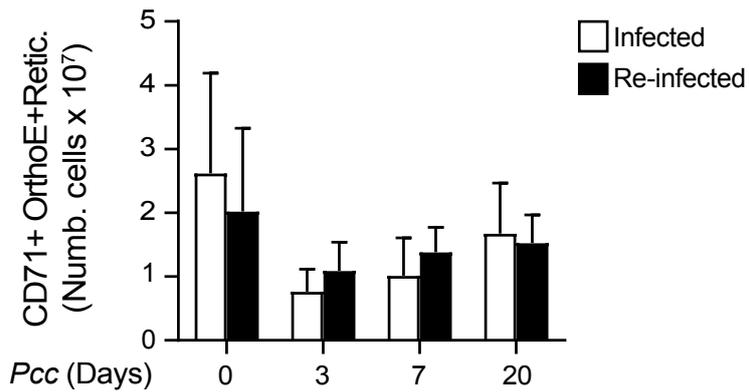
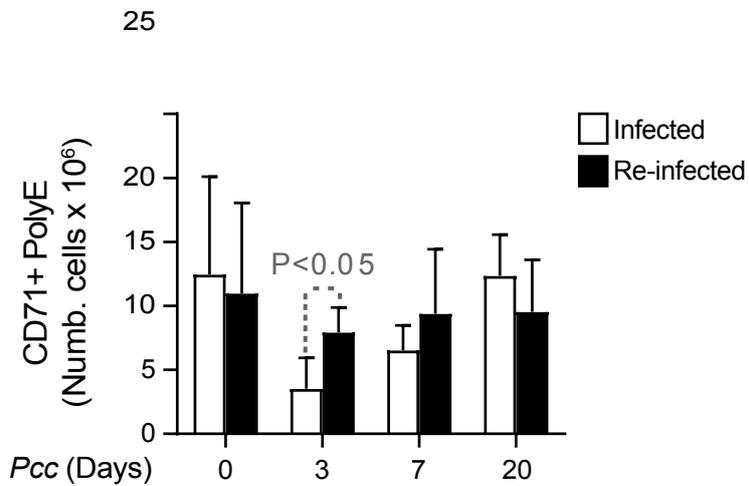
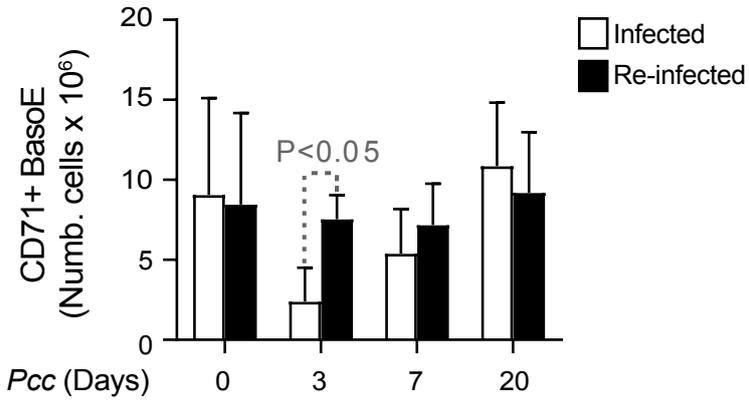
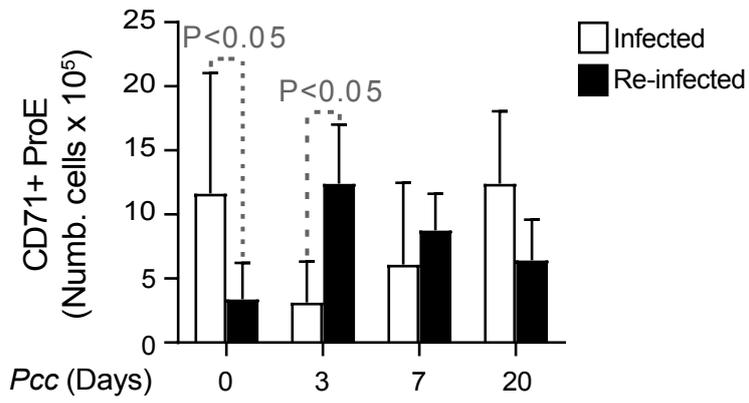
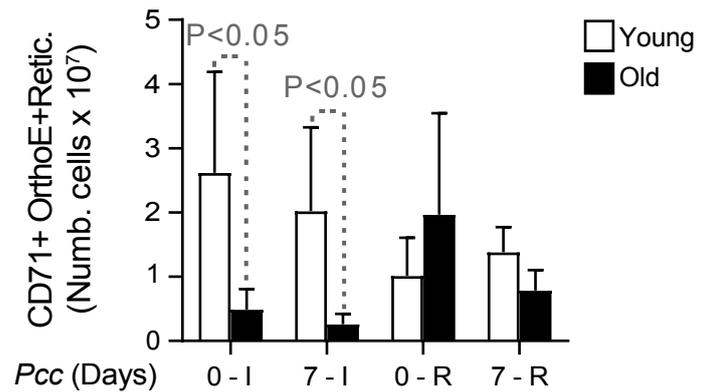
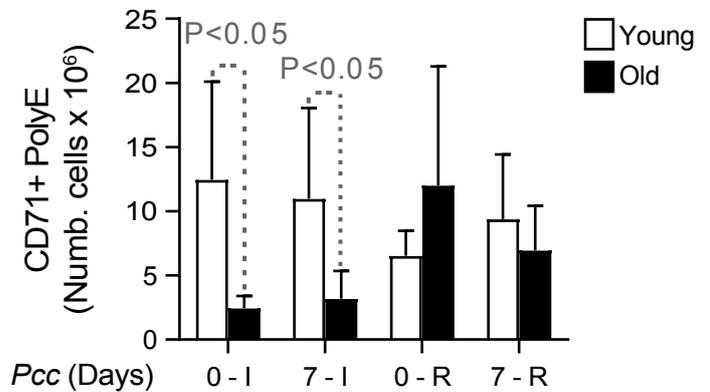
