

Communication

Synthesis of New 1,2,3,4-Tetrahydroquinoline Hybrid of Ibuprofen and Its Biological Evaluation

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Abstract: Herein we report the obtaining of 1-(3,4-dihydroquinolin-1(2H)-yl)-2-(4-isobutylphenyl)propan-1-one and its characterization. The newly obtained hybrid and its derivatives (hybrids of ibuprofen with 1,2,3,4-tetrahydroisoquinoline, and piperidine) were screened for their in vitro antioxidant, antitryptic, and inhibition of albumin denaturation activity. The lipophilicity was established using both reversed-phase thin layer chromatography and in silico calculations.

Keywords: quinoline; ibuprofen; hybrid molecule; in silico; in vitro; antioxidant; antitryptic; anti-inflammatory; lipophilicity; SARS-CoV-2; chloroquine

1. Introduction

Benzo[b]pyridine or quinoline **1** in Figure 1 is one of the most prominent organic skeletons in medicinal chemistry practice and remains its interest until nowadays due to its pharmacological properties.

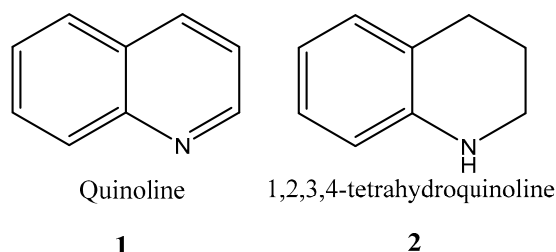


Figure 1. Structural formula of quinoline and 1,2,3,4-tetrahydroquinoline.

The quinoline fragment is part of the vast array of natural and synthetic medicines used in medical practice with diverse biological activities such as anti-inflammatory [1,2], anticancer [3], antimalarial [4,5], etc. The most outstanding performance for the quinoline fragment in drug development reveals its antimalarial and anticancer action, according to a large share of all the bioactive quinoline compounds known.

Recently, few drugs become even more popular in connection with the treatment of SARS-CoV-2 (COVID 19). In between those medicines are quinine **3**, chloroquine **4**, hydroxychloroquine **5**, and others are also part of the family of drugs containing a quinoline (given in red) fragment in its structure (Figure 2).

Discovered in 1934 by H. Andersag chloroquine is a drug primarily used to treat malaria. Together with its derivatives chloroquine were used in the treatment of many other conditions such as HIV, rheumatoid arthritis, and systemic lupus erythematosus [6]. Chloroquine **4** blocks up the production of hemozoin by the parasite inside red blood cells, releasing free heme. Then, the complexed heme with chloroquine exerts a toxic effect on the plasma membrane of the *Plasmodium species* in order to treat malaria [7,8].



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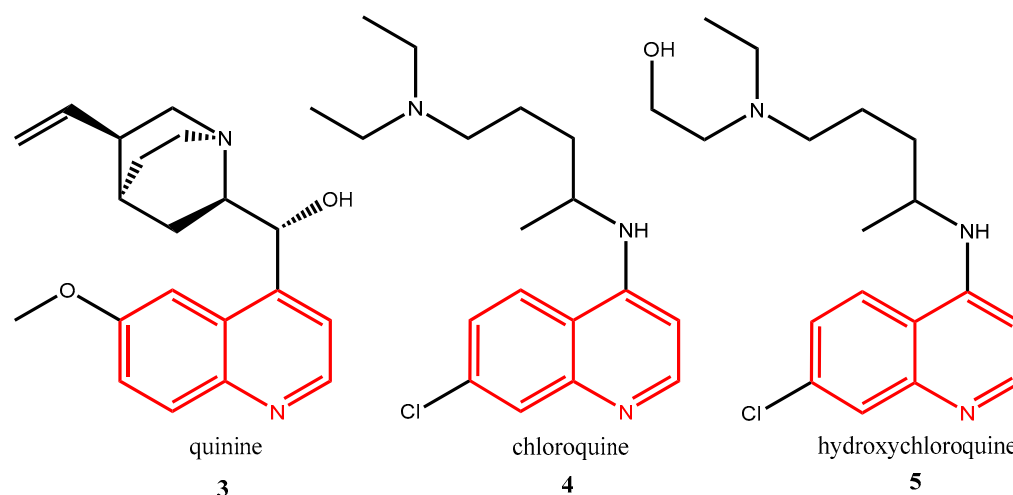


Figure 2. Structural formulas of quinine 3, chloroquine 4, and hydroxychloroquine 5.

Likewise, the 1,2,3,4-tetrahydroquinoline skeleton is prevalent in many pharmacologically active synthetic and natural products [9]. Another interesting example is the 2,4,6-trisubstituted tetrahydroquinoline 6 (Figure 3) isolated from *Martinella iquitosensis*. (*E*)-4-guanidino-2-methylbut-2-en-1-yl(2*S*,4*S*)-4-(1-ethyl-3-(3-methylbut-2-en-1-yl)guanidino-2-(3-(5-methylhex-4-enimidamido)propyl)-1,2,3,4-tetrahydroquinoline-6-carboxylate exhibits activity as bradykinin antagonist and with α -adrenergic, histaminergic, and muscarinic receptors.

