

Review **Coronary Artery Spasm-Related Heart Failure Syndrome: Literature Review**

Ming-Jui Hung 1,†, Chi-Tai Yeh 2,3 [,](https://orcid.org/0000-0001-5189-9755) Nicholas G. Kounis ⁴ [,](https://orcid.org/0000-0002-9751-6710) Ioanna Koniari ⁵ , Patrick Hu 6,7 and Ming-Yow Hung 8,9,10,* ,[†](https://orcid.org/0000-0002-6912-7523)

- ¹ Division of Cardiology, Department of Internal Medicine, Chang Gung Memorial Hospital Keelung, Chang Gung University College of Medicine, Keelung City 24201, Taiwan; hmj1447@cgmh.org.tw
- ² Department of Medical Research and Education, Shuang Ho Hospital, Taipei Medical University, New Taipei City 23561, Taiwan; ctyeh@s.tmu.edu.tw
- ³ Continuing Education Program of Food Biotechnology Applications, College of Science and Engineering, National Taitung University, Taitung 95092, Taiwan
- 4 Department of Cardiology, University of Patras Medical School, 26221 Patras, Greece; ngkounis@otenet.gr
5 Cardiology Department Liverpool Heart and Cheet Hospital Liverpool L14 3PE LIK:
- ⁵ Cardiology Department, Liverpool Heart and Chest Hospital, Liverpool L14 3PE, UK; ioanna.koniari@lhch.nhs.uk
- ⁶ Department of Internal Medicine, School of Medicine, University of California, Riverside, Riverside, CA 92521, USA; patrick.hu@ucr.edu
- 7 Department of Cardiology, Riverside Medical Clinic, Riverside, CA 92506, USA
8 Division of Cardiology, Department of Internal Medicine, Shuang Ho Hospital
- ⁸ Division of Cardiology, Department of Internal Medicine, Shuang Ho Hospital, Taipei Medical University, No.291, Zhongzheng Rd., Zhonghe District, New Taipei City 23561, Taiwan
- ⁹ Taipei Heart Institute, Taipei Medical University, Taipei City 110301, Taiwan
¹⁰ Division of Cardiology Department of Internal Medicine, School of Medicin
- ¹⁰ Division of Cardiology, Department of Internal Medicine, School of Medicine, College of Medicine, Taipei Medical University, New Taipei City 23561, Taiwan
- ***** Correspondence: myhung6@ms77.hinet.net; Tel.: +886-2-224-900-88; Fax: +886-2-827-337-36 (ext. 2714)
- † These authors contributed equally to this work.

Abstract: Although heart failure (HF) is a clinical syndrome that becomes worse over time, certain cases can be reversed with appropriate treatments. While coronary artery spasm (CAS) is still underappreciated and may be misdiagnosed, ischemia due to coronary artery disease and CAS is becoming the single most frequent cause of HF worldwide. CAS could lead to syncope, HF, arrhythmias, and myocardial ischemic syndromes such as asymptomatic ischemia, rest and/or effort angina, myocardial infarction, and sudden death. Albeit the clinical significance of asymptomatic CAS has been undervalued, affected individuals compared with those with classic Heberden's angina pectoris are at higher risk of syncope, life-threatening arrhythmias, and sudden death. As a result, a prompt diagnosis implements appropriate treatment strategies, which have significant life-changing consequences to prevent CAS-related complications, such as HF. Although an accurate diagnosis depends mainly on coronary angiography and provocative testing, clinical characteristics may help decision-making. Because the majority of CAS-related HF (CASHF) patients present with less severe phenotypes than overt HF, it underscores the importance of understanding risk factors correlated with CAS to prevent the future burden of HF. This narrative literature review summarises and discusses separately the epidemiology, clinical features, pathophysiology, and management of patients with CASHF.

Keywords: heart failure with preserved ejection fraction; heart failure with reduced ejection fraction; coronary artery spasm

1. Introduction

Heart failure (HF) is a clinical, heterogeneous syndrome stemming from any structural or functional ventricular impairment of diastolic filling or systolic ejection fraction or both [\[1\]](#page-23-0). Because the left ventricular ejection fraction (LVEF) has a bimodal distribution among HF patients [\[2\]](#page-23-1), LVEF has been a phenotypic marker indicative of idiosyncratic

Citation: Hung, M.-J.; Yeh, C.-T.; Kounis, N.G.; Koniari, I.; Hu, P.; Hung, M.-Y. Coronary Artery Spasm-Related Heart Failure Syndrome: Literature Review. *Int. J. Mol. Sci.* **2023**, *24*, 7530. [https://](https://doi.org/10.3390/ijms24087530) doi.org/10.3390/ijms24087530

Academic Editors: Isabella Russo and Lih Kuo

Received: 15 January 2023 Revised: 4 April 2023 Accepted: 11 April 2023 Published: 19 April 2023

Copyright: © 2023 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license [\(https://](https://creativecommons.org/licenses/by/4.0/) [creativecommons.org/licenses/by/](https://creativecommons.org/licenses/by/4.0/) $4.0/$).

pathophysiological mechanisms [\[3](#page-23-2)[,4\]](#page-23-3) and, most importantly, response to therapies [\[5\]](#page-23-4); however, regardless of LVEF, the prognosis of HF patients has been also correlated with diastolic dysfunction [\[6\]](#page-23-5). In the general population, LV diastolic dysfunction demonstrates a prevalence of \sim 21%, but only 1.1–5.5% of individuals present symptoms [\[7\]](#page-23-6), suggesting the ischemic cascade beginning with clinically silent diastolic dysfunction has been substantially underrecognized. There is no specific, noninvasive diagnostic test that serves as a gold standard for HF diagnosis since it is a clinical entity established upon a careful history, physical examination, laboratory, and imaging data. The clinical diagnostic gold standard of HF is the identification of an elevated pulmonary capillary wedge pressure at rest or exercise on an invasive hemodynamic exercise test in a symptomatic patient [\[8\]](#page-23-7). While most patients with suspected HF do not require invasive testing for diagnosis, an echocardiogram constitutes often the best method for HF diagnosis [\[9\]](#page-23-8).

In the US, approximately 50% of HF patients have LVEF \geq 50%, with the balance having LVEF <50% [\[10,](#page-24-0)[11\]](#page-24-1). While HF with reduced ejection fraction (HFrEF) <50% is a final common pathway of systolic dysfunction due to various etiologies [\[12\]](#page-24-2), numerous drugs and cardiac devices have been reported to enhance outcomes in patients with HFrEF independently of etiology, demonstrating that patients with HFrEF share similar pathophysiological pathways to the progression of systolic dysfunction [\[12\]](#page-24-2). In contrast to HFrEF, such a unifying pathophysiological adaptation is lacking in HF with preserved ejection fraction (HFpEF) \geq 50% [\[13\]](#page-24-3), which has proved to be the main form of HF worldwide because of aging of the general population and the augmenting prevalences of obesity, diabetes mellitus, and hypertension [\[13\]](#page-24-3). Because the relevant therapy should target the different underlying etiologies, pathophysiologies, and comorbidities [\[3,](#page-23-2)[14–](#page-24-4)[16\]](#page-24-5), patients with HFpEF may respond in a less homogenous way to treatment. Furthermore, the long-term survival rates in HFpEF patients are lower than those in HFrEF patients, although mostly driven by non-cardiovascular causes [\[17\]](#page-24-6). Notably, HF is considered as treatment failure rather than an indication for therapy [\[18\]](#page-24-7); future attempts to reduce HF burden should focus not only on reducing or averting exposure to risk factors but also on the management of comorbidities. However, neither the molecular mechanisms underlying HF, irrespective of LVEF, nor effective prevention strategies are fully understood.

Basic science evidence, epidemiological studies, and clinical trials suggest that coronary artery disease (CAD), including epicardial CAD and coronary microvascular disease (CMD) [\[19\]](#page-24-8), is a significant contributor to HF pathogenesis. In patients with HFrEF, CAD constitutes frequently the main cause [\[20\]](#page-24-9). Thus, up to 25% of HF patients classified clinically as "nonischemic cardiomyopathy," might reveal evidence of CAD at autopsy [\[21\]](#page-24-10), and ischemic changes have also been reported in endomyocardial biopsies [\[22\]](#page-24-11) in such patients. On the other hand, coronary artery spasm (CAS), an excessive coronary vasoconstriction leading to total or subtotal vascular obstruction, has been considered one of the causes of HFrEF [\[23–](#page-24-12)[25\]](#page-24-13).

As a separate entity from classic angina pectoris (pectoris dolor) described by Dr. William Heberden (1710–1801) based on 20 cases with this affliction in 1772 [\[26\]](#page-24-14), which started when chest pain was evoked by exercise or emotional stress, and was relieved by rest or nitroglycerin, in 1959, with Dr. Rexford Kennamer and others, Dr. Myron Prinzmetal (1908–1987) published the first landmark report [\[27\]](#page-24-15) of their findings on "A variant form of angina pectoris". In Prinzmetal's first report, among the 32 cases of variant angina between 1931–1956, of which 20 and 12 were personally observed and reported in the literature, respectively, the pain appeared at rest or during daily routine activity but was not caused by exercise or emotional stress. Among the reported 32 patients, 12 had myocardial infarction [MI] at follow-up [\[27\]](#page-24-15). Because both forms of angina pectoris had coronary atherosclerosis in common post-mortem, and the variant angina attack usually happened at rest, when the vascular physiologically hypertonic action is greatest [\[27\]](#page-24-15), the mechanism for variant angina proposed by Prinzmetal et al., or other researchers was arterial hypertonus or CAS, respectively. Of note, following the first report of coronary angiography in 1959, CAS in

variant angina had never been proved angiographically [\[28,](#page-24-16)[29\]](#page-24-17) within a decade until the early 1970s [\[29–](#page-24-17)[31\]](#page-24-18).

It was common for coronary angiography in the 1970s and 1980s to diagnose CAS in the catheterization laboratory. It turned out increasingly clear that CAS could occur in a patient with [\[26](#page-24-14)[,27\]](#page-24-15) or without atherosclerotic obstructive CAD, referred to as "variant of the variant" [\[30\]](#page-24-19). Moreover, CAS is more frequently associated with ST-segment depression rather than non-progressive elevation [\[32,](#page-24-20)[33\]](#page-24-21). Hence, the term "variant angina" is specifically reserved for CAS-induced angina with temporary ST-segment elevation (Figure [1\)](#page-2-0). In addition, CAS-related acute coronary syndrome can be due to anaphylactic reactions, involving the release of inflammatory mediators such as histamine, chymase, leukotrienes, and platelet-activating factor from mast cells upon activation to cause the constriction of coronary vascular smooth muscle cells that constitute the pathophysiologic mechanism of Kounis syndrome [\[34](#page-24-22)[,35\]](#page-24-23). Altogether, in coronary heart disease, atherosclerotic obstructive CAD cannot be regarded as the sole source of angina pectoris [\[36\]](#page-24-24).

Figure 1. The 12-lead electrocardiograms and coronary angiography of variant angina. Angina attack (**A**) and post-sublingual nitroglycerin 0.6 mg (**B**) 12-lead electrocardiograms of a 47-year-old male revealed brief ST-segment elevation in II, III, and aVF leads. Ten months later because of recurrent chest pain, he underwent coronary angiography. The coronary angiography revealed intracoronary methylergonovine-induced CAS in the middle portion of the right coronary artery (**C**, arrow), which was alleviated after intracoronary nitroglycerin 200 µg (**D**). (Reproduced from [\[33\]](#page-24-21), with permission of the publisher.).

Most importantly, the use of nitroglycerin at the beginning of coronary angiography should be avoided [\[37\]](#page-25-0) to prevent inadvertent abrogation of spontaneous CAS. However, nitroglycerin solution has to be fully prepared before performing CAS provocative testing to relieve established CAS promptly through intracoronary infusion [\[33\]](#page-24-21). Therefore, 2 sets of coronary angiograms pre- and post-intracoronary nitroglycerin should be obtained routinely once obstructive lesions are noted. Spontaneous CAS can be misdiagnosed as a candidate for percutaneous coronary intervention unless the alleviation of obstructive stenosis is documented after intracoronary nitroglycerin, emphasizing the importance of intracoronary nitroglycerin infusion before endeavored coronary intervention, and avoiding unnecessary coronary revascularization [\[38\]](#page-25-1). Because coronary revascularization in selected obstructive CAD patients can ameliorate diastolic dysfunction, decrease morbidity and mortality [\[39–](#page-25-2)[42\]](#page-25-3), and enhance systolic function [\[43\]](#page-25-4), likely by improvement of hibernating myocardium, medical treatments in non-obstructive CAD, such as CAS, patients may similarly improve left ventricular diastolic and systolic function.

2. Epidemiology

Framingham Heart Study has demonstrated that, from the 1950s onwards, the role of myocardial ischemia and infarction has developed substantially [\[44](#page-25-5)[,45\]](#page-25-6), placing great emphasis on the prevention of HF through the prevention of myocardial ischemia and infarction. Despite the fact that the cumulative incidence of HF is similar between both genders, women are approximately 65% less likely than men to develop HFrEF, particularly in their younger years [\[45](#page-25-6)[–47\]](#page-25-7), while HFpEF is twice as common in women than men, which results from physiologic differences between the two genders [\[48\]](#page-25-8). On the other hand, survival after a diagnosis of HF, irrespective of HFrEF or HFpEF, has shown modest improvement in the 21st century and lags behind other serious conditions, such as cancer [\[49\]](#page-25-9). Hence, contributing factors require further clarification, among which CAS is becoming important. There are large variations in CAS prevalence across the world, as CAS frequency is greater in Japan than in west countries [\[50\]](#page-25-10). The prevalence of CAS is high (40%) among patients showing evanescent, resolving ST-segment elevation admitted to Japanese hospitals [\[51\]](#page-25-11). Moreover, using provocative testing, CAS of more than two coronary arteries appears more frequently in Japanese (24.3%) [\[52\]](#page-25-12) and Taiwanese populations (19.3%) [\[53\]](#page-25-13) than in Caucasians (7.5%) [\[54\]](#page-25-14). On the other hand, men are more likely than women to develop CAS both in East Asia and Western countries [\[51](#page-25-11)[,53\]](#page-25-13). Most CAS appears in people aged 40 to 70 years and the prevalence declines after the age of 70 years [\[27,](#page-24-15)[51,](#page-25-11)[53\]](#page-25-13). Several studies have demonstrated that CAS prevalence in patients without obstructive CAD is around 50% in angina and, specifically, 57% in acute coronary syndrome in Asia [\[55](#page-25-15)[–57\]](#page-25-16). Among provocative tests using intracoronary acetylcholine for functional vasomotor abnormalities in acute coronary syndrome without obstructive CAD, 79% of individuals demonstrate a positive finding in Japan [\[58\]](#page-25-17), whereas the results are positive in 16% of French [\[59\]](#page-25-18) and 49% of German [\[57\]](#page-25-16) patients. Notably, CAS diagnosis can be challenging due to pretreatment with antispastic nitroglycerin or calcium channel antagonists, refraining from coronary constrictors, and changes in disease activity. Additionally, contemporary trends of CAS prevalence tend to decline in Japan because of reduced performance of misperceived time-consuming provocative tests, or extensive use of statins and calcium channel antagonists [\[58\]](#page-25-17).

CAS is an exceptionally complex multifactorial disease in which smoking, inflammation, metabolic, psychosocial, and physical factors come into play. Although it was reported more than 20 years ago [\[50\]](#page-25-10), the racial differences in coronary vasomotion disorders between Asian and Caucasian populations remain controversial. First, previous studies show that epicardial CAS is more often recognized in Japanese and Taiwanese people than in Caucasian populations, while CMD is typically observed in Caucasian patients, which may be because Japanese and Taiwanese cardiologists have performed spasm provocation testing actively for 30 and 20 years, respectively, in patients with nonobstructive CAD, whereas most Caucasian cardiologists do not perform provocative testing for nonobstructive CAD

in the cardiac catheterization laboratory [\[60\]](#page-25-19). However, for an unknown reason, some Taiwanese cardiologists are resistant to acknowledging the existence of CAS, which affects patients' physical and psychological quality of life, and as a result, oppose performing provocative testing for the diagnosis of CAS. Second, various diagnostic procedures are performed worldwide, such as intravenous ergonovine-provoked >70% luminal reduction in France [\[61\]](#page-26-0) and intracoronary acetylcholine-provoked >75% luminal reduction in Germany [\[57\]](#page-25-16). Third, according to a Japanese study [\[62\]](#page-26-1), intracoronary acetylcholine administration time is crucial to provoke CAS. Slow injection of acetylcholine for 3 min may induce microvascular CAS, whereas rapid injection of acetylcholine for 20–30 s may provoke epicardial CAS, leading to inconsistency in the prevalence and incidence of CAS between Japanese and Caucasian patients. Fourth, Japanese cardiologists have stated that in some European institutions, acetylcholine testing without pacemakers is employed, which may cause bradycardia or cardiac arrest in the right coronary artery rendering difficult interpretation of provocative testing. If Caucasian cardiologists perform provocative testing with pacemakers similar to Japanese cardiologists, the prevalence and incidence of CAS may be higher than ever thought. However, in Taiwanese specialists' experiences without implementing pacemakers when performing provocative testing using the bolus injections of ergonovine, there has been no cardiac arrest but only rarely mild bradycardia, which can quickly return to normal after immediate intracoronary administration of nitroglycerin once CAS occurs [\[63,](#page-26-2)[64\]](#page-26-3). Fifth, the definition of positive epicardial CAS is different among previous Japanese, Taiwanese, and Caucasian studies [\[60\]](#page-25-19). For example, the definition of provoked CAS is a reduction of >50% [\[65\]](#page-26-4), >70% [\[61](#page-26-0)[,66](#page-26-5)[,67\]](#page-26-6), >75% [\[57,](#page-25-16)[68–](#page-26-7)[71\]](#page-26-8), >90% [\[24,](#page-24-25)[51,](#page-25-11)[71,](#page-26-8)[72\]](#page-26-9), or 99–100% [\[73\]](#page-26-10) in luminal diameter compared with postintracoronary nitroglycerin. Sixth, among all the clusters of CAS risk factors, the predominant factors that cause CAS in Asian patients may be different from those in white patients. Notably, it is estimated that nearly 1 billion people globally, most of whom are Asians, carry the Glu504Lys polymorphism in the aldehyde dehydrogenase 2 (ALDH2) gene [\[74\]](#page-26-11). This ALDH2 mutant is significantly associated with a high level of high-sensitivity C-reactive protein [\[75\]](#page-26-12), which is a risk factor for CAS. In conclusion, while previous studies demonstrate that existence of racial heterogeneity in coronary vasomotor response [\[50\]](#page-25-10), the prevalences of CAS and CAS-related HF (CASHF) in different populations are largely unknown.

In the US, while the incidence and prevalence of HF are increased [\[11](#page-24-1)[,76\]](#page-26-13), the agespecific incidence of HF might be reduced, but to a lesser degree in HFpEF compared with HFrEF [\[77\]](#page-26-14). A UK study showed that the age-adjusted incidence of HF fell by 7% between 2002 and 2014, whereas the absolute incidence of HF increased by 12%, and prevalent HF increased by 23% [\[78\]](#page-26-15). This growth in the absolute number indicates population aging, reduced mortality from cardiovascular diseases, including MI [\[78\]](#page-26-15), and the increasing prevalence of risk factors. Given that approximately 50% of HFrEF cases can be attributed to ischemia [\[79\]](#page-26-16), a new diagnosis of HFrEF frequently needs an assessment for underlying CAD. Despite the fact that individual-level factors (eg, old age, serious comorbidities, noncandidates, or no preference for coronary revascularization) should be evaluated before referral, coronary angiography remains the gold standard for diagnosis of obstructive CAD [\[80\]](#page-26-17). In addition to epicardial CAD, microvascular CAD is becoming widespread and often under-recognized [\[81\]](#page-26-18); hence, both epicardial and microvascular CAD, clinically overt or silent, acute or chronic, can result in decreased perfusion, myocardial damage, and further reduced myocardial function.