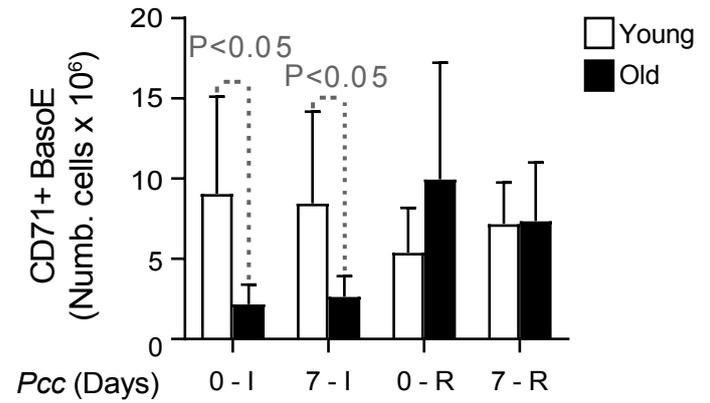
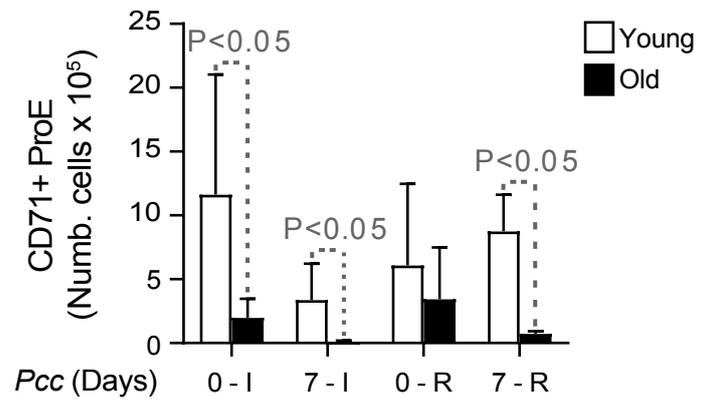
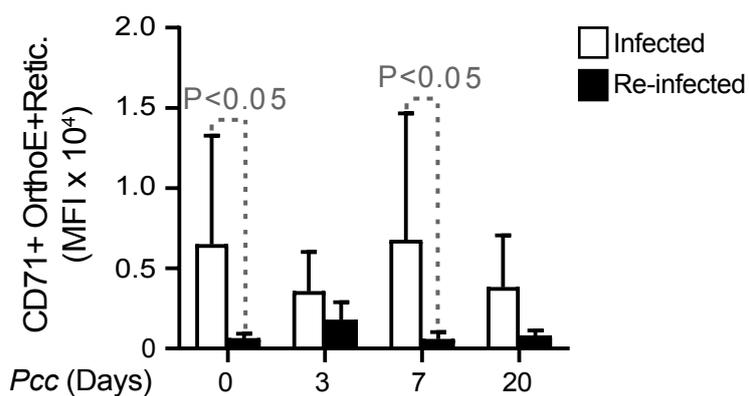
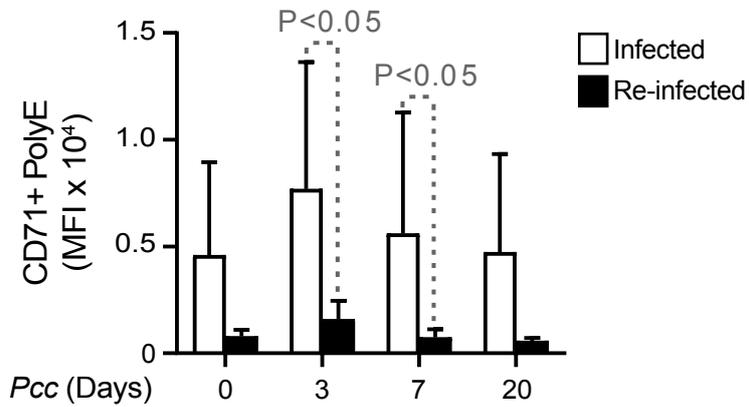
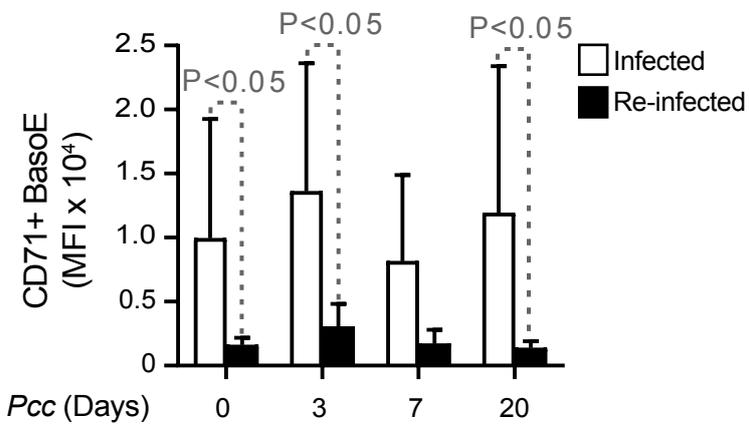
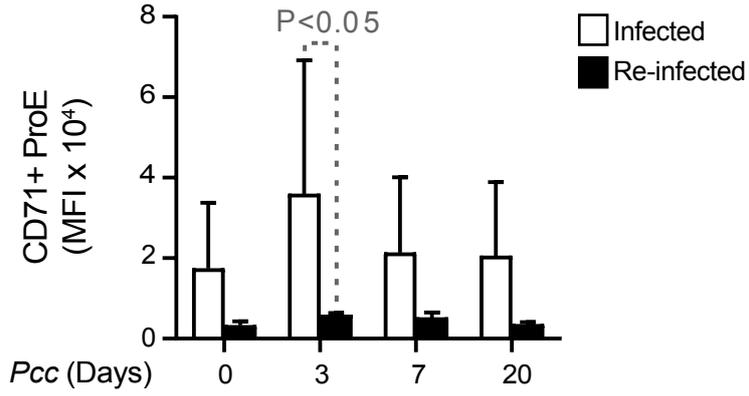
