



Review **Does the Composition of Gut Microbiota Affect Chronic Kidney Disease? Molecular Mechanisms Contributed to Decreasing Glomerular Filtration Rate**

Ewelina Młynarska ^{1,*}^(D), Emilian Budny ¹^(D), Maciej Saar ¹^(D), Ewa Wojtanowska ¹, Justyna Jankowska ¹, Szymon Marciszuk ¹^(D), Marcin Mazur ¹, Jacek Rysz ² and Beata Franczyk ¹

- ¹ Department of Nephrocardiology, Medical University of Lodz, ul. Zeromskiego 113, 90-549 Lodz, Poland; marcin.mazur.publ@gmail.com (M.M.)
- ² Department of Nephrology, Hypertension and Family Medicine, Medical University of Lodz, ul. Zeromskiego 113, 90-549 Lodz, Poland
- * Correspondence: emmlynarska@gmail.com; Tel.: +48-(042)-639-37-50

Abstract: Chronic kidney disease (CKD) is a very prevalent and insidious disease, particularly with initially poorly manifested symptoms that progressively culminate in the manifestation of an advanced stage of the condition. The gradual impairment of kidney function, particularly decreased filtration capacity, results in the retention of uremic toxins and affects numerous molecular mechanisms within the body. The dysbiotic intestinal microbiome plays a crucial role in the accumulation of protein-bound uremic toxins such as p-cresol (pC), indoxyl sulfate (IS), and p-cresyl sulfate (p-CS) through the ongoing fermentation process. The described phenomenon leads to an elevated level of oxidative stress and inflammation, subsequently resulting in tissue damage and complications, particularly an increase in cardiovascular risk, representing the predominant cause of mortality in chronic kidney disease (CKD). Therefore, exploring methods to reduce uremic toxins is currently a pivotal therapeutic strategy aimed at reducing the risk of organ damage in patients with chronic kidney disease (CKD). This review aims to summarize recent discoveries on modifying the composition of the intestinal microbiota through the introduction of special probiotic and synbiotic supplements for CKD therapy. The potential to connect the gut microbiota with CKD opens the possibility for further extensive research in this area, which could lead to the incorporation of synbiotics and probiotics into the fundamental treatment and prevention of CKD.

Keywords: gut microbiota; microbiome; chronic kidney disease (CKD); uremic toxins; probiotics; prebiotics

1. Introduction

Chronic kidney disease (CKD) is a significant global health concern, with a high prevalence, poorly expressed symptoms in the early stages of the disease, and co-morbidity with other disease entities. Approximately 13.4% of the population is affected by CKD, with the majority (10.6%) in the late stages of the disease (stages 3–5) and 0.1% in stage 5 [1]. However, the percentage provided above may be inaccurate as the population suffering from CKD in stages 1–2 may not be aware of their condition, which could lead to its under detection and warping of the data above [2]. The number of patients at stage 5 is low because people with chronic kidney disease (CKD) are five to ten times more likely to die prematurely than to develop kidney failure [3].

Chronic kidney disease is a progressive and irreversible condition that results in the gradual loss of nephrons, leading to an overload of the remaining nephrons through hyperfiltration. This, in turn, causes glomerular hypertrophy, followed by sclerosis and fibrosis of the interstitium [4]. The progressive kidney damage and reduced filtration result in the accumulation of several substances called uremic toxins in the body. It is hypothesized that there is a correlation between the presence of uremic toxins, the



Citation: Młynarska, E.; Budny, E.; Saar, M.; Wojtanowska, E.; Jankowska, J.; Marciszuk, S.; Mazur, M.; Rysz, J.; Franczyk, B. Does the Composition of Gut Microbiota Affect Chronic Kidney Disease? Molecular Mechanisms Contributed to Decreasing Glomerular Filtration Rate. *Int. J. Mol. Sci.* 2024, 25, 10429. https://doi.org/ 10.3390/ijms251910429

Academic Editors: Carolyn M. Ecelbarger and Marijn Speeckaert

Received: 12 August 2024 Revised: 16 September 2024 Accepted: 25 September 2024 Published: 27 September 2024



Copyright: © 2024 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (https:// creativecommons.org/licenses/by/ 4.0/). development of inflammation and immune dysfunction, and the onset of vascular disease. In addition, uremic toxins have been linked to abnormalities in platelet function, altered drug metabolism, dysbiosis in the gut (including increased bacterial translocation), and the advancement of CKD [3,5,6]. Moreover, the 1-alpha-hydroxylation of vitamin D, which occurs in the kidneys, is also impaired, which is the cause of hypocalcemia and secondary hyperparathyroidism [7]. Additionally, in CKD, anemia is caused primarily by reduced erythropoietin production by the kidneys. Damaged kidneys cannot maintain normal blood pH, volume, and adequate electrolyte composition [8].

Furthermore, research findings confirm that patients with chronic kidney disease (CKD) exhibit an altered composition of gut microbiota when correlated to healthy controls (HC) [9–12]. In comparison to the control group at the phylum level, CKD patients demonstrated an increased presence of Proteobacteria and Bacteriodetes while exhibiting reduced levels of Actinobacteria and Firmicutes. In CKD patients, at the genus level, a higher quantity of *Bacteroides, Prevotella*, and *Klebsiella* were detected, while a lowered count of *Bifidobacterium, Collinsella, Dorea, Oscillospira*, and *Akkermansia* were observed in contrast with those of the HC group. In addition, the study demonstrated a reduction in the diversity of the gut microbiota in patients with CKD. These findings suggest a dysbiosis in the gut microbiome of CKD patients [12].

The most common causes of chronic kidney disease are type 2 diabetes and arterial hypertension (AH) [4]. Furthermore, the composition of the gut microbiota and its metabolites could be associated with the onset and progression of each of these diseases [13,14].

Recently, CKD has been gaining popularity amongst scientists and clinicians as their recommended treatment and care are still not up to par [15,16]. This fact emphasizes the demand for further investigation into other treatment options [16]. The connection between CKD and gut microbiota suggests the usefulness of including probiotics and synbiotics in the therapy. The inclusion of such supplements is said to help regulate the balance of the intestinal flora, which may lead to suppression of the progression of CKD [16–19]. As it has been proven in several animal studies, the supplementation of *Lactobacillus* slows the progression of CKD and delays the occurrence of renal failure by altering short-chain fatty acid and nicotinamide metabolism [16,20]. After introducing probiotics to the treatment, the tendency to reduce the levels of tumor necrosis factor- α (TNF- α), interleukin (IL)-6, IL-18, and endotoxin in serum occurred [16,21].

The objective of this review is to describe the role of gut microbiota in renal failure, taking into account the molecular processes involved and new insights into the treatment of chronic kidney disease (CKD).

2. Review Methodology

The comprehensive analysis was performed on papers published between 2004 and 2023. Emphasis was placed on articles pertaining to microbiota composition and probiotics, which were published between 2017 and 2024.

The materials for the review were identified through the use of the following keywords: "gut microbiota", "chronic kidney disease", "uremic toxins", "probiotics", and "prebiotics". The analysis included both English-language and non-English-language publications.

The study population analyzed in the included papers included individuals with chronic kidney disease (CKD) and end-stage renal disease (ESRD), as well as matched control groups. The objective was to analyze the results of changes in the composition of the gut microbiota occurring during CKD and ESRD, including the molecular mechanisms that may influence the development and progression of these diseases and new insights into the use of probiotics and prebiotics in the treatment of kidney diseases.

3. What Is Renal Failure?

3.1. Definition, Types

Chronic kidney disease (CKD) is characterized by a gradual and irreversible progression, with a relatively insidious onset. A significant feature of the disease is the increased risk of complications, with death primarily associated with vascular diseases.

According to Kidney Disease: Improving Global Outcomes (KDIGO), CKD is currently defined as a decreased glomerular filtration rate (GFR) <60 mL/min per 1.73 m² (GFR categories G3a–G5) present for a minimum of 3 months or other abnormalities of kidney structure or function in the form of kidney damage with implications for health, present for a minimum of 3 months [22]. The abnormalities are illustrated in Figure 1.

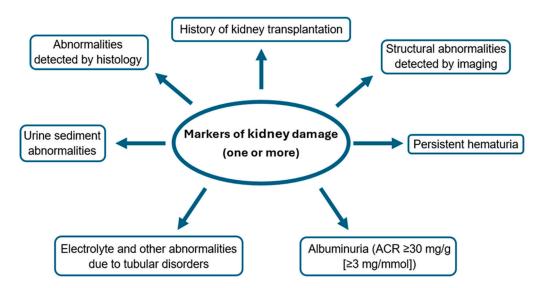


Figure 1. Definition of chronic kidney disease based on markers of kidney damage [22]. ACR—albumin-to-creatinine ratio.

CKD is classified into five stages based on GFR (G1–G5) and three categories based on albuminuria (A1–A3). The divisions are presented in Tables 1 and 2.

Stages	GFR (mL/min/1.73 m ²)	Classification
G1	>90	Normal or high
G2	60–89	Mildly decreased
G3a	45–59	Mildly to moderately decreased
G3b	30–44	Moderately to severely decreased
G4	15–29	Severely decreased
G5	<15	Kidney failure

Table 1.	Categories	of CKD	based or	n GFR leve	l [22].
----------	------------	--------	----------	------------	---------

GFR—glomerular filtration rate. CKD—chronic kidney disease.

Table 2. Categories of CKD based on albuminuria [22].

Category	AER (mg/24 h)	ACR (mg/mmol)	Classification
A1	<30	<3	Normal to mildly increased
A2	30–300	3–30	Moderately increased
A3	>300	>30	Severely increased

ACR—albumin-to-creatinine ratio. AER—albumin excretion ratio.

3.2. Etiology and Causes

The human kidney contains approximately one million nephrons. Following the completion of organogenesis, the body is unable to produce additional nephrons [8,23]. During growth, nephrons also increase in size to align with the body's evolving requirements. However, in situations of prolonged or sustained weight gain (e.g., during pregnancy or obesity), nephron hypertrophy (mainly involving increased dimensions of the glomerular tuft, Bowman's capsule, and proximal tubule) occurs as a compensatory mechanism [8]. The same effect can occur with the loss of healthy nephrons (injury or donation of one kidney), i.e., hypertrophy of the remaining nephrons.

The loss of nephrons results in an increase in intraglomerular pressure, which in turn causes the glomerular walls to stretch, damaging the glomerular cells [24]. Additionally, the appearance of large pores is observed, allowing for increased filtration of plasma proteins [25,26]. Mechanical stress may also increase angiotensin II (AngII) production and angiotensin type 1 receptor expression in podocytes, and AngII may directly impair glomerular barrier screening function, probably through inhibited nephrin expression, independent of its hemodynamic effects [26–28]. Proteinuria is thought to lead to structural and functional damage to the nephron. Furthermore, the loading of podocytes with protein results in the release of transforming growth factor β (TGF- β), which in turn leads to the differentiation of mesangial cells into myofibroblasts [26,29-31]. The increased protein concentration in the tubules causes the release of chemokines, cytokines, vasoactive molecules, and growth factors by the tubule cells, which in turn leads to interstitial fibrosis through the accumulation of inflammatory cells, collagen, fibronectin, and other components [32,33]. Moreover, complement factors accumulate, exerting cytotoxic, pro-inflammatory, and fibrinogenic effects [30]. Additionally, the inflammatory process is driven by tissue damage induced by protein movement, resulting in the production of reactive oxygen species and the endoplasmic reticulum stress response [34]. Through this mechanism, oxidative modification of membrane lipids, proteins, and DNA and induction of cell death occur, resulting in tissue inflammation and local recruitment of macrophages and lymphocytes [33].

The unfavorable microenvironment is thought to promote the mesenchymal transformation of differentiated epithelial cells, possibly also endothelial cells and podocytes, which in turn may further exacerbate proteinuria and glomerular sclerosis [26,35].

