

Communication **GL-II-73, a Positive Allosteric Modulator of** α**5GABA^A Receptors, Reverses Dopamine System Dysfunction Associated with Pilocarpine-Induced Temporal Lobe Epilepsy**

Alexandra M. McCoy 1,2 [,](https://orcid.org/0000-0002-8449-7717) Thomas D. Prevot 3,4, Dishary Sharmin ⁵ , James M. Cook ⁵ [,](https://orcid.org/0000-0001-5512-3022) Etienne L. Sibille 3,4,6 and Daniel J. Lodge 1,2,*

- ¹ Department of Pharmacology and Center for Biomedical Neuroscience, UT Health San Antonio, San Antonio, TX 78229, USA; mccoya1@livemail.uthscsa.edu
- ² South Texas Veterans Health Care System, Audie L. Murphy Division, San Antonio, TX 78229, USA
³ Campbell Family Mantal Health Pessagge Institute of CAMH Texanta ON M5S 251 Canada:
- ³ Campbell Family Mental Health Research Institute of CAMH, Toronto, ON M5S 2S1, Canada; thomas.prevot@camh.ca (T.D.P.); etienne.sibille@camh.ca (E.L.S.)
- ⁴ Department of Psychiatry, University of Toronto, Toronto, ON M5T 1R8, Canada
- ⁵ Department of Chemistry and Biochemistry, University of Wisconsin-Milwaukee, Milwaukee, WI 53201, USA; dsharmin@uwm.edu (D.S.); capncook@uwm.edu (J.M.C.)
- ⁶ Department of Pharmacology and Toxicology, University of Toronto, Toronto, ON M5T 1R8, Canada
- ***** Correspondence: lodged@uthscsa.edu

Abstract: Although seizures are a hallmark feature of temporal lobe epilepsy (TLE), psychiatric comorbidities, including psychosis, are frequently associated with TLE and contribute to decreased quality of life. Currently, there are no defined therapeutic protocols to manage psychosis in TLE patients, as antipsychotic agents may induce epileptic seizures and are associated with severe side effects and pharmacokinetic and pharmacodynamic interactions with antiepileptic drugs. Thus, novel treatment strategies are necessary. Several lines of evidence suggest that hippocampal hyperactivity is central to the pathology of both TLE and psychosis; therefore, restoring hippocampal activity back to normal levels may be a novel therapeutic approach for treating psychosis in TLE. In rodent models, increased activity in the ventral hippocampus (vHipp) results in aberrant dopamine system function, which is thought to underlie symptoms of psychosis. Indeed, we have previously demonstrated that targeting α 5-containing γ-aminobutyric acid receptors (α 5GABA_ARs), an inhibitory receptor abundant in the hippocampus, with positive allosteric modulators (PAMs), can restore dopamine system function in rodent models displaying hippocampal hyperactivity. Thus, we posited that α 5-PAMs may be beneficial in a model used to study TLE. Here, we demonstrate that pilocarpineinduced TLE is associated with increased VTA dopamine neuron activity, an effect that was completely reversed by intra-vHipp administration of GL-II-73, a selective α5-PAM. Further, pilocarpine did not alter the hippocampal α 5GABA_AR expression or synaptic localization that may affect the efficacy of α 5-PAMs. Taken together, these results suggest augmenting α 5GABA_AR function as a novel therapeutic modality for the treatment of psychosis in TLE.

Keywords: dopamine; temporal lobe epilepsy; electrophysiology

1. Introduction

Temporal lobe epilepsy (TLE) is a disorder characterized by the presence of spontaneous seizures, or uncontrolled neural activity, involving the temporal lobe, typically the hippocampal formation. Given that seizures are the hallmark feature of TLE, other neuropsychiatric comorbidities, such as psychosis, are often overlooked, despite the wellestablished relationship between TLE and psychotic symptoms [\[1](#page-8-0)[–4\]](#page-8-1). TLE patients are 6–12 times more likely than the general population to suffer from psychosis [\[5](#page-8-2)[,6\]](#page-8-3); however, the management of psychosis for patients with TLE is currently inadequate. Both firstand second-generation antipsychotics are associated with an increase in seizure incidence,

Citation: McCoy, A.M.; Prevot, T.D.; Sharmin, D.; Cook, J.M.; Sibille, E.L.; Lodge, D.J. GL-II-73, a Positive Allosteric Modulator of α5GABA^A Receptors, Reverses Dopamine System Dysfunction Associated with Pilocarpine-Induced Temporal Lobe Epilepsy. *Int. J. Mol. Sci.* **2023**, *24*, 11588. [https://doi.org/10.3390/](https://doi.org/10.3390/ijms241411588) [ijms241411588](https://doi.org/10.3390/ijms241411588)

Academic Editor: Alexandre Hiroaki Kihara

Received: 25 June 2023 Revised: 15 July 2023 Accepted: 16 July 2023 Published: 18 July 2023

Copyright: © 2023 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license [\(https://](https://creativecommons.org/licenses/by/4.0/) [creativecommons.org/licenses/by/](https://creativecommons.org/licenses/by/4.0/) $4.0/$).

as they can lower seizure thresholds [\[7](#page-9-0)[,8\]](#page-9-1). Additionally, antipsychotics often produce intolerable side effects that lead to the discontinuation of medication [\[9\]](#page-9-2), and have potential pharmacokinetic and pharmacodynamic interactions with antiepileptic drugs [\[7\]](#page-9-0). Thus, there are limited treatment options for the management of psychosis in patients with TLE.

The hippocampus is a region that is central to the pathology of both TLE and psychosis, and is the area of the brain with the lowest seizure threshold [\[10](#page-9-3)[,11\]](#page-9-4). Although psychosis is not present in all patients with TLE, they are 6–12 times more likely to develop psychosis compared to the general population [\[6\]](#page-8-3), with psychosis risk being associated with numerous factors including early onset of epilepsy, a history of seizures, hippocampal sclerosis, and hippocampal abnormalities including cell loss in the CA1 region (for review, see [\[2\]](#page-8-4)). It has been suggested that deficits in GABAergic signaling in the hippocampus contribute to some forms of epilepsy [\[12\]](#page-9-5), perhaps due to epilepsy-associated interneuron death [\[13\]](#page-9-6). Interestingly, hippocampal abnormalities including increased activity at rest have been observed in psychosis patients, and are correlated with psychosis symptom severity [\[14–](#page-9-7)[18\]](#page-9-8). Additionally, a loss of parvalbumin (PV)-positive interneurons in the hippocampus has been reported in post-mortem studies of psychosis patients [\[19\]](#page-9-9) and in rodent models used to study psychosis [\[20\]](#page-9-10). Thus, a loss of inhibitory tone and subsequent hyperactivity in the hippocampus may be a point of pathological convergence for both TLE and psychosis. Indeed, our laboratory, and others, have shown that hyperactivity in the ventral hippocampus (vHipp) drives aberrant dopamine system function in rodents [\[21](#page-9-11)[–23\]](#page-9-12), with increased dopamine neuron activity being critical for generating symptoms of psychosis [\[24](#page-9-13)[,25\]](#page-9-14). Importantly, preclinical studies have demonstrated that restoring inhibitory tone in the vHipp can alleviate dopamine system dysfunction and psychosis-like phenotypes in animal models used to study psychosis [\[26](#page-9-15)[–30\]](#page-9-16). These data provide a strong rationale for targeting GABAergic neurotransmission in the hippocampus as a potential site of therapeutic intervention for the treatment of psychosis in patients with TLE.