3. Clinical Features of CASHF

Until recently, patients with non-obstructive CAD were often inappropriately reassured due to assuming a favorable prognosis without further investigation, although clinical features may require coronary angiography. However, ischemia with non-obstructive CAD (INOCA) is a non-benign condition correlated with an equivalent incidence of adverse events as well as poor quality of life compared to obstructive CAD [\[82\]](#page-26-19). INOCA is defined as when patients present with symptoms and signs suggesting ischemia but are found to

have no obstructive CAD at coronary angiography [\[83\]](#page-26-20). Indeed, this phenomenon is a primary cause of myocardial ischemia and is linked to a high risk of MI, decompensated HF, stroke, and unexpected sudden death [\[84](#page-27-0)[–90\]](#page-27-1). On the other hand, CAS-related angina is common [\[27](#page-24-15)[,91\]](#page-27-2), although not as frequent as classic Heberden's angina. While in the Taiwanese general population, the prevalence of CAS and obstructive CAD over 12 years of follow-up is 0.067% and 8.7%, respectively [\[92\]](#page-27-3), the prevalence of CAS in other racial populations needs to be clarified and the frequency of CAS diagnosis might be increased when careful criteria are applied for its detection [\[91\]](#page-27-2). In Japan, although non-invasive provocation tests such as hyperventilation tests and cold pressor tests decreased remarkably in 2014, diagnosis by invasive provocation tests and the occurrence of CAS-related angina increased in 2014 compared with 2008, albeit not significantly [\[93\]](#page-27-4). CAS provocation tests using left-to-right coronary sequential evaluation were employed in just 30% of the Japanese hospitals; hence, although 40% of the centers were dissatisfied with standard spasm provocation tests, the majority of the hospitals confirmed the necessity of CAS provocation tests in the future [\[93\]](#page-27-4). As a result, considering that (1) CAS can cause resting angina with S-T segment depression and/or pseudonormalization of T waves; (2) asymptomatic CAS-induced angina is common [\[91](#page-27-2)[,94\]](#page-27-5); and (3) cold-induced angina may arise from CAS [\[95\]](#page-27-6), the incidence of CAS-induced myocardial ischemia could be doubtlessly much higher than that of the public perception indicated by the clinical presentation.

In clinical practice, angina pectoris results from a temporary myocardial oxygensupply-demand imbalance [\[96](#page-27-7)[,97\]](#page-27-8), causing 2 types of ischemia, exertional demand ischemia and non-exertional supply ischemia [\[98\]](#page-27-9). Because coronary disorders are dynamic [\[99\]](#page-27-10), nonobstructive CAD might become flow-limiting stenosis if the vascular tone is increased [\[99\]](#page-27-10), and hence increased oxygen demand might not always precede myocardial ischemia [\[100\]](#page-27-11). In this regard, endothelial dysfunction makes up about two-thirds of symptomatic patients of INOCA and a smaller percentage of "MI with non-obstructive CAD" (MINOCA). The clinical spectrum of INOCA includes epicardial CAS, microvascular CAS, or mixed epicardial/microvascular CAS [\[19\]](#page-24-8). Microvascular CAS can cause myocardial necrosis, mild elevations of cardiac troponin, subtle left ventricular contractile abnormalities [\[101\]](#page-27-12), and early-stage HF [\[102\]](#page-27-13). Previous studies showed that microvascular CAS could be demonstrated not only in angina patients with normal epicardial coronary arteries but also in HFpEF [\[4,](#page-23-3)[103–](#page-27-14)[105\]](#page-27-15); hence, angina and dyspnea can appear at 2 extremes in the presence of a continuum of disease contributing to the development of microvascular CAS and HFpEF [\[106\]](#page-27-16). Furthermore, 30–48% of in-patients receiving the optimal treatment for HFrEF <45% have provoked epicardial CAS [\[23](#page-24-12)[,24](#page-24-25)[,67](#page-26-6)[,72\]](#page-26-9). While the prevalence of hypertension and smoking are higher in epicardial CAS-related than non-CAS-related HFrEF [\[24\]](#page-24-25), more research is required to evaluate the risk factors of epicardial CASHFrEF.

Various arrhythmias, especially ventricular premature complex, more often appear in >50% of CAS-induced angina than in classic Heberden's angina pectoris [\[39](#page-25-2)[,107\]](#page-28-0), albeit through unknown mechanisms but possibly involving QT dispersion in CAS-induced cardiac arrest and syncope [\[108\]](#page-28-1). While the severity of CAS has no relationship with the occurrence of these arrhythmias, ventricular arrhythmias occur more frequently during anterior wall ischemia [\[109\]](#page-28-2); however, right CAS-induced ventricular arrhythmias are not uncommon (Figure [2\)](#page-6-0). Besides, ventricular fibrillation (VF) complicating CAS is responsible for sudden death with morphologically normal coronary arteries in autopsies, as previously reported [\[110\]](#page-28-3). Although cardioversion is always required to terminate VF, VF induced by CAS, epicardial or microvascular, rarely terminates spontaneously [\[50,](#page-25-10)[80\]](#page-26-17) (Figures [3](#page-7-0) and [4\)](#page-8-0). Additionally, in a study of patients who had implanted cardioverter defibrillators, VF was asymptomatic in 43% and nonsustained in 40% of episodes [\[111\]](#page-28-4). The probability of syncope or pre-syncope is 25% and 62% when VF is <10 and \geq 10 s, respectively [\[111\]](#page-28-4). On the other hand, about 40% of CAS-related inferolateral J wave and VF does not cause angina at the first VF, and could have been misinterpreted as early repolarization syndrome [\[112\]](#page-28-5), which in the younger age group is associated with features of CAS such as lower systolic blood pressure and lower heart rate [\[113\]](#page-28-6). Therefore, CAS is essential and should be included

in the differential diagnosis of syncope and early repolarization syndrome, prompting optimal medical management.

Figure 2. Epicardial CAS-induced ventricular ectopics: 24 h Holter monitor, electrocardiograms, pressure tracing, and right coronary arteriogram in a 53-year-old female presenting with frequent palpitation and unstable rest angina. (**A**) A 24 h Holter monitor showed sinus rhythm with runs of ventricular ectopics in singles and couplets without preceding ST segment changes; (**B**) simultaneous lead I, II, III electrocardiogram and systemic arterial pressure tracing during intracoronary ergonovine testing; (**C**) baseline angiographically normal right coronary artery with minimal plaquing; (**D**) middle spasm (arrow) immediately after intracoronary administration of 45 µg ergonovine. Ventricular ectopics in singles and one couplet occurred at the same time; (**E**) the CAS and ventricular ectopics were relieved after intracoronary administration of 200 µg nitroglycerin. The patient's consciousness remained clear throughout the examination.

Figure 3. Epicardial CAS-induced VF: electrocardiograms, pressure tracing, and right coronary arteriogram in a 50-year-old male with unstable angina, presenting after wakening with rest angina at night. (**A**) Simultaneous lead I, II, aVR electrocardiogram, and systemic arterial pressure tracing during intracoronary ergonovine testing; (**B**) baseline angiographically normal right coronary artery with minimal plaquing; (**C**) ostial spasm (arrow) immediately after intracoronary administration of 15 µg ergonovine; (**D**) in 10 s, the ostial spasm recovered spontaneously, multi-focal spasms appeared in the proximal and middle portion, and ventricular fibrillation occurred at the same time for 10 s and recovered spontaneously without intervention; (**E**) multi-focal spasms were relieved after intracoronary administration of 100 µg nitroglycerin. The patient's consciousness remained clear throughout the examination. (Reproduced from [\[63\]](#page-26-2), with permission of the publisher).

On the other hand, extreme CAS may cause life-threatening pulseless electrical activity or asystole without the occurrence of ventricular tachycardia or VF [\[50\]](#page-25-10), which, when involving all of the three epicardial coronary arteries, can suddenly stop heartbeat due to pulseless electrical activity and flash freeze the whole myocardium immediately, leading to invisible coronary flow [\[114\]](#page-28-7) and, despite intracoronary administration of nitroglycerin, prolonged contrast retention in the coronary arteries. While extended continuous cardiac massage is effective to resolve CAS-related pulseless electrical activity [\[114\]](#page-28-7), cardiac pacing or implantable cardioverter defibrillator may not be feasible for the recovery of viable muscle from frozen myocardium during pulseless electrical activity, and may result in unexplained death [\[114](#page-28-7)[,115\]](#page-28-8). Furthermore, ischemia of the sinus node or atrioventricular node arteries due to CAS can affect the development of pulseless electrical activity or asystole [\[114\]](#page-28-7). Taken together, without the induction of ventricular arrhythmias, CAS can directly result in pulseless electrical activity or asystole.

Figure 4. Microvascular CAS-induced VF: electrocardiograms (green line), pressure tracing (purple line), and right coronary arteriogram in a 75-year-old male with unstable rest angina. (**A**) Simultaneous lead I, II, III electrocardiogram and systemic arterial pressure tracing during intracoronary ergonovine testing; (**B**,**C**) baseline angiographically normal right coronary artery with minimal plaquing (white and red arrows); (**D**,**E**) microvascular spasm (white and red arrows) immediately after intracoronary administration of 45 µg ergonovine. Ventricular fibrillation occurred at the same time for 17 s and recovered spontaneously without intervention; (**F**,**G**) microvascular spasms were relieved after intracoronary administration of 200 µg nitroglycerin (white and red arrows). The patient's consciousness remained clear throughout the examination.

4. Pathogenesis

CAS not only causes remarkable progression of CAD but also demonstrates a "jumpup" phenomenon in as short as 25 min in a swine model [\[116\]](#page-28-9), in which CAS superimposed on minimal coronary stenosis can rapidly progress to total atherosclerotic obstruction, resulting in MI [\[117\]](#page-28-10). In a pig model, the phenomenon can be explained by that CAS of abrupt rather than gradual onset can cause intramural hemorrhage in the plaque's neovasculature and the subsequent sudden progression of organic coronary stenosis, leading to MI [\[118\]](#page-28-11). Most importantly, while smoking, age, C-reactive protein (CRP) [\[50\]](#page-25-10), ALDH2 deficiency [\[119\]](#page-28-12), and lipoprotein(a) [\[120\]](#page-28-13) are risk factors for CAS, CAS is not associated with the classic risk factors for CAD [\[50](#page-25-10)[,121\]](#page-28-14), such as diabetes mellitus, hypertension [\[53\]](#page-25-13), hypercholesterolemia [\[50,](#page-25-10)[121\]](#page-28-14) and obesity [\[50](#page-25-10)[,121\]](#page-28-14), suggesting pathophysiological differences exist between CAS and CAD. While risk factors in an individual usually exist together and have a cumulative and interactive effect to increase a person's chance of getting CAS (Figure [5\)](#page-9-0), precipitating factors refer to a specific event, which may act in the same patient to cause the onset of CAS in various circumstances. Notably, while older rather than younger people are more likely to develop CAS, smoking in the younger compared with their older analogs has a more powerful effect on CAS occurrence [\[122\]](#page-28-15). In addition, as smoking and age appear to have a more important role in men [\[123\]](#page-28-16), CAS risk factors may be gender-specific.

The relaxation and contraction of vascular smooth muscle cells are regulated primarily through dephosphorylation and phosphorylation of the myosin light chain, respectively. CMD is characterized by impaired microvascular smooth muscle cell dilation, which can ultimately lead to HFpEF [\[124\]](#page-28-17). In addition, elevated Rho-kinase activity of smooth muscle cells favors contraction by directly increasing sensitization of the myosin light chain to Ca^{2+} and indirectly augmenting phosphorylation of the myosin light chain [\[125\]](#page-28-18). The Rho-kinase activity in vascular smooth muscle cells is elevated after wrapping the coronary arteries with interleukin (IL)-1β beads in a pig CAS model [\[125](#page-28-18)[–127\]](#page-28-19). Other animal models of spontaneous CAS include KATP mutant or SUR2 KATP knockout mice, suggesting that loss of function of KATP channels can induce hypercontraction of smooth muscle cells without atherosclerosis [\[128](#page-28-20)[,129\]](#page-28-21). Mice lacking α 1H T-type calcium channels show a

normal contraction of coronary arteries but decreased response in acetylcholine-induced relaxation [\[130\]](#page-28-22). Together, these models reveal that hyperreactivity of vascular smooth muscle cells can cause CAS via various pathways, whereas their clinical relevance in humans remains largely unknown.

Figure 5. Risk factors and precipitating factors are represented by rectangles and circles, respectively, for CAS. (Adapted from [\[33\]](#page-24-21), with permission of the publisher).

The majority of CASHF patients present with a less severe phenotype (stages A and B) than overt HF, among which stage A is for patients at risk for HF but without current or prior symptoms or signs of HF and structural or biomarker evidence of heart disease, and stage B is for patients without current or prior symptoms or signs of HF but evidence of structural heart disease or abnormal cardiac function, or elevated natriuretic peptide levels [\[131\]](#page-29-0), underscoring the importance of understanding risk factors for CAS to prevent the future burden of HF. Among the risk factors of CAS [\[132\]](#page-29-1), smoking (relative risk 1.47) is independently associated with incident HF [\[133\]](#page-29-2). While no circulatory factor impeding oxygen supply to the heart such as fixed coronary stenosis or exercise is responsible for eliciting CAS-related angina, from the onset of the electrocardiographic abnormalities to the start of their reversion, the mean heart rate and arterial blood pressure are decreased, and isovolumic contraction time is lengthened, reducing the left ventricular performance [\[134\]](#page-29-3)

Inflammation has been shown to account for the dissimilarities in cardiac remodeling between HFpEF and HFrEF. While HFpEF is linked to concentric hypertrophy, adverse remodeling in HFrEF is often due to ischemia-induced progressive loss of cardiomyocytes, with a patchy distribution of replacement fibrosis of dead cells by collagen, leading to LV dilatation and maladaptive remodeling [\[135](#page-29-4)[–138\]](#page-29-5). Furthermore, because pathophysiological differences exist between HFpEF and HFrEF, the inflammatory biomarkers, including CRP and IL-6, are higher in HFpEF than in HFrEF, while markers of cardiomyocyte injury, such as high-sensitivity troponin T and brain natriuretic peptides, are higher in HFrEF than in HFpEF [\[139\]](#page-29-6).

Chronic myocardial dysfunction resulting from hypoperfusion, hibernation, or both, may also increase the risk of HF [\[140](#page-29-7)[,141\]](#page-29-8). Subjects with both epicardial CAD and CMD may have chronic hypoperfusion-associated inflammation and fibrosis, resulting in increased myocardial stiffness. Similarly, episodic coronary hypoperfusion such as CAS may cause myocardial functional impairment for hours to days (myocardial stunning) [\[142\]](#page-29-9). Studies with positron emission tomography [\[143\]](#page-29-10) and single photon emission computed tomography [\[144\]](#page-29-11) show decreased blood flow and glucose uptake in myocardial areas that concomitantly have decreased systolic function. In addition, ventricular diastolic dysfunction appears in both experimental [\[145\]](#page-29-12) and clinical ischemia [\[146\]](#page-29-13). Of note, the decrease in coronary vasodilator reserve is proportional to extent of arterial luminal stenosis [\[147](#page-29-14)[,148\]](#page-29-15). Consequently, myocardium with normal resting blood flow may have decreased exercise blood flow and may display decreased glucose metabolism on positron emission tomography during exercise and a concomitant decrease in ventricular function.

As associated researchers at the National Human Genome Research Institute unlock the mysteries of the complete set of the human genome, almost every disease has a genetic component [\[149\]](#page-29-16), which is the case with CAS in that a mutation in the ALDH2 gene is believed to be the cause [\[150\]](#page-29-17) and associated with Asian flush syndrome. Mizuno Y et al., showed that Asians with defective ALDH2*2 alleles have a higher risk of CAS. They also found that the defective gene positively interacts with the detrimental effects of smoking on stronger vasoconstriction than each factor alone by increasing reactive aldehydes [\[151\]](#page-29-18). Furthermore, other Japanese studies show that the mutant ALDH2*2 allele carriers compared with subjects with the ALDH2*1/1 genotype have higher frequencies of more severe CAS-related myocardial injury [\[119](#page-28-12)[,152\]](#page-29-19).

The mechanisms of coronary vasomotor disorders can be endothelium-dependent or endothelium-independent [\[19\]](#page-24-8). While endothelium-dependent dysfunction results from an endothelium-derived disparity between relaxing factors, e.g., nitric oxide (NO), and constrictors, e.g., endothelin [\[19\]](#page-24-8), endothelium-independent function relies on vascular myocyte tone [\[19\]](#page-24-8). Endothelial dysfunction, as in CAS, reduces the bioavailability of nitric oxide (NO), cyclic guanosine monophosphate, and protein kinase G in adjacent cardiomyocytes [\[153\]](#page-29-20), contributing to myocardial fibrosis and HFpEF [\[154](#page-29-21)[,155\]](#page-30-0). While during HF development, the initial inflammatory response is a protective reaction to tissue injury, it may lead to irreversible damage when the inflammation is prolonged. Pathologic features, common to all cardiomyopathies irrespective of origin, include ventricular hypertrophy, fibrosis, scarring, and dilatation [\[102\]](#page-27-13). This phenomenon was investigated in 2 animal models of congestive cardiomyopathy: the hereditary cardiomyopathic Syrian hamster and the hypertensive-diabetic rat [\[102\]](#page-27-13). In both the genetic and the acquired disease models, there was focal myocytolytic necrosis with the subsequent healing with focal scars, ventricular hypertrophy, ventricular dilatation with congestive HF, and, finally, death [\[102\]](#page-27-13). In both diseases, the microcirculation of the animal hearts had been studied by the use of silicone rubber perfusions; microvascular CAS was demonstrated early in the disease associated with small areas of myocytolytic necrosis and subsequent fibrosis [\[102\]](#page-27-13). Because the distance between cardiomyocytes and endothelial cells is fewer than $3 \mu m$ [\[19\]](#page-24-8), allowing for sufficient blood supply and bidirectional influences, both myocardial fibrosis- and hypertrophy-induced subendocardial ischemia may cause left ventricular diastolic dysfunction and longitudinal systolic abnormalities, leading to remodeling and HFpEF, which may reciprocally trigger subendocardial ischemia and endothelial dysfunction in return [\[156\]](#page-30-1). Collectively, although myocardial ischemia directly contributes to HFpEF [\[157\]](#page-30-2), the causes and mechanisms contributing to HF as well as CASHF, albeit largely unknown, are likely multifactorial (Table [1\)](#page-11-0).

Table 1. Proposed mechanisms of coronary artery spasm-related heart failure.

4.1. Microvascular CASHF

Several cardiac and systemic disorders, such as HFpEF, brain small-vessel disease, diabetes, hypertension, chronic inflammatory and autoimmune diseases, and chronic kidney disease can develop INOCA [\[198](#page-31-17)[,199\]](#page-31-18). In most of these patients, close relationships exist between microvascular dysfunction and atherosclerotic epicardial CAD [\[198](#page-31-17)[,199\]](#page-31-18). In

early diabetic rats, the coronary microvascular focal and segmental constrictions occur when prostacyclin and nitric oxide production is prevented, which, if left untreated in advanced diabetes, will progress to irreversible microvascular damage [\[200\]](#page-31-19). On the other hand, while CMD does not develop atheroma in accord with epicardial atherosclerosis, coronary microcirculation in patients carrying cardiovascular risk factors can evolve into structural and functional atherosclerotic-like changes [\[19\]](#page-24-8), presenting as either vasodilator abnormality and/or microvascular CAS [\[19\]](#page-24-8). Furthermore, CMD can occur in the absence or presence of obstructive epicardial CAD [\[201\]](#page-31-20). As a result, the CMD-related myocardial ischemias are unlike those attributable to epicardial flow-limiting stenosis, in which the regional ischemia perfused by the obstructed epicardial artery is homogeneously distributed, resulting in regional wall motion abnormality [\[19\]](#page-24-8). In contrast, myocardial ischemia in CMD may appear as patchy and not entail all microvessels originating from an epicardial artery, causing symptoms without wall motion abnormalities [\[202\]](#page-31-21), or as a generalized phenomenon resulting in diffuse perfusion and wall motion abnormalities, and thereby HFrEF with a normal result on noninvasive stress imaging tests.

Circulating factors, such as fibrocytes, circulating monocyte-derived cells, and fibroblasts, might regulate the effects of microvascular CAS favoring the development of left ventricular fibrosis and hypertrophy [\[203\]](#page-32-0). While fibrocytes are recruited to chronically injured myocardium in cardiac remodeling in mice, treatment with serum amyloid P decreased fibrocyte accumulation and fibrosis [\[204\]](#page-32-1). Another modulatory factor, atrial natriuretic peptide, may induce phosphorylation of Smad proteins, thus inhibiting their nuclear translocation and binding to TGF-Smad responsive elements in the promoter regions of extra-cellular matrix genes [\[205\]](#page-32-2). An auxiliary potential mechanism, as proposed by Pepine et al. [\[206\]](#page-32-3), involves repetitive cycles of ischemia-reperfusion such as in CAS that impede cardiac myocyte relaxation thereby causing diastolic dysfunction and HFpEF.

In addition, endothelial dysfunction associated with inflammation reduces the content of cyclic guanosine monophosphate (cGMP), protein kinase G (PKG), and transforming growth factor (TGF)-β in cardiomyocytes and microvascular NO bioavailability, all of which are involved in the physiological modulation of cardiac hypertrophy and stiffness [\[207](#page-32-4)[–210\]](#page-32-5) Besides, NO reduction inhibits cGMP and TGF-β functions, favoring conversion of endothelial cells into mesenchymal cells such as fibroblasts [\[210–](#page-32-5)[212\]](#page-32-6). Overall, these changes promote hypertrophy, fibrosis, and the subsequent development of left ventricular diastolic dysfunction.

