



Review

The Role of the Oral Microbiome in the Development of Diseases

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Abstract: Periodontal disease (PD) is a complex and infectious illness that begins with a disruption of bacterial homeostasis. This disease induces a host inflammatory response, leading to damage of the soft and connective tooth-supporting tissues. Moreover, in advanced cases, it can contribute to tooth loss. The aetiological factors of PDs have been widely researched, but the pathogenesis of PD has still not been totally clarified. There are a number of factors that have an effect on the aetiology and pathogenesis of PD. It is purported that microbiological, genetic susceptibility and lifestyle can determine the development and severity of the disease. The human body's defence response to the accumulation of plaque and its enzymes is known to be a major factor for PD. The oral cavity is colonised by a characteristic and complex microbiota that grows as diverse biofilms on all mucosal and dental surfaces. The aim of this review was to provide the latest updates in the literature regarding still-existing problems with PD and to highlight the role of the oral microbiome in periodontal health and disease. Better awareness and knowledge of the causes of dysbiosis, environmental risk factors and periodontal therapy can reduce the growing worldwide prevalence of PDs. The promotion of good oral hygiene, limiting smoking, alcohol consumption and exposure to stress and comprehensive treatment to decrease the pathogenicity of oral biofilm can help reduce PD as well as other diseases. Evidence linking disorders of the oral microbiome to various systemic diseases has increased the understanding of the importance of the oral microbiome in regulating many processes in the human body and, thus, its impact on the development of many diseases.

Keywords: periodontal diseases; oral microbiome; oral health; oral diseases; systemic diseases



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1. Introduction

Periodontitis is one of the most common infectious diseases [1], which affects 10–50% of the global population, depending on its severity [2]. Gingivitis is a mild and reversible form of periodontal disease (PD), and if not treated properly, this can progress to periodontitis [3]. PD is a complex and infectious illness that begins with the disruption of bacterial homeostasis. This disease induces a host inflammatory response, leading to damage to the soft and connective tooth-supporting tissues [4–6]. Moreover, in advanced cases, it can contribute to tooth loss [7]. The aetiological factors of PDs have been widely researched, but the pathogenesis of PD has still not been totally clarified. There are a number of factors that have an effect on the aetiology and pathogenesis of PD. It is purported that microbiological, genetic susceptibility and lifestyle can determine the development and severity of the disease [8,9]. The human body's defence response to the accumulation of plaque and its enzymes is known to be a major factor for PD [10]. Microbial plaque is a biofilm that forms on the teeth and gingiva and is one of the crucial causes of PD [11]. There are other individual risk factors contributing to PD such as obesity, poor oral hygiene, stress, a diet low in vitamins C and D and tobacco use [6,10,12–14]. The interplay between cigarette smoking and PD has been analysed in numerous studies, suggesting that smoking is a significant environmental risk factor for PD. One component of cigarette smoke, nicotine, can

be related to changing clinical aspects and development [6,13]. Additionally, many studies noticed that PD has an impact on the progression of various systemic diseases such as osteoporosis, atherosclerosis, diabetes, cardiovascular diseases and ischemic cardiomyopathy, all of which may aggravate the disease [15,16]. Moreover, other studies revealed the opposite situation, showing that systemic disease can exacerbate PD. In addition, taking medications such as steroids, anti-epilepsy drugs and cancer therapy drugs can also increase PD. Susceptibility to this disorder is connected to the triggering of host antibacterial defence mechanisms [17]. Numerous studies have indicated the association between genetic factors with PD. Cytokines and their genetic polymorphisms influence susceptibility to this disease and its severity. Nevertheless, Nibali et al. presented interesting results of their work, with the inheritance of periodontitis being assessed as OR 0.38 (95% CI, 0.34–0.43) in twin studies and OR 0.15 (95% CI, 0.06–0.24) in other family research [18]. It was observed that changes in the oral microbiome composition naturally increase with age [19], which may be related to the greater susceptibility of the elderly to chronic periodontitis [20]. Periodontal disorders intensify problems with chewing, the function of speech and aesthetics, which definitely worsen the quality of life [19].

A new model of the pathogenesis of periodontal disorders assumes that the disease involves a more diverse microflora associated with periodontitis than previously thought. The disease is caused by the synergy of multiple microbes and dysbiosis, which disrupt the ecologically balanced biofilm associated with periodontal homeostasis and are not the result of individual pathogens [21]. Under healthy conditions, the oral microbiome exhibits a well-balanced, dynamic ecosystem [22]. Dysbiosis of the oral microbiome means an imbalance in relative abundance or an impact on microbial species, which contribute to the disease development of susceptible patients [23].

2. Microbiome in the Oral Health

The term “microbiome” was created by Joshua Lederberg and refers to the community of symbiotic, commensal and pathogenic microorganisms [24]. The composition and interactions of any microbiome contribute to overall health, being a key factor in oral health [25]. The oral cavity is colonised by a characteristic and complex microbiota that grows as diverse biofilms on all mucosal and dental surfaces [26]. There are more than 700 species of bacteria, fungi, viruses, archaeobacteria and protozoa in the oral cavity [20]. Bacteria are the most well-researched microorganisms in the oral cavity [27], but only 57% of bacterial species in the oral cavity have been officially named [28]. In health, oral microflora mainly consists of facultative anaerobic Gram-positive bacteria [29]. The oral fungal microbiome (mycobiome) is a significant component of the oral microbiome. The *Candida* genus is present in about 25–75% of healthy individuals as a commensal organism [30]. *Candida albicans* is one of the most crucial, prevalent fungal species. Under certain, favourable conditions, *Candida* species, as opportunistic pathogens, can cause infections of the oral mucosa [31]. The oral microbiota usually live in harmony with the host and provide important benefits that contribute to overall health. The microorganisms in oral biofilms do not exist as single cells but live in close proximity to one another [27]. The microbial interactions can be synergistic or antagonistic [32,33]. Moreover, the oral environment has an impact on the composition of the microbiome. If some changes in local conditions occur, they can influence interactions between microorganisms in the mouth and increase the risk of PD. The composition of the oral microbiome has been widely explored by using metagenomics and metatranscriptomics [2]. Using these methods, Belstrøm et al. observed the transcriptional activity of prevalent *Streptococcus* species under healthy conditions and periodontitis. The researchers discovered that the transcriptional activity of *Streptococcus* species was higher in health and reduced in PD [34]. Of particular note, *Streptococcus* species are Gram-positive, aerobic to facultatively anaerobic bacteria that are part of the normal flora of the oral cavity. In health, new species of *Streptococcus dentisani* and *Streptococcus salivarius*, which have potential probiotic features, are associated with the treatment of miscellaneous oral diseases such as periodontal disorders, among others [35–37].

Recently, there has been growing interest in the use of probiotics to treat PDs. Probiotics are defined as “living microorganisms that can have a beneficial effect on the host when taken in sufficient doses” [38]. Their function is to regulate host immune function, restore balance and maintain homeostasis in the mouth [3]. Good results of probiotics in improving oral health have been noticed not only in periodontal disorders [39] but also in dental caries [40], *Candida* infection [41] and halitosis [42]. The genus *Lactobacillus* is well known as a health-promoting probiotic in periodontal therapy. *Bifidobacterium*, *Streptococcus* and *Weissella* are also known probiotics, which play a positive role in oral care. Other species such as *Bacillus subtilis* and *Saccharomyces cerevisiae* also have a good impact on the oral cavity [3]. In addition, some strains of bacteria isolated from the oral cavity have been produced commercially as probiotics, including *Lactobacillus reuteri*, *Lactobacillus brevis* and *Streptococcus salivarius* [25,43]. Kawai et al. [44] suggested that *Limosilactobacillus (Lactobacillus) fermentum* ALAL020 may be a future probiotic candidate. This bacterium produces a cyclic dipeptide with antibacterial activity against *Porphyromonas gingivalis* and *Prevotella intermedia* [44]. Currently, the beneficial effects of synbiotics on health are of great interest to scientific research. Synbiotics are defined as “a mixture comprising live microorganisms and substrate(s) selectively utilized by host microorganisms that confers a health benefit on the host” [43]. It has been observed that administering a synbiotic in combination with probiotics can help prevent and treat certain metabolic disorders. However, there is little evidence for this [45]. Duraisamy et al. noticed that synbiotics can diminish *Streptococcus mutans* levels in children’s saliva but are less effective compared to probiotics [46].

3. Oral Microbiome in Periodontal Diseases

The triad of oral anaerobic bacteria, the so-called “red complex” (*Porphyromonas gingivalis*, *Treponema denticola* and *Tannerella forsythia*), have historically been considered as the basic infectious organisms associated with periodontitis [27]. However, this has been identified in culture-based studies, and many of the wide variety of bacteria present in samples were overlooked [47]. Nevertheless, only a few bacteria, namely *P. gingivalis*, *Aggregatibacter actinomycetemcomitans*, *Tannerella forsythia*, *Prevotella intermedia* and *Fusobacterium nucleatum*, were confirmed to initiate and progress PDs [48]. In addition, *Candida albicans* is one of the most crucial fungal residents of the oral microbiome. This commensal coloniser protects *P. gingivalis* from being recognised by the host’s immune cells and can contribute to bacterial infections of the gums [29]. Contemporary periodontology concentrates not only on the pathogenicity of dental plaque but also on the interplay between oral microorganisms and the host [3]. Microflora perturbances then lead to gingivitis and, eventually, periodontitis. Factors such as the availability of oxygen, nutrients and changes in pH may contribute to disorders in homeostasis in the oral cavity as well as systemic diseases [47,49]. Modification in the oral microbiome can lead to the expansion of microorganisms and provides excellent conditions for the growth of opportunistic microbes [10]. In addition, perturbations in the periodontal microbiota are associated with an alteration from a symbiotic to dysbiotic microbial community. The symbiotic structure includes facultative bacteria such as *Actinomyces* and *Streptococcus*, which then shifts to mainly anaerobic types (such as the phyla *Firmicutes*, *Proteobacteria*, *Spirochaetes*, *Bacteroidetes* and *Synergistetes*) [48]. The transition of microbial composition precedes the clinical symptoms of PD [50]. It is widely known that factors such as residing microorganisms, age, general health, lifestyle, and nutritional status have an impact on oral health [50].