One potential approach to normalize hippocampal activity may be to target $γ$ -aminobutyric acid type A receptors containing the $α5$ subunit ($α5GABA_ARs$), which have a well-characterized expression profile, and are primarily expressed within the hippocampal formation [\[31,](#page-9-17)[32\]](#page-10-0). Recent studies have demonstrated that positive allosteric modulators (PAMs), selective for α 5GABA_ARs, can produce a host of beneficial effects, including antidepressant, anxiolytic, procognitive [\[33](#page-10-1)[,34\]](#page-10-2), and antipsychotic-like effects [\[28,](#page-9-18)[35\]](#page-10-3). Specifically, α 5-PAMs have been shown to restore aberrant dopamine neuron activity in two different animal models used to study psychosis [\[28,](#page-9-18)[35\]](#page-10-3); however, these antipsychotic-like effects appear to be dependent on the extrasynaptic location of α 5GABA_AR (extrasynaptic versus synaptic [\[36\]](#page-10-4).

Periods of high neural activity, like those seen during seizures, have been shown to shunt hippocampal α 5GABA_ARs into the synapse in vitro [\[37\]](#page-10-5), where an α 5-PAM may be less effective [\[36\]](#page-10-4). For these reasons, we examined whether α 5GABA_ARs translocate into the synapse in the pilocarpine model used to study TLE, and investigated if an α 5-PAM, GL-II-73, was able to restore dopamine system function, using in vivo electrophysiology. GL-II-73 has been characterized as a positive allosteric modulator at the GABA_A receptor, with priority affinity and efficacy at α 5GABA_ARs [\[33\]](#page-10-1). Specifically, Ki values at the α 5 subunit were shown to be 6 times more selective compared to α 2, 11 times more compared to α 1, and 12.5 times more compared to α 3. Efficacy at α 5GABA_ARs also showed PAM activity, showing preferential α 5-PAM activity compared to other α -subunits, most importantly compared to α 1. We observed a significant increase in VTA dopamine neuron activity following recurrent seizures, which was reversed by intra-vHipp administration of the α5-PAM, GL-II-73. We also found that pilocarpine- and saline-treated rats did not differ in their total or synaptic expression of α 5GABA_ARs. These results suggest that restoring hippocampal inhibitory tone with GL-II-73 may have clinical utility for the treatment of psychosis associated with epilepsy. Taken together, these data provide a strong rationale

that targeting GABAergic neurotransmission in the hippocampus may be a potential site of therapeutic intervention for the treatment of psychosis in patients with TLE.

strong rationale that targeting GABAergic neurotransmission in the hippocampus may

2. Results

2.1. Pilocarpine-Induced Seizures Increased VTA Dopamine Neuron Population Activity, Which Was Reversed by GL-II-73 2.1. Pilocarpine-Induced Seizures Increased VTA Dopamine Neuron Population Activity, which 2.1. Puocurpine-inuuceu 5e

To establish a model of TLE in rodents [\[38,](#page-10-6)[39\]](#page-10-7), we unilaterally injected pilocarpine To establish a model of TLE in rodents [38,39], we unilaterally injected pilocarpine directly into the vHipp. A representative brain slice showing the cannula track is depicted directly into the vHipp. A representative brain slice showing the cannula track is depicted ancely the the VIPP. IT representance standard aboving the cannot rated is depicted in Figure [1A](#page-2-0). Consistent with previous reports [\[40\]](#page-10-8), we found that rats that received an In 1 gave 11. Consistent with provides reports $[2\sigma]$, we retain that these that received an intra-vHipp injection of pilocarpine had a significantly higher Racine rating compared to $\frac{1}{2}$ and $\frac{1}{2}$ $\frac{1}{2$

Figure 1. Verification of placements and response to pilocarpine. (A, Left) Representative brain **Figure 1.** Verification of placements and response to pilocarpine. (**A**, **Left**) Representative brain slice containing an electrode track in the ventral tegmental area (VTA; black arrow) and cannula track in the ventral hippocampus (vHipp; white arrow) with corresponding schematic of the brain section (right, −5.40 mm posterior to bregma, [\[41\]](#page-10-9)). (**B**) Pilocarpine-treated rats exhibited significantly higher cantly higher average Racine ratings compared to saline-treated controls, **** p < 0.0001, n = 6– average Racine ratings compared to saline-treated controls, **** *p* < 0.0001, *n* = 6–16/group; circles denote male subjects and squares denote females.

Dopamine system function is known to be altered in patients with [psy](#page-9-13)[ch](#page-9-14)[osis](#page-10-10) Dopamine system function is known to be altered in patients with psychosis [24,25,42] and in rodent models used to study [ps](#page-9-11)[ych](#page-10-11)[osi](#page-10-12)s [21,43–45], as well as in pilocarpine-treated rats [40]. Consistent with previous findings, we observed a significant increase in dopamine neuron activity in pilocarpine-treated rats that received intra-v[Hip](#page-3-0)p vehicle (Figure 2A, $n = 13$ rats; 1.88 \pm 0.10 cells per track; two-way ANOVA; $F_{\text{Pilocarpine (1,33)}} = 15.370; p < 0.001;$ $F_{\text{Drug (1,33)}} = 7.121; p = 0.012; Holm-Sidak; t = 3.977, p = 0.002$) when compared to salinetreated rats that received intra-vHipp vehicle ($n = 6$ rats; 1.13 ± 0.13 cells per track). Interestingly, the pilocarpine-induced increase in dopamine neuron activity was completely reversed by the intra-vHipp administration of GL-II-73 ($n = 12$ rats; 1.30 ± 0.13 cells per track; Holm–Sidak; $t = 3.800$, $p = 0.002$), and had no effect in saline-treated rats ($n = 6$ rats; 1.00 ± 0.09 cells per track). Consistent with a previous observation [\[40\]](#page-10-8), a subset of rats (4 of 17) did not develop a hyperdopaminergic phenotype (*n* = 4 rats; 1.05 ± 0.13 cells per track), were classified as non-responders (defined as >1.5 cells per track), and were not included in

this dataset. As expected, no significant differences were observed in the average firing rate (Figure 2B; F_{Pilocarpine(1,228)} = 0.140; $p = 0.709$; F_{Drug (1,228)} = 0.039; $p = 0.844$) or bursting pat-tern (Figure [2C](#page-3-0); F_{Pilocarpine (1,228)} < 0.001; $p = 0.989$; $F_{\text{Drug (1,228)}} = 0.034$; $p = 0.853$). It should be noted that while both male (circles) and female (squares) subjects were represented, we are underpowered to detect any sex differences.

vation \mathcal{A} subset of \mathcal{A} of \mathcal{A} of \mathcal{A} of \mathcal{A} and develope (n $=$ \mathcal{A} of \mathcal{A} of \mathcal{A} of \mathcal{A}

Figure 2. GL-II-73 reverses pilocarpine-induced increases in VTA dopamine neuron activity. (A) **Figure 2.** GL-II-73 reverses pilocarpine-induced increases in VTA dopamine neuron activity. (**A**) Intraventral hippocampal (vHipp) pilocarpine significantly increased the number of spontaneously active ously active dopamine cells per track (population activity), which was reversed by intra-vHipp dopamine cells per track (population activity), which was reversed by intra-vHipp administration of $\frac{1}{2}$ II-72 (100 ng/ $\frac{1}{2}$, 0.75 $\frac{1}{2}$). Average firing rate (B) and percent burst firing (C) were not abanged GL-II-73 (100 ng/ μ L; 0.75 μ L). Average firing rate (**B**) and percent burst firing (**C**) were not changed by pilocarpine or drug treatment. (**D**) Representative trace of dopamine neuron firing and action potential for saline-treated (left) and pilocarpine-treated (right) rats. ** *p* < 0.005, *n* = 6–13/group; circles denote male subjects and squares denote females.