4.2. Epicardial CASHF

Although epicardial atherosclerosis may induce endothelial dysfunction of CMD, atherosclerotic CMD may in reverse accelerate the development of epicardial atherosclerosis through decreased blood flow and wall shear stress, leading to progressive epicardial endothelial dysfunction [\[213\]](#page-32-7) and thrombus formation [\[214\]](#page-32-8). Similarly, CAS involving epicardial and microvascular arteries is therefore considered the expression of the same CAS development sharing a common pathophysiological milieu that affects the entire coronary circulation [\[215\]](#page-32-9). A substantial body of evidence suggests that subjects with microvascular angina have 2 important extra features contributing to angina symptoms: (1) hyperreactivity of smooth muscle cells to microvascular constrictor stimuli; (2) enhanced awareness of cardiac pain-provoking stimuli. Indeed, a significant number of patients with microvascular angina have microvascular CAS, which is angina accompanied by ST-segment depression after the intracoronary acetylcholine provocative testing [\[106\]](#page-27-16).

A previous study using substance P, a pure endothelial-dependent vasodilator, demonstrates that in patients with variant angina, endothelial dysfunction at sites of CAS is not necessarily present [\[216\]](#page-32-10). Furthermore, in variant angina, several studies fail to show endothelial dysfunction in non-CAS coronary arteries as well as in peripheral arteries [\[217\]](#page-32-11), and other studies also did not show the higher prevalence of NO synthase polymorphismsassociated endothelial dysfunction [\[218\]](#page-32-12). Altogether, an impairment of endotheliummediated vasodilation appears unlikely to cause CAS by itself, although it might facilitate the effects of coronary vasoconstrictors "CAS prone" individuals [\[219\]](#page-32-13).

CAS, particularly multi-focal spasms [\[23\]](#page-24-12), causes myocardial necrosis via reperfusion injury [\[220\]](#page-32-14), leading to reduced diastolic relaxation during angina [\[221\]](#page-32-15), and the subsequent development of HFpEF and HFrEF [\[23–](#page-24-12)[25](#page-24-13)[,67,](#page-26-6)[72\]](#page-26-9). Of note, left ventricular dysfunction may recover in about 2 min to baseline when the electrocardiographic abnormalities start returning to the pre-CAS state [\[36,](#page-24-24)[134\]](#page-29-3).

Takotsubo cardiomyopathy is an acute and reversible form of unexpected physical and emotional distress-related HFrEF featuring symptoms and signs of acute MI without CAD, in which the apex of the left ventricle balloon enlarges to resemble a takotsubo, a Japanese octopus pot [\[222\]](#page-32-16). Despite the syndrome more frequently occurring in older women than in men [\[222\]](#page-32-16), it can affect people of any age, including a newborn after delivery distress with catecholamine-mediated cardiac toxicity [\[223\]](#page-32-17). Precipitating mechanisms are multifactorial and complex, including microvascular and epicardial CAS [\[222\]](#page-32-16), genetics, and thyroid disorders [\[224\]](#page-32-18). Stress activates the sympathetic nervous system to release circulatory catecholamines and the hypothalamic-pituitary-adrenal axis to release circulatory glucocorticoids [\[225\]](#page-32-19). While initially protective for the heart, glucocorticoids not only increase plasma levels of catecholamines by inhibiting uptake but also induce cardiac supersensitivity to catecholamines, leading to an enhanced β-adrenoceptor signal transduction system [\[225\]](#page-32-19). Excessive catecholamines induce diminished apical and enhanced basal wall motion of the left ventricle due to the apicobasal adrenoceptor gradient [\[224\]](#page-32-18). Furthermore, low cate cholamine levels stimulate cardiac $Ca²⁺$ movements, whereas excessive catecholamine levels induce intracellular Ca^{2+} overload in cardiomyocytes, resulting in cardiac dysfunction [\[226\]](#page-32-20). On the other hand, under stressful conditions, high catecholamine levels are oxidized to form oxyradicals, which can cause CAS [\[225\]](#page-32-19). Few cases of Takotsubo cardiomyopathy due to an angiographically confirmed focal, single vessel, or multivessel CAS have been reported. A retrospective analysis in 10 of 48 (21%) Takotsubo cardiomyopathy cases have shown positive provocative CAS, 5 of whom involved both right and left coronary arteries [\[227\]](#page-32-21). Angelini reported 4 cases of Takotsubo cardiomyopathy in which echocardiographic apical ballooning or similar symptoms could be reproduced by provocative CAS [\[228\]](#page-32-22). Moreover, it has also been demonstrated that alternate recurrent CAS and Takotsubo cardiomyopathy can exist in the same individual [\[229\]](#page-33-0). These observations underscore the importance of CAS as a culprit process underlying Takotsubo cardiomyopathy and the targeted treatments accordingly. Further studies will provide critical insights into this unique issue.

4.3. Cellular and Animal Models of Takotsubo Cardiomyopathy, CAS, and Microvascular CASHrEF

Since the late 1800s, because of the similarity in disease processes among animals and humans, animal models began to be developed and help elucidate the connection between dietary cholesterol and atherosclerotic progression [\[230\]](#page-33-1). Since then, the inflammatory and immunological nature of atherosclerosis has been revealed by several studies in patients and experimental models, underscoring the importance of inflammation in CAD, as well as in CAS. Investigation of disease-modifying mechanisms in these models will be crucial for developing future diagnostics and therapy against CAS as well as CASHF. We provide an introduction to experimental models that are used for CAS studies and the research techniques that can be utilized (Table [2\)](#page-15-0). Whether these models can be used for CASHF experiments remains to be elucidated.

Table 2. Proposed models of takotsubo cardiomyopathy and coronary artery spasm.

It stands to reason that the investigator must acknowledge the limitations of animal models so as to construct and interpret relevant experiments cautiously when extrapolated to humans. Problems in cross-species extrapolation and local differences in the arteries are well known. The response of the coronary artery in dogs appeared to be different from that in humans [\[116\]](#page-28-9). Isolated vessels often do not respond in the same way in vitro as in situ, even in the same species. Although, often, only part of the disease is triggered in the animal during an experiment, even studying these partial processes may help understand the course and mechanisms of a disease. Genetically modified mice are playing an increasingly important role in this type of research.

5. Treatment

Traditionally, unless contraindicated, HFrEF should be treated with β-blocker, angiotensin receptor–neprilysin inhibitor, angiotensin-converting enzyme inhibitor, or angiotensin receptor blocker, with the addition of a mineralocorticoid receptor antagonist in patients with prominent symptoms [\[80\]](#page-26-17), while Ivabradine and hydralazine/isosorbide dinitrate may also be considered in the management of HFrEF [\[249\]](#page-33-20). More recently, sodiumglucose cotransporter 2 inhibitors have much-improved disease outcomes, dramatically reducing cardiovascular and all-cause mortality regardless of diabetic state, and vericiguat, a stimulator of soluble guanylate cyclase, reduces inpatient admissions for high-risk HFrEF [\[80\]](#page-26-17). Device therapies may have benefits for specific subpopulations of HFrEF [\[80\]](#page-26-17). On the contrary, medication classes that are efficacious in HFrEF have been less so in HFpEF, decreasing the risk of inpatient admissions but not cardiovascular or all-cause mortality in HFpEF [\[249\]](#page-33-20). These observations underline the significance of non-cardiac comorbidities and underscore the complexity of pathophysiological mechanisms, both cardiac and non-cardiac, underpinning HFpEF [\[249\]](#page-33-20).

CASHF cannot be improved by interventional revascularization, and medications are the cornerstones of treatment. Although the interplay of epicardial and microvascular

CAS and associated risk factors are clinically relevant and represents a critical differentiator for what may constitute specific therapeutic strategies in CASHF, the CAS-induced abnormal regional wall motion, dilated left ventricular and reduced systolic function improved 6 months to >1 year by medications, including calcium channel blockers and nitrate/nicorandil [\[23,](#page-24-12)[72\]](#page-26-9). HFrEF, e.g., dilated cardiomyopathy in Syrian hamsters [\[220,](#page-32-14)[250\]](#page-33-21) and in German patients [\[251\]](#page-33-22), with CMD possibly caused by CAS, can be improved through vasodilator effect by the medical treatment with verapamil and diltiazem, respectively. Of note, while auxiliary diltiazem in suspected CAS-related dilated cardiomyopathy has mortality benefits, improved symptoms, and hemodynamics by reducing afterload, arrhythmias, and catecholamine levels [\[251\]](#page-33-22), diltiazem in individuals with infarction-related HFrEF has a dismal prognosis [\[252\]](#page-33-23). On the other hand, calcium channel blockers in non-ischemic HFrEF are not recommended as first-line therapy. Hence, although first-generation calcium channel blockers (except amlodipine and felodipine), dihydropyridine, and nondihydropyridine, should be limited in non-CAS-induced HFrEF because of no functional, mortality, or outcome benefits [\[253\]](#page-34-0), if HFrEF patients have provoked CAS, calcium channel blockers might improve myocardial ischemia due to CAS [\[23](#page-24-12)[,67\]](#page-26-6). Future research is needed to investigate the potential therapeutic role of calcium channel blockers in CASHFrEF. In contrast, the use of β-blockers in CASHFrEF may aggravate CAS [\[25\]](#page-24-13). Finally, although fasudil, a Rho-kinase inhibitor, prevents acetylcholine-induced CAS and associated myocardial ischemia [\[254\]](#page-34-1), its role in CASHF remains unknown. Additionally, patients with CAS-related dilated cardiomyopathy have a higher prevalence of atrial fibrillation than those without CAS [67% vs. 8% (*p* < 0.05)] [\[67\]](#page-26-6). Therefore, dilated cardiomyopathy with atrial fibrillation is probably an indication to identify CAS [\[67\]](#page-26-6). Taken together, although no guideline addresses the therapeutic significance of calcium channel blockers in CASHF [\[24\]](#page-24-25), the differential diagnosis of dilated cardiomyopathy or HFrEF should include CAS since calcium channel blockers are potentially promising medical options [\[24,](#page-24-25)[72\]](#page-26-9).

Notwithstanding established treatments for CAD, some patients suffer from refractory symptoms. The soluble guanylate cyclase stimulator riociguat, licensed for pulmonary hypertension treatment, has been reported to resolve recurrent and refractory CAS-induced angina [\[255\]](#page-34-2). The drug inhibited the acetylcholine provocation of epicardial CAS, and resulted in a remarkably satisfactory long-term (10 months) effect on perceived wellbeing [\[255\]](#page-34-2), suggesting that the soluble guanylate cyclase pathway is a potential novel therapeutic target in CAS. However, randomized controlled clinical trials are necessary to strengthen this presupposition.

In acute MI, although prompt coronary reperfusion is the most effective way to limit myocardial injury, the subsequent cardiomyocyte apoptosis, adverse left ventricular remodeling, and, finally, ischemic HF, the medical therapies of the subsequent tissue inflammation and its following suppression and resolution, remains largely unknown [\[171\]](#page-30-16). Among dietary phytochemicals that are naturally plant-derived and have been investigated to offer some protection against chronic diseases, garcinol demonstrates potential drug treatment effects in in vitro studies, such as its anti-inflammatory, anti-oxidative, and anti-cancer properties [\[256\]](#page-34-3). In rat models with isoproterenol-induced HFrEF, garcinol treatment increased the heart rate and improved the maximum rate of increase in pressure (+dp/dtmax), maximum rate of decrease in pressure (−dp/dtmax), ejection fraction, and systolic pressure in the left ventricle [\[257\]](#page-34-4). We have previously demonstrated that garcinol suppresses lipoprotein(a)-induced oxidative stress and inflammatory cytokines by α 7-nicotinic acetylcholine receptor-mediated inhibition of p38 MAPK/NF-κB signaling in cardiomyocyte AC16 cells and isoproterenol-induced acute MI mice [\[258\]](#page-34-5). These observations suggest that garcinol may effectively prevent cardiomyocyte apoptosis.

Although inflammation can be a cause–effect event of HF and, hence, a therapeutic target, clinical trials evaluating anti-inflammatory treatments failed to produce adequate relief; however, it is as yet uncertain what targeted anti-inflammatory therapy in distinct sub-phenotypes of HF such as CASHF will prove to be successful [\[259\]](#page-34-6). Potential targeted anti-inflammatory therapies include the inhibition of IL-1β, IL-6, and galectin-3 [\[259\]](#page-34-6). In

CANTOS (Canakinumab Anti-Inflammatory Thrombosis Outcomes Study) trial, subjects who were canakinumab-responsive (as reflected by a decrease in C-reactive protein) had a significant drop in HF hospitalizations and the composite of HF hospitalizations and all-cause mortality by 38% and 32%, respectively, compared with placebo [\[260\]](#page-34-7). Although anti-IL-6 therapies have been approved for rheumatologic and inflammatory disorders, including tocilizumab, siltuximab, and sarilumab, no clinical trial yet investigates the effects of IL-6 inhibitors in HF patients [\[259\]](#page-34-6). In experimental preclinical studies, pharmacologic inhibition of galectin-3 through the utilization of either modified citrus pectin or N-acetyllactosamine avoids myocardial and renal fibrosis and dysfunction [\[261,](#page-34-8)[262\]](#page-34-9), which may warrant further investigation in HF. Among anti-inflammatory agents, NSAID use has previously been linked to an increased risk of hypervolemia, blood pressure elevation [\[263\]](#page-34-10), and HF [\[264\]](#page-34-11). In large-scale clinical trials, anti-TNF- α agents did not prevent HF [\[265\]](#page-34-12). Taken together, to date, clinical trials of directed anticytokine and anti-inflammatory treatments for HF have proved mostly unsuccessful [\[259\]](#page-34-6), and the effects of these therapies have yet studied in CASHF. Of note, because cytokines can become cardioprotective in certain conditions, the timing (acute or chronic phase following MI) and intensity of the cell type-specific inhibition (leukocytes, cardiac fibroblasts or cardiomyocytes) must be taken into account in developing anti-inflammatory therapies [\[169\]](#page-30-14).

6. Conclusions

CAS is common, though it is still unsolved, and deserves the same fast action as CAD. Because CAS can cause rapid plaque progression of CAD and the development of acute coronary syndrome, including MI, underrecognized CAS is concerned with the health of the individuals and population as a whole, as well as with the health implications of the economic and social policies, and investment in health policies. While medical care can prolong survival and improve prognosis after the occurrence of CAD and HF, more important is to identify ill people afflicted with CAS before the potential subsequent development of CAD and HF in the first place. It has been demonstrated that after the HF condition is stabilized, the provocative testing for CAS can be safely performed. Treatment should be started early once CAS is diagnosed. On the other hand, while HF treatment aims to control the symptoms and slow down the progression, CASHF is one of a few conditions in that medical therapy may reverse HF. Furthermore, patients with CASHFrEF may have associated atrial fibrillation.

CAS has been a multifactorial disorder that cannot be attributed to a single factor alone (Figure [6\)](#page-22-0). In addition, because vascular smooth muscle cell hyperreactivity is a nonspecific reaction and CAS-induced angina is not improved by rest, CAS-related angina can occur under different situations in the same patient. As a consequence, identifying CAS is crucial in clinical practice because the therapeutic strategies between CAS and obstructive CAD are different, and calcium channel blockers are needed to improve the left ventricular function of CASHFrEF. Accordingly, it is of paramount importance to administer intracoronary nitroglycerin adequately before coronary interventions to distinguish spontaneous CAS from obstructive CAD, thus limiting vascular damage to all layers and preventing unnecessary interventions. Finally, we agree with the Japanese cardiologists to recommend upgrading the pharmacological CAS provocative testing to Class I in the guidelines in patients with angina but without obstructive CAD throughout the world.

Figure 6. Graphical abstract depicting the multifactorial molecular and cellular mechanisms involved in the initiation and progression of CAS and CAS-related preclinical HF to clinical overt HF. The development of CAS can be contributed to by smoking, CRP, and Lp(a). Fasudil, Tocilizumab, Garcinol, and Riociguat are potential disease-modifying therapies of CAS [\[196,](#page-31-15)[197,](#page-31-16)[201,](#page-31-20)[252\]](#page-33-23). The reversible nature of CASHF is suggested and represented by a reciprocal relationship and a positive feedback loop between epicardial and microvascular CAS. Solid arrows: direct activating interactions; Dashed arrow: indirect activating interactions; Blunt arrows: inhibition. Ach: acetylcholine; α7-nAChR: α7-nicotinic acetylcholine receptor; CHRNA7: α7-nAChR protein coding gene; CamKII: calmodulin-dependent kinase II; CAS: coronary artery spasm; CRP: C-reactive protein; HFpEF: heart failure with reduced ejection fraction; HFrEF: heart failure with reduced ejection fraction; ICAM-1: intercellular adhesion molecule 1; IL-6: Interleukin-6; INOCA: ischemia with non-obstructive coronary artery disease; Lp(a): lipoprotein(a); MINOCA: myocardial infarction with non-obstructive coronary artery disease; p38MAPK: p38 mitogen-activated protein kinase; VCAM-1: vascular cell adhesion molecule 1.

Avoiding cigarette smoking and alcohol with concomitant appropriate dosing and timing of calcium antagonists remain the mainstay of CAS therapy. Besides, instead of treating a specific physical condition, we should focus on the whole person's health. Anxiety and depression confer high risks for CAS-related myocardial ischemia. In Taiwanese patients, anxiety is associated with a remarkably 5-fold increased risk of incident CAS [\[92\]](#page-27-3), suggesting that simple assessment tools can be used for patients at risk for CAS to evaluate mental health well-being, and treatments of psychological disorders can have a beneficial impact on CAS [\[266\]](#page-34-13). Because recurrent angina events are commonly observed in CAS, further investigation is needed and important to help better clarify the responsible molecular mechanisms and manage CAS more effectively.

Author Contributions: Conceptualization and writing—original draft preparation, M.-J.H. and M.-Y.H.; writing—review and editing, C.-T.Y., N.G.K., I.K. and P.H. All authors have read and agreed to the published version of the manuscript.

Funding: Funding was provided by the National Science Council of Taiwan grant to M.-Y.H. (MOST 111-2314-B-038-028), and by the grant from Taipei Medical University (111TMU-SHH-01) to M.-Y.H.

Institutional Review Board Statement: The cases demonstrating epicardial CAS-induced ventricular ectopics and microvascular CAS-induced ventricular fibrillation were approved by the Taipei Medical University Joint Institutional Review Board (approval number: TMU-JIRB 201111004 version 1.3).

Informed Consent Statement: Not applicable.

Data Availability Statement: Not applicable.

Conflicts of Interest: The authors declare no conflict of interest.

Abbreviations

ALDH2: Aldehyde Dehydrogenase 2; CAD: coronary artery disease;CAS: coronary artery spasm; CASHF: CAS-related heart failure; CASHFrEF: CAS-related heart failure with reduced ejection fraction CMD: coronary microvascular disease; CRP: C-reactive protein; HF: heart failure; HFpEF: heart failure with reduced ejection fraction; HFrEF: heart failure with reduced ejection fraction; hiPSC-CMs: induced pluripotent stem cells; IL: Interleukin; INOCA: ischemia with non-obstructive coronary artery disease; LVEF: left ventricular ejection fraction; MI: myocardial infarction; MINOCA: myocardial infarction with non-obstructive coronary artery disease; SMA: α-smooth muscle actin; SMP30: senescence marker protein-30; VF: ventricular fibrillation.