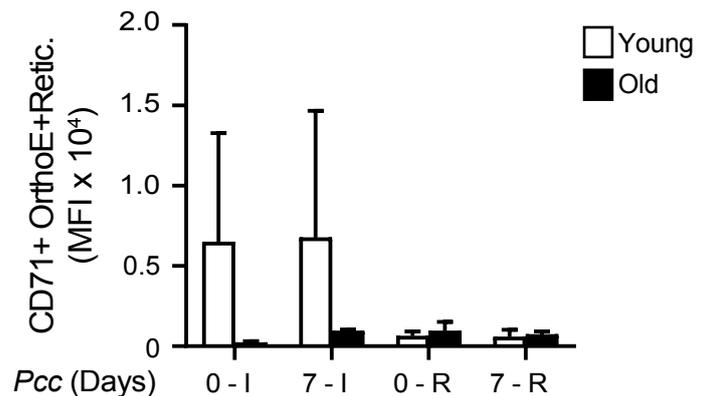
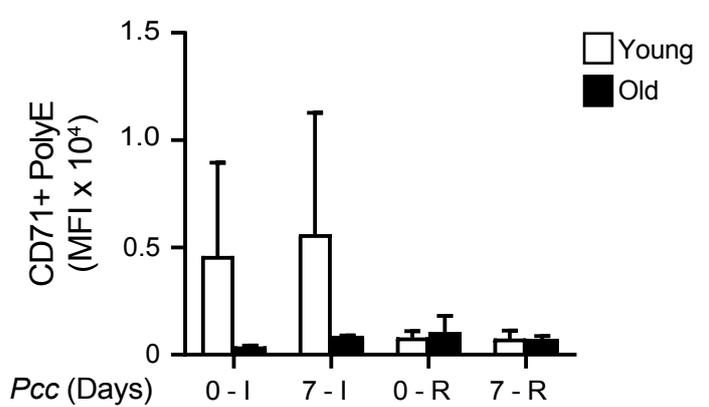
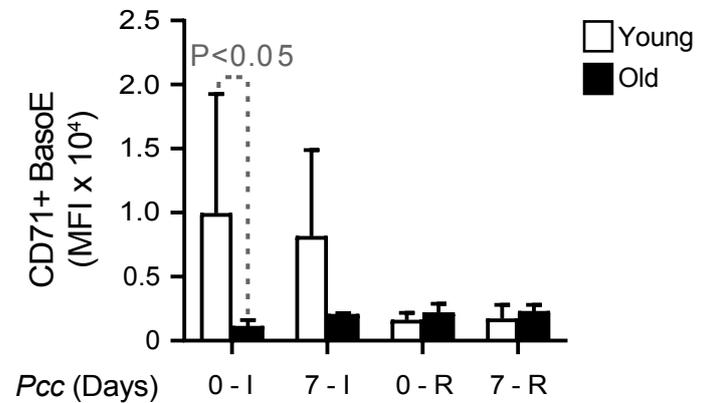
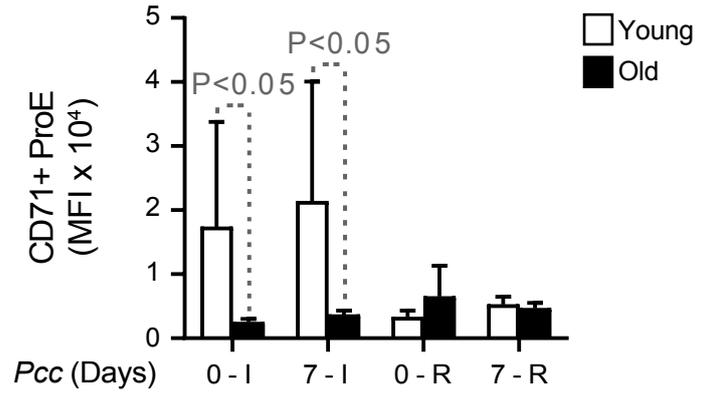
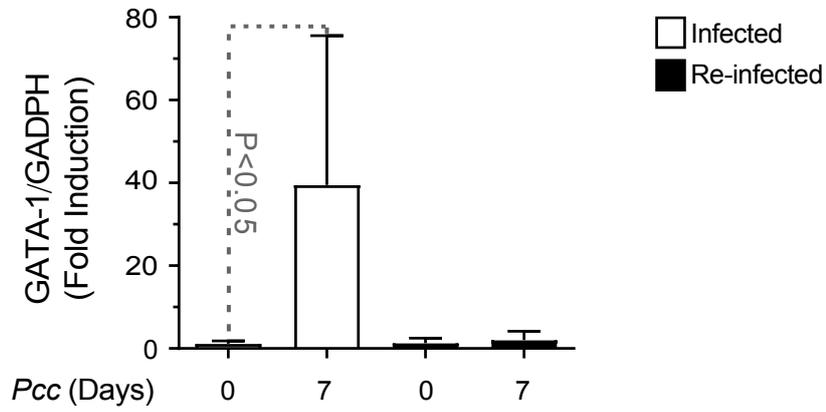
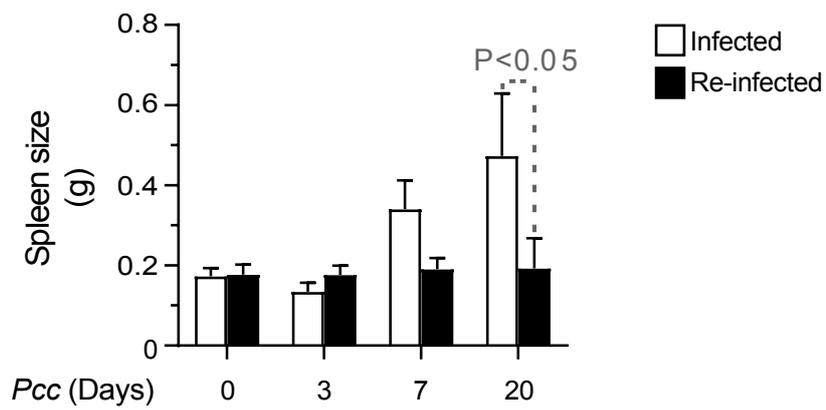
