

Figure S1. Expression profiling of the human hyaluronan synthase (*HAS*) gene family in fibroblast-like synoviocytes (mean + SE) at timepoint 1 = lag phase, timepoints 2–3 = proliferating cells, and timepoints 4–5 = high-density cells. Real-time quantitative PCR was used to determine the ratio of the mRNA expression of the three *HAS* genes relative to the control gene ribosomal protein lateral stalk subunit P0. Means with dissimilar letters indicate significant differences between the isoforms within a timepoint (Kruskal–Wallis ANOVA, $p < 0.05$).

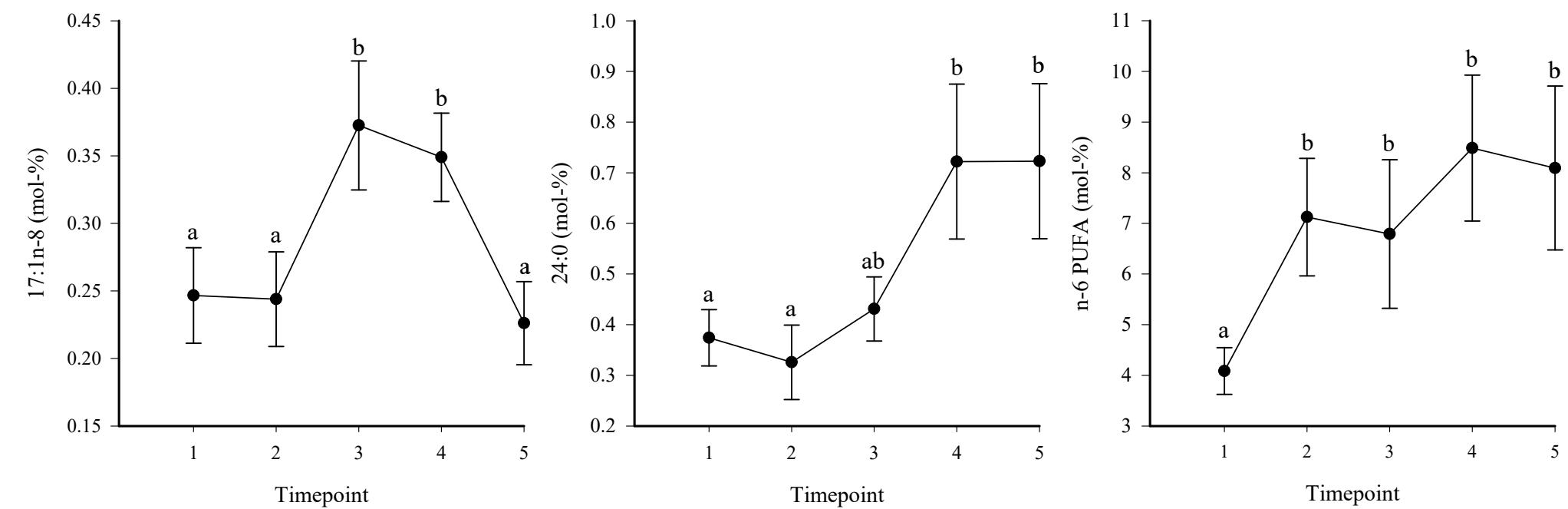


Figure S2. Temporal changes in the proportions of 17:1n-8, 24:0, and the sum of n-6 polyunsaturated fatty acids (PUFA) in fibroblast-like synoviocytes. Timepoint 1 = lag phase, timepoints 2–3 = proliferating cells, timepoints 4–5 = high-density cells. Dissimilar letters above means \pm SE indicate significant differences between timepoints (Kruskal-Wallis ANOVA, $p < 0.05$).

Table S1. Fatty acid composition of the media, cells, and ultracentrifuged fraction (mean \pm SE).

