

Supplemental Information

Supplement Table S1. List of antibodies and working dilutions used in this study.

Antibody name	Source	Cat.no°	Working Dilution WB	Working Dilution ICC	Working Dilution IF
<i><u>Primary Antibodies</u></i>					
SM-calponin	Abcam	Ab78491	1:400	1:250	1:100
ACTB	SantaCruz	sc47778	1:500	-	-
ckit/CD117	Abcam	ab32363	1:200	1:200	4.72µg/ml
PDGFR- α	SantaCruz	sc398206	1:400	1:100	1:100
PDGFR- β	Abcam	ab69506	1:600	1:300	1:300
KLF-4	Abcam	ab75486	1:50	1:50	10µg/ml
VEGF-A	Abcam	ab1316	1:100	1:100	1:100
α SMC	Abcam	ab5694	1:500	1:500	1:500
Vimentin	SantaCruz	sc5565	1:200	1:200	1µg/ml
CD133 (EPR20980-104)	Abcam	ab216323	1:600	1:1000	1:1000
CD63	Abcam	ab68418	1:750	-	-
Integrin β -1 (CD29)	Abcam	Ab183666	1:400	1:200	-
HSP90	CellSignaling	7874S	1:800	-	-
CD34 (B6)	SantaCruz	sc74499	1:400	1:50	4µg/ml
TSG101	SantaCruz	sc7974	1:500	-	-
<i><u>Secondary Antibodies</u></i>					
AF488GAR	molecular probes	A11034	-	1:1000	1:500
AF546GAM	molecular probes	A11030	-	1:1000	1:1000
AF488GAM	molecular probes	A11029	-	1:1000	1:1000
AF546GAR	molecular probes	A11035	-	1:1000	1:1000
AF488DAG	molecular probes	A11055	-	1:500	1:500
HRP-anti-mouse IgG	CellSignaling	7076S	1:2000	-	-
HRP-anti-rabbit IgG	CellSignaling	7074S	1:3000	-	-
<i><u>Isotype-control</u></i>					
	BDPharmigen	550878	-	4 µg/ml	4 µg/ml
purified mouse IgG	Abcam	Ab27478	-	1.89 µg/ml	1.89 µg/ml
purified rabbit IgG					

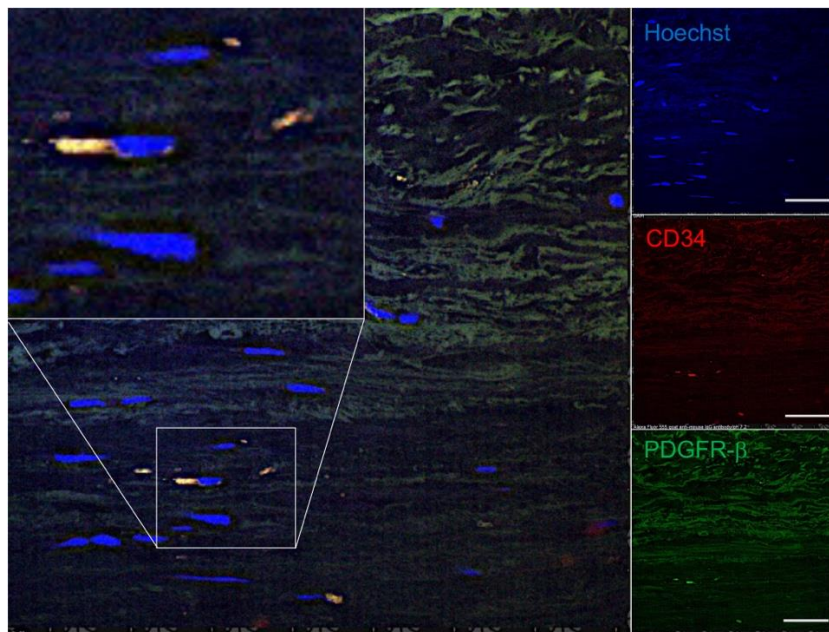
BD Pharmigen™ (BD Biosciences), San Jose, CA; Santa Cruz Biotechnologies, Inc., TX, USA; Invitrogen Molecular Probes, ThermoFisher Scientific, Massachusetts, USA; Novus Biologics, LLC, CO, USA; Dianova GmbH, Hamburg, Germany.

