



*Communication*

# **Benthic Biodiversity by Baited Camera Observations on the Cosmonaut Sea Shelf of East Antarctica**

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**Abstract:** A free-fall baited camera lander was launched for the first time on the Cosmonaut Sea shelf of East Antarctica at a depth of 694 m during the 38th Chinese National Antarctic Research Expedition (CHINARE) in 2022. We identified 31 unique taxa (23 were invertebrates and eight were fish) belonging to eight phyla from 2403 pictures and 40 videos. The Antarctic jonasfish (*Notolepis coatsi*) was the most frequently observed fish taxa. Ten species of vulnerable marine ecosystem (VME) taxa were observed, accounting for 32% of all species. The maximum number (MaxN) of *Natatolana meridionalis* individuals per image frame was ten, and they were attracted to the bait. The macrobenthic community type were sessile suspension feeders with associated fauna (SSFA), which was shaped by the muddy substrata with scattered rocks. Rocks served as the best habitats for sessile fauna. The study reveals the megafauna community and their habitat by image survey in the Cosmonaut Sea for the first time. It helped us obtain Antarctic biodiversity baselines and monitoring data for future ecosystem health assessment and better protection.

**Keywords:** lander; Cosmonaut Sea; megafauna; Antarctica; image survey

# **1. Introduction**

The Southern Ocean is unique among the world's oceans in terms of its linkage with the other major ocean basins, its rich and unusual marine ecosystem, and its interaction with the physical climate system and the biogeochemistry of the region [\[1\]](#page-8-0). It comprises 15% of the world's oceans and is home to thousands of endemic species [\[2–](#page-8-1)[4\]](#page-8-2). The Southern Ocean has become a hot area for global research on climate change and ecological evolution due to its harsh natural environment, fragile ecosystem, and sensitivity to environmental variations. There is a growing need for marine biodiversity baselines and monitoring data to assess ocean ecosystem health, especially around Antarctica, where data are rare [\[5–](#page-8-3)[8\]](#page-8-4).

The Cosmonaut Sea (30◦~60◦ E) is located to the west of Enderby Land in east Antarctica and has been poorly explored [\[9\]](#page-8-5). Therefore, very few biological data have been recorded for the region [\[10](#page-8-6)[–13\]](#page-8-7), and existing ones mainly include the composition and distribution of phytoplankton, mesozooplankton, euphausiid larvae, krill, squid, and Antarctic jonasfish [\[14](#page-8-8)[–16\]](#page-8-9). However, there are still no data about macrobenthos in the Cosmonaut Sea. Antarctic benthic ecosystems in the Cosmonaut Sea may be sentinels for monitoring the effects of climate change [\[17](#page-8-10)[,18\]](#page-8-11). Macrobenthic communities in Antarctica



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differ in biodiversity and ecosystem functioning and are shaped by a variety of physical and biological drivers [\[19\]](#page-8-12). Some of these communities are unique in their occurrence and and biological drivers [19]. Some of these communities are unique in their occurrence and never occupied with the [18]. Bothe of under commutances are analyze in their occurrence and proportions of species and life forms. Some are typical for the entire Antarctic shelf, but never occur with exactly the same proportions or compositions  $[18]$ .

Trawls, sledges, and dredges were historically common facilities for sampling the shelf benthic marine communities of Antarctica [\[12,](#page-8-13)[20,](#page-8-14)[21\]](#page-8-15). These traditional methods can help in understanding the structure and function of the Antarctic benthic systems and are good for species identification of slow-moving macrobenthos. However, they cannot describe species behaviors and interactions and have difficulty in capturing more mobile species [\[4\]](#page-8-2). Video surveys are the emerging methods to estimate the relative abundances of scavenging fishes and invertebrates [\[8,](#page-8-4)[22,](#page-9-0)[23\]](#page-9-1). Such studies can help us develop a better understanding of ecosystem patterns and processes in Antarctica, such as the Antarctic Peninsula, Prydz Bay, and the Amundsen Sea, and the applicability has also been justified [\[4](#page-8-2)[,24](#page-9-2)[–27\]](#page-9-3). [4,24–27].

In this study, a free-fall baited camera lander was deployed for the first time on the In this study, a free-fall baited camera lander was deployed for the first time on the Cosmonaut Sea shelf during the 38th Chinese National Antarctic Research Expedition Cosmonaut Sea shelf during the 38th Chinese National Antarctic Research Expedition (CHINARE) on 22 February 2022. The objective was to preliminarily survey fish and invertebrate communities and basically understand the structure of the benthic community and the interactions among species on the Cosmonaut Sea shelf. and the interactions among species on the Cosmonaut Sea shelf.