Fatty acid	Control medium	Conditioned medium	Fibroblast-like synoviocytes	Extracellular vesicles	<i>p</i>
14:0	4.718 \pm 0.168 ^a	4.678 \pm 0.094 ^a	5.258 \pm 0.166 ^b	5.618 \pm 0.103 ^b	<0.0005
14:1n-5	0.399 \pm 0.040 ^b	0.293 \pm 0.017 ^a	0.434 \pm 0.028 ^b	0.444 \pm 0.024 ^b	<0.0005
15:0 <i>i</i>	0.262 \pm 0.023 ^{bc}	0.220 \pm 0.013 ^b	0.291 \pm 0.018 ^c	0.153 \pm 0.010 ^a	<0.0005
15:0 <i>ai</i>	0.378 \pm 0.034 ^a	0.364 \pm 0.018 ^a	0.563 \pm 0.033 ^b	0.428 \pm 0.023 ^a	<0.0005
15:0	0.288 \pm 0.031 ^b	0.268 \pm 0.013 ^b	0.284 \pm 0.018 ^b	0.205 \pm 0.014 ^a	<0.0005
DMA 16:0	0.651 \pm 0.071 ^a	0.662 \pm 0.055 ^a	1.190 \pm 0.072 ^b	0.779 \pm 0.055 ^a	<0.0005
16:0	30.563 \pm 0.618 ^b	29.849 \pm 0.466 ^b	25.963 \pm 0.446 ^a	30.758 \pm 0.298 ^b	<0.0005
16:1n-9	0.311 \pm 0.035 ^a	0.466 \pm 0.031 ^b	0.612 \pm 0.033 ^c	0.353 \pm 0.022 ^a	<0.0005
16:1n-7	1.309 \pm 0.059	1.392 \pm 0.046	1.479 \pm 0.046	1.413 \pm 0.043	0.125
16:1n-5	0.248 \pm 0.032 ^a	0.349 \pm 0.026 ^{ab}	0.501 \pm 0.035 ^c	0.388 \pm 0.032 ^b	0.001
17:0 <i>i</i>	1.065 \pm 0.060 ^a	1.005 \pm 0.057 ^a	1.306 \pm 0.065 ^b	1.096 \pm 0.042 ^a	0.004
17:0 <i>ai</i>	0.360 \pm 0.048	0.281 \pm 0.014	0.321 \pm 0.017	0.307 \pm 0.016	0.201
17:0	0.923 \pm 0.025 ^c	0.850 \pm 0.022 ^{bc}	0.817 \pm 0.032 ^b	0.735 \pm 0.020 ^a	<0.0005
17:1n-8	0.339 \pm 0.042 ^b	0.309 \pm 0.019 ^b	0.289 \pm 0.018 ^{ab}	0.240 \pm 0.015 ^a	0.025
DMA 18:0	0.558 \pm 0.058 ^a	0.521 \pm 0.035 ^a	1.009 \pm 0.057 ^b	0.612 \pm 0.030 ^a	<0.0005
18:0	19.297 \pm 0.615 ^{ab}	20.478 \pm 0.326 ^b	18.605 \pm 0.233 ^a	19.886 \pm 0.144 ^b	<0.0005
18:1n-9	24.562 \pm 0.252 ^{bc}	23.703 \pm 0.289 ^b	22.264 \pm 0.400 ^a	24.640 \pm 0.223 ^c	<0.0005
18:1n-7	3.578 \pm 0.203 ^b	3.443 \pm 0.111 ^b	3.539 \pm 0.076 ^b	2.850 \pm 0.069 ^a	<0.0005
18:1n-5	1.428 \pm 0.152 ^{ab}	1.433 \pm 0.071 ^a	1.740 \pm 0.082 ^b	1.718 \pm 0.076 ^b	0.009
18:2n-6	1.875 \pm 0.069 ^c	1.693 \pm 0.040 ^b	1.623 \pm 0.032 ^{ab}	1.578 \pm 0.026 ^a	0.001