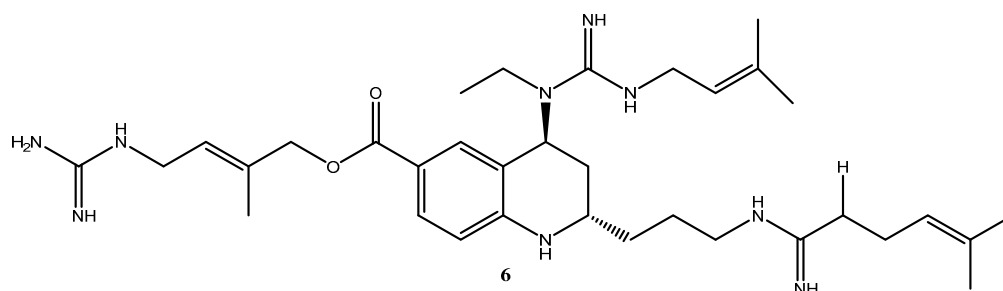


Figure 3. Structural formula of (*E*)-4-guanidino-2-methylbut-2-en-1-yl(2*S*,4*S*)-4-(1-ethyl-3-(3-methylbut-2-en-1-yl) guanidino)-2-(3-(5-methylhex-4-enimidamido)propyl)-1,2,3,4-tetrahydroquinoline-6-carboxylate.

Many tetrahydroquinolines owing various simple or complex substituents have interesting biochemical activity; some are potential pharmaceutical agents. Thus, a very simple derivative, 2-methyl-5-hydroxy-1,2,3,4-tetrahydroquinoline, exhibits analgesic activity one-eighth as potent as morphine. There are many examples that could be given for the rich and varied biological activity of 1,2,3,4-tetrahydroquinoline derivatives [10].

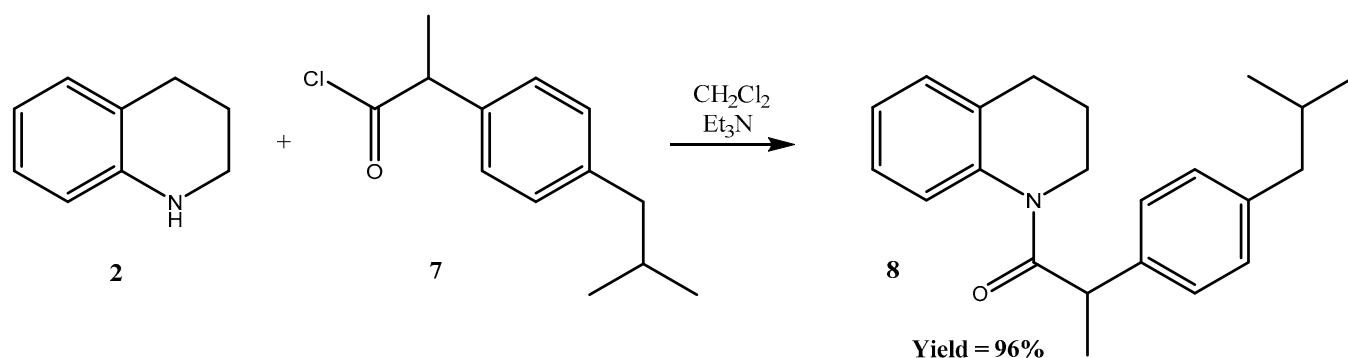
Ibuprofen, on the other hand, is maybe the most popular non-steroidal anti-inflammatory drug, used in the treatment of many musculoskeletal disorders [11], and is the safest traditional choice for utilization during chronic neuroinflammation such as in Alzheimer's [12], Parkinson's [13], and Machado–Joseph disease [14]. Hybrids of ibuprofen with quinoline are interesting [11] in order to study their potential biological activity, and to answer the question of whether they will combine the properties of both parent molecules or not, as we know the carboxylic functional group is not required for the anti-inflammatory properties of profens [15] but for causing gastric toxicity.

Due to the importance of the creation of new hybrid molecules in the medicinal chemistry industry, we are searching for easy, fast, and low-cost ways for the preparation of new drug candidates. The obtaining of exactly this hybrid molecule can lead to the biological properties of the constituent fragments (ibuprofen and 1,2,3,4-tetrahydroquinoline) to be combined in the new one.

2. Results

2.1. Synthesis

Herein we report the successfully synthesized 1-(3,4-dihydroquinolin-1(2H)-yl)-2-(4-isobutylphenyl)propan-1-one **8**, as shown in Scheme 1.



Scheme 1. Synthesis of hybrid molecule **8**.

In order to obtain the new hybrid molecule **8** we have started using coupling agents such as DCC and EDC to activate the carboxylic group of ibuprofen using various conditions, but unsuccessfully. We obtained the target compound **8** successfully by acylation of 1,2,3,4-tetrahydroquinoline **2** with 2-(4-isobutylphenyl)propanoyl chloride **7**. The reaction generally works in a high yield.

The resultant compound is characterized by UV, ^1H - and ^{13}C -NMR, HPLC and HRMS spectra.

Using the same procedure, we have synthesized two other ibuprofen hybrids already described in the literature in order to evaluate and compare their potential biological properties with the newly obtained ibuprofen hybrid **8**. We obtained the 1-(3,4-dihydroisoquinolin-2(1H)-yl)-2-(4-isobutylphenyl)propan-1-one **9** [16], and 2-(4-isobutylphenyl)-1-(piperidin-1-yl)propan-1-one **10** [17,18] (Figure 4).