#### **2. Method and Equipment 2. Method and Equipment**

# *2.1. Location 2.1. Location*

The lander was deployed on the shelf (48◦50.980′ E, 66◦22.226′ S) near the slope of the The lander was deployed on the shelf (48°50.980′ E, 66°22.226′ S) near the slope of the Cosmonaut Sea (Figure [1\)](#page-1-0). The Cosmonaut Sea is the site of an important confluence in Cosmonaut Sea (Figure 1). The Cosmonaut Sea is the site of an important confluence in polar circulation, but poorly investigated, meriting more research efforts [\[9\]](#page-8-5). polar circulation, but poorly investigated, meriting more research efforts [9].

<span id="page-1-0"></span>

**Figure 1.** Sampling station of the lander. **Figure 1.** Sampling station of the lander.

# *2.2. Equipment 2.2. Equipment*

The lander system framework is made of titanium, consisting of batteries, an acoustic The lander system framework is made of titanium, consisting of batteries, an acoustic releaser, cameras, and traps (Figure 2). The batteries are rechargeable lithium batteries releaser, cameras, and traps (Figure [2\)](#page-2-0). The batteries are rechargeable lithium batteries combined with capacity greater than 96 Ah. The releaser is manufactured by iXblue. The combined with capacity greater than 96 Ah. The releaser is manufactured by iXblue. The load and communication distance of the iXblue releaser are not less than 2500 kg and 10 load and communication distance of the iXblue releaser are not less than 2500 kg and 10 km, respectively. iXblue is powered by a No. 1 alkaline battery or lithium battery. The Sea-Bird Scientific SBE 37 is equipped with high-accuracy temperature, conductivity, and pressure sensors with an RS-232 interface, internal batteries, data storage, and pump. The sampling interval was set to 120 s, and the maximum observation depth was 7000 m. Two cameras

were configured: one (CO01-016E,  $3648 \times 2736$ ) was used to take photos, and the other (CO02-016HE, HD1080P) was used to record videos. The lander used a high-efficiency LED lamp (CO04-50LI-6000AA), and the light was angled downward at a constant  $45^{\circ}$  from the horizontal plane. Two traps were used in the study: a large fish and invertebrate trap (82 cm  $\times$  47 cm  $\times$  47 cm) and an invertebrate trap ( $\varphi$ 10 cm  $\times$  50 cm).

<span id="page-2-0"></span>

**Figure 2.** Structure of the free-fall baited camera lander. **Figure 2.** Structure of the free-fall baited camera lander.

# *2.3. Deployment Process 2.3. Deployment Process*

The lander was deployed and recovered during the 38th CHINARE on the  $\rm R/V$ Xuelong2 icebreaker to a depth of 694 m. The equipment was deployed at 20:09 and landed on the sea floor at 20:26 on 22 February 2022 (UTC). on the sea floor at 20:26 on 22 February 2022 (UTC).

The lander deployment process had four stages: setup, deployment, recovery, and data data exporting. Prior to deployment, the cameras were activated through underwater sys-by a computer. The camera was preprogrammed to take 1 picture every 30 s, and the video tems by a computer. The camera was preprogrammed to take 1 picture every 30 s, and the camera was preprogrammed to take 10 min of video every 30 min. The survey time was set exporting. Prior to deployment, the cameras were activated through underwater systems according to the battery charging time. Then, baits (chicken legs and *Silver sillago*) were loaded into the traps. The last step was connecting the acoustic releaser to the cement block. Deployment was completed, and the lander was dropped into the sea. The dropping rate was approximately 1 m/s. Location and bottom depths were based on triangulation of the acoustic releases after the deployment [\[28\]](#page-9-4). The lander was recovered in the daytime when the sea was calm. The ship traveled to the deployment area for lander recovery. To recover the lander, the deck unit was used to search for the signal from the acoustic releaser, and the distance between the equipment and the ship was measured. Then, the deck unit released the signal to recover the lander. The rising rate was approximately 1 m/s. The crew searched for the lander, which was attached to orange balls from the bridge. Finally, the equipment was found and brought up onto the deck.

The lander was washed using fresh water after being placed on the deck. Then, the samples were collected from the trap cage. Following retrieval, an external hard disk was used to store the images and videos from the cameras and the data from SBE 37 for further analysis.