Furthermore, following the damage, the process of epithelial–mesenchymal transition (EMT) is initiated in proximal tubule epithelial cells (TECs). This is regulated by E-cadherin SNAI1 and GSK3β, which is inhibited by phosphorylation of ERK and p90RSK [35–40]. The EMT process occurring in TECs has been referred to as the "failed-repair proximal tubule cell" state, often named partial EMT (pEMT) [40]. The reason for the occurrence of scarring rather than regeneration in the kidney is the prevalence of pEMT over TEC proliferation [41]. Studies of interleukin 11 (IL-11) in a mouse model have demonstrated that it also stimulates epithelial–mesenchymal transition (EMT) of epithelial cells [40,42,43]. In a mouse model of CKD, A. A. Widjaja et al. observed that the diseased mice's kidney weight was reduced by 67%. After the IL-11 neutralizing treatment (X203) was admitted, roughly 50% of the lost kidney mass was regained, while kidney size remained unaltered in the control group. At the same time, the collagen content of the kidneys and the degree of histological fibrosis exhibited a gradual decline throughout the course of X203 therapy. The researchers observed that anti-IL11 treatment in chronic kidney disease promoted TEC regeneration, reversed fibrosis and the pEMT phenotype, and enhanced renal function [40].

3.3. Standard Treatment

The treatment of chronic kidney disease includes identification and treatment of the underlying cause, inhibition of disease progression, prevention and treatment of complications, treatment of comorbidities, prevention of cardiovascular disease, preparation for renal replacement therapy, and renal replacement therapy.

3.3.1. Treatment of Hypertension

It is recommended that the presence and severity of albuminuria be assessed prior to initiating treatment. The use of an angiotensin-converting enzyme inhibitor (ACE-I) or an angiotensin II receptor blocker (ARB) is recommended in adults with diabetes and a urinary albumin-to-creatinine ratio (ACR) above 300 mg/24 h [44–47]. Due to the high risk of hyperkalemia and acute kidney injury, it is advisable to avoid dual therapy with ACE-I and ARB. In certain instances, the use of aldosterone receptor antagonists may be considered [44,48].

3.3.2. Treatment of Diabetes Mellitus

Glycemic control represents a crucial element in the management of CKD, with evidence suggesting that achieving a target hemoglobin A1c of approximately 7.0% can delay the progression of the disease [44,45,49–51]. This is in line with the recommendations set forth by the majority of clinical guidelines. During therapy, drugs that are mainly removed by the kidneys should be avoided, and dose reductions or discontinuation may be required for drugs metabolized by the liver and/or partially by the kidneys, especially when eGFR is <30 mL/min/1.73 m² [44,45,49]. In patients with markedly increased albuminuria, it is advisable to discontinue SGLT-2 inhibitors, if possible.

3.3.3. Cardiovascular Disease Risk Reduction

The prevalence of cardiovascular disease in individuals with chronic kidney disease (CKD) is considerably higher than in those without CKD. Furthermore, individuals with CKD exhibit a more unfavorable cardiovascular prognosis [44]. Consequently, reducing cardiovascular risk is a crucial aspect of CKD treatment. It is recommended that patients aged 50 years or older with CKD should be treated with a low- or moderate-dose statin, regardless of their low-density lipoprotein cholesterol level [52,53]. It is recommended that patients should be encouraged to stop smoking [44]. Kidney Disease: Improving Global Outcomes (KDIGO) recommends a target systolic and diastolic blood pressure of less than 140 mm Hg and less than 90 mm Hg, respectively, among adults with CKD based on expert opinion [22,54].

Furthermore, patients with chronic kidney disease (CKD) should avoid nephrotoxins, which include non-steroidal anti-inflammatory drugs (NSAIDs), phosphate-based intestinal preparations, and proton pump inhibitors [44]. A comprehensive list is beyond the scope of this review.

3.4. Complications

The progressive reduction in the number of nephrons results in the disruption of numerous vital processes in which the kidneys are involved. The functionality of a multitude of systems could be impaired, leading to complications such as anemia, bone mineral disorders (MBD associated with vitamin D deficiency, hyperparathyroidism, hyperkalemia, and hyperphosphatemia), hypertension, increased effective circulating fluid volume, hyperuricemia, and dyslipidemia [8].

Among the complications, cardiovascular disease is of particular significance, being the leading cause of death in CKD patients worldwide. This is influenced by dyslipidemia, arterial hypertension, and hyperuricemia [55].

4. Molecular Basis of Renal Failure

Molecular reactions and mechanisms have the potential to regulate a multitude of processes within the human body, including those that may contribute to the development of disease. Chronic kidney disease is characterized by the destruction of the structural and functional units of the kidney, which results in an irreversible reduction in renal function. This is caused by a series of pathogenic mechanisms that target and damage the kidneys [56]. In response to relevant stimuli, inflammation is considered to be a host defense mechanism against pathogens that can generate pro-inflammatory cytokines to activate innate immunity. Moreover, the triggers of renal inflammation include bacterial

6 of 19

and viral infections, lipid metabolism, high glucose levels, and ischemia–reperfusion injury (IRI) [57]. The inflammatory response, characterized by leukocyte infiltration and tubular cell death, is a consequence of renal (IRI). The underlying mechanisms include necroptosis and ferroptosis, which are responsible for initiating the inflammatory cascade and subsequent tissue damage [58].

4.1. Programmed Cell Death

Necroptosis could initiate the spread of cell death through ferroptosis [59]. Necroptosis to ferroptosis may be achieved by phosphatidylethanolamine-binding protein 1 and 15-lipoxygenase [60]. Due to this fact, the combined small-molecule inhibitor Necrostatin-1f, which has a strong inhibitory effect on necroptosis and a weak inhibitory effect on ferroptosis, was developed based on the relationship between necroptosis and ferroptosis [61]. Pyroptosis is a form of programmed cell death that is induced by the activation of the membrane-targeting, pore-forming gasdermin protein family (GSDM). Moreover, pyroptosis is dependent on the activity of caspases -1/4/5/11 in the context of gasdermin D (GSDMD)-mediated pyroptosis, as well as caspases-3 in the presence of gasdermin E (GSDME)-mediated pyroptosis [62]. The sequence of events leading to the activation of pyroptosis and subsequent renal failure is illustrated in Figure 2. Furthermore, it was initially characterized by the loss of membrane integrity and the secretion of cytokines such as IL-1β. Due to this fact, it was initially described as a caspase 1- and inflammasome-dependent pathway, whereby cells undergo regulated cell death in response to the activation of the NLRP3 inflammasome [63]. What is more, pyroptosis may also be associated with ferroptosis [64]. ROS-Tom20-Caspase3-GSDME-signaling pathway: iron ions and reactive oxygen species (ROS)-inducing drugs induce pyroptosis [65]. Pyroptosis is putatively implicated in the pathogenesis of kidney diseases through two distinct pathways: the caspase-1-mediated canonical pathway and the caspase-4/5/11-mediated noncanonical pathway [66].

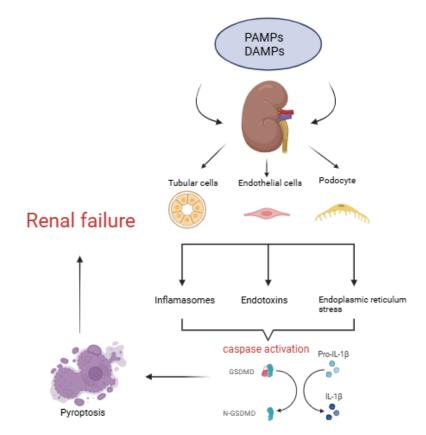


Figure 2. Two distinct molecular mechanisms, pathogen-associated molecular patterns (PAMPs) and damage-associated molecular patterns (DAMPs), are responsible for activating inflammatory bodies [66].

These factors induce cellular stress, which subsequently activates the inflammasome [67]. The inflammasome activates caspase-1, which in turn induces the cleavage and release of the proinflammatory cytokines IL-1 β and IL-18 [68]. Cleaved caspase-1 has been established as a direct activator of GSDMD [63]. Upon cleavage of GSDMD into GSDMD-NT, the resulting fragments migrate to the membrane, where they aggregate and perforate the membrane, thereby releasing inflammatory factors such as IL-1 β and IL-18, which lead to pyroptosis [69]. Several common kidney diseases have the potential to progress to end-stage renal disease (ESRD), including acute kidney injury, diabetic kidney diseases, renal fibrosis, and kidney inflammation. It has been established that all forms of kidney disease are caused by a certain level of inflammatory response. Pyroptosis, which is regulated by a variety of inflammatory bodies, plays a significant role in the progression of kidney disease [66]. PAMPs—pathogen-associated molecular patterns. DAMPs—damage-associated molecular patterns. IL—Interleukin. GSDMD—Gasdermin D.

4.2. The Co-Occurrence of Obesity and Renal Failure

A previous meta-analysis indicated a positive correlation between increased body mass index (BMI) and the risk of developing kidney disease (KD). This suggests that obesity significantly increases the risk of KD in the population [70]. Therefore, it has been documented that obesity-related glomerulopathy (ORG) constitutes a form of chronic kidney disease (CKD) [71]. The adverse effects of lipotoxicity on the progression of ketogenic diets among patients with acute kidney injury (AKI) and chronic kidney disease, including diabetic nephropathy (DN), ORG, and polycystic kidney disease (PKD), have been well documented [72,73]. The de novo formation of lipids is contingent upon an equilibrium between the acquisition of lipids and their subsequent disposal. This equilibrium is regulated due to three major pathways: lipid uptake, lipid synthesis, fatty acid oxidation (FAO), and lipid export. This section will examine the molecular mechanisms that regulate lipid homeostasis in KD [74]. In the kidney, the expression of cluster of differentiation 36 (CD36) can be observed in distal tubular epithelial cells (TECs), mesangial cells, podocytes, microvascular endothelial cells, and interstitial macrophages. However, the expression of CD36 in proximal tubule cells (PTCs) appears to be inconsistent [75–77]. Furthermore, overexpression of CD36 has been demonstrated to result in increased renal tubular damage and renal fibrosis in folic acid-treated mice compared to wild-type controls. However, there was no significant difference in renal inflammation observed [78]. In addition to the well-known inflammatory factors, recent studies have shown that the collagen type I (Col I)-mediated discoidin domain receptor 1 (DDR1) activation induces CD36-mediated podocyte lipotoxic injury, which was previously unrecognized [79].

4.3. Uremic Toxins

End-stage renal disease is inextricably linked to the accumulation of toxic metabolites in the blood and other metabolic compartments. This accumulation has been postulated to be associated with an increased generation of toxins from a dysbiotic microbiome, which is accompanied by a reduction in their elimination by impaired kidneys [80]. The loss of kidney function is associated with the development of intestinal dysbiosis, which is characterized by the secretion of urea into the gastrointestinal tract and the subsequent hydrolysis of urea into carbon dioxide and ammonia by certain gut microbes [81]. Patients with chronic kidney disease exhibit a diminished excretion of uremic toxins, which are hazardous metabolites, due to impaired functioning of the glomerulus and proximal tubules [82]. The presence of chronic kidney disease may be accompanied by the development of intestinal inflammation and epithelial barrier impairment. This can result in accelerated translocation of bacterial-derived uremic toxins from the gut into the systemic circulation, leading to increased oxidative stress and injury to the kidney, cardiovascular, and endocrine systems [80]. It seems that the prevalence of bacteria with fermentative activity may lead to the release and accumulation in the gut and in the blood of several substances, such as p-cresol (p-C), indoxyl sulfate (IS), and p-cresyl sulfate (p-CS) [83,84].

