



# *Article* **Evolutionary Insights from Dental Diversity in Afro-Asian Primates**

Hao Pan <sup>1[,](https://orcid.org/0000-0002-1832-8480)2,†</sup>®, He Zhang <sup>2,3,†</sup>®, Dionisios Youlatos <sup>2,[4](https://orcid.org/0000-0001-8276-727X)</sup>®, Jing Wang <sup>1</sup>, Gang He <sup>1,2</sup>, Songtao Guo <sup>1</sup>, Kang Huang <sup>1</sup>, **Rong Hou 1,2, Ruliang Pan 1,2,5 [,](https://orcid.org/0000-0003-4467-2143) Gu Fang <sup>1</sup> , Yuli Li <sup>1</sup> , Pei Zhang 1,[\\*](https://orcid.org/0000-0003-4433-382X) and Baoguo Li 1,2,\***

- <sup>1</sup> Shaanxi Key Laboratory for Animal Conservation, College of Life Sciences, Northwest University, Xi'an 710069, China; panhao0502@163.com (H.P.); ruliang.pan@uwa.edu.au (R.P.)
- 2 International Centre of Biodiversity and Primate Conservation, Dali University, Dali 671003, China
- 3 Jiangxi Provincial Key Laboratory of Conservation Biology, College of Forestry, Jiangxi Agricultural University, Nanchang 330029, China
- <sup>4</sup> Department of Zoology, School of Biology, Aristotle University of Thessaloniki, GR-54124 Thessaloniki, Greece
- <sup>5</sup> School of Human Sciences, The University of Western Australia, Perth, WA 6009, Australia
- **\*** Correspondence: peizhang@nwu.edu.cn (P.Z.); baoguoli@nwu.edu.cn (B.L.)
- These authors contributed equally to this work.

**Abstract:** The evolutionary development and phylogenetic division between Asian and African cercopithecoids (Cercopithecidae) have attracted significant attention in genetics, molecular biology, behavior, and morphology. However, less emphasis has been placed on how they have evolved morphologically after divergence, approximately 10 million years ago (mya) for Colobinae and 5–7 mya for Cercopithecinae, corresponding to the significant variation and diversity in landscape, climate, habitat, and ecologies between the two continents. This study examines whether such variation and diversity have been reflected in dental morphology. Our findings reveal substantial differences between Hylobatidae and Cercopithecidae, as well as between Colobinae and Cercopithecinae, indicating that size-adjusted dental variation mainly reveals the diversity associated with evolution and phylogenetic inertia. Interestingly, despite the earlier divergence of Afro-Asian colobines, their Euclidean Distance is comparable to that of Afro-Asian cercopithecines. This implies that latecomers (macaques) demonstrate equivalent diversity to colobines due to their extensive dispersion and broader adaptative radiation on the same continent. Colobinae exhibit more developed premolar and molar regions. However, when post-canine teeth are considered alone, Colobinae present a significantly larger molar size than Asian Cercopithecinae but not with the African Cercopihecinae. This contradicts the hypothesis that folivorous primates (Colobinae) have larger post-canine molars than frugivorous ones (Cercopithecinae). The considerable molar size in African Cercopithecinae must be associated with their more protrusive and larger facial structure rather than a specific dietary preference, being less diverse than their Asian counterparts—a trait that has evolved phylogenetically. This study also paves the way for further exploration of facial and cranial differences between the continental groups of Cercopithecinae and Colobinae, delving deeply into diversity variation due to geographical and climatic adaptations.

**Keywords:** Cercopithecidae; dental allometry analysis; environmental and climate changes; Africa and Asia primates; natural selection and environmental adaptation

### **1. Introduction**

The evolution of catarrhine primates (Old World monkeys, gibbons, great apes, and humans) in Asia experienced significant changes during the Neogene and Quaternary following the migrations from Africa to Asia via Europe [\[1–](#page-10-0)[4\]](#page-10-1). Different groups of Asian Cercopithecidae (macaques and colobines) started their journey to varying timeframes from Africa. However, by the end of the Miocene and Early Pliocene, they settled in



**Citation:** Pan, H.; Zhang, H.; Youlatos, D.; Wang, J.; He, G.; Guo, S.; Huang, K.; Hou, R.; Pan, R.; Fang, G.; et al. Evolutionary Insights from Dental Diversity in Afro-Asian Primates. *Diversity* **2024**, *16*, 565. [https://](https://doi.org/10.3390/d16090565) [doi.org/10.3390/d16090565](https://doi.org/10.3390/d16090565)

Academic Editor: Luc Legal

Received: 25 August 2024 Revised: 4 September 2024 Accepted: 9 September 2024 Published: 11 September 2024



**Copyright:** © 2024 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license [\(https://](https://creativecommons.org/licenses/by/4.0/) [creativecommons.org/licenses/by/](https://creativecommons.org/licenses/by/4.0/)  $4.0/$ ).

a Convergence–Divergence Center (CDC) located in southeast Qinghai–Tibet Plateaus and Hengduan Mountains, a region linking East, South, and Southeast Asia [\[1\]](#page-10-0). Fossil hylobatids from India [\[5\]](#page-10-2) and in the Center [\[6](#page-10-3)[–8\]](#page-10-4) also indicate that gibbons used the same paths entering the CDC, through which Asian catarrhines dispersed to various parts of Southeast and East Asia  $[1,9-11]$  $[1,9-11]$  $[1,9-11]$ . Unlike today, the region they colonized was relatively flat. However, it underwent significant changes during the Pliocene and Quaternary, showcasing remarkable diversity in many aspects due to the accelerated uplift of the Qinghai–Tibet Plateau and Himalayas, caused by the intense collision between the Indian and European plates [\[12](#page-10-7)[,13\]](#page-10-8). This created significantly diverse geographic changes in Asia. Thus, similar to modern humans (*Homo erectus* and *H. sapiens*), Afro-Asian monkeys in Asia experienced a period of evolutionary radiation that was driven by the significant landscape diversity and the formation of land bridges among islands due to lowered sea levels of insular geography following the lowering sea level [\[1](#page-10-0)[,14](#page-11-0)[,15\]](#page-11-1).