References

- 1. Jessup, M.; Brozena, S. Heart failure. *N. Engl. J. Med.* **2003**, *348*, 2007–2018. [\[CrossRef\]](https://doi.org/10.1056/NEJMra021498)
- 2. Dunlay, S.M.; Roger, V.L.; Weston, S.A.; Jiang, R.; Redfield, M.M. Longitudinal changes in ejection fraction in heart failure patients with preserved and reduced ejection fraction. *Circ. Heart Fail.* **2012**, *5*, 720–726. [\[CrossRef\]](https://doi.org/10.1161/CIRCHEARTFAILURE.111.966366) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/22936826)
- 3. Borlaug, B.A.; Redfield, M.M. Diastolic and systolic heart failure are distinct phenotypes within the heart failure spectrum. *Circulation* **2011**, *123*, 2006–2013. [\[CrossRef\]](https://doi.org/10.1161/CIRCULATIONAHA.110.954388) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/21555723)
- 4. Paulus, W.J.; Tschöpe, C. A novel paradigm for heart failure with preserved ejection fraction: Comorbidities drive myocardial dysfunction and remodeling through coronary microvascular endothelial inflammation. *J. Am. Coll. Cardiol.* **2013**, *62*, 263–271. [\[CrossRef\]](https://doi.org/10.1016/j.jacc.2013.02.092) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/23684677)
- 5. McMurray, J.J. Clinical practice. Systolic heart failure. *N. Engl. J. Med.* **2010**, *362*, 228–238. [\[CrossRef\]](https://doi.org/10.1056/NEJMcp0909392) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/20089973)
- 6. Brucks, S.; Little, W.C.; Chao, T.; Kitzman, D.W.; Wesley-Farrington, D.; Gandhi, S.; Shihabi, Z.K. Contribution of left ventricular diastolic dysfunction to heart failure regardless of ejection fraction. *Am. J. Cardiol.* **2005**, *95*, 603–606. [\[CrossRef\]](https://doi.org/10.1016/j.amjcard.2004.11.006)
- 7. Lam, C.S.; Lyass, A.; Kraigher-Krainer, E.; Massaro, J.M.; Lee, D.S.; Ho, J.E.; Levy, D.; Redfield, M.M.; Pieske, B.M.; Benjamin, E.J.; et al. Cardiac dysfunction and noncardiac dysfunction as precursors of heart failure with reduced and preserved ejection fraction in the community. *Circulation* **2011**, *124*, 24–30. [\[CrossRef\]](https://doi.org/10.1161/CIRCULATIONAHA.110.979203)
- 8. Eisman, A.S.; Shah, R.V.; Dhakal, B.P.; Pappagianopoulos, P.P.; Wooster, L.; Bailey, C.; Cunningham, T.F.; Hardin, K.M.; Baggish, A.L.; Ho, J.E.; et al. Pulmonary Capillary Wedge Pressure Patterns During Exercise Predict Exercise Capacity and Incident Heart Failure. *Circ. Heart Fail.* **2018**, *11*, e004750. [\[CrossRef\]](https://doi.org/10.1161/CIRCHEARTFAILURE.117.004750)
- 9. Greenberg, B.; Kim, P.J.; Kahn, A.M. Clinical evaluation of heart failure. In *Heart Failure: A Companion to Braunwald's Heart Disease*, 4th ed.; Felker, G.M., Mann, D.L., Eds.; Elsevier: Philadelphia, PA, USA, 2020; Chapter 31.
- 10. Shah, K.S.; Xu, H.; Matsouaka, R.A.; Bhatt, D.L.; Heidenreich, P.A.; Hernandez, A.F.; Devore, A.D.; Yancy, C.W.; Fonarow, G.C. Heart Failure with Preserved, Borderline, and Reduced Ejection Fraction: 5-Year Outcomes. *J. Am. Coll. Cardiol.* **2017**, *70*, 2476–2486. [\[CrossRef\]](https://doi.org/10.1016/j.jacc.2017.08.074)
- 11. Virani, S.S.; Alonso, A.; Benjamin, E.J.; Bittencourt, M.S.; Callaway, C.W.; Carson, A.P.; Chamberlain, A.M.; Chang, A.R.; Cheng, S.; Delling, F.N.; et al. Heart Disease and Stroke Statistics-2020 Update: A Report from the American Heart Association. *Circulation* **2020**, *141*, e139–e596. [\[CrossRef\]](https://doi.org/10.1161/CIR.0000000000000757)
- 12. Dunlay, S.M.; Roger, V.L.; Redfield, M.M. Epidemiology of heart failure with preserved ejection fraction. *Nat. Rev. Cardiol.* **2017**, *14*, 591–602. [\[CrossRef\]](https://doi.org/10.1038/nrcardio.2017.65)
- 13. Borlaug, B.A. Evaluation and management of heart failure with preserved ejection fraction. *Nat. Rev. Cardiol.* **2020**, *17*, 559–573. [\[CrossRef\]](https://doi.org/10.1038/s41569-020-0363-2)
- 14. Obokata, M.; Kane, G.C.; Reddy, Y.N.; Olson, T.P.; Melenovsky, V.; Borlaug, B.A. Role of Diastolic Stress Testing in the Evaluation for Heart Failure With Preserved Ejection Fraction: A Simultaneous Invasive-Echocardiographic Study. *Circulation* **2017**, *135*, 825–838. [\[CrossRef\]](https://doi.org/10.1161/CIRCULATIONAHA.116.024822)
- 15. Shah, S.J.; Katz, D.H.; Deo, R.C. Phenotypic spectrum of heart failure with preserved ejection fraction. *Heart Fail. Clin.* **2014**, *10*, 407–418. [\[CrossRef\]](https://doi.org/10.1016/j.hfc.2014.04.008) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/24975905)
- 16. Shah, S.J.; Kitzman, D.W.; Borlaug, B.A.; van Heerebeek, L.; Zile, M.R.; Kass, D.A.; Paulus, W.J. Phenotype-Specific Treatment of Heart Failure With Preserved Ejection Fraction: A Multiorgan Roadmap. *Circulation* **2016**, *134*, 73–90. [\[CrossRef\]](https://doi.org/10.1161/CIRCULATIONAHA.116.021884)
- 17. Senni, M.; Redfield, M.M. Heart failure with preserved systolic function. A different natural history? *J. Am. Coll. Cardiol.* **2001**, *38*, 1277–1282. [\[CrossRef\]](https://doi.org/10.1016/S0735-1097(01)01567-4)
- 18. Kannel, W.B. Lessons from curbing the coronary artery disease epidemic for confronting the impending epidemic of heart failure. *Med. Clin. N. Am.* **2004**, *88*, 1129–1133. [\[CrossRef\]](https://doi.org/10.1016/j.mcna.2004.02.002)
- 19. Vancheri, F.; Longo, G.; Vancheri, S.; Henein, M. Coronary Microvascular Dysfunction. *J. Clin. Med.* **2020**, *9*, 2880. [\[CrossRef\]](https://doi.org/10.3390/jcm9092880) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/32899944)
- 20. Cowie, M.R.; Wood, D.A.; Coats, A.J.; Thompson, S.G.; Poole-Wilson, P.A.; Suresh, V.; Sutton, G.C. Incidence and aetiology of heart failure; a population-based study. *Eur. Heart J.* **1999**, *20*, 421–428. [\[CrossRef\]](https://doi.org/10.1053/euhj.1998.1280)
- 21. Repetto, A.; Dal Bello, B.; Pasotti, M.; Agozzino, M.; Viganò, M.; Klersy, C.; Tavazzi, L.; Arbustini, E. Coronary atherosclerosis in end-stage idiopathic dilated cardiomyopathy: An innocent bystander? *Eur. Heart J.* **2005**, *26*, 1519–1527. [\[CrossRef\]](https://doi.org/10.1093/eurheartj/ehi342) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/15917275)
- 22. Felker, G.M.; Thompson, R.E.; Hare, J.M.; Hruban, R.H.; Clemetson, D.E.; Howard, D.L.; Baughman, K.L.; Kasper, E.K. Underlying causes and long-term survival in patients with initially unexplained cardiomyopathy. *N. Engl. J. Med.* **2000**, *342*, 1077–1084. [\[CrossRef\]](https://doi.org/10.1056/NEJM200004133421502)
- 23. Sueda, S.; Kohno, H.; Oshita, A.; Izoe, Y.; Nomoto, T.; Fukuda, H. Vasospastic heart failure: Multiple spasm may cause transient heart failure? *J. Cardiol.* **2009**, *54*, 452–459. [\[CrossRef\]](https://doi.org/10.1016/j.jjcc.2009.07.007)
- 24. Inami, T.; Kataoka, M.; Shimura, N.; Ishiguro, H.; Kohshoh, H.; Taguchi, H.; Yanagisawa, R.; Hara, Y.; Satoh, T.; Yoshino, H. Left ventricular dysfunction due to diffuse multiple vessel coronary artery spasm can be concealed in dilated cardiomyopathy. *Eur. J. Heart Fail.* **2012**, *14*, 1130–1138. [\[CrossRef\]](https://doi.org/10.1093/eurjhf/hfs103) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/22713288)
- 25. Oda, S.; Fujii, Y.; Takemoto, H.; Nomura, S.; Nakayama, H.; Toyota, Y.; Nakamura, H.; Teragawa, H. Heart failure in which coronary spasms played an important role. *Intern. Med.* **2014**, *53*, 227–232. [\[CrossRef\]](https://doi.org/10.2169/internalmedicine.53.1217) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/24492691)
- 26. MacAlpin, R.N.; Kattus, A.A.; Alvaro, A.B. Angina pectoris at rest with preservation of exercise capacity: Prinzmetal's variant angina. *Circulation* **1973**, *47*, 946–958. [\[CrossRef\]](https://doi.org/10.1161/01.CIR.47.5.946)
- 27. Prinzmetal, M.; Kennamer, R.; Merliss, R.; Wada, T.; Bor, N. Angina pectoris. I. A variant form of angina pectoris; preliminary report. *Am. J. Med.* **1959**, *27*, 375–388. [\[CrossRef\]](https://doi.org/10.1016/0002-9343(59)90003-8) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/14434946)
- 28. Blumgart, H.L.; Schlesinger, M.J.; Davis, D. Studies on the relation of the clinical manifestations of angina pectoris, coronary thrombosis and myocardial infarction to the pathologic findings. *Am. Heart J.* **1940**, *19*, 1–91. [\[CrossRef\]](https://doi.org/10.1016/S0002-8703(40)90305-2)
- 29. Oliva, P.B.; Potts, D.E.; Pluss, R.G. Coronary arterial spasm in Prinzmetal angina. Documentation by coronary arteriography. *N. Engl. J. Med.* **1973**, *288*, 745–751. [\[CrossRef\]](https://doi.org/10.1056/NEJM197304122881501)
- 30. Cheng, T.O.; Bashour, T.; Kelser, G.A., Jr.; Weiss, L.; Bacos, J. Variant angina of prinzmetal with normal coronary arteriograms. A variant of the variant. *Circulation* **1973**, *47*, 476–485. [\[CrossRef\]](https://doi.org/10.1161/01.CIR.47.3.476)
- 31. Maseri, A.; Mimmo, R.; Chierchia, S.; Marchesi, C.; Pesola, A.; L'Abbate, A. Coronary artery spasm as a cause of acute myocardial ischemia in man. *Chest* **1975**, *68*, 625–633. [\[CrossRef\]](https://doi.org/10.1378/chest.68.5.625)
- 32. Cheng, C.W.; Yang, N.I.; Lin, K.J.; Hung, M.J.; Cherng, W.J. Role of coronary spasm for a positive noninvasive stress test result in angina pectoris patients without hemodynamically significant coronary artery disease. *Am. J. Med. Sci.* **2008**, *335*, 354–362. [\[CrossRef\]](https://doi.org/10.1097/MAJ.0b013e31815681b2)
- 33. Hung, M.J.; Hu, P.; Hung, M.Y. Coronary artery spasm: Review and update. *Int. J. Med. Sci.* **2014**, *11*, 1161–1171. [\[CrossRef\]](https://doi.org/10.7150/ijms.9623) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/25249785)
- 34. Kounis, N.G.; Zavras, G.M. Histamine-induced coronary artery spasm: The concept of allergic angina. *Br. J. Clin. Pract.* **1991**, *45*, 121–128. [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/1793697)
- 35. Kounis, N.G.; Koniari, I.; Velissaris, D.; Tzanis, G.; Hahalis, G. Kounis Syndrome—Not a Single-organ Arterial Disorder but a Multisystem and Multidisciplinary Disease. *Balkan Med. J.* **2019**, *36*, 212–221. [\[CrossRef\]](https://doi.org/10.4274/balkanmedj.galenos.2019.2019.5.62)
- 36. Maseri, A. Pathogenetic mechanisms of angina pectoris: Expanding views. *Br. Heart J.* **1980**, *43*, 648–660. [\[CrossRef\]](https://doi.org/10.1136/hrt.43.6.648)
- 37. Chahine, R.A.; Luchi, R.J. Coronary arterial spasm: Culprit or bystander? *Am. J. Cardiol.* **1976**, *37*, 936–937. [\[CrossRef\]](https://doi.org/10.1016/0002-9149(76)90123-5)
- 38. Nasiri-Partovi, A.; Shafiee, A.; Rahmani, R. Intracoronary injection of nitroglycerine can prevent unnecessary percutaneous coronary intervention. *BMC Cardiovasc. Disord.* **2022**, *22*, 416. [\[CrossRef\]](https://doi.org/10.1186/s12872-022-02823-2)
- 39. O'Connor, C.M.; Velazquez, E.J.; Gardner, L.H.; Smith, P.K.; Newman, M.F.; Landolfo, K.P.; Lee, K.L.; Califf, R.M.; Jones, R.H. Comparison of coronary artery bypass grafting versus medical therapy on long-term outcome in patients with ischemic cardiomyopathy (a 25-year experience from the Duke Cardiovascular Disease Databank). *Am. J. Cardiol.* **2002**, *90*, 101–107. [\[CrossRef\]](https://doi.org/10.1016/S0002-9149(02)02429-3) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/12106836)
- 40. Alderman, E.L.; Fisher, L.D.; Litwin, P.; Kaiser, G.C.; Myers, W.O.; Maynard, C.; Levine, F.; Schloss, M. Results of coronary artery surgery in patients with poor left ventricular function (CASS). *Circulation* **1983**, *68*, 785–795. [\[CrossRef\]](https://doi.org/10.1161/01.CIR.68.4.785)
- 41. Baker, D.W.; Jones, R.; Hodges, J.; Massie, B.M.; Konstam, M.A.; Rose, E.A. Management of heart failure. III. The role of revascularization in the treatment of patients with moderate or severe left ventricular systolic dysfunction. *JAMA* **1994**, *272*, 1528–1534. [\[CrossRef\]](https://doi.org/10.1001/jama.1994.03520190074038)
- 42. Pigott, J.D.; Kouchoukos, N.T.; Oberman, A.; Cutter, G.R. Late results of surgical and medical therapy for patients with coronary artery disease and depressed left ventricular function. *J. Am. Coll. Cardiol.* **1985**, *5*, 1036–1045. [\[CrossRef\]](https://doi.org/10.1016/S0735-1097(85)80003-6)
- 43. Elefteriades, J.A.; Tolis, G., Jr.; Levi, E.; Mills, L.K.; Zaret, B.L. Coronary artery bypass grafting in severe left ventricular dysfunction: Excellent survival with improved ejection fraction and functional state. *J. Am. Coll. Cardiol.* **1993**, *22*, 1411–1417. [\[CrossRef\]](https://doi.org/10.1016/0735-1097(93)90551-B) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/8227799)
- 44. Lloyd-Jones, D.M.; Larson, M.G.; Leip, E.P.; Beiser, A.; D'Agostino, R.B.; Kannel, W.B.; Murabito, J.M.; Vasan, R.S.; Benjamin, E.J.; Levy, D. Lifetime risk for developing congestive heart failure: The Framingham Heart Study. *Circulation* **2002**, *106*, 3068–3072. [\[CrossRef\]](https://doi.org/10.1161/01.CIR.0000039105.49749.6F) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/12473553)
- 45. Levy, D.; Kenchaiah, S.; Larson, M.G.; Benjamin, E.J.; Kupka, M.J.; Ho, K.K.; Murabito, J.M.; Vasan, R.S. Long-term trends in the incidence of and survival with heart failure. *N. Engl. J. Med.* **2002**, *347*, 1397–1402. [\[CrossRef\]](https://doi.org/10.1056/NEJMoa020265)
- 46. Kenchaiah, S.; Vasan, R.S. Heart Failure in Women–Insights from the Framingham Heart Study. *Cardiovasc. Drugs Ther.* **2015**, *29*, 377–390. [\[CrossRef\]](https://doi.org/10.1007/s10557-015-6599-0)
- 47. Ho, J.E.; Lyass, A.; Lee, D.S.; Vasan, R.S.; Kannel, W.B.; Larson, M.G.; Levy, D. Predictors of new-onset heart failure: Differences in preserved versus reduced ejection fraction. *Circ. Heart Fail.* **2013**, *6*, 279–286. [\[CrossRef\]](https://doi.org/10.1161/CIRCHEARTFAILURE.112.972828)
- 48. Piña, I.L.; Kokkinos, P.; Kao, A.; Bittner, V.; Saval, M.; Clare, B.; Goldberg, L.; Johnson, M.; Swank, A.; Ventura, H.; et al. Baseline differences in the HF-ACTION trial by sex. *Am. Heart J.* **2009**, *158* (Suppl. 4), S16–S23. [\[CrossRef\]](https://doi.org/10.1016/j.ahj.2009.07.012)
- 49. Taylor, C.J.; Ordóñez-Mena, J.M.; Roalfe, A.K.; Lay-Flurrie, S.; Jones, N.R.; Marshall, T.; Hobbs, F.D.R. Trends in survival after a diagnosis of heart failure in the United Kingdom 2000–2017: Population based cohort study. *BMJ* **2019**, *364*, l223. [\[CrossRef\]](https://doi.org/10.1136/bmj.l223)
- 50. Beltrame, J.F.; Sasayama, S.; Maseri, A. Racial heterogeneity in coronary artery vasomotor reactivity: Differences between Japanese and Caucasian patients. *J. Am. Coll. Cardiol.* **1999**, *33*, 1442–1452. [\[CrossRef\]](https://doi.org/10.1016/S0735-1097(99)00073-X) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/10334407)
- 51. JCS Joint Working Group. Guidelines for diagnosis and treatment of patients with vasospastic angina (Coronary Spastic Angina) (JCS 2013). *Circ. J.* **2014**, *78*, 2779–2801. [\[CrossRef\]](https://doi.org/10.1253/circj.CJ-66-0098) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/25273915)
- 52. Sueda, S.; Kohno, H.; Fukuda, H.; Ochi, N.; Kawada, H.; Hayashi, Y.; Uraoka, T. Frequency of provoked coronary spasms in patients undergoing coronary arteriography using a spasm provocation test via intracoronary administration of ergonovine. *Angiology* **2004**, *55*, 403–411. [\[CrossRef\]](https://doi.org/10.1177/000331970405500407)
- 53. Hung, M.Y.; Hsu, K.H.; Hung, M.J.; Cheng, C.W.; Cherng, W.J. Interactions among gender, age, hypertension and C-reactive protein in coronary vasospasm. *Eur. J. Clin. Investig.* **2010**, *40*, 1094–1103. [\[CrossRef\]](https://doi.org/10.1111/j.1365-2362.2010.02360.x)
- 54. Bertrand, M.E.; LaBlanche, J.M.; Tilmant, P.Y.; Thieuleux, F.A.; Delforge, M.R.; Carre, A.G.; Asseman, P.; Berzin, B.; Libersa, C.; Laurent, J.M. Frequency of provoked coronary arterial spasm in 1089 consecutive patients undergoing coronary arteriography. *Circulation* **1982**, *65*, 1299–1306. [\[CrossRef\]](https://doi.org/10.1161/01.CIR.65.7.1299) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/7200405)
- 55. Hung, M.J.; Cherng, W.J.; Cheng, C.W.; Li, L.F. Comparison of serum levels of inflammatory markers in patients with coronary vasospasm without significant fixed coronary artery disease versus patients with stable angina pectoris and acute coronary syndromes with significant fixed coronary artery disease. *Am. J. Cardiol.* **2006**, *97*, 1429–1434. [\[CrossRef\]](https://doi.org/10.1016/j.amjcard.2005.12.035) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/16679078)
- 56. Hung, M.J.; Cheng, C.W.; Yang, N.I.; Hung, M.Y.; Cherng, W.J. Coronary vasospasm-induced acute coronary syndrome complicated by life-threatening cardiac arrhythmias in patients without hemodynamically significant coronary artery disease. *Int. J. Cardiol.* **2007**, *117*, 37–44. [\[CrossRef\]](https://doi.org/10.1016/j.ijcard.2006.03.055) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/16844245)
- 57. Ong, P.; Athanasiadis, A.; Hill, S.; Vogelsberg, H.; Voehringer, M.; Sechtem, U. Coronary artery spasm as a frequent cause of acute coronary syndrome: The CASPAR (Coronary Artery Spasm in Patients with Acute Coronary Syndrome) Study. *J. Am. Coll. Cardiol.* **2008**, *52*, 523–527. [\[CrossRef\]](https://doi.org/10.1016/j.jacc.2008.04.050)