In approximately 90% of cases of PD, halitosis (malodour) can be noticed in the mouth [51]. Halitosis is mainly caused by *P. gingivalis*, *T. denticola*, *Fusobacterium* and *T. forsythia*, the same species that have been associated with PD. The process of bacterial degradation of sulphur-containing amino acids into volatile gases leads to oral malodour. Halitosis is caused by the biodegradation of sulphur-containing amino acids and the production of volatile sulphur compounds [52]. Moreover, poor oral hygiene, bacterial coating of the tongue and periodontal disorders such as gingivitis, periodontitis and caries can contribute to halitosis [53]. Malodour can also be affected by dietary habits, such as smoking, alcohol consumption, obesity, diabetes, stress and advanced age [54–56]. In

In addition, risk factors such as age and tooth decay can also lead to malodour in children [57], but it is difficult to distinguish between oral bacteria that cause odour in adolescents. Three types of bacteria including *Fusobacterium*, *Veillonella* and *Prevotella* are dominant in children with halitosis [56]. A noteworthy finding observed by Wu et al. is that the oral microbiome was altered, and more abundant species were present among obese people suffering from malodour compared to healthy persons [58]. Tobacco use has a similar effect and may also alter the diversity of the oral microflora. Yitzhaki et al. observed the relationships between patients wearing dentures and halitosis [59]. The researcher noticed higher bacterial diversity in the oral microflora among patients with halitosis wearing dentures, observing meaningful differences from the control group. There were bacterial taxa, including 117 species, 29 genera (mainly *Leptotrichia*, *Megasphaera*, *Atopobium* and others) and 9 phyla (*Fusobacteria*, *Firmicutes*) detected [59]. An important research finding was that *Candida* species accounted for the largest percentage of microbes among smokers with halitosis [56]. Zhang et al. suggested that halitosis can be detected long before clinical symptoms appear as a result of changes in the microbiome of the tongue coating. Nevertheless, alterations in the tongue coating microbiome can be used as biomarkers of an early stage of halitosis and can help find better strategies for the diagnosis, prognosis and treatment [60]. Although periodontitis is associated with a variety of microorganisms, *Fusobacterium nucleatum* is a known, predominant periodontal pathogen that can influence other bacteria and form an inflammatory microenvironment. In addition, *F. nucleatum* may modulate and enhance the invasive potential of *P. gingivalis* [61]. Thus, it can promote and accelerate the development of periodontitis. Moreover, *Fusobacterium nucleatum* can cause local halitosis and pulp infections and may systemically promote the development or progression of oral cancer and other extraoral diseases [62]. Kang et al. [63] explored and identified three types of *Weissella cibaria* from human saliva, which produce hydrogen peroxides. These isolates can inhibit volatile sulphur compounds formed by *F. nucleatum*. In addition, *W. cibaria* can block the production of interleukin-6 and interleukin-8 by oral epithelial cells caused by *F. nucleatum*. Suzuki et al. [64] showed that *Lactobacillus saliva* WB21 buccal tablets can specifically reduce the amount of *F. nucleatum* in patients with halitosis. This means that the use of *W. cibaria* and *Lactobacillus saliva* WB21 as probiotics can be a beneficial method of combating bad breath and controlling PD [62,64].

Subgingival communities of microbes, including fungi, archaea and viruses, can lead to periodontitis-related dysbiosis [8]. The microbial communities are responsible for the pathological processes that have an impact on the periodontium. Although alterations in the composition and function of subgingival bacteria have been extensively explored, how the succession of microorganisms and the transformation of health into disease proceeds is still not fully understood. Periodontal health is frequently defined as opposite to PD, in the absence of any clinical symptoms of disorder [65]. The development of the oral microflora is known to involve interactions between the host's genetics and the host's immune system, and changes in the composition of the microbiome depend on exposure to environmental factors [10].

During periodontal health, the health-associated species dominate the local microbiome. The development of gingivitis or PD is associated with increased biomass and the appearance of pathogenic species. The further progression of the disease is characterized by a shift in the microbiome balance into PD-associated pathogens. Alterations in composition and species diversity may lead to the identification of potential biomarkers in the diagnosis of PDs [55]. In addition, biomarkers in saliva may reflect various conditions in the oral cavity connected with periodontitis. Saliva easily collects and can show the condition of the entire mouth. For example, nitric oxide (NO) is known as a biological marker, which is related to the aetiopathogenesis of oral diseases [66]. Reher et al. observed increased levels of NO in patients with periodontal disorders in comparison to healthy people [67]. Furthermore, it was noticed that the level of NO was correlated with periodontitis severity. The researcher also suggested that the levels of NO were linked with the deterioration

of periodontal parameters such as probing depth and were the result of an inflammatory response induced by bacteria.

4. Oral Microbiome in Systemic Diseases

Under different conditions, bacterial flora has the ability to change the balance between health and sickness, both locally and systemically. Microorganisms present in the mouth interact with themselves and with the host in illness and health [68]. The oral cavity is the entry point and direct way to the lungs and the digestive system, so the microbiomes of these structures are interconnected across the human body. That explains why the oral microbiome is associated with many systemic diseases [47]. Homeostasis disorders due to food habits and poor oral hygiene not only lead to oral diseases such as caries or periodontal problems but also to other systemic diseases and cancers. The direct route and good access of the oral microbiome to the respiratory system explain the link between oral disorders and lung diseases, such as respiratory tract infection, bacterial pneumonia, chronic obstructive pulmonary disease (COPD) and cystic fibrosis [69].

Previous studies have shown that disturbances in the oral microbiome can lead to abnormalities in the airway microbiome, which can cause an abnormal local immune response and chronic inflammation in the airways and the onset of chronic obstructive pulmonary disease (COPD) [70]. The inflammatory process caused by bacterial infection leads to impaired lung defence mechanisms, contributing to progressive lung damage and the loss of lung function, which is characteristic of COPD [71]. In COPD, there is a higher incidence of bacterial colonization by potential respiratory pathogens, such as *Pseudomonas aeruginosa* and bacteria of the genus *Actinomyces* [72].

Pseudomonas aeruginosa is an important pathogen in cystic fibrosis patients. Whiley et al. have found that in vitro *Streptococcus* species could modulate the production of virulence factors (elastase and pyocyanin) by *P. aeruginosa* [73]. It has also been shown that the pathogenicity of *P. aeruginosa* can be inhibited by *S. oralis* through the production of hydrogen peroxide (H₂O₂) [74].

Furthermore, Haran et al. presented intriguing findings that the dysbiosis of the oral microbiome may affect the duration of COVID-19 symptoms [75]. *Prevotella* species have been found in abundance in COVID-19 patients. Furthermore, these species are considered to produce proteins that can contribute to SARS-CoV-2 infection and may aggravate COVID-19 [76]. *Veillonella* strains are also capable of eliciting inflammatory responses. This genus induces IL-6 (Interleukin-6) [77], whereas *Prevotella* species activate TLR-2 and the expression of IL-23 and IL-1 [78].

Studies in recent years have shown that bacteria produce a number of compounds that cause the development of systemic inflammatory responses that impair the blood–brain barrier (BBB), exacerbating neuroinflammation and ultimately neurodegeneration [79–81]. The microbiota produces a number of neuromediators such as serotonin, kynurenine, melatonin, GABA (gamma-aminobutyric acid), tryptophan, catecholamines, histamine and acetylcholine [82,83]. Abnormalities in the serotonin and kynurenine pathway of tryptophan metabolism have been detected in patients with neurodegenerative diseases including Alzheimer's disease (AD) [84]. Dysregulation of the kynurenine pathway of tryptophan metabolism may be one of the main factors contributing to AD development [85,86]. Other metabolites produced by the microbiota that can affect brain function and blood–brain barrier permeability are short-chain fatty acids (SCFAs): acetate, butyrate and propionate [87,88]. SCFAs can affect transmission processes in the central nervous system and thus regulate cognitive function. LPS and amyloids secreted by bacteria are involved in the process of neurodegeneration [89,90]. Many bacterial species produce extracellular amyloid fibres to form a biofilm [91]. Bacterial and brain amyloids are biologically similar in structure, composition and physicochemical characteristics [90]. Previous studies have shown that secretory products from the microbiome exert strong pro-inflammatory effects by activating complement and other components of the immune response, leading to an increased synthesis of pro-inflammatory cytokines and the development of neuroinflamma-

tion in the brain [92,93]. This intensifies amyloid aggregation and inflammatory responses. Both bacterial amyloid proteins and LPS are potent activators of the chronic inflammation in the cerebral rim observed in AD patients [94]. Dominy et al. [95] observed a correlation between *P. gingivalis* and the progression of Alzheimer's disease. The presence of this bacterium was noticed in the brain of AD patients. During studies in mice, it was also discovered that the infection of *P. gingivalis* led to brain colonisation and the growth of components of amyloid plaques. Nevertheless, *P. gingivalis* produces neurotoxic gingipain proteases, which inhibit tau function (a hallmark of AD). This means that gingipain inhibitors could be used to treat neurodegeneration in AD. Furthermore, oral species of the phylum *Spirochaetes* also form amyloid plaques and play a role in the progression of dementia in AD. In addition, these organisms avoid host defence and create more atypical, resistant forms and biofilms, which contribute to the maintenance of chronic infection and higher resistance to treatment [96] (Figure 1, Tables 1 and 2).