Supplement Table S2. Primer sequences for qPCR

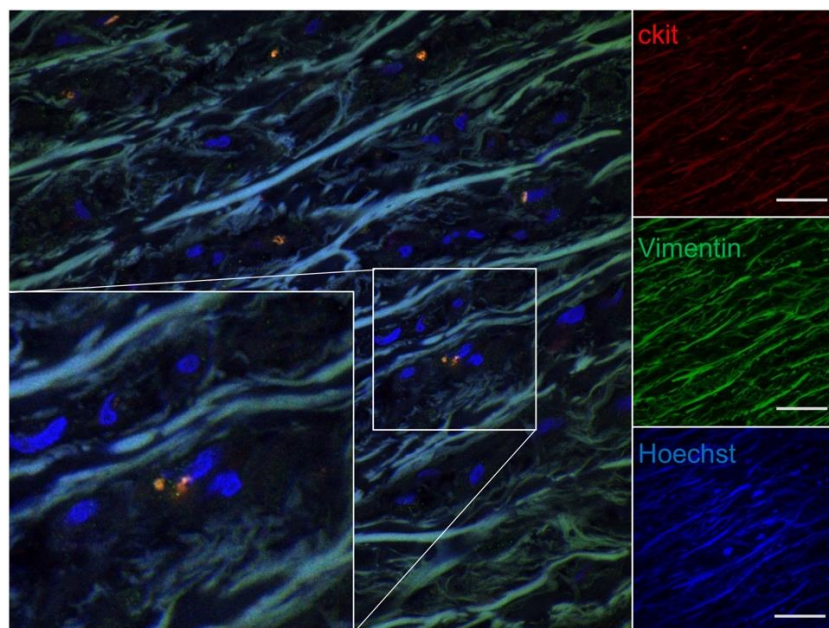
Target	Forward Primer	Reverse Primer
<i>miRNA primer sequences</i>		
<i>RNU6</i>	CGCTTCGGCAGCACATATAC	AGGGGCCATGCTAATCTTCT
<i>SNORD44</i>	TGATGATAAGCAAATGCTGACTG	GAGCTAATTAAGACCTTCATGTTTCAAG
<i>hsa-mir-21-5p</i>	GCAGTAGCTTATCAGACTGATG	GGTCCAGTTTTTTTTTTTTTTTCAAC
<i>hsa-mir-24-3p</i>	GATCCTGGCTCAGTTCAGCAGGAACA GC	TCGAGCTGTTCTGCTGAACTGAGCCAG
<i>hsa-mir-143-3p</i>	GCAGTGCTGCATCTCTG	GAACATGTCTGCGTATCTC
<i>hsa-mir-145-5p</i>	GTCCAGTTTTCCAGGA	GAACATGTCTGCGTATCTC
<i>hsa-mir-146a</i>	GAGAACTGAATTCCATGG	GAACATGTCTGCGTATCTC
<i>hsa-mir-221-3p</i>	GCCGAGAGCTACATTGTCTG	GTCGTATCCAGTGCAGGG
<i>hsa-mir-221-5p</i>	TCCGCGCCCTTGCCAGACC	GTGCCTGGTGCTCTCTTACC
<i>hsa-mir-222-5p</i>	GGGCTCAGTAGCCAGTGTA	CAGTGCGTGTCGTGGAGT
<i>mRNA primer sequences</i>		
<i>VIM</i>	AGGCAAAGCAGGAGTCCACTGA	ATCTGGCGTTCCAGGGACTCAT
<i>KLF4</i>	CGACGCGTGCTCCCATCTT	GGCAGTGTTGGTTCATATCCA
<i>KIT</i>	TCATCGAGTGTGATGGGAAA	GGTGACTTGTTCAGGCAACA
<i>MYH11</i>	GTCCAGGAGATGAGGCAGAAAC	GTCTGCGTTCTCTTTCTCCAGC
<i>COL1A1</i>	GATTCCCTGGACCTAAAGGTGC	AGCCTCTCCATCTTTGCCAGCA
<i>ACTA</i>	CTATGCCTCTGGACGCACAAC	CAGATCCAGACGCATGATGGCA
<i>PDGFRA</i>	GACTTTCGCCAAAGTGGAGGAG	AGCCACCGTGAGTTCAGAACGC
<i>PDGFRB</i>	AGGACAAACCGTACCTTGGGTGACT	CAGTTCTGACACGTACCGGGTCTC
<i>CNN1</i>	CTGGCTGCAGCTTATTGATG	CTGAGAGAGTGGATCGAGGG
<i>INTGR</i>	AGAAGCTCAAGCCAGAGG	GCATCTGTGGAACACACC
<i>RPLPO</i> (housekeeping gene)	AGCCCAGAACACTGGTCTC	ACTCAGGATTTCAATGGTGCC
<i>GAPDH</i> (housekeeping gene)	TGCACCACCAACTGCTTAGC	GGCATGGACTGTGGTCATGAG