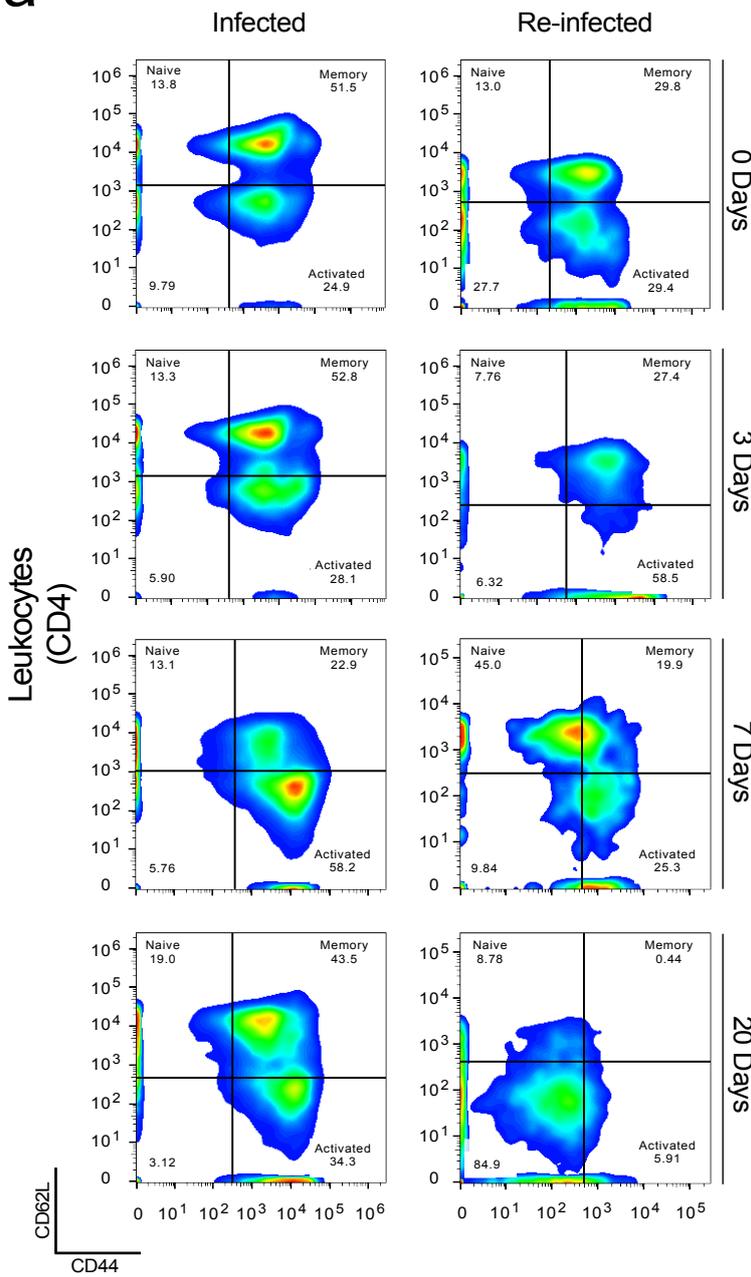
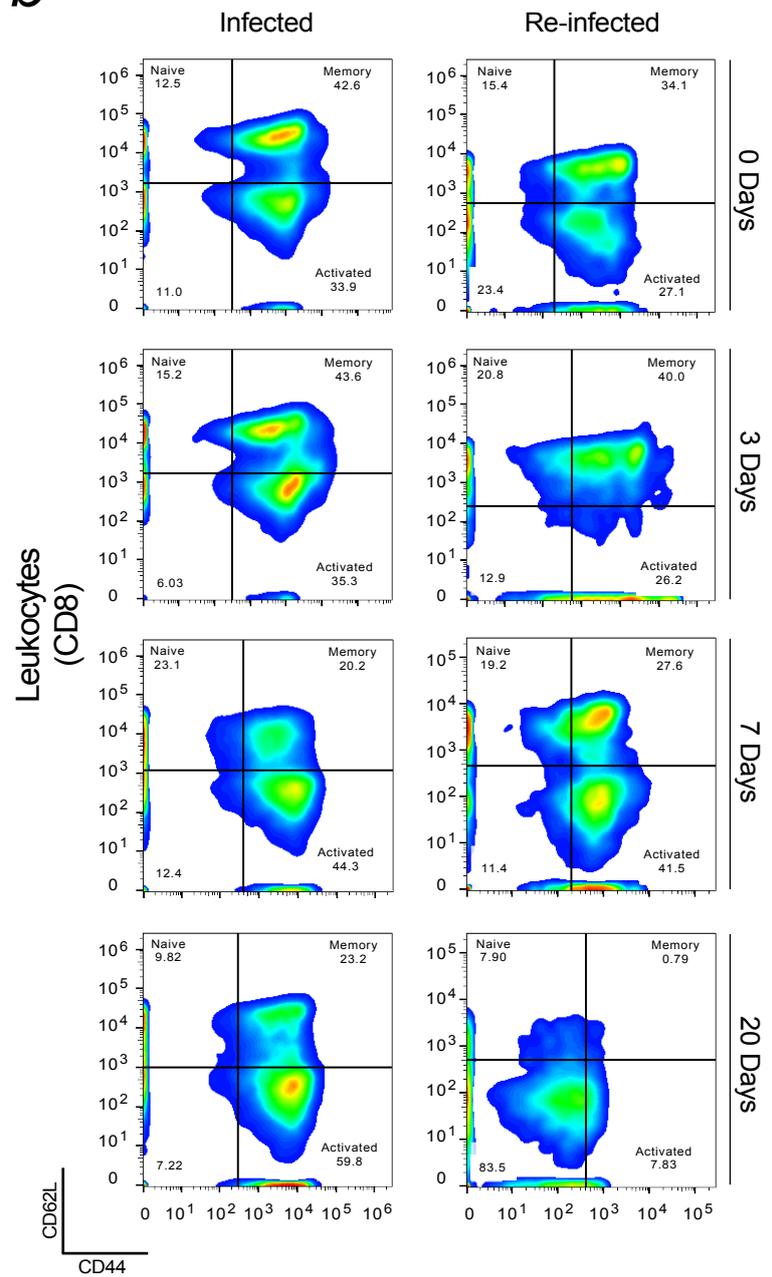
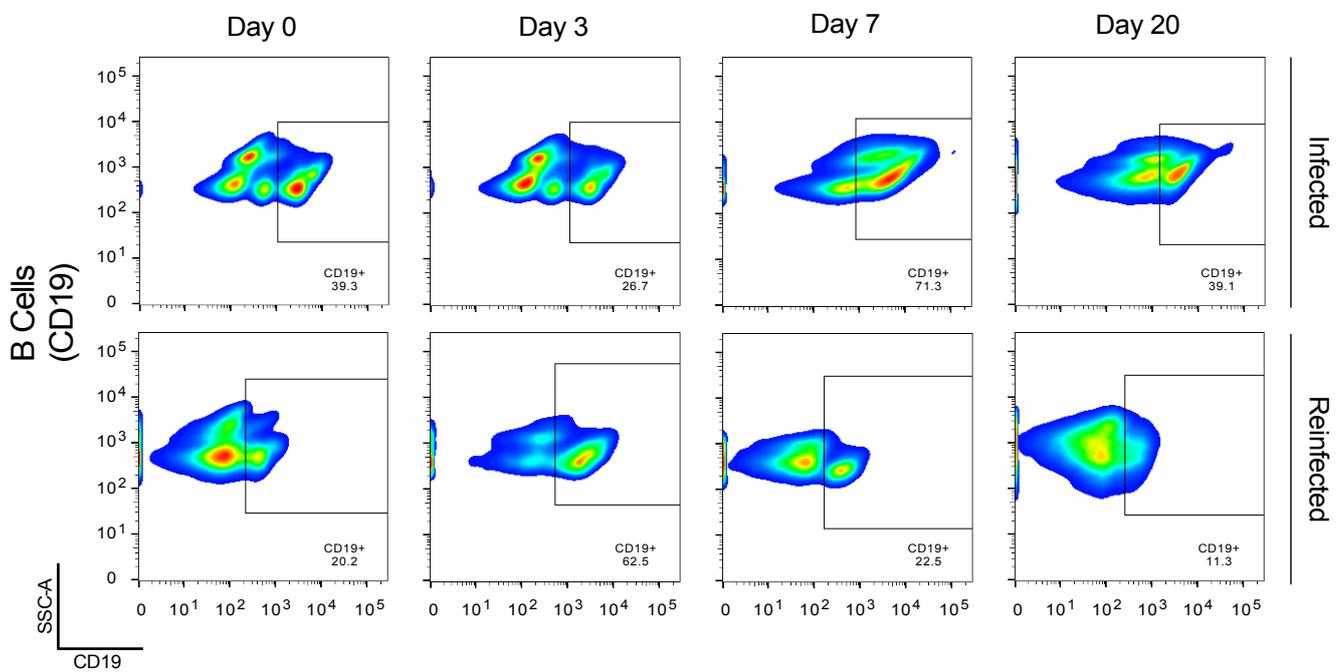
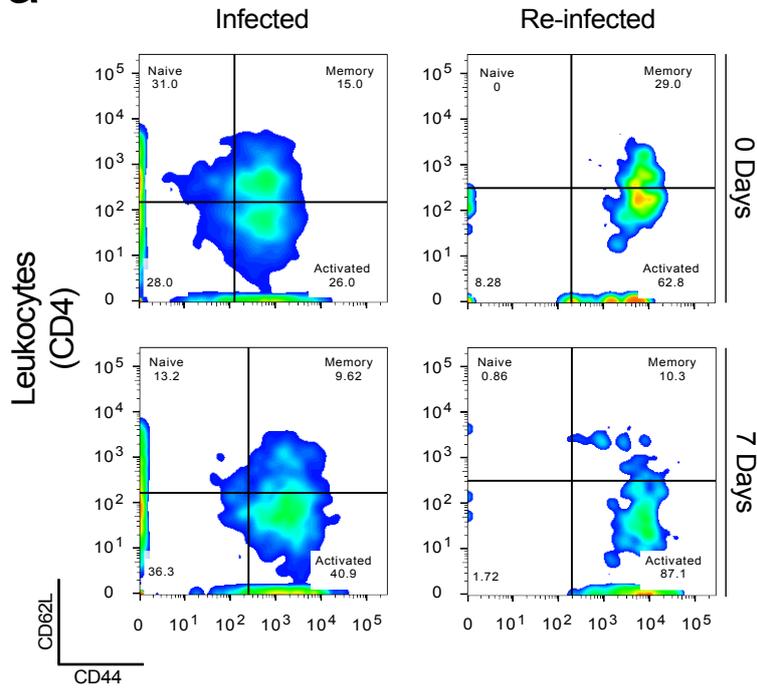
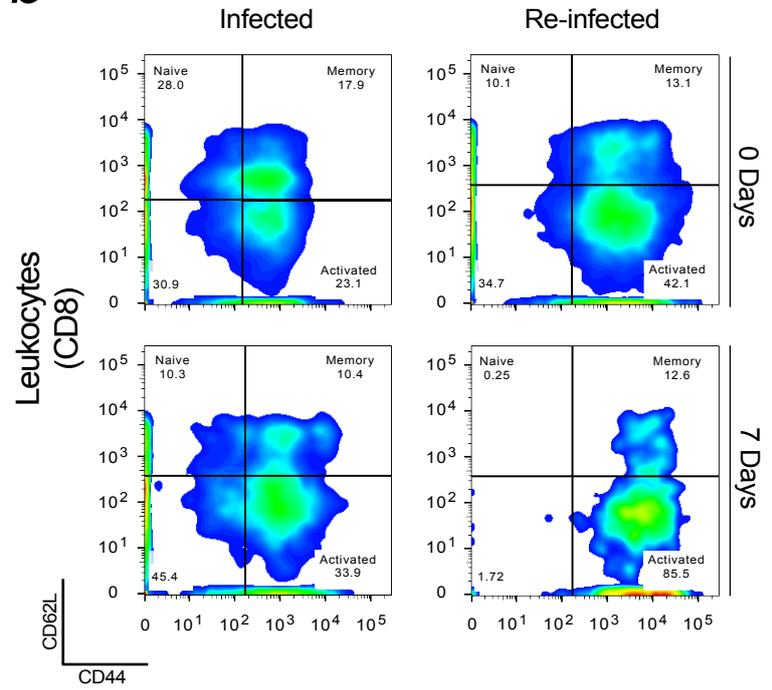
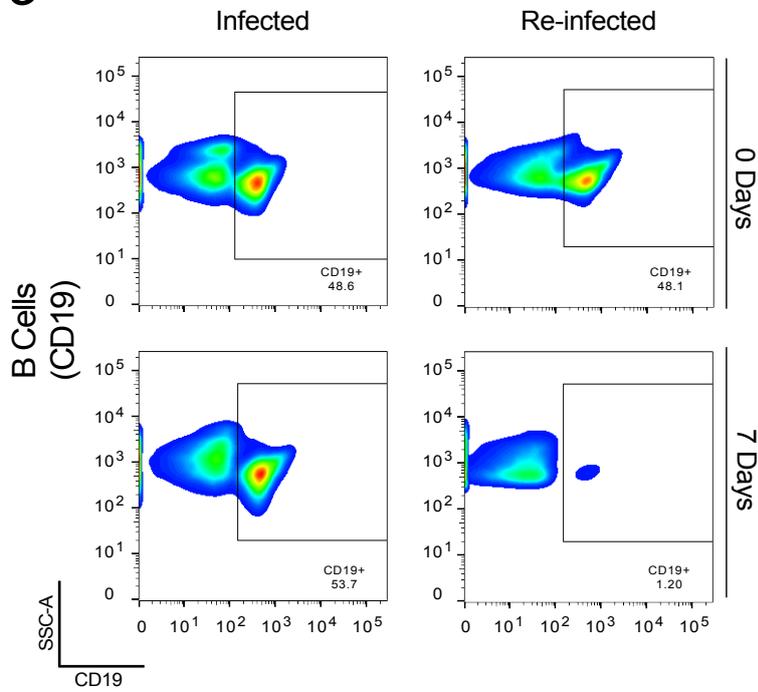


