

Interesting Images

Feeding Behavior of *Coralliophila* sp. on Corals Affected by Caribbean Ciliate Infection (CCI): A New Possible Vector?

Simone Montano ^{1,2,*} , Greta Aeby ³ , Paolo Galli ^{1,2} and Bert W. Hoeksema ^{4,5} 

¹ Department of Earth and Environmental Sciences (DISAT), University of Milan–Bicocca, Piazza della Scienza, 20126 Milan, Italy; paolo.galli@unimib.it

² MaRHE Center (Marine Research and High Education Center), Magoodhoo Island, Faafu Atoll 12030, Maldives

³ Department of Biological and Environmental Sciences, Qatar University, Doha 2713, Qatar; greta@hawaii.edu

⁴ Taxonomy, Systematics and Geodiversity Group, Naturalis Biodiversity Center, 2300 RA Leiden, The Netherlands; bert.hoeksema@naturalis.nl

⁵ Groningen Institute for Evolutionary Life Sciences, University of Groningen, 9700 CC Groningen, The Netherlands

* Correspondence: simone.montano@unimib.it

Abstract: Coral reefs in the Caribbean are known to be affected by many coral diseases, yet the ecology and etiology of most diseases remain understudied. The Caribbean ciliate infection (CCI) caused by ciliates belonging to the genus *Halofolliculina* is a common disease on Caribbean reefs, with direct contact considered the most likely way through which the ciliates can be transmitted between infected and healthy colonies. Here we report an observation regarding a *Coralliophila* sp. snail feeding in proximity to a cluster of ciliates forming the typical disease band of CCI. The result of this observation is twofold. The feeding behavior of the snail may allow the passive attachment of ciliates on the body or shell of the snail resulting in indirect transport of the ciliates among colonies, which makes it eligible as a possible disease vector. Alternatively, the lesions created from snail feeding may enhance the progression of the ciliates already present on the coral as well as promoting additional infections allowing pathogens to enter through the feeding scar.

Keywords: coral disease; *Halofolliculina*; transmission mechanism; Bonaire; *Acropora*



Citation: Montano, S.; Aeby, G.; Galli, P.; Hoeksema, B.W. Feeding Behavior of *Coralliophila* sp. on Corals Affected by Caribbean Ciliate Infection (CCI): A New Possible Vector? *Diversity* **2022**, *14*, 363. <https://doi.org/10.3390/d14050363>

Academic Editors: Andrew Bauman and Harilaos Lessios

Received: 7 April 2022

Accepted: 2 May 2022

Published: 4 May 2022

Publisher's Note: MDPI stays neutral with regard to jurisdictional claims in published maps and institutional affiliations.



Copyright: © 2022 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (<https://creativecommons.org/licenses/by/4.0/>).

Coral diseases represent a serious threat for coral reefs worldwide, with the Caribbean considered a “hotspot” of disease outbreaks [1]. Currently, the coral reefs of the Caribbean are experiencing an outbreak of stony coral tissue loss disease (SCTLD) that originated on the reefs of Florida in 2014 [2] and is spreading across the Caribbean resulting in extensive colony mortality [3–6]. Despite the devastating effect this disease has had on coral reefs, our understanding of its ecology, pathogenesis, and etiologies is limited, making it difficult for resource managers to make decisions on how best to maintain these critical resources. As example, understanding disease spread among individual colonies or coral populations would be a critical factor in developing management actions to slow down or stop disease and the resultant mortality, yet disease transmission dynamics are still understudied [7].

Many studies have suggested that coral-feeding animals can promote disease transmission among colonies on a reef. Several families of coral reef fishes have been observed in the field feeding on coral disease lesions including black band disease (BBD) [8,9], brown band disease (BrB) [10] and stony coral tissue loss disease [11]. Corallivores feeding on disease lesions could transmit the pathogen by subsequently feeding or defaecating on a non-diseased colony and this has been suggested as a mechanism of black band disease transfer by butterflyfishes [9,12] as well as spreading the trematode parasite that causes *Porites trematodiasis* in Hawaii [13]. Numerous types of invertebrate corallivores (snails, nudibranchs, fireworm, crown-of-thorns seastars) have also been implicated in disease

transmission, either directly [1,14,15] or indirectly via feeding scars which subsequently develop disease [16–21].