- 58. Goto, A.; Ito, S.; Kondo, H.; Nomura, Y.; Yasue, N.; Suzumura, H.; Takeda, Y.; Tomimoto, S.; Yamada, Y.; Horio, T.; et al. Evaluation of adjunctive intracoronary administration of acetylcholine following intravenous infusion of ergonovine to provoke coronary artery spasm. *J. Cardiol.* **1999**, *34*, 309–316. [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/10642927)
- 59. Yasue, H.; Mizuno, Y.; Harada, E. Coronary artery spasm—Clinical features, pathogenesis and treatment. *Proc. Jpn. Acad. Ser. B* **2019**, *95*, 53–66. [\[CrossRef\]](https://doi.org/10.2183/pjab.95.005) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/30745502)
- 60. Sueda, S.; Sakaue, T. Racial Differences in Patients with Coronary Vasomotion Disorders. *J. Coron. Artery Dis.* **2021**, *27*, 7–17. [\[CrossRef\]](https://doi.org/10.7793/jcad.27.002)
- 61. Da, C.A.; Isaaz, K.; Faure, E.; Mourot, S.; Cerisier, A.; Lamaud, M. Clinical characteristics, aetiological factors and long-term prognosis of myocardial infarction with an absolutely normal coronary angiogram; a 3-year follow-up study of 91 patients. *Eur. Heart J.* **2001**, *22*, 1459–1465.
- 62. Sueda, S.; Kohno, H. The acetylcholine administration time plays the key role for provoked spasm in the spasm provocation test. *J. Cardiol.* **2017**, *70*, 141–146. [\[CrossRef\]](https://doi.org/10.1016/j.jjcc.2016.11.003) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/27988074)
- 63. Hung, M.Y.; Hung, M.J.; Cheng, C.W.; Yang, N.I.; Cherng, W.J. Safety and predictors of a positive result of intracoronary ergonovine testing in patients with ischemic heart disease without hemodynamically significant coronary artery stenosis in Taiwan. *Acta Cardiol. Sin.* **2007**, *23*, 150–159.
- 64. Takagi, Y.; Yasuda, S.; Takahashi, J.; Tsunoda, R.; Ogata, Y.; Seki, A.; Sumiyoshi, T.; Matsui, M.; Goto, T.; Tanabe, Y.; et al. Clinical implications of provocation tests for coronary artery spasm: Safety, arrhythmic complications, and prognostic impact: Multicentre registry study of the Japanese Coronary Spasm Association. *Eur. Heart J.* **2013**, *34*, 258–267. [\[CrossRef\]](https://doi.org/10.1093/eurheartj/ehs199) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/22782943)
- 65. Scanlon, P.J.; Faxon, D.P.; Audet, A.M.; Carabello, B.; Dehmer, G.J.; Eagle, K.A.; Legako, R.D.; Leon, D.F.; Murray, J.A.; Nissen, S.E.; et al. ACC/AHA guidelines for coronary angiography. A report of the American College of Cardiology/American Heart Association Task Force on practice guidelines (Committee on Coronary Angiography). Developed in collaboration with the Society for Cardiac Angiography and Interventions. *J. Am. Coll. Cardiol.* **1999**, *33*, 1756–1824.
- 66. Hung, M.Y.; Kounis, N.G.; Lu, M.Y.; Hu, P. Myocardial Ischemic Syndromes, Heart Failure Syndromes, Electrocardiographic Abnormalities, Arrhythmic Syndromes and Angiographic Diagnosis of Coronary Artery Spasm: Literature Review. *Int. J. Med. Sci.* **2020**, *17*, 1071–1082. [\[CrossRef\]](https://doi.org/10.7150/ijms.43472)
- 67. Baim, D.S. Coronary angiography. In *Grossman's Cardiac Catheterization, Angiography, and Intervention*, 7th ed.; Baim, D.S., Ed.; Lippincott Williams & Wilkins: Philadelphia, PA, USA, 2006.
- 68. Nishi, I.; Ilda, K.; Kawano, S.; Masumi, T.; Fumikura, Y.; Ohtsuka, S.; Watanabe, S.; Yamaguchi, I. Effects of anti-vasospastic agents in Japanese patients with dilated cardiomyopathy and coronary vasospasm. *Jpn. Heart J.* **2002**, *43*, 333–342. [\[CrossRef\]](https://doi.org/10.1536/jhj.43.333) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/12227709)
- 69. Heupler, F.A., Jr. Provocative testing for coronary arterial spasm: Risk, method and rationale. *Am. J. Cardiol.* **1980**, *46*, 335–337. [\[CrossRef\]](https://doi.org/10.1016/0002-9149(80)90081-8) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/7405847)
- 70. Bertrand, M.E.; Lablanche, J.M.; Tilmant, P.Y.; Thieuleux, F.A.; Delforge, M.G.; Chahine, R.A. The provocation of coronary arterial spasm in patients with recent transmural myocardial infarction. *Eur. Heart J.* **1983**, *4*, 532–535. [\[CrossRef\]](https://doi.org/10.1093/oxfordjournals.eurheartj.a061518) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/6641747)
- 71. Montalescot, G.; Sechtem, U.; Achenbach, S.; Andreotti, F.; Arden, C.; Budaj, A.; Bugiardini, R.; Crea, F.; Cuisset, T.; Di Mario, C.; et al. 2013 ESC guidelines on the management of stable coronary artery disease: The Task Force on the management of stable coronary artery disease of the European Society of Cardiology. *Eur. Heart J.* **2013**, *34*, 2949–3003.
- 72. Beltrame, J.F.; Crea, F.; Kaski, J.C.; Ogawa, H.; Ong, P.; Sechtem, U.; Shimokawa, H.; Bairey Merz, C.N.; Coronary Vasomotion Disorders International Study Group (COVADIS). International standardization of diagnostic criteria for vasospastic angina. *Eur. Heart J.* **2017**, *38*, 2565–2568. [\[CrossRef\]](https://doi.org/10.1093/eurheartj/ehv351)
- 73. Sakata, K.; Nawada, R.; Ohbayashi, K.; Tamekiyo, H.; Yoshida, H. Diffuse and severe left ventricular dysfunction induced by epicardial coronary artery spasm. *Angiology* **2000**, *51*, 837–847. [\[CrossRef\]](https://doi.org/10.1177/000331970005101006) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/11108328)
- 74. Fujioka, K.; Gordon, S. Effects of "Essential AD2" Supplement on Blood Acetaldehyde Levels in Individuals Who Have Aldehyde Dehydrogenase (ALDH2) Deficiency. *Am. J. Ther.* **2019**, *26*, 583–588. [\[CrossRef\]](https://doi.org/10.1097/MJT.0000000000000744) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/29509552)
- 75. Xu, F.; Chen, Y.G.; Xue, L.; Li, R.J.; Zhang, H.; Bian, Y.; Zhang, C.; Lv, R.J.; Feng, J.B.; Zhang, Y. Role of aldehyde dehydrogenase 2 Glu504lys polymorphism in acute coronary syndrome. *J. Cell. Mol. Med.* **2011**, *15*, 1955–1962. [\[CrossRef\]](https://doi.org/10.1111/j.1582-4934.2010.01181.x)
- 76. Mozaffarian, D.; Benjamin, E.J.; Go, A.S.; Arnett, D.K.; Blaha, M.J.; Cushman, M.; de Ferranti, S.; Després, J.P.; Fullerton, H.J.; Howard, V.J.; et al. Heart disease and stroke statistics—2015 update: A report from the American Heart Association. *Circulation* **2015**, *131*, e29–e322. [\[CrossRef\]](https://doi.org/10.1161/CIR.0000000000000152) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/25520374)
- 77. Gerber, Y.; Weston, S.A.; Redfield, M.M.; Chamberlain, A.M.; Manemann, S.M.; Jiang, R.; Killian, J.M.; Roger, V.L. A contemporary appraisal of the heart failure epidemic in Olmsted County, Minnesota, 2000 to 2010. *JAMA Intern. Med.* **2015**, *175*, 996–1004. [\[CrossRef\]](https://doi.org/10.1001/jamainternmed.2015.0924)
- 78. Conrad, N.; Judge, A.; Tran, J.; Mohseni, H.; Hedgecott, D.; Crespillo, A.P.; Allison, M.; Hemingway, H.; Cleland, J.G.; McMurray, J.J.V.; et al. Temporal trends and patterns in heart failure incidence: A population-based study of 4 million individuals. *Lancet* **2018**, *391*, 572–580. [\[CrossRef\]](https://doi.org/10.1016/S0140-6736(17)32520-5)
- 79. Maggioni, A.P.; Dahlström, U.; Filippatos, G.; Chioncel, O.; Crespo Leiro, M.; Drozdz, J.; Fruhwald, F.; Gullestad, L.; Logeart, D.; Fabbri, G.; et al. EURObservational Research Programme: Regional differences and 1-year follow-up results of the Heart Failure Pilot Survey (ESC-HF Pilot). *Eur. J. Heart Fail.* **2013**, *15*, 808–817. [\[CrossRef\]](https://doi.org/10.1093/eurjhf/hft050)
- 80. Murphy, S.P.; Ibrahim, N.E.; Januzzi, J.L. Heart Failure with Reduced Ejection Fraction: A Review. *JAMA* **2020**, *324*, 488–504. [\[CrossRef\]](https://doi.org/10.1001/jama.2020.10262) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/32749493)
- 81. Mohri, M.; Takeshita, A. Coronary microvascular disease in humans. *Jpn. Heart J.* **1999**, *40*, 97–108. [\[CrossRef\]](https://doi.org/10.1536/jhj.40.97)
- 82. Kunadian, V.; Chieffo, A.; Camici, P.G.; Berry, C.; Escaned, J.; Maas, A.H.E.M.; Prescott, E.; Karam, N.; Appelman, Y.; Fraccaro, C.; et al. An EAPCI Expert Consensus Document on Ischaemia with Non-Obstructive Coronary Arteries in Collaboration with European Society of Cardiology Working Group on Coronary Pathophysiology & Microcirculation Endorsed by Coronary Vasomotor Disorders International Study Group. *Eur. Heart J.* **2020**, *41*, 3504–3520. [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/32626906)
- 83. Bairey Merz, C.N.; Pepine, C.J.; Walsh, M.N.; Fleg, J.L. Ischemia and No Obstructive Coronary Artery Disease (INOCA): Developing Evidence-Based Therapies and Research Agenda for the Next Decade. *Circulation* **2017**, *135*, 1075–1092. [\[CrossRef\]](https://doi.org/10.1161/CIRCULATIONAHA.116.024534)
- 84. Jespersen, L.; Hvelplund, A.; Abildstrøm, S.Z.; Pedersen, F.; Galatius, S.; Madsen, J.K.; Jørgensen, E.; Kelbæk, H.; Prescott, E. Stable angina pectoris with no obstructive coronary artery disease is associated with increased risks of major adverse cardiovascular events. *Eur. Heart J.* **2012**, *33*, 734–744. [\[CrossRef\]](https://doi.org/10.1093/eurheartj/ehr331)
- 85. Maddox, T.M.; Stanislawski, M.A.; Grunwald, G.K.; Bradley, S.M.; Ho, P.M.; Tsai, T.T.; Patel, M.R.; Sandhu, A.; Valle, J.; Magid, D.J.; et al. Nonobstructive coronary artery disease and risk of myocardial infarction. *JAMA* **2014**, *312*, 1754–1763. [\[CrossRef\]](https://doi.org/10.1001/jama.2014.14681)
- 86. Petersen, J.W.; Johnson, B.D.; Kip, K.E.; Anderson, R.D.; Handberg, E.M.; Sharaf, B.; Mehta, P.K.; Kelsey, S.F.; Merz, C.N.; Pepine, C.J. TIMI frame count and adverse events in women with no obstructive coronary disease: A pilot study from the NHLBI-sponsored Women's Ischemia Syndrome Evaluation (WISE). *PLoS ONE* **2014**, *9*, e96630. [\[CrossRef\]](https://doi.org/10.1371/journal.pone.0096630)
- 87. Gulati, M.; Cooper-DeHoff, R.M.; McClure, C.; Johnson, B.D.; Shaw, L.J.; Handberg, E.M.; Zineh, I.; Kelsey, S.F.; Arnsdorf, M.F.; Black, H.R.; et al. Adverse cardiovascular outcomes in women with nonobstructive coronary artery disease: A report from the Women's Ischemia Syndrome Evaluation Study and the St James Women Take Heart Project. *Arch. Intern. Med.* **2009**, *169*, 843–850. [\[CrossRef\]](https://doi.org/10.1001/archinternmed.2009.50)
- 88. Brainin, P.; Frestad, D.; Prescott, E. The prognostic value of coronary endothelial and microvascular dysfunction in subjects with normal or non-obstructive coronary artery disease: A systematic review and meta-analysis. *Int. J. Cardiol.* **2018**, *254*, 1–9. [\[CrossRef\]](https://doi.org/10.1016/j.ijcard.2017.10.052) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/29407076)
- 89. Herscovici, R.; Sedlak, T.; Wei, J.; Pepine, C.J.; Handberg, E.; Bairey Merz, C.N. Ischemia and No Obstructive Coronary Artery Disease (INOCA): What Is the Risk? *J. Am. Heart Assoc.* **2018**, *7*, e008868. [\[CrossRef\]](https://doi.org/10.1161/JAHA.118.008868) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/30371178)
- 90. Lind, L.; Berglund, L.; Larsson, A.; Sundström, J. Endothelial function in resistance and conduit arteries and 5-year risk of cardiovascular disease. *Circulation* **2011**, *123*, 1545–1551. [\[CrossRef\]](https://doi.org/10.1161/CIRCULATIONAHA.110.984047)
- 91. Maseri, A.; Severi, S.; Nes, M.D.; L'Abbate, A.; Chierchia, S.; Marzilli, M.; Ballestra, A.M.; Parodi, O.; Biagini, A.; Distante, A. "Variant" angina: One aspect of a continuous spectrum of vasospastic myocardial ischemia. Pathogenetic mechanisms, estimated incidence and clinical and coronary arteriographic findings in 138 patients. *Am. J. Cardiol.* **1978**, *42*, 1019–1035. [\[CrossRef\]](https://doi.org/10.1016/0002-9149(78)90691-4) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/727129)
- 92. Hung, M.Y.; Mao, C.T.; Hung, M.J.; Wang, J.K.; Lee, H.C.; Yeh, C.T.; Hu, P.; Chen, T.H.; Chang, N.C. Coronary Artery Spasm as Related to Anxiety and Depression: A Nationwide Population-Based Study. *Psychosom. Med.* **2019**, *81*, 237–245. [\[CrossRef\]](https://doi.org/10.1097/PSY.0000000000000666)
- 93. Sueda, S.; Kohno, H.; Yoshino, H. The real world in the clinic before and after the establishment of guidelines for coronary artery spasm: A questionnaire for members of the Japanese Cine-angio Association. *Heart Vessel.* **2017**, *32*, 637–643. [\[CrossRef\]](https://doi.org/10.1007/s00380-016-0916-9)
- 94. Schang, S.J., Jr.; Pepine, C.J. Transient asymptomatic S-T segment depression during daily activity. *Am. J. Cardiol.* **1977**, *39*, 396–402. [\[CrossRef\]](https://doi.org/10.1016/S0002-9149(77)80095-7)
- 95. Mudge, G.H., Jr.; Grossman, W.; Mills, R.M., Jr.; Lesch, M.; Braunwald, E. Reflex increase in coronary vascular resistance in patients with ischemic heart disease. *N. Engl. J. Med.* **1976**, *295*, 1333–1337. [\[CrossRef\]](https://doi.org/10.1056/NEJM197612092952401)
- 96. Priedberg, C.K. Some comments and reflections on changing interests and new developments in angina pectoris. *Circulation* **1972**, *46*, 1037–1047. [\[CrossRef\]](https://doi.org/10.1161/01.CIR.46.6.1037)
- 97. Gibbons, R.J.; Abrams, J.; Chatterjee, K.; Daley, J.; Deedwania, P.C.; Douglas, J.S.; Ferguson, T.B., Jr.; Fihn, S.D.; Fraker, T.D., Jr.; Gardin, J.M.; et al. ACC/AHA 2002 guideline update for the management of patients with chronic stable angina—Summary article: A report of the American College of Cardiology/American Heart Association Task Force on Practice Guidelines (Committee on the Management of Patients With Chronic Stable Angina). *Circulation* **2003**, *107*, 149–158.
- 98. Apstein, C.S.; Grossman, W. Opposite initial effects of supply and demand ischemia on left ventricular diastolic compliance: The ischemia-diastolic paradox. *J. Mol. Cell. Cardiol.* **1987**, *19*, 119–128. [\[CrossRef\]](https://doi.org/10.1016/S0022-2828(87)80551-5)
- 99. Wilson, R.F.; Marcus, M.L.; White, C.W. Prediction of the physiologic significance of coronary arterial lesions by quantitative lesion geometry in patients with limited coronary artery disease. *Circulation* **1987**, *75*, 723–732. [\[CrossRef\]](https://doi.org/10.1161/01.CIR.75.4.723) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/3829334)
- 100. Maseri, A.; Parodi, O.; Severi, S.; Pesola, A. Transient transmural reduction of myocardial blood flow demonstrated by thallium-201 scintigraphy, as a cause of variant angina. *Circulation* **1976**, *54*, 280–288. [\[CrossRef\]](https://doi.org/10.1161/01.CIR.54.2.280)
- 101. Arrebola-Moreno, A.L.; Arrebola, J.P.; Moral-Ruiz, A.; Ramirez-Hernandez, J.A.; Melgares-Moreno, R.; Kaski, J.C. Coronary microvascular spasm triggers transient ischemic left ventricular diastolic abnormalities in patients with chest pain and angiographically normal coronary arteries. *Atherosclerosis* **2014**, *236*, 207e–214e. [\[CrossRef\]](https://doi.org/10.1016/j.atherosclerosis.2014.07.009) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/25075937)
- 102. Sonnenblick, E.H.; Fein, F.; Capasso, J.M.; Factor, S.M. Microvascular spasm as a cause of cardiomyopathies and the calciumblocking agent verapamil as potential primary therapy. *Am. J. Cardiol.* **1985**, *55*, 179B–184B. [\[CrossRef\]](https://doi.org/10.1016/0002-9149(85)90629-0) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/3881912)
- 103. Pries, A.R.; Reglin, B. Coronary microcirculatory pathophysiology: Can we afford it to remain a black box? *Eur. Heart J.* **2017**, *38*, 478–488. [\[CrossRef\]](https://doi.org/10.1093/eurheartj/ehv760) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/26843279)
- 104. Lee, J.F.; Barrett-O'Keefe, Z.; Garten, R.S.; Nelson, A.D.; Ryan, J.J.; Nativi, J.N.; Richardson, R.S.; Wray, D.W. Evidence of microvascular dysfunction in heart failure with preserved ejection fraction. *Heart* **2016**, *102*, 278–284. [\[CrossRef\]](https://doi.org/10.1136/heartjnl-2015-308403)
- 105. De Boer, R.A.; Pinto, Y.M.; Van Veldhuisen, D.J. The imbalance between oxygen demand and supply as a potential mechanism in the pathophysiology of heart failure: The role of microvascular growth and abnormalities. *Microcirculation* **2003**, *10*, 113–126. [\[CrossRef\]](https://doi.org/10.1080/713773607)
- 106. Ong, P.; Athanasiadis, A.; Borgulya, G.; Mahrholdt, H.; Kaski, J.C.; Sechtem, U. High prevalence of a pathological response to acetylcholine testing in patients with stable angina pectoris and unobstructed coronary arteries. The ACOVA Study (Abnormal COronaryVAsomotion in patients with stable angina and unobstructed coronary arteries). *J. Am. Coll. Cardiol* **2013**, *59*, 655–662. [\[CrossRef\]](https://doi.org/10.1016/j.jacc.2011.11.015) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/22322081)
- 107. Higgins, C.B.; Wexler, L.; Silverman, J.F.; Schroeder, J.S. Clinical and arteriographic features of Prinzmetal's variant angina: Documentation of etiologic factors. *Am. J. Cardiol.* **1976**, *37*, 831–839. [\[CrossRef\]](https://doi.org/10.1016/0002-9149(76)90106-5)
- 108. Parchure, N.; Batchvarov, V.; Malik, M.; Camm, A.J.; Kaski, J.C. Increased QT dispersion in patients with Prinzmetal's variant angina and cardiac arrest. *Cardiovasc. Res.* **2001**, *50*, 379–385. [\[CrossRef\]](https://doi.org/10.1016/S0008-6363(00)00290-X) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/11334842)
- 109. Selzer, A.; Langston, M.; Ruggeroli, C.; Cohn, K. Clinical syndrome of variant angina with normal coronary arteriogram. *N. Engl. J. Med.* **1976**, *295*, 1343–1347. [\[CrossRef\]](https://doi.org/10.1056/NEJM197612092952403)