2.2. Pilocarpine-Induced Seizures Did Not Alter α5GABAAR Expression within the vHipp *2.2. Pilocarpine-Induced Seizures Did Not Alter α5GABAAR Expression within the vHipp*

The localization of α5GABAARs (extrasynaptic vs. synaptic) plays a key role in the The localization of α5GABAARs (extrasynaptic vs. synaptic) plays a key role in the hippocampal modulation of dopamine neuron activity [36] and activity-dependent hippocampal modulation of dopamine neuron activity [\[36\]](#page-10-4) and activity-dependent changes in α 5GABA_AR localization are known to occur [\[37\]](#page-10-5). Thus, we verified the expression and localization of α 5GABA_ARs between saline and pilocarpine-treated rats. To examine synaptic localization, we measured gephyrin-associated α5GABA_ARs using coimmunoprecipitation. We found no differences in total α5 protein levels (Figure [3A](#page-4-0), *t* test; *t* = 1.075

 $p = 0.314$) or gephyrin-pulldown α5 protein levels (Figure [3B](#page-4-0), t test; t = 1.631, p = 0.142) between pilocarpine-treated and saline-treated rats.

Figure 3. Pilocarpine does not alter total or synaptic α5 protein. There were no differences in total **Figure 3.** Pilocarpine does not alter total or synaptic α5 protein. There were no differences in total α5 protein expression; (A) co-immunoprecipitation of α 5 and gephyrin was used to measure synaptic α 5 protein (B) between saline- and pilocarpine-treated rats. Representative bands from saline- (left) α 5 protein (**B**) between saline- and pilocarpine-treated rats. Representative bands from saline- (**left**) and pilocarpine-treated rats (**right**) are below graphs. $n = 5$ rats/group, circles denote male subjects and squares denote females.

3. Discussion **3. Discussion**

Despite the fact that temporal lobe epilepsy (TLE) patients are 6–12 times more Despite the fact that temporal lobe epilepsy (TLE) patients are 6–12 times more likely than the general population to suffer from psychosis, treatments for psychosis in this patient population remain inadequate [\[5](#page-8-2)[,6\]](#page-8-3). Antipsychotic medications incompletely treat psychosis symptoms and are associated with substantial adverse effects [\[9\]](#page-9-2). Additionally, there is evidence to suggest that antipsychotics can lower seizure threshold and increase seizure incidence [\[7](#page-9-0)[,8\]](#page-9-1), and undergo potential pharmacokinetic and pharmacodynamic interactions with antiepileptic drugs $[7]$. Thus, there is an immediate need for novel therapeutics to treat psychosis in TLE patients.

Augmented hippocampal activity appears to be central to the pathology of both TLE Augmented hippocampal activity appears to be central to the pathology of both TLE and psychosis [16,21,46], with diminished GABAergic signaling within the hippocampus and psychosis [\[16,](#page-9-19)[21,](#page-9-11)[46\]](#page-10-13), with diminished GABAergic signaling within the hippocampus likely contributing to pathophysiology $[20,47-50]$ $[20,47-50]$ $[20,47-50]$. α5GABA_A receptors are inhibitory receptors highly enriched in the hippocampus [\[31](#page-9-17)[,32\]](#page-10-0). Thus, we were interested to see if \overline{F} restoring hippocampal inhibition by administering an α5-PAM, GL-II-73, could reverse restoring hippocampal inhibition by administering an α5-PAM, GL-II-73, could reverse aberrant dopamine system function in a rodent model used to study TLE. aberrant dopamine system function in a rodent model used to study TLE.

In this study, we induced spontaneous recurrent seizures by injecting the political services of π compound, pilocarpine, directly into the vHipp [\[38](#page-10-6)[,39](#page-10-7)[,51\]](#page-10-16). The administration of pilo-
complex a museuminism exercise a partial is a suall astablished medal word to study TLE in etaphic) a mascurinic receptor agonist, is a well-established model used to stady TEE in rodents [\[13,](#page-9-6)[38](#page-10-6)[,39\]](#page-10-7). Both systemic and intra-vHipp administration of pilocarpine can genstudy TLE in rodents [13,38,39]. Both systemic and intra-vHipp administration of pilo-erate status epilepticus within minutes, and can produce spontaneous recurrent seizures effective status epilepticus within minutes, and can produce spontaneous recentrum schlares after a latent period of several weeks, but intracranial administration is associated with a much lower mortality rate [\[51\]](#page-10-16), hence our choice to use this route of administration. Inducing TLE with pro-epileptic compounds has been shown to result in behavioral and neurophysiological alterations associated with psychosis, including hypersensitivity to amphetamine [\[52\]](#page-10-17) and increased VTA dopamine neuron activity [\[40\]](#page-10-8). Consistent with these studies, we observed increases in VTA dopamine neuron population activity, without changes to firing rate or burst firing (Figure [2\)](#page-3-0). This is consistent with the broader body of literature demonstrating a link between hippocampal hyperactivity, aberrant dopamine neuron activity, and psychosis symptoms. Specifically, clinical studies have indicated In this study, we induced spontaneous recurrent seizures by injecting the pro-epileptic carpine, a muscarinic receptor agonist, is a well-established model used to study TLE in that psychosis patients display hippocampal abnormalities [\[16](#page-9-19)[,50](#page-10-15)[,53\]](#page-10-18). Additionally, our

laboratory and others have demonstrated that, in rodents, vHipp hyperactivity can profoundly augment VTA dopamine neuron activity via a multisynaptic circuit [\[21–](#page-9-11)[23,](#page-9-12)[54\]](#page-10-19). VTA population activity, rather than firing rate or burst firing, is consistently altered in animal models of psychosis, and is regulated by the hippocampus [\[21](#page-9-11)[,22](#page-9-20)[,54\]](#page-10-19), whereas other aspects of VTA dopamine neuron activity, such as burst firing, are regulated by regions such as the laterodorsal tegmental and pedunculopontine tegmental nuclei [\[43,](#page-10-11)[55](#page-10-20)[,56\]](#page-10-21). Thus, we believe that the increased VTA dopamine neuron population activity observed in pilocarpine-treated rats was due to increased vHipp activity.

Consistent with previous reports [\[40\]](#page-10-8), a subset of pilocarpine-treated rats (25%) did not display elevated dopamine neuron activity (<1.5 spontaneously active cells/track). This segregation of "responders" and "non-responders" was based on the clear bimodal distribution of dopamine neuron population activity observed in previous studies [\[57\]](#page-11-0). However, the presence of non-responders is consistent with clinical outcomes, in which only a portion of TLE patients develop psychosis [\[5,](#page-8-2)[6\]](#page-8-3). While the origin of the non-responders is unknown, it has been suggested to be due to the use of anesthetic during dopamine neuron recordings, which acts on the GABAergic system [\[58,](#page-11-1)[59\]](#page-11-2), similar to many antiepileptic drugs [\[60\]](#page-11-3), and may decrease hippocampal seizures [\[40\]](#page-10-8).