#### *2.4. Metrics and Biodiversity Analyses*

Annotations were made from the video footage to identify species to the highest taxonomic resolution [\[8\]](#page-8-4). A relative abundance metric, the maximum number (MaxN), was used in the video survey. Counts of MaxN for individuals of each species in a video frame were performed rather than the total tally per deployment to avoid double-counting [\[29\]](#page-9-5). Other variables were also recorded, such as the time for a taxon's first arrival and the time when the maximum number of a taxon was observed upon the landing of the device on the seafloor [\[8,](#page-8-4)[23](#page-9-1)[,30\]](#page-9-6).

### **3. Results**

A total of 2403 photos and 40 videos were recorded underwater by the lander during approximately 20 h.

## *3.1. Substrate Type*

The images were used to determine substrate characteristics. The area was dominated by muddy substrate containing some rocks. The rocks were scattered on the sea floor and served as substrata for different taxa, such as sponges, bryozoans, and corals. These taxa, in turn, served as living substrata for ophiuroids, asteroids, holothurians, and others (Figure [3\)](#page-3-0).

<span id="page-3-0"></span>

**Figure 3.** Substrate type of the Cosmonaut Sea shelf. **Figure 3.** Substrate type of the Cosmonaut Sea shelf.

## *3.2. Observation of the Hydrological Environment 3.2. Observation of the Hydrological Environment*

The SBE 37 started to record data on salinity, temperature, and pressure at the bottom The SBE 37 started to record data on salinity, temperature, and pressure at the bottom at 21:00. The linear trend of pressure is 0.0528 dbar per hour, which is significant at a 95% confidence level. The highest pressure was 701.545 dbar at 16:06, and the lowest one 700.542 at 23:42. The average value of salinity and temperature were 34.6096 psu and was 700.542 at 23:42. The average value of salinity and temperature were 34.6096 psu and 0.1091 °C, respectively. The highest salinity was 34.6446 psu at 5:42, and the highest value 0.1091 ◦C, respectively. The highest salinity was 34.6446 psu at 5:42, and the highest value of temperature was  $0.1584 °C$  at 8:20. Salinity and temperature varied strongly (Figure [4\)](#page-4-0), with a sharp fall at 13:00. The salinity reached its lowest value (34.5511 psu) at 13:42. The with a sharp fall at 13:00. The salinity reached its lowest value (34.5511 psu) at 13:42. The temperature reached its lowest value (−0.0478 °C) at 13:32. The lower temperature and temperature reached its lowest value (−0.0478 ◦C) at 13:32. The lower temperature and salinity lasted approximately one hour, and they quickly increased at 14:20. These data salinity lasted approximately one hour, and they quickly increased at 14:20. These data may imply that there was a low-temperature and low-salinity water mass passing over may imply that there was a low-temperature and low-salinity water mass passing over the bottom. the bottom.

<span id="page-4-0"></span>

**Figure 4.** Data collected by the SBE 37 recorder. **Figure 4.** Data collected by the SBE 37 recorder.

## *3.3. Recorded Taxa*

*3.3. Recorded Taxa*  Thirty-one species were identified, representing eight phyla, twenty orders, and thirty families (Table [1\)](#page-5-0). Invertebrates accounted for 23 species, and fishes accounted for 8 species  $t_{\text{Fion}}$   $\frac{1}{2}$ . Bryozoa dominated in the images (Figure [5\)](#page-4-1). Bryozoa dominated in the images.

<span id="page-4-1"></span>

**Figure 5.** Fish images from the shelf of Cosmonaut Sea.

Three fish were the most frequently occurring fish taxa. *Notolepis coatsi* appeared in 37 videos with up to four individuals in one frame and in 29 of 2403 photos with up to three individuals in one photo. *Trematomus lepidorhinus* was found in 16 of 40 videos and 809 of 2403 photos. *Melanostigma gelatinosum* appeared in 117 of 2403 photos. *Dissostichus mawsoni* was recorded only once by the cameras at 3:35, 23 February 2022.

Ten species of VME (vulnerable marine ecosystem) taxa were observed, accounting for 32% of all species. VME taxa include sponges, sea mats, and cold-water corals, which can provide an important habitat for diversity of marine organisms [\[31\]](#page-9-7). Sea pens, sea anemones, and sea squirts were the VME taxa commonly observed (Table [1\)](#page-5-0).



<span id="page-5-0"></span>**Table 1.** Maximum number (MaxN) of individuals of each species.

*Nothoadmete* sp. was first observed at 21:09, and the MaxN per image frame was four. *Natatolana meridionalis* was observed at 21:16, and the MaxN per image frame was ten. These two species were the main ones influencing the MaxN of individuals per image frame each hour.