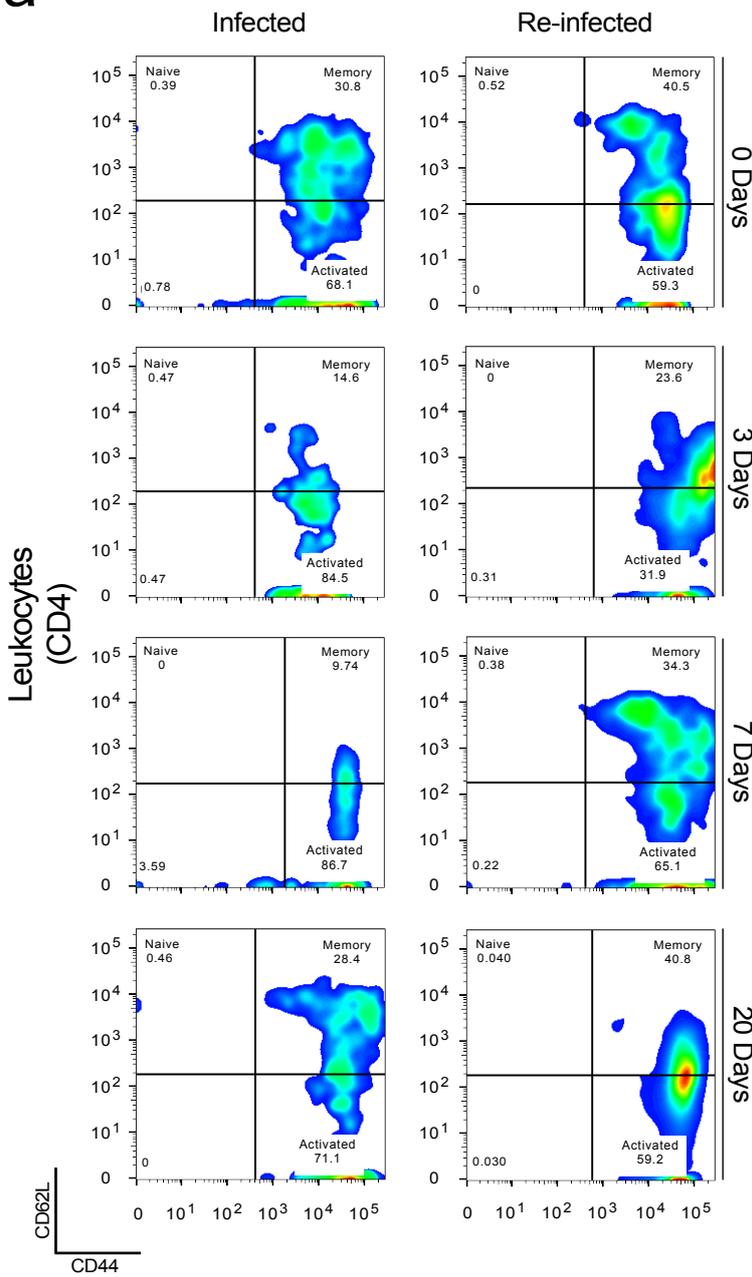
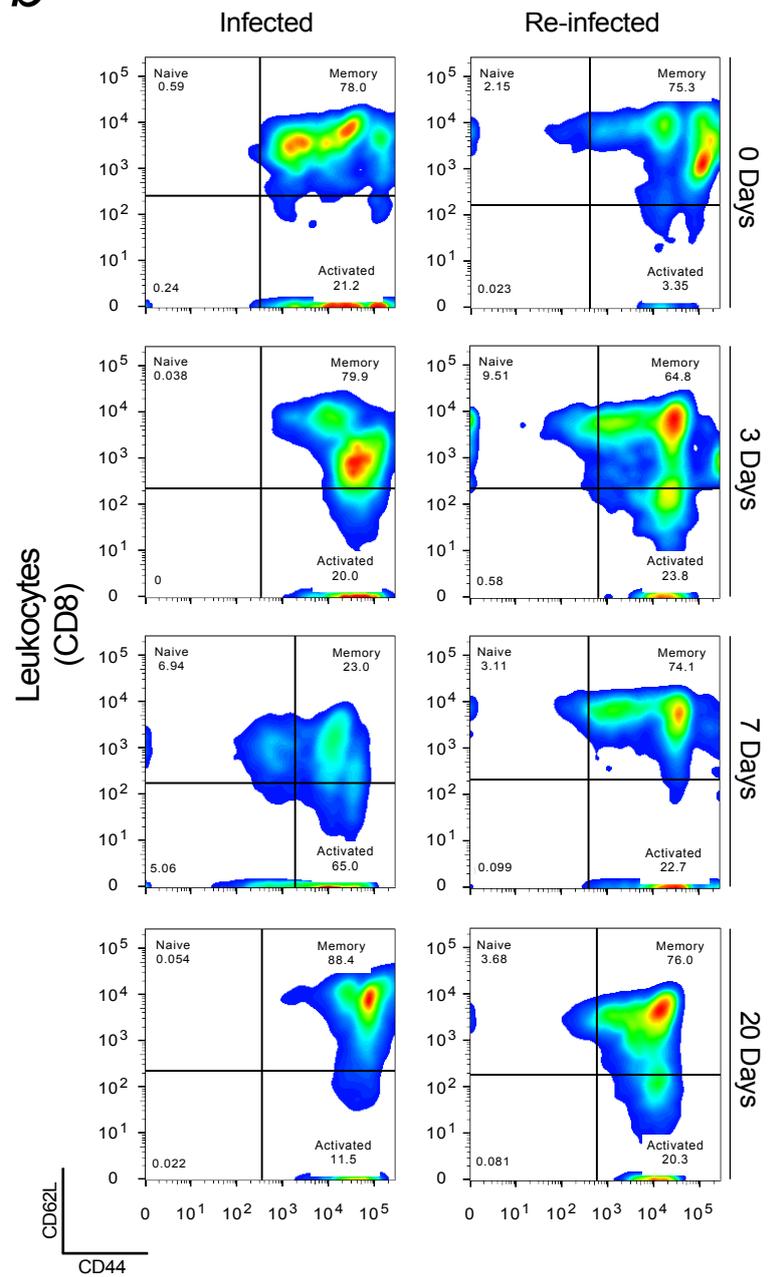
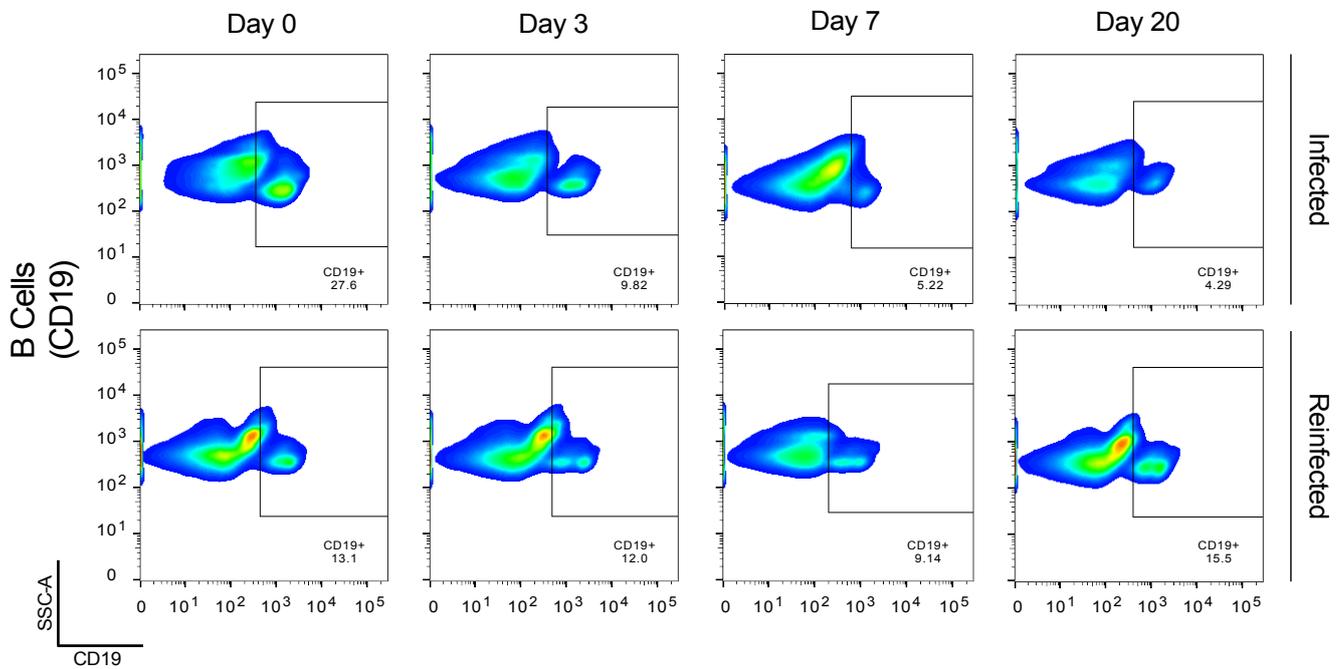
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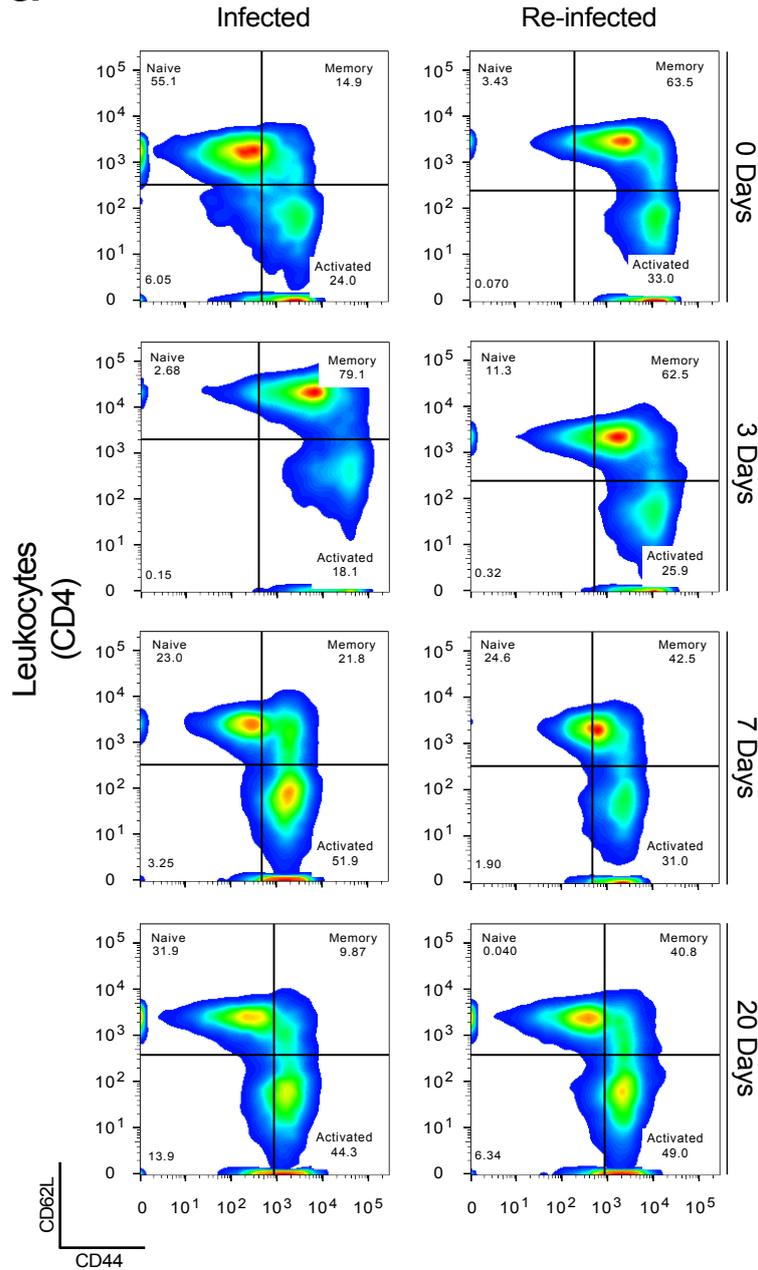
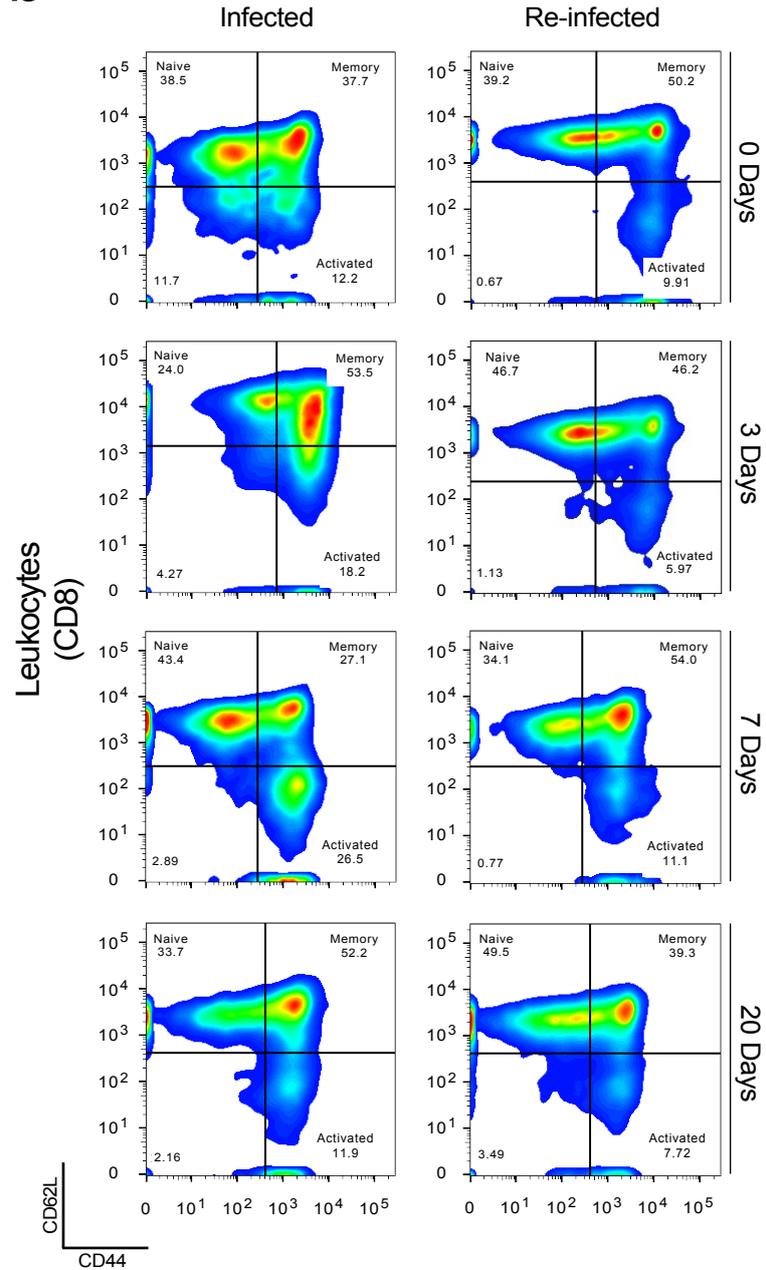
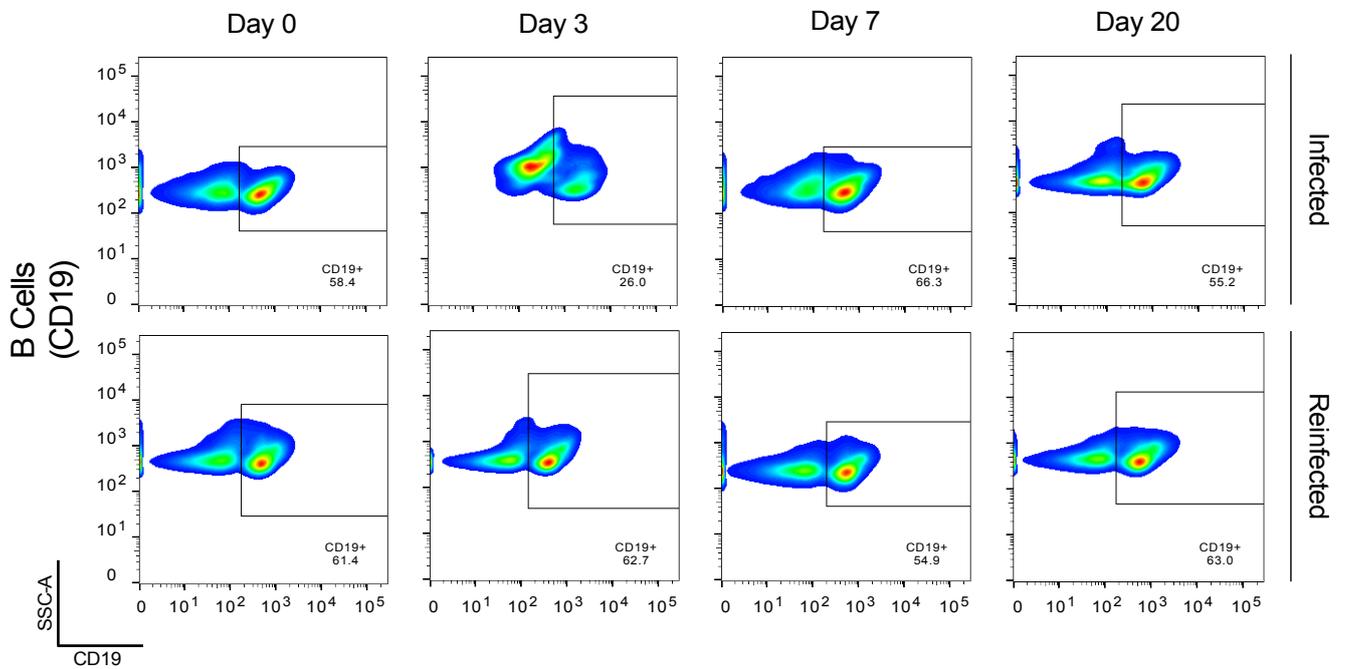
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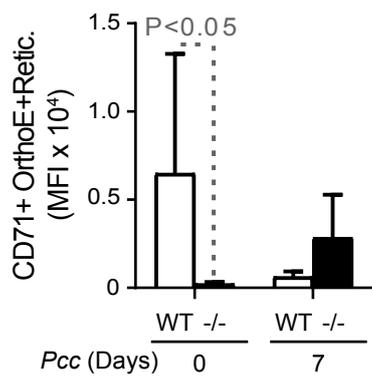