Here we demonstrate that intra-vHipp pilocarpine can produce aberrant dopamine neuron activity, and that the intra-vHipp administration of GL-II-73 fully reverses dopamine system dysfunction caused by TLE induction. This is consistent with the growing body of literature demonstrating that attenuating activity in the vHipp using a variety of methods including pharmacological [\[23\]](#page-9-12), surgical [\[30\]](#page-9-16) or cell-based approaches [\[29\]](#page-9-21) can restore aberrant dopamine system function in animal models used to study psychosis. Further, GL-II-73 and other α5-PAMs have shown promising therapeutic efficacy in multiple animal models used to study psychosis [\[28,](#page-9-18)[35,](#page-10-3)[61\]](#page-11-4). In the current study, we provide evidence that the therapeutic utility of GL-II-73 extends to epilepsy-related psychosis as well. This is an important discovery, as it has been shown that $GABA_AR$ expression may be altered in some rat models of TLE, and a reduction in α 5GABA_A receptor expression may decrease drug efficacy [\[12](#page-9-5)[,48,](#page-10-22)[49,](#page-10-23)[62\]](#page-11-5). Specifically, a downregulation of α 5GABA_ARs in the CA1 and CA2 regions of the hippocampus has been observed in pilocarpine-treated rats [\[47\]](#page-10-14). While we did not observe any differences in α 5GABA_AR expression here, methodological differences could be responsible for this discrepancy. We took tissue following our electrophysiology experiments, which occurred mere days after the first spontaneous seizures appeared (which ranged from 2 to 6 weeks after pilocarpine administration), whereas others have observed differences in α 5GABA_AR in rats that had been treated with pilocarpine 3–4 months prior [\[47\]](#page-10-14). It is possible that the downregulation of α 5GABA_ARs occurs as part of the more chronic stages of pilocarpine-induced epilepsy, and at our timepoint, changes are undetectable. While α 5GABA_AR expression may influence the efficacy of α 5-PAMs, we have recently demonstrated that shifting α 5GABA_ARs from the extrasynaptic space to the synapse abolishes the ability of GL-II-73 to modulate VTA dopamine neuron activity (McCoy et al., in submission). Since periods of high-frequency activity have been reported to shunt α 5GABA_ARs to the synapse [\[37\]](#page-10-5), we wanted to ensure that this does not occur in models of epilepsy. We found no differences in gephyrin-associated α 5GABA_ARs or in total α 5GABA_ARs, suggesting that the proportion of synaptic α 5GABA_ARs also does not change with pilocarpine treatment. These results are consistent with the electrophysiology data, which demonstrate that intra-vHipp GL-II-73 can still restore aberrant VTA dopamine neuron activity, even in a model of hippocampal hyperactivity.

4. Materials and Methods

All experiments were performed in accordance with the guidelines outlined in the USPH Guide for the Care and Use of Laboratory Animals and were approved by the Institutional Animal Care and the Use Committees of UT Health San Antonio and the U.S. Department of Veterans Affairs.

4.1. Animals

Adult male ($n = 11$) and female ($n = 11$) Sprague Dawley rats (250–300 g) purchased from Envigo (Indianapolis, IN, USA) were used for all experiments. Rats were group housed until surgery and kept on a 12 h light/dark cycle. Food and water were provided *ad libitum*.

4.2. Seizure Induction

All survival procedures were performed under general anesthesia in a semi sterile environment. Rats were anesthetized with Fluriso (2–5% isoflurane, USP, with oxygen flow at 1 L/min) and placed in a stereotaxic frame (Kopf, Tujunga, CA, USA) with blunt, atraumatic ear bars. A core body temperature of $37 °C$ was maintained. For the intracranial administration of drugs, standard guide cannulas (23 gauge; PlasticsOne, Roanoke, VA, USA) with an internal cannula (30 gauge; PlasticsOne) extending 1 mm past the end of the guide were used. Rats were randomly assigned to receive either pilocarpine $(2.4 \text{ mg}/\mu\text{L})$; 1 µL) or vehicle (1 µL; Dulbecco's phosphate-buffered saline (PBS)) and unilaterally injected (∼0.5 µL per min) in the right vHipp (from bregma: AP −5.3, ML +5.0, from brain surface: DV −6.0 mm). The dose of pilocarpine was selected based on previously published work demonstrating that a majority of rats (~75%) given this dose display a hyperdopaminergic phenotype [\[40\]](#page-10-8). This route of administration (intra-vHipp) is preferred to systemic administration as it is associated with significantly lower mortality [\[51\]](#page-10-16). Even so, three rats that were administered pilocarpine died before electrophysiology could be performed. Rats were administered ketoprofen as an analgesic (5 mg/kg, s.c.) and monitored closely until conscious. Within 5 min of waking, the majority of rats displayed status epilepticus, which was quantified using a modified Racine motor scale [\[63\]](#page-11-6) defined as the following: $0 =$ no abnormal behavior, 1 = mouth and facial spasms, 2 = head nodding/wet dog shakes, $3 =$ forelimb clonus, $4 =$ rearing, $5 =$ rearing and falling, racing, jumping. Animals exhibiting convulsions lasting more than 120 min were injected with midazolam (5 mg/kg; i.p.) to stop seizure activity. Rats were monitored by an experimenter daily in the morning starting at 8:00 am for two hours for the development of spontaneous seizures, which appeared 2–6 weeks following injection.

4.3. In Vivo Extracellular Dopamine Neuron Electrophysiology

As early as the day following the development of spontaneous seizures, but no later than 1 week after development, in vivo extracellular dopamine neuron electrophysiology was performed. Rats were anesthetized with 8% chloral hydrate (400 mg/kg, i.p.) and placed in a stereotaxic apparatus for the duration of the experiment, typically lasting no longer than three hours. GL-II-73 (100 ng/ μ L; 0.75 uL) or vehicle (85% H₂O, 14% propylene glycol, 1% Tween80; 0.75 μ L) were administered directly into the vHipp unilaterally such that each rat served as its own vehicle control. The vehicle side was always recorded first, and the hemispheres receiving each drug were counterbalanced. The dose of GL-II-73 was selected based on previous characterization [\[28](#page-9-18)[,33](#page-10-1)[,35\]](#page-10-3). Extracellular glass microelectrodes (impedance ~6–10 M Ω) were lowered into the VTA (from bregma: AP −5.3, ML ± 0.6 , and from brain surface: DV -6.5 to -9.0 mm) using a hydraulic micro-positioner (Model 640, Kopf). Spontaneously active dopamine neurons were identified using open filter settings (low-frequency cutoff: 30 Hz; high-frequency cutoff: 30 kHz) according to previously established electrophysiological criteria [\[64](#page-11-7)[,65\]](#page-11-8). Three parameters of dopamine activity were measured and analyzed: (1) the number of dopamine neurons firing spontaneously per track (population activity), (2) average firing rate, and (3) proportion of action potentials occurring in bursts (defined as the incidence of spikes with <80 ms between them; termination of the burst is defined by >160 ms between spikes). Electrophysiological data were collected with commercially available software (LabChart software; Version 8 ADInstruments, Sydney, Australia).

4.4. Verification of Electrode and Cannula Placement

At the completion of all experiments, rats were sacrificed via chloral hydrate overdose and rapidly decapitated. Brains were extracted and cut down the midsagittal line into hemispheres. One hemisphere was fixed for at least 24 h (4% phosphate-buffered formaldehyde), cryoprotected (10% *w/v* sucrose in phosphate-buffered saline) until saturated and used to verify electrode (VTA) and cannula (vHipp) placement. Brains were coronally sectioned (25 µm) using a cryostat (Leica, Buffalo Grove, IL, USA). Sections containing electrode or cannula tracks were mounted onto gelatin-coated slides, stained with neutral red (0.1%) and thionin acetate (0.01%), and cover-slipped with DPX Mountant for histochemical confirmation within the VTA or vHipp (Figure [1A](#page-2-0)). The other hemisphere was stored and used for protein expression analysis.