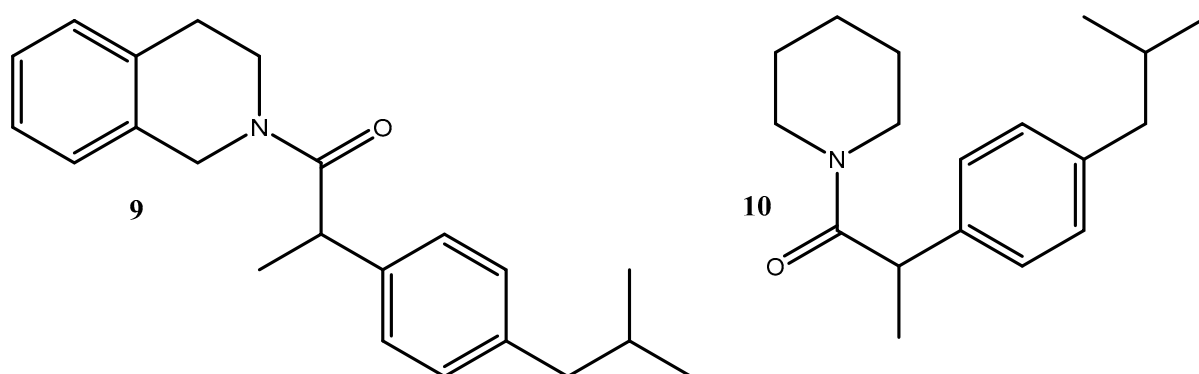


Figure 4. Structural formulas of hybrid molecules of ibuprofen with 1,2,3,4-tetrahydroisoquinoline **9** and piperidine **10**.

2.2. Biological Evaluation

All three compounds **8**, **9**, and **10** were tested for their *in vitro* inhibition albumin denaturation (IAD), antioxidant, and antitryptic activity (ATA). The obtained *in vitro* results were compared to the predicted *in silico* ones. The results are presented in Table 1. In this study, we have investigated the effect of substituents in the structure of pharmacophore I, while pharmacophore II did not change structurally.

Table 1. In vitro and in silico biological activity results. Hydrogen peroxide scavenging activity (HPSA), inhibition of albumin denaturation (IAD), and antitryptic activity (ATA) were expressed as IC_{50} . Ascorbic acid (AA), quercetin (Qrc), ibuprofen (Ibu) and ketoprofen (Ket) were used as standards. R_M is a non-dimensional quantity because it is a function of R_f and is determined by thin-layer chromatography. cAnti-I and cAnti-A are expressed as P_a (probability “to be active”) estimates the chance that the studied compound is belonging to the sub-class of active compounds (resembles the structures of molecules, which are the most typical in a sub-set of “actives” in PASS training set). The value of the most active compound is 1; R_M -lipophilicity; cAnti-I – calculated anti-inflammatory activity; cAnti-A–calculated anti-arthritic activity.

Compounds	$IC_{50} \pm SD, \mu g/mL$			$R_M \pm SD$	cLogP	P_a	
	HPSA	IAD	ATA			cAnti-I	cAnti-A
AA	24.84 ± 0.35	-	-	-	-	-	-
Qrc	69.25 ± 1.82	-	-	-	-	-	-
Ibu	-	81.50 ± 4.95	259.82 ± 9.14	1.11 ± 0.010	3.72	0.903	0.573
Ket	-	126.58 ± 5.00	720.57 ± 19.78	1.54 ± 0.015	3.59	0.925	0.469
H1	112.55 ± 2.32	90.23 ± 0.32	631.03 ± 41.88	1.04 ± 0.015	4.47	0.565	-
H2	103.76 ± 2.61	77.38 ± 0.55	270.36 ± 20.85	1.33 ± 0.015	5.78	0.526	-
H3	98.06 ± 7.17	92.08 ± 1.21	263.00 ± 14.48	1.14 ± 0.015	4.96	0.505	-

For the purpose of the biological evaluation study, we have named the three compounds hybrid 1 (H1), hybrid 2 (H2), and hybrid 3 (H3) as shown in Figure 5. Pharmacophore II in all molecules is ibuprofen residue while pharmacophore I varies. Pharmacophore I at hybrid 1 is piperidine, at hybrid 2 is 1,2,3,4-tetrahydroquinoline, and 1,2,3,4-tetrahydroisoquinoline is pharmacophore I at the third hybrid.

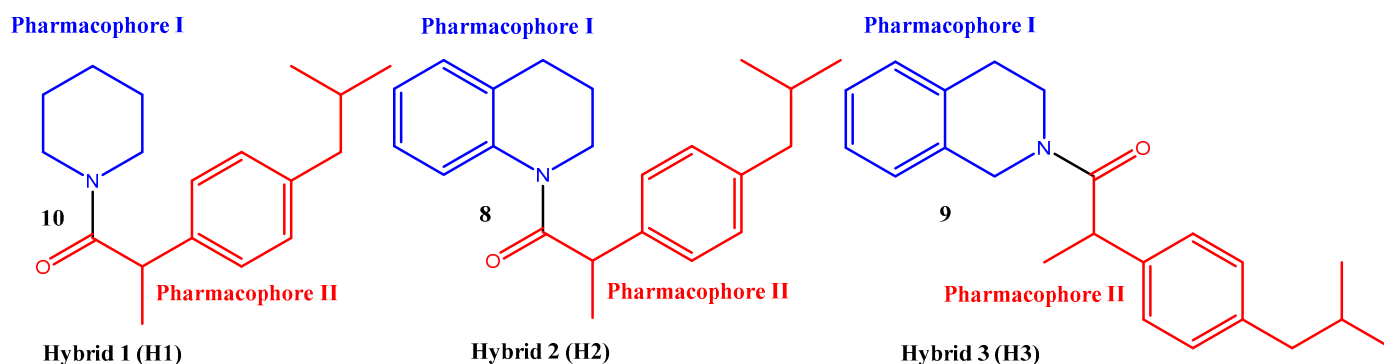


Figure 5. Structure of ibuprofen hybrids formed by two pharmacophores. Pharmacophore I (blue)–as the structures of piperidine, 1,2,3,4-tetrahydroquinoline and 1,2,3,4-tetrahydroisoquinoline, Pharmacophore II (red)–the structure of ibuprofen.

