



Near Infrared Photoimmunotherapy: A Review of Recent Progress and Their Target Molecules for Cancer Therapy

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Abstract: Near infrared photoimmunotherapy (NIR-PIT) is a newly developed molecular targeted cancer treatment, which selectively kills cancer cells or immune-regulatory cells and induces therapeutic host immune responses by administrating a cancer targeting moiety conjugated with IRdye700. The local exposure to near-infrared (NIR) light causes a photo-induced ligand release reaction, which causes damage to the target cell, resulting in immunogenic cell death (ICD) with little or no side effect to the surrounding normal cells. Moreover, NIR-PIT can generate an immune response in distant metastases and inhibit further cancer attack by combing cancer cells targeting NIR-PIT and immune regulatory cells targeting NIR-PIT or other cancer treatment modalities. Several recent improvements in NIR-PIT have been explored such as catheter-driven NIR light delivery, real-time monitoring of cancer, and the development of new target molecule, leading to NIR-PIT, their mechanism and design strategies for cancer treatment. Furthermore, the overall possible targeting molecules for NIR-PIT with their application for cancer treatment are briefly summarised.

Keywords: cancer therapy; NIR-PIT; ICD; antitumor host immunity; target molecule

1. Introduction

Cancer is one of the top global health problems. The primary cancer treatments are surgery, chemotherapy, and/or radiation, which are well known to cause considerable damage to normal cells. Over the last three decades, several newly developed modalities have been developed for cancer treatment including immunotherapy, targeted therapy, photoimmunotherapy (PIT), photothermal therapy, and photodynamic therapy (PDT) [1–4]. These treatment modalities aim to mitigate the cancer burden with fewer side effects. PDT is a medical treatment that mostly depends on the generation of cytotoxic singlet oxygen and other reactive oxygen species (ROS) that directly destroy tumor cells via the activation of a photosensitizer (Ps) by light irradiation at a corresponding wavelength [5–7]. Over the last 40 years, PDT has been extensively used to treat cancer cells, and several approaches have been developed to increase PDT activity and specificity. One of these approaches is the use of nanoparticle as a Ps delivery vehicle. Nanoparticle-based PDT takes advantage of the enhanced permeability and retention (EPR) effect, and thus has the potential for better Ps accumulation into tumor tissues and less side effects in the surrounding cells compared with conventional PDT [6]. Moreover, the incapability of destroying deep tumors using PDT can be overcome by applying fiber optic inserted interstitial photodynamic therapy (iPDT) [8]. PIT emerges as a novel promising approach, which is a combination of phototherapy and immunotherapy that leads to effective and specific eradication of primary tumors and distant metastasis, and that inhibits cancer recurrence [9]. Similar to PDT, PIT approaches kill the treated cells through the local activation of Ps with an



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Copyright: © 2023 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (https:// creativecommons.org/licenses/by/ 4.0/). appropriate light wavelength, but they have the advantages of a higher efficacy and less side effects due to the properties of the targeting moiety of PIT agents such as antibodies, peptides, and small ligands.

The fastest growing PIT approach in this field is the NIR-PIT approach. The cell death mechanism in NIR-PIT is distinctive, with features of cell swelling, the formation of bleb, and membrane rupturing, resulting in vigorous anti-tumor immune responses [10–12]. It has great potential to selectively and locally destroy cancer cells, inducing immunogenic cell death (ICD) and triggering an antitumor immune response [13]. This is mainly due to the release of damage-associated molecular patterns (DAMPs) and tumor-associated antigens (TAAs) after PIT, which activate antitumor immune responses [14,15]. However, this response is insufficient to treat the metastatic tumor as the immunosuppression of the tumor microenvironment could limit the antitumor immunity [3,16]. To induce the antitumor immune response, a combination of PIT with immune checkpoint inhibitors and immunoadjuvants were applied as a synergistic treatment against tumor cells [16]. NIR-PIT has advantages over conventional PDT as it does not depend on ROS generation, has a higher efficacy and little or no side effects on the surrounding cells [17].

For these reasons, NIR-PIT has gained encouraging success both in preclinical and clinical applications for different cancers [18]. Different targeting moieties were employed in NIR-PIT to enhance the tumor immunogenicity, targeting capability, stability, and flexibility of NIR-PIT agents [9,19–21].

2. Antibody and Antibody Mimetic-Based NIR-PIT

Tumor cell growth and progression are associated with either the up or down regulation of certain pathways, which in some cases requires an increased expression of related receptors. Therefore, targeting these receptors using specific ligands such as antibodies, their fragments, and mimetics can enhance the tumor cell selectivity and spare the surrounding healthy tissue [22]. In NIR-PIT, monoclonal antibodies and antibody derivatives such as fragment antigen-binding (Fab), $F(ab')_2$, single chain antibody fragment (scFv), diabody, minibody, and nanobody, as well as antibody mimetics, were used as delivery vehicles for the IRdye700 producing photo-immunoconjugates [23–26]. Several studies have demonstrated the success of photo-immunoconjugates using antibodies targeting receptors overexpressed in tumor cells [27].

Most clinical and preclinical studies involve cetuximab and panitumumab conjugated with IRdye700 (cetuximab-IRdye700 and panitumumab-IRdye700) for targeting the membrane receptor epidermal growth factor receptor (EGFR) overexpressing cancer cells [13,28]. After NIR light irradiation, cetuximab-NIR-PIT and panitumumab-NIR-PIT agents can selectively kill target cells and elicit host anti-tumor immune responses [28–31].

For homogeneous micro distribution of antibody-conjugates, a cocktail of panitumumab and basiliximab conjugated IRdye700 was used for NIR-PIT that significantly reduced tumor growth and prolonged the survival of tumor-bearing mice compared with the single antibody-IRdye700 conjugate [32]. When combining NIR-PIT reagents, panitumumab-IRdye700 and anti-CD25-F(ab')₂-IRdye700 demonstrated the effective inhibition of tumor growth and an enhanced antitumor immune response by depleting regulatory T (T_{regs}) cells from the tumor microenvironment [33]. Furthermore, conjugating IRdye700 to pertuzumab and trastuzumab that targets human epidermal growth factor receptor 2 (Her2)-expressing cancer cells revealed antitumor effects after NIR light irradiation [25,34–37]. In addition, other Ps, for instance porphyrin and chlorin e6 conjugated trastuzumab, were also used to eliminate Her2 positive cancer cells with improvements in specific tissue access and penetration [38,39].

Besides targeting the EGFR and Her2, several antibodies targeting different TAAs have been used to generate NIR-PIT reagents. These include, for example, rituximab (anti-CD20) [40], anti-prostate-specific membrane antigen (PMSA) antibody [41], anti-epithelial cell adhesion molecule (EpCAM) antibody [42,43], anti-CD44 antibody [44], anti-CD47 antibody [45,46], and anti-tumor-associated calcium signal transducer 2 (TROP2) antibody [13,47,48]. Moreover, AC133 monoclonal antibody (mAb) [49] and anti-CD44 anti-

tibody [44,50–52] conjugated with IRdye700 agents were used for NIR-PIT against different cancers by targeting cancer stem cell marker AC133 and CD44, respectively.

In addition to the full-length mAb, their fragments have also been used to develop NIR-PIT reagents because of their smaller molecular size, more efficient tumor penetration, and faster tissue clearance. These fragments, such scFv and Fab, are lacking the antibody Fc region, which is responsible for initiating antibody-dependent cellular cytotoxicity (ADCC) and complement-dependent cytotoxicity (CDC) immune responses. For example, scFvs against EGFR [23,53], EpCAM, and chondroitin sulphate proteoglycan 4 (CSPG4) [54] were used to generate NIR-PIT agents. Furthermore, Sato et al. demonstrated that conjugating IRdye700 to $F(ab')_2$ targeting CD25 can selectively eliminate tumor-infiltrating T_{regs} [55]. A recent study reported that combining anti-CD44 antibody and anti-CD25 antibody conjugated with IRdye700 led to significant tumor growth inhibition and a prolonged survival rate [44]. Anti-CD25-IgG or its anti-CD25-F(ab')₂ derivative [44,55,56] and anticytotoxic T-lymphocyte-associated protein 4 (CTLA4) antibody [57] conjugated NIR-PIT agents can selectively eliminate the local Tregs only within the tumor microenvironment (TME), resulting in upregulation of the anti-tumor immune responses [47]. The effectiveness of NIR-PIT was demonstrated by using $F(ab')_2$ targeting programmed death-ligand 1 (PD-L1) and CD25 to generate anti-PD-L1-F(ab')2-IRdye700 and anti-CD25-F(ab')2-IRdye700, respectively [57,58]. Avelumab (human anti-PD-L1 mAb) [59,60] and F(ab')₂ fragments of the anti-PD-L1 antibody [60] conjugated with IRdye700 show significant therapeutic effects against different cancers.

A minibody named MS5 was generated by fusing its scFv to the human IgG1 Fc domain [61]. As MS5 can bind to solid and blood cancer cells, IRdye700 was conjugated to the MS5 and used for targeting various cancer cells. MS5-IRdye700 is able to kill target cells in both monolayer and tumor spheroid cultures by inducing the immunogenic cell death pathway [62]. Moreover, NIR-PIT targeting PSMA was generated by conjugating IRdye700 with anti-PMSA diabody and minibody. These PSMA-diabody-IRdye700 and PSMA-minibody-IRdye700 show significant inhibition of tumor growth and enhance survival rate [63]. Nanobodies (12–15 kDa) are antibody-fragments derived from heavy-chain antibodies, which are highly soluble and physically stable [64]. Nanobody targeting G protein-coupled receptor (GPCR) was site-specifically conjugated to IRdye700. This nanobody-IRdye700 selectively kills US28-expressing glioblastoma cells [65,66].

In addition, affibody molecules, which are small (6.5-kDa) affinity proteins that can bind protein targets with a high affinity and selectivity, were used to generate NIR-PIT [66,67]. Because of its small size, the affibody has rapid clearance, and good tumor penetration, and can even pass the blood-brain barrier [68]. Recently, the Her2 affibody $(Z_{Her2:2395})$ -IRdye700 was shown to have selective cell death and trigger the release of all hallmarks of immunogenic cell death, resulting in the maturation of dendritic cell (DC) and the augmentation of anticancer immunity [69]. Another study demonstrated that IRdye700-Her2 affibody induced the necrotic cell death of Her2-overexpressing breast cancer cells [70]. In addition to the Her2 affibody, a dimeric platelet-derived growth factor receptor β (PDGFR β) affibody ($Z_{PDGFR\beta}$) was conjugated with IRdye700 and showed the specific binding of PDGFRB overexpressed pericytes of many types of tumors. After NIR light irradiation, Z_{PDGFR}-IRdye700 specifically kills pericytes, resulting in damage to the tumor blood vessels, thereby inducing tumor destruction [71]. Furthermore, EGFR-specific affibody molecule ($Z_{EGFR:03115}$) conjugated with IRdye700 shows brain tumor destruction by inducing ICD and turning an immunosuppressive tumor environment into an immunevulnerable one [72,73]. The above-mentioned studies suggest that affibody-based NIR-PIT could be an attractive alternative to mAb-based NIR-PIT.

3. Peptide and Small Ligand-Based NIR-PIT

Besides antibodies, short peptide targeting TAAs were used to deliver IRdye700 specifically to the target cells. For example, IRdye700 was covalently conjugated to the arginine–glycine–aspartic acid (RGD) peptide linked with human serum albumin to target

the integrins, which are highly expressed in several cancers. The nanoconjugate shows effective cancer-specific delivery and massive cancer cell killing [74]. The nanoconjugate is further developed by conjugating IRdye700 with the RGD peptide to 8-arm polyethylene glycol (PEG). The RGD–8PEG–IRdye700 demonstrates significant cancer cell reduction after light irradiation [75]. In addition, IRdye700 was conjugated to multiple cyclic RGD peptides (cRGD) to obtain better accumulation into the target sites with an increased number of cyclic RGD. The 15 cyclic RGD peptide polymer revealed significant inhibition of the tumor growth [76].