Halofolliculinid ciliates can cause progressive tissue loss on corals; this disease is termed skeletal eroding band in the Indo-Pacific and Caribbean ciliate infection in the Caribbean [22]. Caribbean ciliate infection (CCI) was first reported in 2006 [23], and it can affect ~4 to 8 % of corals as observed in Venezuela and Curaçao [22]. It manifests as a dark-grey band 1–10 cm thick, located at the interface between recently exposed skeleton and apparently healthy coral tissue showing the characteristic spotted appearance of the clustering ciliates [24,25] (Figure 1a,b). *Halofolliculina* ciliates have a life cycle represented by two distinct phases: a sessile ciliate (encased within a lorica), and a motile larval phase. During replication, the de-differentiation of the sessile feeding trophont results into a simple motile phase, which then divides asexually into two motile swimmers that may move using ciliary locomotion and disperse [26]. Transmission of ciliate infection among coral colonies occurs on direct contact between a healthy and infected colony [26] and through the water column if the healthy colony has a prior injury of any origin [27,28]. Ciliates at the sessile stage have also been found embedded in the shells of a number of corallivorous gastropods which may serve as passive vectors of the disease [22].

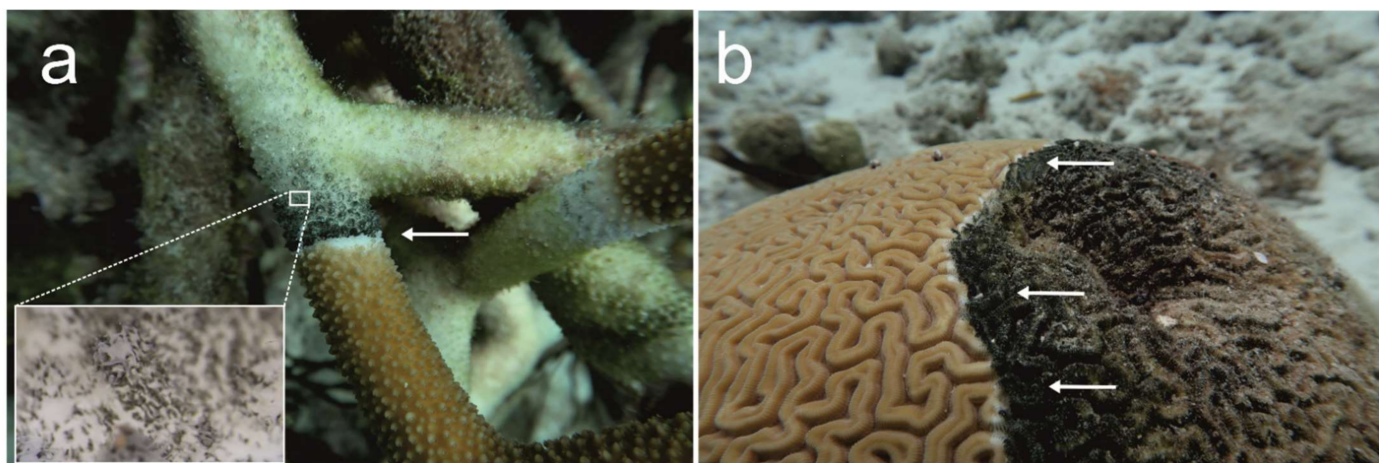


Figure 1. It shows the typical appearance of corals affected by CCI. The arrows indicate the cluster of ciliates forming CCI dark-grey bands located at the interface between recently exposed skeleton and apparently healthy coral tissue on (a) *Acropora cervicornis* and (b) *Diploria labyrinthiformis*.

Here we report on an observation made in Bonaire in 2019, in which a *Coralliophila* sp. snail was observed feeding on coral tissue at the edge of the cluster of ciliates forming typical disease bands of CCI (Figure 2a,b). The snail species in question is probably *C. galea* (Dillwyn, 1823), previously misidentified as *C. abbreviata* (Lamarck, 1816), which is so far the only *Coralliophila* species reported from Caribbean *Acropora* spp. [29–31]. This observation creates the possibility that transmission of CCI may also be facilitated by snail activities. Lesions created by snail predation may open-up wounds in the coral which can then be colonized by the ciliates at the swarmer stage. Alternatively, snails are attracted to injured coral tissue [32] so coral lesions created by CCI could attract snails and allow passive attachment of ciliates on the body, or shell, of the snail resulting in indirect transport of the ciliates among colonies. *Coralliophila* species have been implicated as a potential vector of white band disease [15,33], white pox disease [1], and white plague disease [34] in the Caribbean, as well as to disease development in *Porites cylindrica* in the Indo-Pacific [20]. Our observation adds to the growing body of evidence on the role that snails play in disease transmission, however, the extent to which *Coralliophila* may be involved in the pathogenesis of halofolliculinid ciliate infection in Bonaire needs further investigation.