The species caught in the traps were Isopoda (*N. meridionalis*), Amphipoda (*Abyssorchomene* sp.), and fish (*N. coatsi*), with nine, thirteen, and two individuals (Figure [6\)](#page-6-0), respectively.

<span id="page-6-0"></span>

**Figure 6.** The pictures of *N. coatsi* caught. **Figure 6.** The pictures of *N. coatsi* caught.

# **4. Discussion 4. Discussion**

# *4.1. Substrate Type and Assemblages 4.1. Substrate Type and Assemblages*

Seabed imagery was used to determine substrate characteristics. The distribution of benthic biota on the Antarctic shelf have frequently been associated with substrate type and water depth [\[32,](#page-9-8)[33\]](#page-9-9). Muddy substrates with scattered rocks were found in this type and water depth [32,33]. Muddy substrates with scattered rocks were found in this study. Many rocks may be dropstones from iceberg scouring [\[33\]](#page-9-9). Rocks are important for buildbuilding benthic communities and they provide a hard attachment site and an elevated position is the position of the position off the bottom, enhancing the food supply for sessile suspension feeders, such as sponges, sponges, bryozoans, pennatulaceun, and actiniae [\[32\]](#page-9-8). Deposit feeders are often associated bryozoans, pennatulaceun, and actiniae [32]. Deposit feeders are often associated with soft with soft substrates, where they are able to feed on fine particles [\[34\]](#page-9-10). Amphipods were which substrates, where they are able to feed on fine particles [34]. Amphipods were associated with muddy substrates from the images recorded, and the phytodetritus in woten the muddy substrates from the images recorded, and the phytodetritus in  $m$ muddy substrates may be the food source for them. position off the bottom, enhancing the food supply for sessile suspension feeders, such as

#### *4.2. Hydrological Environment and Assemblages*

*Meijers et al.* [\[9\]](#page-8-5) found seasonal Antarctic Bottom Water (AABW) produced in the region ( $60°$  E), which moved down the slope and was deflected westward due to the Coriolis force. The mass water mixed with Antarctic Circumpolar Current (ACC) and Weddell Gyre waters above it as it moved westward across the regions 30° E, 40° E, and  $50°$  E, eroding the strong characteristics observed in the region (60 $°$  E). Lowered acoustic Doppler current profiler (LADCP) data showed that the bottom water had large velocities near the deployment area [9]. The different postures of *U. carpenteri* showed high-strength and disordered current at the bottom. The current at the bottom brought a large number of organic debris, which provided the food source for sessile suspension feeders, such as sponges, colder-water corals, and sea pens. This was important for structuring the benthic community. This was important for sea pens. This was important for structuring the benthic community.

#### *4.3. Comments on Fishes and Invertebrates in Images*

*Notolepis coatsi* was the most frequently encountered fish taxon and the second-largest fish biomass in the Cosmonaut Sea of this study, like that in Prydz Bay [\[15](#page-8-16)[,16\]](#page-8-9). It is a midsize bathypelagic fish widely distributed around the Antarctic continental shelf [\[16\]](#page-8-9). *N. coatsi* may feed only on krill [\[35\]](#page-9-11) and account for 50% of the food consumed by Antarctic fur seals (*Arctocephalus gazella*) [\[36\]](#page-9-12). Therefore, *N. coatsi* plays an important role in the marine food web, which directly links krill and marine mammals in the Southern Ocean. N. *coatsi* is regarded as an ideal organism to evaluate the role of the Cosmonaut Sea because of its large population size, position in the food web, and specialized diet [\[16\]](#page-8-9). Additionally, we not only explored *N. coatsi* behavior but also trapped the individuals using the lander.

*Dissostichus mawsoni* was observed by photographic survey for the first time in the Cosmonaut Sea and is a large nototheniid species endemic to the Antarctic continental shelf. The Antarctic toothfish is an important fishery resource in the Southern Ocean and plays an important role in Antarctic ecosystems [\[11\]](#page-8-17). Sufficient data and stock information are needed to establish fisheries, but such information is rare in the Cosmonaut Sea. In the future, the lander can help us assess the relative abundance and population size structure of the Antarctic toothfish in the Cosmonaut Sea.