In contrast, similar changes in geography, climate, and habitats were relatively less noticeable in Africa [\[16–](#page-11-2)[18\]](#page-11-3), such as expanding savannas and grasslands in East Africa. Under humid global conditions, savannas spread northwards and eastwards over desert areas. Between 6.3 and 6.0 mya, a significant shift in vegetation was marked by a notable decrease in woodlands. Concurrently, West and East Africa experienced very arid conditions, evidenced by the sparse tree cover in the tropical regions. These changes were linked to global climate variations caused by the formation of the Arctic ice sheet, which induced intense tropical aridity. Finally, savannas expanded at the expense of rain forests in both West and East Africa [\[19\]](#page-11-4).

These significant environmental changes and diversity would have affected the cercopithecid primates' ecology in Africa and Asia. Dental diversity, linked to dietary selection and habits, represents a vital tool for exploring how dental structures have evolved phylogenetically and shaped biomechanically following the divergence in biology, climate, and geology between the two continents. Hence, exploring the magnitude of dental diversity related to environmental adaptations between the Cercopithecinae and Colobinae in Africa and Asia is essential. Previous studies have shown that frugivorous primates have a smaller post-canine dental size relative to body size than folivorous primates [\[20](#page-11-5)[–23\]](#page-11-6). It is interesting to determine whether the African representatives, Colobinae or Cercopithecinae, exhibit larger molar size than their Asian counterparts or vice versa due to climatic, environmental, and dietary differences. This may raise the hypothesis that there may be significant continental differences in selective constraints and environmental adaptations among Cercopithecinae and Colobinae, particularly with their dental structures. A similar approach was developed by [\[24\]](#page-11-7), but focusing only on Colobinae.

Considering the importance of dental morphology for understanding phylogenetic and adaptive patterns, the present study aims to (a) comprehend the dental variation in African and Asian Cercopithecidae regarding the divergent evolutionary patterns and environmental changes specific to each continent; (b) examine whether Colobinae exhibit more significant dental variation due to the earlier continental separation, around 10.7 mya [\[25\]](#page-11-8), compared to Cercopithecinae, which separated approximately 7.0–5.5 mya [\[26\]](#page-11-9) or 5 mya [\[11\]](#page-10-6); and (c) determine whether there is a significant post-canine dental size variation between Cercopithecinae and Colobinae.

### **2. Material and Methods**

### *2.1. Material*

The studied material consists of the upper and lower teeth of African and Asian Cercopithecidae, representing different family clades with divergent ecologies (Table [1\)](#page-2-0). For comparative purposes, we also included the teeth of Asian Hylobatidae as an outgroup. Hylobatidae (i.e., gibbons or lesser apes) are quite different from great apes in terms of facial structure, showing the smallest facial size with insignificant sexual dimorphism [\[27\]](#page-11-10). They also exhibit significantly lower canine sexual dimorphism than great apes [\[28\]](#page-11-11). However, studies of gibbons' canine sexual dimorphism and comparison with

other catarrhines are still relatively uncommon (see [\[29–](#page-11-12)[33\]](#page-11-13)). Thus, they represent an excellent example for determining canine sexual dimorphism compared to that observed within the Cercopithecidae.

Continent	Family	Subfamily	Genus	Species (Sex)
Africa	Cercopithecidae	Colobinae	Cercocebus Colobus	torquatus (6F,10M) guereza (18F, 16M, 4U) polykomos (2F, 1M, 1U)
			Piliocolobus	badius (18F, 16M, 2U)
		Cercopithecinae	Procolobus Cercopithecus	<i>verus</i> (21F, 14M, 7U) mona(2F) nictitans (3M, 1U)
			Mandrillus	leucophaeus (11F, 11M)
			Papio	hamadryas (2F, 10M) ursinus (7F, 11M)
			Масаса	sylvanus (10F, 11M)
Asia		Colobinae	<b>Nasalis</b> Presbytis	larvatus (10F, 12M) chrysomelas (1F) comata (3F, 4M) melalophos (4F, 5M) rubicunda (7F, 5M)
			Pygathrix Rhinopithecus	nemaeus (11F, 16M) avunculus (3F, 3M) bieti (12F, 9M) brelichi (1F, 4M) roxellana (34F, 32M, 9U)
			Semnopithecus Trachypithecus	entellus (5F, 5M) cristatus (19F, 15M) francoisi (8F, 10M) <i>obscurus</i> (10F, 10M) phayrei (26F, 10M) vetulus (5F, 5M)
			<i>Simias</i>	concolor (2F, 1M)
		Cercopithecinae	Масаса	arctoides (3F, 3M) assamensis (3F, 3M) <i>fascicularis</i> (3F, 3M) fuscata (4F, 3M) mulatta (4F, 3M) nemestrina (3F, 3M) nigra (3F, 3M) radiata (3F, 3M) silenus (3F, 3M) sinica (4F, 4M) thibetana (2F, 4M)
	Hylobatidae		Hylobates	agilis (10F, 10M, 2U) <i>lar</i> (13F, 20M, 4U) muelleri (13U) pileatus (1F, 2M)
			Hoolock	hoolock (8F, 11M)

<span id="page-2-0"></span>**Table 1.** The Cercopithecidae and Hylobatidae studied \*.

\* Please see the Acknowledgments for data collection. F: female; M: male; and U: unknown sex.

The materials used in this study are housed in various institutes, universities, and museums (see Acknowledgments for details). All specimens included are adults, identified by the full eruption of M3s.