2.3. Hydrogen Peroxide Scavenging Activity (HPSA)

Reactive oxygen systems (ROS) are chemically reactive oxygen-containing radicals and molecules [super oxide ($O_2^{\bullet-}$), hydroxylene ($\bullet OH$), peroxy (ROO^{\bullet}) and alkoxy (RO^{\bullet}), HOCl, ozone (O_3), peroxyxynitrite $ONOO^-$], singlet oxygen (1O_2) and H_2O_2]. They are naturally formed as a by-product of cellular metabolism. Under physiological conditions, enzyme systems regulate ROS levels.

They are known for damaging vital molecules of biological importance such as phospholipids, proteins, and DNA. The damage caused by them has been shown to be involved in the development of a number of diseases (cancer, cardiovascular disease, atherosclerosis, and Alzheimer’s disease) [19]. Even in a state of physiological health, the toxic effects of oxygen and its derivatives, accumulating in the body, lead to a reduction in life expectancy [20].

In the present work, research has focused on the deactivation of hydrogen peroxide. Hydrogen peroxide is an oxidant that is continuously formed in living tissues as a result of several metabolic processes. However, its detoxification is very important in preventing it from getting into harmful reactions, such as the Fenton reaction [21].

The inflammatory process also causes and accelerates the formation of ROS. Mostly the formation of a superoxide anionic radical at the site of inflammation and this is related to the formation of other ROS species, such as H_2O_2 .

It is also involved in reductive decomposition reactions of hydrogen peroxide (so-called Haber–Weiss reaction) and organic hydroperoxides ROOH, assuming that at least some of the oxygen produced in these reactions is in the singlet state [22,23]. For this reason, the removal of H_2O_2 is very important to prevent the generation of $\bullet OH$ and the protection of biological systems.

The obtained results of antioxidant activity of the synthetic analogs of ibuprofen were compared to the standards of ascorbic acid and quercetin, as they are natural compounds with proven antioxidant properties.

Compared to ascorbic acid ($24.84 \mu\text{g/mL}$) and quercetin ($69.25 \mu\text{g/mL}$) the obtained ibuprofen derivatives demonstrated lower in vitro antioxidant activity. Compound **H3** ($98.06 \mu\text{g/mL}$) demonstrates higher antioxidant activity, compared to the rest of the synthesized compound (**H1**, **H2**) (Table 1, Figure 6). In the structure of the evaluated compounds, the presence of benzene nuclei increases the antioxidant effect. Therefore, compounds **H2**, and **H3** show better activity compared to **H1**.

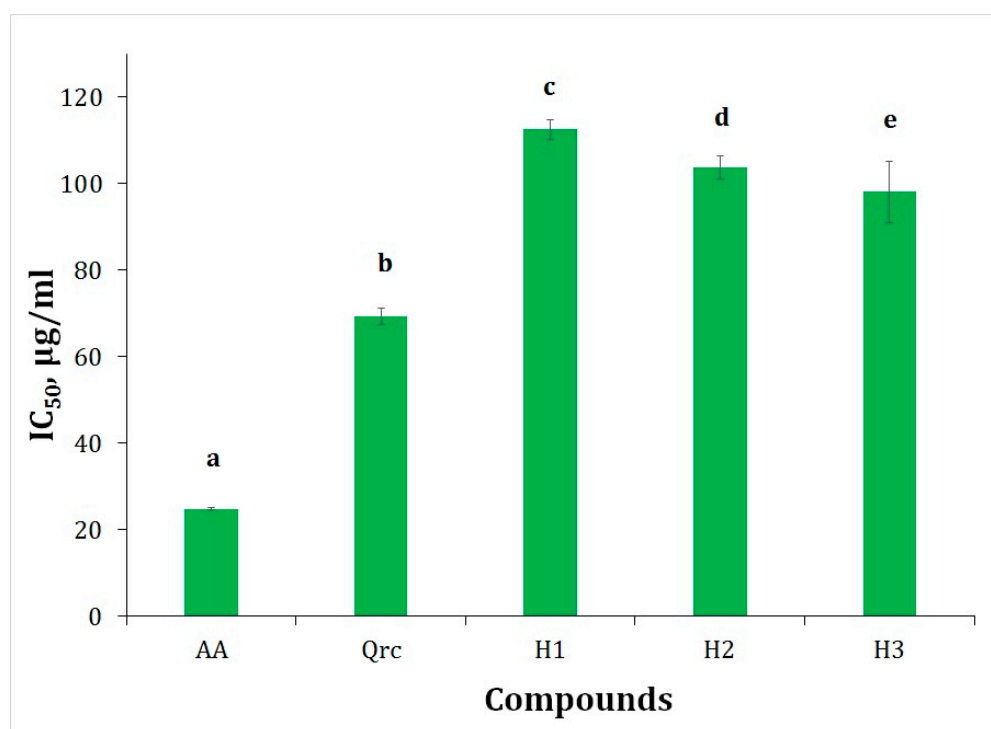


Figure 6. HPSA of ibuprofen derivatives. Ascorbic acid (AA) and quercetin (Qrc) were used as standards. The results of antioxidant activity were expressed as IC_{50} . Different letters for the same method indicate significant difference at $p < 0.05$ levels by Duncan's test.