In total, 10 VME taxonomic categories were observed from the lander imagery, accounting for 32% of all species. The VME taxa were distributed sparsely, and most of them lived on rocks. The presence of benthic invertebrates contributes to the creation of complex three-dimensional structures and provides substrata for other organisms called sessile suspension feeders with associated fauna (SSFA) [\[32](#page-9-8)[,33\]](#page-9-9). The main sessile suspension feeders recorded in this survey were Bryozoa. Associated fauna included Ophiuroidea, Asteroidea, Gastropoda, Isopoda, and others. Isopoda and Amphipoda were the main taxa in the trap, so the bait is possibly attractive to them. *Natatolana meridionalis* first arrived at the bait fifty-one minutes after the lander arrived at the seafloor. The bait was consumed for approximately eight hours, until it was fully exhausted. Data from the lander can help assess changes in benthic community diversity and their associated habitat structure.

#### *4.4. Observation Advantages and Limitations of the Lander*

A major advantage of the lander is that it can explore fauna behavior, including the interactions between species. It is a powerful tool for the detection of rare predatory fish species [\[37\]](#page-9-13), which are difficult to find through physical sampling. The structure of the community should be visually determined to better understand vulnerable marine ecosystems. The lander, thus, allows us to clarify anthropogenic impacts, such as fishing and climate change. Also, the density of data for macrobenthos is low, especially in the Cosmonaut Sea. Although traditional trawling is advisable for the collection of all faunal types for higher resolution taxonomic investigations, the lander adds an existing monitoring program to extend biological observing capacity into the Antarctic Ocean [\[8\]](#page-8-4). Moreover, the lander is an unnamed vehicle that falls to the seafloor unattached to any cable, subsequently operating autonomously at the bottom, and it can carry diverse instruments for environmental parameter collections [\[38\]](#page-9-14).

However, the lander cannot acquire images of small sessile fauna (meiofauna and infauna). Quantitative analysis like density per unit cannot be achieved, and only the relative measures of abundance (MaxN) can be applied at this moment. As the lander could help to better understand the current spatial variability on the Cosmonaut Sea shelf fauna, data here would serve as a supplement to the baseline for future comparisons [\[4\]](#page-8-2).

**Author Contributions:** J.M.: investigation, data curation, writing—original draft, and validation. X.H.: conceptualization, software, and writing—review and editing. K.L.: data curation and software. Y.H.: data curation and writing—review & editing. S.Z.: data curation. Y.Z.: data curation and writing—review and editing. Y.L.: data curation and writing—review and editing. S.C.: investigation and supervision. M.L.: investigation and supervision. X.M.: investigation. H.L.: supervision and conceptualization. W.L.: supervision, conceptualization, and resources. All authors have read and agreed to the published version of the manuscript.

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**Institutional Review Board Statement:** Although our research was about benthos in Antarctica, all the data were from the images of the lander. Therefore, we believe an ethical review process was not required for our study.

**Informed Consent Statement:** Informed consent was obtained from all individual participants included in the study.

**Data Availability Statement:** The raw data supporting the conclusions of this article will be made available by the authors on request.

