

Supplementary Materials

Employing engineered enolase promoter for efficient expression of *Thermomyces lanuginosus* lipase in *Yarrowia lipolytica* via a self-excisable vector

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Supplementary Tables

Table S1. the extracellular protein concentration of recombinant strains.

Strains	the extracellular protein concentration (g/L)	Annotation
Polf	0.129 \pm 0.003	-
Polf/tll	0.307 \pm 0.018	-
Polf/2tll	0.380 \pm 0.012	-
Polf/3tll	0.438 \pm 0.025	Before fermentation conditions were optimized
Polf/4tll	0.422 \pm 0.010	-
Polf/3tll	0.530 \pm 0.024	After optimization of fermentation conditions
Polf/3tll-hp4e ₁₀₀	0.523 \pm 0.016	-
Polf/3tll-hac1	0.620 \pm 0.038	-
Polf/3tll-kar2	0.560 \pm 0.020	-
Polf/3tll-pdi	0.572 \pm 0.024	-
Polf/3tll-hac1-kar2	0.720 \pm 0.027	-
Polf/3tll-hac1-pdi	0.724 \pm 0.016	-
Polf/3tll-hac1-pdi-kar2	0.779 \pm 0.024	-

Table S2. Strains and plasmids used in this study.

Plasmids/strains	Description	Reference or source
Plasmids		
hp4d-rml	pINA 1296 derivative, harboring <i>rml</i> gene under the control of hp4d promoter	[18]
Cre-Y3	harboring <i>cre</i> expression cassette under the control of pPOX2 promoter	[42]
Cre-axp1	harboring <i>rol</i> gene expression cassette	[42]
T-egfp	harboring <i>egfp</i> gene expression cassette	Our laboratory
Mlu-vgb	harboring <i>vgb</i> gene expression cassette	Our laboratory
pMD19-act1	harboring <i>act1</i> gene expression cassette	Our laboratory
hp4d-egfp	hp4d-rml derivative, harboring <i>egfp</i> gene under the control of hp4d promoter	This study
hp4d-tll	hp4d-rml derivative, harboring <i>tll</i> gene under the control of hp4d promoter	This study
pENOn-egfp (n = 75, 100, 125, 150, 175, 200, 300, 400, 700, 1000, and 1300 bp)	hp4d-egfp derivative, harboring <i>egfp</i> gene under the control of different length of enolase promoter	This study
pENOn-tll (n = 100, 125, 200, 400, 700, 1000, and 1300 bp)	pENOn-egfp derivative, harboring <i>tll</i> gene under the control of different length of enolase promoter	This study
hp4e _n -tll (n = 100, 125, 200, 400, 700, 1000, and 1300 bp)	hp4d-tll derivative, harboring <i>tll</i> gene under the control of different length of enolase hybrid promoter	This study
hp4e ₁₀₀ -tll (x = 8, 12, 16, 20, 24, 28, and 32)	hp4e ₁₀₀ -tll derivative, harboring <i>tll</i> gene under the control of enolase hybrid promoter containing different copies of UAS1B	This study
pUAXp7166-hp16e ₁₀₀ -tll	Cre-Y3 and hp16e ₁₀₀ -tll derivative, harboring upAxp homologous fragment, a self-excising vector for <i>tll</i> gene markerless integration	This study
pUXpr7166-hp4e ₁₀₀	pUAXp7166-hp16e ₁₀₀ -tll derivative, harboring upXpr homologous fragment	This study
pUXpr7166-hac1	pUXpr7166-hp4e ₁₀₀ derivative, a self-excising vector for <i>hac1</i> gene markerless integration	This study
pUXpr7166-kar2	pUXpr7166-hp4e ₁₀₀ derivative, a self-excising vector for <i>kar2</i> gene markerless integration	This study
pUXpr7166-pdi	pUXpr7166-hp4e ₁₀₀ derivative, a self-excising vector for <i>pdi</i> gene markerless integration	This study