- 110. Dalen, J.E.; Ockene, I.S.; Alpert, J.S. Coronary spasm, coronary thrombosis, and myocardial infarction: A hypothesis concerning the pathophysiology of acute myocardial infarction. *Am. Heart J.* **1982**, *104 Pt 1*, 1119–1124. [\[CrossRef\]](https://doi.org/10.1016/0002-8703(82)90458-6)
- 111. Farmer, D.M.; Swygman, C.A.; Wang, P.J.; Mark Estes, N.A., 3rd; Link, M.S. Evidence that nonsustained polymorphic ventricular tachycardia causes syncope (data from implantable cardioverter defibrillators). *Am. J. Cardiol.* **2003**, *91*, 606–609. [\[CrossRef\]](https://doi.org/10.1016/S0002-9149(02)03319-2)
- 112. Kamakura, T.; Wada, M.; Ishibashi, K.; Inoue, Y.Y.; Miyamoto, K.; Okamura, H.; Nagase, S.; Noda, T.; Aiba, T.; Yasuda, S.; et al. Significance of Coronary Artery Spasm Diagnosis in Patients with Early Repolarization Syndrome. *J. Am. Heart Assoc.* **2018**, *7*, e007942. [\[CrossRef\]](https://doi.org/10.1161/JAHA.117.007942)
- 113. Ali, A.; Butt, N.; Sheikh, A.S. Early repolarization syndrome: A cause of sudden cardiac death. *World J. Cardiol.* **2015**, *7*, 466–475. [\[CrossRef\]](https://doi.org/10.4330/wjc.v7.i8.466)
- 114. Sueda, S.; Fujimoto, K.; Sasaki, Y.; Habara, H.; Kohno, H. Cardiogenic Shock due to Pulseless Electrical Activity Arrest Associated with Severe Coronary Artery Spasm. *Intern. Med.* **2018**, *57*, 2853–2857. [\[CrossRef\]](https://doi.org/10.2169/internalmedicine.0196-17)
- 115. Letsas, K.P.; Filippatos, G.S.; Efremidis, M.; Sideris, A.; Kardaras, F. Secondary prevention of sudden cardiac death in coronary artery spasm: Is implantable cardioverter defibrillator always efficient? *Int. J. Cardiol.* **2007**, *117*, 141–143. [\[CrossRef\]](https://doi.org/10.1016/j.ijcard.2006.06.055)
- 116. Nakamura, M. Our animal model of coronary spasm—My personal view. *J. Atheroscler. Thromb.* **2000**, *6*, 1–12. [\[CrossRef\]](https://doi.org/10.5551/jat1994.6.1)
- 117. Nobuyoshi, M.; Tanaka, M.; Nosaka, H.; Kimura, T.; Yokoi, H.; Hamasaki, N.; Kim, K.; Shindo, T.; Kimura, K. Progression of coronary atherosclerosis: Is coronary spasm related to progression? *J. Am. Coll. Cardiol.* **1991**, *18*, 904–910. [\[CrossRef\]](https://doi.org/10.1016/0735-1097(91)90745-U)
- 118. Kuga, T.; Tagawa, H.; Tomoike, H.; Mitsuoka, W.; Egashira, S.; Ohara, Y.; Takeshita, A.; Nakamura, M. Role of coronary artery spasm in progression of organic coronary stenosis and acute myocardial infarction in a swine model. Importance of mode of onset and duration of coronary artery spasm. *Circulation* **1993**, *87*, 573–582. [\[CrossRef\]](https://doi.org/10.1161/01.CIR.87.2.573) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/8425301)
- 119. Mizuno, Y.; Hokimoto, S.; Harada, E.; Kinoshita, K.; Nakagawa, K.; Yoshimura, M.; Ogawa, H.; Yasue, H. Variant Aldehyde Dehydrogenase 2 (ALDH2*2) Is a Risk Factor for Coronary Spasm and ST-Segment Elevation Myocardial Infarction. *J. Am. Heart Assoc.* **2016**, *5*, e003247. [\[CrossRef\]](https://doi.org/10.1161/JAHA.116.003247) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/27153870)
- 120. Miwa, K.; Nakagawa, K.; Yoshida, N.; Taguchi, Y.; Inoue, H. Lipoprotein(a) is a risk factor for occurrence of acute myocardial infarction in patients with coronary vasospasm. *J. Am. Coll. Cardiol.* **2000**, *35*, 1200–1205. [\[CrossRef\]](https://doi.org/10.1016/S0735-1097(00)00550-7)
- 121. Sugiishi, M.; Takatsu, F. Cigarette smoking is a major risk factor for coronary spasm. *Circulation* **1993**, *87*, 76–79. [\[CrossRef\]](https://doi.org/10.1161/01.CIR.87.1.76) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/8419026)
- 122. Hung, M.Y.; Hsu, K.H.; Hung, M.J.; Cheng, C.W.; Kuo, L.T.; Cherng, W.J. Interaction between cigarette smoking and highsensitivity C-reactive protein in the development of coronary vasospasm in patients without hemodynamically significant coronary artery disease. *Am. J. Med. Sci.* **2009**, *338*, 440–446. [\[CrossRef\]](https://doi.org/10.1097/MAJ.0b013e3181b9147f)
- 123. Hung, M.J.; Hsu, K.H.; Hu, W.S.; Chang, N.C.; Hung, M.Y. C-reactive protein for predicting prognosis and its gender-specific associations with diabetes mellitus and hypertension in the development of coronary artery spasm. *PLoS ONE* **2013**, *8*, e77655. [\[CrossRef\]](https://doi.org/10.1371/journal.pone.0077655)
- 124. D'Amario, D.; Migliaro, S.; Borovac, J.A.; Restivo, A.; Vergallo, R.; Galli, M.; Leone, A.M.; Montone, R.A.; Niccoli, G.; Aspromonte, N.; et al. Microvascular Dysfunction in Heart Failure with Preserved Ejection Fraction. *Front. Physiol.* **2019**, *10*, 1347. [\[CrossRef\]](https://doi.org/10.3389/fphys.2019.01347)
- 125. Kandabashi, T.; Shimokawa, H.; Miyata, K.; Kunihiro, I.; Kawano, Y.; Fukata, Y.; Higo, T.; Egashira, K.; Takahashi, S.; Kaibuchi, K.; et al. Inhibition of myosin phosphatase by upregulated rho-kinase plays a key role for coronary artery spasm in a porcine model with interleukin-1beta. *Circulation* **2000**, *101*, 1319–1323. [\[CrossRef\]](https://doi.org/10.1161/01.CIR.101.11.1319)
- 126. Shimokawa, H.; Tomoike, H.; Nabeyama, S.; Yamamoto, H.; Ishii, Y.; Tanaka, K.; Nakamura, M. Coronary artery spasm induced in miniature swine: Angiographic evidence and relation to coronary atherosclerosis. *Am. Heart J.* **1985**, *110*, 300–310. [\[CrossRef\]](https://doi.org/10.1016/0002-8703(85)90148-6) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/4025107)
- 127. Kandabashi, T.; Shimokawa, H.; Miyata, K.; Kunihiro, I.; Eto, Y.; Morishige, K.; Matsumoto, Y.; Obara, K.; Nakayama, K.; Takahashi, S.; et al. Evidence for protein kinase C-mediated activation of Rho-kinase in a porcine model of coronary artery spasm. *Arterioscler. Thromb. Vasc. Biol.* **2003**, *23*, 2209–2214. [\[CrossRef\]](https://doi.org/10.1161/01.ATV.0000104010.87348.26)
- 128. Chutkow, W.A.; Pu, J.; Wheeler, M.T.; Wada, T.; Makielski, J.C.; Burant, C.F.; McNally, E.M. Episodic coronary artery vasospasm and hypertension develop in the absence of Sur2 K(ATP) channels. *J. Clin. Investig.* **2002**, *110*, 203–208. [\[CrossRef\]](https://doi.org/10.1172/JCI0215672) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/12122112)
- 129. Kakkar, R.; Ye, B.; Stoller, D.A.; Smelley, M.; Shi, N.Q.; Galles, K.; Hadhazy, M.; Makielski, J.C.; McNally, E.M. Spontaneous coronary vasospasm in KATP mutant mice arises from a smooth muscle-extrinsic process. *Circ. Res.* **2006**, *98*, 682–689. [\[CrossRef\]](https://doi.org/10.1161/01.RES.0000207498.40005.e7)
- 130. Chen, C.C.; Lamping, K.G.; Nuno, D.W.; Barresi, R.; Prouty, S.J.; Lavoie, J.L.; Cribbs, L.L.; England, S.K.; Sigmund, C.D.; Weiss, R.M.; et al. Abnormal coronary function in mice deficient in α1H T-type Ca2+ channels. *Science* **2003**, *302*, 1416–1418. [\[CrossRef\]](https://doi.org/10.1126/science.1089268)
- 131. Bozkurt, B.; Coats, A.J.S.; Tsutsui, H.; Abdelhamid, C.M.; Adamopoulos, S.; Albert, N.; Anker, S.D.; Atherton, J.; Böhm, M.; Butler, J.; et al. Universal definition and classification of heart failure: A report of the Heart Failure Society of America, Heart Failure Association of the European Society of Cardiology, Japanese Heart Failure Society and Writing Committee of the Universal Definition of Heart Failure: Endorsed by the Canadian Heart Failure Society, Heart Failure Association of India, Cardiac Society of Australia and New Zealand, and Chinese Heart Failure Association. *Eur. J. Heart Fail.* **2021**, *23*, 352–380. [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/33605000)
- 132. Hung, M.J.; Hsu, K.H.; Chang, N.C.; Hung, M.Y. Increased Numbers of Coronary Events in Winter and Spring Due to Coronary Artery Spasm: Effect of Age, Sex, Smoking, and Inflammation. *J. Am. Coll. Cardiol.* **2015**, *65*, 2047–2048. [\[CrossRef\]](https://doi.org/10.1016/j.jacc.2015.02.060)
- 133. Hoffman, R.M.; Psaty, B.M.; Kronmal, R.A. Modifiable Risk Factors for Incident Heart Failure in the Coronary Artery Surgery Study. *Arch. Intern. Med.* **1994**, *154*, 417–423. [\[CrossRef\]](https://doi.org/10.1001/archinte.1994.00420040081012) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/8117174)
- 134. Guazzi, M.; Polese, A.; Fiorentini, C.; Magrini, F.; Bartorelli, C. Left ventricular performance and related haemodynamic changes in Prinzmetal's variant angina pectoris. *Br. Heart J.* **1971**, *33*, 84–94. [\[CrossRef\]](https://doi.org/10.1136/hrt.33.1.84)
- 135. Westermann, D.; Lindner, D.; Kasner, M.; Zietsch, C.; Savvatis, K.; Escher, F.; von Schlippenbach, J.; Skurk, C.; Steendijk, P.; Riad, A.; et al. Cardiac inflammation contributes to changes in the extracellular matrix in patients with heart failure and normal ejection fraction. *Circ. Heart Fail.* **2011**, *4*, 44–52. [\[CrossRef\]](https://doi.org/10.1161/CIRCHEARTFAILURE.109.931451) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/21075869)
- 136. Mohammed, S.F.; Hussain, S.; Mirzoyev, S.A.; Edwards, W.D.; Maleszewski, J.J.; Redfield, M.M. Coronary microvascular rarefaction and myocardial fibrosis in heart failure with preserved ejection fraction. *Circulation* **2015**, *131*, 550–559. [\[CrossRef\]](https://doi.org/10.1161/CIRCULATIONAHA.114.009625)
- 137. Van Heerebeek, L.; Borbély, A.; Niessen, H.W.; Bronzwaer, J.G.; van der Velden, J.; Stienen, G.J.; Linke, W.A.; Laarman, G.J.; Paulus, W.J. Myocardial structure and function differ in systolic and diastolic heart failure. *Circulation* **2006**, *113*, 1966–1973. [\[CrossRef\]](https://doi.org/10.1161/CIRCULATIONAHA.105.587519)
- 138. Burchfield, J.S.; Xie, M.; Hill, J.A. Pathological ventricular remodeling: Mechanisms: Part 1 of 2. *Circulation* **2013**, *128*, 388–400. [\[CrossRef\]](https://doi.org/10.1161/CIRCULATIONAHA.113.001878)
- 139. Sanders-van Wijk, S.; van Empel, V.; Davarzani, N.; Maeder, M.T.; Handschin, R.; Pfisterer, M.E.; Brunner-La Rocca, H.P.; TIME-CHF investigators. Circulating biomarkers of distinct pathophysiological pathways in heart failure with preserved vs. reduced left ventricular ejection fraction. *Eur. J. Heart Fail.* **2015**, *17*, 1006–1014. [\[CrossRef\]](https://doi.org/10.1002/ejhf.414)
- 140. Wijns, W.; Vatner, S.F.; Camici, P.G. Hibernating myocardium. *N. Engl. J. Med.* **1998**, *339*, 173–181. [\[CrossRef\]](https://doi.org/10.1056/NEJM199807163390307)
- 141. Kloner, R.A.; Przyklenk, K. Hibernation and stunning of the myocardium. *N. Engl. J. Med.* **1991**, *325*, 1877–1879. [\[CrossRef\]](https://doi.org/10.1056/NEJM199112263252610)
- 142. Bolli, R. Myocardial 'stunning' in man. *Circulation* **1992**, *86*, 1671–1691. [\[CrossRef\]](https://doi.org/10.1161/01.CIR.86.6.1671) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/1451239)
- 143. Auerbach, M.A.; Schöder, H.; Hoh, C.; Gambhir, S.S.; Yaghoubi, S.; Sayre, J.W.; Silverman, D.; Phelps, M.E.; Schelbert, H.R.; Czernin, J. Prevalence of myocardial viability as detected by positron emission tomography in patients with ischemic cardiomyopathy. *Circulation* **1999**, *99*, 2921–2926. [\[CrossRef\]](https://doi.org/10.1161/01.CIR.99.22.2921)
- 144. Wei, L.; Kadoya, M.; Momose, M.; Kurozumi, M.; Matsushita, T.; Yamada, A. Serial assessment of left ventricular function in various patient groups with Tl-201 gated myocardial perfusion SPECT. *Radiat. Med.* **2007**, *25*, 65–72. [\[CrossRef\]](https://doi.org/10.1007/s11604-006-0105-3) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/17541515)
- 145. Bell, S.P.; Fabian, J.; Watkins, M.W.; LeWinter, M.M. Decrease in forces responsible for diastolic suction during acute coronary occlusion. *Circulation* **1997**, *96*, 2348–2352. [\[CrossRef\]](https://doi.org/10.1161/01.CIR.96.7.2348) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/9337210)
- 146. Firstenberg, M.S.; Smedira, N.G.; Greenberg, N.L.; Prior, D.L.; McCarthy, P.M.; Garcia, M.J.; Thomas, J.D. Relationship between early diastolic intraventricular pressure gradients, an index of elastic recoil, and improvements in systolic and diastolic function. *Circulation* **2001**, *104* (Suppl. S1), I330–I335. [\[CrossRef\]](https://doi.org/10.1161/hc37t1.094834)
- 147. Uren, N.G.; Melin, J.A.; De Bruyne, B.; Wijns, W.; Baudhuin, T.; Camici, P.G. Relation between myocardial blood flow and the severity of coronary-artery stenosis. *N. Engl. J. Med.* **1994**, *330*, 1782–1788. [\[CrossRef\]](https://doi.org/10.1056/NEJM199406233302503) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/8190154)
- 148. Di Carli, M.; Czernin, J.; Hoh, C.K.; Gerbaudo, V.H.; Brunken, R.C.; Huang, S.C.; Phelps, M.E.; Schelbert, H.R. Relation among stenosis severity, myocardial blood flow, and flow reserve in patients with coronary artery disease. *Circulation* **1995**, *91*, 1944–1951. [\[CrossRef\]](https://doi.org/10.1161/01.CIR.91.7.1944)
- 149. Jackson, M.; Marks, L.; May, G.H.W.; Wilson, J.B. The genetic basis of disease. *Essays Biochem.* **2018**, *62*, 643–723. [\[CrossRef\]](https://doi.org/10.1042/EBC20170053)
- 150. Franczyk, B.; Dybiec, J.; Frąk, W.; Krzemińska, J.; Kućmierz, J.; Młynarska, E.; Szlagor, M.; Wronka, M.; Rysz, J. Cellular Mechanisms of Coronary Artery Spasm. *Biomedicines* **2022**, *10*, 2349. [\[CrossRef\]](https://doi.org/10.3390/biomedicines10102349)
- 151. Mizuno, Y.; Hokimoto, S.; Harada, E.; Kinoshita, K.; Yoshimura, M.; Yasue, H. Variant Aldehyde Dehydrogenase 2 (ALDH2*2) in East Asians Interactively Exacerbates Tobacco Smoking Risk for Coronary Spasm-Possible Role of Reactive Aldehydes. *Circ. J.* **2016**, *81*, 96–102. [\[CrossRef\]](https://doi.org/10.1253/circj.CJ-16-0969)
- 152. Mizuno, Y.; Harada, E.; Morita, S.; Kinoshita, K.; Hayashida, M.; Shono, M.; Morikawa, Y.; Murohara, T.; Nakayama, M.; Yoshimura, M.; et al. East asian variant of aldehyde dehydrogenase 2 is associated with coronary spastic angina: Possible roles of reactive aldehydes and implications of alcohol flushing syndrome. *Circulation* **2015**, *131*, 1665–1673. [\[CrossRef\]](https://doi.org/10.1161/CIRCULATIONAHA.114.013120) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/25759460)
- 153. Schulz, E.; Jansen, T.; Wenzel, P.; Daiber, A.; Münzel, T. Nitric oxide, tetrahydrobiopterin, oxidative stress, and endothelial dysfunction in hypertension. *Antioxid. Redox Signal.* **2008**, *10*, 1115–1126. [\[CrossRef\]](https://doi.org/10.1089/ars.2007.1989) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/18321209)
- 154. Vettel, C.; Lämmle, S.; Ewens, S.; Cervirgen, C.; Emons, J.; Ongherth, A.; Dewenter, M.; Lindner, D.; Westermann, D.; Nikolaev, V.O.; et al. PDE2-mediated cAMP hydrolysis accelerates cardiac fibroblast to myofibroblast conversion and is antagonized by exogenous activation of cGMP signaling pathways. *Am. J. Physiol. Heart Circ. Physiol.* **2014**, *306*, H1246–H1252. [\[CrossRef\]](https://doi.org/10.1152/ajpheart.00852.2013) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/24531807)
- 155. Crea, F.; Bairey Merz, C.N.; Beltrame, J.F.; Kaski, J.C.; Ogawa, H.; Ong, P.; Sechtem, U.; Shimokawa, H.; Camici, P.G.; Coronary Vasomotion Disorders International Study Group (COVADIS). The parallel tales of microvascular angina and heart failure with preserved ejection fraction: A paradigm shift. *Eur. Heart J.* **2017**, *38*, 473–477. [\[CrossRef\]](https://doi.org/10.1093/eurheartj/ehw461)
- 156. Tschöpe, C.; Post, H. Latent ischaemia as a trigger for a circulus vitiosus of inflammation, fibrosis, and stiffness in HFPEF. *Eur. J. Heart Fail.* **2015**, *17*, 1210–1212. [\[CrossRef\]](https://doi.org/10.1002/ejhf.439) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/26647213)
- 157. Silverman, M.G.; Patel, B.; Blankstein, R.; Lima, J.A.; Blumenthal, R.S.; Nasir, K.; Blaha, M.J. Impact of Race, Ethnicity, and Multimodality Biomarkers on the Incidence of New-Onset Heart Failure with Preserved Ejection Fraction (from the Multi-Ethnic Study of Atherosclerosis). *Am. J. Cardiol.* **2016**, *117*, 1474–1481. [\[CrossRef\]](https://doi.org/10.1016/j.amjcard.2016.02.017) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/27001449)
- 158. Hein, S.; Arnon, E.; Kostin, S.; Schönburg, M.; Elsässer, A.; Polyakova, V.; Bauer, E.P.; Klövekorn, W.P.; Schaper, J. Progression from compensated hypertrophy to failure in the pressure-overloaded human heart: Structural deterioration and compensatory mechanisms. *Circulation* **2003**, *107*, 984–991. [\[CrossRef\]](https://doi.org/10.1161/01.CIR.0000051865.66123.B7)
- 159. Schirone, L.; Forte, M.; Palmerio, S.; Yee, D.; Nocella, C.; Angelini, F.; Pagano, F.; Schiavon, S.; Bordin, A.; Carrizzo, A.; et al. A Review of the Molecular Mechanisms Underlying the Development and Progression of Cardiac Remodeling. *Oxidative Med. Cell. Longev.* **2017**, *2017*, 3920195. [\[CrossRef\]](https://doi.org/10.1155/2017/3920195)
- 160. Lehnart, S.E.; Maier, L.S.; Hasenfuss, G. Abnormalities of calcium metabolism and myocardial contractility depression in the failing heart. *Heart Fail. Rev.* **2009**, *14*, 213–224. [\[CrossRef\]](https://doi.org/10.1007/s10741-009-9146-x)
- 161. Jugdutt, B.I. Ventricular remodeling after infarction and the extracellular collagen matrix: When is enough enough? *Circulation* **2003**, *108*, 1395–1403. [\[CrossRef\]](https://doi.org/10.1161/01.CIR.0000085658.98621.49)