Low-molecular-weight (LMW) ligands are used also for targeting cancer antigens because of their efficient tumor penetration, reduced immunogenicity, and lack of Fcmediated side effects [77,78]. A recent study demonstrated that the LMW ligand, IRdye700, targeting PSMA decreased cell viability via different cytotoxic mechanisms [79].

4. Therapeutic Mechanism of NIR-PIT

4.1. Physicochemical Therapeutic Activity

NIR-PIT induces cell death after NIR light treatment, which activates the photochemical ligand reaction that releases the hydrophobic side chain of IRdye700, which in turn makes the remaining molecule hydrophobic. This photochemical reaction changes the NIR-PIT bound to the cell membrane by forming water insoluble aggregates of NIR-PIT or a Z-stack multimer of silicon-phthalocyanine IRdye700 rings [80]. The reduction in the cell membrane integrity of the NIR-PIT-antigen complex can damage the target proteins, which causes immediate swelling of the cells and subsequently releases the intracellular materials [10,12,17,81]. Moreover, it is reported that NIR-PIT agents are aggregated on the plasma membrane and lead to necrotic cell death. Internalized NIR-PIT agents also have cytotoxic effects following NIR light irradiation. It can induce necrotic cell death via ROS generation, which causes significant leakage of the lysosomal contents into the cytosol (Figure 1). Nevertheless, cytotoxicity by NIR-PIT is weaker in lysosomes than in the plasma membranes [82].



Figure 1. Mechanism of the NIR-PIT approach and its immunological consequences.

4.2. ICD and Anti-Tumor Immune Augmentation

The chemical and physical damages associated with NIR-PIT lead to rapid disruption of the cell membrane, which is a characteristic feature of ICD. ICD is a local immune reaction initiated by releasing cancer antigens from the dying cancer cells [17,30]. NIR-PIT releases DAMPs hallmarks, such as calreticulin (CRT), high-mobility group box 1 (HMGB1), adenosine triphosphate (ATP), heat shock protein (Hsp) 70, and Hsp 90 [25,51]. These DAMPs markers are responsible for the maturation of DCs, which prime the naive CD8⁺

T cells. Primed CD8⁺ T cells are then proliferated due to the rapid release of multiple neoantigens and have the ability to attack the residual cancer cells (Figure 1). The induced immune cells could induce systemic anticancer immune response via their migration throughout the body and by attacking distant metastatic sites. In addition, host immunity might be enhanced by NIR-PIT targeting immune suppressor cells, resulting in the selective depletion of inhibitory immune cells [47] (Figure 1).

4.3. Super-Enhanced Permeability and Retention (SUPR)

NIR-PIT selectively kills the tumor cells without affecting healthy cells by targeting the overexpressed antigen on the target cells. The light treatment causes no damage to surrounding normal cells that lack the targeted antigen [25]. It is reported that the repeated administration of NIR-PIT and NIR light suppress residual cell growth and recurrence attacks [83]. After applying NIR light, the NIR-PIT bound perivascular cells undergo necrosis, generating a space between the vessels and the remaining tumor mass wall, which enhances the vascular permeability of the nanosized drug in the treated tumor bed. The nanotherapeutic agents can remain in the tumor bed for several days. This SUPR of NIR-PIT enables enhanced delivery of nanodrugs into the tumor bed. Several recent studies have shown that the therapeutic effect is more efficient when a combination of NIR-PIT and anticancer nano drugs are applied compared with corresponding therapies individually [33,44].

5. Designing Strategies for NIR-PIT

In general, NIR-PIT is generated by conjugating IRdye700 with tumor-specific ligands. Various strategies are reported for binding antibodies to IRdye700; however, the direct conjugation methods via lysine or cysteine residues of the antibody are the most commonly used methods [13]. These methods, also known as randomly conjugation methods, are associated with generating heterogeneous NIR-PIT agent populations resulting in an unufied therapeutic and safety profile of NIR-PIT [84]. Site-specific conjugation methods are applied to generate homogeneous NIR-PIT. One promising approach is based on exploiting the simple, controlled, and robust site-specific conjugation properties of the SNAP-tag conjugation method. Here, several scFv molecules were genetically fused with the SNAP-tag protein to allow for the conjugation of bengylguanine (BG) modified IRdye700 [54,85,86]. This approach generates highly homogenous NIR-PIT agents with unified pharmacokinetic properties. This study demonstrates the potent phototherapeutic activities of NIR-PIT against breast cancer, ovarian cancer, and skin cancer cells in vitro [23,53,54].

6. NIR Light Delivery Method for NIR-PIT

The delivery of NIR light to the target tissue is very crucial in order to achieve effective therapeutic activities. The penetration depth of NIR light into the tissue is approximately 2 cm from the surface [47]. NIR light can be applied with a conventional extracorporeal apparatus that has a frontal diffuser to the tumor site, for instance in the mouth and skin [87]. Nagaya et al. (2018) first described the efficacy of a fiber optic diffuser under endoscopic guidance that could deliver NIR light into the deeply located tumor, such as peritoneum and thorax tumors [88,89]. Recently, a novel catheter mounted with light emitting diodes (LEDs) was developed to overcome the limitations of conventional external irradiation devices and endoscope diffusers. This catheter has the ability of deep insertion and irradiation, nonkinking properties, and a temperature sensor that can assist in avoiding thermal burn. By using this catheter, tumor growth is significantly reduced in cholangiocarcinoma xenografts in mice treated with NIR-PIT [90]. More recently, an endovascular-therapy-based light illumination technology (ET-BLIT) was developed to provide deep light irradiation within the body. In ET-BLIT, a single lumen catheter system was used that contains a tip partial transparent catheter with a transparent distal end attached to the thermocouple head and an optical light diffuser selective for lateral light irradiation. By using ET-BLIT, the

NIR light can reach the liver and kidneys without damaging the blood vessel or other side effects [91].

7. Monitoring the Therapeutic Effects of NIR-PIT

Real-time monitoring of the tumor accumulation, therapeutic response, and appropriate NIR light irradiation in NIR-PIT is important for precise treatment [47,92]. Several imaging methods are used to evaluate the effectiveness of NIR-PIT directly after treatment. Bioluminescence imaging (BLI) can be used preclinically to monitor the efficacy of NIR-PIT [93], but clinically, tumor cells cannot be transfected with the luciferase gene in advance [92]. One of the imaging methods during NIR-PIT is IRdye700 fluorescence signal of IRdye700 disappears after NIR light irradiation at 690 nm because the photochemical ligand-release reaction causes the precipitation of conjugated proteins [17,80,92]. As the glucose uptake of cancer cells increases dramatically, 18F-fluorodeoxyglucose positron emission tomography (18F-FDG PET) imaging provides early metabolic changes in the tumors, serving as an excellent method for evaluating the immediate treatment success in NIR-PIT-treated tumors. The 18F-FDG PET can be sustained for at least 24 h [94].

In addition to BLI and fluorescence imaging, optical coherence tomography (OCT) is used as a biomedical imaging tool that reveals dramatic hemodynamic changes in tumor vessels during NIR-PIT. This imaging system shows a significant difference in treated tumors compared with untreated tumors [95]. Moreover, the micro-distribution of the NIR-PIT agent from the tumor surface to the deep tumor during and after treatment and its therapeutic effects is monitored using a two-channel fluorescence fiber imaging system and two-photon microscopy with and without a microprism [96].

Recently, a customized camera system (LIGHTVISION) was designed to detect indocyanine green, and it can also detect the fluorescence arising from IRdye700 during NIR-PIT, which is implemented for intraoperative imaging of NIR-PIT. This camera system can monitor the NIR-PIT in real-time at wavelengths of 830 nm, which is far from the intense laser excitation light at 690 nm [97]. In addition, magnetic resonance imaging (MRI) is a widely used imaging modality, which can also be used to detect early therapeutic effects after NIR-PIT. The tumoricidal effects and hemodynamic changes induced by NIR-PIT can be monitored by ¹³C MRI, BOLD MRI and photoacoustic imaging [92,98].

8. Targeting Molecules for NIR-PIT and Their Application for Cancer Treatment

In the past few years, many NIR-PIT agents were developed for targeting different types of cancers. Initially, NIR-PIT was developed for targeting EGFR and Her2, and then expanded to target diverse cell surface proteins (such as EpCAM, CD44, CD47, CD25, PD-L1, and CTLA4) by using th mAb, antibody fragment, and nanobody. However, NIR-PIT can be applied to any cancer or regulatory cell in the tumor microenvironment if the available tumor antigens are overexpressed [47]. Here, we briefly summarized recently developed NIR-PIT agents (Table 1), as well as their clinical and preclinical applications.

Cancer Type	Target Molecule	Photo-Immunoconjugates	Therapeutic Outcome	References
Head and neck cancer, skin cancer, lung cancer, breast cancer, uterine cervical cancer, glioblastoma, bladder cancer	EGFR	* Cetuximab-IRdye700 * Cetuximab-IRdye700 + anti-PD-L1 scFv-425-SNAP-IRdye700 Affibody (Z _{EGFR:03115})-IRdye700 Panitumab-IRdye700 Panitumab-IRdye700 + Trastuzumab-IRdye700 Panitumab-IRdye700 + anti-CD25-F(ab') ₂ -IRdye700	TVR, ProS, AIA CCE CCE TVR, ProS TVR TVR, ProS, AIA, IMD	[19,30] [19] [53] [72] [99] [100] [33]
Gastric cancer, breast cancer, lung cancer	Her2	Trastuzumab-IRdye700 Trastuzumab-emtansine-IRdye700 Affibody (Z _{Her2-2395)} -IRdye700	TVR, ProS Cytotoxic photo-bystander effect TVR, AIA	[25,36,101–104] [69]
Colon cancer, lung cancer, head and neck cancer, oral cancer	CD44	Anti-CD44-IRdye700 + Interleukin-15 Anti-CD44-IRdye700 + anti-CTLA4-mAb Anti-CD44-IRdye700 + anti-PD-1	TVR, ProS, AIA TVR, ProS TVR, ProS, AIA	[105] [106] [51,52]
Brain cancer	CD133	AC133 mAb-IRdye700	TVR, ProS	[49]
Esophageal cancer	CAFs	Anti-FAP-IRdye700 + 5-fluorouracil (FU)	CCE	[107]
Prostate cancer	PSMA	Anti-PSMA mAb-IRdye700	TVR, ProS	[41]
Colon cancer, biliary tract cancer, pancreatic cancer	CEA	Anti-human CEA mAb-IRdye700	TVR	[108,109]
Lymphoma	CD20	Rituximab-IRdye700	TVR, ProS	[40]
Colon cancer, gastric cancer, pancreatic cancer	TROP2	Humanized anti-TROP2-IRdye700	TVR	[48]
Melanoma	CD146	Anti-CD146 mAb (YY146)-IRdye700	TVR	[110]
Colorectal cancer	GPA33	Anti-GPA33-scFv-IRdye700	TVR	[111]
Hepatocellular carcinoma	GPC3	Anti-GPC3-IRdye700 + nab-paclitaxel	TVR	[112]
Lung cancer	DLL3	Rovalpituzumab-IRdye700	TVR, ProS	[113]
Gastrointestinal cancer	Cadherin-17	Anti-cadherin-17 mAb-IRdye700	TVR	[114]
Gastrointestinal cancer	c-KIT	Anti-c-KIT-IRdye700	TVR	[115]
Mesothelioma	PDPN	Anti-PDPN mAb-IRdye700	TVR, ProS, AIA	[116]
Mesothelioma, pancreatic cancer, ovarian cancer	Mesothelin	Anti-mesothelin-IRdye700	TVR, ProS	[117]
Melanoma	CD29	Anti-CD29-IRdye700 + anti-CD44- IRdye700 and anti-CTLA4 mAb	TVR, ProS	[118]
Lung cancer, mesothelioma	GPR87	Anti-GPR87-IRdye700	TVR	[119]
Prostate, bladder, brain, ovarian cancer	GPC-1	Miltuximab-IRdye700	TVR	[120]
Breast cancer	ICAM-1	Anti-ICAM-1-IRdye700	TVR, ProS	[121]
Prostate and breast tumor	JAM-A	Anti-JAM-A mAb-IRdye700	Decreases number of mitotic cells in cancer tissue	[122]
Lung cancer	CD276	Anti-CD276-F(ab)-IRdye700 + anti-PD-1/PD-L1 blockade	TVR, ProS, AIA	[123]

Table 1. NIR-PIT agents and their clinical and preclinical applications.