The presence of OH groups in the structure of the compounds affects their antioxidant activity. That is the reason why ascorbic acid and quercetin are characterized by very good antioxidant activity. In addition, there are free ortho and para positions in both phenolic nuclei in the quercetin structure, while in the structure of the tetrahydroquinoline derivative next to the nitrogen atom there is only one para and ortho position. That is why the analyzed hybrid molecules are showing lower HPSA compared to the quercetin standard.

Sroka reported that the substitution of hydroxyls is the most important factor in increasing the antioxidant and hydrogen peroxide scavenging activities of compounds with two OH groups substituted to the phenolic ring [24].

2.4. Inhibition of Albumin Denaturation (IAD)

Inflammation is the response of living tissues to injury. It involves a complex array of enzyme activation, mediator release, fluid extravasation, cell migration, tissue breakdown, and repair [25]. The denaturation of proteins is a well-documented cause of inflammation in rheumatoid arthritis. Several anti-inflammatory drugs have shown a dose-dependent ability to inhibit thermally induced protein denaturation [26]. The obtained ibuprofen derivatives were screened for the inhibition of albumin denaturation. This method provides the extent information on to which albumin is protected from denaturation when heated. For this purpose, we have used human albumin. The percentages of inhibition of synthesized ibuprofen derivatives are presented in Figure 7. The results of the study are presented as IC_{50} . As ibuprofen and ketoprofen have proven properties, we have decided to use them as a benchmark to compare the activities of newly synthesized ibuprofen derivatives. The IC_{50} values of ibuprofen and ketoprofen estimated as IAD is 81.50 $\mu\text{g}/\text{mL}$ and 126.58 $\mu\text{g}/\text{mL}$, respectively, (Table 1, Figure 7). All obtained results show the IC_{50} values of ibuprofen derivatives are in the range from 77.38 to 92.08 $\mu\text{g}/\text{mL}$ (Table 1, Figure 7). The IAD of ibuprofen derivatives was assessed using the Duncan test. This test allows a statistical assessment to be made between the mean IADs of the compounds (**H1**, **H2**, and **H3**), and the standards (ibuprofen and ketoprofen). The observed data show that ibuprofen derivatives are more active than ketoprofen. The analysis of variance using Duncan test (with a significance level $p < 0.05$) revealed that there were not significant differences of the IAD values between ibuprofen and **H2**, and between **H1** and **H3**.

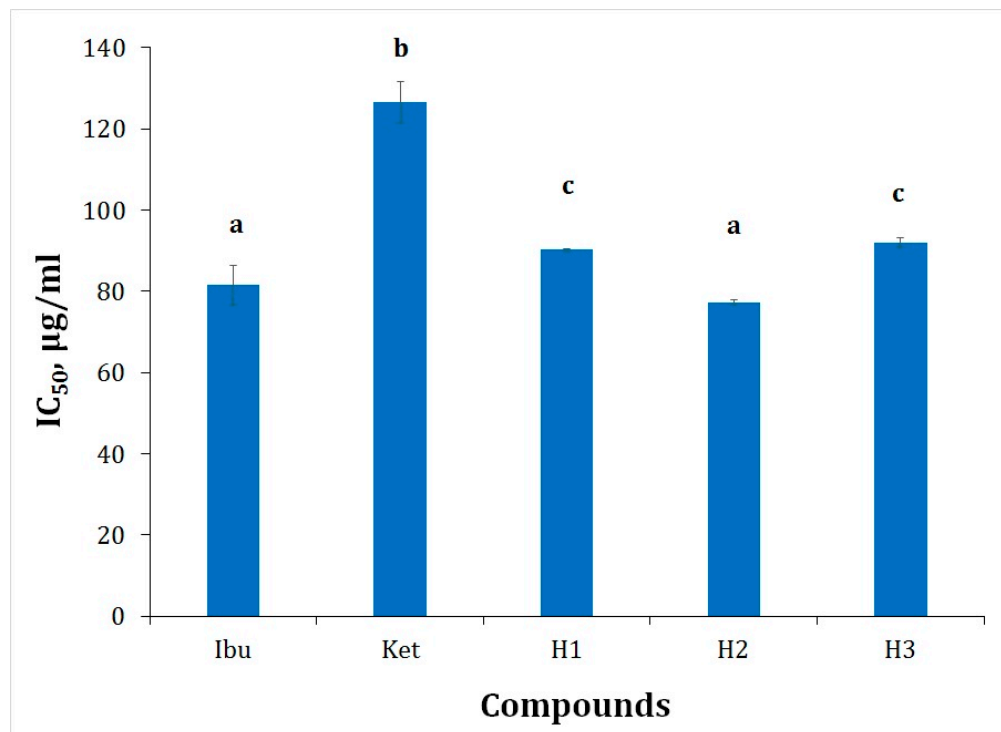


Figure 7. IAD of ibuprofen derivatives. Ibuprofen (Ibu) and ketoprofen (Ket) were used as standards. The results of IAD were expressed as IC_{50} . Different letters for the same method indicate significant difference at $p < 0.05$ levels by Duncan's test.

In general, the resulting compounds (**H1-3**) exhibit high IAD activity as ibuprofen. The calculated values for anti-inflammatory activity (cAnti-A) show that the standards

(ibuprofen and ketoprofen) are characterized by higher activity than the synthesized derivatives (**H1-3**), i.e., an inverse dependence is observed between in vitro and in silico (Table 1). There is a well-defined relationship between R_M and cLogP (Table 1).

Furthermore, IAD analysis reveals that lipophilicity is a major physicochemical parameter. The studied synthetic ibuprofen derivatives show lipophilicity (R_M) in the range between 1.04 and 1.33, which to some extent affects albumin protection. R_M value of **H2** (1.33) shows a greater effect on the stabilization of the albumin molecule (Table 1).