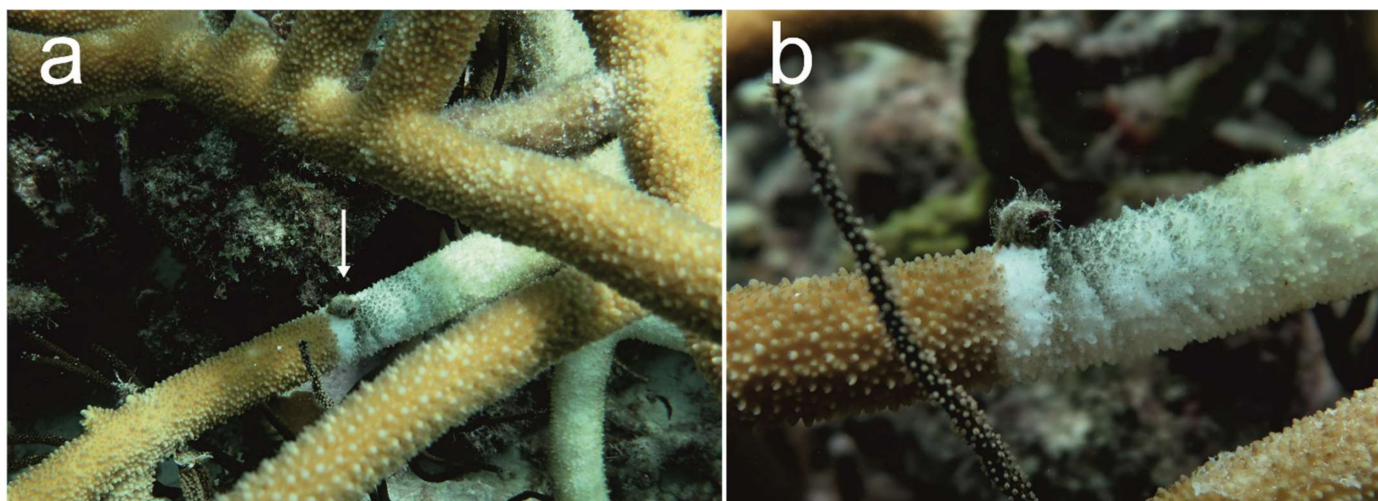


Figure 2. (a) *Coralliophila* sp. snail feeding in proximity of cluster of ciliates forming the typical disease band of CCI; (b) close-up of the snail's feeding behavior.

Author Contributions: Investigation, S.M., B.W.H.; data curation, G.A.; writing—original draft preparation, S.M., G.A., B.W.H.; supervision, P.G. All authors have read and agreed to the published version of the manuscript.

Funding: Fieldwork at Bonaire was supported by the World Wildlife Fund (WWF) Netherlands Biodiversity Fund, the Treub Maatschappij—Society for the Advancement of Research in the Tropics, and by the Nature of the Netherlands program of Naturalis Biodiversity Center.

Acknowledgments: S.M. is grateful to the Naturalis Biodiversity Center for providing Martin Fellowships, which supported fieldwork in Bonaire (2019). We are grateful to the Stichting Nationale Parken (STINAPA) and Dutch Caribbean Nature Alliance (DCNA) at Bonaire for assistance in the submission of the research proposal and the research permit. A special thanks to the Magnificent 7 team for its unforgettable support.

Conflicts of Interest: The authors declare no conflict of interest.