Table 1. Cont.

Cancer Type	Target Molecule	Photo-Immunoconjugates	Therapeutic Outcome	References
Colorectal cancer	PDGFRβ	PDGFR β affibody (Z _{PDGFRβ})-IRdye700	Killed pericytes, damaged tumor blood vessels, antitumor effect	[71]
Lung cancer	MRP1	Anti-MRP1 mAb-IRdye700	TVR	[124]
Lymphoma	CLA	Anti-CLA-IRdye700	AIA	[125]
Chemoresistant tumors	Pgp	Anti-Pgp mAb-IRdye700 or Anti-Pgp-F(ab)-IRdye700	TVR, ProS	[126]
Gastric cancer	VEGFR-2	Anti-VEGFR-2 mAb-IRdye700	Damage in tumor neovasculature	[35]
Lung cancer, ovarian cancer	PD-L1	Avelumab-IRdye700 Anti-PD-L1-F(ab)2-IRdye 700	TVR, ProS AIA, AAE TVR, ProS	[59,60] [58]
Colon cancer, lung cancer, prostate cancer, head and neck cancer	CD25	Anti-CD25 mAb-IRdye700 or anti-CD25-F(ab')2-IRdye700 Anti-CD25 mAb-IRdye700 + anti-CD44 mAb-IRdye700	TD, AIA TVR, ProS, AIA	[55] [44]
Breast cancer, lung cancer	CD206	Anti-CD206 mAb-IRdye700	Suppressed sorafenib-resistant tumors	[127]
Colon cancer, lung cancer, head and neck cancer	CTLA4	Anti-CTLA4-IRdye700 Anti-CTLA4-IRdye700 + anti-hEGFR-IRdye700 Anti-CTLA4-IgG-IRdye700 or anti-CTLA4-F(ab')2-IRdye700	TVR, ProS, TD, AIA TVR, ProS, TD, AIA TVR, ProS, TD, AIA	[57] [128] [129]
Colon cancer, lung cancer	VISTA	Anti-VISTA mAb-IRdye700	TVR, ProS	[130]
Breast cancer	Gr-1	Anti-GR-1-IRdye700	Eliminate tumor-induced splenic MDSCs	[131]
Oral cancers, colon cancer	Ly6G	Anti-Ly6G mAb-IRdye700 + Panitumab-IRdye700	Eliminate MDSCs, TVR, ProS, AAE	[132]

*, Clinical trials in human; EGFR, epidermal growth factor receptor; Her2, human epidermal growth factor receptor-2; CAFs, cancer-associated fibroblasts; PSMA, prostate-specific membrane antigen; CEA, carcinoembryonic antigen; TROP2, tumor-associated calcium signal transducer 2; GPA33, glycoprotein A33 antigen; GPC3, Glypican-3; DLL3, delta-like protein 3; PDPN, podoplanin; GPR87, G-protein receptor; GPC-1, glypican-1; ICAM-1, intercellular adhesion molecule-1; JAM-A, junctional adhesion molecule-A; PDGFRβ, platelet-derived growth factor receptor β; MRP1, multidrug resistance protein 1; CLA, cutaneous lymphocyte antigen; Pgp, P-glycoprotein; VEGFR-2, vascular endothelial growth factor receptor 2; PD-L1, programmed death-ligand 1; CTLA4, cytotoxic T-lymphocyte-associated protein 4; VISTA, V-domain immunoglobulin suppressor of T cell activation; *GR-1*, granulocyte receptor-1 antigen; Ly6G, lymphocyte antigen 6 complex locus G6D; TVR, tumor volume reduction; ProS, prolonged survival; CCE, cancer cell elimination; AIA, antitumor immune augmentation; TD, T_{regs} depletion; AAE, abscopal antitumor effect; IMD, immunologic memory development; MDSCs, myeloid-derived suppressor cells.

8.1. EGFR

EGFR overexpression is observed in numerous solid tumors, including head-and-neck, prostate, ovarian, breast, renal, colon, lung, pancreas, skin, and esophageal cancers [33,47,133,134]. Therefore, EGFR is considered to be an excellent target for NIR-PIT and its therapeutic properties have been investigated by several studies [23,33,53,54,72,83,99,100,135,136]. In 2020, the first NIR-PIT drug (cetuximab-IRdye700) was conditionally registered and approved for clinical use in Japan [47].

8.2. Her2

Her2, a membrane tyrosine kinase, has been an established therapeutic target for several years and is overexpressed in different cancer cells including breast cancer [137], gastric cancer [138], and esophageal cancer [139]. Her2-targeted NIR-PIT has exhibited an effective cytotoxicity and significant reduction in tumor volume in Her2 expressing breast cancer [25], lung metastasis [36], non-small cell lung carcinoma [102], and ovarian cancer [101] both in vitro and in vivo. A more recent study showed that Her2 targeting NIR-PIT is more effective in cisplatin (CDDP) chemoresistance small cell lung cancer because Her2 expression is upregulated due to CDDP resistance [103]. In addition, a combination of Her2-targeted NIR-PIT combined with chemotherapy (5-fluorouracil) significantly reduced tumor growth in gastric cancer by inducing necrotic and apoptotic cell death [140].

8.3. PSMA and CEA

The overexpression of the membrane protein PSMA provided an efficient target for cancer treatment. Anti-PSMA-IRdye700 has been employed in NIR-PIT as a promising candidate for the treatment of PSMA-expressing prostate tumor [41], resulting in significant inhibition and prolonged survival of prostate tumor bearing mice. Several NIR-PIT studies were also performed in gastric and pancreatic cancer by targeting CEA, which showed a significant suppression of tumor growth and an improved survival rate [108,109,141,142].

8.4. CD44

CD44 is a well-characterized cancer stem cell marker [143] used as a potential therapeutic target for NIR-PIT. NIR-PIT with anti-CD44-IRdye700 has shown decreased tumor growth and an increased survival rate in a CD44 expressing oral cancer bearing mouse model [50]. Additionally, CD44-targeted NIR-PIT combined with immune checkpoint inhibitor (PD-1 or CTLA4) decreased tumor progression and significantly prolonged survival rate compared with single therapies in colon and lung tumors [51,52,60,106]. Tumor growth inhibition, prolonged survival, and increased anti-tumor immunity have been observed after combined treatment of CD44-targeted NIR-PIT and short-term IL-15 administration compared with the single-agent therapy in colon, lung, and oral tumor bearing subcutaneous mouse models [105].

8.5. PD-L1

PD-L1 is a transmembrane protein, and is overexpressed in numerous cancer cells including ovarian cancer, renal cell carcinoma, and melanoma. PD-L1 induces T cell tolerance by binding to PD-1, expressed on tumor-infiltrating lymphocytes [144]. Tumor growth was significantly inhibited after in vitro NIR-PIT treatment using anti-PD-L1-IRdye700 in a papillary adenocarcinoma bearing mouse model [60]. PD-L1 targeted NIR-PIT also eliminated PD-L1 expressing TAMs and cancer cells in ovarian cancer xenografts [59]. In addition, PD-L1 targeted NIR-PIT resulted in the depletion of the tumor and augmentation of the antitumor immunity in syngeneic mouse tumor. Although a limited cytotoxic effect was found in the in vitro study, an enhanced antitumor effect was observed in the colon, lung, and prostate tumor mouse models [58].

8.6. CTLA4

CTLA4-targeted NIR-PIT depletes intratumorally CTLA4 expressing T_{regs} , resulting in an enhanced T cell mediated antitumor immunity. This local depletion of T_{regs} cells in the tumor bed combined with cancer targeting NIR-PIT is a promising antitumor therapy [57]. Combining NIR-PIT targeting CTLA4 expressing cells and cancer cells is highly effective at suppressing tumor growth and the depletion of CTLA4 expressing T_{regs} . Thus, the antitumor immune response in distant untreated tumors is enhanced [128].

8.7. CD25

Successful cancer therapy is hindered by expanding tumor induced T_{regs} cells. Tumorinfiltrating CD25⁺ T_{reg} cell depletion was contemplated as a vital step for enhancing the anticancer immunity [145]. Local CD25 targeted NIR-PIT selectively eradicated intratumorally T_{regs} , resulting in rapid activation of CD8 T and NK cells, leading to cell-mediated cancer killing [55]. Both antibody fragments and full-length antibody generated NIR-PIT (anti-CD25-F(ab')₂-IRdye700 and anti-CD25-IgG-IRdye700) targeting CD25 were able to significantly inhibit tumor growth in mice with the superior effect of anti-CD25-F(ab')₂-NIR-PIT [56]. Combining CD25-targeted NIR-PIT with CD44 or hEGFR-targeted NIR-PIT showed an improvement in tumor growth inhibition and prolonged survival in tumors compared with the single therapeutic agents [33,44].

8.8. Others

Several TAAs are targeted by NIR-PIT, including CD206, CD47, CD20, vascular endothelial growth factor receptor-2 (VEGF2), and fibroblast activation protein (FAP). TAMtargeted NIR-PIT using an anti-CD206 antibody inhibited the progression of a subcutaneous tumor [127]. NIR-PIT utilizing anti-VEGF2-IRdye700 selectively damaged the tumor vascular endothelium, exhibiting antitumor effects by angiogenesis inhibition in gastric cancer [126]. Repeated CD47-targeted NIR-PIT further suppressed tumor growth and improved survival compared with a single round of treatment [45,46]. FAP-targeted NIR-PIT suppressed tumor growth and reduced chemoresistance in an esophageal tumor model [107,146]. Another study demonstrated that FAP- α - targeted NIR-PIT was able to kill engineered cancer cells effectively and specifically [147]. Moreover, arming anti-CD20 mAb (rituximab) with IRdye700 led to significant inhibition of B-cell lymphoma growth [40].

8.9. Artificially Induced Antigens Targeting NIR-PIT

NIR-PIT relies on targeting highly expressed cell surface antigens, which are not expressed homogenously in all cancer types. Therefore, artificially induced antigen expression for targeting NIR-PIT could be an alternative approach. Yoshida et al. (2012) expressed exogenous Her2 extracellular domain (Her2-ECD) expression on Her2-negative breast cancer cells by transducing the Her2-ECD gene containing the adenoviral vector [148]. Her2-targeted NIR-PIT showed destruction of the Her2-negative breast cancer cells, inhibition of metastasis, and prolongation of survival in the generated cells in vivo [148–150]. A very recent study revealed that ephrin type-A receptor 2 (EphA2) encoding lentiviral vector was transduced to stably express EphA2 for EphA2 targeting NIR-PIT. The anti-EphA2-IRdye700 exhibited notable tumor growth inhibition in EphA2 transduced tumor cells. In addition, EphA2 targeted NIR-PIT elicited innate and adaptive immune responses within the treated tumor [151].