4.5. Immunoprecipitation and Western Blot

After electrophysiology was completed, rats were sacrificed as described above and the vHipp from either the right or left hemisphere was dissected on ice. The vHipp was then homogenized with lysis buffer, centrifuged at $14,000 \times g$ for 2 min and the supernatants were aliquoted and stored at -80 °C until further use. The α 5 subunit and its binding partners were immunoprecipitated using Dynabeads protein G magnetic beads (ThermoFisher, Waltham, MA, USA) according to the manufacturer's protocol. Western blots were used to quantify α 5 and gephyrin protein levels in both the immunoprecipitated samples and hippocampal homogenates. In short, proteins in the total lysate were separated in an SDS-PAGE gel followed by blotting onto a 0.2 μ m nitrocellulose membrane using a trans-blot turbo transfer system (Bio-Rad, Hercules, CA, USA). Membranes were then incubated with a primary antibody against α5-GABAARs (1:1000), gephyrin (1:3000), or GAPDH (1:1000) overnight at 4 \degree C, and washed 3 times (10 min) with Tris-buffered saline with 0.1% Tween 20 (TBST) prior to incubation with a horseradish peroxidase-conjugated antimouse antibody (1:5000; for GAPDH only) or an anti-rabbit secondary antibody (1:5000) for 1 h at room temperature. Membranes were washed with TBST and incubated with Pierce™ ECL western blotting substrate (ThermoFisher) followed by exposure to X-ray film for detection. Antibodies were stripped using a commercially available stripping buffer (ThermoFisher). Densitometry analyses of immunoreactive bands were performed using the NIH Image J software version 1.53e from the scanned films. Densitometric arbitrary units obtained for α 5GABA_AR immunoreactive bands were normalized to GAPDH. For coimmunoprecipitation experiments, gephyrin bands were normalized to α 5GABA_AR levels.

4.6. Statistical Analysis

All data are represented as mean \pm SEM with *n* values representing the number of rats per experimental group unless otherwise indicated. Electrophysiology data were analyzed using LabChart Pro Version 8 (ADInstruments, Colorado Springs, CO, USA). A two-way ANOVA of these data (factors: pilocarpine \times drug) was performed and plotted using Prism software version 9.5.1 (GraphPad Software Inc.; San Diego, CA, USA). When significant main effects or interactions were detected the Holm–Sidak post-hoc test was used. Western blots and Racine ratings were analyzed by t-test and plotted in Prism. All tests were two-tailed, and significance was determined at *p* < 0.05.

4.7. Materials

The proprietary compound, GL-II-73, was supplied by the Centre for Addiction and Mental Health, Campbell Family Mental Health Research Institute (Toronto, ON, Canada). Pilocarpine hydrochloride (P6503), chloral hydrate (C8383), propylene glycol (P4347), DPX (06522) and Tween80 (P1754) were obtained from Sigma-Aldrich (St. Louis, MO, USA). Midazolam hydrochloride was purchased from Akorn Pharmaceuticals (Lake Forest, IL, USA). The α 5 antibody was from R&D Systems (#PPS027; Minneapolis, MN USA) and the anti-rabbit-HRP secondary was from Cell signaling (#7074; Danvers, MA, USA). All other antibodies were from Abcam (Cambridge, UK), including gephryin (ab181382) and

GAPDH (#9484). Dynabeads protein G magnetic beads (10003D), ECL substrate (32106), stripping buffer (21059), and coverslips (174942) were from ThermoFisher (Waltham, MA, USA). Gels (4569035) and membranes (1704158) were from Bio-Rad. All other chemicals or reagents were of either analytical or laboratory grade and purchased from various suppliers.

5. Conclusions

As with other subunit-selective compounds, the question arises of whether the biological effects that are observed are really mediated by the subunit for which the compound has some relative selectivity. Based on in vitro data, GL-II-73 displays significant selectivity for α 5GABA_ARs, but we acknowledge that we did not formally demonstrate that the effects observed with GL-II-73 are truly mediated by α 5GABA_ARs in this study. Future research could resolve this using transgenic approaches. Another interesting area of research would be to examine if the increased tonic inhibition cause by α 5-PAMs may attenuate seizure activity in the hippocampus [\[66\]](#page-11-9). Thus, α5-PAMs may provide relief for both epileptic and psychotic symptoms associated with TLE. Future studies should examine the potential antiseizure effects of α 5-PAMs, as GL-II-73 may be beneficial for treating multiple pathologies associated with TLE [\[33,](#page-10-1)[67\]](#page-11-10). As there are currently no viable treatment options for managing psychosis in epilepsy patients, this study provides promising preclinical evidence that GL-II-73 may be able to provide relief for those suffering from TLE with comorbid psychosis.

Author Contributions: A.M.M. designed the study, collected the data, wrote and edited the manuscript. D.J.L. designed the study, edited and approved the final manuscript. D.S. synthesized the drug. E.L.S., T.D.P. and J.M.C. edited and approved the final manuscript. All authors have read and agreed to the published version of the manuscript.

Funding: This work was supported by Merit Awards from the United States Department of Veterans Affairs, Biomedical Laboratory Research (BX004693 and BX004646 to D.J.L.) and Development Service and National Institutes of Health grants (DA054177, DA043204, AA029023 to J.C., and T32-NS082145 to A.M.M.).

Institutional Review Board Statement: The animal study protocol was approved by the UTHSCSA IACUC on 2022-01-20 (Approval code—20150106AR).

Informed Consent Statement: Not applicable.

Data Availability Statement: The data presented in this study are available on request from the corresponding author.

Conflicts of Interest: E.S., T.D.P and J.C. are co-inventors or listed on US patent applications that cover GABAergic ligands, including GL-II-73, and their use in brain disorders. E.S. is co-founder and CSO, and T.D.P is Director of Operation of DAMONA Pharmaceuticals, a biopharmaceutical company dedicated to treating cognitive deficits in brain disorders.