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#### **References**

- <span id="page-8-0"></span>1. Busalacchi, A.J. The role of the Southern Ocean in global processes: An earth system science approach. *Antarct. Sci.* **2004**, *16*, 363–368. [\[CrossRef\]](https://doi.org/10.1017/S0954102004002196)
- <span id="page-8-1"></span>2. Convey, P.; Stevens, M.I. Antarctic Biodiversity. *Science* **2007**, *317*, 1877–1878. [\[CrossRef\]](https://doi.org/10.1126/science.1147261)
- 3. Griffiths, H.J. Antarctic marine biodiversity—What do we know about the distribution of life in the southern ocean? *PLoS ONE* **2010**, *5*, e11683. [\[CrossRef\]](https://doi.org/10.1371/journal.pone.0011683) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/20689841)
- <span id="page-8-2"></span>4. Friedlander, A.M.; Goodell, W.; Salinas-de-León, P.; Ballesteros, E.; Berkenpas, E.; Capurro, A.P.; Cárdenas, C.A.; Hüne, M.; Lagger, C.; Landaeta, M.F.; et al. Spatial patterns of continental shelf faunal community structure along the Western Antarctic Peninsula. *PLoS ONE* **2020**, *15*, e0239895. [\[CrossRef\]](https://doi.org/10.1371/journal.pone.0239895) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/33002046)
- <span id="page-8-3"></span>5. Costello, M.J.; Coll, M.; Danovaro, R.; Halpin, P.; Ojaveer, H.; Miloslavich, P. A census of marine biodiversity knowledge, resources, and future challenges. *PLoS ONE* **2010**, *5*, e12110. [\[CrossRef\]](https://doi.org/10.1371/journal.pone.0012110)
- 6. Levin, L.A.; Bett, B.J.; Gates, A.R.; Heimbach, P. Global observing needs in the deep ocean. *Front. Mar. Sci.* **2019**, *6*, 241. [\[CrossRef\]](https://doi.org/10.3389/fmars.2019.00241)
- 7. Rogers, A.; Aburto-Oropeza, O.; Appeltans, W.; Assis, J. *Critical Habitats and Biodiversity: Inventory, Thresholds and Governance*; World Resources Institute: Washington, DC, USA, 2020.
- <span id="page-8-4"></span>8. Giddens, J.; Turchik, A.; Goodell, W.; Rodriguez, M.; Delaney, D. The National Geographic Society Deep-Sea Camera System: A Low-Cost Remote Video Survey Instrument to Advance Biodiversity Observation in the Deep Ocean. *Front. Mar. Sci.* **2021**, *7*, 601411. [\[CrossRef\]](https://doi.org/10.3389/fmars.2020.601411)
- <span id="page-8-5"></span>9. Meijers, A.J.S.; Klocker, A.; Bindoff, N.L. The circulation and water masses of the Antarctic shelf and continental slope between 30 and 80◦E. *Deep. Sea Res. Part II Top. Stud. Oceanogr.* **2010**, *57*, 723–737. [\[CrossRef\]](https://doi.org/10.1016/j.dsr2.2009.04.019)
- <span id="page-8-6"></span>10. Barnes, D.; Peck, L. Vulnerability of Antarctic shelf biodiversity to predicted regional warming. *Clim. Res.* **2008**, *37*, 149–163. [\[CrossRef\]](https://doi.org/10.3354/cr00760)
- <span id="page-8-17"></span>11. Barnes, D.K.A.; Clarke, A. Antarctic marine biology. *Curr. Biol.* **2011**, *21*, 451–457. [\[CrossRef\]](https://doi.org/10.1016/j.cub.2011.04.012) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/21683895)
- <span id="page-8-13"></span>12. Linse, K.; Griffiths, H.J.; Barnes, D.K.A.; Brandt, A.; Davey, N.; David, B.; Grave, S.D.; D'Acoz, C.D.; Eléaume, M.; Glover, A.G.; et al. The macro- and megabenthic fauna on the continental shelf of the eastern Amundsen Sea, Antarctica. *Cont. Shelf Res.* **2013**, *68*, 80–90. [\[CrossRef\]](https://doi.org/10.1016/j.csr.2013.08.012)
- <span id="page-8-7"></span>13. Mou, J.F.; Liu, K.; Huang, Y.Q.; Lin, J.H.; He, X.B.; Zhang, S.Y.; Li, D.; Zu, Y.C.; Chen, Z.H.; Fu, S.J.; et al. Species Diversity and Community Structure of Macrobenthos in the Cosmonaut Sea, East Antarctica. *Diversity* **2023**, *15*, 1197. [\[CrossRef\]](https://doi.org/10.3390/d15121197)
- <span id="page-8-8"></span>14. Hunt, B.; Pakhomov, E.; Trotsenko, B.J. The macrozooplankton of the Cosmonaut Sea, east Antarctica (30◦E–60◦E), 1987–1990. *Deep. Sea Res. Oceanogr. Res. Pap.* **2007**, *54*, 1042–1069. [\[CrossRef\]](https://doi.org/10.1016/j.dsr.2007.04.002)