18:3n-6	0.358 \pm 0.043 ^{ab}	0.416 \pm 0.049 ^a	0.556 \pm 0.036 ^c	0.604 \pm 0.079 ^{bc}	0.002
18:3n-3	0.675 \pm 0.056 ^a	0.711 \pm 0.053 ^a	0.989 \pm 0.054 ^b	1.063 \pm 0.060 ^b	<0.0005
20:0	0.374 \pm 0.045	0.448 \pm 0.029	0.474 \pm 0.029	0.385 \pm 0.017	0.178
20:1n-9	0.196 \pm 0.017	0.235 \pm 0.021	0.270 \pm 0.018	0.219 \pm 0.012	0.100
20:3n-6	0.675 \pm 0.117 ^{ab}	0.742 \pm 0.070 ^b	1.184 \pm 0.072 ^c	0.453 \pm 0.033 ^a	<0.0005
20:4n-6	0.872 \pm 0.200 ^b	1.086 \pm 0.107 ^b	3.294 \pm 0.543 ^c	0.355 \pm 0.023 ^a	<0.0005
20:5n-3	0.394 \pm 0.064 ^b	0.396 \pm 0.050 ^b	0.435 \pm 0.040 ^b	0.227 \pm 0.023 ^a	0.001
22:0	0.762 \pm 0.075	0.713 \pm 0.041	0.781 \pm 0.049	0.802 \pm 0.038	0.767
22:1n-11	0.061 \pm 0.011 ^{bc}	0.045 \pm 0.005 ^{ab}	0.068 \pm 0.005 ^c	0.031 \pm 0.003 ^a	<0.0005
22:1n-9	0.151 \pm 0.033 ^{ab}	0.125 \pm 0.016 ^a	0.194 \pm 0.018 ^b	0.098 \pm 0.007 ^a	<0.0005
22:1n-7	0.042 \pm 0.010 ^a	0.046 \pm 0.007 ^a	0.053 \pm 0.004 ^b	0.031 \pm 0.003 ^a	0.001
22:4n-6	0.258 \pm 0.079 ^{ab}	0.395 \pm 0.076 ^b	0.323 \pm 0.049 ^b	0.151 \pm 0.023 ^a	<0.0005
22:5n-3	0.992 \pm 0.239 ^a	1.156 \pm 0.119 ^a	1.583 \pm 0.114 ^b	0.901 \pm 0.059 ^a	<0.0005
22:6n-3	0.611 \pm 0.186 ^b	0.795 \pm 0.117 ^b	1.055 \pm 0.093 ^c	0.268 \pm 0.017 ^a	<0.0005
24:0	0.325 \pm 0.040 ^{bc}	0.357 \pm 0.027 ^b	0.518 \pm 0.054 ^c	0.172 \pm 0.019 ^a	<0.0005
24:1n-9	0.141 \pm 0.051 ^{bc}	0.079 \pm 0.014 ^b	0.137 \pm 0.016 ^c	0.037 \pm 0.003 ^a	<0.0005
SFA	59.314 \pm 1.028 ^b	59.510 \pm 0.657 ^b	55.183 \pm 0.499 ^a	60.547 \pm 0.265 ^b	<0.0005
MUFA	32.767 \pm 0.536	31.918 \pm 0.278	31.578 \pm 0.393	32.463 \pm 0.159	0.219
PUFA	6.710 \pm 0.823 ^{ab}	7.390 \pm 0.546 ^b	11.041 \pm 0.749 ^c	5.600 \pm 0.204 ^a	<0.0005
n-6 PUFA	4.037 \pm 0.417 ^{ab}	4.332 \pm 0.279 ^b	6.981 \pm 0.609 ^c	3.140 \pm 0.113 ^a	<0.0005
n-3 PUFA	2.672 \pm 0.442 ^a	3.058 \pm 0.284 ^a	4.061 \pm 0.219 ^b	2.460 \pm 0.127 ^a	<0.0005
n-3/n-6 PUFA	0.647 \pm 0.056 ^{ab}	0.670 \pm 0.030 ^a	0.663 \pm 0.038 ^a	0.799 \pm 0.043 ^b	0.021

