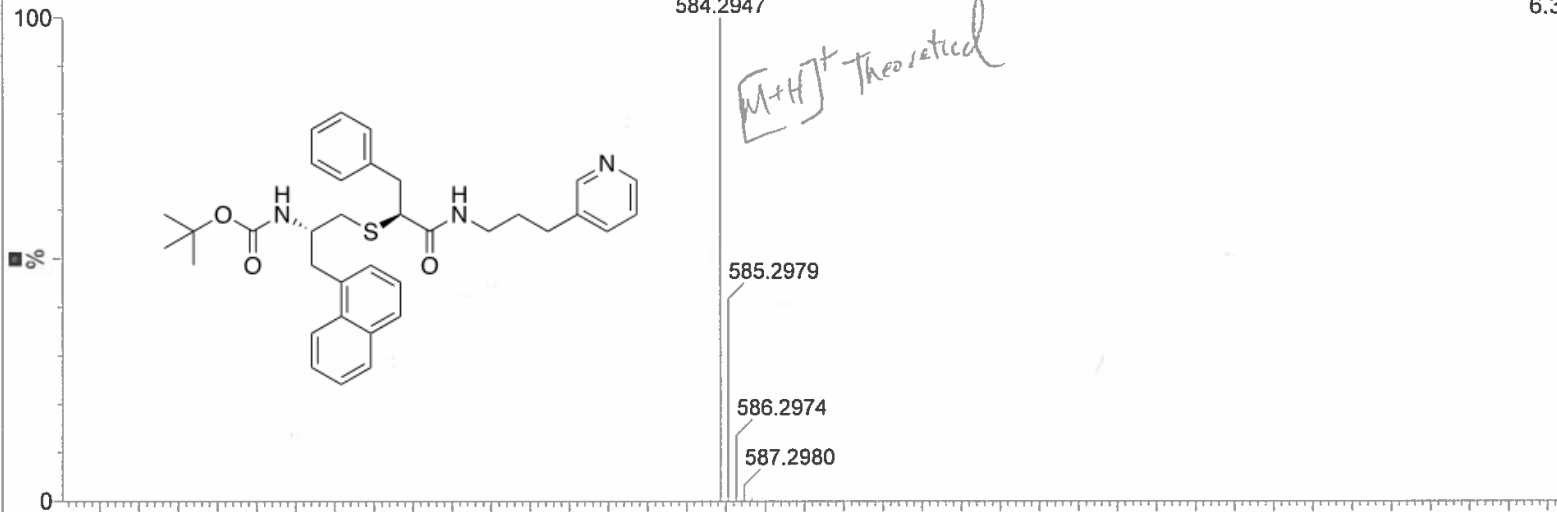
TOF MS ES+
1.20e4



$[M+H]^+$ observed

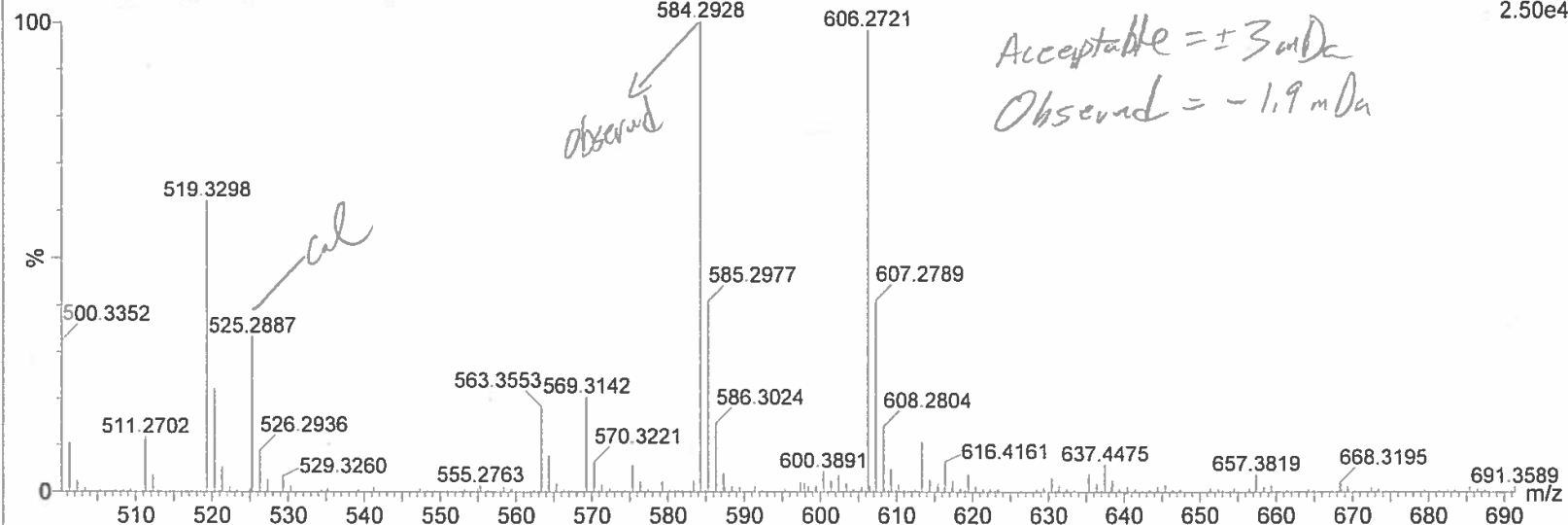
4129_a (0.019) Is (1.00,0.01) C35H42N3O3S

TOF MS ES+
6.30e12