- <span id="page-8-16"></span>15. Van de Putte, A.P.; Jackson, G.D.; Pakhomov, E.; Flores, H.; Volckaert, F.A.M. Distribution of squid and fish in the pelagic zone of the Cosmonaut Sea and Prydz Bay region during the BROKE-West campaign. *Deep. Sea Res. Part II Top. Stud. Oceanogr.* **2010**, *57*, 956–967. [\[CrossRef\]](https://doi.org/10.1016/j.dsr2.2008.02.015)
- <span id="page-8-9"></span>16. Ran, Q.; Duan, M.G.; Wang, P.C.; Ye, Z.J.; Mou, J.F.; Wang, X.Q.; Tian, Y.J.; Zhang, C.; Qiao, H.J.; Zhang, J. Predicting the current habitat suitability and future habitat changes of Antarctic jonasfish *Notolepis coatsorum* in the Southern Ocean. *Deep. Sea Res. Part II Top. Stud. Oceanogr.* **2022**, *199*, 105077. [\[CrossRef\]](https://doi.org/10.1016/j.dsr2.2022.105077)
- <span id="page-8-10"></span>17. Sahade, R.; Lagger, C.; Torre, L.; Momo, F.; Monien, P.; Schloss, I.; Barnes, D.K.A.; Servetto, N.; Tarantelli, S.; Tatián, M.; et al. Climate change and glacier retreat drive shifts in an Antarctic benthic ecosystem. *Sci. Adv.* **2015**, *1*, e1500050. [\[CrossRef\]](https://doi.org/10.1126/sciadv.1500050)
- <span id="page-8-11"></span>18. Mou, J.F.; Liu, K.; Huang, Y.Q.; He, X.B.; Zhang, S.Y.; Wang, J.J.; Lin, J.H.; Lin, H.S.; Liu, W.H. The macro-and megabenthic fauna on the continental shelf of Prydz Bay, east Antarctica. *Deep. Sea Res. Part II Top. Stud. Oceanogr.* **2022**, *198*, 105052. [\[CrossRef\]](https://doi.org/10.1016/j.dsr2.2022.105052)
- <span id="page-8-12"></span>19. Gutt, J. Some "driving forces" structuring communities of the sublittoral Antarctic macrobenthos. *Antarct. Sci.* **2000**, *12*, 297–313. [\[CrossRef\]](https://doi.org/10.1017/S0954102000000365)
- <span id="page-8-14"></span>20. Arnaud, P.M.; Lo'pez, C.M.; Olaso, I.; Ramil, F.; Ramos-Esplá, A.A.; Ramos, A. Semi-quantitative study of macrobenthic fauna in the region of the South Shetland Islands and the Antarctic Peninsula. *Polar Biol.* **1998**, *19*, 160–166. [\[CrossRef\]](https://doi.org/10.1007/s003000050229)
- <span id="page-8-15"></span>21. Malyutina, M. Russian deep-sea investigations of Antarctic fauna. *Deep. Res Part II Top. Stud. Oceanogr.* **2004**, *51*, 1551–1570. [\[CrossRef\]](https://doi.org/10.1016/j.dsr2.2004.07.012)
- <span id="page-9-0"></span>22. Linley, T.D.; Gerringer, M.E.; Yancey, P.H.; Drazen, J.C.; Weinstock, C.L.; Jamieson, A.J. Fishes of the hadal zone including new species, in situ observations and depth records of Liparidae. *Deep. Sea Res. Part I Oceanogr. Res. Pap.* **2016**, *114*, 99–110. [\[CrossRef\]](https://doi.org/10.1016/j.dsr.2016.05.003)
- <span id="page-9-1"></span>23. Leitner, A.B.; Neuheimer, A.B.; Donlon, E.; Smith, C.R.; Drazen, J.C. Environmental and bathymetric influences on abyssal bait-attending communities of the Clarion Clipperton Zone. *Deep. Sea Res. Part I Oceanogr. Res. Pap.* **2017**, *125*, 65–80. [\[CrossRef\]](https://doi.org/10.1016/j.dsr.2017.04.017)
- <span id="page-9-2"></span>24. Riddle, M.J.; Craven, M.; Goldsworthy, P.; Carsey, F. A diverse benthic assemblage 100 km from open water under the Amery Ice Shelf, Antarctica. *Paleoceanogr. Paleoclimatol.* **2007**, *22*, PA1204. [\[CrossRef\]](https://doi.org/10.1029/2006PA001327)
- 25. Sumida, P.Y.G.; Bernardino, A.F.; Stedall, V.P.; Glover, A.G.; Smith, C.R. Temporal changes in benthic mega-faunal abundance and composition across the West Antarctic Peninsula shelf: Results from video surveys. *Deep. Res Part II Top. Stud. Oceanogr.* **2008**, *55*, 2465–2477. [\[CrossRef\]](https://doi.org/10.1016/j.dsr2.2008.06.006)
- 26. Eastman, J.T.; Amsler, M.O.; Aronson, R.B.; Thatje, S.; McClintock, J.B.; Vos, S.C.; Kaeli, J.W.; Singh, H.; Mesa, M.L. Photographic survey of benthos provides insights into the Antarctic fish fauna from the Marguerite Bay slope and the Amundsen Sea. *Antarct. Sci.* **2013**, *25*, 31–43. [\[CrossRef\]](https://doi.org/10.1017/S0954102012000697)