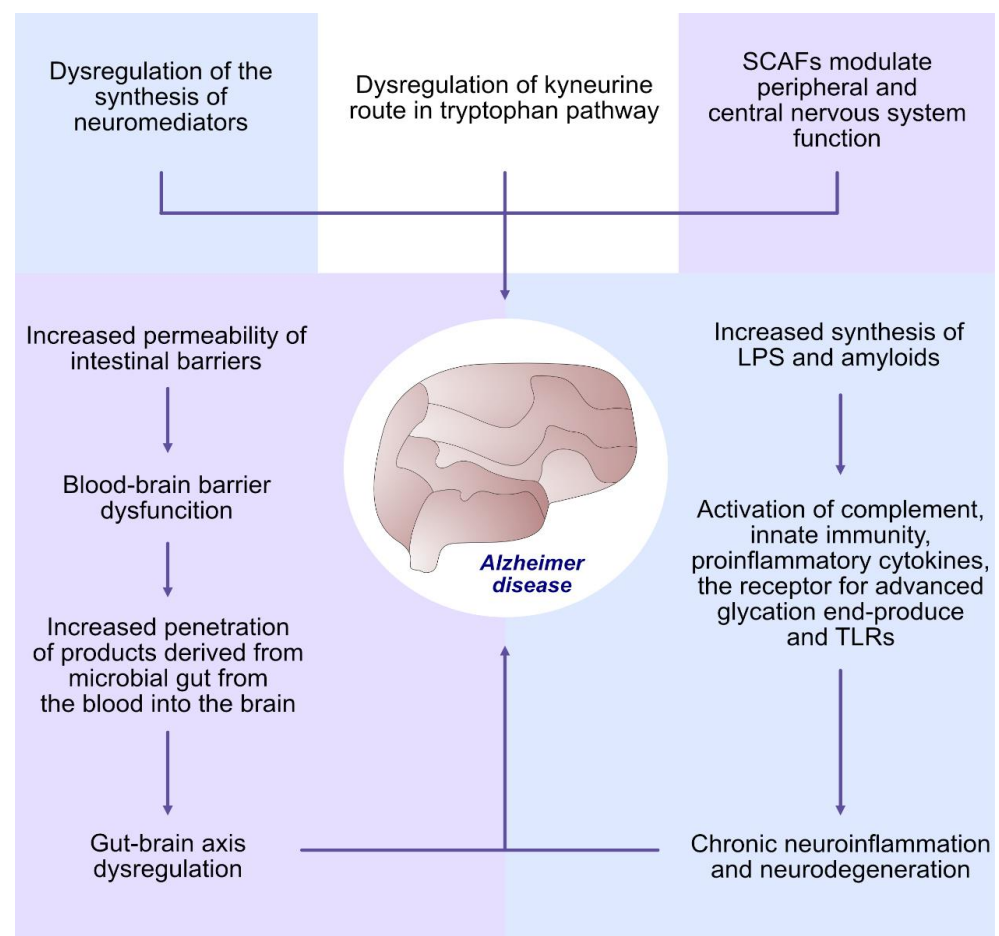


Figure 1. The involvement of oral microbiome dysbiosis in the development of Alzheimer's disease. SCFAs—short-chain fatty acids, TLRs—toll-like receptors, LPS—lipopolysaccharide.

Table 1. The associations between oral bacteria and diseases.

General Diseases	Bacteria	References
Respiratory tract infection Chronic obstructive pulmonary disease Cystic fibrosis	<i>Porphyromonas gingivalis</i> <i>Aggregatibacter actinomycetemcomitans</i> <i>Fusobacterium nucleatum</i> <i>Chlamydia pneumoniae</i>	[69–73]
Alzheimer’s disease	<i>Prevotella intermedia</i> <i>Tannerella forsythia</i> <i>Aggregatibacter actinomycetemcomitans</i> <i>Porphyromonas gingivalis</i> <i>Fusobacterium nucleatum</i>	[79–93]
Cardiovascular diseases (atherosclerosis/coronary diseases)	<i>Porphyromonas gingivalis</i> <i>Treponema denticola</i> <i>Aggregatibacter actinomycetemcomitans</i> <i>Prevotella intermedia</i> <i>Tannerella forsythia</i>	[97–106]
Diabetes and insulin resistance	<i>Porphyromonas gingivalis</i> <i>Aggregatibacter actinomycetemcomitans</i> <i>Fusobacterium nucleatum</i>	[107–110]
Rheumatoid arthritis	<i>Porphyromonas gingivalis</i> <i>Aggregatibacter actinomycetemcomitans.</i>	[111–125]
Pancreatic cancer Colorectal carcinoma	<i>Neisseria elongata</i> <i>Granulicatella adiacens</i> <i>Porphyromonas gingivalis</i> <i>Fusobacterium nucleatum</i>	[126–130]

The potential impact of periodontal infection on cardiovascular diseases has been the subject of much research [97,98]. Teles et al. undertook the challenge of explaining possible associations with an increased risk of cardiovascular diseases and periodontal disorders. However, this process is still poorly clarified [99]. Much is known about individual pathogens connected with periodontitis, rather than the mechanisms describing the association between PDs and cardiovascular disease. It has been shown that a number of compounds produced by the microbiome can enhance the development of atherosclerotic lesions in the vessels. Such compounds include trimethylamine (TMA), which is oxidized by monooxygenase to TMAO (Trimethylamine N-oxide) [100]. TMAO enhances foam cell formation and increases VCAM-1 (Vascular cell adhesion molecule-1) expression, which enhances monocyte adhesion to the endothelium [101]. There is also the activation of the protein kinase C (PKC) and nuclear factor- κ B (NF- κ B) pathway, which disrupts endothelial cell function and results in the development of atherosclerotic lesions [102,103]. Proatherogenic effects are also exhibited by SCFAs, which affect the processes of chemotaxis and phagocytosis, induce the formation of reactive oxygen species and activate monocytes and macrophages [104,105]. Another component that exacerbates atherogenesis may be LPS present on the cell membrane of bacteria. LPS increases the expression of chemokines and adhesion molecules, enhances the formation of foam cells and increases the adhesion of monocytes to endothelial cells [106]. In addition, LPS can also bind to toll-like receptor 4 (TLR4) on the surface of immunocompetent cells and induce the secretion of pro-inflammatory cytokines (TNF, IL-6) [105]. This leads to the development of inflammation in the vessels and the formation of atherosclerotic plaque, affecting its stability (Figure 2, Table 2).

Table 2. The involvement of oral microbiome dysbiosis in the development of diseases.

General Diseases	Mechanisms	References
Atherosclerosis and coronary artery disease	LPS induces the expression of chemokines and cell adhesion molecules Increased production of trimethylamine Increased foam cell generation Promoting monocyte adherence by up-regulating the level of vascular cell adhesion molecule-1 (VCAM-1) Activation of the protein kinase C (PKC) and nuclear factor- κ B (NF- κ B) pathways Damage to endothelial cells Disturbances in mitochondrial repair and myocardial metabolism Disturbances in bile acid circulation Disturbances in cholesterol and lipid metabolism Enhanced synthesis of pro-inflammatory cytokines Inflammatory response in endothelium Promotion of atherosclerotic plaque formation Plaque instability	[97–106]
Alzheimer's disease	Dysregulation of the synthesis of neuromediators: serotonin, kynurenine, melatonin, GABA, catecholamines, histamine and acetylcholine Dysregulation of kynurenine route in tryptophan pathway Short-chain fatty acids (SCFAs), (acetate, butyrate and propionate) modulate peripheral and central nervous system function Increased permeability of intestinal barriers Blood–brain barrier dysfunction Increased penetration of products derived from microbial gut from the blood into the brain Gut–brain axis dysregulation Increased synthesis of LPS and amyloids Activation of complement, innate immunity, pro-inflammatory cytokines, the receptor for advanced glycation end-products (RAGE) and TLRs. Chronic neuroinflammation and neurodegeneration	[79–93]
Diabetes and insulin resistance	Short-chain fatty acids act on parasympathetic activity to increase food intake Stimulation of TLR-4 by bacterial LPS induces inflammatory response Disturbances in bile acid circulation Disturbances in cholesterol and lipid metabolism Enhanced synthesis of pro-inflammatory cytokines Chronic systemic inflammation Enhanced oxidative stress Insulin resistance	[107–110]
Rheumatoid arthritis	Induction of anti-CCP antibodies Hypercitrullination in neutrophils Increased production of IL-17 Formation of Th17 cells	[111–125]
Colorectal carcinoma	Dysfunction in mucosal homeostasis Dysfunction in the gut epithelial barrier Increased intestinal permeability Increased synthesis of pro-inflammatory cytokines Increased cellular proliferation Changes in β -catenin and Wnt signalling	[126–130]

In addition, the microbiome has been shown to affect lipid and carbohydrate metabolism. It has been shown that SCFAs can modulate pancreatic β -cell function and insulin production, contributing to the development of diabetes [106]. SCFAs stimulate parasympathetic nervous system functions to increase food intake [107]. Products of the microbiome can also cause disturbances in bile acid circulation [108]. Microbiome dysbiosis may also stimulate chronic systemic inflammation and enhance oxidative stress, leading to insulin resistance and the development of diabetes [109] (Figure 3, Table 2).

Xiao et al. presented the interplay between the oral microbiome, diabetes and PDs [110]. The researcher observed that the pro-inflammatory cytokine IL-17 is associated with periodontitis and that its inhibition has an impact on the pathogenicity of the diabetic microbiome. Moreover, diabetes exacerbates periodontitis, while periodontitis causes a pathogenic alteration in the microbiome. This biome change contributes to the susceptibility and severity of PD. Treatment with anti-IL-17 antibodies may alleviate symptoms.

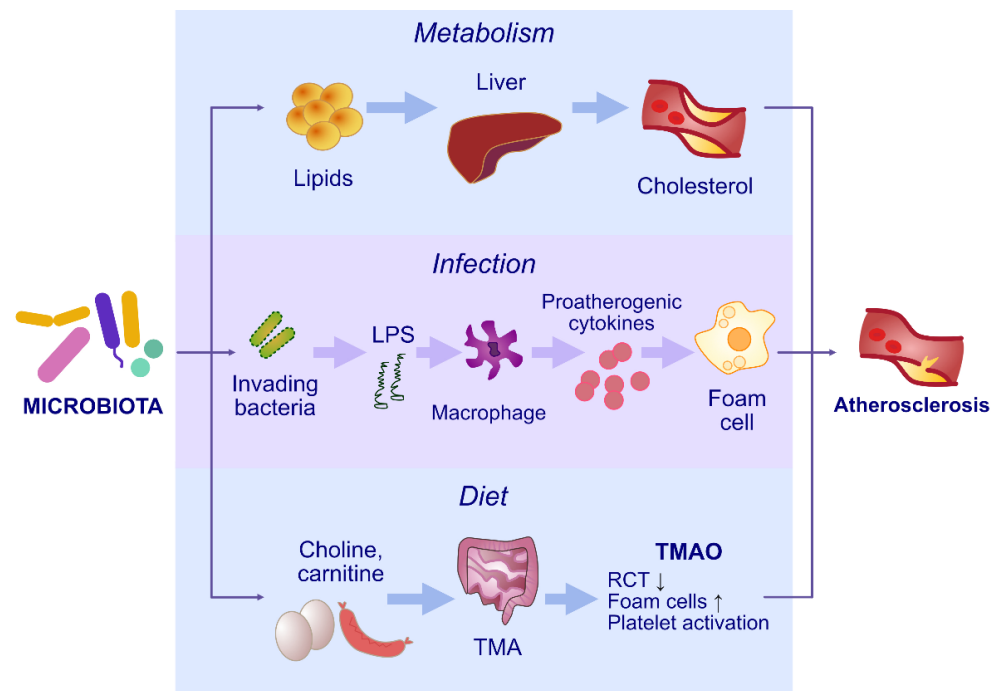


Figure 2. The involvement of oral microbiome dysbiosis in the development of atherosclerosis. LPS—lipopolysaccharide, TMAO—trimethylamine N-oxide, TMA—trimethylamine, RCT—reverse cholesterol transport.