### *2.2. Methods*

A digital caliper (Mitutoyo Inc., Tokyo, Japan) with an accuracy of 0.01 mm was used to take the maximum mesiodistal (length) and buccolingual (width) measurements for each tooth on the maxilla and mandible. Tooth size (crown area) was multiplied by length with width. Cranial length from the tip of occipital protuberance to the alveolar between the right and left I<sup>1</sup> was also measured as a proxy for body size, e.g., [\[20,](#page-11-5)[34\]](#page-11-14).

Allometric analyses ( $Y = aX<sup>b</sup>$ ) have been widely used across various fields, such as biology, ecology, paleontology, and anthropology, to explore how an organism's structure or function changes relative to its size. These analyses provide valuable insights into growth patterns, physiological constraints, evolutionary adaptations, and ecological diversity [\[35\]](#page-11-15). When transformed using logarithms, this allometric formula becomes a linear regression,  $logY = loga + blogX$ , where Y (the dependent variable) represents the square root of the tooth crown area, and X (the independent variable) represents body size, approximated by its proxy, cranial length.

To delve into dental variation, we used the residual, a size-adjusted value, to indicate how far an individual deviates from the regression line of the allometric analysis. The following equation was used:

Residual = antilog (observed value-predicted value).

The observed values are those of the raw measurements, and the predicted values are those calculated from the allometric formula [\[24,](#page-11-7)[36](#page-11-16)[–38\]](#page-11-17).

Ordinary least-square (OLS) regression analysis was used for allometric analysis (log-transformed data). OLS is considered more suitable for calculating residuals for size adjustment (e.g., [\[39,](#page-11-18)[40\]](#page-11-19)).

Three types of residuals were calculated:

- (1) Comparison between Cercopithecidae and Hylobatidae aims to explore the relationship between teeth and body size of catarrhines in Africa and Asia. As seen below, residuals were further analyzed using multivariate discriminant function analysis (DFA).
- (2) Comparison within the Cercopithecidae aims to decipher differences between the representatives of Colobinae and Cercopithecinae in Africa and Asia. Residuals of the four groups (African Colobinae, African Cercopithecinae, Asian Colobinae, and Asian Cercopithecinae) were also analyzed using DFA.
- (3) Comparing the post-canine teeth size within the Cercopithecidae aims to determine the magnitude of dental variation for Colobinae and Cercopithecinae in Africa and Asia.

DFA is a standard multivariate analysis for classification and dimensionality reduction, used to identify a linear combination of variables that best separates two or more groups or classes. Its main objective is to maximize the between-class and within-class variance ratio, finding a linear combination of variables that maximizes the differences between the means of the groups while minimizing the variation within each category [\[41\]](#page-11-20). We performed DFA to analyze residuals generated in comparisons (1) and (2).

Furthermore, Analysis of Variance (ANOVA) was employed to evaluate differences between the studied groups based on DFA scores (coordinates).

All the analyses were carried out by IBM SPSS Statistics v. 20.

### **3. Results**

The results of the first two types of allometric analyses indicate that each tooth size (area) presents a significant positive relationship (*p* < 0.001) with body size, represented by cranial length. The primary purpose of this study is to explore the variation between the different groups of Cercopithecidae, correlation coefficients, and regression slopes, indicating that the allometric scale between teeth and body size is not reported here. Their corresponding residuals were further explored via DFAs. The results (eigenvalues, percentage of explained variance, coefficients of eigenvectors on the first two axes) of the three-group DFA (Colobinae, Cercopithecinae, Hylobatidae) and the four-group DFA

(African Colobinae, African Cercopithecinae, Asian Colobinae, Asian Cercopithecinae) are presented in Table [2.](#page-4-0)



<span id="page-4-0"></span>**Table 2.** Eigenvalues, percentages of explained variance, and coefficients of eigenvectors of the first two DFA axes for the three-group DFA (Colobinae, Cercopithecinae, Hylobatidae) and the four-group DFA (African Colobinae, African Cercopithecinae, Asian Colobinae, Asian Cercopithecinae).

U: upper teeth areas; L: lower teeth areas.

Regarding the three-group analysis, the first and second DFs explain 63.0% and 37.0% of the total variance. On the positive side of DF1, the variables with the highest contribution are UM3 (0.66), LP3 (0.62), and UP4 (0.50), and on the negative side of DF1, UP3 ( $-0.56$ ) and LC (−0.61). Figure [1](#page-5-0) shows the dispersal pattern of the three groups along DF1 and DF2. The ANOVA results of the discriminant scores show that Hylobatidae, principally located on the negative side, are significantly distinct  $(p < 0.001)$  from both Colobinae and Cercopithecinae, both latter groups showing significant overlap along DF1.

Along DF2, the positive side of the axis is characterized by UM1 (0.89) and LM3 (0.50), whereas the negative side is characterized by UM2 (-1.12) and LP4 (-0.676). Along this axis, Hylobatidae shows substantial separation from Colobinae ( $p = 0.002$ ) and Cercopithecinae (*p* < 0.001). Colobinae and Cercopithecinae are also significantly separated (*p* < 0.001). Our results indicate that Hylobatidae are distinct from Cercopithecidae in DF1 and DF2, whereas the monkey subfamilies are separated only along DF2.

Concerning the four-group analysis, the first and second axes account for 53.0% and 31.8%, respectively. UM1(0.84) and LP3 (0.75) contribute significantly to the positive side of DF1, whereas LM1 (-0.82) and UP4 (-0.70) are significant contributors to the negative side of the axis. The dispersal pattern of the four groups of Asian and African Cercopithecidae along DF1 and DF2 are illustrated in Figure [2.](#page-5-1) Along DF1, Cercopithecinae (African and Asian) appear significantly distinct  $(p < 0.001)$  from Colobinae (African and Asian). However, no significant separation between African and Asian Colobinae and African and Asian Cercopithecinae is observed.