9. Limitation of NIR-PIT

Applying a single PIT agent with mAb against TAAs is unable to fully cure cancer. Moreover, the recurrence of cancer has also been observed after single PIT agent treatment. Multiple NIR-PIT targeting different TAAs in a broad range of cancer types is complicated, expensive, time-consuming, and unpractical. To generate versatile NIR-PIT agents, Shirasu et al. (2019) first developed novel NIR-PIT agents by combining biotinylated antibodies (BioAbs) with AvIR (IRdye700-conjugated NeutrAvidin) to facilitate unlimited target specificities. This approach can overcome the limitation of the repetitive preparation of IRdye700-mAb conjugates [152].

The recurrence of cancer attack in NIR-PIT-treated mice occurs because of the inhomogeneous intratumoral distribution of mAb-IRdye700 due to its relatively large molecular size. To overcome this limitation, systematic repeated NIR light exposure was administrated to the cancer cells in a fractionated manner of mAb-IRdye700, under the guidance of IRdye700 fluorescence signal [83]. Alternatively, a second mAb with a lower affinity was added to overcome the "binding site barrier" phenomenon, thereby facilitating the penetration of the second mAb into deep tumor cells [32].

The labelling of tumor-specific ligands with IRdye700 is generally associated with several drawbacks including, free IRdye700 impurities, heterogenous IRdye700-ligand conjugates from batch to batch, poor reproducibility, and aggregation [13,153,154]. Moreover, direct conjugation of ligands with IRdye700 may occur at the recognition site, which can hamper or even lose the targeting activity [13]. NIR-PIT relies on specific ligands for targeting highly expressed cell surface antigens, which are not always overexpressed in continuously developing cancers. Thereby, this technology is limited to tumors that express a high level of cell surface antigens [18]. However, a recent study demonstrated that NIR-PIT targeting low-expressed PD-L1 generated sufficient antitumor effects in tumor cells and an increased antitumor immunity [58].

The limitation of NIR-PIT also includes the difficulty of delivering NIR light to large tumor masses and to deeply located tumors such as lung tumor, intraperitoneal cancers, and cholangiocarcinoma [37,38,51,99,155]. Recently, an ET-BLIT was developed for light irradiation that can be accessible in the whole body [91]. Moreover, realistic, immunocompetent mouse models suitable for human diseases are required for studying the systemic antitumor immune responses [18,119,156]. As the growth of tumors is faster in mice than humans [157], no apparent side effects of repeated NIR-PIT were observed in a mouse model. In addition, the long term side effects of repeated NIR-PIT were not investigated [36]. A very recent study demonstrated that light doses at 100 J/cm² could cause extensive local edema and weight loss [158].

NIR-PIT is mostly unable to induce durable antitumor responses due to the adaptive immune resistance of humans [51]. The single NIR-PIT agent was reported to be effective at killing cancer cells, but tumors can develop resistance to monotherapy [152]. To overcome this limitation, a combination of immune-checkpoint inhibitors [52,108], multiple NIR-PIT agents targeting different antigens [100], and T_{regs} targeting additional NIR-PIT agents [44] can be applied. NIR-PIT targeting T_{reg} cell antigens show tumor regression by depleting intratumoral T_{reg} cells. However, the activated T cells can also be depleted by ADCC and CDC due to the Fc region of the antibodies used in NIR-PIT [47]. Systemic depletion of T_{regs} may also induce adverse autoimmune events, which can be overcome by depleting the T_{reg} cells locally without eliminating T_{regs} in other organs [55,56].

10. Conclusions

NIR-PIT has become a promising treatment modality for any type of cancer by targeting cancer cells, immune-regulatory cells, or both, where NIR can reach. NIR-PIT has advantages over traditional therapy by eliminating the cancer cells in a highly specific manner and enhancing the host immune response with minimal off-target effects. When used in combination with immune-suppressor targeted NIR-PIT or adjuvant therapies, NIR-PIT can generate a robust antitumor immune response. The development of a new NIR light delivery system, uniform NIR light treatment methods, and real-time monitoring of the treatment effect has made NIR-PIT a potential treatment for a wide variety of cancers.

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References

- Twomey, J.D.; Zhang, B. Cancer Immunotherapy Update: FDA-Approved Checkpoint Inhibitors and Companion Diagnostics. AAPS J. 2021, 23, 39. [CrossRef] [PubMed]
- 2. Yan, L.; Rosen, N.; Arteaga, C. Targeted cancer therapies. Chin. J. Cancer 2011, 30, 1–4. [CrossRef] [PubMed]
- Li, X.; Lovell, J.F.; Yoon, J.; Chen, X. Clinical development and potential of photothermal and photodynamic therapies for cancer. Nat. Rev. Clin. Oncol. 2020, 17, 657–674. [CrossRef] [PubMed]
- Baskaran, R.; Lee, J.; Yang, S.G. Clinical development of photodynamic agents and therapeutic applications. *Biomater. Res.* 2018, 22, 25. [CrossRef] [PubMed]
- 5. Gunaydin, G.; Gedik, M.E.; Ayan, S. Photodynamic Therapy-Current Limitations and Novel Approaches. *Front. Chem.* **2021**, *9*, 691697. [CrossRef]
- 6. Liu, Z.; Xie, Z.; Li, W.; Wu, X.; Jiang, X.; Li, G.; Cao, L.; Zhang, D.; Wang, Q.; Xue, P.; et al. Photodynamic immunotherapy of cancers based on nanotechnology: Recent advances and future challenges. *J. Nanobiotechnol.* **2021**, *19*, 160. [CrossRef] [PubMed]
- 7. Wang, M.; Rao, J.; Wang, M.; Li, X.; Liu, K.; Naylor, M.F.; Nordquist, R.E.; Chen, W.R.; Zhou, F.J.T. Cancer photo-immunotherapy: From bench to bedside. *Theranostics* **2021**, *11*, 2218. [CrossRef]
- 8. Shafirstein, G.; Bellnier, D.; Oakley, E.; Hamilton, S.; Potasek, M.; Beeson, K.; Parilov, E.J.C. Interstitial photodynamic therapy—A focused review. *Cancers* 2017, *9*, 12. [CrossRef]
- 9. Zou, J.; Li, L.; Yang, Z.; Chen, X.J.N. Phototherapy meets immunotherapy: A win-win strategy to fight against cancer. *Nanophotonics* 2021, *10*, 3229–3245. [CrossRef]
- 10. Kobayashi, H.; Furusawa, A.; Rosenberg, A.; Choyke, P.L. Near-infrared photoimmunotherapy of cancer: A new approach that kills cancer cells and enhances anti-cancer host immunity. *Int. Immunol.* **2021**, *33*, 7–15. [CrossRef]
- 11. Monaco, H.; Yokomizo, S.; Choi, H.S.; Kashiwagi, S.J.V. Quickly evolving near-infrared photoimmunotherapy provides multifaceted approach to modern cancer treatment. *VIEW* **2022**, *3*, 20200110. [CrossRef]
- 12. Ogawa, M.; Takakura, H.J.B.; Chemistry, M. Photoimmunotherapy: A new cancer treatment using photochemical reactions. *Bioorg. Med. Chem.* **2021**, 43, 116274. [CrossRef] [PubMed]
- Mussini, A.; Uriati, E.; Bianchini, P.; Diaspro, A.; Cavanna, L.; Abbruzzetti, S.; Viappiani, C.J.B.C. Targeted photoimmunotherapy for cancer. *Biomol. Concepts* 2022, 13, 126–147. [CrossRef] [PubMed]
- 14. Castano, A.P.; Mroz, P.; Hamblin, M.R. Photodynamic therapy and anti-tumour immunity. *Nat. Rev. Cancer* **2006**, *6*, 535–545. [CrossRef]
- Chen, Z.; Liu, L.; Liang, R.; Luo, Z.; He, H.; Wu, Z.; Tian, H.; Zheng, M.; Ma, Y.; Cai, L. Bioinspired hybrid protein oxygen nanocarrier amplified photodynamic therapy for eliciting anti-tumor immunity and abscopal effect. ACS Nano 2018, 12, 8633–8645. [CrossRef] [PubMed]
- 16. Shen, Z.; Ma, Q.; Zhou, X.; Zhang, G.; Hao, G.; Sun, Y.; Cao, J. Strategies to improve photodynamic therapy efficacy by relieving the tumor hypoxia environment. *NPG Asia Mater.* **2021**, *13*, 39. [CrossRef]
- Kobayashi, H.; Choyke, P.L. Near-Infrared Photoimmunotherapy of Cancer. Acc. Chem. Res. 2019, 52, 2332–2339. [CrossRef] [PubMed]
- Paraboschi, I.; Turnock, S.; Kramer-Marek, G.; Musleh, L.; Barisa, M.; Anderson, J.; Giuliani, S. Near-InfraRed PhotoImmunoTherapy (NIR-PIT) for the local control of solid cancers: Challenges and potentials for human applications. *Crit. Rev. Oncol. Hematol.* 2021, 161, 103325. [CrossRef]
- 19. Liu, Y.; Zhang, L.; Chang, R.; Yan, X. Supramolecular cancer photoimmunotherapy based on precise peptide self-assembly design. *Chem. Commun.* **2022**, *58*, 2247–2258. [CrossRef]
- Peng, Z.; Lv, X.; Huang, S. Photoimmunotherapy: A New Paradigm in Solid Tumor Immunotherapy. *Cancer Control* 2022, 29, 10732748221088825. [CrossRef]
- Xu, X.; Lu, H.; Lee, R. Near infrared light triggered photo/immuno-therapy toward cancers. Front. Bioeng. Biotechnol. 2020, 8, 488. [CrossRef]
- 22. Even-Desrumeaux, K.; Baty, D.; Chames, P.J.C. State of the art in tumor antigen and biomarker discovery. *Cancers* 2011, *3*, 2554–2596. [CrossRef]
- Von Felbert, V.; Bauerschlag, D.; Maass, N.; Bräutigam, K.; Meinhold-Heerlein, I.; Woitok, M.; Barth, S.; Hussain, A.F.J. A specific photoimmunotheranostics agent to detect and eliminate skin cancer cells expressing EGFR. *J. Cancer Res. Clin. Oncol.* 2016, 142, 1003–1011. [CrossRef] [PubMed]