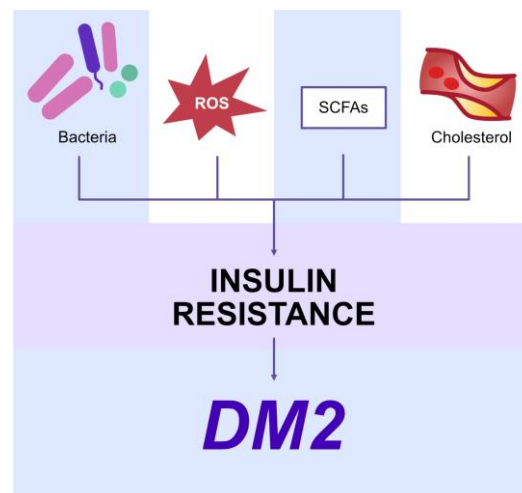


Figure 3. The involvement of oral microbiome dysbiosis in the development of diabetes mellitus. ROS—reactive oxygen species, SCFAs—short-chain fatty acids, DM2—diabetes mellitus type 2.

Recent studies have shown that periodontal disease is correlated with an increased risk of rheumatoid arthritis (RA) in humans and in mouse models of arthritis [111]. Rheumatoid arthritis is a systemic inflammatory disease that leads to joint destruction. The pathogenesis of this disease has been shown to be complex, with both genetic and environmental factors involved in its development. One of the environmental factors involved in the development of RA has been shown to be the microbiome [112]. Studies in recent years have shown a higher incidence of RA in patients with periodontal disease [113,114]. In addition, it has been shown that the severity of periodontal disease correlates with the activity of the disease process in RA patients and that treating the symptoms of periodontal disease results in the alleviation of RA symptoms [115,116]. These data suggest that bacteria associated with the development of periodontal disease may be involved in the pathogenesis of RA.

One of the main bacteria involved in the development of periodontal disease is *Porphyromonas gingivalis*. It has been shown that this bacterium can induce the formation of anti-CCP antibodies in RA patients [117–120]. In addition, *Porphyromonas gingivalis* can enhance the production of IL-17, a cytokine that plays an important role in the development of inflammation in RA [121]. *Porphyromonas gingivalis* was also found to enhance the formation of Th17 cells involved in the pathogenesis of RA [121]. Another bacterium that is thought to be associated with the pathogenesis of RA is *Aggregatibacter actinomycetemcomitans*. This bacterium has been shown to induce the formation of citrullinated autoantigens, which play a key role in the development of RA [122,123]. König et al. showed that leukotoxin-A produced by *A. actinomycetemcomitans* enhances hypercitrullination in neutrophils, an important element in the development of a pathological immune response in RA [124]. In addition, antibodies to *A. actinomycetemcomitans* and leukotoxin-A have been found in RA patients [125]. In conclusion, periodontal bacteria such as *Porphyromonas gingivalis* and *Aggregatibacter actinomycetemcomitans* may be involved in the development of RA by contributing to the production of autoantibodies and the process of autoimmunity. Further research is needed on the involvement of the microbiome in the pathogenesis of RA (Tables 1 and 2, Figure 4).

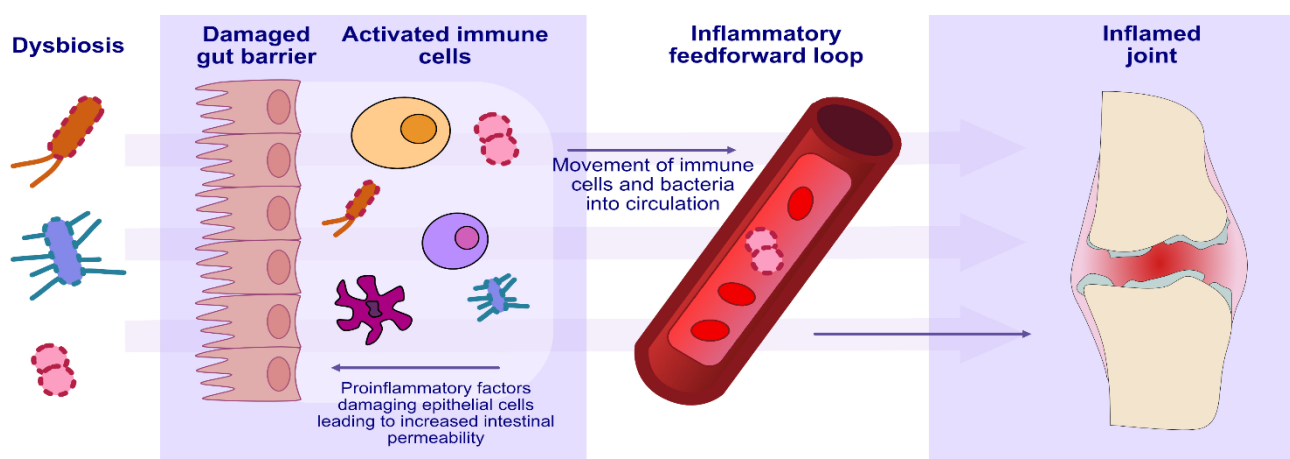


Figure 4. The involvement of oral microbiome dysbiosis in the development of rheumatoid arthritis.

The oral microbiome plays a significant role not only in maintaining oral health but also in maintaining balance throughout the body. Many studies presented findings showing that the dysbiosis of oral microbiota may also contribute to the development of cancers [126,127]. Farrell et al. [128] noticed reduced levels of *Neisseria elongata* and *Streptococcus mitis* in the salivary microbiome in patients with pancreatic cancer compared to healthy individuals, while the levels of *Granulicatella adiacens* were significantly higher in patients with pancreatic cancer. Previous studies have also shown that the dysbiosis of the oral microbiome may be involved in the development of colorectal cancer. The disturbances in the oral microbiome may cause dysfunction in the gut epithelial barrier, increased intestinal permeability, increased synthesis of pro-inflammatory cytokines, increased cellular proliferation, as well as changes in β -catenin and Wnt signalling, leading to enhanced carcinogenesis [129,130] (Tables 1 and 2, Figure 5).

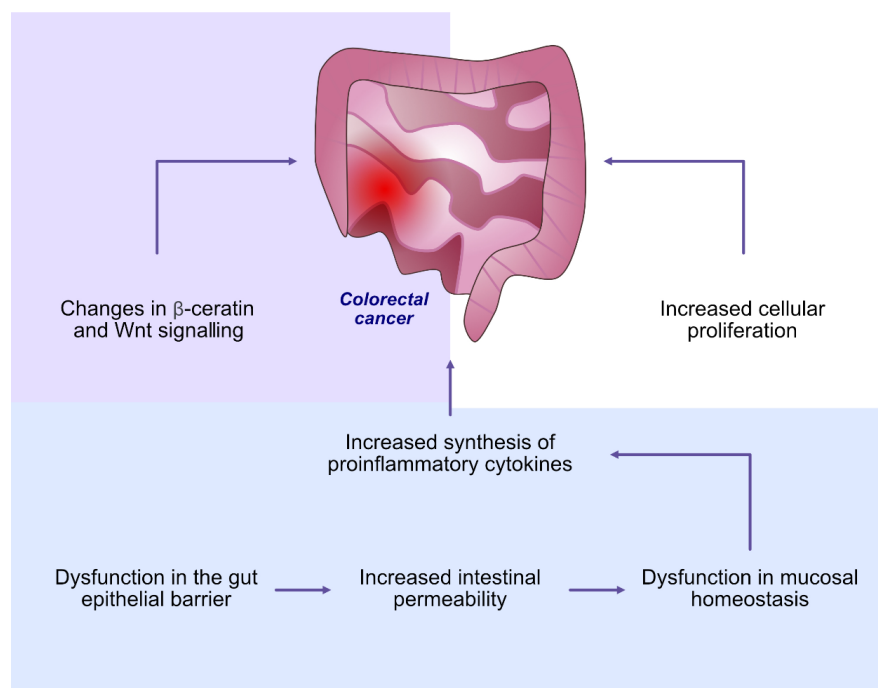


Figure 5. The involvement of oral microbiome dysbiosis in the development of colorectal cancer.

5. Conclusions

Previous studies highlight the role of the oral microbiome in periodontal health and disease. Greater awareness and knowledge of the causes of dysbiosis, environmental risk factors and periodontal therapy can reduce the increasing prevalence of PD worldwide. The promotion of proper oral hygiene, a reduction in smoking, alcohol consumption and exposure to stress and comprehensive treatment to reduce the pathogenicity of the oral biofilm can help reduce the incidence of PD. Recent research on the human microbiome has led to increased interest in the oral microbiome and its impact on the normal course of oral processes and the development of disease states, including many systemic diseases. Evidence linking the relationship between disorders of the oral microbiome and various systemic diseases has increased awareness of the importance of the oral microbiome in regulating numerous processes in the human body and, thus, its impact on the development of many diseases. Full knowledge of the role of the oral microbiome in health and the development of disease processes can contribute to the prevention and treatment of diseases.