<span id="page-5-0"></span>

Figure 1. Individual scatter plots of the Hylobatidae, Colobinae, and Cercopithecinae along DFA's first and second axes.

<span id="page-5-1"></span>

**Figure 2.** DFA among Asian Colobinae, African Colobinae, Asian Cercopithecinae, and African Cercopithecinae. The Euclidean Distance (Eu) between Asian and African Colobinae is 2.04, and between Asian and African Cercopithecinae, it is 2.06.

Along DF2, LM3 (0.72) and UM1 (0.61) are major contributors on the positive side of the axis, whereas UM2 ( $-1.14$ ), LP4 ( $-0.68$ ), and UI1 ( $-0.58$ ) show noticeable negative contributions. Asian Colobinae and Cercopithecinae show significant separation from their African counterparts (Colobinae: *p* < 0.001; Cercopithecinae: *p* < 0.05). The Euclidean Distances reflecting the magnitude of separation between the African and Asian representatives of the two subfamilies on the first two DFA axes are comparable: Colobinae = 2.04; Cercopithecinae = 2.06. Our results indicate that DF1 separates between the two subfamilies, whereas DF2 separates the Asian and African representatives.

Figure [3](#page-6-0) illustrates the residual differences between Cerocopithecidae groups from different continents, derived from an allometric analysis of both post-canine teeth and body size. The slope for both upper and lower molars exceeds 0.80 against body size, with a correlation coefficient of 0.86, indicating a strong correlation (*p* < 0.001). The ANOVA results indicate that African Cercopithecinae have significantly higher values than the other groups. Both African and Asian Colobinae show substantially higher values than Asian Cercopithecinae, which display the lowest variation compared to the different groups. Finally, the difference between Asian and African Colobinae is not statistically significant. significant.

results indicate that African Cercopithecinae have significantly higher values than the

<span id="page-6-0"></span>

**Figure 3.** Comparison of residuals among the Cercopithecidae groups derived from the allometric **Figure 3.** Comparison of residuals among the Cercopithecidae groups derived from the allometric analysis between post-canine teeth and body size (proxied by cranial length). analysis between post-canine teeth and body size (proxied by cranial length).

Regarding canine sexual dimorphism, we calculated the degree of lower canines among the three studied groups. Hylobatidae are characterized by the lowest degree among the three studied groups. Hylobatidae are characterized by the lowest degree of sexual dimorphism  $(SD = 0.0596)$ , whereas Cercopithecinae show the highest degree  $\frac{\partial G}{\partial \rho}$  = 0.0696),  $\frac{\partial G}{\partial \rho}$  = 0.05966), whereas Cercopithecinae show the highest degree (SD  $\frac{\partial G}{\partial \rho}$ ),  $\frac{\partial G}{\partial \rho}$  $(SD = 0.6062)$  (Figure [4\)](#page-6-1). Colobinae show moderate lower canine sexual dimorphism  $(SD = 0.2195)$  $(SD = 0.3185).$ Regarding canine sexual dimorphism, we calculated the degree of lower canines

<span id="page-6-1"></span>

**Figure 4.** Sexual dimorphism (SD = 1 − Female/Male) of mandibular canines in Cercopithecinae, **Figure 4.** Sexual dimorphism (SD = 1 − Female/Male) of mandibular canines in Cercopithecinae, Colobinae, and Hylobatidae. Colobinae, and Hylobatidae.

## **4. Discussion 4. Discussion**

Dental morphology, combined with the osteo-muscular morphology of the skull and Dental morphology, combined with the osteo-muscular morphology of the skull and mandible, strongly correlates with body size  $[42]$  and is tightly linked to food items' selection, acquisition, and process. Therefore, it provides insightful and diverse information on the dietary ecology of organisms and, ultimately, on the climate, geography, and environment where they dwell. Moreover, teeth are the most available fossil elements frequently used to explore evolutionary patterns, phylogenetic relationships, and dietary diversity selection. In this study, we used basic metrics (tooth length and width) to analyze the dental variation in African and Asian Cercopithecidae concerning the divergent evolutionary patterns and environmental changes specific to each continent to determine whether Colobi-<br> nae exhibit more significant dental variation compared to Cercopithecinae and to identify whether there is a significant post-canine dental size variation between Cercopithecinae and Colobinae.

### *4.1. Evolutionary and Phylogenetic Diversity*

Regarding dental morphology, the two studied primate catarrhine families appeared well separated. Our analyses showed that mandibular canines (LCs), premolars (LP3 and LP4), and maxillary molars (UM2 and UM3) are responsible for the differences between Cercopithecidae and Hylobatidae (Figure [1,](#page-5-0) Table [1\)](#page-2-0). Fossil evidence indicates that hominoids (Hylobatidae + Hominidae) and Cercopithecoids (Victoriapithecidae + Cercopithecidae) most likely diverged around 23.0 mya in the Early Miocene [\[43\]](#page-11-22). During this differentiation, there was an evolutionary reduction in prognathism and tooth size, with gibbons exhibiting smaller teeth relative to body size, especially the mandibular canines, and very low canine sexual dimorphism (Figure [4\)](#page-6-1), most likely related to their monogamous mating system [\[44,](#page-11-23)[45\]](#page-11-24). These differences may reflect maxillary, mandibular, and dental morphological selections that can trace the divergent evolutionary histories of the two groups, which have adopted distinct behavioral and ecological selections in Africa and Asia.