Strains

Po1f	<i>MatA</i> , <i>leu2-270</i> , <i>ura3-302</i> , <i>xpr2-322</i> , <i>axp1-2</i> , <i>Leu⁻</i> , <i>Ura⁻</i> , Δ AEF, Δ AXP	[41]
Po1f/pENOn-egfp (n = 75, 100, 125, 150, 175, 200, 300, 400, 700, 1000, and 1300 bp)	eGFP expressed in Po1f using pENOn-egfp	This study
Po1f/hp4d-egfp	eGFP expressed in Po1f using hp4d-egfp	This study
Po1f/pENOn-tll (n = 100, 125, 200, 400, 700, 1000, and 1300 bp)	TLL expressed in Po1f using pENOn-tll	This study
Po1f/hp4d-tll	TLL expressed in Po1f using hp4d-tll	This study
Po1f/hp4en-tll (n = 100, 125, 200, 400, 700, 1000, and 1300 bp)	TLL expressed in Po1f using hp4en-tll	This study
Po1f/hpxe100-tll (x = 8, 12, 16, 20, 24, 28, and 32)	TLL expressed in Po1f using hpxe100-tll	This study
Po1f/1tll	Po1f derivative, harboring 1 copy of <i>tll</i> expression cassette	This study
Po1f/2tll	Po1f/1tll derivative, harboring 2 copies of <i>tll</i> expression cassettes	This study
Po1f/3tll	Po1f/2tll derivative, harboring 3 copies of <i>tll</i> expression cassettes	This study
Po1f/4tll	Po1f/3tll derivative, harboring 4 copies of <i>tll</i> expression cassettes	This study
Po1f/3tll-hp4e100	Po1f/3tll derivative, harboring an empty expression cassette	This study
Po1f/3tll-hac1	Po1f/3tll harboring <i>hac1</i> gene cassette	This study
Po1f/3tll-kar2	Po1f/3tll harboring <i>kar2</i> gene cassette	This study
Po1f/3tll-pdi	Po1f/3tll harboring <i>pdi</i> gene cassette	This study
Po1f/3tll-hac1-pdi	Po1f/3tll harboring <i>hac1</i> gene cassette and <i>pdi</i> gene cassette	This study
Po1f/3tll-hac1-kar2	Po1f/3tll harboring <i>hac1</i> gene cassette and <i>kar2</i> gene cassette	This study
Po1f/3tll-hac1-pdi-kar2	Po1f/3tll harboring <i>hac1</i> gene cassette, <i>pdi</i> gene cassette, and <i>kar2</i> gene cassette	This study

Table S3. Primers used in this study.

Primers	Sequence (5'-3')	Annotation
egfp-F	CCCA <u>AGCTT</u> ATGGTGAGCAAGGGCGAGGAG	<i>Hind</i> III site (underlined)
egfp-R	GGGGT <u>ACCTT</u> ACTTGTACAGCTCGTCC	<i>Kpn</i> I site (underlined)
TLL-F	ACCGCCTTTACTATTCTCAC <u>GGCCGTTCTGGCCAT</u> GGAGGTCTCCCAGGACCTGTTC	<i>Sfi</i> I site (underlined)
TLL-R	CGGCAACGTGGGGACAGGCCATGGAG <u>GTACCTTA</u> CAGGCAGGTGCCGATCAGGCCG	<i>Kpn</i> I site (underlined)
pENO ₇₅ -F1	GAGCACCGCCGCCGCAAGGAATGGT <u>GGATCCTAA</u> AAGGAGCAATCCCCACTAG	<i>Bam</i> HI site (underlined)
pENO ₁₀₀ -F1	GAGCACCGCCGCCGCAAGGAATGGT <u>GGATCCAGC</u> GCAATTGGGTTTGCATCATGTA	<i>Bam</i> HI site (underlined)
pENO ₁₂₅ -F1	GAGCACCGCCGCCGCAAGGAATGGT <u>GGATCCGAT</u> GACTATTTGGGACCAATCCAA	<i>Bam</i> HI site (underlined)
pENO ₁₅₀ -F1	GAGCACCGCCGCCGCAAGGAATGGT <u>GGATCCTGA</u> CGAAAGCACCCCCCTTTGTACA	<i>Bam</i> HI site (underlined)
pENO ₁₇₅ -F1	GAGCACCGCCGCCGCAAGGAATGGT <u>GGATCCCC</u> CCAATGGCTTTTAACTTTTCGAA	<i>Bam</i> HI site (underlined)
pENO ₂₀₀ -F1	GAGCACCGCCGCCGCAAGGAATGGT <u>GGATCCAGT</u> CCTTCAACAATCCATCTCACC	<i>Bam</i> HI site (underlined)
pENO ₃₀₀ -F1	GAGCACCGCCGCCGCAAGGAATGGT <u>GGATCCGTA</u> TTTGAGAGCAAGTTTGGGATT	<i>Bam</i> HI site (underlined)
pENO ₄₀₀ -F1	GAGCACCGCCGCCGCAAGGAATGGT <u>GGATCCTTT</u> CACTTGCCAGAACTCTAAGCG	<i>Bam</i> HI site (underlined)
pENO ₇₀₀ -F1	GAGCACCGCCGCCGCAAGGAATGGT <u>GGATCCGTC</u> CAGTGCAACCCAGACAGCTGAG	<i>Bam</i> HI site (underlined)
pENO ₁₀₀₀ -F1	GAGCACCGCCGCCGCAAGGAATGGT <u>GGATCCCCA</u> TGGTGCGTGGAGGCTTTGGCA	<i>Bam</i> HI site (underlined)
pENO ₁₃₀₀ -F1	GAGCACCGCCGCCGCAAGGAATGGT <u>GGATCCATC</u> TACCTCGGCCCTTGTTTCTTGA	<i>Bam</i> HI site (underlined)