- 24. Fernandes, S.R.; Fernandes, R.; Sarmento, B.; Pereira, P.M.; Tomé, J.P.J.O.; Chemistry, B. Photoimmunoconjugates: Novel synthetic strategies to target and treat cancer by photodynamic therapy. *Org. Biomol. Chem.* **2019**, *17*, 2579–2593. [CrossRef] [PubMed]
- Mitsunaga, M.; Ogawa, M.; Kosaka, N.; Rosenblum, L.T.; Choyke, P.L.; Kobayashi, H. Cancer cell–selective in vivo near infrared photoimmunotherapy targeting specific membrane molecules. *Nat. Med.* 2011, *17*, 1685–1691. [CrossRef]
- Wollschlaeger, C.; Meinhold-Heerlein, I.; Cong, X.; Bräutigam, K.; Di Fiore, S.; Zeppernick, F.; Klockenbring, T.; Stickeler, E.; Barth, S.; Hussain, A.F. Simultaneous and independent dual site-specific self-labeling of recombinant antibodies. *Bioconjug. Chem.* 2018, 29, 3586–3594. [CrossRef] [PubMed]
- 27. Wakiyama, H.; Kato, T.; Furusawa, A.; Choyke, P.L.; Kobayashi, H.J.N. Near infrared photoimmunotherapy of cancer; possible clinical applications. *Nanophotonics* **2021**, *10*, 3135–3151. [CrossRef] [PubMed]
- Okada, R.; Kato, T.; Furusawa, A.; Inagaki, F.; Wakiyama, H.; Fujimura, D.; Okuyama, S.; Furumoto, H.; Fukushima, H.; Choyke, P.L. Immunotherapy. Selection of antibody and light exposure regimens alters therapeutic effects of EGFR-targeted near-infrared photoimmunotherapy. *Cancer Immunol. Immunother.* 2022, 71, 1877–1887. [CrossRef] [PubMed]
- 29. Nagaya, T.; Nakamura, Y.; Sato, K.; Harada, T.; Choyke, P.L.; Kobayashi, H.J. Improved micro-distribution of antibody-photon absorber conjugates after initial near infrared photoimmunotherapy (NIR-PIT). J. Control. Release 2016, 232, 1–8. [CrossRef]
- Ogawa, M.; Tomita, Y.; Nakamura, Y.; Lee, M.-J.; Lee, S.; Tomita, S.; Nagaya, T.; Sato, K.; Yamauchi, T.; Iwai, H.J.O. Immunogenic cancer cell death selectively induced by near infrared photoimmunotherapy initiates host tumor immunity. *Oncotarget* 2017, *8*, 10425. [CrossRef]
- 31. Sato, K.; Watanabe, R.; Hanaoka, H.; Harada, T.; Nakajima, T.; Kim, I.; Paik, C.H.; Choyke, P.L.; Kobayashi, H. Photoimmunotherapy: Comparative effectiveness of two monoclonal antibodies targeting the epidermal growth factor receptor. *Mol. Oncol.* **2014**, *8*, 620–632. [CrossRef]
- Nakajima, T.; Sano, K.; Choyke, P.L.; Kobayashi, H.J.T. Improving the efficacy of Photoimmunotherapy (PIT) using a cocktail of antibody conjugates in a multiple antigen tumor model. *Theranostics* 2013, *3*, 357. [CrossRef] [PubMed]
- Okada, R.; Furusawa, A.; Vermeer, D.W.; Inagaki, F.; Wakiyama, H.; Kato, T.; Nagaya, T.; Choyke, P.L.; Spanos, W.C.; Allen, C.T. Near-infrared photoimmunotherapy targeting human-EGFR in a mouse tumor model simulating current and future clinical trials. *EBioMedicine* 2021, 67, 103345. [CrossRef] [PubMed]
- 34. Nishimura, T.; Mitsunaga, M.; Ito, K.; Kobayashi, H.; Saruta, M. Cancer neovasculature-targeted near-infrared photoimmunotherapy (NIR-PIT) for gastric cancer: Different mechanisms of phototoxicity compared to cell membrane-targeted NIR-PIT. *Gastric Cancer* 2020, *23*, 82–94. [CrossRef]
- 35. Sato, K.; Nakajima, T.; Choyke, P.L.; Kobayashi, H. Selective cell elimination in vitro and in vivo from tissues and tumors using antibodies conjugated with a near infrared phthalocyanine. *RSC Adv.* **2015**, *5*, 25105–25114. [CrossRef]
- Sato, K.; Nagaya, T.; Mitsunaga, M.; Choyke, P.L.; Kobayashi, H. Near infrared photoimmunotherapy for lung metastases. *Cancer Lett.* 2015, 365, 112–121. [CrossRef]
- Hirata, H.; Kuwatani, M.; Nakajima, K.; Kodama, Y.; Yoshikawa, Y.; Ogawa, M.; Sakamoto, N. Near-infrared photoimmunotherapy (NIR-PIT) on cholangiocarcinoma using a novel catheter device with light emitting diodes. *Cancer Sci.* 2021, 112, 828–838. [CrossRef] [PubMed]
- Kim, K.S.; Kim, J.; Kim, D.H.; Hwang, H.S.; Na, K. Multifunctional trastuzumab-chlorin e6 conjugate for the treatment of HER2-positive human breast cancer. *Biomater. Sci.* 2018, 6, 1217–1226. [CrossRef] [PubMed]
- Korsak, B.; Almeida, G.M.; Rocha, S.; Pereira, C.; Mendes, N.; Osório, H.; Pereira, P.M.R.; Rodrigues, J.M.M.; Schneider, R.J.; Sarmento, B.; et al. Porphyrin modified trastuzumab improves efficacy of HER2 targeted photodynamic therapy of gastric cancer. *Int. J. Cancer* 2017, 141, 1478–1489. [CrossRef]
- 40. Nagaya, T.; Nakamura, Y.; Sato, K.; Harada, T.; Choyke, P.L.; Kobayashi, H. Near infrared photoimmunotherapy of B-cell lymphoma. *Mol. Oncol.* **2016**, *10*, 1404–1414. [CrossRef]
- Nagaya, T.; Nakamura, Y.; Okuyama, S.; Ogata, F.; Maruoka, Y.; Choyke, P.L.; Kobayashi, H. Near-Infrared Photoimmunotherapy Targeting Prostate Cancer with Prostate-Specific Membrane Antigen (PSMA) Antibody. *Mol. Cancer Res.* 2017, 15, 1153–1162. [CrossRef] [PubMed]
- 42. Isoda, Y.; Piao, W.; Taguchi, E.; Iwano, J.; Takaoka, S.; Uchida, A.; Yoshikawa, K.; Enokizono, J.; Arakawa, E.; Tomizuka, K.; et al. Development and evaluation of a novel antibody-photon absorber conjugate reveals the possibility of photoimmunotherapyinduced vascular occlusion during treatment in vivo. *Oncotarget* 2018, *9*, 31422–31431. [CrossRef]
- Han, Y.; An, Y.; Jia, G.; Wang, X.; He, C.; Ding, Y.; Tang, Q. Theranostic micelles based on upconversion nanoparticles for dual-modality imaging and photodynamic therapy in hepatocellular carcinoma. *Nanoscale* 2018, 10, 6511–6523. [CrossRef]
- Maruoka, Y.; Furusawa, A.; Okada, R.; Inagaki, F.; Fujimura, D.; Wakiyama, H.; Kato, T.; Nagaya, T.; Choyke, P.L.; Kobayashi, H. Combined CD44- and CD25-Targeted Near-Infrared Photoimmunotherapy Selectively Kills Cancer and Regulatory T Cells in Syngeneic Mouse Cancer Models. *Cancer Immunol. Res.* 2020, *8*, 345–355. [CrossRef]
- Kiss, B.; van den Berg, N.S.; Ertsey, R.; McKenna, K.; Mach, K.E.; Zhang, C.A.; Volkmer, J.P.; Weissman, I.L.; Rosenthal, E.L.; Liao, J.C. CD47-Targeted Near-Infrared Photoimmunotherapy for Human Bladder Cancer. *Clin. Cancer Res.* 2019, 25, 3561–3571. [CrossRef] [PubMed]
- 46. Yang, Y.; Yan, X.; Li, J.; Liu, C.; Yang, X. CD47-targeted optical molecular imaging and near-infrared photoimmunotherapy in the detection and treatment of bladder cancer. *Mol. Ther. Oncolytics* **2022**, *24*, 319–330. [CrossRef] [PubMed]

- Maruoka, Y.; Wakiyama, H.; Choyke, P.L.; Kobayashi, H. Near infrared photoimmunotherapy for cancers: A translational perspective. *EBioMedicine* 2021, 70, 103501. [CrossRef]
- Nishimura, T.; Mitsunaga, M.; Sawada, R.; Saruta, M.; Kobayashi, H.; Matsumoto, N.; Kanke, T.; Yanai, H.; Nakamura, K. Photoimmunotherapy targeting biliary-pancreatic cancer with humanized anti-TROP2 antibody. *Cancer Med.* 2019, *8*, 7781–7792. [CrossRef]
- Jing, H.; Weidensteiner, C.; Reichardt, W.; Gaedicke, S.; Zhu, X.; Grosu, A.L.; Kobayashi, H.; Niedermann, G. Imaging and Selective Elimination of Glioblastoma Stem Cells with Theranostic Near-Infrared-Labeled CD133-Specific Antibodies. *Theranostics* 2016, 6, 862–874. [CrossRef]
- 50. Nagaya, T.; Nakamura, Y.; Okuyama, S.; Ogata, F.; Maruoka, Y.; Choyke, P.L.; Allen, C.; Kobayashi, H. Syngeneic Mouse Models of Oral Cancer Are Effectively Targeted by Anti-CD44-Based NIR-PIT. *Mol. Cancer Res.* **2017**, *15*, 1667–1677. [CrossRef]
- Nagaya, T.; Friedman, J.; Maruoka, Y.; Ogata, F.; Okuyama, S.; Clavijo, P.E.; Choyke, P.L.; Allen, C.; Kobayashi, H. Host Immunity Following Near-Infrared Photoimmunotherapy Is Enhanced with PD-1 Checkpoint Blockade to Eradicate Established Antigenic Tumors. *Cancer Immunol. Res.* 2019, 7, 401–413. [CrossRef] [PubMed]
- Wakiyama, H.; Furusawa, A.; Okada, R.; Inagaki, F.; Kato, T.; Maruoka, Y.; Choyke, P.L.; Kobayashi, H. Increased Immunogenicity of a Minimally Immunogenic Tumor after Cancer-Targeting Near Infrared Photoimmunotherapy. *Cancers* 2020, 12, 3747. [CrossRef]
- Bauerschlag, D.; Meinhold-Heerlein, I.; Maass, N.; Bleilevens, A.; Bräutigam, K.; Al Rawashdeh, W.; Di Fiore, S.; Haugg, A.M.; Gremse, F.; Steitz, J.; et al. Detection and Specific Elimination of EGFR(+) Ovarian Cancer Cells Using a Near Infrared Photoimmunotheranostic Approach. *Pharm. Res.* 2017, 34, 696–703. [CrossRef] [PubMed]
- 54. Amoury, M.; Bauerschlag, D.; Zeppernick, F.; von Felbert, V.; Berges, N.; Di Fiore, S.; Mintert, I.; Bleilevens, A.; Maass, N.; Bräutigam, K.; et al. Photoimmunotheranostic agents for triple-negative breast cancer diagnosis and therapy that can be activated on demand. *Oncotarget* **2016**, *7*, 54925–54936. [CrossRef]
- Sato, K.; Sato, N.; Xu, B.; Nakamura, Y.; Nagaya, T.; Choyke, P.L.; Hasegawa, Y.; Kobayashi, H. Spatially selective depletion of tumor-associated regulatory T cells with near-infrared photoimmunotherapy. *Sci. Transl. Med.* 2016, *8*, 352ra110. [CrossRef] [PubMed]
- Okada, R.; Maruoka, Y.; Furusawa, A.; Inagaki, F.; Nagaya, T.; Fujimura, D.; Choyke, P.L.; Kobayashi, H. The Effect of Antibody Fragments on CD25 Targeted Regulatory T Cell Near-Infrared Photoimmunotherapy. *Bioconjug. Chem.* 2019, 30, 2624–2633. [CrossRef] [PubMed]
- 57. Okada, R.; Kato, T.; Furusawa, A.; Inagaki, F.; Wakiyama, H.; Choyke, P.L.; Kobayashi, H. Local Depletion of Immune Checkpoint Ligand CTLA4 Expressing Cells in Tumor Beds Enhances Antitumor Host Immunity. *Adv. Ther.* **2021**, *4*, 2000269. [CrossRef]
- Taki, S.; Matsuoka, K.; Nishinaga, Y.; Takahashi, K.; Yasui, H.; Koike, C.; Shimizu, M.; Sato, M.; Sato, K. Spatiotemporal depletion of tumor-associated immune checkpoint PD-L1 with near-infrared photoimmunotherapy promotes antitumor immunity. *J. Immunother. Cancer* 2021, 9, e003036. [CrossRef]
- 59. Jin, J.; Sivakumar, I.; Mironchik, Y.; Krishnamachary, B.; Wildes, F.; Barnett, J.D.; Hung, C.F.; Nimmagadda, S.; Kobayashi, H.; Bhujwalla, Z.M.; et al. PD-L1 near Infrared Photoimmunotherapy of Ovarian Cancer Model. *Cancers* **2022**, *14*, 619. [CrossRef]
- 60. Nagaya, T.; Nakamura, Y.; Sato, K.; Harada, T.; Choyke, P.L.; Hodge, J.W.; Schlom, J.; Kobayashi, H. Near infrared photoimmunotherapy with avelumab, an anti-programmed death-ligand 1 (PD-L1) antibody. *Oncotarget* **2017**, *8*, 8807–8817. [CrossRef]
- 61. Sioud, M.; Westby, P.; Vasovic, V.; Fløisand, Y.; Peng, Q. Development of a new high-affinity human antibody with antitumor activity against solid and blood malignancies. *Faseb J.* **2018**, *32*, 5063–5077. [CrossRef] [PubMed]
- 62. Sioud, M.; Juzenas, P.; Zhang, Q.; Kleinauskas, A.; Peng, Q. Evaluation of In Vitro Phototoxicity of a Minibody-IR700 Conjugate Using Cell Monolayer and Multicellular Tumor Spheroid Models. *Cancers* **2021**, *13*, 3356. [CrossRef] [PubMed]
- Watanabe, R.; Hanaoka, H.; Sato, K.; Nagaya, T.; Harada, T.; Mitsunaga, M.; Kim, I.; Paik, C.H.; Wu, A.M.; Choyke, P.L.; et al. Photoimmunotherapy targeting prostate-specific membrane antigen: Are antibody fragments as effective as antibodies? *J. Nucl. Med.* 2015, 56, 140–144. [CrossRef]
- 64. Jovčevska, I.; Muyldermans, S. The Therapeutic Potential of Nanobodies. BioDrugs 2020, 34, 11–26. [CrossRef]
- De Groof, T.W.M.; Mashayekhi, V.; Fan, T.S.; Bergkamp, N.D.; Sastre Toraño, J.; van Senten, J.R.; Heukers, R.; Smit, M.J.; Oliveira, S. Nanobody-Targeted Photodynamic Therapy Selectively Kills Viral GPCR-Expressing Glioblastoma Cells. *Mol. Pharm.* 2019, 16, 3145–3156. [CrossRef]
- 66. Frejd, F.Y.; Kim, K.T. Affibody molecules as engineered protein drugs. Exp. Mol. Med. 2017, 49, e306. [CrossRef]
- Ståhl, S.; Gräslund, T.; Eriksson Karlström, A.; Frejd, F.Y.; Nygren, P.; Löfblom, J. Affibody Molecules in Biotechnological and Medical Applications. *Trends Biotechnol.* 2017, 35, 691–712. [CrossRef] [PubMed]
- Tolmachev, V.; Orlova, A.; Nilsson, F.Y.; Feldwisch, J.; Wennborg, A.; Abrahmsén, L. Affibody molecules: Potential for in vivo imaging of molecular targets for cancer therapy. *Expert Opin. Biol. Ther.* 2007, 7, 555–568. [CrossRef]
- 69. Mączyńska, J.; Da Pieve, C.; Burley, T.A.; Raes, F.; Shah, A.; Saczko, J.; Harrington, K.J.; Kramer-Marek, G. Immunomodulatory activity of IR700-labelled affibody targeting HER2. *Cell Death Dis.* **2020**, *11*, 886. [CrossRef]
- Yamaguchi, H.; On, J.; Morita, T.; Suzuki, T.; Okada, Y.; Ono, J.; Evdokiou, A. Combination of Near-Infrared Photoimmunotherapy Using Trastuzumab and Small Protein Mimetic for HER2-Positive Breast Cancer. *Int. J. Mol. Sci.* 2021, 22, 12213. [CrossRef]
- Shi, Q.; Tao, Z.; Yang, H.; Fan, Q.; Wei, D.; Wan, L.; Lu, X. PDGFRβ-specific affibody-directed delivery of a photosensitizer, IR700, is efficient for vascular-targeted photodynamic therapy of colorectal cancer. *Drug Deliv.* 2017, 24, 1818–1830. [CrossRef] [PubMed]