Even though Cercopithecidae appeared relatively homogeneous compared to Hylobatidae, our within-family analysis demonstrated two exciting findings. First, there was a clear distinction between the two subfamilies, Colobinae and Cercopithecinae (Figure [2,](#page-5-1) DF1), most likely reflecting evolutionary and phylogenetic disparities between the two groups. Secondly, we identified geographic separation (Africa vs. Asia) within each subfamily (Figure [2,](#page-5-1) DF2), probably related to the climatic, vegetational, and environmental differences and diversities between the two continents during the Neogene and Quaternary. The evolutionary history of Asian and African cercopithecids, after their divergence during the Neogene, has been significantly influenced by climatic and geographic diversity [\[46](#page-11-25)[,47\]](#page-12-0), followed by remarkable environmental shifts during their dispersal from Africa to Asia [\[1,](#page-10-0)[25,](#page-11-8)[26\]](#page-11-9).

As addressed above, the ancestors of Asian cercopithecids left Africa and moved eastwards via Europe following the collision of the Afro-Arabian plate with Eurasia during the early Miocene (23.8–18 mya). These events allowed different groups of cercopithecids to disperse towards Asia in various periods. Colobinae most likely migrated first in the Middle Miocene, around 10.9 mya (9.6–12.3) [\[43\]](#page-11-22), whereas *Macaca* probably migrated much later in the Late Miocene (7.0~6.7 mya) [\[26\]](#page-11-9). Ultimately, they reached a Convergence– Divergence Center (CDC) linking South, East, and Southeast Asia at the southeast corner of the Qinghai–Tibet Plateau and Hengduan Mountains during the Miocene–Pliocene [\[1\]](#page-10-0). Colobinae spread and diverged in South, East, and Southeast Asia [\[48\]](#page-12-1). On the other hand, the cercopithecine genus *Macaca* (except *M. sylvanus* in North Africa) was widely spread in East and South Asia through alternative dispersal paths [\[1,](#page-10-0)[49\]](#page-12-2).

Subsequently, environmental and climatic changes in Africa and Asia during the Pliocene and Quaternary further shaped the divergent evolution of these groups. Over the last 3 mya, East Africa has experienced a long-term drying trend related to the formation of the East African Rift Valley [\[50\]](#page-12-3). Throughout the Pliocene and later during the Pleistocene, tectonic changes and glaciation in the northern hemisphere increased climate variability, reduced the rainfall, and caused shifts from woodlands to grasslands and even more open areas [\[51](#page-12-4)[,52\]](#page-12-5), leading to the formation of modern African environments [\[53\]](#page-12-6). Consequently, about 90% of Africa has a tropical climate with higher temperatures than the other continents, characterized by high seasonal and diurnal temperature variation and wet seasons [\[54\]](#page-12-7).

In Asia, changes were more drastic. The collision between the Indian and the Eurasian plates generated the accelerated uplift of the Qinghai–Tibet Plateau (with an average altitude of 4500 m), linking it to the Himalayas and the Hengduan Mountains [\[55](#page-12-8)[–57\]](#page-12-9), profoundly influencing Asian climate and environmental and biodiversity evolution [\[58](#page-12-10)[–61\]](#page-12-11). These changes were especially evident in areas such as South, East, and Southeast Asia, where the ancestors of extant Asian Cercopithecids dispersed and radiated [\[11,](#page-10-6)[26,](#page-11-9)[62\]](#page-12-12).

In South Asia, the environment of the Indian subcontinent experienced a shift from forests and shrubs to grasslands during the Miocene and Pliocene [\[63](#page-12-13)[–65\]](#page-12-14) with intense monsoonal rains, especially during the Early and Middle Pleistocene [\[66\]](#page-12-15). Its position is pivotal for animal migrations and the only path for Asian macaques and humans toward East and Southeast Asia [\[11](#page-10-6)[,26](#page-11-9)[,67\]](#page-12-16).

During the Quaternary, in East Asia, the environmental and geographic diversity was higher than in other continents, providing suitable ecological and environmental conditions for the dispersal of Cercopithecids [\[4\]](#page-10-1). The eastern coastal plains were humid, with moist tropical and subtropical forests covering southern regions and deserts with meadows and alpine and subalpine woodland habitats [\[61](#page-12-11)[,68\]](#page-12-17). That created a tremendous convergence– divergence faunal and floral center [\[1](#page-10-0)[,69\]](#page-12-18). Finally, Southeast Asia was dominated by a mix of savanna, open woodlands, and evergreen forests throughout the Pleistocene. These conditions were ideal for early hominin and nonhuman primate survival [\[70\]](#page-12-19). Moreover, during the Last Glacial Period (LGP; 125–10 kya) and particularly in the Last Glacial Maximum (LGM; 23–19 kya), a north–south savanna land bridge connecting the Malaysian Peninsula to the major Indonesian islands established shelter for "rain forest" refugia [\[71](#page-12-20)[,72\]](#page-12-21). These were most likely nurtured by the double monsoon system, southwest between May and August, transforming moisture from the Indian Ocean, and northeast between September and April, conveying water from the South China Sea [\[15\]](#page-11-1).

Considering the geological, climatic, and environmental differences between the two continents, it is unsurprising that both Cercopithecinae and Colobinae exhibit the observed **Distribution** and *Diversion* and *Diversion Diversion and Asian representatives. These differences* are also evident in their similarly high Euclidean Distances (Figure [2\)](#page-5-1), even though the divergence time of the Colobinae is much older than that of the Cercopithecinae [\[11](#page-10-6)[,26](#page-11-9)[,43\]](#page-11-22). This may be related to the broader past and present Asian distribution of the cercopithecine genus *Macaca* ([Fig](#page-8-0)ure 5), currently found in 20 Asian countries, covering more than  $5 \times 10^6$  km<sup>2</sup> in primary, secondary, deciduous, coniferous, riverine, and mangrove f[ores](#page-12-22)ts [73] and displaying a significant phylogenetic div[ersi](#page-12-2)ty [49].