- <span id="page-9-3"></span>27. Ambroso, S.; Salazar, J.; Zapata-Guardiola, R.; Federwisch, L.; Richter, C.; Gili, J.M.; Teixidó, N. Pristine populations of habitatforming gorgonian species on the Antarctic continental shelf. *Sci. Rep.* **2017**, *7*, 12251. [\[CrossRef\]](https://doi.org/10.1038/s41598-017-12427-y) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/28947777)
- <span id="page-9-4"></span>28. Alberty, M.; Sprintall, J.; Mackinnon, J.; Germineaud, C.; Cravatte, S.; Ganachaud, A. Moored Observations of Transport in the Solomon Sea. *J. Geophys. Res. Oceans* **2019**, *124*, 8166–8192. [\[CrossRef\]](https://doi.org/10.1029/2019JC015143)
- <span id="page-9-5"></span>29. Langlois, T.; Williams, J.; Monk, J.; Bouchet, P.; Currey, L.; Goetze, J.; Harasti, D.; Huveneers, C.; Lerodiaconou, D.; Malcolm, H.; et al. Marine sampling field manual for benthic stereo BRUVS (Baited Remote Underwater Videos). In *Field Manuals for Marine Sampling to Monitor Australian Waters*; Przeslawski, R., Foster, S., Eds.; National Environmental Science Programme (NESP): Canberra, Australia, 2018; pp. 82–104.
- <span id="page-9-6"></span>30. Linley, T.D.; Craig, J.; Jamieson, A.J.; Priede, I.G. Bathyal and abyssal demersal bait-attending fauna of the Eastern Mediterranean Sea. *Mar. Biol.* **2018**, *165*, 159. [\[CrossRef\]](https://doi.org/10.1007/s00227-018-3413-0)
- <span id="page-9-7"></span>31. Brasier, M.J.; Grant, S.M.; Trathan, P.N.; Allcock, L.; Ashford, O.; Blagbrough, H.; Brandt, A.; Danis, B.; Downey, R.; Eléaume, M.P.; et al. Benthic biodiversity in the South Orkney Islands Southern Shelf Marine Protected Area. *Biodiversity* **2018**, *19*, 5–19. [\[CrossRef\]](https://doi.org/10.1080/14888386.2018.1468821)
- <span id="page-9-8"></span>32. Smith, J.; O'Brien, P.E.; Stark, J.S.; Johnstone, G.J.; Riddle, M. Integrating multibeam sonar and underwater video data to map benthic habitats in an East Antarctic nearshore environment. *Estuar. Coast. Shelf Sci.* **2015**, *164*, 520–536. [\[CrossRef\]](https://doi.org/10.1016/j.ecss.2015.07.036)
- <span id="page-9-9"></span>33. Post, A.L.; Lavoie, C.; Domack, E.W.; Leventer, A.; Shevenell, A.E.; Fraser, A.D. Environmental drivers of benthic communities and habitat heterogeneity on an East Antarctic shelf. *Antarct. Sci.* **2017**, *29*, 17–32. [\[CrossRef\]](https://doi.org/10.1017/S0954102016000468)
- <span id="page-9-10"></span>34. Jones, D.O.B.; Bett, B.J.; Tyler, P.A. Depth-related changes to 860 density, diversity and structure of benthic megafaunal assemblages 861 in the Fimbul ice shelf region, Weddell Sea, Antarctica. *Polar Biol.* **2007**, *862*, 1579–1592. [\[CrossRef\]](https://doi.org/10.1007/s00300-007-0319-6)
- <span id="page-9-11"></span>35. Gon, O.; Heemstra, P.C. *Fishes of the Southern Ocean*; J.L.B. Smith Institute of Ichthyology Grahamstown: Grahamstown, South Africa, 1990.
- <span id="page-9-12"></span>36. Ciaputa, P.; Siciński, J. Seasonal and annual changes in Antarctic Fur seal (*Arctocephalus gazella*) diet in the area of Admiralty Bay, King George Island, South Shetland Islands. *Pol. Polar Res.* **2006**, *27*, 171–184.
- <span id="page-9-13"></span>37. Fujiwara, Y.; Tsuchida, S.; Kawato, M.; Masuda, K. Detection of the Largest Deep-Sea-Endemic Teleost Fish at Depths of Over 2000 m Through a Combination of eDNA Metabarcoding and Baited Camera Observations. *Front. Mar. Sci.* **2022**, *9*, 945758. [\[CrossRef\]](https://doi.org/10.3389/fmars.2022.945758)
- <span id="page-9-14"></span>38. Brandt, A.; Gutt, J.; Hildebrandt, M.; Pawlowski, J.; Schwendner, J.; Soltwedel, T.; Thomsen, L. Cutting the Umbilical: New Technological Perspectives in Benthic Deep-Sea Research. *J. Mar. Sci. Eng.* **2016**, *4*, 36. [\[CrossRef\]](https://doi.org/10.3390/jmse4020036)

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