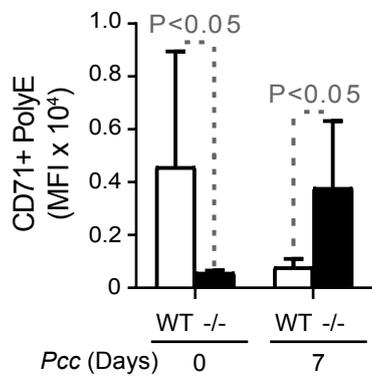
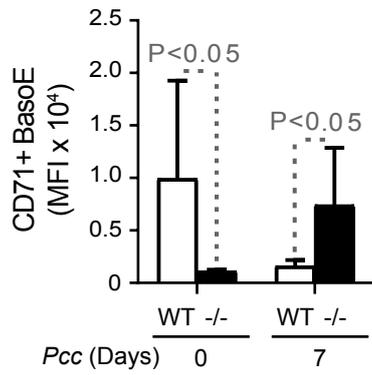
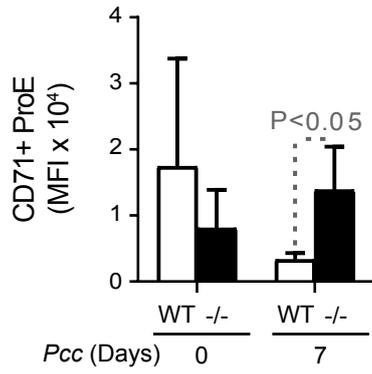
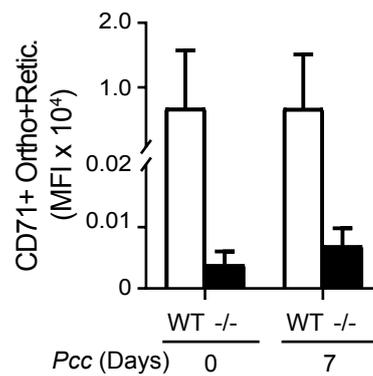
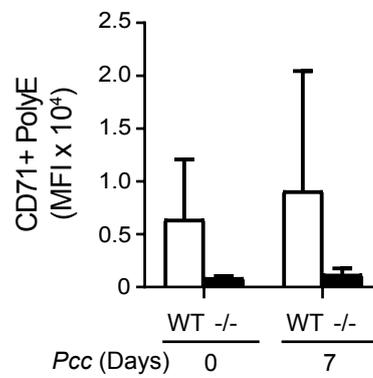
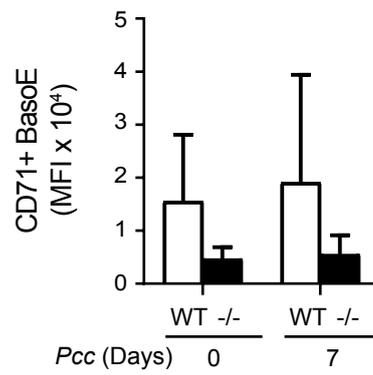
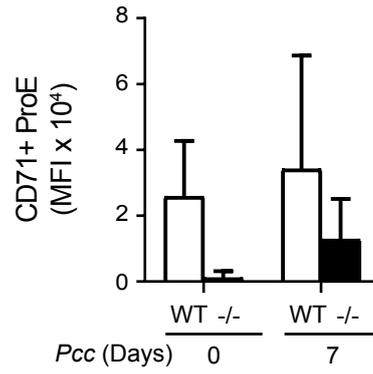
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Supplementary Figure legends