IS is a protein-bound uremic toxin that is derived from indole, a metabolite produced by the gut microbiota through the degradation of tryptophan. It has been demonstrated that IS causes intracellular oxidative stress and influences transcription via the aryl hydrocarbon receptor [85]. The presence of indoxyl sulfate has been demonstrated to induce pro-inflammatory responses in endothelial cells [86]. In tubular epithelial cells, indoxyl sulfate is excreted from the blood into the urine via organic anion transporters (OATs) and organic anion transporting polypeptides (OATPs), which are encoded by genes in the SLCO superfamily [87,88]. IS has been observed to induce the expression of intercellular adhesion molecule 1 (ICAM-1) in the kidney, which has been demonstrated to enhance monocyte infiltration. Additionally, it has been shown to stimulate the production of monocyte chemoattractant protein (MCP-1), a chemokine that plays a crucial role in macrophage recruitment and activation [89,90]. Furthermore, macrophage induction is a consequence of the acceleration of the aryl hydrocarbon-NF-κB/MAPK receptor cascades [91]. The transcription factor NF-κB plays a pivotal role in the pathological effects of IS in the kidneys. In human proximal tubule cells (HK-2), it has been observed that the activation of NF-κB by IS inhibits cell proliferation, induces and accelerates aging through the induction of p53, and promotes fibrosis through the induction of TGF- β 1 and plasminogen activator inhibitor-1 (PAI-1) expression [92]. Furthermore, it was postulated that p53 induction may also be involved in renal fibrosis through the stimulation of TGF- β 1 expression, which in turn activates the Smad3 pathway [93]. The remaining two uremic toxins, p-C and p-CS are produced as metabolites of tyrosine and phenylalanine. These are directly produced by intestinal bacteria when there is dysbiosis [94]. Furthermore, P-C has been demonstrated to induce vascular damage and genotoxicity in enterocytes [95]. It has been suggested that also p-CS may contribute to immune dysfunction by suppressing the activity of macrophages, which are pivotal in the immune system. Additionally, p-CS has been linked to the progression of renal injury and has also been demonstrated to increase ROS production and induce immunosuppression [83].

5. Association between Chronic Kidney Disease and Gut Microbiota

5.1. Basic Microbiological Information and Molecular Aspects

The influence of the gut microbiota on the human body has been a subject of scientific investigation for a considerable period [13].

Scientists show that the composition of gut microbiota could be divided into two distinct enterotypes. The first enterotype was characterized by a higher abundance of *Prevotella*, whereas the second enterotype exhibited a higher prevalence of *Bacterioides* [96]. However, some scientists have proposed a third enterotype, which is dominated by *Ruminococcus* [97]. The findings of Jiang et al. show that at the end stage of renal disease (ESRD), the total amount of gut bacteria may be reduced. Furthermore, the authors suggest that there is a possibility of a higher prevalence of *Prevotella* in the healthy group, whereas Bacteroides could be present in greater quantities in patients with ESRD [97].

The scientific literature indicates that the gut microbiota may be responsible for the generation of metabolic products, including gut-derived uremic toxins such as α -phenylacetyll-glutamine, 5-hydroxyindole, indoxyl glucuronide, p-cresyl sulfate (PCS), and indoxyl sulfate (IS) [98]. Of these, p-cresyl sulfate and indoxyl sulfate have been the subject of extensive study to date, which has demonstrated that they may be elevated in patients with kidney diseases [99]. Moreover, a higher presence of bacteria is correlated with higher IS and PCS levels, as evidenced by the findings of studies that have examined the relationship between the two variables. *Bacteroides* and *Blautia* appear to be associated with a high IS level, while *Enterococcus*, *Akkermansia*, *Dialister*, and *Ruminococcus* are linked to a higher PCS level [13]. Furthermore, these metabolites may exert a deleterious effect on renal tubular cells and endothelium, which could lead to their damage [84,100]. Therefore, the accumulation of uremic toxins may contribute to the progression of chronic kidney disease [84]. The possible link between gut-derived uremic toxins was also noted by S.C. Guldris et al. The authors observed clinical consequences in the form of progression of CKD due to increased inflammation and oxidative stress [101].

5.2. Microbiological Assessment and Used Technical Methods

The composition of the microbiota was identified from fecal samples in the analyzed papers. Bacterial DNA was isolated from the samples after purification from human DNA and amplified by a PCR reaction. Subsequently, 16S rRNA gene sequencing or metagenome shotgun sequencing was performed, followed by metagenetic analysis and gene mapping to identify specific species.

5.3. Quantitative and Qualitative Gut Microbiota Analysis in Patients with CKD and ESRD Compared to the Healthy Population

Some bacteria may be less abundant in chronic kidney disease. This correlation is illustrated in Table 3. Wang et al. suggested that there is a probability of lower quantities of the Clostridiaceae family without specifying a particular type of genus [11]. Moreover, greater presence of *Roseburia* and *Faecalibacterium* in the population without chronic kidney disease has been noticed in many scientific research studies [11,97,102–105]. Furthermore, a lower capacity of *Eubacterium* and *Ruminococcus* could be non-linearly correlated with a decrease in glomerular filtration rate (GFR) [11,104,105]. Studies have reported that *Bacteroides*, *Prevotella*, *Blautia*, and *Lactobacillus* could decrease in the population with CKD in comparison to the healthy population [97,102,106]. Additionally, the *Lachnospira*, *Veillonella*, and *Dialister* populations within the guts of patients diagnosed with chronic kidney disease may exhibit a reduction in abundance when compared to healthy populations. However, the extent of this reduction may vary depending on the stage of the disease [107].

Table 3. Presentation of the lower quantity of bacterial populations in patients with chronic kidney disease in comparison to healthy individuals. The table presents the distribution of successive taxonomic levels, including cluster, class, order, family, genus and species, as well as Gram classification In consideration of the detail provided in the referenced works, when uncertainty existed regarding a specific taxonomic level, a cell in the table was left filled with the symbol '-' to indicate that species was not known with certainty [11,97,102–107].

Phylum	Class	Order	Family	Genus	Species	Gram Classification	Presence in CKD
Bacteroidota	Bacteroidia	Bacteroidales	Bacteroidaceae	Bacteroides	Fragilis	Negative	Lower
Bacteroidetes	Bacteroidia	Bacteroidales	Prevotellaceae *	Prevotella	-	Negative	Lower
Bacillota	Bacilli	Lactobacillales	Lactobacillaceae *	Lactobacillus	-	Positive	Lower
Bacillota	Clostridia	Eubacteriales	Lachnospiraceae	Roseburia	Hominis	Positive	Lower
Bacillota	Clostridia	Eubacteriales	Oscillospiraceae	Faecalibacterium	Prausnitzii	Positive	Lower
Bacillota	Negativicutes	Vellionellales	Veillonellaceae *	Veillonella	Parvula	Negative	Lower
Bacillota	Clostridia	Eubacteriales	Lachnospiraceae	Lachnospira	-	Positive	Lower
Bacillota	Negativicutes	Veillonellales	Veillonellaceae *	Dialister	Succinatiphilus	Negative	Lower
Firmicutes	Clostridia	Clostridiales	Clostridiaceae	-	-	Positive	Lower
Bacillota	Clostridia	Clostridiales	Eubacteriaceae *	Eubacterium	Rectale	Positive	Lower
Firmicutes	Clostridia	Clostridiales	Ruminococcaceae	Ruminococcus	Bromii, Callidus	Positive	Lower
Bacillota	Clostridia	Eubacteriales	Lachnospiraceae	Blautia	-	Positive	Lower

* A lower presence of this family may also occur in patients undergoing dialysis in comparison to the healthy population [11,99].

Researchers have demonstrated that a particular bacterial population may be more prevalent in individuals with chronic kidney disease. The correlation is illustrated in Table 4. S. T. Gryp et al. showed that an increased *Escherichia* population may occur in people with CKD [108]. Furthermore, a lower abundance of *Enterococcus* and *Klebsiella* could have been observed in people who do not suffer from chronic kidney disease [97,99,106]. Furthermore, H. Wang et al. observed that patients with chronic kidney disease exhibited

an elevated prevalence of the *Flavonifractor* and *Citrobacter* genera [11]. These outcomes were also observed in other independent studies [104,105]. Nevertheless, an augmented prevalence of the Selenomonadaceae family as well as the *Akkermansia* genus has also been observed [11,106]. Additionally, the populations of *Desulfovibrio* and *Streptococcus* appeared to be diminished in healthy controls [103,109]. Additionally, an increased prevalence of *Oscillibacter* and *Alistipes* has been observed in individuals with chronic kidney disease, with a potential correlation between this elevation and the severity of the disease [107].

The scientific studies presented in Table 5 show that in patients with chronic kidney disease, the composition of the intestinal microbiota varies depending on the section of the gastrointestinal tract examined. Moreover, scientific researchers observed variations in the gut bacterial population in patients undergoing dialysis, including hemodialysis and peritoneal dialysis. In a study presented by G. P. Hobby et al., an elevated prevalence of bacteria belonging to the genera *Brachybacterium* and *Catenibacterium*, as well as the families Halomonadaceae and Pseudomonadaceae, was observed in dialysis patients [99]. Moreover, a potential decrease in the Prevotellaceae population was noted in patients with chronic kidney disease and those undergoing dialysis [97,99]. Furthermore, H. Wang et al. showed that in dialysis patients, the population of bacteria from the Proteobacteria phylum and the Enterobacteriaceae family could be increased, and the number of bacteria from the Euryarchaeota phylum and the Veillonellaceae, Lactobacillaceae, and Eubacteriaceae families could be decreased [11].

Table 4. Presentation of the higher quantity of bacterial populations in patients with chronic kidney disease in comparison to healthy individuals. The table presents the distribution of successive taxonomic levels, including cluster, class, order, family, genus, and species, as well as Gram classification. In consideration of the detail provided in the referenced works, when uncertainty existed regarding a specific taxonomic level, a cell in the table was left filled with the symbol '-' to indicate that the species was not known with certainty [11,97,99,103–109].

Phylum	Class	Order	Family	Genus	Species	Gram Classification	Presence in CKD
Firmicutes	Bacilli	Lactobacillales	Enterococcaceae *	Enterococcus	-	Positive	Higher
Pseudomonadota	Gammaproteobacteria	Enterobacteriales	Enterobacteriaceae *	Klebsiella	-	Negative	Higher
Bacillota	Bacilli	Lactobacillales	Streptococcaceae	Streptococcus	-	Positive	Higher
Pseudomonadota	Gammaproteobacteria	Enterobacterales	Enterobacteriaceae *	Escherichia	-	Negative	Higher
Proteobacteria *	Deltaproteobacteria	Desulfovibrionales	Desulfovibrionaceae	Desulfovibrio	-	Negative	Higher
Firmicutes	Clostridia	Clostridiales	Ruminococcaceae	Oscillibacter	-	Positive	Higher
Bacillota	Negativicutes	Selenomonadales	Selenomonadaceae	-	-	Negative	Higher
Bacillota	Clostridia	Eubacteriales	Oscillospiraceae	Flavonifractor	Plautii	Positive	Higher
Pseudomonadota	Gammaproteobacteria	Enterobacterales	Enterobacteriaceae *	Citrobacter	Freundii	Negative	Higher
Bacteroidota	Bacteroidia	Bacteroidales	Rikenellaceae	Alistipes	Werkmanii	Negative	Higher
Verrucomicrobiota	Verrucomicrobiae	Verrucomicrobiales	Akkermansiaceae	Akkermansia	-	Negative	Higher

* A higher presence of this family may also occur in patients undergoing dialysis in comparison to the healthy population [70,73].

Table 5. Presentation of the alterations in the composition of the gut microbiota in relation to the specific section of the digestive tract [110].

Intestinal Tract	Normal	ACKD/CKD
Stomach	Helicobacter, Lactobacillus	No change observed
Duodenum	Lactococcus, Streptococcus, Staphylococcus	Increased
Jejunum	Streptococcus, Lactobacillus, Enterococcus	Increased
Ileum	Clostridium, Enterobacteriaceae, Bacteroides	Increased
Colon	Fusobacterium Prevotellaceae Proteus Actinobacteria Bacteroides Firmicutes	Aerobic overgrowth c.a. 100 times Bifidobacterium spp., Lactobacillus Acinetobcter, Proteus spp. Enterobacteria, E. coli Proteobacteria Increased

CKD-chronic kidney disease. ACKD-advanced chronic kidney disease.