Supplemental Figures

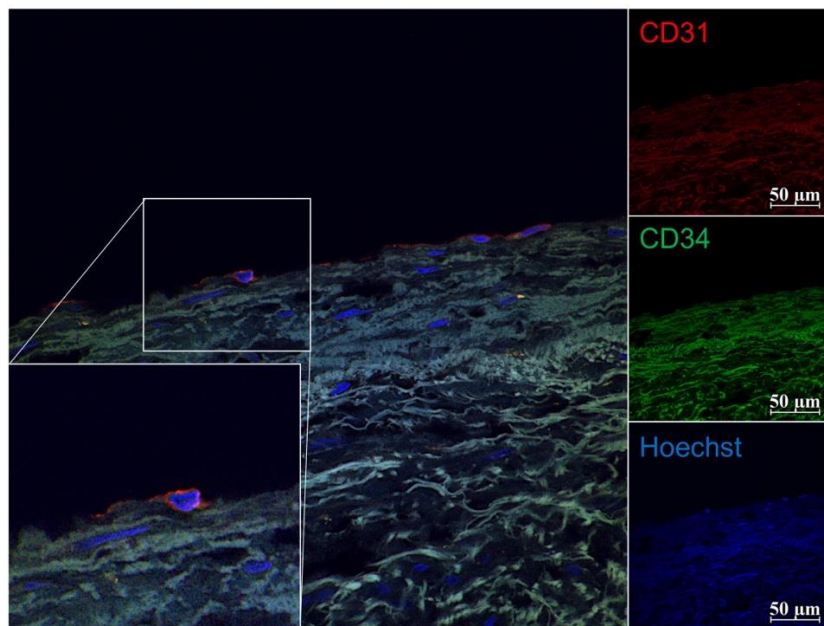
A



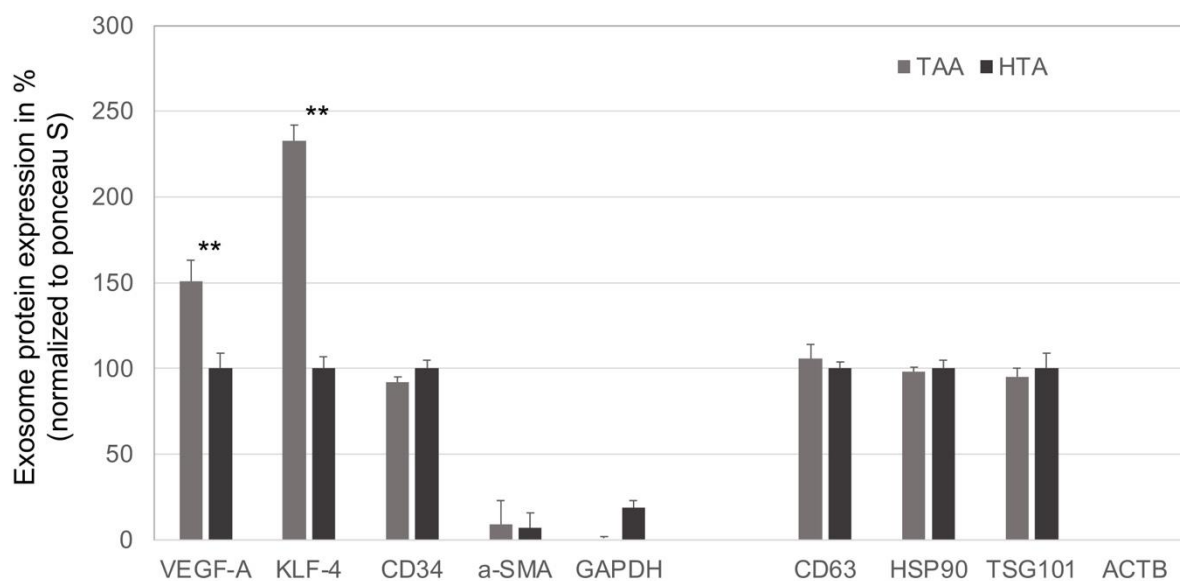
B



Supplemental Figure S1. CD34, ckit, vimentin and PDGFR-β staining confirmed aortic TC in TAA. Double staining of well-known aortic TC markers in (A) CD34 (red) and PDGFR-β (green), and (B) ckit (red) and vimentin (green), were done to identified TC specificity. Scalebar, 50 μm.

**Supplemental Figure S2. Validation of TC specificity by CD34/CD31 double staining.**

CD34 negative and CD31 positive immunostaining were also described for vasa vasorum or endothelial cell population and should be distinct from aortic TCs. The lack of endothelial marker CD31 were performed to validate aortic TC specificity. Red, CD31; green, CD34; blue, nuclei (Hoechst). Scalebar, 50 µm.



Supplemental Figure S3. Exosome-specific soluble factors VEGF and KLF-4 were increased in exosomes derived from TAA-TCs (gray bars) compared to HTA-TCs (black bars) samples. Soluble factors VEGF-A, KLF-4, CD34 and α -SMA, as well as surface proteins, CD63, HSP90 and TSG101, were analyzed by Western Blot (Figure 2H-I). ACTB and GAPDH were used as loading control. S-Figure 1 shows statistical calculation of protein expression after normalization to ponceau S signals. **, $p < 0.01$. Data are mean \pm SD of three independent experiments.