2.5. Antitryptic Activity (ATA)

Proteinases have been implicated in arthritic reactions. Neutrophils are known to be a rich source of proteinase which carries many serine proteinases in their lysosomal granules. It was previously reported that leukocyte proteinase plays an important role in the development of tissue damage during inflammatory reactions and that a significant level of protection was provided by proteinase inhibitors [26,27]. In vitro anti-arthritic activity was assessed as antitryptic activity [27]. The IC_{50} results for the ATA range from 263 to 631.03 $\mu\text{g}/\text{mL}$.

The analysis of variance using Duncan test (with a significance level $p < 0.05$) revealed that there were no significant statistical differences in the ATA values between ibuprofen and **H2** and **H3**. The results present that the ibuprofen derivatives (**H2** and **H3**) show better antitryptic activity compared to ketoprofen and **H1** (Table 1, Figure 8).

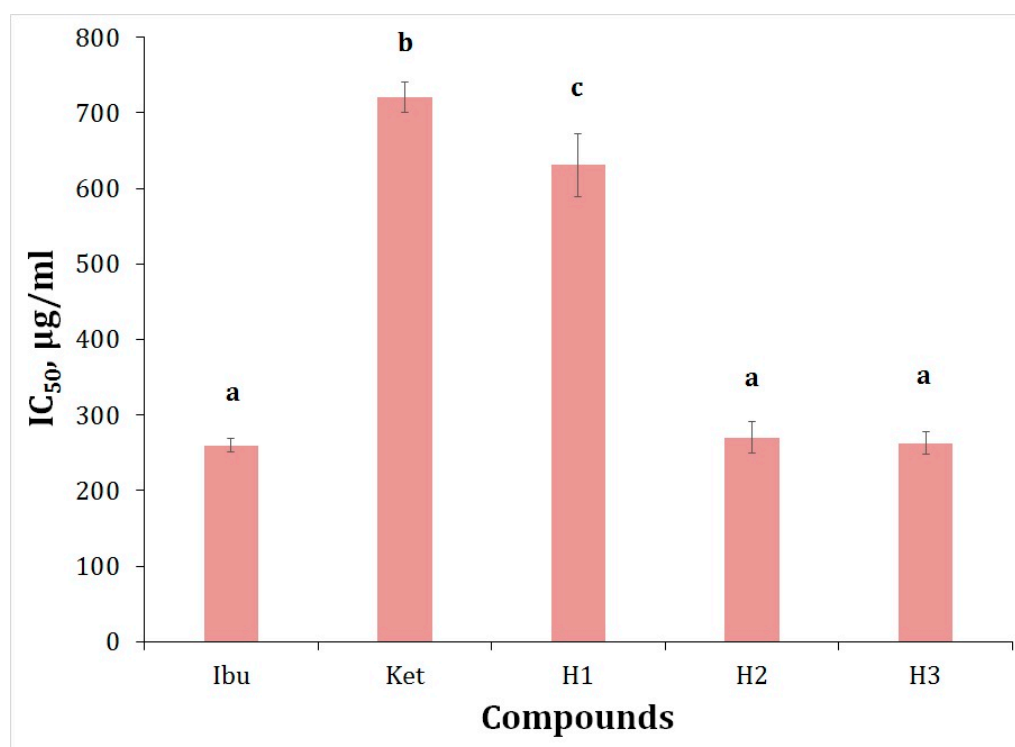


Figure 8. ATA of ibuprofen derivatives. Ibuprofen (Ibu) and ketoprofen (Ket) were used as standards. The results of ATA were expressed as IC_{50} . Different letters for the same method indicate significant difference at $p < 0.05$ levels by Duncan's test.

2.6. Lipophilicity

Lipophilicity is the most regularly applied parameter used in SAR drug discovery studies. It can be experimentally determined or calculated. Lipophilicity has been correlated with permeability, solubility, and increases in target potency and toxicity. We determined the lipophilicity by reverse phase thin layer chromatography (RPTLC) method as R_M values. This is considered to be a reliable, fast, and convenient method for expressing lipophilicity [28]. Aside from the essential role of lipophilicity for the kinetics of biologically

active compounds, antioxidants of hydrophilic or lipophilic character are both needed to act as radical scavengers in the aqueous phase or as chain-breaking antioxidants in biological membranes [29].

In the present work, we have investigated the antioxidant and in vitro biological activity of the newly synthesized ibuprofen derivatives. Lipophilicity proved to be an important factor in their activity. The results have shown that compounds (**H1-3**) are lipophilic with low antioxidant activity. However, it should not be taken as a criterion for whether the compounds will exhibit biological activity. However, the use of lipophilic antioxidants is necessary to neutralize harmful radicals in cell membranes [29].

In general, the in vitro studies results show that the ibuprofen hybrid molecules exhibit IAD and ATA. From both experiments, we derive important information about the properties of potential new drugs, and both experiments are related to preserving the integrity of the albumin molecule. HSA is known to have two major binding sites for drugs: Sudlow sites I and II. According to Sudlow's nomenclature, bulky heterocyclic molecules bind to Sudlow's site I (located in subdomain IIA), whereas Sudlow's site II (located in subdomain IIIA) is preferred by aromatic carboxylates with an extended conformation [30].

Ibuprofen has been shown to have good anti-inflammatory activity. However, it is also known that N-containing 6-membered alkaloids have biological effects such as anti-HIV and anti-tumor [31–33]. Therefore, we combined the two biologically active compounds in a new hybrid molecule. From all designed and carried experiments, we found out that the new hybrid molecules exhibit significant IAD and ATA activity.