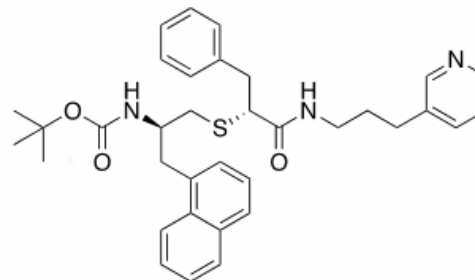
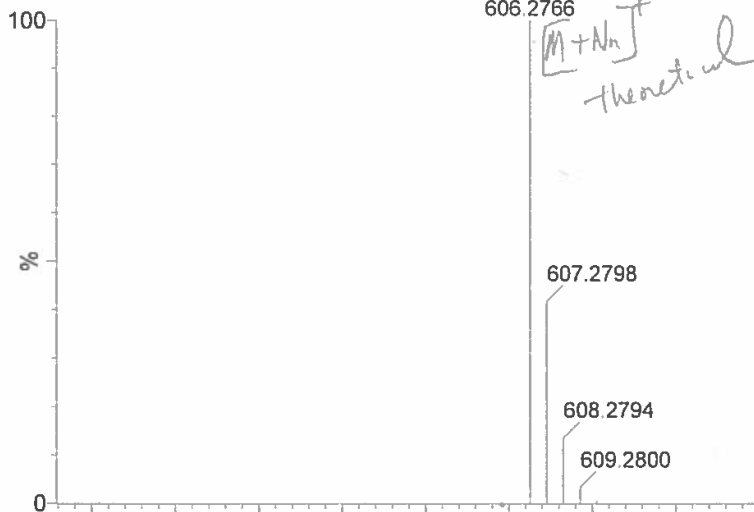
4129_a 20 (0.367) AM (Cen,5, 80.00, Ar,8000.0,525.29,0.70); Sm (SG, 2x3.00); Sb (1,40.00); Cm (20:39)