References

1. Sutherland, K.P.; Shaban, S.; Joyner, J.L.; Porter, J.W.; Lipp, E.K. Human pathogen shown to cause disease in the threatened elkhorn coral *Acropora palmata*. *PLoS ONE* **2011**, *6*, e23468. [[CrossRef](#)] [[PubMed](#)]
2. Precht, W.F.; Gintert, B.E.; Robbart, M.L.; Fura, R.; Van Woesik, R. Unprecedented disease-related coral mortality in Southeastern Florida. *Sci. Rep.* **2016**, *6*, 31374. [[CrossRef](#)] [[PubMed](#)]
3. Alvarez-Filip, L.; Estrada-Saldivar, N.; Pérez-Cervantes, E.; Molina-Hernandez, A.; Gonzalez-Barrios, F.J. A rapid spread of the stony coral tissue loss disease outbreak in the Mexican Caribbean. *PeerJ* **2019**, *7*, e8069. [[CrossRef](#)] [[PubMed](#)]
4. Meiling, S.; Muller, E.M.; Smith, T.B.; Brandt, M.E. 3D photogrammetry reveals dynamics of stony coral tissue loss disease (SCTLD) lesion progression across a thermal stress event. *Front. Mar. Sci.* **2020**, *7*, 1128. [[CrossRef](#)]
5. Heres, M.M.; Farmer, B.H.; Elmer, F.; Hertler, H. Ecological consequences of stony coral tissue loss disease in the Turks and Caicos Islands. *Coral Reefs* **2021**, *40*, 609–624. [[CrossRef](#)]
6. Thome, P.E.; Rivera-Ortega, J.; Rodríguez-Villalobos, J.C.; Cerqueda-García, D.; Guzmán-Urieta, E.O.; García-Maldonado, J.Q.; Carabantes, N.; Jordán-Dahlgren, E. Local dynamics of a white syndrome outbreak and changes in the microbial community associated with colonies of the scleractinian brain coral *Pseudodiploria strigosa*. *PeerJ* **2021**, *9*, e10695. [[CrossRef](#)]
7. Shore, A.; Caldwell, J.M. Modes of coral disease transmission: How do diseases spread between individuals and among populations? *Mar. Biol.* **2019**, *166*, 1–14. [[CrossRef](#)]
8. Cole, A.J.; Chong-Seng, K.M.; Pratchett, M.S.; Jones, G.P. Coral-feeding fishes slow progression of black-band disease. *Coral Reefs* **2009**, *28*, 965. [[CrossRef](#)]
9. Chong-Seng, K.M.; Cole, A.J.; Pratchett, M.S.; Willis, B.L. Selective feeding by coral reef fishes on coral lesions associated with brown band and black band disease. *Coral Reefs* **2011**, *30*, 473–481. [[CrossRef](#)]
10. Nicolet, K.J.; Hoogenboom, M.O. The corallivorous invertebrate *Drupella* aids in transmission of brown band disease on the Great Barrier Reef. *Coral Reefs* **2013**, *32*, 585–595. [[CrossRef](#)]
11. Noonan, K.R.; Childress, M.J. Association of butterflyfishes and stony coral tissue loss disease in the Florida Keys. *Coral Reefs* **2020**, *39*, 1581–1590. [[CrossRef](#)]