pENO-R1	TCCTCGCCCTTGCTCACCAT <u>AAGCTT</u> GGTGATAAA TGTGTGGTTAGACGGG	<i>Hind</i> III site (underlined)
pENO ₁₀₀ -F2	GGA <u>AAGATCT</u> AGCGCAATTGGGTTTGCATCATGTA	<i>Bgl</i> II site (underlined)
pENO ₁₂₅ -F2	GGA <u>AAGATCT</u> GATGACTATTTGGGACCAATCCAA	<i>Bgl</i> II site (underlined)
pENO ₂₀₀ -F2	GGA <u>AAGATCT</u> AGTCCTTCAACAATCCATCTCACC	<i>Bgl</i> II site (underlined)
pENO ₄₀₀ -F2	GGA <u>AAGATCT</u> TTTCACTTGCCAGAACTCTAAGCG	<i>Bgl</i> II site (underlined)
pENO ₇₀₀ -F2	GGA <u>AAGATCT</u> GTCCAGTGCAACCCAGACAGCTGAG	<i>Bgl</i> II site (underlined)
pENO ₁₀₀₀ -F2	GGA <u>AAGATCT</u> CCATGGTGCGTGGAGGCTTTGGCA	<i>Bgl</i> II site (underlined)
pENO ₁₃₀₀ -F2	GGA <u>AAGATCT</u> ATCTACCTCGGCCCTTGTTCCTTGA	<i>Bgl</i> II site (underlined)
dwCre-F	GCCTGCTGGAAGATGGCGATTAG	-
lipTT66-R1	GTGTGAGATACCGTTCGTATAATGTATGCTATACG AAGTTATCTCCACCTGTGTCAATC	-
66upAxp-F	CATTATACGAACGGTATCTCACACCCACCTCCAAG	-
upAxp-R	GGGAGAGCTCTAGAGTCGACACGCGTCG <u>ACTAGT</u> CTTTAGAATTGGAAGCTGATC	<i>Spe</i> I site (underlined)
hp4d-F	CTAGTCGACGCGTGTGCGACTCTAGAGCTC	-
Xpr2pre-R	GGCCAGAACGGCCGTGAGAATAGTAAAG	-
ROL-F	CTATTCTCACGGCCGTTCTGGCCGTGCCTGTGTCT GGCAAGTC	-
kpnROL-R	GAGGT <u>ACCCT</u> ACAGACAGGAGCCCTCGTTG	<i>Kpn</i> I site (underlined)
XprTT-F	CAACGAGGGCTCCTGTCTGTAGGGTACCTCCATGG CCTGTCCCCACGTTG	-
Xprlox71-R	CACGAGGCCCTTTCTGTCTTCAAGA <u>AATTC</u> GAGACCT CATGTTTGACAGCTTATCATAAC	<i>Eco</i> RI site (underlined)
Yvgb-F	CCACA <u>AAGCTT</u> GCCACCATGTTGGATCAACAGAC CATTAAC	<i>Hind</i> III site (underlined)
Yvgb-R	CAACGTGGGGACAGGCCATGGAG <u>GCGGCCG</u> CTTAT TCAACAGCTTGAGCG	<i>Not</i> I site (underlined)
mLeu-F	GA <u>AAGATCT</u> ACTGATCACGGGCAAAAG	<i>Bgl</i> II site (underlined)

