

## **Supplementary Figure Legends**

S1: Cytotoxicity testing of each of the single compounds from the 10 hit well. Compounds were tested over a period of 72 h at a final concentration of 0.1  $\mu$ M and 0.5  $\mu$ M. Cells were treated with 0.1 % DMSO as a vehicle control. Cell toxicity was assessed at 72 h using CellTox Green Cytotoxicity Assay which was added to the cell suspension and cells were incubated for 20 minutes at room temperature before reading. Relative fluorescence unit (RFU) (Ex: 485 nm, Em: 520 nm) was measured using a Synergy HTX Multi- mode Microplate reader. Data is representative of N=3  $\pm$ SEM. \* =  $p < 0.05$ . \*\* =  $p < 0.01$ , \*\*\*\* =  $p < 0.0001$ .

S2: Dose response curves to determine IC<sub>50</sub> values to optimize the ideal dosing for the double and triple combinations. Cell viability was determined using CellTiter® Glo following 72 h. Luminescence was measured using a Synergy HTX Multi- mode Microplate reader. Data is representative of N=3  $\pm$ SEM.

S3: Corresponding densometric quantification of PARP, cleaved-PARP, procaspase-9, cleaved caspase-9, procaspase-3, and cleaved caspase-3 in Kasumi-1 and MV4-11 cell lines following 72 h treatment following 72 h treatment as single agents and as a combination. GAPDH was used as a loading control. Data is representative of representative of N=3  $\pm$ SEM.