TOF MS ES+
2.50e4



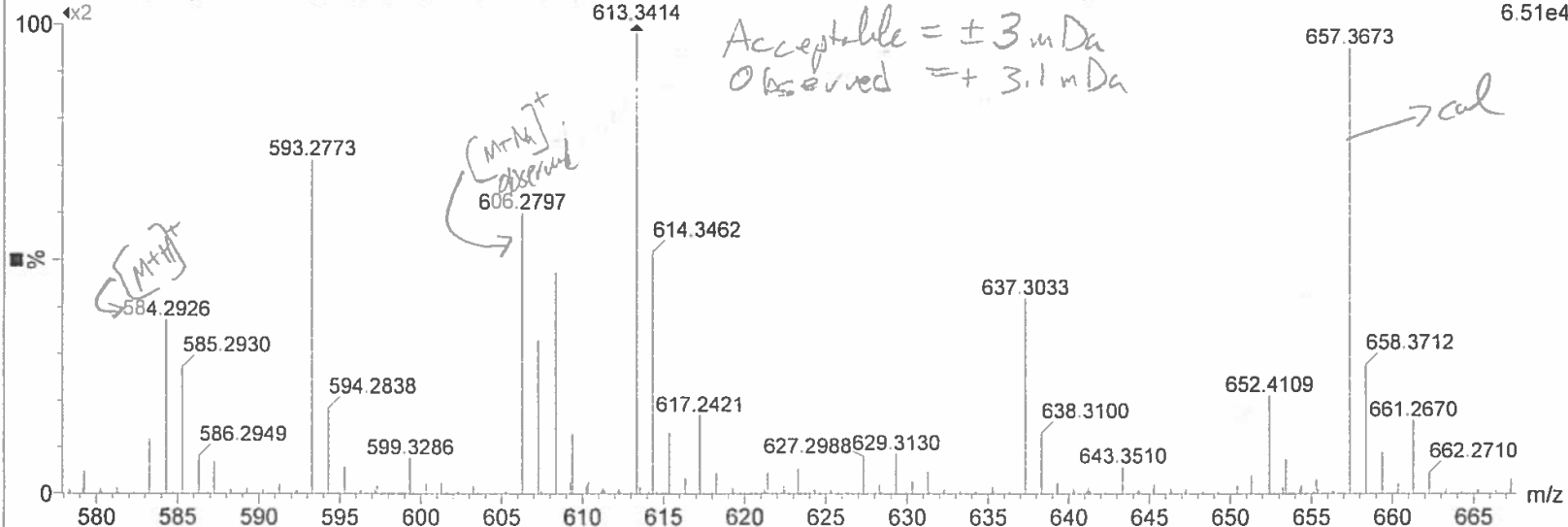
4145_a (0.018) Is (1.00,0.01) C₃₅H₄₁N₃O₃SNa