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References

- Hernández, M.; Mayer, M.P.A.; Santi-Rocca, J. Editorial: The Human Microbiota in Periodontitis. *Front. Cell. Infect. Microbiol.* **2022**, *12*, 952205. [[CrossRef](#)]
- Greenwood, D.; Afacan, B.; Emingil, G.; Bostanci, N.; Belibasakis, G.N. Salivary Microbiome Shifts in Response to Periodontal Treatment Outcome. *Proteom. Clin. Appl.* **2020**, *14*, e2000011. [[CrossRef](#)]
- Zhang, Y.; Ding, Y.; Guo, Q. Probiotic Species in the Management of Periodontal Diseases: An Overview. *Front. Cell. Infect. Microbiol.* **2022**, *12*, 806463. [[CrossRef](#)]

4. Van Dyke, T.E. Pro-resolving mediators in the regulation of periodontal disease. *Mol. Asp. Med.* **2017**, *58*, 21–36. [[CrossRef](#)] [[PubMed](#)]
5. Herrero, E.R.; Fernandes, S.; Verspecht, T.; Ugarte-Berzal, E.; Boon, N.; Proost, P.; Teughels, W. Dysbiotic biofilms deregulate the periodontal inflammatory response. *J. Dent. Res.* **2018**, *97*, 547–555. [[CrossRef](#)]
6. Kozak, M.; Dabrowska-Zamojcin, E.; Mazurek-Mochol, M.; Pawlik, A. Cytokines and Their Genetic Polymorphisms Related to Periodontal Disease. *J. Clin. Med.* **2020**, *9*, 4045. [[CrossRef](#)]
7. Kirst, M.E.; Li, E.C.; Alfant, B.; Chi, Y.Y.; Walker, C.; Magnusson, I.; Wang, G.P. Dysbiosis and alterations in predicted functions of the subgingival microbiome in chronic periodontitis. *Appl. Environ. Microbiol.* **2015**, *81*, 783–793. [[CrossRef](#)] [[PubMed](#)]
8. Heidari, Z. The association between proinflammatory gene polymorphisms and level of gingival tissue degradation in chronic periodontal disease. *Gene Cell Tissue* **2014**, *1*, e21898. [[CrossRef](#)]
9. Heidari, Z.; Moudi, B.; Mahmoudzadeh-Sagheb, H. Immunomodulatory factors gene polymorphisms in chronic periodontal disease: An overview. *BMC Oral Health* **2019**, *19*, 29. [[CrossRef](#)]
10. Di Stefano, M.; Polizzi, A.; Santonocito, S.; Romano, A.; Lombardi, T.; Isola, G. Impact of Oral Microbiome in Periodontal Health and Periodontitis: A Critical Review on Prevention and Treatment. *Int. J. Mol. Sci.* **2022**, *23*, 5142. [[CrossRef](#)]
11. Cheng, W.C.; van Asten, S.D.; Burns, L.A.; Evans, H.G.; Walter, G.J.; Hashim, A.; Hughesand, F.J.; Taams, L.S. Periodontal disease-associated pathogens *P. gingivalis* and *A. actinomycetemcomitans* activate human CD14+ monocytes leading to enhanced Th17/IL-17 responses. *Eur. J. Immunol.* **2016**, *46*, 2211–2221. [[CrossRef](#)]
12. Johnson, G.K.; Guthmiller, J.M. The impact of cigarette smoking on periodontal disease and treatment. *Periodontol. 2000* **2007**, *44*, 178–194. [[CrossRef](#)]
13. Popa, G.V.; Costache, A.; Badea, O.; Cojocaru, M.O.; Mitroi, G.; Lazăr, A.C.; Olimid, D.-A.; Mogoantă, L. Histopathological and immunohistochemical study of periodontal changes in chronic smokers. *Romanian J. Morphol. Embryol.* **2021**, *62*, 209–217. [[CrossRef](#)] [[PubMed](#)]
14. Sawant, S.; Dugad, J.; Parikh, D.; Srinivasan, S.; Singh, H. Oral Microbial Signatures of Tobacco Chewers and Oral Cancer Patients in India. *Pathogens* **2023**, *12*, 78. [[CrossRef](#)] [[PubMed](#)]
15. Genco, R.J.; Grossi, S.G.; Ho, A.; Nishimura, F.; Murayama, Y. A proposed model linking inflammation to obesity, diabetes, and periodontal infections. *J. Periodontol.* **2005**, *76* (Suppl. S11), 2075–2084. [[CrossRef](#)] [[PubMed](#)]
16. Carrizales-Sepúlveda, E.F.; Ordaz-Farías, A.; Vera-Pineda, R.; Flores-Ramírez, R. Periodontal disease, systemic inflammation and the risk of cardiovascular disease. *Heart Lung Circ.* **2018**, *27*, 1327–1334. [[CrossRef](#)]
17. Kim, J.; Amar, S. Periodontal disease and systemic conditions: A bidirectional relationship. *Odontology* **2006**, *94*, 10–21. [[CrossRef](#)]
18. Nibali, L.; Bayliss-Chapman, J.; Almoftareh, S.A.; Zhou, Y.; Divaris, K.; Vieira, A.R. What is the heritability periodontitis? A systematic review. *J. Dent. Res.* **2019**, *98*, 632–641. [[CrossRef](#)] [[PubMed](#)]
19. López, R.; Smith, P.; Göstemeyer, G.; Schwendicke, F. Ageing, dental caries and periodontal diseases. *J. Clin. Periodontol.* **2017**, *44*, S145–S152. [[CrossRef](#)]
20. Belibasakis, G.N. Microbiological changes of the ageing oral cavity. *Arch. Oral Biol.* **2018**, *96*, 230–232. [[CrossRef](#)]
21. Hajishengallis, G.; Lamont, R.J. Beyond the red complex and into more complexity: The Polymicrobial Synergy and Dysbiosis (PSD) model of periodontal disease etiology. *Mol. Oral Microbiol.* **2012**, *27*, 409–419. [[CrossRef](#)]
22. Najmanova, L.; Sabova, L.; Lenartova, M.; Janatova, T.; Mysak, J.; Vetrovsky, T.; Tesinska, B.; Novotna, G.B.; Koberska, M.; Broukal, Z.; et al. R/G value—A numeric index of periodontal health. *Front. Cell. Infect. Microbiol.* **2021**, *11*, 602643. [[CrossRef](#)]
23. Hajishengallis, G. Periodontitis: From microbial immune subversion to systemic inflammation. *Nat. Rev. Immunol.* **2015**, *15*, 30–44. [[CrossRef](#)]
24. Deo, P.N.; Deshmukh, R. Oral microbiome: Unveiling the fundamentals. *J. Oral. Maxillofac Pathol. JOMFP* **2019**, *23*, 122–128. [[CrossRef](#)] [[PubMed](#)]
25. Mahasneh, S.A.; Mahasneh, A.M. Probiotics: A Promising Role in Dental Health. *Dent J.* **2017**, *5*, 26. [[CrossRef](#)] [[PubMed](#)]
26. Thuy, D.; Devine, D.; Marsh, P. Oral biofilms: Molecular analysis, challenges, and future prospects in dental diagnostics. *Clin. Cosmet. Investig. Dent.* **2013**, *5*, 11–19.
27. Marsh, P.D.; Zaura, E. Dental biofilm: Ecological interactions in health and disease. *J. Clin. Periodontol.* **2017**, *44* (Suppl. S18), S12–S22. [[CrossRef](#)] [[PubMed](#)]
28. Escapa, I.F.; Chen, T.; Huang, Y.; Gajare, P.; Dewhirst, F.E.; Lemon, K.P. New insights into human nostril microbiome from the expanded Human Oral Microbiome Database (eHOMD): A resource for the microbiome of the human aerodigestive tract. *MSystems* **2018**, *3*, e00187–18. [[CrossRef](#)]
29. Bartnicka, D.; Gonzalez-Gonzalez, M.; Sykut, J.; Koziel, J.; Ciaston, I.; Adamowicz, K.; Bras, G.; Zawrotniak, M.; Karkowska-Kuleta, J.; Satala, D.; et al. *Candida albicans* Shields the Periodontal Killer *Porphyromonas gingivalis* from Recognition by the Host Immune System and Supports the Bacterial Infection of Gingival Tissue. *Int. J. Mol. Sci.* **2020**, *21*, 1984. [[CrossRef](#)]
30. Barros, P.P.; Ribeiro, F.C.; Rossoni, R.D. Influence of *Candida krusei* and *Candida glabrata* on *Candida albicans* gene expression In Vitro biofilms. *Arch. Oral Biol.* **2016**, *64*, 92–101. [[CrossRef](#)]
31. Jorgensen, M.R.; Kragelund, C.; Jansen, P.; Keller, M.K.; Twetman, S. Probiotic *Lactobacillus reuteri* has antifungal effects on oral *Candida* species In Vitro. *Arch. Oral Biol.* **2017**, *9*, 127–135. [[CrossRef](#)]
32. Nobbis, A.H.; Jenkinson, H.F. Interkingdom networking within the oral micro-biome. *Microbes and Infection.* **2015**, *17*, 484–492. [[CrossRef](#)]