mLeu-R	CATGGTGGC <u>AAGCTT</u> TGTGG	<i>Hind</i> III site (underlined)
XprTT-F2	TCCATGGCCTGTCCCCACGTTG	-
Ampup-R	GAAGCATTTATCAGGGTTATTG	-
lipTT66-R2	CTGTCTACCGTTCGTATAATGTATGCTATACGAAG TTATCTCCACCTGTGTCAATC	-
66upXpr-F	CATTATACGAACGGTAGACAGTTAGAGCAGCAAC GCG	-
upXpr-R	CTCTAGAGTCGACACGCGTCG <u>ACTAGT</u> GGGCCCCGT CGATGGGGTTTATATC	<i>Spe</i> I site (underlined)
pENO ₁₀₀ -F3	GGA <u>AAGATCT</u> AGCGCAATTGGGTTTGCATCATGTA	<i>Bgl</i> II site (underlined)
pENO ₁₀₀ -F4	CGCGGATCCAGCGCAATTGGGTTTGCATCATGTA	<i>Bam</i> HI site (underlined)
pENO-R2	CATGGTGGC <u>AAGCTT</u> GGTGATAAATGTGTGGTTAG ACG	<i>Hind</i> III site (underlined)
pENO-R3	ATTTGCGGCCGCGGTGATAAATGTGTGGTTAGACG	<i>Not</i> I site (underlined)
Kar2-F	ATCACCA <u>AAGCTT</u> GCCACCATGAAGTTCTCTATGCC TTCGTGGG	<i>Hind</i> III site (underlined)
Kar2-R	AAGGAAAAAAGCGGCCGCTTAAAGCTCATCGTGG AAAGGAGCC	<i>Not</i> I site (underlined)
Pdi-F	ATCACCA <u>AAGCTT</u> GCCACCATGAAGTTCACTGCCCT CACAATTG	<i>Hind</i> III site (underlined)
Pdi-R	AAGGAAAAAAGCGGCCGCTTAAAGCTCATCATCA ATCTTGCCT	<i>Not</i> I site (underlined)
Hac1-F	ATCACCA <u>AAGCTT</u> GCCACCATGTCTATCAAGCGAGA AGAGTCCT	<i>Hind</i> III site (underlined)
Hac1-R	AAGGAAAAAAGCGGCCGCTCACTCCAATCCCCCA AACAGGTTG	<i>Not</i> I site (underlined)
Axpout-R	CGACCTGGAGAAGATCTGTG	-
qact1-F	CTGGCCGAGATCTTACCGAC	RT-PCR for <i>act1</i> gene
qact1-R	CATCGGGAAGCTCGTAGGAC	RT-PCR for <i>act1</i> gene

qtll-F	CTGGACAACACCAACAAGCTG	RT-PCR for <i>tll</i> gene
qtll-R	GTCCTCAACCTTCTGTCGGAG	RT-PCR for <i>tll</i> gene

Supplementary Figures

Figure captions

Figure S1. PCR identification of several Polf/tll strains. (A) PCR using primers XprTT-F2 and Axpout-R, CK: Polf (control); (B) PCR using primers TLL-F and TLL-R, CK: pUAxp7166-hp16e₁₀₀-tll (control); H₂O: water (control).

Figure S2. (A) Effect of glycosylation on TLL expressed by *Y. lipolytica*; lane M, molecular weight marker; lane 1, Polf/tll fermentation supernatant; lane 2, deglycosylation of TLL expressed by Polf/tll; lane 3, Endo H; (B) Mass spectrometry results for TLL, partially matched peptides are shown in red.

Figure S3. Optimization of TLL fermentation in 500-mL shake flasks. (A) Effects of different D-sorbitol concentrations on TLL production; (B) Relationship between TLL activity and inoculation density; (C) Influences of different initial pH on TLL production; (D) Effects of culture medium volume on TLL expression. (E) Time course of TLL expression.