UFA/SFA	0.671 ± 0.031^a	0.669 ± 0.018^a	0.778 ± 0.016^b	0.630 ± 0.006^a	<0.0005
DMA	1.209 ± 0.097^a	1.182 ± 0.082^a	2.198 ± 0.102^b	1.391 ± 0.079^a	<0.0005
DBI	0.568 ± 0.037^{ab}	0.594 ± 0.023^b	0.739 ± 0.028^c	0.513 ± 0.007^a	<0.0005
TACL	17.068 ± 0.037^{bc}	17.112 ± 0.018^c	17.059 ± 0.032^b	16.938 ± 0.011^a	<0.0005

i = iso, *ai* = anteiso, DMA = dimethyl acetal, *i.e.*, plasmalogen alkenyl chain derivative, SFA = saturated fatty acid, MUFA = monounsaturated fatty acid, PUFA = polyunsaturated fatty acid, UFA = unsaturated fatty acid (MUFA + PUFA), DBI = double bond index, TACL = total average chain length

Dissimilar superscript letters indicate significant differences between sample types within a row (Kruskal–Wallis ANOVA, $p < 0.05$)

Table S2. Primer sequences of the studied genes.

Gene		Sequence (5'-3')	Length
<i>RPLP0</i>	FOR	5' AGATGCAGCAGATCCGCAT 3'	319 bp
	REV	5' GTGGTGATACTAAAGCTG 3'	
<i>HASI</i>	FOR	5' CAAGATTCTCAGTCTGGAC 3'	124 bp
	REV	5' TAAGAACGAGGAGAAAGCAG 3'	
<i>HAS2</i>	FOR	5' CAGAATCCAAACAGACAGTTC 3'	187 bp
	REV	5' TAAGGTGTTGTGTGACTG 3'	
<i>HAS3</i>	FOR	5' CTTAAGGGTTGCTGCTTGC 3'	194 bp
	REV	5' GTTCGTGGGAGATGAAGGAA 3'	
<i>CD44</i>	FOR	5' CATCTACCCCAGCAACCCTA 3'	153 bp
	REV	5' CTGTCTGTCCTGTCGGTGAT 3'	
<i>IL-1β</i>	FOR	5' TGGAGCAACAAAGTGGTGT 3'	157 bp
	REV	5' TTGGGATCTACACTCTCCAGC 3'	
<i>IL-6</i>	FOR	5' TGCAATAACCACCCCTGACC 3'	163 bp
	REV	5' GTGCCCATGCTACATTGCC 3'	
<i>TNF-α</i>	FOR	5' CCCAGGGACCTCTCTAATCA 3'	80 bp
	REV	5' GCTACAGGCTTGTCACTCGG 3'	
<i>COX-2</i>	FOR	5' ATCTACGGTTGCTGTGGGG 3'	156 bp
	REV	5' GCCTGCTTGTCTGGAACAAAC 3'	
<i>PPAR-α</i>	FOR	5' GCTGTCACCACAGTAGCTTG 3'	197 bp
	REV	5' TTCCAGAACTATCCTCGCCG 3'	
<i>PPAR-γ</i>	FOR	5' TTCTCAAACGAGAGTCAGCCT 3'	126 bp
	REV	5' CAAAGGAGTGGAGTGGTCT 3'	

RPLP0 = ribosomal protein lateral stalk subunit P0, *HASI-3* = hyaluronan synthase 1–3, *CD44* = cluster of differentiation 44, *IL-1 β /6* = interleukin-1 β /6, *TNF- α* = tumor necrosis factor α , *COX-2* = cyclooxygenase 2, *PPAR- α/γ* = peroxisome proliferator-activated receptor α/γ