References

- 1. Butler, T.; Weisholtz, D.; Isenberg, N.; Harding, E.; Epstein, J.; Stern, E.; Silbersweig, D. Neuroimaging of frontal–limbic dysfunction in schizophrenia and epilepsy-related psychosis: Toward a convergent neurobiology. *Epilepsy Behav.* **2012**, *23*, 113–122. [\[CrossRef\]](https://doi.org/10.1016/j.yebeh.2011.11.004) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/22209327)
- 2. Irwin, L.G.; Fortune, D.G. Risk Factors for Psychosis Secondary to Temporal Lobe Epilepsy: A Systematic Review. *J. Neuropsychiatry Clin. Neurosci.* **2014**, *26*, 5–23. [\[CrossRef\]](https://doi.org/10.1176/appi.neuropsych.12120403) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/24515674)
- 3. Maguire, M.; Singh, J.; Marson, A. Epilepsy and psychosis: A practical approach. *Pract. Neurol.* **2018**, *18*, 106–114. [\[CrossRef\]](https://doi.org/10.1136/practneurol-2017-001775) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/29273616)
- 4. Sachdev, P.S. Alternating and Postictal Psychoses: Review and a Unifying Hypothesis. *Schizophr. Bull.* **2007**, *33*, 1029–1037. [\[CrossRef\]](https://doi.org/10.1093/schbul/sbm012)
- 5. Clancy, M.J.; Clarke, M.C.; Connor, D.J.; Cannon, M.; Cotter, D.R. The prevalence of psychosis in epilepsy; a systematic review and meta-analysis. *BMC Psychiatry* **2014**, *14*, 75. [\[CrossRef\]](https://doi.org/10.1186/1471-244X-14-75)
- 6. Slater, E.; Beard, A.W.; Glithero, E. The Schizophrenialike Psychoses of Epilepsy. *Br. J. Psychiatry J. Ment. Sci.* **1963**, *109*, 95–150. [\[CrossRef\]](https://doi.org/10.1192/bjp.109.458.95)
- 7. Hedges, D.; Jeppson, K.; Whitehead, P. Antipsychotic medication and seizures: A review. *Drugs Today Barc. Spain 1998* **2003**, *39*, 551–557. [\[CrossRef\]](https://doi.org/10.1358/dot.2003.39.7.799445)
- 8. Jeon, S.M.; Park, H.-Y.; Park, S.; Chung, U.S.; Kwon, J.-W. Association of Treatment with Antipsychotics, Antidepressants, or Both with Movement Disorders and Seizures among Children and Adolescents with Depression in Korea. *JAMA Netw. Open* **2022**, *5*, e227074. [\[CrossRef\]](https://doi.org/10.1001/jamanetworkopen.2022.7074)
- 9. Lieberman, J.A.; Stroup, T.S.; McEvoy, J.P.; Swartz, M.S.; Rosenheck, R.A.; Perkins, D.O.; Keefe, R.S.E.; Davis, S.M.; Davis, C.E.; Lebowitz, B.D.; et al. Effectiveness of Antipsychotic Drugs in Patients with Chronic Schizophrenia. *N. Engl. J. Med.* **2005**, *353*, 1209–1223. [\[CrossRef\]](https://doi.org/10.1056/NEJMoa051688)
- 10. Burnham, W.M. Why Are Complex Partial Seizures Intractable? In *Intractable Seizures: Diagnosis, Treatment, and Prevention*; Burnham, W.M., Carlen, P.L., Hwang, P.A., Eds.; Advances in Experimental Medicine and Biology; Springer: Boston, MA, USA, 2002; pp. 107–110. ISBN 978-1-4615-1335-3.
- 11. Racine, R.; Rose, P.A.; Burnham, W.M. Afterdischarge Thresholds and Kindling Rates in Dorsal and Ventral Hippocampus and Dentate Gyrus. *Can. J. Neurol. Sci. J. Can. Sci. Neurol.* **1977**, *4*, 273–278. [\[CrossRef\]](https://doi.org/10.1017/S0317167100025117)
- 12. Brooks-Kayal, A.R.; Shumate, M.D.; Jin, H.; Rikhter, T.Y.; Coulter, D.A. Selective changes in single cell GABAA receptor subunit expression and function in temporal lobe epilepsy. *Nat. Med.* **1998**, *4*, 1166–1172. [\[CrossRef\]](https://doi.org/10.1038/2661) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/9771750)
- 13. Wang, L.; Liu, Y.-H.; Huang, Y.-G.; Chen, L.-W. Time-Course of neuronal death in the mouse pilocarpine model of chronic epilepsy using fluoro-jade c staining. *Brain Res.* **2008**, *1241*, 157–167. [\[CrossRef\]](https://doi.org/10.1016/j.brainres.2008.07.097) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/18708038)
- 14. Heckers, S.; Konradi, C. Hippocampal neurons in schizophrenia. *J. Neural Transm. Vienna Austria* **1996**, *109*, 891–905. [\[CrossRef\]](https://doi.org/10.1007/s007020200073) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/12111476)
- 15. Malaspina, D.; Storer, S.; Furman, V.; Esser, P.; Printz, D.; Berman, A.; Lignelli, A.; Gorman, J.; Van Heertum, R. SPECT study of visual fixation in schizophrenia and comparison subjects. *Biol. Psychiatry* **1999**, *46*, 89–93. [\[CrossRef\]](https://doi.org/10.1016/S0006-3223(98)00306-0) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/10394477)
- 16. Schobel, S.A.; Lewandowski, N.M.; Corcoran, C.M.; Moore, H.; Brown, T.; Malaspina, D.; Small, S.A. Differential targeting of the CA1 subfield of the hippocampal formation by schizophrenia and related psychotic disorders. *Arch. Gen. Psychiatry* **2009**, *66*, 938–946. [\[CrossRef\]](https://doi.org/10.1001/archgenpsychiatry.2009.115) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/19736350)
- 17. Weiss, A.P.; Schacter, D.L.; Goff, D.C.; Rauch, S.L.; Alpert, N.M.; Fischman, A.J.; Heckers, S. Impaired hippocampal recruitment during normal modulation of memory performance in schizophrenia. *Biol. Psychiatry* **2003**, *53*, 48–55. [\[CrossRef\]](https://doi.org/10.1016/S0006-3223(02)01541-X)
- 18. Molina, V.; Reig, S.; Pascau, J.; Sanz, J.; Sarramea, F.; Gispert, J.D.; Luque, R.; Benito, C.; Palomo, T.; Desco, M. Anatomical and functional cerebral variables associated with basal symptoms but not risperidone response in minimally treated schizophrenia. *Psychiatry Res. Neuroimaging* **2003**, *124*, 163–175. [\[CrossRef\]](https://doi.org/10.1016/S0925-4927(03)00107-0) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/14623068)
- 19. Lewis, D.A.; Hashimoto, T.; Volk, D.W. Cortical inhibitory neurons and schizophrenia. *Nat. Rev. Neurosci.* **2005**, *6*, 312–324. [\[CrossRef\]](https://doi.org/10.1038/nrn1648)
- 20. Lodge, D.J.; Behrens, M.M.; Grace, A.A. A Loss of Parvalbumin-Containing Interneurons Is Associated with Diminished Oscillatory Activity in an Animal Model of Schizophrenia. *J. Neurosci.* **2009**, *29*, 2344–2354. [\[CrossRef\]](https://doi.org/10.1523/JNEUROSCI.5419-08.2009)
- 21. Lodge, D.J.; Grace, A.A. Aberrant hippocampal activity underlies the dopamine dysregulation in an animal model of schizophrenia. *J. Neurosci. Off. J. Soc. Neurosci.* **2007**, *27*, 11424–11430. [\[CrossRef\]](https://doi.org/10.1523/JNEUROSCI.2847-07.2007)
- 22. Lodge, D.J.; Grace, A.A. Hippocampal dysregulation of dopamine system function and the pathophysiology of schizophrenia. *Trends Pharmacol. Sci.* **2011**, *32*, 507–513. [\[CrossRef\]](https://doi.org/10.1016/j.tips.2011.05.001) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/21700346)
- 23. Perez, S.M.; Lodge, D.J. Convergent Inputs from the Hippocampus and Thalamus to the Nucleus Accumbens Regulate Dopamine Neuron Activity. *J. Neurosci.* **2018**, *38*, 10607–10618. [\[CrossRef\]](https://doi.org/10.1523/JNEUROSCI.2629-16.2018)