- 162. Barouch, L.A.; Gao, D.; Chen, L.; Miller, K.L.; Xu, W.; Phan, A.C.; Kittleson, M.M.; Minhas, K.M.; Berkowitz, D.E.; Wei, C.; et al. Cardiac myocyte apoptosis is associated with increased DNA damage and decreased survival in murine models of obesity. *Circ. Res.* **2006**, *98*, 119–124. [\[CrossRef\]](https://doi.org/10.1161/01.RES.0000199348.10580.1d)
- 163. Kleinbongard, P.; Schulz, R.; Heusch, G. TNFα in myocardial ischemia/reperfusion, remodeling and heart failure. *Heart Fail. Rev.* **2011**, *16*, 49–69. [\[CrossRef\]](https://doi.org/10.1007/s10741-010-9180-8)
- 164. Koitabashi, N.; Danner, T.; Zaiman, A.L.; Pinto, Y.M.; Rowell, J.; Mankowski, J.; Zhang, D.; Nakamura, T.; Takimoto, E.; Kass, D.A. Pivotal role of cardiomyocyte TGF-β signaling in the murine pathological response to sustained pressure overload. *J. Clin. Investig.* **2011**, *121*, 2301–2312. [\[CrossRef\]](https://doi.org/10.1172/JCI44824)
- 165. Yamauchi-Takihara, K.; Ihara, Y.; Ogata, A.; Yoshizaki, K.; Azuma, J.; Kishimoto, T. Hypoxic stress induces cardiac myocytederived interleukin-6. *Circulation* **1995**, *91*, 1520–1524. [\[CrossRef\]](https://doi.org/10.1161/01.CIR.91.5.1520)
- 166. Shioi, T.; Matsumori, A.; Kihara, Y.; Inoko, M.; Ono, K.; Iwanaga, Y.; Yamada, T.; Iwasaki, A.; Matsushima, K.; Sasayama, S. Increased expression of interleukin-1 beta and monocyte chemotactic and activating factor/monocyte chemoattractant protein-1 in the hypertrophied and failing heart with pressure overload. *Circ. Res.* **1997**, *81*, 664–671. [\[CrossRef\]](https://doi.org/10.1161/01.RES.81.5.664)
- 167. Kandalam, V.; Basu, R.; Moore, L.; Fan, D.; Wang, X.; Jaworski, D.M.; Oudit, G.Y.; Kassiri, Z. Lack of tissue inhibitor of metalloproteinases 2 leads to exacerbated left ventricular dysfunction and adverse extracellular matrix remodeling in response to biomechanical stress. *Circulation* **2011**, *124*, 2094–2105. [\[CrossRef\]](https://doi.org/10.1161/CIRCULATIONAHA.111.030338)
- 168. Wang, J.; Chen, H.; Seth, A.; McCulloch, C.A. Mechanical force regulation of myofibroblast differentiation in cardiac fibroblasts. *Am. J. Physiol. Heart Circ. Physiol.* **2003**, *285*, H1871–H1881. [\[CrossRef\]](https://doi.org/10.1152/ajpheart.00387.2003) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/12842814)
- 169. Porter, K.E.; Turner, N.A. Cardiac fibroblasts: At the heart of myocardial remodeling. *Pharmacol. Ther.* **2009**, *123*, 255–278. [\[CrossRef\]](https://doi.org/10.1016/j.pharmthera.2009.05.002) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/19460403)
- 170. Segura, A.M.; Frazier, O.H.; Buja, L.M. Fibrosis and heart failure. *Heart Fail. Rev.* **2014**, *19*, 173–185. [\[CrossRef\]](https://doi.org/10.1007/s10741-012-9365-4) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/23124941)
- 171. Prabhu, S.D.; Frangogiannis, N.G. The Biological Basis for Cardiac Repair After Myocardial Infarction: From Inflammation to Fibrosis. *Circ. Res.* **2016**, *119*, 91–112. [\[CrossRef\]](https://doi.org/10.1161/CIRCRESAHA.116.303577) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/27340270)
- 172. Fujita, S.; Shimojo, N.; Terasaki, F.; Otsuka, K.; Hosotani, N.; Kohda, Y.; Tanaka, T.; Nishioka, T.; Yoshida, T.; Hiroe, M.; et al. Atrial natriuretic peptide exerts protective action against angiotensin II-induced cardiac remodeling by attenuating inflammation via endothelin-1/endothelin receptor A cascade. *Heart Vessel.* **2013**, *28*, 646–657. [\[CrossRef\]](https://doi.org/10.1007/s00380-012-0311-0)
- 173. Sarkar, S.; Vellaichamy, E.; Young, D.; Sen, S. Influence of cytokines and growth factors in ANG II-mediated collagen upregulation by fibroblasts in rats: Role of myocytes. *Am. J. Physiol. Heart Circ. Physiol.* **2004**, *287*, H107–H117. [\[CrossRef\]](https://doi.org/10.1152/ajpheart.00763.2003) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/15059775)
- 174. Zeisberg, E.M.; Tarnavski, O.; Zeisberg, M.; Dorfman, A.L.; McMullen, J.R.; Gustafsson, E.; Chandraker, A.; Yuan, X.; Pu, W.T.; Roberts, A.B.; et al. Endothelial-to-mesenchymal transition contributes to cardiac fibrosis. *Nat. Med.* **2007**, *13*, 952–961. [\[CrossRef\]](https://doi.org/10.1038/nm1613) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/17660828)
- 175. Appari, M.; Breitbart, A.; Brandes, F.; Szaroszyk, M.; Froese, N.; Korf-Klingebiel, M.; Mohammadi, M.M.; Grund, A.; Scharf, G.M.; Wang, H.; et al. C1q-TNF-Related Protein-9 Promotes Cardiac Hypertrophy and Failure. *Circ. Res.* **2017**, *120*, 66–77. [\[CrossRef\]](https://doi.org/10.1161/CIRCRESAHA.116.309398)
- 176. Mortensen, R.M. Immune cell modulation of cardiac remodeling. *Circulation* **2012**, *125*, 1597–1600. [\[CrossRef\]](https://doi.org/10.1161/CIRCULATIONAHA.112.097832) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/22388322)
- 177. Zhang, W.; Chancey, A.L.; Tzeng, H.P.; Zhou, Z.; Lavine, K.J.; Gao, F.; Sivasubramanian, N.; Barger, P.M.; Mann, D.L. The development of myocardial fibrosis in transgenic mice with targeted overexpression of tumor necrosis factor requires mast cell-fibroblast interactions. *Circulation* **2011**, *124*, 2106–2116. [\[CrossRef\]](https://doi.org/10.1161/CIRCULATIONAHA.111.052399)
- 178. Lakhani, I.; Wong, M.V.; Hung, J.K.F.; Gong, M.; Waleed, K.B.; Xia, Y.; Lee, S.; Roever, L.; Liu, T.; Tse, G.; et al. Diagnostic and prognostic value of serum C-reactive protein in heart failure with preserved ejection fraction: A systematic review and meta-analysis. *Heart Fail. Rev.* **2021**, *26*, 1141–1150. [\[CrossRef\]](https://doi.org/10.1007/s10741-020-09927-x) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/32030562)
- 179. Mann, D.L. Innate immunity and the failing heart: The cytokine hypothesis revisited. *Circ. Res.* **2015**, *116*, 1254–1268. [\[CrossRef\]](https://doi.org/10.1161/CIRCRESAHA.116.302317) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/25814686)
- 180. Bartekova, M.; Radosinska, J.; Jelemensky, M.; Dhalla, N.S. Role of cytokines and inflammation in heart function during health and disease. *Heart Fail. Rev.* **2018**, *23*, 733–758. [\[CrossRef\]](https://doi.org/10.1007/s10741-018-9716-x)
- 181. Frati, G.; Schirone, L.; Chimenti, I.; Yee, D.; Biondi-Zoccai, G.; Volpe, M.; Sciarretta, S. An overview of the inflammatory signalling mechanisms in the myocardium underlying the development of diabetic cardiomyopathy. *Cardiovasc. Res.* **2017**, *113*, 378–388. [\[CrossRef\]](https://doi.org/10.1093/cvr/cvx011)
- 182. Neubauer, S. The failing heart—An engine out of fuel. *N. Engl. J. Med.* **2007**, *356*, 1140–1151. [\[CrossRef\]](https://doi.org/10.1056/NEJMra063052)
- 183. Desvergne, B.; Wahli, W. Peroxisome proliferator-activated receptors: Nuclear control of metabolism. *Endocr. Rev.* **1999**, *20*, 649–688.
- 184. Duncan, J.G.; Finck, B.N. The PPARalpha-PGC-1alpha Axis Controls Cardiac Energy Metabolism in Healthy and Diseased Myocardium. *PPAR Res.* **2008**, *2008*, 253817. [\[CrossRef\]](https://doi.org/10.1155/2008/253817)
- 185. Arany, Z.; He, H.; Lin, J.; Hoyer, K.; Handschin, C.; Toka, O.; Ahmad, F.; Matsui, T.; Chin, S.; Wu, P.H.; et al. Transcriptional coactivator PGC-1 alpha controls the energy state and contractile function of cardiac muscle. *Cell Metab.* **2005**, *1*, 259–271. [\[CrossRef\]](https://doi.org/10.1016/j.cmet.2005.03.002)
- 186. Ventura-Clapier, R.; Garnier, A.; Veksler, V. Transcriptional control of mitochondrial biogenesis: The central role of PGC-1alpha. *Cardiovasc. Res.* **2008**, *79*, 208–217. [\[CrossRef\]](https://doi.org/10.1093/cvr/cvn098)
- 187. Hsieh, M.C.F.; Das, D.; Sambandam, N.; Zhang, M.Q.; Nahlé, Z. Regulation of the PDK4 isozyme by the Rb-E2F1 complex. *J. Biol. Chem.* **2008**, *283*, 27410–27417. [\[CrossRef\]](https://doi.org/10.1074/jbc.M802418200)
- 188. Oudit, G.Y.; Sun, H.; Kerfant, B.G.; Crackower, M.A.; Penninger, J.M.; Backx, P.H. The role of phosphoinositide-3 kinase and PTEN in cardiovascular physiology and disease. *J. Mol. Cell. Cardiol.* **2004**, *37*, 449–471. [\[CrossRef\]](https://doi.org/10.1016/j.yjmcc.2004.05.015) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/15276015)
- 189. Song, X.; Kusakari, Y.; Xiao, C.Y.; Kinsella, S.D.; Rosenberg, M.A.; Scherrer-Crosbie, M.; Hara, K.; Rosenzweig, A.; Matsui, T. mTOR attenuates the inflammatory response in cardiomyocytes and prevents cardiac dysfunction in pathological hypertrophy. *Am. J. Physiol. Cell Physiol.* **2010**, *299*, C1256–C1266. [\[CrossRef\]](https://doi.org/10.1152/ajpcell.00338.2010) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/20861467)
- 190. Castoldi, G.; Di Gioia, C.R.; Bombardi, C.; Catalucci, D.; Corradi, B.; Gualazzi, M.G.; Leopizzi, M.; Mancini, M.; Zerbini, G.; Condorelli, G.; et al. MiR-133a regulates collagen 1A1, potential role of miR-133a in myocardial fibrosis in angiotensin II-dependent hypertension. *J. Cell. Physiol.* **2012**, *227*, 850–856. [\[CrossRef\]](https://doi.org/10.1002/jcp.22939) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/21769867)
- 191. Dai, D.F.; Johnson, S.C.; Villarin, J.J.; Chin, M.T.; Nieves-Cintrón, M.; Chen, T.; Marcinek, D.J.; Dorn, G.W., 2nd; Kang, Y.J.; Prolla, T.A.; et al. Mitochondrial oxidative stress mediates angiotensin II-induced cardiac hypertrophy and Galphaq overexpressioninduced heart failure. *Circ. Res.* **2011**, *108*, 837–846. [\[CrossRef\]](https://doi.org/10.1161/CIRCRESAHA.110.232306)
- 192. Arany, Z.; Novikov, M.; Chin, S.; Ma, Y.; Rosenzweig, A.; Spiegelman, B.M. Transverse aortic constriction leads to accelerated heart failure in mice lacking PPAR-gamma coactivator 1alpha. *Proc. Natl. Acad. Sci. USA* **2006**, *103*, 10086–10091. [\[CrossRef\]](https://doi.org/10.1073/pnas.0603615103)
- 193. Riehle, C.; Wende, A.R.; Zaha, V.G.; Pires, K.M.; Wayment, B.; Olsen, C.; Bugger, H.; Buchanan, J.; Wang, X.; Moreira, A.B.; et al. PGC-1β deficiency accelerates the transition to heart failure in pressure overload hypertrophy. *Circ. Res.* **2011**, *109*, 783–793. [\[CrossRef\]](https://doi.org/10.1161/CIRCRESAHA.111.243964)
- 194. Amorim, P.A.; Nguyen, T.D.; Shingu, Y.; Schwarzer, M.; Mohr, F.W.; Schrepper, A.; Doenst, T. Myocardial infarction in rats causes partial impairment in insulin response associated with reduced fatty acid oxidation and mitochondrial gene expression. *J. Thorac. Cardiovasc. Surg.* **2010**, *140*, 1160–1167. [\[CrossRef\]](https://doi.org/10.1016/j.jtcvs.2010.08.003) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/20850803)
- 195. Ikeuchi, M.; Matsusaka, H.; Kang, D.; Matsushima, S.; Ide, T.; Kubota, T.; Fujiwara, T.; Hamasaki, N.; Takeshita, A.; Sunagawa, K.; et al. Overexpression of mitochondrial transcription factor a ameliorates mitochondrial deficiencies and cardiac failure after myocardial infarction. *Circulation* **2005**, *112*, 683–690. [\[CrossRef\]](https://doi.org/10.1161/CIRCULATIONAHA.104.524835)
- 196. Sciarretta, S.; Yee, D.; Shenoy, V.; Nagarajan, N.; Sadoshima, J. The importance of autophagy in cardioprotection. *High Blood Press. Cardiovasc. Prev.* **2014**, *21*, 21–28. [\[CrossRef\]](https://doi.org/10.1007/s40292-013-0029-9) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/24235024)
- 197. Erickson, J.R.; Joiner, M.L.; Guan, X.; Kutschke, W.; Yang, J.; Oddis, C.V.; Bartlett, R.K.; Lowe, J.S.; O'Donnell, S.E.; Aykin-Burns, N.; et al. A dynamic pathway for calcium-independent activation of CaMKII by methionine oxidation. *Cell* **2008**, *133*, 462–474. [\[CrossRef\]](https://doi.org/10.1016/j.cell.2008.02.048)
- 198. Lee, B.K.; Lim, H.S.; Fearon, W.F.; Yong, A.S.; Yamada, R.; Tanaka, S.; Lee, D.P.; Yeung, A.C.; Tremmel, J.A. Invasive evaluation of patients with angina in the absence of obstructive coronary artery disease. *Circulation* **2015**, *131*, 1054–1060. [\[CrossRef\]](https://doi.org/10.1161/CIRCULATIONAHA.114.012636) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/25712205)
- 199. Khuddus, M.A.; Pepine, C.J.; Handberg, E.M.; Bairey Merz, C.N.; Sopko, G.; Bavry, A.A.; Denardo, S.J.; McGorray, S.P.; Smith, K.M.; Sharaf, B.L.; et al. An intravascular ultrasound analysis in women experiencing chest pain in the absence of obstructive coronary artery disease: A substudy from the National Heart, Lung and Blood Institute-Sponsored Women's Ischemia Syndrome Evaluation (WISE). *J. Interv. Cardiol.* **2010**, *23*, 511–519. [\[CrossRef\]](https://doi.org/10.1111/j.1540-8183.2010.00598.x)
- 200. Jenkins, M.J.; Edgley, A.J.; Sonobe, T.; Umetani, K.; Schwenke, D.O.; Fujii, Y.; Brown, R.D.; Kelly, D.J.; Shirai, M.; Pearson, J.T. Dynamic synchrotron imaging of diabetic rat coronary microcirculation in vivo. *Arterioscler. Thromb. Vasc. Biol.* **2012**, *32*, 370–377. [\[CrossRef\]](https://doi.org/10.1161/ATVBAHA.111.237172)
- 201. Konst, R.E.; Guzik, T.J.; Kaski, J.C.; Maas, A.H.E.M.; Elias-Smale, S.E. The pathogenic role of coronary microvascular dysfunction in the setting of other cardiac or systemic conditions. *Cardiovasc. Res.* **2020**, *116*, 817–828. [\[CrossRef\]](https://doi.org/10.1093/cvr/cvaa009)
- 202. Lanza, G.A.; Crea, F. Primary coronary microvascular dysfunction: Clinical presentation, pathophysiology, and management. *Circulation* **2010**, *121*, 2317–2325. [\[CrossRef\]](https://doi.org/10.1161/CIRCULATIONAHA.109.900191)
- 203. Reilkoff, R.A.; Bucala, R.; Herzog, E.L. Fibrocytes: Emerging effector cells in chronic inflammation. *Nat. Rev. Immunol.* **2011**, *11*, 427–435. [\[CrossRef\]](https://doi.org/10.1038/nri2990)
- 204. Pilling, D.; Buckley, C.D.; Salmon, M.; Gomer, R.H. Inhibition of fibrocyte differentiation by serum amyloid P. *J. Immunol.* **2003**, *171*, 5537–5546. [\[CrossRef\]](https://doi.org/10.4049/jimmunol.171.10.5537)
- 205. Li, P.; Wang, D.; Lucas, J.; Oparil, S.; Xing, D.; Cao, X.; Novak, L.; Renfrow, M.B.; Chen, Y.F. Atrial natriuretic peptide inhibits transforming growth factor beta-induced Smad signaling and myofibroblast transformation in mouse cardiac fibroblasts. *Circ. Res.* **2008**, *102*, 185–192. [\[CrossRef\]](https://doi.org/10.1161/CIRCRESAHA.107.157677) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/17991884)
- 206. Pepine, C.J.; Petersen, J.W.; Bairey Merz, C.N. A microvascular-myocardial diastolic dysfunctional state and risk for mental stress ischemia: A revised concept of ischemia during daily life. *JACC Cardiovasc. Imaging* **2014**, *7*, 362–365. [\[CrossRef\]](https://doi.org/10.1016/j.jcmg.2013.11.009)
- 207. Bork, N.I.; Molina, C.E.; Nikolaev, V.O. cGMP signalling in cardiomyocyte microdomains. *Biochem. Soc. Trans.* **2019**, *47*, 1327–1339. [\[CrossRef\]](https://doi.org/10.1042/BST20190225) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/31652306)
- 208. Van Heerebeek, L.; Hamdani, N.; Falcão-Pires, I.; Leite-Moreira, A.F.; Begieneman, M.P.; Bronzwaer, J.G.; van der Velden, J.; Stienen, G.J.; Laarman, G.J.; Somsen, A.; et al. Low myocardial protein kinase G activity in heart failure with preserved ejection fraction. *Circulation* **2012**, *126*, 830–839. [\[CrossRef\]](https://doi.org/10.1161/CIRCULATIONAHA.111.076075) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/22806632)
- 209. Liu, G.; Ma, C.; Yang, H.; Zhang, P.Y. Transforming growth factor β and its role in heart disease. *Exp. Ther. Med.* **2017**, *13*, 2123–2128. [\[CrossRef\]](https://doi.org/10.3892/etm.2017.4246)
- 210. Harvey, A.; Montezano, A.C.; Lopes, R.A.; Rios, F.; Touyz, R.M. Vascular Fibrosis in Aging and Hypertension: Molecular Mechanisms and Clinical Implications. *Can. J. Cardiol.* **2016**, *32*, 659–668. [\[CrossRef\]](https://doi.org/10.1016/j.cjca.2016.02.070) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/27118293)
- 211. O'Riordan, E.; Mendelev, N.; Patschan, S.; Patschan, D.; Eskander, J.; Cohen-Gould, L.; Chander, P.; Goligorsky, M.S. Chronic NOS inhibition actuates endothelial-mesenchymal transformation. *Am. J. Physiol. Heart Circ. Physiol.* **2007**, *292*, H285–H294. [\[CrossRef\]](https://doi.org/10.1152/ajpheart.00560.2006)
- 212. Tschöpe, C.; Van Linthout, S. New insights in (inter)cellular mechanisms by heart failure with preserved ejection fraction. *Curr. Heart Fail. Rep.* **2014**, *11*, 436–444. [\[CrossRef\]](https://doi.org/10.1007/s11897-014-0219-3)
- 213. Samady, H.; Eshtehardi, P.; McDaniel, M.C.; Suo, J.; Dhawan, S.S.; Maynard, C.; Timmins, L.H.; Quyyumi, A.A.; Giddens, D.P. Coronary artery wall shear stress is associated with progression and transformation of atherosclerotic plaque and arterial remodeling in patients with coronary artery disease. *Circulation* **2011**, *124*, 779–788. [\[CrossRef\]](https://doi.org/10.1161/CIRCULATIONAHA.111.021824)
- 214. Lerman, A.; Holmes, D.R.; Herrmann, J.; Gersh, B.J. Microcirculatory dysfunction in ST-elevation myocardial infarction: Cause, consequence, or both? *Eur. Heart J.* **2007**, *28*, 788–797. [\[CrossRef\]](https://doi.org/10.1093/eurheartj/ehl501)
- 215. Martínez Pereyra, V.; Hubert, A.; Seitz, A.; Bekeredjian, R.; Sechtem, U.; Ong, P. Epicardial and microvascular coronary spasm in the same patient?-acetylcholine testing pointing towards a common pathophysiological background. *Coron. Artery Dis.* **2020**, *31*, 398–399. [\[CrossRef\]](https://doi.org/10.1097/MCA.0000000000000829) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/31658150)