33. Ng, H.M.; Kin, L.X.; Dashper, S.G.; Slakeski, N.; Butler, C.A.; Reynolds, E.C. Bacterial interactions in pathogenic subgingival plaque. *Microb. Pathog.* **2016**, *94*, 60–69. [[CrossRef](#)]
34. Belstrøm, D.; Constancias, F.; Markvart, M.; Sikora, M.; Sorensen, C.E.; Givskov, M. Transcriptional Activity of Predominant Streptococcus Species at Multiple Oral Sites Associate With Periodontal Status. *Front. Cell. Infect. Microbiol.* **2021**, *11*, 752664. [[CrossRef](#)] [[PubMed](#)]
35. López-López, A.; Camelo-Castillo, A.; Ferrer, M.D.; Simon-Soro, A.; Mira, A. Health-Associated Niche Inhabitants as Oral Probiotics: The Case of *Streptococcus dentisani*. *Front. Microbiol.* **2017**, *8*, 379. [[CrossRef](#)]
36. Esteban-Fernandez, A.; Ferrer, M.D.; Zorraquin-Pena, I.; Lopez-Lopez, A.; Moreno-Arribas, M.V.; Mira, A. In Vitro Beneficial Effects of *Streptococcus dentisani* as Potential Oral Probiotic for Periodontal Diseases. *J. Periodontol.* **2019**, *90*, 1346–1355. [[CrossRef](#)] [[PubMed](#)]
37. Di Pierro, F.; Zanvit, A.; Nobili, P.; Risso, P.; Fornaini, C. Cariogram Outcome After 90 Days of Oral Treatment With *Streptococcus salivarius* M18 in Children at High Risk for Dental Caries: Results of a Randomized, Controlled Study. *Clin. Cosmet. Investig. Dent.* **2015**, *7*, 107–113. [[CrossRef](#)] [[PubMed](#)]
38. Hill, C.; Guarner, F.; Reid, G.; Gibson, G.R.; Merenstein, D.J.; Pot, B.; Morelli, L.; Canani, R.B.; Flint, H.J.; Salminen, S.; et al. Expert Consensus Document. The International Scientific Association for Probiotics and Prebiotics Consensus Statement on the Scope and Appropriate Use of the Term Probiotic. *Nat. Rev. Gastroenterol. Hepatol.* **2014**, *11*, 506–514. [[CrossRef](#)] [[PubMed](#)]
39. Ince, G.; Gursoy, H.; Ipci, S.D.; Cakar, G.; Emekli-Alturfan, E.; Yilmaz, S. Clinical and Biochemical Evaluation of Lozenges Containing *Lactobacillus reuteri* as an Adjunct to Non-Surgical Periodontal Therapy in Chronic Periodontitis. *J. Periodontol.* **2015**, *86*, 746–754. [[CrossRef](#)]
40. Sivamaruthi, B.S.; Kesika, P.; Chaiyasut, C. A Review of the Role of Probiotic Supplementation in Dental Caries. *Probiotics Antimicrob. Proteins* **2020**, *12*, 1300–1309. [[CrossRef](#)]
41. Ohshima, T.; Kojima, Y.; Seneviratne, C.J.; Maeda, N. Therapeutic Application of Synbiotics, a Fusion of Probiotics and Prebiotics, and Biogenics as a New Concept for Oral Candida Infections: A Mini Review. *Front. Microbiol.* **2016**, *7*, 10. [[CrossRef](#)] [[PubMed](#)]
42. Yoo, J.I.; Shin, I.S.; Jeon, J.G.; Yang, Y.M.; Kim, J.G.; Lee, D.W. The Effect of Probiotics on Halitosis: A Systematic Review and Meta-Analysis. *Probiotics Antimicrob. Proteins* **2019**, *11*, 150–157. [[CrossRef](#)] [[PubMed](#)]
43. Allaker, R.P.; Stephen, A.S. Use of Probiotics and Oral Health. *Curr. Oral. Health Rep.* **2017**, *4*, 309–318. [[CrossRef](#)]
44. Kawai, T.; Ohshima, T.; Tanaka, T.; Ikawa, S.; Tani, A.; Inazumi, N.; Shin, R.; Itoh, Y.; Meyer, K.; Maeda, N. *Limosi lactobacillus* (*Lactobacillus fermentum* ALAL020), a Probiotic Candidate Bacterium, Produces a Cyclic Dipeptide That Suppresses the Periodontal Pathogens *Porphyromonas gingivalis* and *Prevotella intermedia*. *Front Cell. Infect Microbiol.* **2022**, *12*, 804334. [[CrossRef](#)] [[PubMed](#)]
45. Swanson, K.S.; Gibson, G.R.; Hutkins, R.; Reimer, R.A.; Reid, G.; Verbeke, K.; Scott, K.P.; Holscher, H.D.; Azad, M.B.; Delzenne, N.M.; et al. The International Scientific Association for Probiotics and Prebiotics (ISAPP) consensus statement on the definition and scope of synbiotics. *Nat. Rev. Gastroenterol. Hepatol.* **2020**, *17*, 687–701. [[CrossRef](#)] [[PubMed](#)]
46. Duraisamy, V.; Geethapriya, P.R.; Bharath, C.; Niveditha, R.S.; John, J.B. Role of probiotics and synbiotics on inhibiting *Streptococcus mutans* level in saliva of children: A randomized controlled trial. *J. Indian Soc. Pedod. Prev. Dent.* **2021**, *39*, 275–278. [[CrossRef](#)]
47. Willis, J.R.; Gabaldón, T. The Human Oral Microbiome in Health and Disease: From Sequences to Ecosystems. *Microorganisms* **2020**, *8*, 308. [[CrossRef](#)]
48. Wilson, M. *Bacteriology of Humans: An Ecological Perspective*; Blackwell Publishing Ltd.: Hoboken, NJ, USA, 2008.
49. Liu, B.; Faller, L.L.; Klitgord, N.; Mazumdar, V.; Ghodsi, M.; Sommer, D.D.; Gibbons, T.R.; Treangen, T.J.; Chang, Y.-C.; Li, S.; et al. Deep sequencing of the oral microbiome reveals signatures of periodontal disease. *PLoS ONE* **2012**, *7*, e37919. [[CrossRef](#)]
50. Stamatova, I.; Meurman, J.H. Probiotics: Health benefits in the mouth. *Am. J. Dent.* **2009**, *22*, 329–338.
51. De Geest, S.; Laleman, I.; Teughels, W.; Dekeyser, C.; Quirynen, M. Periodontal diseases as a source of halitosis: A review of the evidence and treatment approaches for dentists and dental hygienists. *Periodontol. 2000* **2016**, *71*, 213–227. [[CrossRef](#)]
52. Sterer, N.; Shaharabany, M.; Rosenberg, M. Beta-Galactosidase activity and H₂S production in an experimental oral biofilm. *J. Breath Res.* **2009**, *3*, 016006. [[CrossRef](#)] [[PubMed](#)]
53. Bornstein, M.M.; Kislig, K.; Hoti, B.B.; Seemann, R.; Lussi, A. Prevalence of halitosis in the population of the city of Bern, Switzerland: A study comparing self-reported and clinical data. *Eur. J. Oral Sci.* **2009**, *117*, 261–267. [[CrossRef](#)] [[PubMed](#)]
54. De Lima, P.O.; Nani, B.D.; Rolim, G.S.; Groppo, F.C.; Franz-Montan, M.; Alves De Moraes, A.B.; Cogo-Müller, K.; Marcondes, F.K. Effects of academic stress on the levels of oral volatile sulfur compounds, halitosis-related bacteria and stress biomarkers of healthy female undergraduate students. *J. Breath Res.* **2020**, *14*, 036005. [[CrossRef](#)] [[PubMed](#)]
55. Wu, J.; Cannon, R.D.; Ji, P.; Farella, M.; Mei, L. Halitosis: Prevalence, risk factors, sources, measurement and treatment—A review of the literature. *Aust. Dent. J.* **2020**, *65*, 4–11. [[CrossRef](#)]
56. Hampelska, K.; Jaworska, M.M.; Babalska, Z.Ł.; Karpiński, T.M. The Role of Oral Microbiota in Intra-Oral Halitosis. *J. Clin. Med.* **2020**, *9*, 2484. [[CrossRef](#)]
57. Nalcaci, R.; Sonmez, I.S. Evaluation of oral malodor in children. *Oral. Surg. Oral. Med. Oral. Pathol. Oral. Radiol. Endod.* **2008**, *106*, 384–388. [[CrossRef](#)]
58. Wu, Y.; Chi, X.; Zhang, Q.; Chen, F.; Deng, X. Characterization of the salivary microbiome in people with obesity. *PeerJ* **2018**, *6*, e4458. [[CrossRef](#)]
59. Yitzhaki, S.; Reshef, L.; Gophna, U.; Rosenberg, M.; Sterer, N. Microbiome associated with denture malodour. *J. Breath Res.* **2018**, *12*, 027103. [[CrossRef](#)]