6. Potential Role of Probiotics in the Treatment of Chronic Kidney Disease

6.1. Gut Dysbiosis and Chronic Kidney Disease—Can We Treat Both?

Presumably, probiotics can alter gut microbiota in patients with chronic kidney disease (CKD). In CKD, the composition of gut microbiota often becomes imbalanced, leading to overgrowth of harmful bacteria and accumulation of uremic toxins (e.g., indoxyl sulfate, p-cresyl sulfate). The said imbalance, known as dysbiosis, promotes inflammation, oxidative stress, and eventually the progression of CKD [111].

Although small-scale studies and animal-based models have shown promising results, larger, well-controlled clinical trials are required to confirm the efficacy of probiotics in CKD management. The impact of probiotics on clinical outcomes like CKD progression and mortality in CKD and cardiovascular health patients remains an area of active research [112].

6.2. What Are Probiotics?

Probiotics are defined as live microorganisms that provide health benefits when consumed regularly in adequate amounts. Probiotics might be found in fermented foods such as yogurt and are also available as dietary supplements. Those microorganisms have the potential to support gut health by maintaining a healthy balance of gut microbiota, which influences various bodily functions, including the immune response, blood pressure, digestion, and even kidney function. Recent scientific research suggests the potential benefits of using probiotics in chronic kidney disease (CKD). The potential efficacy of probiotics in reducing uremic toxin levels and delaying CKD progression has been investigated in in vitro models, animal models, and CKD patients [113]. Moreover, synbiotics are a combination of both probiotics and prebiotics, which may facilitate the survival and implantation of live microbial food supplements in the gastrointestinal tract. The objective of synbiotics is to improve host health by enhancing the beneficial effects of probiotics and prebiotics [114]. Although the potential of probiotics and synbiotics in the treatment and prevention of diseases represents an exciting area of research, several challenges remain, as outlined by the authors in Table 6 [110].

Table 6. Challenges and difficulties related to developing probiotic- and synbiotic-based therapy for chronic kidney disease [110].

Name of a Challenge	Importance	
Strain-specific effects	Identifying a highly specific strain that could be useful and excluding those that are ineffective	
Long-term safety and efficacy	Long-term studies are necessary to provide definitive evidence regarding the efficacy of the proposed treatment	
Individual variability	The organism's response to the probiotic is variable	
Regulatory and quality issues	Ensuring the optimal quality and quantity of the product	

6.3. Probiotics and Renal Failure: Mechanisms of Action

A review of scientific literature reveals a multitude of potential mechanisms through which probiotics may exert an effect on kidney failure. These include the following:

- Reduction in uremic toxins: Probiotics could support a reduction in the production and absorption of uremic toxins by modifying the gut microbiota composition. Research has shown that specific probiotic strains, including Lactobacillus and Bifidobacterium, have the potential to decrease the levels of indoxyl sulfate and p-cresyl sulfate, which may potentially slow the progression of kidney disease [92].
- 2. **Improvement of gut barrier function:** Renal failure could compromise the integrity of the gut barrier, leading to increased intestinal permeability (leaky gut). Moreover, probiotics could strengthen the gut barrier by promoting the growth of beneficial bacteria that enhance gut integrity. These microorganisms might reduce the translo-

cation of harmful bacteria and toxins from the gut into the bloodstream while also potentially mitigating systemic inflammation and further kidney damage [115].

- 3. **Metabolic benefits:** Probiotics have the potential to enhance metabolic profiles by increasing insulin sensitivity, reducing oxidative stress, and lowering lipid levels [21].
- 4. Anti-inflammatory effects: Chronic inflammation represents a pivotal element in the advancement of renal failure. Probiotics could exert anti-inflammatory effects by modulating the immune response and reducing the production of pro-inflammatory cytokines, thereby aiding in the protection of kidney tissues from further damage [116].
- 5. **Regulation of blood pressure:** Hypertension is both a cause and a consequence of renal failure. Some probiotic strains could help regulate blood pressure by producing bioactive peptides that inhibit angiotensin-converting enzyme (ACE), a key regulator of blood pressure. Improved blood pressure control can reduce the stress on the kidneys and slow the progression of kidney disease [117].

6.4. The Use of Probiotics and Synbiotics in Kidney Diseases

Several studies have yielded encouraging results in the treatment of renal diseases. A study published in the Journal of Renal Nutrition showed that patients with chronic kidney disease who received a probiotic supplement containing Lactobacillus and Bifidobacterium strains could exhibit a notable reduction in serum urea and creatinine levels compared to the control group. These findings suggest that probiotics may facilitate improvements in renal function and delay the necessity for dialysis [118]. Furthermore, research in patients with end-stage renal disease undergoing hemodialysis has indicated that probiotic supplementation may reduce inflammatory markers and improve quality of life. For example, a study published in the American Journal of Kidney Diseases reported that patients receiving a multi-strain probiotic exhibited decreased levels of C-reactive protein (CRP), a marker of inflammation [119]. Moreover, research in the field of acute kidney injury (AKI) remains scarce. However, animal studies have indicated that probiotics could reduce kidney injury by modulating the immune response and decreasing oxidative stress. These findings suggest potential benefits in the prevention of AKI in high-risk patients, such as those undergoing major surgery or experiencing severe infections.

6.5. The Use of Probiotics and Synbiotics in Chronic Kidney Disease

The scientific literature indicates that from the earliest stages of chronic kidney disease, there could be a quantitative and qualitative alteration of the intestinal microflora (dysbiosis), which could result in changes in the composition and metabolic activities of the microflora [120]. A review of the literature reveals that a multitude of observational, randomized, placebo-controlled, and double-blind studies have documented the beneficial effects of probiotics and synbiotics on CKD patients, even at the molecular level. The authors have compiled a comprehensive summary of these findings in Tables 7 and 8.

Authors	Synbiotic	Results
T. Ogawa et al. [121]	Bifidobacterium longum JBL01 oligosaccharides	Decrease phosphorous levels that returned to baseline 2 weeks later
I. Nakabayashi et al. [122]	Bifidobacterium breve Yakult Lactobacillus casei Shirota galactooligosaccharides	Decrease in p-cresol in plasma Returning to correct bowel movement Connection of p-cresol level and constipation
J. Cruz-Mora et al. [123]	Bifidobacterium lactis Lactobacillus acidophilus inulin	Increase in <i>Bifidobacteria</i> in feces Decrease of <i>Lactobacilli</i> in feces Alleviation of gastrointestinal symptoms
D. Viramontes-Hörner et al. [124]	Bifidobacterium lactis Lactobacillus acidophilus Inulin	Diminishing of CRP and TNF-alpha levels. Alleviation of gastrointestinal symptoms

Table 7. The impact of synbiotic usage on the gut microbiota in patients with chronic kidney disease.

Authors	Synbiotic	Results
B. Guida et al. [125]	Lactobacillus casei subsp. Rhamnosus, Lactobacillus plantarum, Bifidobacterium infantis, Lactobacillus gasseri, Lactobacillus salivarius, Streptococcus thermophilus, Lactobacillus sporogenes, resistant tapioca starch and inulin	Decrease in p-cresol in plasma
M. Rossi et al. [126]	Bifidobacteria Lactobacillus Streptococcus Inulin Galactooligosaccharides Fructooligosaccharides	Increased <i>Bifidobacteria</i> Decreased Ruminococcaceae Slight increase in albuminuria No alteration in inflammation markers and oxidative stress Decrease PCS

Table 7. Cont.

Table 8. The impact of probiotic usage on the gut microbiota in patients with chronic kidney disease.

Authors	Probiotic	Results
M. L. Simenhoff et al. [127]	Lactobacillus acidophilus	↓ Dimethylamine, ↓Nitrosodimethylamine
F. Takayama et al. [128]	Bifidobacterium longum JCM008	\downarrow Indoxyl sulfate
K. Taki et al. [129]	Bifidobacterium longum	↓ Homocysteine, indoxyl sulfate, and triglycerides
Y. Ando et al. [130]	Bifidobacterium longum	Lowering of CKD's progression in patients with $con Cr \ge 4 mg/dl \text{ or } P \ge 4 mg/dl$
M. Hida et al. [131]	Lebenin	\downarrow p-cresol in feces and in serum
R. Natarajan et al. [132]	Renadyl	Reduction in CRP, leucocyte count, and indoxyl glucuronide
IK. Wang et al. [133]	Bifidobacterium catenulatum A302, Lactobacillus plantarum A87, Bifidobacterium longum A101, Bifidobacterium bifidum A218,	\uparrow IL-10 Slight preservation of kidney function ↓ TNF-α, IL-5, IL-6, and endotoxin
P. V. M. Alatriste et al. [110]	Lactobacillus casei shirota	↓ Urea
N. Ranganathan et al. [134]	Streptococcus thermophilus KB27, Bifidobacterium longum KB35, Lactobacillus acidophilus KB31	↑ Quality of life
N. Ranganathan et al. [135]	Bifidobacterium longum KB35, Lactobacillus acidophilus KB31, Streptococcus thermophilus KB27,	↑ Quality of life

 \downarrow —Decreased level of. \uparrow —Increased level of.

7. Conclusions

The gut microbiota significantly impacts the development of various diseases, including chronic kidney disease. Extensive research has been conducted to investigate the relationship between the composition of the gut microbiota and the development of chronic kidney disease (CKD). It has been established that individuals with chronic kidney disease may exhibit dysbiosis in the composition of the intestinal microbiome, leading to the accumulation and accelerated translocation of uremic toxins due to compromised integrity of the intestinal epithelial barrier. This may result in an increase in oxidative stress and the onset of renal inflammation, which ultimately leads to renal damage. Patients with chronic kidney disease could derive benefit from dietary intervention incorporating prebiotics, probiotics, and synbiotics. This approach aims to modify the dysbiotic intestinal microbiome, which represents the primary goal in the treatment of uremic toxins in CKD. Furthermore, they have been shown to have a beneficial influence on the enhancement of epithelial barrier function and the regulation of blood pressure. The potential to alleviate symptoms and provide information about their impact on the pathogenesis and progression of chronic kidney disease is a source of considerable optimism.

It is important to note that there are outstanding unresolved issues: the short observation period, the lack of identification of the most suitable strains for intervention, and the variable individual response to probiotics. These factors make it impossible to reach a definitive conclusion on this matter now. Moreover, the significant individual variability in the intestinal microbiota of CKD patients, along with additional confounding factors such as diverse dietary patterns or other supplements, hinders the formation of objective conclusions. These challenges could enable researchers to ascertain the therapy's full potential and contribute to groundbreaking results in the future. To improve the quality of future studies, it is important to define inclusion criteria related to patients' diets and other supplements to eliminate any additional variables. In this review, the authors also propose to expand the study assessment to answer the question of whether dysbiosis within the microbiome occurs before or after laboratory manifestations of chronic kidney disease. If these changes manifest prior to the onset of CKD, we should monitor the microbiome in at-risk individuals to implement prevention earlier and modify patient-related risk factors before symptoms develop. Further research is imperative to evaluate the potential impact of introducing probiotic therapy, prebiotic therapy, or synbiotic therapy in delaying the onset of initial symptoms or progression of chronic kidney disease.

Author Contributions: Conceptualization: E.B. and E.M.; methodology: E.M. and E.B.; software: E.M.; validation: E.M., B.F. and J.R.; formal analysis: E.M., E.B., M.S., E.W., J.J., S.M. and M.M.; investigation: E.B., M.S., E.W., J.J., S.M. and M.M.; investigation: E.B., M.S., E.W., J.J., S.M. and E.M.; resources: E.M., B.F. and J.R.; data curation: E.M.; writing original draft preparation: E.B., M.S., E.W., J.J., M.M. and S.M.; writing—review and editing: E.M.; visualization: E.M. and E.B.; supervision: E.M.; project administration: E.M., B.F. and J.R.; funding acquisition: B.F. and J.R. All authors have read and agreed to the published version of the manuscript.