TOF MS ES+
6.30e12



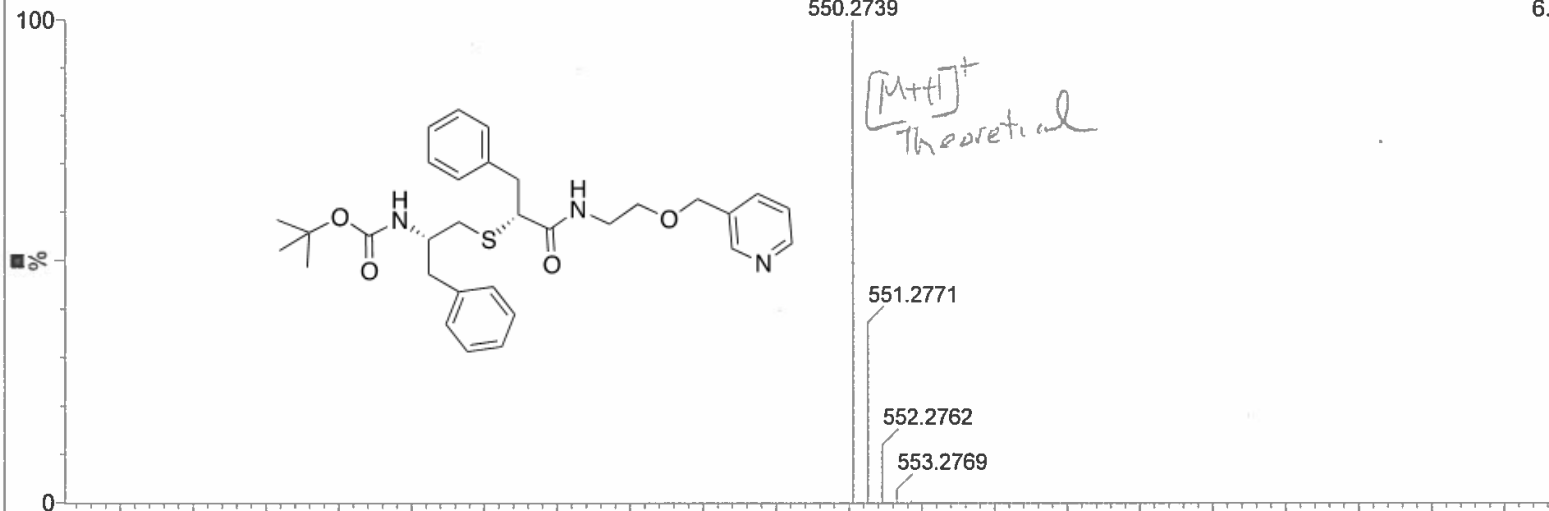
4145_a 17 (0.312) AM (Cen,5, 80.00, Ar,8000.0,657.37,0.70); Sm (SG, 2x3.00); Sb (1,40.00); Cm (11:22)