- Burley, T.A.; Mączyńska, J.; Shah, A.; Szopa, W.; Harrington, K.J.; Boult, J.K.R.; Mrozek-Wilczkiewicz, A.; Vinci, M.; Bamber, J.C.; Kaspera, W.; et al. Near-infrared photoimmunotherapy targeting EGFR-Shedding new light on glioblastoma treatment. *Int. J. Cancer* 2018, 142, 2363–2374. [CrossRef]
- Mączyńska, J.; Raes, F.; Da Pieve, C.; Turnock, S.; Boult, J.K.R.; Hoebart, J.; Niedbala, M.; Robinson, S.P.; Harrington, K.J.; Kaspera, W.; et al. Triggering anti-GBM immune response with EGFR-mediated photoimmunotherapy. *BMC Med.* 2022, 20, 16. [CrossRef]
- 74. Li, F.; Zhao, Y.; Mao, C.; Kong, Y.; Ming, X. RGD-Modified Albumin Nanoconjugates for Targeted Delivery of a Porphyrin Photosensitizer. *Mol. Pharm.* 2017, 14, 2793–2804. [CrossRef]
- Zhao, Y.; Li, F.; Mao, C.; Ming, X. Multiarm Nanoconjugates for Cancer Cell-Targeted Delivery of Photosensitizers. *Mol. Pharm.* 2018, 15, 2559–2569. [CrossRef]
- Dou, X.; Nomoto, T.; Takemoto, H.; Matsui, M.; Tomoda, K.; Nishiyama, N. Effect of multiple cyclic RGD peptides on tumor accumulation and intratumoral distribution of IRDye 700DX-conjugated polymers. *Sci. Rep.* 2018, *8*, 8126. [CrossRef] [PubMed]
- 77. Liu, H.; Zhao, Z.; Zhang, L.; Li, Y.; Jain, A.; Barve, A.; Jin, W.; Liu, Y.; Fetse, J.; Cheng, K. Discovery of low-molecular weight anti-PD-L1 peptides for cancer immunotherapy. *J. Immunother. Cancer* **2019**, *7*, 270. [CrossRef]
- 78. Sasikumar, P.G.; Ramachandra, M. Small-molecule antagonists of the immune checkpoint pathways: Concept to clinic. *Future Med. Chem.* **2017**, *9*, 1305–1308. [CrossRef]
- Nakajima, K.; Miyazaki, F.; Terada, K.; Takakura, H.; Suzuki, M.; Ogawa, M. Comparison of low-molecular-weight ligand and whole antibody in prostate-specific membrane antigen targeted near-infrared photoimmunotherapy. *Int. J. Pharm.* 2021, 609, 121135. [CrossRef]
- Sato, K.; Ando, K.; Okuyama, S.; Moriguchi, S.; Ogura, T.; Totoki, S.; Hanaoka, H.; Nagaya, T.; Kokawa, R.; Takakura, H.; et al. Photoinduced Ligand Release from a Silicon Phthalocyanine Dye Conjugated with Monoclonal Antibodies: A Mechanism of Cancer Cell Cytotoxicity after Near-Infrared Photoimmunotherapy. ACS Cent. Sci. 2018, 4, 1559–1569. [CrossRef]
- Kobayashi, H.; Griffiths, G.L.; Choyke, P.L. Near-Infrared Photoimmunotherapy: Photoactivatable Antibody-Drug Conjugates (ADCs). *Bioconjug. Chem.* 2020, 31, 28–36. [CrossRef]
- Nakajima, K.; Ogawa, M. Phototoxicity in near-infrared photoimmunotherapy is influenced by the subcellular localization of antibody-IR700. *Photodiagnosis Photodyn. Ther.* 2020, 31, 101926. [CrossRef] [PubMed]
- 83. Mitsunaga, M.; Nakajima, T.; Sano, K.; Choyke, P.L.; Kobayashi, H. Near-infrared theranostic photoimmunotherapy (PIT): Repeated exposure of light enhances the effect of immunoconjugate. *Bioconjug. Chem.* **2012**, *23*, 604–609. [CrossRef] [PubMed]
- 84. Perez, H.L.; Cardarelli, P.M.; Deshpande, S.; Gangwar, S.; Schroeder, G.M.; Vite, G.D.; Borzilleri, R.M. Antibody-drug conjugates: Current status and future directions. *Drug Discov. Today* **2014**, *19*, 869–881. [CrossRef]
- 85. Hussain, A.F.; Heppenstall, P.A.; Kampmeier, F.; Meinhold-Heerlein, I.; Barth, S. One-step site-specific antibody fragment auto-conjugation using SNAP-tag technology. *Nat. Protoc.* **2019**, *14*, 3101–3125. [CrossRef] [PubMed]
- Chouman, K.; Woitok, M.; Mladenov, R.; Kessler, C.; Weinhold, E.; Hanz, G.; Fischer, R.; Meinhold-Heerlein, I.; Bleilevens, A.; Gresch, G.; et al. Fine Tuning Antibody Conjugation Methods using SNAP-tag Technology. *Anticancer Agents Med. Chem.* 2017, 17, 1434–1440. [CrossRef]
- 87. Henderson, T.A.; Morries, L.D. Near-infrared photonic energy penetration: Can infrared phototherapy effectively reach the human brain? *Neuropsychiatr. Dis. Treat.* **2015**, *11*, 2191–2208. [CrossRef]
- 88. Nagaya, T.; Okuyama, S.; Ogata, F.; Maruoka, Y.; Choyke, P.L.; Kobayashi, H. Endoscopic near infrared photoimmunotherapy using a fiber optic diffuser for peritoneal dissemination of gastric cancer. *Cancer Sci.* **2018**, *109*, 1902–1908. [CrossRef]
- Nagaya, T.; Okuyama, S.; Ogata, F.; Maruoka, Y.; Choyke, P.L.; Kobayashi, H. Near infrared photoimmunotherapy using a fiber optic diffuser for treating peritoneal gastric cancer dissemination. *Gastric Cancer* 2019, 22, 463–472. [CrossRef]
- Okada, R.; Furusawa, A.; Inagaki, F.; Wakiyama, H.; Kato, T.; Okuyama, S.; Furumoto, H.; Fukushima, H.; Choyke, P.L.; Kobayashi, H. Endoscopic near-infrared photoimmunotherapy in an orthotopic head and neck cancer model. *Cancer Sci.* 2021, *112*, 3041–3049. [CrossRef]
- Tsukamoto, T.; Fujita, Y.; Shimogami, M.; Kaneda, K.; Seto, T.; Mizukami, K.; Takei, M.; Isobe, Y.; Yasui, H.; Sato, K. Inside-the-body light delivery system using endovascular therapy-based light illumination technology. *EBioMedicine* 2022, 85, 104289. [CrossRef] [PubMed]
- 92. Zhang, X.; Nakajima, T.; Mizoi, K.; Tsushima, Y.; Ogihara, T. Imaging modalities for monitoring acute therapeutic effects after near-infrared photoimmunotherapy in vivo. *J. Biophotonics* **2022**, *15*, e202100266. [CrossRef] [PubMed]
- 93. Nakajima, T.; Sato, K.; Hanaoka, H.; Watanabe, R.; Harada, T.; Choyke, P.L.; Kobayashi, H. The effects of conjugate and light dose on photo-immunotherapy induced cytotoxicity. *BMC Cancer* **2014**, *14*, 389. [CrossRef] [PubMed]
- Sano, K.; Mitsunaga, M.; Nakajima, T.; Choyke, P.L.; Kobayashi, H. Acute cytotoxic effects of photoimmunotherapy assessed by 18F-FDG PET. J. Nucl. Med. 2013, 54, 770–775. [CrossRef]
- Liang, C.P.; Nakajima, T.; Watanabe, R.; Sato, K.; Choyke, P.L.; Chen, Y.; Kobayashi, H. Real-time monitoring of hemodynamic changes in tumor vessels during photoimmunotherapy using optical coherence tomography. *J. Biomed. Opt.* 2014, *19*, 98004. [CrossRef]
- 96. Tang, Q.; Nagaya, T.; Liu, Y.; Lin, J.; Sato, K.; Kobayashi, H.; Chen, Y. Real-time monitoring of microdistribution of antibody-photon absorber conjugates during photoimmunotherapy in vivo. *J. Control. Release* **2017**, *260*, 154–163. [CrossRef]