Funding: This research received no external funding.

Institutional Review Board Statement: Not applicable.

Informed Consent Statement: Not applicable.

Data Availability Statement: The data used in this article is sourced from materials mentioned in the References section.

Conflicts of Interest: The authors declare no conflicts of interest.

References

- 1. Hill, N.R.; Fatoba, S.T.; Oke, J.L.; Hirst, J.A.; O'Callaghan, C.A.; Lasserson, D.S.; Hobbs, F.D.R. Global Prevalence of Chronic Kidney Disease—A Systematic Review and Meta-Analysis. *PLoS ONE* **2016**, *11*, e0158765. [CrossRef]
- Evans, M.; Lewis, R.D.; Morgan, A.R.; Whyte, M.B.; Hanif, W.; Bain, S.C.; Davies, S.; Dashora, U.; Yousef, Z.; Patel, D.C.; et al. A Narrative Review of Chronic Kidney Disease in Clinical Practice: Current Challenges and Future Perspectives. *Adv. Ther.* 2021, 39, 33–43. [CrossRef]
- 3. Webster, A.C.; Nagler, E.V.; Morton, R.L.; Masson, P. Chronic Kidney Disease. Lancet 2017, 389, 1238–1252. [CrossRef]
- 4. Charles, C.; Ferris, A.H. Chronic Kidney Disease. Prim. Care Clin. Off. Pr. 2020, 47, 585–595. [CrossRef]
- 5. Anders, H.-J.; Andersen, K.; Stecher, B. The intestinal microbiota, a leaky gut, and abnormal immunity in kidney disease. *Kidney Int.* **2013**, *83*, 1010–1016. [CrossRef]
- 6. Vanholder, R.; Baurmeister, U.; Brunet, P.; Cohen, G.; Glorieux, G.; Jankowski, J. A Bench to Bedside View of Uremic Toxins. *J. Am. Soc. Nephrol.* **2008**, *19*, 863–870. [CrossRef]
- Zaimi, M.; Grapsa, E. Current therapeutic approach of chronic kidney disease-mineral and bone disorder. *Ther. Apher. Dial.* 2024, 28, 671–689. [CrossRef]
- Romagnani, P.; Remuzzi, G.; Glassock, R.; Levin, A.; Jager, K.J.; Tonelli, M.; Massy, Z.; Wanner, C.; Anders, H.-J. Chronic kidney disease. *Nat. Rev. Dis. Primers.* 2017, 3, 17088. [CrossRef]

- 9. Krukowski, H.; Valkenburg, S.; Madella, A.-M.; Garssen, J.; van Bergenhenegouwen, J.; Overbeek, S.A.; Huys, G.R.B.; Raes, J.; Glorieux, G. Gut microbiome studies in CKD: Opportunities, pitfalls and therapeutic potential. *Nat. Rev. Nephrol.* **2022**, *19*, 87–101. [CrossRef]
- 10. Zhang, P.; Wang, X.; Li, S.; Cao, X.; Zou, J.; Fang, Y.; Shi, Y.; Xiang, F.; Shen, B.; Li, Y.; et al. Metagenome-wide analysis uncovers gut microbial signatures and implicates taxon-specific functions in end-stage renal disease. *Genome Biol.* 2023, 24, 226. [CrossRef]
- 11. Wang, H.; Ainiwaer, A.; Song, Y.; Qin, L.; Peng, A.; Bao, H.; Qin, H. Perturbed gut microbiome and fecal and serum metabolomes are associated with chronic kidney disease severity. *Microbiome* **2023**, *11*, 3. [CrossRef]
- 12. Li, J.; Shen, Y.; Yan, K.; Wang, S.; Jiao, J.; Chi, H.; Zhong, J.-C.; Dong, Y.; Wang, P. The compositional and functional imbalance of the gut microbiota in CKD linked to disease patterns. *J. Transl. Med.* **2024**, *22*, 773. [CrossRef]
- 13. Tokarek, J.; Budny, E.; Saar, M.; Kućmierz, J.; Młynarska, E.; Rysz, J.; Franczyk, B. Does the Composition of Gut Microbiota Affect Hypertension? Molecular Mechanisms Involved in Increasing Blood Pressure. *Int. J. Mol. Sci.* 2023, 24, 1377. [CrossRef]
- 14. Gurung, M.; Li, Z.; You, H.; Rodrigues, R.; Jump, D.B.; Morgun, A.; Shulzhenko, N. Role of gut microbiota in type 2 diabetes pathophysiology. *EBioMedicine* **2020**, *51*, 102590. [CrossRef]
- Neuen, B.L.; Heerspink, H.J.; Vart, P.; Claggett, B.L.; Fletcher, R.A.; Arnott, C.; Costa, J.d.O.; Falster, M.O.; Pearson, S.-A.; Mahaffey, K.W.; et al. Estimated Lifetime Cardiovascular, Kidney, and Mortality Benefits of Combination Treatment With SGLT2 Inhibitors, GLP-1 Receptor Agonists, and Nonsteroidal MRA Compared With Conventional Care in Patients With Type 2 Diabetes and Albuminuria. *Circulation* 2024, 149, 450–462. [CrossRef]
- 16. Liu, C.; Yang, L.; Wei, W.; Fu, P. Efficacy of probiotics/synbiotics supplementation in patients with chronic kidney disease: A systematic review and meta-analysis of randomized controlled trials. *Front. Nutr.* **2024**, *11*, 1434613. [CrossRef]
- 17. Meijers, B.; Evenepoel, P.; Anders, H.-J. Intestinal microbiome and fitness in kidney disease. Nat. Rev. Nephrol. 2019, 15, 531–545. [CrossRef]
- Cosola, C.; Rocchetti, M.M.; di Bari, I.; Acquaviva, P.M.; Maranzano, V.; Corciulo, S.; Di Ciaula, A.; Di Palo, D.M.; La Forgia, F.M.; Fontana, S.; et al. An Innovative Synbiotic Formulation Decreases Free Serum Indoxyl Sulfate, Small Intestine Permeability and Ameliorates Gastrointestinal Symptoms in a Randomized Pilot Trial in Stage IIIb-IV CKD Patients. *Toxins* 2021, 13, 334. [CrossRef]
- Lim, P.S.; Wang, H.F.; Lee, M.C.; Chiu, L.-S.; Wu, M.-Y.; Chang, W.-C.; Wu, T.K. The Efficacy of Lactobacillus-Containing Probiotic Supplementation in Hemodialysis Patients: A Randomized, Double-Blind, Placebo-Controlled Trial. *J. Ren. Nutr.* 2021, 31, 189–198. [CrossRef]
- 20. Barrows, I.R.; Ramezani, A.; Raj, D.S. Gut Feeling in AKI. J. Am. Soc. Nephrol. 2015, 26, 1755–1757. [CrossRef]
- de Araújo, É.M.R.; Meneses, G.C.; Carioca, A.A.F.; Martins, A.M.C.; Daher, E.D.F.; da Silva Junior, G.B. Use of probiotics in patients with chronic kidney disease on hemodialysis: A randomized clinical trial. *Braz. J. Nephrol.* 2023, 45, 152–161. [CrossRef]
- Stevens, P.E.; Ahmed, S.B.; Carrero, J.J.; Foster, B.; Francis, A.; Hall, R.K.; Herrington, W.G.; Hill, G.; Inker, L.A.; Kazancıoğlu, R.; et al. KDIGO 2024 Clinical Practice Guideline for the Evaluation and Management of Chronic Kidney Disease. *Kidney Int.* 2024, 105, S117–S314.
- 23. Bertram, J.F.; Douglas-Denton, R.N.; Diouf, B.; Hughson, M.D.; Hoy, W.E. Human nephron number: Implications for health and disease. *Pediatr. Nephrol.* 2011, 26, 1529–1533. [CrossRef]
- Ibrahim, H.N.; E Rosenberg, M.; Hostetter, T.H. Role of the renin-angiotensin-aldosterone system in the progression of renal disease: A critical review. *Semin. Nephrol.* 1997, 17, 431–440.
- 25. Gagliardini, E.; Conti, S.; Benigni, A.; Remuzzi, G.; Remuzzi, A. Imaging of the Porous Ultrastructure of the Glomerular Epithelial Filtration Slit. *J. Am. Soc. Nephrol.* **2010**, *21*, 2081–2089. [CrossRef]
- Ruggenenti, P.; Cravedi, P.; Remuzzi, G. Mechanisms and Treatment of CKD. *J. Am. Soc. Nephrol.* 2012, 23, 1917–1928. [CrossRef]
 Durvasula, R.V.; Petermann, A.T.; Hiromura, K.; Blonski, M.; Pippin, J.; Mundel, P.; Pichler, R.; Griffin, S.; Couser, W.G.; Shankland, S.J. Activation of a local tissue angiotensin system in podocytes by mechanical strain. *Kidney Int.* 2004, *65*, 30–39. [CrossRef]
- Benigni, A.; Gagliardini, E.; Remuzzi, G. Changes in glomerular perm-selectivity induced by angiotensin II imply podocyte dysfunction and slit diaphragm protein rearrangement. *Semin. Nephrol.* 2004, 24, 131–140. [CrossRef]
- 29. Barnes, J.L.; Gorin, Y. Myofibroblast differentiation during fibrosis: Role of NAD(P)H oxidases. Kidney Int. 2011, 79, 944–956. [CrossRef]
- 30. Abbate, M.; Zoja, C.; Remuzzi, G. How Does Proteinuria Cause Progressive Renal Damage? J. Am. Soc. Nephrol. 2006, 17, 2974–2984. [CrossRef]
- Abbate, M.; Zoja, C.; Morigi, M.; Rottoli, D.; Angioletti, S.; Tomasoni, S.; Zanchi, C.; Longaretti, L.; Donadelli, R.; Remuzzi, G. Transforming Growth Factor-β1 Is Up-Regulated by Podocytes in Response to Excess Intraglomerular Passage of Proteins. *Am. J. Pathol.* 2002, *161*, 2179–2193. [CrossRef]
- 32. Johnson, D.W.; Saunders, H.J.; Baxter, R.C.; Field, M.J.; Pollock, C.A. Paracrine stimulation of human renal fibroblasts by proximal tubule cells. *Kidney Int.* **1998**, *54*, 747–757. [CrossRef]
- 33. Zeisberg, M.; Neilson, E.G. Mechanisms of Tubulointerstitial Fibrosis. J. Am. Soc. Nephrol. 2010, 21, 1819–1834. [CrossRef]
- 34. Schlondorff, D.O. Overview of factors contributing to the pathophysiology of progressive renal disease. *Kidney Int.* **2008**, 74, 860–866. [CrossRef]
- 35. Liu, Y. New Insights into Epithelial-Mesenchymal Transition in Kidney Fibrosis. J. Am. Soc. Nephrol. 2010, 21, 212–222. [CrossRef]
- Lovisa, S.; LeBleu, V.S.; Tampe, B.; Sugimoto, H.; Vadnagara, K.; Carstens, J.L.; Wu, C.-C.; Hagos, Y.; Burckhardt, B.C.; Pentcheva-Hoang, T.; et al. Epithelial-to-mesenchymal transition induces cell cycle arrest and parenchymal damage in renal fibrosis. *Nat. Med.* 2015, 21, 998–1009. [CrossRef]