Suppl. Fig. S1 – CD71 assessment in erythroid populations of *Pcc* Infected and Reinfected young vs. aged mice.

Flow cytometry was used to study the abundance of CD71 in erythroid cells in bone marrow of *Pcc* infected and reinfected mice at days 3, 7, and 20. The non-manipulated group was compared to recovered animals (R, day 60 post-first infection). (a) Histogram representations of the number of CD71+ cells in erythroid proerythroblast (ProE), basophilic (BasoE), polychromatic (PolyE), and orthochromatic and reticulocytes (OrthoE+Retic), collected from the bone marrow of young (8-12 weeks) mice. (b) Histogram representations of the number of CD71+ cells in erythroid proerythroblast (ProE), basophilic (BasoE), polychromatic (PolyE), and orthochromatic and reticulocytes (OrthoE+Retic), collected from the bone marrow of young (8-12 weeks) and old (52-60 weeks) infected and reinfected mice, at indicated time points. Data were presented as mean values \pm standard deviations (n=8-13). Statistically significant differences were calculated by one-way ANOVA and indicated accordingly.

Suppl. Fig. S2 – CD71 assessment in erythroid populations of *Pcc* Infected and Reinfected young vs. aged mice.

Flow cytometry was used to study the median fluorescen intensity (MFI) of CD71 in erythroid cells in bone marrow of *Pcc* infected and reinfected mice at days 3, 7, and 20. The non-manipulated group was compared to recovered animals (R, day 60 post-infection). (a) Histogram representations of the number of CD71+ cells in erythroid proerythroblast (ProE), basophilic (BasoE), polychromatic (PolyE), and orthochromatic and reticulocytes (OrthoE+Retic), collected from the bone marrow of young (8-12 weeks) mice. (b) Histogram representations show the MFI of CD71+ cells in erythroid proerythroblast (ProE), basophilic (BasoE), polychromatic (PolyE), and orthochromatic and reticulocytes (OrthoE+Retic), collected from the spleen of young (8-12 weeks) and old (52-60 weeks) infected and reinfected mice, at indicated time points. Data were presented as mean values \pm standard deviations (n=8-13). Statistically significant differences were calculated by one-way ANOVA and indicated accordingly.