- Inagaki, F.F.; Fujimura, D.; Furusawa, A.; Okada, R.; Wakiyama, H.; Kato, T.; Choyke, P.L.; Kobayashi, H. Diagnostic imaging in near-infrared photoimmunotherapy using a commercially available camera for indocyanine green. *Cancer Sci.* 2021, 112, 1326–1330. [CrossRef]
- Kishimoto, S.; Oshima, N.; Yamamoto, K.; Munasinghe, J.; Ardenkjaer-Larsen, J.H.; Mitchell, J.B.; Choyke, P.L.; Krishna, M.C. Molecular imaging of tumor photoimmunotherapy: Evidence of photosensitized tumor necrosis and hemodynamic changes. *Free Radic. Biol. Med.* 2018, 116, 1–10. [CrossRef]
- Nakamura, Y.; Ohler, Z.W.; Householder, D.; Nagaya, T.; Sato, K.; Okuyama, S.; Ogata, F.; Daar, D.; Hoa, T.; Choyke, P.L.; et al. Near Infrared Photoimmunotherapy in a Transgenic Mouse Model of Spontaneous Epidermal Growth Factor Receptor (EGFR)expressing Lung Cancer. *Mol. Cancer Ther.* 2017, *16*, 408–414. [CrossRef]
- 100. Siddiqui, M.R.; Railkar, R.; Sanford, T.; Crooks, D.R.; Eckhaus, M.A.; Haines, D.; Choyke, P.L.; Kobayashi, H.; Agarwal, P.K. Targeting Epidermal Growth Factor Receptor (EGFR) and Human Epidermal Growth Factor Receptor 2 (HER2) Expressing Bladder Cancer Using Combination Photoimmunotherapy (PIT). *Sci. Rep.* 2019, *9*, 2084. [CrossRef]
- 101. Sato, K.; Hanaoka, H.; Watanabe, R.; Nakajima, T.; Choyke, P.L.; Kobayashi, H. Near infrared photoimmunotherapy in the treatment of disseminated peritoneal ovarian cancer. *Mol. Cancer. Ther.* **2015**, *14*, 141–150. [CrossRef] [PubMed]
- 102. Sato, K.; Nagaya, T.; Choyke, P.L.; Kobayashi, H. Near infrared photoimmunotherapy in the treatment of pleural disseminated NSCLC: Preclinical experience. *Theranostics* **2015**, *5*, 698–709. [CrossRef] [PubMed]
- 103. Takahashi, K.; Taki, S.; Yasui, H.; Nishinaga, Y.; Isobe, Y.; Matsui, T.; Shimizu, M.; Koike, C.; Sato, K. HER2 targeting near-infrared photoimmunotherapy for a CDDP-resistant small-cell lung cancer. *Cancer Med.* **2021**, *10*, 8808–8819. [CrossRef] [PubMed]
- 104. Takahashi, K.; Yasui, H.; Taki, S.; Shimizu, M.; Koike, C.; Taki, K.; Yukawa, H.; Baba, Y.; Kobayashi, H.; Sato, K.J.B.; et al. Near-infrared-induced drug release from antibody–drug double conjugates exerts a cytotoxic photo-bystander effect. *Bioeng. Transl. Med.* 2022, 7, e10388. [CrossRef]
- 105. Maruoka, Y.; Furusawa, A.; Okada, R.; Inagaki, F.; Wakiyama, H.; Kato, T.; Nagaya, T.; Choyke, P.L.; Kobayashi, H. Interleukin-15 after Near-Infrared Photoimmunotherapy (NIR-PIT) Enhances T Cell Response against Syngeneic Mouse Tumors. *Cancers* 2020, 12, 2575. [CrossRef]
- 106. Maruoka, Y.; Furusawa, A.; Okada, R.; Inagaki, F.; Fujimura, D.; Wakiyama, H.; Kato, T.; Nagaya, T.; Choyke, P.L.; Kobayashi, H. Near-Infrared Photoimmunotherapy Combined with CTLA4 Checkpoint Blockade in Syngeneic Mouse Cancer Models. *Vaccines* 2020, *8*, 528. [CrossRef]
- 107. Katsube, R.; Noma, K.; Ohara, T.; Nishiwaki, N.; Kobayashi, T.; Komoto, S.; Sato, H.; Kashima, H.; Kato, T.; Kikuchi, S.; et al. Fibroblast activation protein targeted near infrared photoimmunotherapy (NIR PIT) overcomes therapeutic resistance in human esophageal cancer. *Sci. Rep.* 2021, *11*, 1693. [CrossRef]
- Shirasu, N.; Yamada, H.; Shibaguchi, H.; Kuroki, M.; Kuroki, M. Potent and specific antitumor effect of CEA-targeted photoimmunotherapy. Int. J. Cancer 2014, 135, 2697–2710. [CrossRef]
- 109. Maawy, A.A.; Hiroshima, Y.; Zhang, Y.; Heim, R.; Makings, L.; Garcia-Guzman, M.; Luiken, G.A.; Kobayashi, H.; Hoffman, R.M.; Bouvet, M. Near infra-red photoimmunotherapy with anti-CEA-IR700 results in extensive tumor lysis and a significant decrease in tumor burden in orthotopic mouse models of pancreatic cancer. *PLoS ONE* 2015, *10*, e0121989. [CrossRef]
- 110. Wei, W.; Jiang, D.; Ehlerding, E.B.; Barnhart, T.E.; Yang, Y.; Engle, J.W.; Luo, Q.Y.; Huang, P.; Cai, W. CD146-Targeted Multimodal Image-Guided Photoimmunotherapy of Melanoma. *Adv. Sci.* **2019**, *6*, 1801237. [CrossRef]
- 111. Wei, D.; Tao, Z.; Shi, Q.; Wang, L.; Liu, L.; She, T.; Yi, Q.; Wen, X.; Liu, L.; Li, S.; et al. Selective Photokilling of Colorectal Tumors by Near-Infrared Photoimmunotherapy with a GPA33-Targeted Single-Chain Antibody Variable Fragment Conjugate. *Mol. Pharm.* 2020, 17, 2508–2517. [CrossRef]
- 112. Hanaoka, H.; Nakajima, T.; Sato, K.; Watanabe, R.; Phung, Y.; Gao, W.; Harada, T.; Kim, I.; Paik, C.H.; Choyke, P.L.; et al. Photoimmunotherapy of hepatocellular carcinoma-targeting Glypican-3 combined with nanosized albumin-bound paclitaxel. *Nanomedicine* 2015, 10, 1139–1147. [CrossRef] [PubMed]
- 113. Isobe, Y.; Sato, K.; Nishinaga, Y.; Takahashi, K.; Taki, S.; Yasui, H.; Shimizu, M.; Endo, R.; Koike, C.; Kuramoto, N.; et al. Near infrared photoimmunotherapy targeting DLL3 for small cell lung cancer. *EBioMedicine* **2020**, *52*, 102632. [CrossRef] [PubMed]
- 114. Lum, Y.L.; Luk, J.M.; Staunton, D.E.; Ng, D.K.P.; Fong, W.P. Cadherin-17 Targeted Near-Infrared Photoimmunotherapy for Treatment of Gastrointestinal Cancer. *Mol. Pharm.* 2020, *17*, 3941–3951. [CrossRef]
- 115. Fujimoto, S.; Muguruma, N.; Okamoto, K.; Kurihara, T.; Sato, Y.; Miyamoto, Y.; Kitamura, S.; Miyamoto, H.; Taguchi, T.; Tsuneyama, K.; et al. A Novel Theranostic Combination of Near-infrared Fluorescence Imaging and Laser Irradiation Targeting c-KIT for Gastrointestinal Stromal Tumors. *Theranostics* **2018**, *8*, 2313–2328. [CrossRef]
- 116. Nishinaga, Y.; Sato, K.; Yasui, H.; Taki, S.; Takahashi, K.; Shimizu, M.; Endo, R.; Koike, C.; Kuramoto, N.; Nakamura, S.; et al. Targeted Phototherapy for Malignant Pleural Mesothelioma: Near-Infrared Photoimmunotherapy Targeting Podoplanin. *Cells* 2020, 9, 1019. [CrossRef] [PubMed]
- 117. Nagaya, T.; Nakamura, Y.; Sato, K.; Zhang, Y.F.; Ni, M.; Choyke, P.L.; Ho, M.; Kobayashi, H. Near infrared photoimmunotherapy with an anti-mesothelin antibody. *Oncotarget* **2016**, *7*, 23361–23369. [CrossRef] [PubMed]
- Furusawa, A.; Okada, R.; Inagaki, F.; Wakiyama, H.; Kato, T.; Furumoto, H.; Fukushima, H.; Okuyama, S.; Choyke, P.L.; Kobayashi, H. CD29 targeted near-infrared photoimmunotherapy (NIR-PIT) in the treatment of a pigmented melanoma model. *Oncoimmunology* 2022, 11, 2019922. [CrossRef]