60. Zhang, Y.; Zhu, C.; Cao, G.; Zhan, J.; Feng, X.; Chen, X. Dynamic Alterations of Oral Microbiota Related to Halitosis in Preschool Children. *Front. Cell. Infect. Microbiol.* **2021**, *11*, 599467. [[CrossRef](#)]
61. Saito, A.; Kokubu, E.; Inagaki, S.; Imamura, K.; Kita, D.; Lamont, R.J.; Ishihara, K. Porphyromonas Gingivalis Entry Into Gingival Epithelial Cells Modulated by *Fusobacterium nucleatum* is Dependent on Lipid Rafts. *Microb. Pathog.* **2012**, *53*, 234–242. [[CrossRef](#)]
62. Chen, Y.; Huang, Z.; Tang, Z.; Huang, Y.; Huang, M.; Liu, H.; Ziebolz, D.; Schmalz, G.; Jia, B.; Zhao, J. More Than Just a Periodontal Pathogen -the Research Progress on *Fusobacterium nucleatum*. *Front. Cell. Infect. Microbiol.* **2022**, *12*, 815318. [[CrossRef](#)]
63. Kang, M.S.; Lim, H.S.; Kim, S.M.; Lee, H.C.; Oh, J.S. Effect of Weissella cibaria on *Fusobacterium nucleatum* induced Interleukin-6 and Interleukin-8 Production in KB Cells. *J. Bacteriol. Virol.* **2011**, *41*, 9–18. [[CrossRef](#)]
64. Suzuki, N.; Yoneda, M.; Tanabe, K.; Fujimoto, A.; Iha, K.; Seno, K.; Yamada, K.; Iwamoto, T.; Masuo, Y.; Hirofujii, T. *Lactobacillus salivarius* WB21-containing Tablets for the Treatment of Oral Malodor: A Double-Blind, Randomized, Placebo-Controlled Crossover Trial. *Oral. Surg. Oral. Med. Oral. Pathol. Oral. Radiol.* **2014**, *117*, 462–470. [[CrossRef](#)]
65. Lenartova, M.; Tesinska, B.; Janatova, T.; Hrebicek, O.; Mysak, J.; Janata, J.; Najmanova, L. The Oral Microbiome in Periodontal Health. *Front. Cell. Infect. Microbiol.* **2021**, *11*, 629723. [[CrossRef](#)]
66. Pignatelli, P.; Fabietti, G.; Ricci, A.; Piattelli, A.; Curia, M.C. How Periodontal Disease and Presence of Nitric Oxide Reducing Oral Bacteria Can Affect Blood Pressure. *Int. J. Mol. Sci.* **2020**, *21*, 7538. [[CrossRef](#)]
67. Reher, V.G.; Zenóbio, E.G.; Costa, F.O.; Reher, P.; Soares, R.V. Nitric oxide levels in saliva increase with severity of chronic periodontitis. *J. Oral Sci.* **2007**, *49*, 271–276. [[CrossRef](#)]
68. Sampaio-Maia, B.; Caldas, I.M.; Pereira, M.L.; Perez-Mongiovi, D.; Araujo, R. The oral microbiome in health and its implication in oral and systemic diseases. *Adv. Appl. Microbiol.* **2016**, *97*, 171–210. [[PubMed](#)]
69. Melo-Dias, S.; Cabral, M.; Furtado, A.; Souto-Miranda, S.; Mendes, M.A.; Cravo, J.; Almeida, C.R.; Marques, A.; Sousa, A. Responsiveness to pulmonary rehabilitation in COPD is associated with changes in microbiota. *Respir Res.* **2023**, *24*, 29. [[CrossRef](#)] [[PubMed](#)]
70. Millares, L.; Monso, E. The Microbiome in COPD: Emerging Potential for Microbiome-Targeted Interventions. *Int. J. Chron. Obstruct. Pulmon. Dis.* **2022**, *17*, 1835–1845. [[CrossRef](#)]
71. Tiew, P.Y.; Jaggi, T.K.; Chan, L.L.Y.; Chotirmall, S.H. The airway microbiome in COPD, bronchiectasis and bronchiectasis-COPD overlap. *Clin. Respir. J.* **2021**, *15*, 123–133. [[CrossRef](#)] [[PubMed](#)]
72. Mammen, M.J.; Sethi, S. COPD and the microbiome. *Respirology* **2016**, *21*, 590–599. [[CrossRef](#)]
73. Whiley, R.A.; Fleming, E.V.; Makhija, R.; Waite, R.D. Environment and colonisation sequence are key parameters driving cooperation and competition between *Pseudomonas aeruginosa* cystic fibrosis strains and oral commensal streptococci. *PLoS ONE* **2015**, *10*, e0115513. [[CrossRef](#)] [[PubMed](#)]
74. Zhu, L.; Kreth, J. The role of hydrogen peroxide in environmental adaptation of oral microbial communities. *Oxidative Med. Cell. Longev.* **2012**, *2012*, 717843. [[CrossRef](#)] [[PubMed](#)]
75. Haran, J.P.; Bradley, E.; Zeamer, A.L.; Cincotta, L.; Salive, M.C.; Dutta, P.; Mutaawe, S.; Anya, O.; Meza-Segura, M.; Moormann, A.M.; et al. Inflammation-type dysbiosis of the oral microbiome as sociates with the duration of COVID-19 symptoms and long COVID. *JCI Insight* **2021**, *6*, e152346. [[CrossRef](#)] [[PubMed](#)]
76. Khan, A.A.; Khan, Z. COVID-2019-associated overexpressed Prevotella proteins mediated host-pathogen interactions and their role in coronavirus outbreak. *Bioinformatics* **2020**, *36*, 4065–4069. [[CrossRef](#)]
77. Van den Bogert, B.; Meijerink, M.; Zoetendal, E.G.; Wells, J.M.; Kleerebezem, M. Immunomodulatory properties of Streptococcus and Veillonella isolates from the human small intestine microbiota. *PLoS ONE* **2014**, *9*, e114277. [[CrossRef](#)]
78. Segal, L.N.; Clemente, J.C.; Tsay, J.-C.J.; Koralov, S.B.; Keller, B.C.; Wu, B.G.; Alison, M.; Shen, N.; Ghedin, E.; Morris, A.; et al. Enrichment of the lung microbiome with oral taxa is associated with lung inflammation of a Th17 phenotype. *Nat. Microbiol.* **2016**, *1*, 16031. [[CrossRef](#)]
79. Quigley, E.M.M. Microbiota-brain-gut axis and neurodegenerative diseases. *Curr. Neurol. Neurosci. Rep.* **2017**, *17*, 94. [[CrossRef](#)]
80. Nagarajan, A.; Srivastava, H.; Morrow, C.D.; Sun, L.Y. Characterizing the gut microbiome changes with aging in a Novel Alzheimer's disease rat model. *Aging* **2023**, *15*, 59–471. [[CrossRef](#)]
81. Jin, J.; Xu, Z.; Zhang, L.; Zhang, C.; Zhao, X.; Mao, Y.; Zhang, H.; Liang, X.; Wu, J.; Yang, Y.; et al. Gut-derived β -amyloid: Likely a centerpiece of the gut-brain axis contributing to Alzheimer's pathogenesis. *Gut Microbes* **2023**, *15*, 2167172. [[CrossRef](#)]
82. Barrett, E.; Ross, R.P.; O'Toole, P.W.; Fitzgerald, G.F.; Stanton, C. γ -Aminobutyric acid production by culturable bacteria from the human intestine. *J. Appl. Microbiol.* **2012**, *113*, 411–417. [[CrossRef](#)]
83. Lyte, M. Probiotics function mechanistically as delivery vehicles for neuroactive compounds: Microbial endocrinology in the design and use of probiotics. *Bioessays* **2011**, *33*, 574–581. [[CrossRef](#)]
84. Ruddick, J.P.; Evans, A.K.; Nutt, D.J.; Lightman, S.; Rook, G.; Lowry, C.A. Tryptophan metabolism in the central nervous system: Medical implications. *Expert Rev. Mol. Med.* **2006**, *31*, 1–27. [[CrossRef](#)] [[PubMed](#)]
85. Widner, B.; Leblhuber, F.; Walli, J.; Titz, G.P.; Demel, U.; Fuchs, D. Tryptophan degradation and immune activation in Alzheimer's disease. *J. Neural. Transm.* **1996**, *107*, 343–353. [[CrossRef](#)] [[PubMed](#)]
86. Gulaj, E.; Pawlak, K.; Bien, B.; Pawlak, D. Kynurenine and its metabolites in Alzheimer's disease patients. *Adv. Med. Sci.* **2010**, *55*, 204–211. [[CrossRef](#)]
87. Du, X.; Wang, X.; Geng, M. Alzheimer's disease hypothesis and related therapies. *Transl. Neurodegener.* **2018**, *7*, 2. [[CrossRef](#)] [[PubMed](#)]
88. Bienenstock, J.; Kunze, W.; Forsythe, P. Microbiota and the gut-brain axis. *Nutr. Rev.* **2015**, *73* (Suppl. S1), 28–31. [[CrossRef](#)]

89. Frost, G.; Sleeth, M.L.; Sahuri-Arisoylu, M.; Lizarbe, B.; Cerdan, S.; Brody, L.; Anastasovska, J.; Ghourab, S.; Hankir, M.; Zhang, S.; et al. The short-chain fatty acid acetate reduces appetite via a central homeostatic mechanism. *Nat. Commun.* **2014**, *5*, 3611. [[CrossRef](#)] [[PubMed](#)]
90. Zhao, Y.; Lukiw, W.J. Microbiome-generated amyloid and potential impact on amyloidogenesis in Alzheimer's disease (AD). *Nat. Sci.* **2015**, *1*, e138.
91. Hill, J.M.; Lukiw, W.J. Microbial-generated amyloids and Alzheimer's disease (AD). *Front. Aging Neurosci.* **2015**, *7*, 9. [[CrossRef](#)]
92. Sanchez-Ramos, J.; Song, S.; Sava, V.; Catlow, B.; Lin, X.; Mori, T.; Cao, C.; Arendash, G.W. Granulocyte colony stimulating factor decreases brain amyloid burden and reverses cognitive impairment in Alzheimer's mice. *Neuroscience* **2009**, *163*, 55–72. [[CrossRef](#)] [[PubMed](#)]
93. Hassan, W.M.; Dostal, V.; Huemann, B.N.; Yerg, J.E.; Link, C.D. Identifying A β -specific pathogenic mechanisms using a nematode model of Alzheimer's disease. *Neurobiol. Aging* **2015**, *36*, 857–866. [[CrossRef](#)] [[PubMed](#)]
94. Arimatsu, K.; Yamada, H.; Miyazawa, H.; Minagawa, T.; Nakajima, M.; Ryder, M.I.; Gotoh, K.; Motooka, D.; Nakamura, S.; Iida, T.; et al. Oral pathobiont induces systemic inflammation and metabolic changes associated with alteration of gut microbiota. *Sci. Rep.* **2014**, *4*, 4828. [[CrossRef](#)] [[PubMed](#)]
95. Dominy, S.S.; Lynch, C.; Ermini, F.; Benedyk, M.; Marczyk, A.; Konradi, A.; Nguyen, M.; Haditsch, U.; Raha, D.; Griffin, C.; et al. *Porphyromonas gingivalis* in Alzheimer's disease brains: Evidence for disease causation and treatment with small-molecule inhibitors. *Sci. Adv.* **2019**, *5*, eaau3333. [[CrossRef](#)] [[PubMed](#)]
96. Miklossy, J. Bacterial amyloid and DNA are important constituents of senile plaques: Further evidence of the spirochetal and biofilm nature of senile plaques. *J. Alzheimers Dis.* **2016**, *53*, 1459–1473. [[CrossRef](#)]
97. Meregildo-Rodríguez, E.D.; Robles-Arce, L.G.; Chunga-Chávez, E.V.; Asmat-Rubio, M.G.; Zavaleta-Alaya, P.; Vásquez-Tirado, G.A. Periodontal disease as a non-traditional risk factor for acute coronary syndrome: A systematic review and meta-analysis. *Infez. Med.* **2022**, *30*, 501–515. [[CrossRef](#)]
98. Larvin, H.; Kang, J.; Aggarwal, V.R.; Pavitt, S.; Wu, J. Risk of incident cardiovascular disease in people with periodontal disease: A systematic review and meta-analysis. *Clin. Exp. Dent. Res.* **2021**, *7*, 109–122. [[CrossRef](#)]
99. Teles, R.; Wang, C.Y. Mechanisms involved in the association between periodontal diseases and cardiovascular disease. *Oral Dis.* **2011**, *17*, 450–461. [[CrossRef](#)]
100. Wang, Z.; Klipfell, E.; Bennett, B.J.; Koeth, R.; Levison, B.S.; DuGar, B.; Feldstein, A.E.; Britt, E.B.; Fu, X.; Chung, Y.-M.; et al. Gut flora metabolism of phosphatidylcholine promotes cardiovascular disease. *Nature* **2011**, *472*, 57–63. [[CrossRef](#)]
101. Ma, G.; Pan, B.; Chen, Y.; Guo, C.; Zhao, M.; Zheng, L.; Chen, B. Trimethylamine N-oxide in atherogenesis: Impairing endothelial self-repair capacity and enhancing monocyte adhesion. *Biosci. Rep.* **2017**, *37*, BSR20160244. [[CrossRef](#)]
102. Makrecka-Kuka, M.; Volska, K.; Antone, U.; Vilskersts, R.; Grinberga, S.; Bandere, D.; Liepinsh, E.; Dambrova, M. Trimethylamine N-oxide impairs pyruvate and fatty acid oxidation in cardiac mitochondria. *Toxicol. Lett.* **2017**, *267*, 32–38. [[CrossRef](#)]
103. Ohira, H.; Tsutsui, W.; Fujioka, Y. Are Short Chain Fatty Acids in Gut Microbiota Defensive Players for Inflammation and Atherosclerosis? *J. Atheroscler. Thromb.* **2017**, *24*, 660–672. [[CrossRef](#)]
104. Lan, T.H.; Huang, X.Q.; Tan, H.M. Vascular fibrosis in atherosclerosis. *Cardiovasc. Pathol.* **2013**, *22*, 401–407. [[CrossRef](#)]
105. Chacón, M.R.; Lozano-Bartolomé, J.; Portero-Otín, M.; Rodríguez, M.; Xifra, G.; Puig, J.; Blasco, G.; Ricart, W.; Chaves, F.; Fernández-Real, J. The gut mycobiome composition is linked to carotid atherosclerosis. *Benef. Microbes* **2018**, *9*, 185–198. [[CrossRef](#)]
106. Chambers, E.S.; Preston, T.; Frost, G.; Morrison, D.J. Role of Gut Microbiota-Generated Short-Chain Fatty Acids in Metabolic and Cardiovascular Health. *Curr. Nutr. Rep.* **2018**, *7*, 198–206. [[CrossRef](#)]
107. Hernández, M.A.G.; Canfora, E.E.; Jocken, J.W.E.; Blaak, E.E. The Short-Chain Fatty Acid Acetate in Body Weight Control and Insulin Sensitivity. *Nutrients* **2019**, *11*, 1943. [[CrossRef](#)]
108. Guo, X.; Okpara, E.S.; Hu, W.; Yan, C.; Wang, Y.; Liang, Q.; Chiang, J.Y.L.; Han, S. Interactive Relationships between Intestinal Flora and Bile Acids. *Int. J. Mol. Sci.* **2022**, *23*, 8343. [[CrossRef](#)]
109. Tanase, D.M.; Gosav, E.M.; Neculae, E.; Costea, C.F.; Ciocoiu, M.; Hurjui, L.L.; Tarniceriu, C.C.; Maranduca, M.A.; Lacatusu, C.M.; Floria, M.; et al. Role of Gut Microbiota on Onset and Progression of Microvascular Complications of Type 2 Diabetes (T2DM). *Nutrients* **2020**, *12*, 3719. [[CrossRef](#)] [[PubMed](#)]
110. Xiao, E.; Mattos, M.; Vieira, G.H.A.; Chen, S.; Corrêa, J.D.; Wu, Y.; Albiero, M.L.; Bittinger, K.; Graves, D.T. Diabetes enhances IL-17 expression and alters the oral microbiome to increase its pathogenicity. *Cell Host Microbe* **2017**, *22*, 120–128. [[CrossRef](#)] [[PubMed](#)]
111. Dissick, A.; Redman, R.S.; Jones, M.; Rangan, B.V.; Reimold, A.; Griffiths, G.R.; Mikuls, T.R.; Amdur, R.L.; Richards, J.S.; Kerr, G.S. Association of periodontitis with rheumatoid arthritis: A pilot study. *J. Periodontol.* **2010**, *81*, 223–230. [[CrossRef](#)] [[PubMed](#)]
112. Scher, J.U.; Ubeda, C.; Equinda, M.; Khanin, R.; Buischi, Y.; Viale, A.; Lipuma, L.; Attur, M.; Pillinger, M.H.; Weissmann, G.; et al. Periodontal disease and the oral microbiota in new-onset rheumatoid arthritis. *Arthritis Rheum.* **2012**, *64*, 3083–3094. [[CrossRef](#)] [[PubMed](#)]
113. Nik-Azis, N.M.; Mohd, N.; Baharin, B.; Said, M.S.M.; Fadzilah, F.M.; Haflah, N.H.M. Periodontal disease in sero-positive rheumatoid arthritis: Scoping review of the epidemiological evidence. *Germs* **2021**, *11*, 266–286. [[CrossRef](#)]
114. De Smit, M.; Westra, J.; Vissink, A.; Doornbos-van der Meer, B.; Brouwer, E.; van Winkelhoff, A.J. Periodontitis in established rheumatoid arthritis patients: A cross-sectional clinical, microbiological and serological study. *Arthritis Res. Ther.* **2012**, *14*, R222. [[CrossRef](#)]