- Grande, M.T.; Sánchez-Laorden, B.; López-Blau, C.; A De Frutos, C.; Boutet, A.; Arévalo, M.; Rowe, R.G.; Weiss, S.J.; López-Novoa, J.M.; Nieto, M.A. Snail1-induced partial epithelial-to-mesenchymal transition drives renal fibrosis in mice and can be targeted to reverse established disease. *Nat. Med.* 2015, *21*, 989–997. [CrossRef]
- Cano, A.; Pérez-Moreno, M.A.; Rodrigo, I.; Locascio, A.; Blanco, M.J.; Del Barrio, M.G.; Portillo, F.; Nieto, M.A. The transcription factor Snail controls epithelial–mesenchymal transitions by repressing E-cadherin expression. *Nat. Cell Biol.* 2000, 2, 76–83. [CrossRef]
- 39. Zhou, B.P.; Deng, J.; Xia, W.; Xu, J.; Li, Y.M.; Gunduz, M.; Hung, M.-C. Dual regulation of Snail by GSK-3β-mediated phosphorylation in control of epithelial–mesenchymal transition. *Nat. Cell Biol.* **2004**, *6*, 931–940. [CrossRef]
- 40. Widjaja, A.A.; Viswanathan, S.; Shekeran, S.G.; Adami, E.; Lim, W.-W.; Chothani, S.; Tan, J.; Goh, J.W.T.; Chen, H.M.; Lim, S.Y.; et al. Targeting endogenous kidney regeneration using anti-IL11 therapy in acute and chronic models of kidney disease. *Nat. Commun.* **2022**, *13*, 7497. [CrossRef]
- 41. Huang, S.; Susztak, K. Epithelial Plasticity versus EMT in Kidney Fibrosis. Trends Mol. Med. 2015, 22, 4–6. [CrossRef]
- Widjaja, A.A.; Viswanathan, S.; Ting, J.G.W.; Tan, J.; Shekeran, S.G.; Carling, D.; Lim, W.-W.; Cook, S.A. IL11 stimulates ERK/P90RSK to inhibit LKB1/AMPK and activate mTOR initiating a mesenchymal program in stromal, epithelial, and cancer cells. *iScience* 2022, 25, 104806. [CrossRef]
- 43. Strikoudis, A.; Cieślak, A.; Loffredo, L.; Chen, Y.-W.; Patel, N.; Saqi, A.; Lederer, D.J.; Snoeck, H.-W. Modeling of Fibrotic Lung Disease Using 3D Organoids Derived from Human Pluripotent Stem Cells. *Cell Rep.* **2019**, *27*, 3709–3723.e5. [CrossRef]
- 44. Chen, T.K.; Knicely, D.H.; Grams, M.E. Chronic Kidney Disease Diagnosis and Management. JAMA 2019, 322, 1294–1304. [CrossRef]
- 45. Inker, L.A.; Astor, B.C.; Fox, C.H.; Isakova, T.; Lash, J.P.; Peralta, C.A.; Kurella Tamura, M.; Feldman, H.I. KDOQI US Commentary on the 2012 KDIGO Clinical Practice Guideline for the Evaluation and Management of CKD. *Am. J. Kidney Dis.* **2014**, *63*, 713–735. [CrossRef]
- Stevens, P.E.; Levin, A. Evaluation and Management of Chronic Kidney Disease: Synopsis of the kidney disease: Improving Global Outcomes 2012 Clinical Practice Guideline. *Ann. Intern. Med.* 2013, 158, 825–830. [CrossRef]
- Whelton, P.K.; Carey, R.M.; Aronow, W.S.; Casey, D.E.; Collins, K.J.; Dennison Himmelfarb, C.; DePalma, S.M.; Gidding, S.; Jamerson, K.A.; Jones, D.W.; et al. 2017 ACC/AHA/AAPA/ABC/ACPM/AGS/APhA/ASH/ASPC/NMA/PCNA Guideline for the Prevention, Detection, Evaluation, and Management of High Blood Pressure in Adults: A Report of the American College of Cardiology/American Heart Association Task Force on Clinical Practice Guidelines. J. Am. Coll. Cardiol. 2018, 71, e127–e248.
- Fried, L.F.; Emanuele, N.; Zhang, J.H.; Brophy, M.; Conner, T.A.; Duckworth, W.; Leehey, D.J.; McCullough, P.A.; O'COnnor, T.; Palevsky, P.M.; et al. Combined Angiotensin Inhibition for the Treatment of Diabetic Nephropathy. *N. Engl. J. Med.* 2013, 369, 1892–1903. [CrossRef]
- 49. Guideline Development Group; Bilo, H.; Coentrão, L.; Couchoud, C.; Covic, A.; De Sutter, J.; Drechsler, C.; Gnudi, L.; Goldsmith, D.; Heaf, J.; et al. Clinical Practice Guideline on management of patients with diabetes and chronic kidney disease stage 3b or higher (eGFR < 45 mL/min). *Nephrol. Dial. Transplant.* 2015, 30, ii1–ii142.
- Shurraw, S.; Hemmelgarn, B.; Lin, M.; Majumdar, S.R.; Klarenbach, S.; Manns, B.; Bello, A.; James, M.; Turin, T.C.; Tonelli, M.; et al. Association Between Glycemic Control and Adverse Outcomes in People with Diabetes Mellitus and Chronic Kidney Disease. *Arch. Intern. Med.* 2011, 171, 1920–1927. [CrossRef]
- 51. UK Prospective Diabetes Study (UKPDS) Group. Intensive blood-glucose control with sulphonylureas or insulin compared with conventional treatment and risk of complications in patients with type 2 diabetes (UKPDS 33). *Lancet* **1998**, 352, 837–853. [CrossRef]
- Anderson, T.J.; Grégoire, J.; Pearson, G.J.; Barry, A.R.; Couture, P.; Dawes, M.; Francis, G.A.; Genest, J., Jr.; Grover, S.; Gupta, M.; et al. 2016 Canadian Cardiovascular Society Guidelines for the Management of Dyslipidemia for the Prevention of Cardiovascular Disease in the Adult. *Can. J. Cardiol.* 2016, 32, 1263–1282. [CrossRef]
- 53. Tonelli, M.; Wanner, C. Lipid Management in Chronic Kidney Disease: Synopsis of the kidney disease: Improving Global Outcomes 2013 Clinical Practice Guideline. *Ann. Intern. Med.* 2014, *160*, 182–189. [CrossRef]
- 54. James, P.A.; Oparil, S.; Carter, B.L.; Cushman, W.C.; Dennison-Himmelfarb, C.; Handler, J.; Lackland, D.T.; Lefevre, M.L.; MacKenzie, T.D.; Ogedegbe, O.; et al. 2014 Evidence-Based Guideline for the Management of High Blood Pressure in Adults: Report from the panel members appointed to the Eighth Joint National Committee (JNC 8). JAMA 2014, 311, 507–520. [CrossRef]
- 55. Thomas, B.; Matsushita, K.; Abate, K.H.; Al-Aly, Z.; Ärnlöv, J.; Asayama, K.; Atkins, R.; Badawi, A.; Ballew, S.H.; Banerjee, A.; et al. Global Cardiovascular and Renal Outcomes of Reduced GFR. J. Am. Soc. Nephrol. 2017, 28, 2167–2179. [CrossRef]
- 56. Agapova, O.A.; Fang, Y.; Sugatani, T.; Seifert, M.E.; Hruska, K.A. Ligand trap for the activin type IIA receptor protects against vascular disease and renal fibrosis in mice with chronic kidney disease. *Kidney Int.* **2016**, *89*, 1231–1243. [CrossRef]
- 57. Lau, A.; Rahn, J.J.; Chappellaz, M.; Chung, H.; Benediktsson, H.; Bihan, D.; von Mässenhausen, A.; Linkermann, A.; Jenne, C.N.; Robbins, S.M.; et al. Dipeptidase-1 governs renal inflammation during ischemia reperfusion injury. *Sci. Adv.* **2022**, *8*, eabm0142. [CrossRef]
- 58. Linkermann, A. Nonapoptotic cell death in acute kidney injury and transplantation. Kidney Int. 2016, 89, 46–57. [CrossRef]
- 59. Belavgeni, A.; Meyer, C.; Stumpf, J.; Hugo, C.; Linkermann, A. Ferroptosis and Necroptosis in the Kidney. *Cell Chem. Biol.* **2020**, 27, 448–462. [CrossRef]
- Lamade, A.M.; Wu, L.; Dar, H.H.; Mentrup, H.L.; Shrivastava, I.H.; Epperly, M.W.; Croix, C.M.S.; Tyurina, Y.Y.; Anthonymuthu, T.S.; Yang, Q.; et al. Inactivation of RIP3 kinase sensitizes to 15LOX/PEBP1-mediated ferroptotic death. *Redox Biol.* 2022, 50, 102232. [CrossRef]
- Tonnus, W.; Meyer, C.; Steinebach, C.; Belavgeni, A.; von Mässenhausen, A.; Gonzalez, N.Z.; Maremonti, F.; Gembardt, F.; Himmerkus, N.; Latk, M.; et al. Dysfunction of the key ferroptosis-surveilling systems hypersensitizes mice to tubular necrosis during acute kidney injury. *Nat. Commun.* 2021, *12*, 4402. [CrossRef] [PubMed]