- 119. Yasui, H.; Nishinaga, Y.; Taki, S.; Takahashi, K.; Isobe, Y.; Sato, K. Near Infrared Photoimmunotherapy for Mouse Models of Pleural Dissemination. *J. Vis. Exp.* **2021**, *23*, e61593. [CrossRef]
- Polikarpov, D.M.; Campbell, D.H.; Lund, M.E.; Lu, Y.; Lu, Y.; Wu, J.; Walsh, B.J.; Zvyagin, A.V.; Gillatt, D.A. The feasibility of Miltuximab®-IRDye700DX-mediated photoimmunotherapy of solid tumors. *Photodiagnosis Photodyn. Ther.* 2020, 32, 102064. [CrossRef]
- 121. Fukushima, H.; Kato, T.; Furusawa, A.; Okada, R.; Wakiyama, H.; Furumoto, H.; Okuyama, S.; Kondo, E.; Choyke, P.L.; Kobayashi, H. Intercellular adhesion molecule-1-targeted near-infrared photoimmunotherapy of triple-negative breast cancer. *Cancer Sci.* 2022, 113, 3180–3192. [CrossRef]
- 122. Walker, E.; Turaga, S.M.; Wang, X.; Gopalakrishnan, R.; Shukla, S.; Basilion, J.P.; Lathia, J.D. Development of near-infrared imaging agents for detection of junction adhesion molecule-A protein. *Transl. Oncol.* **2021**, *14*, 101007. [CrossRef] [PubMed]
- 123. Bao, R.; Wang, Y.; Lai, J.; Zhu, H.; Zhao, Y.; Li, S.; Li, N.; Huang, J.; Yang, Z.; Wang, F.; et al. Enhancing Anti-PD-1/PD-L1 Immune Checkpoint Inhibitory Cancer Therapy by CD276-Targeted Photodynamic Ablation of Tumor Cells and Tumor Vasculature. *Mol. Pharm.* 2019, *16*, 339–348. [CrossRef] [PubMed]
- 124. Li, F.; Mao, C.; Yeh, S.; Sun, Y.; Xin, J.; Shi, Q.; Ming, X. MRP1-targeted near infrared photoimmunotherapy for drug resistant small cell lung cancer. *Int. J. Pharm.* 2021, 604, 120760. [CrossRef]
- 125. Silic-Benussi, M.; Saponeri, A.; Michelotto, A.; Russo, I.; Colombo, A.; Pelizzo, M.G.; Ciminale, V.; Alaibac, M. Near infrared photoimmunotherapy targeting the cutaneous lymphocyte antigen for mycosis fungoides. *Expert Opin. Biol. Ther.* **2021**, *21*, 977–981. [CrossRef] [PubMed]
- 126. Mao, C.; Zhao, Y.; Li, F.; Li, Z.; Tian, S.; Debinski, W.; Ming, X. P-glycoprotein targeted and near-infrared light-guided depletion of chemoresistant tumors. *J. Control. Release* 2018, 286, 289–300. [CrossRef]
- 127. Zhang, C.; Gao, L.; Cai, Y.; Liu, H.; Gao, D.; Lai, J.; Jia, B.; Wang, F.; Liu, Z. Inhibition of tumor growth and metastasis by photoimmunotherapy targeting tumor-associated macrophage in a sorafenib-resistant tumor model. *Biomaterials* **2016**, *84*, 1–12. [CrossRef]
- 128. Kato, T.; Okada, R.; Furusawa, A.; Inagaki, F.; Wakiyama, H.; Furumoto, H.; Okuyama, S.; Fukushima, H.; Choyke, P.L.; Kobayashi, H. Simultaneously Combined Cancer Cell- and CTLA4-Targeted NIR-PIT Causes a Synergistic Treatment Effect in Syngeneic Mouse Models. *Mol. Cancer Ther.* 2021, 20, 2262–2273. [CrossRef]
- 129. Kato, T.; Okada, R.; Furusawa, A.; Wakiyama, H.; Furumoto, H.; Fukushima, H.; Okuyama, S.; Choyke, P.L.; Kobayashi, H. Comparison of the Effectiveness of IgG Antibody versus F(ab')(2) Antibody Fragment in CTLA4-Targeted Near-Infrared Photoimmunotherapy. *Mol. Pharm.* 2022, *19*, 3600–3611. [CrossRef]
- Wakiyama, H.; Furusawa, A.; Okada, R.; Inagaki, F.; Kato, T.; Furumoto, H.; Fukushima, H.; Okuyama, S.; Choyke, P.L.; Kobayashi, H. Opening up new VISTAs: V-domain immunoglobulin suppressor of T cell activation (VISTA) targeted nearinfrared photoimmunotherapy (NIR-PIT) for enhancing host immunity against cancers. *Cancer Immunol. Immunother.* 2022, 71, 2869–2879. [CrossRef]
- 131. Barnett, J.D.; Jin, J.; Penet, M.F.; Kobayashi, H.; Bhujwalla, Z.M. Phototheranostics of Splenic Myeloid-Derived Suppressor Cells and Its Impact on Spleen Metabolism in Tumor-Bearing Mice. *Cancers* **2022**, *14*, 3578. [CrossRef] [PubMed]
- 132. Kato, T.; Fukushima, H.; Furusawa, A.; Okada, R.; Wakiyama, H.; Furumoto, H.; Okuyama, S.; Takao, S.; Choyke, P.L.; Kobayashi, H. Selective depletion of polymorphonuclear myeloid derived suppressor cells in tumor beds with near infrared photoimmunotherapy enhances host immune response. *Oncoimmunology* **2022**, *11*, 2152248. [CrossRef] [PubMed]
- 133. Martinelli, E.; De Palma, R.; Orditura, M.; De Vita, F.; Ciardiello, F. Anti-epidermal growth factor receptor monoclonal antibodies in cancer therapy. *Clin. Exp. Immunol.* **2009**, *158*, 1–9. [CrossRef]
- 134. Maennling, A.E.; Tur, M.K.; Niebert, M.; Klockenbring, T.; Zeppernick, F.; Gattenlöhner, S.; Meinhold-Heerlein, I.; Hussain, A.F. Molecular Targeting Therapy against EGFR Family in Breast Cancer: Progress and Future Potentials. *Cancers* 2019, 11, 1826. [CrossRef] [PubMed]
- 135. Nagaya, T.; Sato, K.; Harada, T.; Nakamura, Y.; Choyke, P.L.; Kobayashi, H. Near Infrared Photoimmunotherapy Targeting EGFR Positive Triple Negative Breast Cancer: Optimizing the Conjugate-Light Regimen. *PLoS ONE* **2015**, *10*, e0136829. [CrossRef]
- 136. Nagaya, T.; Okuyama, S.; Ogata, F.; Maruoka, Y.; Knapp, D.W.; Karagiannis, S.N.; Fazekas-Singer, J.; Choyke, P.L.; LeBlanc, A.K.; Jensen-Jarolim, E.; et al. Near infrared photoimmunotherapy targeting bladder cancer with a canine anti-epidermal growth factor receptor (EGFR) antibody. *Oncotarget* 2018, *9*, 19026–19038. [CrossRef]
- 137. Oh, D.Y.; Bang, Y.J. HER2-targeted therapies-A role beyond breast cancer. Nat. Rev. Clin. Oncol. 2020, 17, 33–48. [CrossRef]
- 138. Gravalos, C.; Jimeno, A. HER2 in gastric cancer: A new prognostic factor and a novel therapeutic target. *Ann. Oncol.* 2008, 19, 1523–1529. [CrossRef]
- 139. Zhan, N.; Dong, W.G.; Tang, Y.F.; Wang, Z.S.; Xiong, C.L. Analysis of HER2 gene amplification and protein expression in esophageal squamous cell carcinoma. *Med. Oncol.* 2012, *29*, 933–940. [CrossRef]
- Ito, K.; Mitsunaga, M.; Arihiro, S.; Saruta, M.; Matsuoka, M.; Kobayashi, H.; Tajiri, H. Molecular targeted photoimmunotherapy for HER2-positive human gastric cancer in combination with chemotherapy results in improved treatment outcomes through different cytotoxic mechanisms. *BMC Cancer* 2016, *16*, 37. [CrossRef]
- 141. Hiroshima, Y.; Maawy, A.; Zhang, Y.; Guzman, M.G.; Heim, R.; Makings, L.; Luiken, G.A.; Kobayashi, H.; Tanaka, K.; Endo, I.; et al. Photoimmunotherapy Inhibits Tumor Recurrence After Surgical Resection on a Pancreatic Cancer Patient-Derived Orthotopic Xenograft (PDOX) Nude Mouse Model. Ann. Surg. Oncol. 2015, 22 (Suppl. S3), S1469–S1474. [CrossRef] [PubMed]

- Maawy, A.A.; Hiroshima, Y.; Zhang, Y.; Garcia-Guzman, M.; Luiken, G.A.; Kobayashi, H.; Hoffman, R.M.; Bouvet, M. Photoimmunotherapy lowers recurrence after pancreatic cancer surgery in orthotopic nude mouse models. *J. Surg. Res.* 2015, 197, 5–11. [CrossRef] [PubMed]
- 143. Ponta, H.; Sherman, L.; Herrlich, P.A. CD44: From adhesion molecules to signalling regulators. *Nat. Rev. Mol. Cell Biol.* 2003, 4, 33–45. [CrossRef]
- 144. Patel, S.P.; Kurzrock, R. PD-L1 Expression as a Predictive Biomarker in Cancer Immunotherapy. *Mol. Cancer Ther.* 2015, 14, 847–856. [CrossRef]
- 145. Colombo, M.P.; Piconese, S. Regulatory-T-cell inhibition versus depletion: The right choice in cancer immunotherapy. *Nat. Rev. Cancer* 2007, *7*, 880–887. [CrossRef]
- 146. Watanabe, S.; Noma, K.; Ohara, T.; Kashima, H.; Sato, H.; Kato, T.; Urano, S.; Katsube, R.; Hashimoto, Y.; Tazawa, H.; et al. Photoimmunotherapy for cancer-associated fibroblasts targeting fibroblast activation protein in human esophageal squamous cell carcinoma. *Cancer Biol. Ther.* 2019, 20, 1234–1248. [CrossRef]
- 147. Jin, J.; Barnett, J.D.; Krishnamachary, B.; Mironchik, Y.; Luo, C.K.; Kobayashi, H.; Bhujwalla, Z.M. Eliminating fibroblast activation protein-α expressing cells by photoimmunotheranostics. *bioRxiv* **2021**. [CrossRef]
- 148. Yoshida, R.; Tazawa, H.; Hashimoto, Y.; Yano, S.; Onishi, T.; Sasaki, T.; Shirakawa, Y.; Kishimoto, H.; Uno, F.; Nishizaki, M.; et al. Mechanism of resistance to trastuzumab and molecular sensitization via ADCC activation by exogenous expression of HER2-extracellular domain in human cancer cells. *Cancer Immunol. Immunother.* **2012**, *61*, 1905–1916. [CrossRef]
- 149. Shimoyama, K.; Kagawa, S.; Ishida, M.; Watanabe, S.; Noma, K.; Takehara, K.; Tazawa, H.; Hashimoto, Y.; Tanabe, S.; Matsuoka, J.; et al. Viral transduction of the HER2-extracellular domain expands trastuzumab-based photoimmunotherapy for HER2-negative breast cancer cells. *Breast Cancer Res. Treat* 2015, 149, 597–605. [CrossRef] [PubMed]
- Ishida, M.; Kagawa, S.; Shimoyama, K.; Takehara, K.; Noma, K.; Tanabe, S.; Shirakawa, Y.; Tazawa, H.; Kobayashi, H.; Fujiwara, T. Trastuzumab-Based Photoimmunotherapy Integrated with Viral HER2 Transduction Inhibits Peritoneally Disseminated HER2-Negative Cancer. *Mol. Cancer Ther.* 2016, 15, 402–411. [CrossRef]
- 151. Hsu, M.A.; Okamura, S.M.; De Magalhaes Filho, C.D.; Bergeron, D.M.; Rodriguez, A.; West, M.; Yadav, D.; Heim, R.; Fong, J.J.; Garcia-Guzman, M. Cancer-targeted photoimmunotherapy induces antitumor immunity and can be augmented by anti-PD-1 therapy for durable anticancer responses in an immunologically active murine tumor model. *Cancer Immunol. Immunother.* 2022, 72, 151–168. [CrossRef] [PubMed]
- 152. Shirasu, N.; Shibaguchi, H.; Yamada, H.; Kuroki, M.; Yasunaga, S. Highly versatile cancer photoimmunotherapy using photosensitizer-conjugated avidin and biotin-conjugated targeting antibodies. *Cancer Cell Int.* 2019, 19, 299. [CrossRef] [PubMed]
- Bullous, A.J.; Alonso, C.M.; Boyle, R.W. Photosensitiser-antibody conjugates for photodynamic therapy. *Photochem. Photobiol. Sci.* 2011, 10, 721–750. [CrossRef] [PubMed]
- 154. Serebrovskaya, E.O.; Edelweiss, E.F.; Stremovskiy, O.A.; Lukyanov, K.A.; Chudakov, D.M.; Deyev, S.M. Targeting cancer cells by using an antireceptor antibody-photosensitizer fusion protein. *Proc. Natl. Acad. Sci. USA* 2009, 106, 9221–9225. [CrossRef] [PubMed]
- 155. Matsuoka, K.; Sato, M.; Sato, K. Hurdles for the wide implementation of photoimmunotherapy. *Immunotherapy* **2021**, *13*, 1427–1438. [CrossRef]
- 156. Hoffman, R.M. Patient-derived orthotopic xenografts: Better mimic of metastasis than subcutaneous xenografts. *Nat. Rev. Cancer* **2015**, *15*, 451–452. [CrossRef]
- 157. Francia, G.; Cruz-Munoz, W.; Man, S.; Xu, P.; Kerbel, R.S. Mouse models of advanced spontaneous metastasis for experimental therapeutics. *Nat. Rev. Cancer* **2011**, *11*, 135–141. [CrossRef]
- Furumoto, H.; Okada, R.; Kato, T.; Wakiyama, H.; Inagaki, F.; Fukushima, H.; Okuyama, S.; Furusawa, A.; Choyke, P.L.; Kobayashi, H. Optimal Light Dose for hEGFR-Targeted Near-Infrared Photoimmunotherapy. *Cancers* 2022, 14, 4042. [CrossRef]

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