Figure S4. Schematic diagram of recombinant plasmids. (A) hp4d-egfp; (B) pENOn-egfp (n = 75, 100, 125, 150, 175, 200, 300, 400, 700, 1000, and 1300 bp); (C) hp4d-tll; (D) pENOn-tll (n = 100, 125, 200, 400, 700, 1000, and 1300 bp); (E) hp4e_n-tll (n = 100, 125, 200, 400, 700, 1000, and 1300 bp).

Figure S5. Flow diagram of the steps used for construction of expression vectors $hp4e_{100}$ -tll ($x = 8, 12, 16, 20, 24, 28, \text{ and } 32$) harboring different copies of UAS1B.

Figure S6. Schematic diagram of recombinant plasmids. (A) pUAXp7166- $hp16e_{100}$ -tll; (B) pUXpr7166-type plasmids, helper gene means $hp4e_{100}$ (control), *hac1*, *kar2*, and *pdi*, respectively.

Figure S1

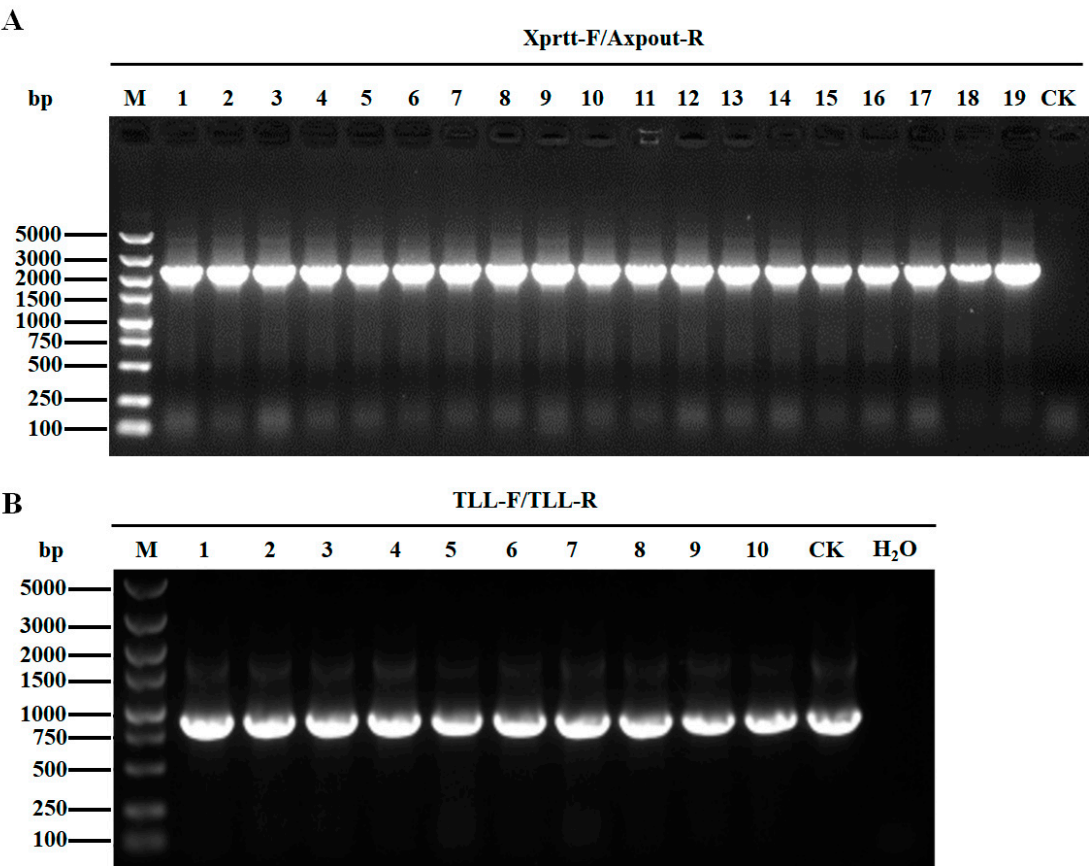