- 62. Wang, Y.; Li, Y.; Xu, Y. Pyroptosis in Kidney Disease. J. Mol. Biol. 2021, 434, 167290. [CrossRef] [PubMed]
- 63. Elias, E.E.; Lyons, B.; Muruve, D.A. Gasdermins and pyroptosis in the kidney. Nat. Rev. Nephrol. 2023, 19, 337–350. [CrossRef] [PubMed]
- Tong, X.; Tang, R.; Xiao, M.; Xu, J.; Wang, W.; Zhang, B.; Liu, J.; Yu, X.; Shi, S. Targeting cell death pathways for cancer therapy: Recent developments in necroptosis, pyroptosis, ferroptosis, and cuproptosis research. J. Hematol. Oncol. 2022, 15, 174. [CrossRef] [PubMed]
- 65. Zhou, B.; Zhang, J.-Y.; Liu, X.-S.; Chen, H.-Z.; Ai, Y.-L.; Cheng, K.; Sun, R.-Y.; Zhou, D.; Han, J.; Wu, Q. Tom20 senses iron-activated ROS signaling to promote melanoma cell pyroptosis. *Cell Res.* **2018**, *28*, 1171–1185. [CrossRef]
- Zhang, K.-J.; Wu, Q.; Jiang, S.-M.; Ding, L.; Liu, C.-X.; Xu, M.; Wang, Y.; Zhou, Y.; Li, L. Pyroptosis: A New Frontier in Kidney Diseases. Oxidative Med. Cell. Longev. 2021, 2021, 6686617. [CrossRef]
- 67. Świerczyński, M.; Fichna, J. Inflamasom jako czynnik sprawczy i ochronny w patogenezie nieswoistych chorób zapalnych jelit. *Postępy Biochem.* **2021**, *67*, 44–53. [CrossRef]
- Shahzad, K.; Fatima, S.; Khawaja, H.; Elwakiel, A.; Gadi, I.; Ambreen, S.; Zimmermann, S.; Mertens, P.R.; Biemann, R.; Isermann, B. Podocyte-specific Nlrp3 inflammasome activation promotes diabetic kidney disease. *Kidney Int.* 2022, 102, 766–779. [CrossRef]
- 69. Liu, X.; Zhang, Z.; Ruan, J.; Pan, Y.; Magupalli, V.G.; Wu, H.; Lieberman, J. Inflammasome-activated gasdermin D causes pyroptosis by forming membrane pores. *Nature* **2016**, *535*, 153–158. [CrossRef]
- Wang, Y.; Chen, X.; Song, Y.; Caballero, B.; Cheskin, L. Association between obesity and kidney disease: A systematic review and meta-analysis. *Kidney Int.* 2008, 73, 19–33. [CrossRef]
- Sharma, I.; Liao, Y.; Zheng, X.; Kanwar, Y.S. New Pandemic: Obesity and Associated Nephropathy. Front. Med. 2021, 8, 673556. [CrossRef] [PubMed]
- 72. Câmara, N.O.S.; Iseki, K.; Kramer, H.; Liu, Z.-H.; Sharma, K. Kidney disease and obesity: Epidemiology, mechanisms and treatment. *Nat. Rev. Nephrol.* 2017, 13, 181–190. [CrossRef] [PubMed]
- 73. Sun, Y.; Ge, X.; Li, X.; He, J.; Wei, X.; Du, J.; Sun, J.; Li, X.; Xun, Z.; Liu, W.; et al. High-fat diet promotes renal injury by inducing oxidative stress and mitochondrial dysfunction. *Cell Death Dis.* **2020**, *11*, 914. [CrossRef] [PubMed]
- 74. Ren, L.; Cui, H.; Wang, Y.; Ju, F.; Cai, Y.; Gang, X.; Wang, G. The role of lipotoxicity in kidney disease: From molecular mechanisms to therapeutic prospects. *Biomed. Pharmacother.* **2023**, *161*, 114465. [CrossRef] [PubMed]
- 75. Gai, Z.; Wang, T.; Visentin, M.; Kullak-Ublick, G.A.; Fu, X.; Wang, Z. Lipid Accumulation and Chronic Kidney Disease. *Nutrients* 2019, *11*, 722. [CrossRef]
- 76. Kennedy, D.J.; Chen, Y.; Huang, W.; Viterna, J.; Liu, J.; Westfall, K.; Tian, J.; Bartlett, D.J.; Tang, W.W.; Xie, Z.; et al. CD36 and Na/K-ATPase-α1 Form a Proinflammatory Signaling Loop in Kidney. *Hypertension* **2013**, *61*, 216–224. [CrossRef]
- 77. Hua, W.; Huang, H.-Z.; Tan, L.-T.; Wan, J.-M.; Gui, H.-B.; Zhao, L.; Ruan, X.-Z.; Chen, X.-M.; Du, X.-G. CD36 Mediated Fatty Acid-Induced Podocyte Apoptosis via Oxidative Stress. *PLoS ONE* **2015**, *10*, e0127507. [CrossRef]
- Jung, J.H.; Choi, J.E.; Song, J.H.; Ahn, S.-H. Human CD36 overexpression in renal tubules accelerates the progression of renal diseases in a mouse model of folic acid-induced acute kidney injury. *Kidney Res. Clin. Pract.* 2018, 37, 30–40. [CrossRef]
- Kim, J.-J.; David, J.M.; Wilbon, S.S.; Santos, J.V.; Patel, D.M.; Ahmad, A.; Mitrofanova, A.; Liu, X.; Mallela, S.K.; Ducasa, G.M.; et al. Discoidin domain receptor 1 activation links extracellular matrix to podocyte lipotoxicity in Alport syndrome. *EBioMedicine* 2020, 63, 103162. [CrossRef]
- 80. Rysz, J.; Franczyk, B.; Ławiński, J.; Olszewski, R.; Ciałkowska-Rysz, A.; Gluba-Brzózka, A. The Impact of CKD on Uremic Toxins and Gut Microbiota. *Toxins* 2021, *13*, 252. [CrossRef]
- Ramezani, A.; Raj, D.S. The Gut Microbiome, Kidney Disease, and Targeted Interventions. J. Am. Soc. Nephrol. 2014, 25, 657–670. [CrossRef] [PubMed]
- Rapa, S.F.; Prisco, F.; Popolo, A.; Iovane, V.; Autore, G.; Di Iorio, B.R.; Dal Piaz, F.; Paciello, O.; Nishijima, F.; Marzocco, S. Pro-Inflammatory Effects of Indoxyl Sulfate in Mice: Impairment of Intestinal Homeostasis and Immune Response. *Int. J. Mol. Sci.* 2021, 22, 1135. [CrossRef] [PubMed]
- 83. Di Paola, R.; De, A.; Izhar, R.; Abate, M.; Zappavigna, S.; Capasso, A.; Perna, A.F.; La Russa, A.; Capasso, G.; Caraglia, M.; et al. Possible Effects of Uremic Toxins p-Cresol, Indoxyl Sulfate, p-Cresyl Sulfate on the Development and Progression of Colon Cancer in Patients with Chronic Renal Failure. *Genes* **2023**, *14*, 1257. [CrossRef] [PubMed]
- Vanholder, R.; Schepers, E.; Pletinck, A.; Nagler, E.V.; Glorieux, G. The Uremic Toxicity of Indoxyl Sulfate and p-Cresyl Sulfate: A Systematic Review. J. Am. Soc. Nephrol. 2014, 25, 1897–1907. [CrossRef] [PubMed]
- Liu, W.-C.; Tomino, Y.; Lu, K.-C. Impacts of Indoxyl Sulfate and p-Cresol Sulfate on Chronic Kidney Disease and Mitigating Effects of AST-120. *Toxins* 2018, 10, 367. [CrossRef] [PubMed]
- Ito, S.; Osaka, M.; Higuchi, Y.; Nishijima, F.; Ishii, H.; Yoshida, M. Indoxyl Sulfate Induces Leukocyte-Endothelial Interactions through Up-regulation of E-selectin. J. Biol. Chem. 2010, 285, 38869–38875. [CrossRef]
- Nakano, T.; Katsuki, S.; Chen, M.; Decano, J.L.; Halu, A.; Lee, L.H.; Pestana, D.V.S.; Kum, A.S.T.; Kuromoto, R.K.; Golden, W.S.; et al. Uremic Toxin Indoxyl Sulfate Promotes Proinflammatory Macrophage Activation Via the Interplay of OATP2B1 and Dll4-Notch Signaling. *Circulation* 2019, 139, 78–96. [CrossRef]
- 88. Watanabe, H.; Miyamoto, Y.; Otagiri, M.; Maruyama, T. Update on the Pharmacokinetics and Redox Properties of Protein-Bound Uremic Toxins. *J. Pharm. Sci.* 2011, 100, 3682–3695. [CrossRef]
- Shimizu, H.; Bolati, D.; Higashiyama, Y.; Nishijima, F.; Shimizu, K.; Niwa, T. Indoxyl sulfate upregulates renal expression of MCP-1 via production of ROS and activation of NF-κB, p53, ERK, and JNK in proximal tubular cells. *Life Sci.* 2012, 90, 525–530. [CrossRef] [PubMed]

- Matsuo, K.; Yamamoto, S.; Wakamatsu, T.; Takahashi, Y.; Kawamura, K.; Kaneko, Y.; Goto, S.; Kazama, J.J.; Narita, I. Increased Proinflammatory Cytokine Production and Decreased Cholesterol Efflux Due to Downregulation of ABCG1 in Macrophages Exposed to Indoxyl Sulfate. *Toxins* 2015, 7, 3155–3166. [CrossRef]
- Chang, J.-F.; Hsieh, C.-Y.; Liou, J.-C.; Liu, S.-H.; Hung, C.-F.; Lu, K.-C.; Lin, C.-C.; Wu, C.-C.; Ka, S.-M.; Wen, L.-L.; et al. Scavenging Intracellular ROS Attenuates *p*-Cresyl Sulfate-Triggered Osteogenesis through MAPK Signaling Pathway and NF-κB Activation in Human Arterial Smooth Muscle Cells. *Toxins* 2020, *12*, 472. [CrossRef] [PubMed]
- 92. Lim, Y.J.; Sidor, N.A.; Tonial, N.C.; Che, A.; Urquhart, B.L. Uremic Toxins in the Progression of Chronic Kidney Disease and Cardiovascular Disease: Mechanisms and Therapeutic Targets. *Toxins* **2021**, *13*, 142. [CrossRef] [PubMed]
- Shimizu, H.; Yisireyili, M.; Nishijima, F.; Niwa, T. Indoxyl Sulfate Enhances p53-TGF-β1-Smad3 Pathway in Proximal Tubular Cells. Am. J. Nephrol. 2013, 37, 97–103. [CrossRef]
- Shafi, T.; Meyer, T.W.; Hostetter, T.H.; Melamed, M.L.; Parekh, R.S.; Hwang, S.; Banerjee, T.; Coresh, J.; Powe, N.R. Free Levels of Selected Organic Solutes and Cardiovascular Morbidity and Mortality in Hemodialysis Patients: Results from the Retained Organic Solutes and Clinical Outcomes (ROSCO) Investigators. *PLoS ONE* 2015, *10*, e0126048. [CrossRef]
- 95. Al Hinai, E.A.; Kullamethee, P.; Rowland, I.R.; Swann, J.; Walton, G.E.; Commane, D.M. Modelling the role of microbial p-cresol in colorectal genotoxicity. *Gut Microbes* **2018**, *10*, 398–411. [CrossRef]
- 96. Li, J.; Zhao, F.; Wang, Y.; Chen, J.; Tao, J.; Tian, G.; Wu, S.; Liu, W.; Cui, Q.; Geng, B.; et al. Gut microbiota dysbiosis contributes to the development of hypertension. *Microbiome* **2017**, *5*, 14. [CrossRef] [PubMed]
- 97. Jiang, S.; Xie, S.; Lv, D.; Wang, P.; He, H.; Zhang, T.; Zhou, Y.; Lin, Q.; Zhou, H.; Jiang, J.; et al. Alteration of the gut microbiota in Chinese population with chronic kidney disease. *Sci. Rep.* **2017**, *7*, 2870. [CrossRef] [PubMed]
- Aronov, P.A.; Luo, F.J.-G.; Plummer, N.S.; Quan, Z.; Holmes, S.; Hostetter, T.H.; Meyer, T.W. Colonic Contribution to Uremic Solutes. J. Am. Soc. Nephrol. 2011, 22, 1769–1776. [CrossRef]
- 99. Hobby, G.P.; Karaduta, O.; Dusio, G.F.; Singh, M.; Zybailov, B.L.; Arthur, J.M. Chronic kidney disease and the gut microbiome. *Am. J. Physiol.* 2019, 316, F1211–F1217. [CrossRef]
- 100. Dou, L.; Bertrand, E.; Cerini, C.; Faure, V.; Sampol, J.; Vanholder, R.; Berland, Y.; Brunet, P. The uremic solutes p-cresol and indoxyl sulfate inhibit endothelial proliferation and wound repair. *Kidney Int.* **2004**, *65*, 442–451. [CrossRef]
- 101. Guldris, S.C.; Parra, E.G.; Amenós, A.C. Gut microbiota in chronic kidney disease. Nefrología 2017, 37, 9–19. [CrossRef]
- Vaziri, N.D.; Wong, J.; Pahl, M.; Piceno, Y.M.; Yuan, J.; DeSantis, T.Z.; Ni, Z.; Nguyen, T.-H.; Andersen, G.L. Chronic kidney disease alters intestinal microbial flora. *Kidney Int.* 2013, *83*, 308–315. [CrossRef] [PubMed]
- 103. Zhao, J.; Ning, X.; Liu, B.; Dong, R.; Bai, M.; Sun, S. Specific alterations in gut microbiota in patients with chronic kidney disease: An updated systematic review. *Ren. Fail.* 2021, 43, 102–112. [CrossRef] [PubMed]
- Hu, X.; Ouyang, S.; Xie, Y.; Gong, Z.; Du, J. Characterizing the gut microbiota in patients with chronic kidney disease. *Postgrad. Med.* 2020, 132, 495–505. [CrossRef] [PubMed]
- 105. Wu, I.-W.; Lin, C.-Y.; Chang, L.-C.; Lee, C.-C.; Chiu, C.-Y.; Hsu, H.-J.; Sun, C.-Y.; Chen, Y.-C.; Kuo, Y.-L.; Yang, C.-W.; et al. Gut Microbiota as Diagnostic Tools for Mirroring Disease Progression and Circulating Nephrotoxin Levels in Chronic Kidney Disease: Discovery and Validation Study. Int. J. Biol. Sci. 2020, 16, 420–434. [CrossRef]
- 106. Ren, Z.; Fan, Y.; Li, A.; Shen, Q.; Wu, J.; Ren, L.; Lu, H.; Ding, S.; Ren, H.; Liu, C.; et al. Alterations of the Human Gut Microbiome in Chronic Kidney Disease. *Adv. Sci.* 2020, *7*, 2001936. [CrossRef]
- 107. Kim, J.E.; Kim, H.-E.; Park, J.I.; Cho, H.; Kwak, M.-J.; Kim, B.-Y.; Yang, S.H.; Lee, J.P.; Kim, D.K.; Joo, K.W.; et al. The Association between Gut Microbiota and Uremia of Chronic Kidney Disease. *Microorganisms* **2020**, *8*, 907. [CrossRef]
- 108. Gryp, T.; Huys, G.R.; Joossens, M.; Van Biesen, W.; Glorieux, G.; Vaneechoutte, M. Isolation and Quantification of Uremic Toxin Precursor-Generating Gut Bacteria in Chronic Kidney Disease Patients. *Int. J. Mol. Sci.* 2020, 21, 1986. [CrossRef]
- Zhao, Z.; Zhang, W.; Guo, R.; Cui, W.; Zhang, F.; Geng, Y.; Liu, C.; Shang, J.; Xiao, J.; Wen, X. Gut Microbiome Alterations Predict Diabetic Kidney Disease in General Population. *Res. Sq.* 2020, 10. [CrossRef]
- 110. Alatriste, P.V.M.; Arronte, R.U.; Espinosa, C.O.G.; Espinosa Cuevas, M.D.L.A. Efecto de lactobacillus casei shirota sobre concentraciones de urea en la enfermedad renal crónica. *Nutr. Hosp.* **2014**, *29*, 582–590.
- 111. Neto, M.P.C.; Aquino, J.d.S.; Silva, L.d.F.R.d.; Silva, R.d.O.; Guimarães, K.S.d.L.; de Oliveira, Y.; de Souza, E.L.; Magnani, M.; Vidal, H.; Alves, J.L.d.B. Gut microbiota and probiotics intervention: A potential therapeutic target for management of cardiometabolic disorders and chronic kidney disease? *Pharmacol. Res.* 2018, 130, 152–163. [CrossRef] [PubMed]
- 112. Lopes, R.d.C.S.O. Modulation of Intestinal Microbiota, Control of Nitrogen Products and Inflammation by Pre/Probiotics in Chronic Kidney Disease: A Systematic Review. *Nutr. Hosp.* **2018**, *35*, 722–730. [CrossRef] [PubMed]
- Chudzik, A.; Orzyłowska, A.; Rola, R.; Stanisz, G.J. Probiotics, Prebiotics and Postbiotics on Mitigation of Depression Symptoms: Modulation of the Brain–Gut–Microbiome Axis. *Biomolecules* 2021, *11*, 1000. [CrossRef] [PubMed]
- 114. Kolida, S.; Gibson, G.R. Synbiotics in Health and Disease. Annu. Rev. Food Sci. Technol. 2011, 2, 373–393. [CrossRef]
- Gou, H.-Z.; Zhang, Y.-L.; Ren, L.-F.; Li, Z.-J.; Zhang, L. How do intestinal probiotics restore the intestinal barrier? *Front. Microbiol.* 2022, 13, 929346. [CrossRef]
- 116. Virk, M.S.; Virk, M.A.; He, Y.; Tufail, T.; Gul, M.; Qayum, A.; Rehman, A.; Rashid, A.; Ekumah, J.-N.; Han, X.; et al. The Anti-Inflammatory and Curative Exponent of Probiotics: A Comprehensive and Authentic Ingredient for the Sustained Functioning of Major Human Organs. *Nutrients* 2024, *16*, 546. [CrossRef]