115. Ortiz, P.; Bissada, N.F.; Palomo, L.; Han, Y.W.; Al-Zahrani, M.S.; Panneerselvam, A.; Askari, A. Periodontal therapy reduces the severity of active rheumatoid arthritis in patients treated with or without tumor necrosis factor inhibitors. *J. Periodontol.* **2009**, *80*, 535–540. [[CrossRef](#)]
116. Al-Katma, M.K.; Bissada, N.F.; Bordeaux, J.M.; Sue, J.; Askari, A.D. Control of periodontal infection reduces the severity of active rheumatoid arthritis. *J. Clin. Rheumatol.* **2007**, *13*, 134–137. [[CrossRef](#)] [[PubMed](#)]
117. Arvikar, S.L.; Collier, D.S.; Fisher, M.C.; Unizony, S.; Cohen, G.L.; McHugh, G.; Kawai, T.; Strle, K.; Steere, A.C. Clinical correlations with *Porphyromonas gingivalis* antibody responses in patients with early rheumatoid arthritis. *Arthritis Res. Ther.* **2013**, *15*, R109. [[CrossRef](#)] [[PubMed](#)]
118. Wegner, N.; Wait, R.; Sroka, A.; Eick, S.; Nguyen, K.A.; Lundberg, K.; Kinloch, A.; Culshaw, S.; Potempa, J.; Venables, P.J. Peptidylarginine deiminase from *Porphyromonas gingivalis* citrullinates human fibrinogen and alpha-enolase: Implications for autoimmunity in rheumatoid arthritis. *Arthritis Rheum.* **2010**, *62*, 2662–2672. [[CrossRef](#)] [[PubMed](#)]
119. Mikuls, T.R.; Payne, J.B.; Yu, F.; Thiele, G.M.; Reynolds, R.J.; Cannon, G.W.; Markt, J.; McGowan, D.; Kerr, G.S.; Redman, R.S.; et al. Periodontitis and *Porphyromonas gingivalis* in patients with rheumatoid arthritis. *Arthritis Rheumatol.* **2014**, *66*, 1090–1100. [[CrossRef](#)]
120. Quirke, A.M.; Quirke, A.M.; Lugli, E.B.; Wegner, N.; Hamilton, B.C.; Charles, P.; Chowdhury, M.; Ytterberg, A.J.; Zubarev, R.A.; Potempa, J.; et al. Heightened immune response to autocitrullinated *Porphyromonas gingivalis* peptidylarginine deiminase: A potential mechanism for breaching immunologic tolerance in rheumatoid arthritis. *Ann. Rheum. Dis.* **2014**, *73*, 263–269. [[CrossRef](#)]
121. De Aquino, S.G.; Abdollahi-Roodsaz, S.; Koenders, M.; Van De Loo, F.A.J.; Pruijn, G.J.M.; Marijnissen, R.J.; Walgreen, B.; Helsen, M.M.; van den Berselaar, L.A.; de Molon, R.S.; et al. Periodontal pathogens directly promote autoimmune experimental arthritis by inducing a TLR2- and IL-1-driven Th17 response. *J. Immunol.* **2014**, *192*, 4103–4111. [[CrossRef](#)]
122. Kulkarni, A.; Beckler, M.D.; Amini, S.S.; Kesselman, M.M. Oral Microbiome in Pre-Rheumatoid Arthritis: The Role of *Aggregatibacter actinomycetemcomitans* in Bacterial Composition. *Cureus* **2022**, *14*, e32201. [[CrossRef](#)] [[PubMed](#)]
123. Looh, S.C.; Soo, Z.M.P.; Wong, J.J.; Yam, H.C.; Chow, S.K.; Hwang, J.S. *Aggregatibacter actinomycetemcomitans* as the Aetiological Cause of Rheumatoid Arthritis: What Are the Unsolved Puzzles? *Toxins* **2022**, *14*, 50. [[CrossRef](#)] [[PubMed](#)]
124. Konig, M.F.; Abusleme, L.; Reinholdt, J.; Palmer, R.J.; Teles, R.P.; Sampson, K.; Rosen, A.; Nigrovic, P.A.; Sokolove, J.; Giles, J.T.; et al. *Aggregatibacter actinomycetemcomitans*-induced hypercitrullination links periodontal infection to autoimmunity in rheumatoid arthritis. *Sci. Transl. Med.* **2016**, *8*, 369ra176. [[CrossRef](#)] [[PubMed](#)]
125. Gómez-Bañuelos, E.; Mukherjee, A.; Darrah, E.; Andrade, F. Rheumatoid Arthritis-Associated Mechanisms of *Porphyromonas gingivalis* and *Aggregatibacter actinomycetemcomitans*. *J. Clin. Med.* **2019**, *8*, 1309. [[CrossRef](#)] [[PubMed](#)]
126. He, J.; Li, Y.; Cao, Y.; Xue, J.; Zhou, X. The oral microbiome diversity and its relation to human diseases. *Folia Microbiol.* **2015**, *60*, 69–80. [[CrossRef](#)]
127. Sammallahti, H.; Kokkola, A.; Rezasoltani, S.; Ghanbari, R.; Asadzadeh Aghdaei, H.; Knuutila, S.; Puolakkainen, P.; Sarhadi, V.K. Microbiota Alterations and Their Association with Oncogenomic Changes in Pancreatic Cancer Patients. *Int. J. Mol. Sci.* **2021**, *22*, 12978. [[CrossRef](#)]
128. Farrell, J.J.; Zhang, L.; Zhou, H.; Chia, D.; Elashoff, D.; Akin, D.; Paster, B.J.; Joshipura, K.; Wong, D.T. Variations of oral microbiota are associated with pancreatic diseases including pancreatic cancer. *Gut* **2012**, *61*, 582–588. [[CrossRef](#)]
129. Feng, Q.; Liang, S.; Jia, H.; Stadlmayr, A.; Tang, L.; Lan, Z.; Zhang, D.; Xia, H.; Xu, X.; Jie, Z.; et al. Gut microbiome development along the colorectal adenoma-carcinoma sequence. *Nat. Commun.* **2015**, *6*, 6528. [[CrossRef](#)]
130. Saus, E.; Iraola-Guzmán, S.; Willis, R.; Brunet-Vega, A.; Gabaldón, T. Microbiome and colorectal cancer. Roles in carcinogenesis and clinical potential. *Mol. Aspects Med.* **2019**, *69*, 93–106. [[CrossRef](#)]

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