12. Aeby, G.S.; Santavy, D.L. Factors affecting susceptibility of the coral *Montastraea faveolata* to black-band disease. *Mar. Ecol. Prog. Ser.* **2006**, *318*, 103–110. [[CrossRef](#)]
13. Aeby, G.S. Interactions of the Digenetic Trematode, *Podocotyloides stenometra*, with Its Coral Intermediate Host and Butterflyfish Definitive Host: Ecology and Evolutionary Implications. Ph.D. Thesis, University of Hawaii at Manoa, Honolulu, HI, USA, 1998.
14. Sussman, M.; Loya, Y.; Fine, M.; Rosenberg, E. The marine fireworm *Hermodice carunculata* is a winter reservoir and spring-summer vector for the coral-bleaching pathogen *Vibrio shiloi*. *Environ. Microbiol.* **2003**, *5*, 250–255. [[CrossRef](#)] [[PubMed](#)]
15. Williams, D.E.; Miller, M.W. Coral disease outbreak: Pattern, prevalence and transmission in *Acropora cervicornis*. *Mar. Ecol. Prog. Ser.* **2005**, *301*, 119–128. [[CrossRef](#)]
16. Dalton, S.J.; Godwin, S. Progressive coral tissue mortality following predation by a corallivorous nudibranch (*Phestilla* sp.). *Coral Reefs* **2006**, *25*, 529. [[CrossRef](#)]
17. Nugues, M.M.; Bak, R.P.M. Brown-band syndrome on feeding scars of the crown-of-thorn starfish *Acanthaster planci*. *Coral Reefs* **2009**, *28*, 507–510. [[CrossRef](#)]
18. Rypien, K.L.; Baker, D.M. Isotopic labeling and antifungal resistance as tracers of gut passage of the sea fan pathogen *Aspergillus sydowii*. *Dis. Aquat. Organ.* **2009**, *86*, 1–7. [[CrossRef](#)]
19. Katz, S.M.; Pollock, F.J.; Bourne, D.G.; Willis, B.L. Crown-of-thorns starfish predation and physical injuries promote brown band disease on corals. *Coral Reefs* **2014**, *33*, 705–716. [[CrossRef](#)]
20. Raymundo, L.J.; Work, T.M.; Miller, R.L.; Lozada-Misa, P.L. Effects of *Coralliophila violacea* on tissue loss in the scleractinian corals *Porites* spp. depend on host response. *Dis. Aquat. Organ.* **2016**, *119*, 75–83. [[CrossRef](#)]
21. Nicolet, K.J.; Chong-Seng, K.M.; Pratchett, M.S.; Willis, B.L.; Hoogenboom, M.O. Predation scars may influence host susceptibility to pathogens: Evaluating the role of corallivores as vectors of coral disease. *Sci. Rep.* **2018**, *8*, 5258. [[CrossRef](#)]
22. Page, C.A.; Cróquer, A.; Bastidas, C.; Rodríguez, S.; Neale, S.J.; Weil, E.; Willis, B.L. Halofolliculina ciliate infections on corals (skeletal eroding disease). In *Diseases of Coral*; Woodley, C.M., Downs, C.A., Bruckner, A.W., Porter, J.W., Galloway, S.B., Eds.; Wiley-Blackwell: Hoboken, NJ, USA, 2016; pp. 361–375.
23. Cróquer, A.; Bastidas, C.; Lipscomb, D. Folliculinid ciliates: A new threat to Caribbean corals? *Dis. Aquat. Org.* **2006**, *69*, 75–78. [[CrossRef](#)] [[PubMed](#)]
24. Cróquer, A.; Bastidas, C.; Lipscomb, D.; Rodríguez-Martínez, R.E.; JordanDahlgren, E.; Guzman, H.M. First report of folliculinid ciliates affecting Caribbean scleractinian corals. *Coral Reefs* **2006**, *25*, 187–191. [[CrossRef](#)]
25. Montano, S.; Maggioni, D.; Liguori, G.; Arrigoni, R.; Berumen, M.L.; Seveso, D.; Galli, P.; Hoeksema, B.W. Morpho-molecular traits of Indo-Pacific and Caribbean *Halofolliculina* ciliate infections. *Coral Reefs* **2020**, *39*, 375–386. [[CrossRef](#)]
26. Antonius, A.A.; Lipscomb, D. First protozoan coral-killer identified in the Indo-Pacific. *Atoll Res. Bull.* **2001**, *481*, 1–21. [[CrossRef](#)]
27. Page, C.A.; Willis, B.L. Epidemiology of skeletal eroding band on the Great Barrier Reef and the role of injury in the initiation of this widespread coral disease. *Coral Reefs* **2008**, *27*, 257–272. [[CrossRef](#)]
28. Rodríguez, S.; Croquer, A.; Guzmán, H.M.; Bastidas, C. A mechanism of transmission and factors affecting coral susceptibility to *Halofolliculina* sp. infection. *Coral Reefs* **2009**, *28*, 67–77. [[CrossRef](#)]
29. Knowlton, N.; Lang, J.C.; Keller, B.D. Fates of staghorn coral isolates on hurricane-damaged reefs in Jamaica: The role of predators. In Proceedings of the 6th International Coral Reef Symposium, Townsville, Australia, 8–12 August 1988; Volume 2, pp. 83–88.
30. Hayes, J.A. Distribution, movement and impact of the corallivorous gastropod *Coralliophila abbreviata* (Lamarck) on a Panamian patch reef. *J. Exp. Mar. Biol. Ecol.* **1990**, *142*, 25–42. [[CrossRef](#)]
31. Potkamp, G.; Vermeij, M.J.A.; Hoeksema, B.W. Genetic and morphological variation in corallivorous snails (*Coralliophila* spp.) living on different host corals at Curaçao, southern Caribbean. *Contrib. Zool.* **2017**, *86*, 111–144. [[CrossRef](#)]
32. Morton, B.; Blackmore, G.; Kwok, C.T. Corallivory and prey choice by *Drupella rugosa* (Gastropoda: Muricidae) in Hong Kong. *J. Molluscan Stud.* **2002**, *68*, 217–223. [[CrossRef](#)]
33. Gignoux-Wolfsohn, S.A.; Marks, C.J.; Vollmer, S.V. White band disease transmission in the threatened coral *Acropora cervicornis*. *Sci. Rep.* **2012**, *2*, 804. [[CrossRef](#)]
34. Clemens, E.; Brandt, M.E. Multiple mechanisms of transmission of the Caribbean coral disease white plague. *Coral Reefs* **2015**, *34*, 1179–1188. [[CrossRef](#)]