- 117. Tokarek, J.; Budny, E.; Saar, M.; Stańczak, K.; Wojtanowska, E.; Młynarska, E.; Rysz, J.; Franczyk, B. Molecular Processes Involved in the Shared Pathways between Cardiovascular Diseases and Diabetes. *Biomedicines* **2023**, *11*, 2611. [CrossRef] [PubMed]
- 118. E Cooper, T.; Khalid, R.; Chan, S.; Craig, J.C.; Hawley, C.M.; Howell, M.; Johnson, D.W.; Jaure, A.; Teixeira-Pinto, A.; Wong, G. Synbiotics, prebiotics and probiotics for people with chronic kidney disease. *Cochrane Database Syst. Rev.* **2023**, 2023, CD013631.
- Kuo, Y.-W.; Huang, Y.-Y.; Tsai, S.-Y.; Wang, J.-Y.; Lin, J.-H.; Syu, Z.-J.; Wang, H.-S.; Hsu, Y.-C.; Chen, J.-F.; Hsia, K.-C.; et al. Probiotic Formula Ameliorates Renal Dysfunction Indicators, Glycemic Levels, and Blood Pressure in a Diabetic Nephropathy Mouse Model. *Nutrients* 2023, 15, 2803. [CrossRef]
- Chávez-Íñiguez, J.S.; Ibarra-Estrada, M.; Gallardo-González, A.M.; Cisneros-Hernández, A.; Granado, R.C.-D.; Chávez-Alonso, G.; Hernández-Barajas, E.M.; Romero-Muñoz, A.C.; Ramos-Avellaneda, F.; Prieto-Magallanes, M.L.; et al. Probiotics in septic acute kidney injury, a double blind, randomized control trial. *Ren. Fail.* 2023, 45, 2260003. [CrossRef]
- Ogawa, T.; Shimada, M.; Nagano, N.; Ito, K.; Ando, T.; Shimomura, Y.; Ando, Y.; Otsuka, K. Oral administration of Bifidobacterium longum in a gastro-resistant seamless capsule decreases serum phosphate levels in patients receiving haemodialysis. *Clin. Kidney J.* 2012, 5, 373–374. [CrossRef] [PubMed]
- 122. Nakabayashi, I.; Nakamura, M.; Kawakami, K.; Ohta, T.; Kato, I.; Uchida, K.; Yoshida, M. Effects of synbiotic treatment on serum level of p-cresol in haemodialysis patients: A preliminary study. *Nephrol. Dial. Transplant.* **2010**, *26*, 1094–1098. [CrossRef]
- 123. Cruz-Mora, J.; Martínez-Hernández, N.E.; Martindel Campo-Lopez, F.; Viramontes-Hörner, D.; Vizmanos-Lamotte, B.; Muñoz-Valle, J.F.; García-García, G.; Parra-Rojas, I.; Castro-Alarcón, N. Effects of a Symbiotic on Gut Microbiota in Mexican Patients with End-Stage Renal Disease. J. Ren. Nutr. 2014, 24, 330–335. [CrossRef] [PubMed]
- 124. Viramontes-Hörner, D.; Márquez-Sandoval, F.; Martín-Del-Campo, F.; Vizmanos-Lamotte, B.; Sandoval-Rodríguez, A.; Armendáriz-Borunda, J.; García-Bejarano, H.; Renoirte-López, K.; García-García, G. Effect of a Symbiotic Gel (Lactobacillus acidophilus + Bifidobacterium lactis + Inulin) on Presence and Severity of Gastrointestinal Symptoms in Hemodialysis Patients. *J. Ren. Nutr.* 2015, 25, 284–291. [CrossRef]
- 125. Guida, B.; Germanò, R.; Trio, R.; Russo, D.; Memoli, B.; Grumetto, L.; Barbato, F.; Cataldi, M. Effect of short-term synbiotic treatment on plasma p-cresol levels in patients with chronic renal failure: A randomized clinical trial. *Nutr. Metab. Cardiovasc. Dis.* 2014, 24, 1043–1049. [CrossRef]
- 126. Rossi, M.; Johnson, D.W.; Morrison, M.; Pascoe, E.M.; Coombes, J.S.; Forbes, J.M.; Szeto, C.-C.; McWhinney, B.C.; Ungerer, J.P.; Campbell, K.L. Synbiotics Easing Renal Failure by Improving Gut Microbiology (SYNERGY): A Randomized Trial. *Clin. J. Am. Soc. Nephrol.* 2016, 11, 223–231. [CrossRef]
- 127. Simenhoff, M.L.; Dunn, S.R.; Zollner, G.P.; Fitzpatrick, M.E.; Emery, S.M.; Sandine, W.E.; Ayres, J.W. Biomodulation of the Toxic and Nutritional Effects of Small Bowel Bacterial Overgrowth in End-Stage Kidney Disease Using Freeze-Dried Lacto-bacillus Acidophilus. *Miner. Electrolyte Metab.* **1996**, *22*, 92–96. [PubMed]
- 128. Takayama, F.; Taki, K.; Niwa, T. Bifidobacterium in gastro-resistant seamless capsule reduces serum levels of indoxyl sulfate in patients on hemodialysis. *Am. J. Kidney Dis.* 2003, *41*, S142–S145. [CrossRef] [PubMed]
- 129. Taki, K.; Takayama, F.; Niwa, T. Beneficial effects of Bifidobacteria in a gastroresistant seamless capsule on hyperhomocysteinemia in hemodialysis patients. *J. Ren. Nutr.* **2005**, *15*, 77–80. [CrossRef]
- 130. Ando, Y.; Miyata, Y.; Tanba, K.; Saito, O.; Muto, S.; Kurosu, M.; Homma, S.; Kusano, E.; Asano, Y. Effect of oral intake of an enteric capsule preparation containing Bifidobacterium longum on the progression of chronic renal failure. *Nihon Jinzo Gakkai Shi* **2003**, 45, 759–764.
- 131. Hida, M.; Aiba, Y.; Sawamura, S.; Suzuki, N.; Satoh, T.; Koga, Y. Inhibition of the Accumulation of Uremic Toxins in the Blood and Their Precursors in the Feces after Oral Administration of Lebenin[®], a Lactic Acid Bacteria Preparation, to Uremic Patients Undergoing Hemodialysis. *Nephron* **1996**, *74*, 349–355. [CrossRef] [PubMed]
- Natarajan, R.; Pechenyak, B.; Vyas, U.; Ranganathan, P.; Weinberg, A.; Liang, P.; Mallappallil, M.C.; Norin, A.J.; Friedman, E.A.; Saggi, S.J. Randomized Controlled Trial of Strain-Specific Probiotic Formulation (Renadyl) in Dialysis Patients. *BioMed Res. Int.* 2014, 2014, 568571. [CrossRef] [PubMed]
- 133. Wang, I.-K.; Wu, Y.-Y.; Yang, Y.-F.; Ting, I.-W.; Lin, C.-C.; Yen, T.-H.; Chen, J.-H.; Wang, C.-H.; Huang, C.-C.; Lin, H.-C. The effect of probiotics on serum levels of cytokine and endotoxin in peritoneal dialysis patients: A randomised, double-blind, placebo-controlled trial. *Benef. Microbes* **2015**, *6*, 423–430. [CrossRef] [PubMed]
- 134. Ranganathan, N.; Friedman, E.A.; Tam, P.; Rao, V.; Ranganathan, P.; Dheer, R. Probiotic dietary supplementation in patients with stage 3 and 4 chronic kidney disease: A 6-month pilot scale trial in Canada. *Curr. Med. Res. Opin.* 2009, 25, 1919–1930. [CrossRef]
- 135. Ranganathan, N.; Ranganathan, P.; Friedman, E.A.; Joseph, A.; Delano, B.; Goldfarb, D.S.; Tam, P.; Rao, A.V.; Anteyi, E.; Musso, C.G. Pilot study of probiotic dietary supplementation for promoting healthy kidney function in patients with chronic kidney disease. *Adv. Ther.* 2010, 27, 634–647. [CrossRef]

Disclaimer/Publisher's Note: The statements, opinions and data contained in all publications are solely those of the individual author(s) and contributor(s) and not of MDPI and/or the editor(s). MDPI and/or the editor(s) disclaim responsibility for any injury to people or property resulting from any ideas, methods, instructions or products referred to in the content.