Suppl. Fig. S3 – Assessment of *GATA-1* expression in the spleen and organ size in infected and reinfected mice.

(a) *GATA-1* mRNA expression, quantified by qRT-PCR, in spleen samples of infected and reinfected mice, at days 0, 3, 7 and 20 post-*Pcc* inoculation. (b). Spleen size, measured in grams (g), upon harvesting from infected and reinfected mice, at indicated time points. Data were presented as mean values \pm standard deviations (n=8-13). Statistically significant differences were calculated by one-way ANOVA and indicated accordingly.

Suppl. Fig. S4 – T and B immune cell activation in the blood of infected and reinfected mice.

Gating strategy of freshly isolated blood immune cells, at days 0, 3, 7 and 20 post-*Pcc* inoculation. (a) CD4+; (b) CD8+ T cells. The activation of T cell subtypes was determined through the expression of CD44 and CD62L, allowing to identify naïve (CD44–CD62L+), effector or activated (CD44+CD62L–), and memory cells (CD44+CD62L+). (c) Circulating B cells (CD19+). Data are expressed as mean values \pm standard deviations (n=7-12). Statistically significant differences were calculated by one-way ANOVA and indicated accordingly.

Suppl. Fig. S5 – T and B immune cell activation in the blood of infected and reinfected young vs. old mice.

Gating strategy of freshly isolated blood immune cells, at days 0 and 7 post-*Pcc* inoculation, from young (8-12 weeks old) and aged (52-60 weeks old) mice: (a) CD4+; (b) CD8+ T cells. The activation of T cell subtypes was determined through the expression of CD44 and CD62L, allowing to identify naïve (CD44–CD62L+), effector or activated (CD44+CD62L–), and memory cells (CD44+CD62L+). (c) Circulating B cells (CD19+). Data are expressed as mean values \pm standard deviations (n=7-10). Statistically significant differences were calculated by one-way ANOVA and indicated accordingly.

Suppl. Fig. S6 – T and B immune cell activation in the bone marrow of infected and reinfected mice.

Gating strategy of freshly isolated bone marrow immune cells, at days 0, 3, 7 and 20 post-*Pcc* inoculation. (a) CD4+; (b) CD8+ T cells. The activation of T cell subtypes was determined through the expression of CD44 and CD62L, allowing to identify naïve (CD44–CD62L+), effector or activated (CD44+CD62L–), and memory cells (CD44+CD62L+). (c) Circulating B cells (CD19+). Data are expressed as mean values \pm standard deviations (n=7-12). Statistically significant differences were calculated by one-way ANOVA and indicated accordingly.

Suppl. Fig. S7 – T and B immune cell activation in the spleen of infected and reinfected mice.

Gating strategy of freshly isolated spleen immune cells, at days 0, 3, 7 and 20 post-*Pcc* inoculation. (a) CD4+; (b) CD8+ T cells. The

activation of T cell subtypes was determined through the expression of CD44 and CD62L, allowing to identify naïve (CD44⁺CD62L⁺), effector or activated (CD44⁺CD62L⁻), and memory cells (CD44⁺CD62L⁺). (c) Circulating B cells (CD19⁺). Data are expressed as mean values \pm standard deviations (n=7-12). Statistically significant differences were calculated by one-way ANOVA and indicated accordingly.

Suppl. Fig. S8.– CD71 assessment in erythroid populations of *Pcc* Infected wild-type and Rag2-deficient mice.

Flow cytometry was used to study the median fluorescen intensity (MFI) of CD71 in erythroid cells in bone marrow of *Pcc* infected wild-type and Rag2-knockout mice at day 7. (a) Histogram representations of the number of CD71⁺ cells in erythroid proerythroblast (ProE), basophilic (BasoE), polychromatic (PolyE), and orthochromatic and reticulocytes (OrthoE+Retic), collected from the bone marrow. (b) Histogram representations show the MFI of CD71⁺ cells in erythroid proerythroblast (ProE), basophilic (BasoE), polychromatic (PolyE), and orthochromatic and reticulocytes (OrthoE+Retic), collected from the spleen. Data were presented as mean values \pm standard deviations (n=8-13). Statistically significant differences were calculated by one-way ANOVA and indicated accordingly.