Figure S2

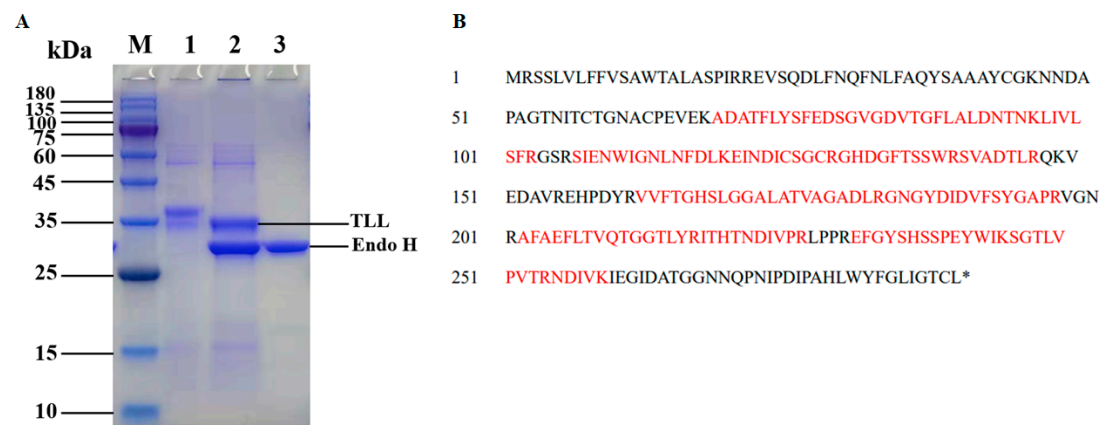


Figure S3

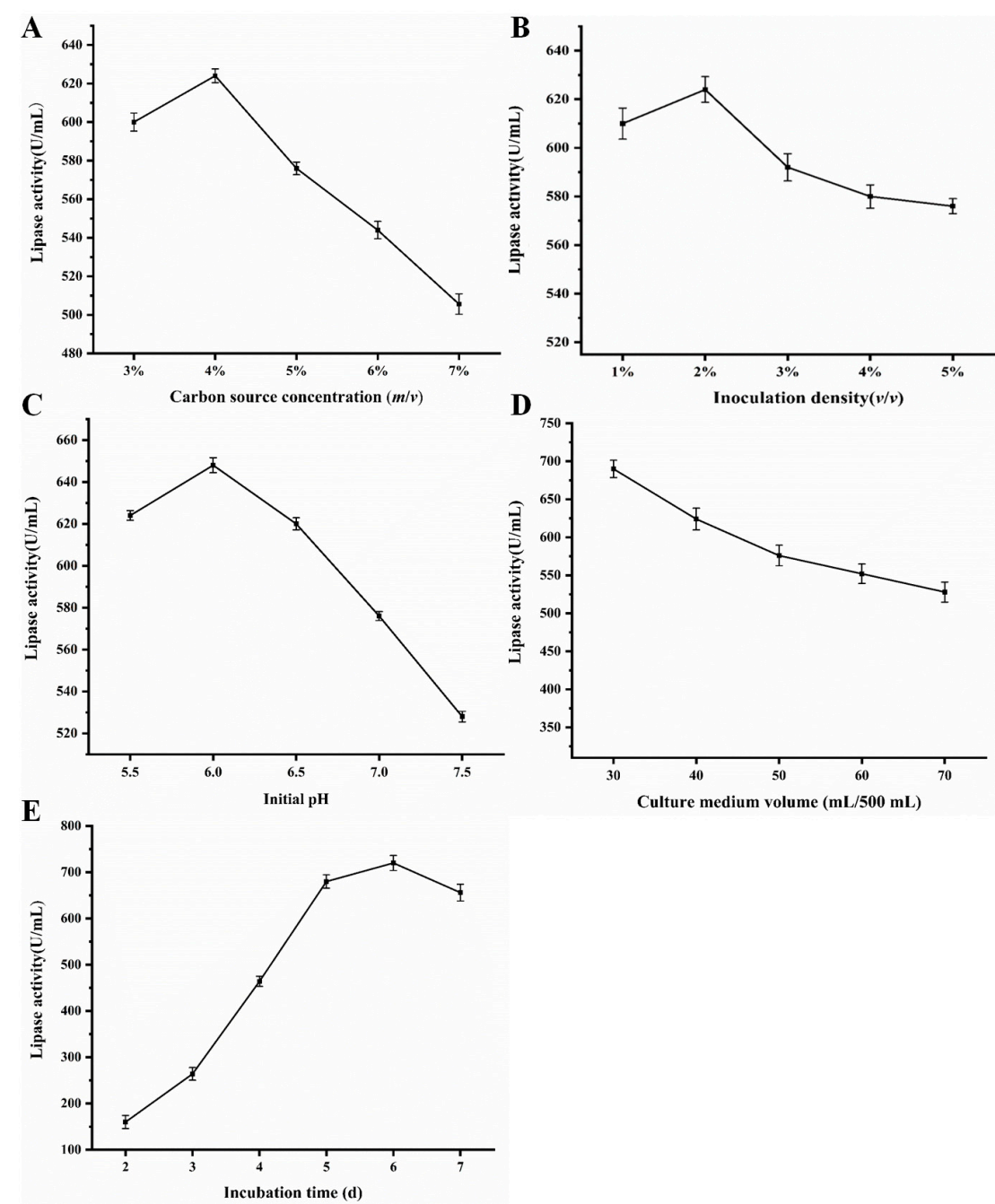


Figure S4

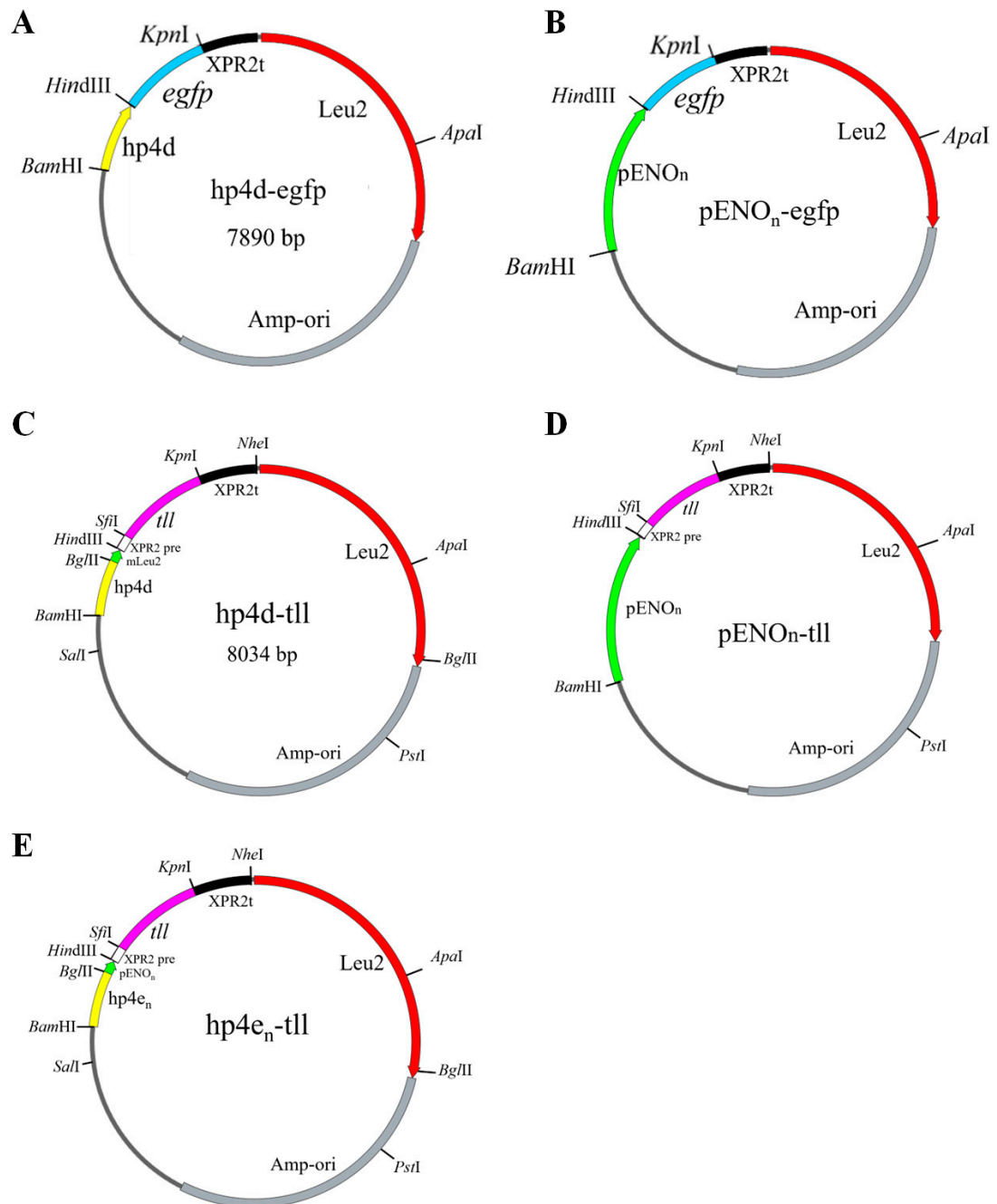


Figure S5

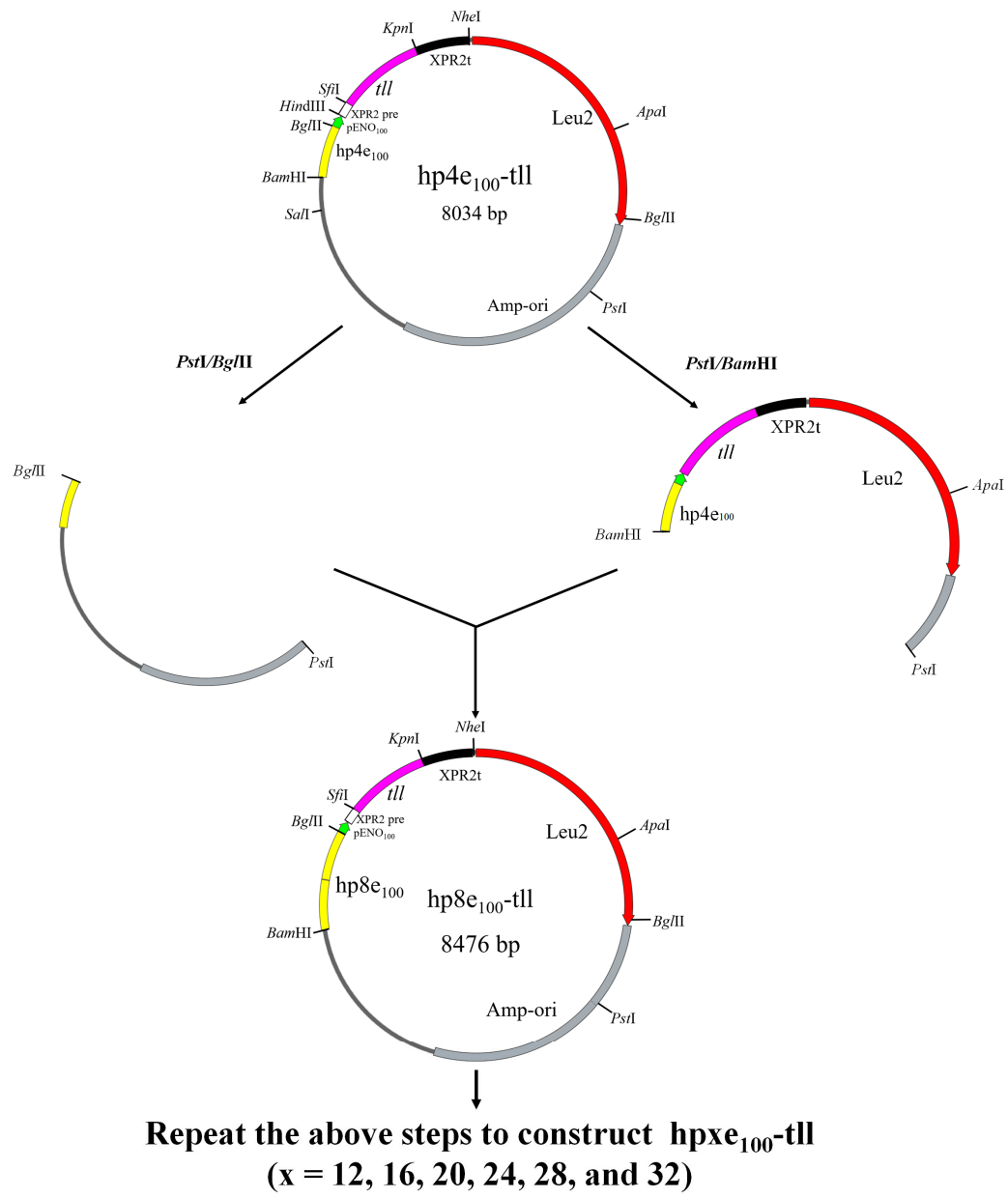


Figure S6