TOF MS ES+
6.51e4



4102_a (0.019) Is (1.00,0.01) C₃₁H₄₀N₃O₄S

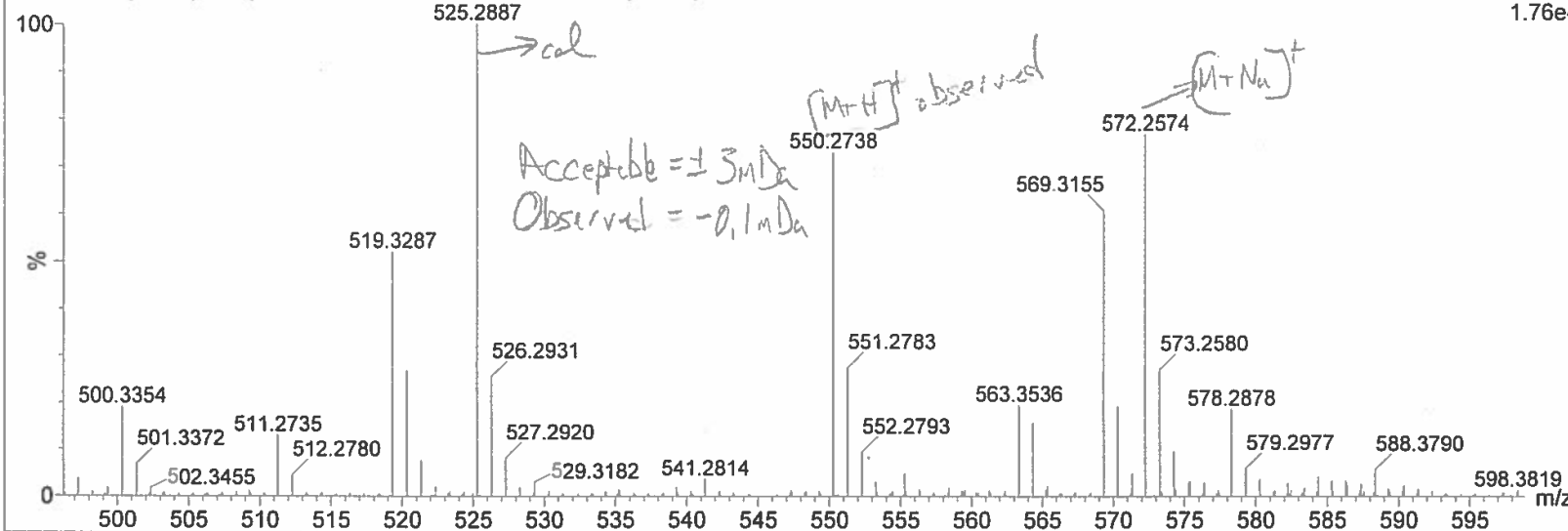
TOF MS ES+
6.57e12



$[M+H]^+$
Theoretical

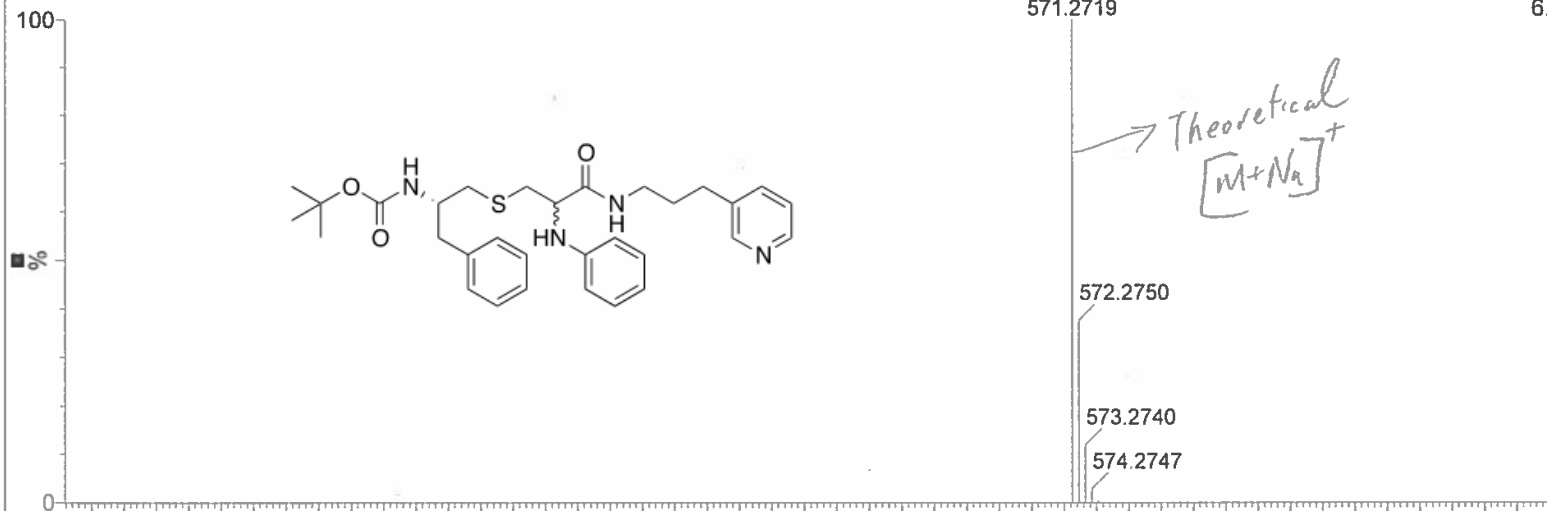
4102_a 17 (0.312) AM (Cen,5, 80.00, Ar,8000.0,525.29,0.70); Sm (SG, 2x3.00); Sb (1.40.00); Cm (11:26)

TOF MS ES+
1.76e4



390_b (0.019) Is (1.00,0.01) C₃₁H₄₀N₄O₃Na

TOF MS ES+
6.56e12



390_b 18 (0.330) AM (Cen,5, 80.00, Ar,8000.0,481.26,0.70); Sm (SG, 2x3.00); Sb (1,40.00); Cm (14:25)

TOF MS ES+
1.31e4