<span id="page-8-0"></span>

**Figure 5.** Fossil distribution of Colobinae and Cercopithecinae in Asia [3,7,8,11,74–100]. **Figure 5.** Fossil distribution of Colobinae and Cercopithecinae in Asia [\[3,](#page-10-9)[7,](#page-10-10)[8](#page-10-4)[,11,](#page-10-6)[74](#page-12-23)[–100\]](#page-13-0).

#### *4.2. Dental Structure and Dietary Selection*

Our multivariate analyses showed that the separation between Colobinae and Cercopithecinae (Figure [2\)](#page-5-1) was mainly due to the contribution of several dental variables, such as UM1 (0.84), LP3 (0.75), LM1 (−0.82), and UP4 (−0.70) along DF1, as well as LM3  $(0.72)$ , UM1  $(0.61)$ , UM2  $(-1.14)$ , and LP4  $(-0.68)$  along DF2. These variables represent the size of the post-canine teeth, premolars, and molars, suggesting that Colobinae have larger premolars and molars than Cercopithecinae. This difference may be related to the overall folivorous diet of Colobinae. Compared to frugivorous and insectivorous primates, folivorous monkeys have larger premolars. This increase may result from premolar molarization [\[101\]](#page-13-1) or accommodate considerable loading while taking leaves and biting stiff leaf fibers during mastication [\[102\]](#page-13-2). This post-canine morphology is closely related to this dietary specialization. As leaves have low nutritional quality, folivorous primates must ingest large quantities to meet their metabolic demands [e.g., [\[103\]](#page-13-3)], implying the frequent use of premolars and molars to leverage food loading, leading to repetitive crushing, chewing, and grinding [\[104\]](#page-13-4). On the other hand, Cercopithecinae are mainly frugivorous [\[105](#page-13-5)[–107\]](#page-13-6) or omnivorous with a flexible dietary strategy [\[108\]](#page-14-0). They tend to display great ecological flexibility and adjust their diet, group size, habitat use, and home range size according to environmental shifts [\[109\]](#page-14-1).

Additionally, our residual analysis of molars (Figure [3\)](#page-6-0) showed that Colobinae have larger molars, as suggested by previous studies [\[20](#page-11-5)[–23\]](#page-11-6), when compared to Asian Cercopithecinae, but not when compared to African Cercopithecinae. The latter possess massive molars, most likely linked to their more pronounced and prognathic facial structure, particularly in *Mandrillus* and *Papio*. This observation aligns with previous studies [\[23\]](#page-11-6), indicating a derived phylogenetic trait. Thus, the hypothesis regarding post-canine tooth differentiation seems applicable between Colobinae and Asian Cercopithecinae but not their African counterparts, primarily known for consuming fruits and seeds [\[110](#page-14-2)[,111\]](#page-14-3). Plio-Pleistocene papionin fossil evidence, particularly for *Papio* and *Theropithecus*, suggests that their diet closely resembled that of contemporary species, mainly comprising the herbaceous layer and mostly monocotyledons [\[112\]](#page-14-4). Thus, the well-developed post-canine molars in African Cercopithecinae may be advantageous for accommodating hard-object foods and may be linked to evolutionary and phylogenetic factors.

Finally, another interesting finding of this study is the lack of significant dental variation between African and Asian Colobinae (Figure [2;](#page-5-1) DF1), exhibiting very similar postcanine molar size (Figure [4\)](#page-6-1). This contrasts with results from studies based on absolute dental size [\[24\]](#page-11-7). These findings imply that, despite the significant diversity in dietary habits among African and Asian Colobinae related to different environmental adaptations [\[113–](#page-14-5)[115\]](#page-14-6), they display very similar molar patterns relative to body size, supporting their classification as folivorous primates [\[103](#page-13-3)[,116–](#page-14-7)[118\]](#page-14-8).

### **5. Conclusions**

Based on an extensive morphometric database and references, the present study explored the variations in size-adjusted dental diversity between Hylobatidae and Cercopithecidae and between African and Asian Colobinae and Cercopithecinae. The findings primarily highlight differences associated with evolutionary history, phylogeny, geographic separation, and environmental adaptation. Notably, although Colobinae split earlier than Cercopithecinae, they exhibit less diversity variation. This is likely due to their more restrained geographic distribution and less varied environments. The previously suggested hypothesis that folivorous primates show more prominent post-canine teeth appears valid only when comparing Colobinae to Asian Cercopithecinae. In contrast, African Cercopithecinae, such as *Mandrillus* and *Papio*, exhibit significantly larger post-canine teeth than the other Cercopithecidae, probably linked to their more pronounced and prognathic facial structure. Cercopithecinae are also characterized by a high degree of canine sexual dimorphism, implying different physiological, ecological, and behavioral adaptations between sexes within the same social systems [\[119,](#page-14-9)[120\]](#page-14-10). Thus, further studies on dental variation

and its relationship with body size across sexes are needed to better understand their evolutionary and adaptive interplay [\[121–](#page-14-11)[123\]](#page-14-12). Additionally, further studies on the differences in facial and cranial structure between Cercopithecinae and Colobinae, as well as the continental groups within each subfamily, are vital for clarifying the patterns of post-canine differentiation observed in this study.

**Author Contributions:** H.P. and H.Z. designed and executed the experiments. R.P., D.Y. and H.P. wrote and revised the manuscript. P.Z. and B.L. carried out project administration. J.W., R.H., G.F. and Y.L. collected the data. H.P. and H.Z. edited a manuscript draft. G.H., S.G., K.H., P.Z. and B.L. organized the funding acquisition: All authors have read and agreed to the published version of the manuscript.

**Funding:** This research was funded by National Natural Science Foundation of China (32371563, 32170507, 32400414, 32170515, 32370534, 32070450, 32071495, 32101238, 32300413), and the Key Research and Development Program of Shaanxi (2024NC-YBXM-116).