3. Materials and Methods

3.1. Synthesis

All reagents and chemicals were purchased from commercial sources (Sigma-Aldrich S.A. and Riedel-de Haën, Sofia, Bulgaria) and used as received. Melting points were determined on a Boetius hot stage apparatus and are uncorrected. The NMR spectral data were recorded on a Bruker Avance II+600 spectrometer (BAS-IOCCP—Sofia, Bruker, Billerica, MA, USA). $^1\text{H-NMR}$ and $^{13}\text{C-NMR}$ spectra for compound **8** were taken in CDCl_3 at 600 MHz and 151 MHz, respectively. Chemical shifts are given in relative ppm and were referenced to tetramethylsilane (TMS) ($\delta = 0.00$ ppm) as an internal standard; the coupling constants are indicated in Hz. The NMR spectra were recorded at room temperature (ac. 295 K). Mass analyses were carried out on a Q Exactive Plus mass spectrometer (Thermo Fisher Scientific, Waltham, MA, USA). HPLC analysis consisted of quaternary mixer Smartline Manager 5000, pump Smartline 1000, and PDA 2800 detector (Knauer, Germany). Chromatographic conditions used: column Purosphere C18, 25 cm \times 4.6 mm i.d., 5 μm particle size (Merck, Germany); mobile phase flow rate was set by 1.0 mL/min; sample volume was 20 μL . TLC was carried out on precoated 0.2 mm Fluka silica gel 60 plates (Merck KGaA, Darmstadt, Germany).

3.2. Synthesis of 1-(3,4-Dihydroquinolin-1(2H)-yl)-2-(4-isobutylphenyl)propan-1-one **8**

To a solution of amine **2** (1 mmol, 0.133 g) in dichloromethane (15 mL), an equal amount of 2-(4-isobutylphenyl)propanoyl chloride (1 mmol, 0.224 g) **7** was added. After 10 min a little excess of triethylamine was added. After 30 min the solution was washed with diluted hydrochloric acid ($\text{HCl}:\text{H}_2\text{O} = 1:4$, v/v), saturated solution of Na_2CO_3 and water. The organic layer was dried (Na_2SO_4), concentrated and filtered on short column with neutral Al_2O_3 .

1-(3,4-dihydroquinolin-1(2H)-yl)-2-(4-isobutylphenyl)propan-1-one **8**: bright yellow oil, yield 96% (0.308 g), $^1\text{H-NMR}$ (600 MHz, CDCl_3) δ 7.20–6.77 (m, 8H), 4.13 (s, 1H), 3.78 (d, $J = 68.9$ Hz, 1H), 3.52 (d, $J = 5.1$ Hz, 1H), 2.42 (d, $J = 24.5$ Hz, 1H), 2.33 (d, $J = 7.2$ Hz, 2H), 1.82–1.61 (m, 3H), 1.39 (d, $J = 6.8$ Hz, 3H), 0.79 (dd, $J = 6.6, 1.7$ Hz, 6H). $^{13}\text{C-NMR}$ (151 MHz, CDCl_3) δ 140.02 (C=O), 138.90 (C, Ar), 129.34 (C, Ar), 129.22 (C, Ar), 128.33 (C, Ar), 128.31 (C, Ar), 127.05 (C, Ar), 126.03 (C, Ar), 125.99 (C, Ar), 125.95 (C, Ar), 125.14 (C, Ar), 44.99 (N-CH₂), 30.23 (-CH), 26.22 (-CH₂), 23.92 (-CH₂), 23.90 (-CH), 22.36 (-CH₃), 22.31 (-CH₂),

20.34 (–CH₃). UV λ_{\max} , MeOH: 250 ($\epsilon = 4.9 \times 10^5$) nm. HRMS Electrospray ionization (ESI) m/z calcd for C₂₂H₂₈NO⁺ = 322.2172, found 322.2175 (mass error $\Delta m = 0.93$ ppm).

Copies of all spectra, HPLC chromatogram, and ESI-HRMS (Figures S1–S5) are provided in the Supplementary Materials file.

3.3. Biological Evaluation

Chemicals and Reagents

Chromatographic grade methanol for HPLC analyses was used (VWR, Vienna, Austria). Water for HPLC was prepared with a Millipore purifier (Millipore, USA). Ibuprofen, ketoprofen, potassium dihydrogen phosphate, dipotassium hydrogen phosphate, sodium chloride, potassium chloride, hydrogen peroxide, ascorbic acid, trypsin, Tris-HCl buffer, and perchloric acid were purchased from Sigma-Aldrich, Taufkirchen, Germany. Human albumin 20%-BB, 200 g/L was purchased from BB-NCIPD Ltd., Sofia, Bulgaria. Chromatographic plates Kieselgel 60 F₂₅₄ were purchased from Merck (Darmstadt, Germany).