- 216. Egashira, K.; Inou, T.; Yamada, A.; Hirooka, Y.; Takeshita, A. Preserved endothelium-dependent vasodilation at the vasospastic site in patients with variant angina. *J. Clin. Investig.* **1992**, *89*, 1047–1052. [\[CrossRef\]](https://doi.org/10.1172/JCI115646) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/1371774)
- 217. Ito, K.; Akita, H.; Kanazawa, K.; Yamada, S.; Shiga, N.; Terashima, M.; Matsuda, Y.; Takai, E.; Iwai, C.; Takaoka, H.; et al. Systemic endothelial function is preserved in men with both active and inactive variant angina pectoris. *Am. J. Cardiol.* **1999**, *84*, 1347–1349. [\[CrossRef\]](https://doi.org/10.1016/S0002-9149(99)00571-8)
- 218. Casas, J.P.; Cavalleri, G.L.; Bautista, L.E.; Smeeth, L.; Humphries, S.E.; Hingorani, A.D. Endothelial nitric oxide synthase gene polymorphisms and cardiovascular disease: A HuGE review. *Am. J. Epidemiol.* **2006**, *164*, 921–935. [\[CrossRef\]](https://doi.org/10.1093/aje/kwj302) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/17018701)
- 219. Lanza, G.A.; Careri, G.; Crea, F. Mechanisms of coronary artery spasm. *Circulation* **2011**, *124*, 1774–1782. [\[CrossRef\]](https://doi.org/10.1161/CIRCULATIONAHA.111.037283)
- 220. Factor, S.M.; Minase, T.; Cho, S.; Dominitz, R.; Sonnenblick, E.H. Microvascular spasm in the cardiomyopathic Syrian hamster: A preventable cause of focal myocardial necrosis. *Circulation* **1982**, *66*, 342–354. [\[CrossRef\]](https://doi.org/10.1161/01.CIR.66.2.342)
- 221. Gaasch, W.H.; Adyanthaya, A.V.; Wang, V.H.; Pickering, E.; Quinones, M.A.; Alexander, J.K. Prinzmetal's variant angina: Hemodynamic and angiographic observations during pain. *Am. J. Cardiol.* **1975**, *35*, 683–690. [\[CrossRef\]](https://doi.org/10.1016/0002-9149(75)90057-0)
- 222. Awad, H.H.; McNeal, A.R.; Goyal, H. Reverse Takotsubo cardiomyopathy: A comprehensive review. *Ann. Transl. Med.* **2018**, *6*, 460. [\[CrossRef\]](https://doi.org/10.21037/atm.2018.11.08)
- 223. Greco, C.A.; De Rito, V.; Petracca, M.; Garzya, M.; Donateo, M.; Magliari, F. Takotsubo syndrome in a newborn. *J. Am. Soc. Echocardiogr.* **2011**, *24*, 471.e5–471.e7. [\[CrossRef\]](https://doi.org/10.1016/j.echo.2010.08.002) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/20810241)
- 224. Fan, X.; Yang, G.; Kowitz, J.; Akin, I.; Zhou, X.; El-Battrawy, I. Takotsubo Syndrome: Translational Implications and Pathomechanisms. *Int. J. Mol Sci.* **2022**, *23*, 1951. [\[CrossRef\]](https://doi.org/10.3390/ijms23041951)
- 225. Adameova, A.; Abdellatif, Y.; Dhalla, N.S. Role of the excessive amounts of circulating catecholamines and glucocorticoids in stress-induced heart disease. *Can. J. Physiol. Pharmacol.* **2009**, *87*, 493–514. [\[CrossRef\]](https://doi.org/10.1139/Y09-042)
- 226. Zhou, S.; Paz, O.; Cao, J.M.; Asotra, K.; Chai, N.N.; Wang, C.; Chen, L.S.; Fishbein, M.C.; Sharifi, B.; Chen, P.S. Differential beta-adrenoceptor expression induced by nerve growth factor infusion into the canine right and left stellate ganglia. *Heart Rhythm.* **2005**, *2*, 1347–1355. [\[CrossRef\]](https://doi.org/10.1016/j.hrthm.2005.08.027)
- 227. Tsuchihashi, K.; Ueshima, K.; Uchida, T.; Oh-mura, N.; Kimura, K.; Owa, M.; Yoshiyama, M.; Miyazaki, S.; Haze, K.; Ogawa, H.; et al. Transient left ventricular apical ballooning without coronary artery stenosis: A novel heart syndrome mimicking acute myocardial infarction. Angina Pectoris-Myocardial Infarction Investigations in Japan. *J. Am. Coll. Cardiol.* **2001**, *38*, 11–18. [\[CrossRef\]](https://doi.org/10.1016/S0735-1097(01)01316-X) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/11451258)
- 228. Angelini, P. Transient left ventricular apical ballooning: A unifying pathophysiologic theory at the edge of Prinzmetal angina. *Catheter. Cardiovasc. Interv.* **2008**, *71*, 342–352. [\[CrossRef\]](https://doi.org/10.1002/ccd.21338)
- 229. Jin, Y.; Li, Q.; Guo, X. Alternate recurrent coronary artery spasm and stress cardiomyopathy: A case report. *BMC Cardiovasc. Disord.* **2020**, *20*, 476. [\[CrossRef\]](https://doi.org/10.1186/s12872-020-01760-2)
- 230. Gisterå, A.; Ketelhuth, D.F.J.; Malin, S.G.; Hansson, G.K. Animal Models of Atherosclerosis-Supportive Notes and Tricks of the Trade. *Circ. Res.* **2022**, *130*, 1869–1887. [\[CrossRef\]](https://doi.org/10.1161/CIRCRESAHA.122.320263)
- 231. Borchert, T.; Hübscher, D.; Guessoum, C.I.; Lam, T.D.; Ghadri, J.R.; Schellinger, I.N.; Tiburcy, M.; Liaw, N.Y.; Li, Y.; Haas, J.; et al. Catecholamine-Dependent β-Adrenergic Signaling in a Pluripotent Stem Cell Model of Takotsubo Cardiomyopathy. *J. Am. Coll. Cardiol.* **2017**, *70*, 975–991. [\[CrossRef\]](https://doi.org/10.1016/j.jacc.2017.06.061)
- 232. Hung, M.Y.; Wu, Y.H.; Bamodu, O.A.; Chen, X.; Lin, Y.K.; Hu, P.; Chang, N.C.; Pang, J.S.; Yeh, C.T. Activation of the monocytic α7 nicotinic acetylcholine receptor modulates oxidative stress and inflammation-associated development of coronary artery spasm via a p38 MAP-kinase signaling-dependent pathway. *Free Radic. Biol. Med.* **2018**, *120*, 266–276. [\[CrossRef\]](https://doi.org/10.1016/j.freeradbiomed.2018.03.050) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/29609021)
- 233. Lin, Y.K.; Yeh, C.T.; Kuo, K.T.; Fong, I.H.; Yadav, V.K.; Kounis, N.G.; Hu, P.; Hung, M.Y. Apolipoprotein (a)/Lipoprotein(a)- Induced Oxidative-Inflammatory α7-nAChR/p38 MAPK/IL-6/RhoA-GTP Signaling Axis and M1 Macrophage Polarization Modulate Inflammation-Associated Development of Coronary Artery Spasm. *Oxidative Med. Cell. Longev.* **2022**, *2022*, 9964689. [\[CrossRef\]](https://doi.org/10.1155/2022/9964689)
- 234. Henry, P.D.; Yokoyama, M. Supersensitivity of atherosclerotic rabbit aorta to ergonovine. Mediation by a serotonergic mechanism. *J. Clin. Investig.* **1980**, *66*, 306–313. [\[CrossRef\]](https://doi.org/10.1172/JCI109858)
- 235. Noguchi, K.; Tomoike, H.; Kawachi, Y.; Maruoka, Y.; Nakamura, M. Angiographic evaluation of the coronary asodilating effect of trapidil in anesthetized closed-chest dogs. *J. Cardiovasc. Pharmacol.* **1982**, *4*, 1049–1054. [\[CrossRef\]](https://doi.org/10.1097/00005344-198211000-00027)
- 236. Kawachi, Y.; Tomoike, H.; Maruoka, Y.; Kikuchi, Y.; Araki, H.; Ishii, Y.; Tanaka, K.; Nakamura, M. Selective hypercontraction caused by ergonovine in the canine coronary artery under conditions of induced atherosclerosis. *Circulation* **1984**, *69*, 441–450. [\[CrossRef\]](https://doi.org/10.1161/01.CIR.69.2.441)
- 237. Shimokawa, H.; Tomoike, H.; Nabeyama, S.; Yamamoto, H.; Araki, H.; Nakamura, M.; Ishii, Y.; Tanaka, K. Coronary artery spasm induced in atherosclerotic miniature swine. *Science* **1983**, *221*, 560–562. [\[CrossRef\]](https://doi.org/10.1126/science.6408736)
- 238. Egashira, K.; Tomoike, H.; Yamamoto, Y.; Yamada, A.; Hayashi, Y.; Nakamura, M. Histamine-induced coronary spasm in regions of intimal thickening in miniature pigs: Roles of serum cholesterol and spontaneous or induced intimal thickening. *Circulation* **1986**, *74*, 826–837. [\[CrossRef\]](https://doi.org/10.1161/01.CIR.74.4.826) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/3530524)
- 239. Shimokawa, H.; Ito, A.; Fukumoto, Y.; Kadokami, T.; Nakaike, R.; Sakata, M.; Takayanagi, T.; Egashira, K.; Takeshita, A. Chronic treatment with interleukin-1 beta induces coronary intimal lesions and vasospastic responses in pigs in vivo. The role of platelet-derived growth factor. *J. Clin. Investig.* **1996**, *97*, 769–776. [\[CrossRef\]](https://doi.org/10.1172/JCI118476) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/8609234)
- 240. Kozai, T.; Shimokawa, H.; Fukumoto, Y.; Kobayashi, S.; Owada, M.K.; Kadokami, T.; Ito, A.; Kuwata, K.; Egashira, K.; Shiraishi, T.; et al. Tyrosine kinase inhibitor markedly suppresses the development of coronary lesions induced by long-term treatment with platelet-derived growth factor in pigs in vivo. *J. Cardiovasc. Pharmacol.* **1997**, *29*, 536–545. [\[CrossRef\]](https://doi.org/10.1097/00005344-199704000-00016) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/9156365)
- 241. Pearson, J.T.; Jenkins, M.J.; Edgley, A.J.; Sonobe, T.; Joshi, M.; Waddingham, M.T.; Fujii, Y.; Schwenke, D.O.; Tsuchimochi, H.; Yoshimoto, M.; et al. Acute Rho-kinase inhibition improves coronary dysfunction in vivo, in the early diabetic microcirculation. *Cardiovasc. Diabetol.* **2013**, *12*, 111. [\[CrossRef\]](https://doi.org/10.1186/1475-2840-12-111) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/24059472)
- 242. Kinjo, T.; Tanaka, M.; Osanai, T.; Shibutani, S.; Narita, I.; Tanno, T.; Nishizaki, K.; Ichikawa, H.; Kimura, Y.; Ishida, Y.; et al. Enhanced p122RhoGAP/DLC-1 Expression Can Be a Cause of Coronary Spasm. *PLoS ONE* **2015**, *10*, e0143884. [\[CrossRef\]](https://doi.org/10.1371/journal.pone.0143884)
- 243. Shibutani, S.; Osanai, T.; Oya, H.; Sagara, S.; Izumiyama, K.; Yamamoto, Y.; Hanada, K.; Tomita, H.; Okumura, K. Mutation analysis ABCC9 gene in Japanese patients with coronary spastic angina. *Hirosaki Med. J.* **2011**, *62*, 27–33.
- 244. Malester, B.; Tong, X.; Ghiu, I.; Kontogeorgis, A.; Gutstein, D.E.; Xu, J.; Hendricks-Munoz, K.D.; Coetzee, W.A. Transgenic expression of a dominant negative K(ATP) channel subunit in the mouse endothelium: Effects on coronary flow and endothelin-1 secretion. *FASEB J.* **2007**, *21*, 2162–2172. [\[CrossRef\]](https://doi.org/10.1096/fj.06-7821com) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/17341678)
- 245. Tomita, H.; Sasaki, S.; Osanai, T.; Nakano, T.; Higuma, T.; Yokoyama, J.; Hanada, H.; Yasujima, M.; Okumura, K. Mutational analysis of Kir6.1 in Japanese patients with coronary spastic angina. *Int. J. Mol. Med.* **2006**, *18*, 589–591. [\[CrossRef\]](https://doi.org/10.3892/ijmm.18.4.589)
- 246. Emanuele, E.; Falcone, C.; Carabela, M.; Minoretti, P.; D'Angelo, A.; Montagna, L.; Geroldi, D. Absence of Kir6.1/KCNJ8 mutations in Italian patients with abnormal coronary vasomotion. *Int. J. Mol. Med.* **2003**, *12*, 509–512. [\[CrossRef\]](https://doi.org/10.3892/ijmm.12.4.509) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/12964027)
- 247. Shibutani, S.; Osanai, T.; Ashitate, T.; Sagara, S.; Izumiyama, K.; Yamamoto, Y.; Hanada, K.; Echizen, T.; Tomita, H.; Fujita, T.; et al. Coronary vasospasm induced in transgenic mouse with increased phospholipase C-δ1 activity. *Circulation* **2012**, *125*, 1027–1036. [\[CrossRef\]](https://doi.org/10.1161/CIRCULATIONAHA.111.064303)
- 248. Yamada, S.; Saitoh, S.; Machii, H.; Mizukami, H.; Hoshino, Y.; Misaka, T.; Ishigami, A.; Takeishi, Y. Coronary artery spasm related to thiol oxidation and senescence marker protein-30 in aging. *Antioxid. Redox Signal.* **2013**, *19*, 1063–1073. [\[CrossRef\]](https://doi.org/10.1089/ars.2012.4903) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/23320823)
- 249. Gevaert, A.B.; Kataria, R.; Zannad, F.; Sauer, A.J.; Damman, K.; Sharma, K.; Shah, S.J.; Van Spall, H.G.C. Heart failure with preserved ejection fraction: Recent concepts in diagnosis, mechanisms and management. *Heart* **2022**, *108*, 1342–1350. [\[CrossRef\]](https://doi.org/10.1136/heartjnl-2021-319605)
- 250. Figulla, H.R.; Vetterlein, F.; Glaubitz, M.; Kreuzer, H. Inhomogenous capillary flow and its prevention by verapamil and hydralazine in the cardiomyopathic Syrian hamster. *Circulation* **1987**, *76*, 208–216. [\[CrossRef\]](https://doi.org/10.1161/01.CIR.76.1.208)
- 251. Figulla, H.R.; Rechenberg, J.V.; Wiegand, V.; Soballa, R.; Kreuzer, H. Beneficial effects of long-term diltiazem treatment in dilated cardiomyopathy. *J. Am. Coll. Cardiol.* **1989**, *13*, 653–658. [\[CrossRef\]](https://doi.org/10.1016/0735-1097(89)90607-4) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/2918172)
- 252. Multicenter Diltiazem Postinfarction Trial Research Group. The effect of diltiazem on mortality and reinfarction after myocardial infarction. *N. Engl. J. Med.* **1988**, *319*, 385–392. [\[CrossRef\]](https://doi.org/10.1056/NEJM198808183190701) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/2899840)
- 253. Yancy, C.W.; Jessup, M.; Bozkurt, B.; Butler, J.; Casey, D.E., Jr.; Drazner, M.H.; Fonarow, G.C.; Geraci, S.A.; Horwich, T.; Januzzi, J.L.; et al. 2013 ACCF/AHA guideline for the management of heart failure: A report of the American College of Cardiology Foundation/American Heart Association Task Force on practice guidelines. *Circulation* **2013**, *128*, e240–e327. [\[CrossRef\]](https://doi.org/10.1161/CIR.0b013e31829e8776) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/23741058)
- 254. Masumoto, A.; Mohri, M.; Shimokawa, H.; Urakami, L.; Usui, M.; Takeshita, A. Suppression of coronary artery spasm by the Rho-kinase inhibitor fasudil in patients with vasospastic angina. *Circulation* **2002**, *105*, 1545–1547. [\[CrossRef\]](https://doi.org/10.1161/hc1002.105938) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/11927519)
- 255. Martínez Pereyra, V.; Seitz, A.; Hubert, A.; Beck, S.; Hofmann, U.; Schwab, M.; Bekeredjian, R.; Sechtem, U.; Ong, P. Repurposing Riociguat for Treatment of Refractory Angina Resulting From Coronary Spasm. *JACC Case Rep.* **2021**, *3*, 392–396. [\[CrossRef\]](https://doi.org/10.1016/j.jaccas.2020.11.043) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/34317544)
- 256. Liu, C.; Ho, P.C.; Wong, F.C.; Sethi, G.; Wang, L.Z.; Goh, B.C. Garcinol: Current status of its anti-oxidative, anti-inflammatory and anti-cancer effects. *Cancer Lett.* **2015**, *362*, 8–14. [\[CrossRef\]](https://doi.org/10.1016/j.canlet.2015.03.019) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/25796441)
- 257. Li, M.; Li, X.; Yang, L. Cardioprotective effects of garcinol following myocardial infarction in rats with isoproterenol-induced heart failure. *AMB Express* **2020**, *10*, 137. [\[CrossRef\]](https://doi.org/10.1186/s13568-020-01065-9)
- 258. Chang, N.C.; Yeh, C.T.; Lin, Y.K.; Kuo, K.T.; Fong, I.H.; Kounis, N.G.; Hu, P.; Hung, M.Y. Garcinol Attenuates Lipoprotein(a)- Induced Oxidative Stress and Inflammatory Cytokine Production in Ventricular Cardiomyocyte through α7-Nicotinic Acetylcholine Receptor-Mediated Inhibition of the p38 MAPK and NF-κB Signaling Pathways. *Antioxidants* **2021**, *10*, 461. [\[CrossRef\]](https://doi.org/10.3390/antiox10030461)
- 259. Murphy, S.P.; Kakkar, R.; McCarthy, C.P.; Januzzi, J.L., Jr. Inflammation in Heart Failure: JACC State-of-the-Art Review. *J. Am. Coll. Cardiol.* **2020**, *75*, 1324–1340. [\[CrossRef\]](https://doi.org/10.1016/j.jacc.2020.01.014)
- 260. Everett, B.M.; Cornel, J.H.; Lainscak, M.; Anker, S.D.; Abbate, A.; Thuren, T.; Libby, P.; Glynn, R.J.; Ridker, P.M. Anti-Inflammatory Therapy With Canakinumab for the Prevention of Hospitalization for Heart Failure. *Circulation* **2019**, *139*, 1289–1299. [\[CrossRef\]](https://doi.org/10.1161/CIRCULATIONAHA.118.038010)
- 261. Yu, L.; Ruifrok, W.P.; Meissner, M.; Bos, E.M.; van Goor, H.; Sanjabi, B.; van der Harst, P.; Pitt, B.; Goldstein, I.J.; Koerts, J.A.; et al. Genetic and pharmacological inhibition of galectin-3 prevents cardiac remodeling by interfering with myocardial fibrogenesis. *Circ. Heart Fail.* **2013**, *6*, 107–117. [\[CrossRef\]](https://doi.org/10.1161/CIRCHEARTFAILURE.112.971168)
- 262. Calvier, L.; Martinez-Martinez, E.; Miana, M.; Cachofeiro, V.; Rousseau, E.; Sádaba, J.R.; Zannad, F.; Rossignol, P.; López-Andrés, N. The impact of galectin-3 inhibition on aldosterone-induced cardiac and renal injuries. *JACC Heart Fail.* **2015**, *3*, 59–67. [\[CrossRef\]](https://doi.org/10.1016/j.jchf.2014.08.002)
- 263. Varga, Z.; Sabzwari, S.R.A.; Vargova, V. Cardiovascular Risk of Nonsteroidal Anti-Inflammatory Drugs: An Under-Recognized Public Health Issue. *Cureus* **2017**, *9*, e1144. [\[CrossRef\]](https://doi.org/10.7759/cureus.1144) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/28491485)
- 264. Arfè, A.; Scotti, L.; Varas-Lorenzo, C.; Nicotra, F.; Zambon, A.; Kollhorst, B.; Schink, T.; Garbe, E.; Herings, R.; Straatman, H.; et al. Non-steroidal anti-inflammatory drugs and risk of heart failure in four European countries: Nested case-control study. *BMJ* **2016**, *354*, i4857. [\[CrossRef\]](https://doi.org/10.1136/bmj.i4857) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/27682515)
- 265. Fildes, J.E.; Shaw, S.M.; Yonan, N.; Williams, S.G. The immune system and chronic heart failure: Is the heart in control? *J. Am. Coll. Cardiol.* **2009**, *53*, 1013–1020. [\[CrossRef\]](https://doi.org/10.1016/j.jacc.2008.11.046)
- 266. Levine, G.N.; Cohen, B.E.; Commodore-Mensah, Y.; Fleury, J.; Huffman, J.C.; Khalid, U.; Labarthe, D.R.; Lavretsky, H.; Michos, E.D.; Spatz, E.S.; et al. Psychological Health, Well-Being, and the Mind-Heart-Body Connection: A Scientific Statement From the American Heart Association. *Circulation* **2021**, *143*, e763–e783. [\[CrossRef\]](https://doi.org/10.1161/CIR.0000000000000947) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/33486973)

Disclaimer/Publisher's Note: The statements, opinions and data contained in all publications are solely those of the individual author(s) and contributor(s) and not of MDPI and/or the editor(s). MDPI and/or the editor(s) disclaim responsibility for any injury to people or property resulting from any ideas, methods, instructions or products referred to in the content.