**Institutional Review Board Statement:** Not applicable.

**Data Availability Statement:** The corresponding authors' data supporting this study's findings are available upon reasonable request.

**Acknowledgments:** We thank the following institutes, universities, and museums for their support of data collection: Kunming Institute of Zoology, and Institute of Zoology, Chinese Academy of Sciences, Field Museum of Natural History, Chicago, IL, USA, Guangxi Forest Department, Nanning, Guangxi, China; National Museum of Natural History, Washington, DC, USA, Guangdong Institute of Entomology, Guangzhou, Guangdong, China, Guangxi Forest Department, Nanning, Guangxi, China, Guangxi Medical College, Nanning, Guangxi, China; Muséum National d'Histoire Naturelle, Paris; Nanchong Teacher's College, Nanchong, Sichuan, China; Yunnan University, Kunming, Yunnan, China; Royal College of Surgeons, London, Department of Mammalogy, British Museum (Natural History), London; Zoological Reference Collection, National University of Singapore. We thank the institutes, universities, and museums for their support in data collection. Our final thanks go to John Smith for his generous help editing the manuscript.

**Conflicts of Interest:** The authors declare no conflicts of interest.

### **References**

- <span id="page-10-0"></span>1. Zhang, H.; Lu, J.; Tang, S.; Huang, Z.; Cui, L.; Lan, D.; Wang, H.; Hou, R.; Xiao, W.; Guo, S.; et al. Southwest China, the Last Refuge of Continental Primates in East Asia. *Biol. Conserv.* **2022**, *273*, 109681. [\[CrossRef\]](https://doi.org/10.1016/j.biocon.2022.109681)
- 2. Delson, E. Hominidae other than Ponginae in Eastern Asia: An Updated Survey. *Asian Paleoprimatol.* **2003**, *3*, 48–51.
- <span id="page-10-9"></span>3. Wu, X.Z.; Poirier, F.E. *Human Evolution of China. A Metric Description of the Fossils and a Review of the Sites*; Oxford University Press: New York, NY, USA, 1995.
- <span id="page-10-1"></span>4. Jablonski, N.G.; Whitfort, M.J. Environmental change during the Quaternary in East Asia and its consequences for mammals. *Rec. West. Aust. Mus.* **1999**, *57*, 307–315.
- <span id="page-10-2"></span>5. Gilbert, C.C.; Ortiz, A.; Pugh, K.D.; Campisano, C.J.; Patel, B.A.; Singh, N.P.; Fleagle, J.G.; Patnaik, R. New Middle Miocene Ape (Primates: Hylobatidae) from Ramnagar, India Fills Major Gaps in the Hominoid Fossil Record. *Proc. R. Soc. B Biol. Sci.* **2020**, *287*, 20201655. [\[CrossRef\]](https://doi.org/10.1098/rspb.2020.1655) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/32900315)
- <span id="page-10-3"></span>6. Ma, S.L. Probe on the Chinese Origin of Gibbons (*Hylobates*). *Acta Theriol. Sin.* **1979**, *17*, 13–23.
- <span id="page-10-10"></span>7. Jablonski, N.G.; Chaplin, G. The Fossil Record of Gibbons. In *The Gibbons. Developments in Primatology: Progress and Prospects*; Whittaker, D.J., Lappan, S., Eds.; Springer: New York, NY, USA, 2009; pp. 111–130.
- <span id="page-10-4"></span>8. Harrison, T. The Fossil Record and Evolutionary History of Hylobatids. In *Evolution of Gibbons and Siamang. Developments in Primatology: Progress and Prospects*; Reichard, U.H., Hirai, H., Barelli, C., Eds.; Springer: New York, NY, USA, 2016; pp. 91–110.
- <span id="page-10-5"></span>9. Iwamoto, M.; Hasegawa, Y.; Koizumi, A. A Pliocene colobine from the Nakatsu Group, Kanagawa, Japan. *Anthropol. Sci.* **2005**, *113*, 123–127. [\[CrossRef\]](https://doi.org/10.1537/ase.04S017)
- 10. Harrison, T.; Jin, C.Z.; Zhang, Y.Q.; Wang, Y.; Zhu, M. Fossil Pongo from the Early Pleistocene Gigantopithecus Fauna of Chongzuo, Guangxi, Southern China. *Quat. Int.* **2014**, *354*, 59–67. [\[CrossRef\]](https://doi.org/10.1016/j.quaint.2014.01.013)
- <span id="page-10-6"></span>11. Delson, E. Fossil Macaques, Phyletic Relationships and a Scenario of Deployment. In *The Macaques: Studies in Ecology, Behavior and Evolution*; Lindburg, D., Ed.; Van Nostrand Reinhold: New York, NY, USA, 1980; Volume 10, pp. 10–30.
- <span id="page-10-7"></span>12. Rowley, D.B. Age of Initiation of Collision Between India and Asia: A Viewer of Stratigraphic Data. *Earth Planet. Sci. Lett.* **1996**, *145*, 1–13. [\[CrossRef\]](https://doi.org/10.1016/S0012-821X(96)00201-4)
- <span id="page-10-8"></span>13. Bouilhol, P.; Jagoutz, O.; Hanchar, J.M.; Dudas, F.O. Dating the India–Eurasia collision through arc magmatic records. *Earth Planet. Sci. Lett.* **2013**, *366*, 163–175. [\[CrossRef\]](https://doi.org/10.1016/j.epsl.2013.01.023)