3.4. Biological Experiments

Hydrogen Peroxide Scavenging Activity (HPSA)

The ability of ibuprofen derivatives to scavenge hydrogen peroxide was assessed according to the method reported by Ruch [34] with minor modification as described by Manolov et al. [35]. The solution of hydrogen peroxide (43 mM) was prepared in potassium phosphate-buffered solution (0.2 M, pH 7.4). Sample analysis was performed as follows: in test tubes were mixed 0.6 mL hydrogen peroxide (43 mM), 1 mL sample/standard with different concentrations (20–1000 $\mu\text{g}/\text{mL}$) and 2.4 mL potassium phosphate-buffered solution. The mixture was stirred and incubated in dark for 10 min at 37 °C. Absorbance was measured at 230 nm with a spectrophotometer (Camspec M508, Leeds, UK) against a blank solution containing phosphate buffer and hydrogen peroxide without the sample. Ascorbic acid and quercetin were used as standards. The percentage HPSA of the samples was evaluated by comparing with a blank sample and calculated using the following formula:

$$\text{I\% (HPSA)} = \left[\frac{A_{\text{blank}} - (A_{\text{TS}} - A_{\text{CS}})}{A_{\text{blank}}} \right] \times 100$$

where A_{blank} is the absorbance of the blank sample (phosphate buffer and hydrogen peroxide), A_{CS} is the absorbance of the control sample (test sample + phosphate buffer) and A_{TS} is the absorbance of the test sample (test sample + phosphate buffer + hydrogen peroxide). The mean IC₅₀ value was estimated based on three replicates by means of interpolating the graphical dependence of scavenging hydrogen peroxide on concentration.

3.5. Inhibition of Albumin Denaturation (IAD)

In vitro analysis of anti-inflammatory activity was assessed as inhibition of albumin denaturation (IAD). The analysis was performed according Sakat method [36] with minor modification [35]. The experiment was performed with human albumin. The solution of albumin (1%) was prepared in distilled water (pH 7.4). The tested compounds/standard were dissolved firstly in 1.2 mL DMF and PBS up to 25 mL so the final concentration of the stock solution is 1000 $\mu\text{g}/\text{mL}$. Then, a series of working solutions with different concentrations (20–500 $\mu\text{g}/\text{mL}$) in PBS were prepared. The reaction mixture was containing 2 mL test sample/standard of different concentrations and 1 mL albumin (1%). The mixture was incubated at 37 °C for 15 min and then heated at 70 °C for 15 min in water bath. After cooling the turbidity was measured at 660 nm with a spectrophotometer (Camspec M508, Leeds, UK). The experiment was performed three times. Percentage inhibition of albumin denaturation (IAD) was calculated against control. The control sample is albumin with the same concentration dissolved in distilled water.

$$\text{\%IAD} = \left[\frac{A_{\text{control}} - A_{\text{sample}}}{A_{\text{control}}} \right] \times 100$$

3.6. Antitryptic Activity (ATA)

This method is known also as an *in vitro* anti-arthritis activity. The analysis was performed according to the method of Oyedapo and Femurewa [27] with minor modification as described by Manolov et al. [35]. The reaction mixture was containing 2 mL 0.06 mg/mL trypsin, 1 mL Tris-HCl buffer (20 mM, pH 7.4) and 1 mL test sample/standard (in methanol) of different concentrations (20–1000 µg/mL). The mixture was incubated at 37 °C for 5 min. Then, 1 mL of human albumin (4% *v/v*) was added. The mixture was incubated for an additional 20 min. To the mixture 2 mL of 70% perchloric acid was added for termination of the reaction. The cloudy suspension was cooled and centrifuged at 5000 rpm for 20 min. The absorbance of the supernatant was measured at 280 nm with a spectrophotometer (Camspec M508, Leeds, UK) against the control solution. The control solution was sample/standard in methanol with different concentrations. Ibuprofen was used as standard. The analysis was performed three times. The percentage of antitryptic activity (ATA) of the samples was evaluated by comparing with a blank sample. The blank sample is prepared as the test sample but with a small exception—perchloric acid is added before albumin.

$$\%ATA = \left[\frac{A_{\text{blank}} - (A_{\text{TS}} - A_{\text{CS}})}{A_{\text{blank}}} \right] \times 100$$

where A_{blank} is the absorbance of the blank sample, A_{CS} is the absorbance of the control solution (test sample in different concentrations) and A_{TS} is the absorbance of the test samples. The mean IC_{50} values were estimated by means of interpolating the graphical dependence of ATA on concentration.

3.7. Physicochemical Characterization

3.7.1. Determination of Lipophilicity as R_M Values

Determination of lipophilicity of ibuprofen derivatives was estimated according to the method reported by Pontiki and Hadjipavlou-Litina [29].

3.7.2. Prediction of Anti-Inflammatory and Anti-Arthritis Activity

A computerized prediction of biological activity (anti-inflammatory and anti-arthritis) for the obtained compounds was performed using the PASS Online program [37,38].

3.8. Statistical Analysis

All the analyses were made in triplicates. Data were expressed as mean \pm SD. The level of significance was set at $p < 0.05$. Statistical program SPSS 19.0 software was used for data analysis by one-way ANOVA followed by Duncan's post hoc test to evaluate differences between mean values of activities (SPSS Inc., Chicago, IL, USA).

4. Conclusions

In conclusion, we have successfully synthesized and characterized the new hybrid molecule 1-(3,4-dihydroquinolin-1(2H)-yl)-2-(4-isobutylphenyl)propan-1-one, which containing two pharmacologically active fragments—1,2,3,4-tetrahydroquinoline and ibuprofen. The newly obtained hybrid **H2** was biologically assessed for its *in vitro* antioxidant, antitryptic, and inhibition of albumin denaturation activity and compared to its structurally similar derivatives **H1** and **H3**. The lipophilicity was also determined experimentally using reversed-phase thin layer chromatography and *in silico* calculations. The analyzed hybrid molecules showed significant results, which makes them interesting for further biological assessments.

Supplementary Materials: The following supporting information can be downloaded online. Figure S1: $^1\text{H-NMR}$ spectrum of compound **8**, Figure S2: $^{13}\text{C-NMR}$ spectrum of compound **8**, Figure S3: UV spectrum of compound **8**, Figure S4: ESI-HRMS of compound **8**, Figure S5: HPLC chromatogram of compound **8**.

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