- 24. Laruelle, M.; Abi-Dargham, A. Dopamine as the wind of the psychotic fire: New evidence from brain imaging studies. *J. Psychopharmacol. Oxf. Engl.* **1999**, *13*, 358–371. [\[CrossRef\]](https://doi.org/10.1177/026988119901300405) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/10667612)
- 25. Abi-Dargham, A. Do we still believe in the dopamine hypothesis? New data bring new evidence. *Int. J. Neuropsychopharmacol.* **2004**, *7* (Suppl. 1), S1–S5. [\[CrossRef\]](https://doi.org/10.1017/S1461145704004110) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/14972078)
- 26. Donegan, J.J.; Tyson, J.A.; Branch, S.Y.; Beckstead, M.J.; Anderson, S.A.; Lodge, D.J. Stem cell-derived interneuron transplants as a treatment for schizophrenia: Preclinical validation in a rodent model. *Mol. Psychiatry* **2017**, *22*, 1492–1501. [\[CrossRef\]](https://doi.org/10.1038/mp.2016.121)
- 27. Donegan, J.J.; Boley, A.M.; Yamaguchi, J.; Toney, G.M.; Lodge, D.J. Modulation of extrasynaptic GABA a alpha 5 receptors in the ventral hippocampus normalizes physiological and behavioral deficits in a circuit specific manner. *Nat. Commun.* **2019**, *10*, 2819. [\[CrossRef\]](https://doi.org/10.1038/s41467-019-10800-1)
- 28. McCoy, A.M.; Prevot, T.D.; Mian, M.Y.; Cook, J.M.; Frazer, A.; Sibille, E.L.; Carreno, F.R.; Lodge, D.J. Positive allosteric modulation of α5-GABAA receptors reverses stress-induced alterations in dopamine system function and prepulse inhibition of startle. *Int. J. Neuropsychopharmacol.* **2022**, *25*, 688–698. [\[CrossRef\]](https://doi.org/10.1093/ijnp/pyac035)
- 29. Perez, S.M.; Lodge, D.J. Hippocampal interneuron transplants reverse aberrant dopamine system function and behavior in a rodent model of schizophrenia. *Mol. Psychiatry* **2013**, *18*, 1193–1198. [\[CrossRef\]](https://doi.org/10.1038/mp.2013.111)
- 30. Perez, S.M.; Shah, A.; Asher, A.; Lodge, D.J. Hippocampal Deep Brain Stimulation Reverses Physiological and Behavioral Deficits in a Rodent Model of Schizophrenia. *Int. J. Neuropsychopharmacol. Off. Sci. J. Coll. Int. Neuropsychopharmacol. CINP* **2013**, *16*, 1331–1339. [\[CrossRef\]](https://doi.org/10.1017/S1461145712001344)
- 31. Fritschy, J.-M.; Mohler, H. GABAA-Receptor heterogeneity in the adult rat brain: Differential regional and cellular distribution of seven major subunits. *J. Comp. Neurol.* **1995**, *359*, 154–194. [\[CrossRef\]](https://doi.org/10.1002/cne.903590111)
- 32. Sur, C.; Fresu, L.; Howell, O.; McKernan, R.M.; Atack, J.R. Autoradiographic Localization of Alpha5 Subunit-Containing GABAA Receptors in Rat Brain. *Brain Res.* **1999**, *822*, 265–270. [\[CrossRef\]](https://doi.org/10.1016/S0006-8993(99)01152-X)
- 33. Prevot, T.D.; Li, G.; Vidojevic, A.; Misquitta, K.A.; Fee, C.; Santrac, A.; Knutson, D.E.; Stephen, M.R.; Kodali, R.; Zahn, N.M.; et al. Novel Benzodiazepine-Like Ligands with Various Anxiolytic, Antidepressant, or Pro-Cognitive Profiles. *Mol. Neuropsychiatry* **2019**, *5*, 84–97. [\[CrossRef\]](https://doi.org/10.1159/000496086) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/31192221)
- 34. Prevot, T.D.; Sumitomo, A.; Tomoda, T.; Knutson, D.E.; Li, G.; Mondal, P.; Banasr, M.; Cook, J.M.; Sibille, E. Reversal of Age-Related Neuronal Atrophy by A5-GABAA Receptor Positive Allosteric Modulation. *Cereb. Cortex N. Y. N 1991* **2021**, *31*, 1395–1408. [\[CrossRef\]](https://doi.org/10.1093/cercor/bhaa310) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/33068001)
- 35. Perez, S.M.; McCoy, A.M.; Prevot, T.D.; Mian, Y.; Carreno, F.R.; Frazer, A.; Cook, J.M.; Sibille, E.; Lodge, D.J. Hippocampal α5-GABAA receptors modulate dopamine neuron activity in the rat ventral tegmental area. *Biol. Psychiatry Glob. Open Sci.* **2022**, *3*, 78–86. [\[CrossRef\]](https://doi.org/10.1016/j.bpsgos.2021.12.010)
- 36. McCoy, A.M.; Prevot, T.D.; Mian, M.Y.; Sharmin, D.; Ahmad, A.; Cook, J.M.; Sibille, E.L.; Lodge, D.J. Extrasynaptic localization is essential for α5GABAA receptor modulation of dopamine system function. *BioRxiv* **2023**. [\[CrossRef\]](https://doi.org/10.1101/2023.07.12.548744)
- 37. Davenport, C.M.; Rajappa, R.; Katchan, L.; Taylor, C.R.; Tsai, M.-C.; Smith, C.M.; de Jong, J.W.; Arnold, D.B.; Lammel, S.; Kramer, R.H. Relocation of an Extrasynaptic GABAA Receptor to Inhibitory Synapses Freezes Excitatory Synaptic Strength and Preserves Memory. *Neuron* **2021**, *109*, 123–134.e4. [\[CrossRef\]](https://doi.org/10.1016/j.neuron.2020.09.037) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/33096025)
- 38. Cavalheiro, E.A. The Pilocarpine Model of Epilepsy. *Ital. J. Neurol. Sci.* **1995**, *16*, 33–37. [\[CrossRef\]](https://doi.org/10.1007/BF02229072)
- 39. Curia, G.; Longo, D.; Biagini, G.; Jones, R.S.G.; Avoli, M. The pilocarpine model of temporal lobe epilepsy. *J. Neurosci. Methods* **2008**, *172*, 143–157. [\[CrossRef\]](https://doi.org/10.1016/j.jneumeth.2008.04.019)
- 40. Cifelli, P.; Grace, A.A. Pilocarpine Induced Temporal Lobe Epilepsy in the Rat Is Associated with Increased Dopamine Neuron Activity. *Int. J. Neuropsychopharmacol. Off. Sci. J. Coll. Int. Neuropsychopharmacol. CINP* **2012**, *15*, 957–964. [\[CrossRef\]](https://doi.org/10.1017/S1461145711001106)
- 41. Paxinos, G.; Watson, C. *The Rat Brain in Stereotaxic Coordinates*, 6th ed.; Elsevier Academic Press: Amsterdam, The Netherlands, 2007; ISBN 9780123919496.
- 42. Howes, O.D.; Montgomery, A.J.; Asselin, M.-C.; Murray, R.M.; Valli, I.; Tabraham, P.; Bramon-Bosch, E.; Valmaggia, L.; Johns, L.; Broome, M.; et al. Elevated striatal dopamine function linked to prodromal signs of schizophrenia. *Arch. Gen. Psychiatry* **2009**, *66*, 13–20. [\[CrossRef\]](https://doi.org/10.1001/archgenpsychiatry.2008.514)
- 43. Grace, A.A.; Gomes, F.V. The Circuitry of Dopamine System Regulation and Its Disruption in Schizophrenia: Insights Into Treatment and Prevention. *Schizophr. Bull.* **2019**, *45*, 148–157. [\[CrossRef\]](https://doi.org/10.1093/schbul/sbx199) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/29385549)
- 44. Perez, S.M.; Elam, H.B.; Lodge, D.J. Increased Presynaptic Dopamine Synthesis Capacity Is Associated with Aberrant Dopamine Neuron Activity in the Methylazoxymethanol Acetate Rodent Model Used to Study Schizophrenia-Related Pathologies. *Schizophr. Bull. Open* **2022**, *3*, sgac067. [\[CrossRef\]](https://doi.org/10.1093/schizbullopen/sgac067) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/36387971)
- 45. Elam, H.B.; Perez, S.M.; Donegan, J.J.; Lodge, D.J. Orexin receptor antagonists reverse aberrant dopamine neuron activity and related behaviors in a rodent model of stress-induced psychosis. *Transl. Psychiatry* **2021**, *11*, 114. [\[CrossRef\]](https://doi.org/10.1038/s41398-021-01235-8) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/33558469)
- 46. Choy, M.; Wells, J.A.; Thomas, D.L.; Gadian, D.G.; Scott, R.C.; Lythgoe, M.F. Cerebral blood flow changes during pilocarpineinduced status epilepticus activity in the rat hippocampus. *Exp. Neurol.* **2010**, *225*, 196–201. [\[CrossRef\]](https://doi.org/10.1016/j.expneurol.2010.06.015)
- 47. Houser, C.R.; Esclapez, M. Downregulation of the α5 subunit of the GABAA receptor in the pilocarpine model of temporal lobe epilepsy. *Hippocampus* **2003**, *13*, 633–645. [\[CrossRef\]](https://doi.org/10.1002/hipo.10108)
- 48. Schwarzer, C.; Tsunashima, K.; Wanzenböck, C.; Fuchs, K.; Sieghart, W.; Sperk, G. GABAA Receptor Subunits in the Rat Hippocampus II: Altered Distribution in Kainic Acid-Induced Temporal Lobe Epilepsy. *Neuroscience* **1997**, *80*, 1001–1017. [\[CrossRef\]](https://doi.org/10.1016/S0306-4522(97)00145-0)
- 49. Tsunashima, K.; Schwarzer, C.; Kirchmair, E.; Sieghart, W.; Sperk, G. GABA(A) receptor subunits in the rat hippocampus III: Altered messenger RNA expression in kainic acid-induced epilepsy. *Neuroscience* **1997**, *80*, 1019–1032. [\[CrossRef\]](https://doi.org/10.1016/S0306-4522(97)00144-9)
- 50. Zhang, Z.J.; Reynolds, G.P. A Selective decrease in the relative density of parvalbumin-immunoreactive neurons in the hippocampus in schizophrenia. *Schizophr. Res.* **2002**, *55*, 1–10. [\[CrossRef\]](https://doi.org/10.1016/S0920-9964(01)00188-8)
- 51. Furtado, M.d.A.; Braga, G.K.; Oliveira, J.A.C.; Del Vecchio, F.; Garcia-Cairasco, N. Behavioral, morphologic, and electroencephalographic evaluation of seizures induced by intrahippocampal microinjection of pilocarpine. *Epilepsia* **2002**, *43* (Suppl. 5), 37–39. [\[CrossRef\]](https://doi.org/10.1046/j.1528-1157.43.s.5.41.x)
- 52. Ando, N.; Morimoto, K.; Watanabe, T.; Ninomiya, T.; Suwaki, H. Enhancement of Central Dopaminergic Activity in the Kainate Model of Temporal Lobe Epilepsy: Implication for the Mechanism of Epileptic Psychosis. *Neuropsychopharmacology* **2004**, *29*, 1251–1258. [\[CrossRef\]](https://doi.org/10.1038/sj.npp.1300427)
- 53. Tamminga, C.A.; Stan, A.D.; Wagner, A.D. The hippocampal formation in schizophrenia. *Am. J. Psychiatry* **2010**, *167*, 1178–1193. [\[CrossRef\]](https://doi.org/10.1176/appi.ajp.2010.09081187) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/20810471)
- 54. Floresco, S.B.; Todd, C.L.; Grace, A.A. Glutamatergic Afferents from the Hippocampus to the Nucleus Accumbens Regulate Activity of Ventral Tegmental Area Dopamine Neurons. *J. Neurosci.* **2001**, *21*, 4915–4922. [\[CrossRef\]](https://doi.org/10.1523/JNEUROSCI.21-13-04915.2001) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/11425919)
- 55. Floresco, S.B.; West, A.R.; Ash, B.; Moore, H.; Grace, A.A. Afferent modulation of dopamine neuron firing differentially regulates tonic and phasic dopamine transmission. *Nat. Neurosci.* **2003**, *6*, 968–973. [\[CrossRef\]](https://doi.org/10.1038/nn1103) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/12897785)
- 56. Lodge, D.J.; Grace, A.A. The laterodorsal tegmentum is essential for burst firing of ventral tegmental area dopamine neurons. *Proc. Natl. Acad. Sci. USA* **2006**, *103*, 5167–5172. [\[CrossRef\]](https://doi.org/10.1073/pnas.0510715103)
- 57. Aguilar, D.D.; Giuffrida, A.; Lodge, D.J. Adolescent Synthetic Cannabinoid Exposure Produces Enduring Changes in Dopamine Neuron Activity in a Rodent Model of Schizophrenia Susceptibility. *Int. J. Neuropsychopharmacol.* **2018**, *21*, 393–403. [\[CrossRef\]](https://doi.org/10.1093/ijnp/pyy003)
- 58. Gauillard, J.; Cheref, S.; Vacherontrystram, M.N.; Martin, J.C. Chloral hydrate: A hypnotic best forgotten? *L'Encephale* **2002**, *28.3 Pt 1*, 200–204.
- 59. Peoples, R.W.; Weight, F.F. Trichloroethanol potentiation of gamma-aminobutyric acid-activated chloride current in mouse hippocampal neurones. *Br. J. Pharmacol.* **1994**, *113*, 555–563. [\[CrossRef\]](https://doi.org/10.1111/j.1476-5381.1994.tb17025.x)
- 60. Davies, J.A. Mechanisms of action of antiepileptic drugs. *Seizure* **1995**, *4*, 267–271. [\[CrossRef\]](https://doi.org/10.1016/S1059-1311(95)80003-4)
- 61. Gill, K.M.; Lodge, D.J.; Cook, J.M.; Aras, S.; Grace, A.A. A Novel A5GABAAR-Positive Allosteric Modulator Reverses Hyperactivation of the Dopamine System in the MAM Model of Schizophrenia. *Neuropsychopharmacology* **2011**, *36*, 1903–1911. [\[CrossRef\]](https://doi.org/10.1038/npp.2011.76)
- 62. Nishimura, T.; Schwarzer, C.; Gasser, E.; Kato, N.; Vezzani, A.; Sperk, G. Altered expression of GABA(A) and GABA(B) receptor subunit mRNAs in the hippocampus after kindling and electrically induced status epilepticus. *Neuroscience* **2005**, *134*, 691–704. [\[CrossRef\]](https://doi.org/10.1016/j.neuroscience.2005.04.013)
- 63. Racine, R.J. Modification of seizure activity by electrical stimulation: II. Motor seizure. *Electroencephalogr. Clin. Neurophysiol.* **1972**, *32*, 281–294. [\[CrossRef\]](https://doi.org/10.1016/0013-4694(72)90177-0) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/4110397)
- 64. Grace, A.A.; Bunney, B.S. Intracellular and Extracellular Electrophysiology of Nigral Dopaminergic Neurons—1. Identification and Characterization. *Neuroscience* **1983**, *10*, 301–315. [\[CrossRef\]](https://doi.org/10.1016/0306-4522(83)90135-5)
- 65. Ungless, M.A.; Grace, A.A. Are you or aren't you? Challenges associated with physiologically identifying dopamine neurons. *Trends Neurosci.* **2012**, *35*, 422–430. [\[CrossRef\]](https://doi.org/10.1016/j.tins.2012.02.003) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/22459161)
- 66. Schipper, S.; Aalbers, M.W.; Rijkers, K.; Swijsen, A.; Rigo, J.M.; Hoogland, G.; Vles, J.S.H. Tonic GABAA Receptors as Potential Target for the Treatment of Temporal Lobe Epilepsy. *Mol. Neurobiol.* **2016**, *53*, 5252–5265. [\[CrossRef\]](https://doi.org/10.1007/s12035-015-9423-8) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/26409480)
- 67. Harden, C.L. Depression and Anxiety in Epilepsy Patients. *Epilepsy Behav. EB* **2002**, *3*, 296. [\[CrossRef\]](https://doi.org/10.1016/S1525-5050(02)00018-5)

Disclaimer/Publisher's Note: The statements, opinions and data contained in all publications are solely those of the individual author(s) and contributor(s) and not of MDPI and/or the editor(s). MDPI and/or the editor(s) disclaim responsibility for any injury to people or property resulting from any ideas, methods, instructions or products referred to in the content.