- <span id="page-11-0"></span>14. Chauhan, P.R. The Indian Subcontinent and 'Out of Africa I'. In *Out of Africa I: The First Hominin Colonization of Eurasia*; Fleagle, J.G., Shea, J.J., Grine, F.E., Baden, A.L., Leakey, R.E., Eds.; Springer: Dordrecht, The Netherlands, 2010; pp. 145–164.
- <span id="page-11-1"></span>15. Patterson, E.W.; Johnson, K.R.; Griffiths, M.L.; Kinsley, C.W.; McGee, D.; Du, X.; Pico, T.; Wolf, A.; Ersek, V.; Mortlock, R.A.; et al. Glacial Changes in Sea Level Modulated Millennial-scale Variability of Southeast Asian Autumn Monsoon Rainfall. *Proc. Natl. Acad. Sci. USA* **2023**, *120*, e2219489120. [\[CrossRef\]](https://doi.org/10.1073/pnas.2219489120)
- <span id="page-11-2"></span>16. Burke, K.; Wilkinson, M.J. Landscape evolution in Africa during the Cenozoic and Quaternary—The legacy and limitations of Lester C. King. *Can. J. Earth Sci.* **2016**, *53*, 1089–1102. [\[CrossRef\]](https://doi.org/10.1139/cjes-2016-0099)
- 17. Stokes, C.R.; Abram, N.J.; Bentley, M.J.; Edwards, T.L.; England, M.H.; Foppert, A.; Jamieson, S.S.R.; Jones, R.S.; King, M.A.; Lenaerts, J.T.M.; et al. Response of the East Antarctic Ice Sheet to Past and Future Climate Change. *Nature* **2022**, *608*, 275–286. [\[CrossRef\]](https://doi.org/10.1038/s41586-022-04946-0) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/35948707)
- <span id="page-11-3"></span>18. Elbelrhiti, H. Initiation and early development of barchan dunes: A case study of the Moroccan Atlantic Sahara desert. *Geomorphology* **2012**, *138*, 181–188. [\[CrossRef\]](https://doi.org/10.1016/j.geomorph.2011.08.033)
- <span id="page-11-4"></span>19. Bonnefille, R. Cenozoic vegetation, climate changes and hominid evolution in tropical Africa. *Glob. Planet. Chang.* **2010**, *72*, 390–411. [\[CrossRef\]](https://doi.org/10.1016/j.gloplacha.2010.01.015)
- <span id="page-11-5"></span>20. Kay, R.F. Allometry and Early Hominids. *Science* **1975**, *189*, 61–64.
- 21. Kay, R.F. The functional adaptations of primate molar teeth. *Am. J. Phys. Anthropol.* **1975**, *43*, 195–216. [\[CrossRef\]](https://doi.org/10.1002/ajpa.1330430207)
- 22. Pirie, P.L. Allometric scaling in the postcanine dentition with reference to primate diets. *Primates* **1978**, *19*, 583–591. [\[CrossRef\]](https://doi.org/10.1007/BF02373318)
- <span id="page-11-6"></span>23. Scott, J.E. Folivory, frugivory, and postcanine size in the cercopithecoidea revisited. *Am. J. Phys. Anthropol.* **2011**, *146*, 20–27. [\[CrossRef\]](https://doi.org/10.1002/ajpa.21535)
- <span id="page-11-7"></span>24. Pan, R.L. Dental morphometric variation between African and Asian colobines, with special reference to the other Old World monkeys. *J. Morphol.* **2006**, *267*, 1087–1098. [\[CrossRef\]](https://doi.org/10.1002/jmor.10463)
- <span id="page-11-8"></span>25. Roos, C.; Zinner, D.; Kubatko, L.S.; Schwarz, C.; Yang, M.Y.; Meyer, D.; Nash, S.D.; Xing, J.; Batzer, M.A.; Brameier, M.; et al. Nuclear Versus Mitochondrial DNA: Evidence for Hybridization in Colobine Monkeys. *BMC Evol. Biol.* **2011**, *11*, 77. [\[CrossRef\]](https://doi.org/10.1186/1471-2148-11-77)
- <span id="page-11-9"></span>26. Roos, C.; Kothe, M.; Alba, D.M.; Delson, E.; Zinner, D. The Radiation of Macaques out of Africa: Evidence from Mitogenome Divergence Times and the Fossil Record. *J. Hum. Evol.* **2019**, *133*, 114–132. [\[CrossRef\]](https://doi.org/10.1016/j.jhevol.2019.05.017)
- <span id="page-11-10"></span>27. Balolia, K.L.; Soligo, C.; Lockwood, C.A. Sexual Dimorphism and Facial Growth Beyond Dental Maturity in Great Apes and Gibbons. *Int. J. Primatol.* **2013**, *34*, 361–387. [\[CrossRef\]](https://doi.org/10.1007/s10764-013-9666-z)
- <span id="page-11-11"></span>28. Kelley, J. Sexual dimorphism in canine shape among extant great apes. *Am. J. Phys. Anthropol.* **1995**, *96*, 365–389. [\[CrossRef\]](https://doi.org/10.1002/ajpa.1330960405) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/7604892)
- <span id="page-11-12"></span>29. Balolia, K. Craniodental Sexual Dimorphism Among Hylobatids. *Int. J. Primatol.* **2021**, *42*, 737–758. [\[CrossRef\]](https://doi.org/10.1007/s10764-021-00233-3)
- 30. Leutenegger, W.; Shell, B. Variability and sexual dimorphism in canine size of Australopithecus and extant hominoids. *J. Hum. Evol.* **1987**, *16*, 359–367. [\[CrossRef\]](https://doi.org/10.1016/0047-2484(87)90066-2)
- 31. Frisch, J.E. Sex-differences in the canines of the gibbon (Hylobates lar). *Primates* **1963**, *4*, 1–10. [\[CrossRef\]](https://doi.org/10.1007/BF01659148)
- 32. Plavcan, J.M. *Sexual Selection, Measures of Sexual Selection, and Sexual Dimorphism in Primates*; Cambridge University Press: Cambridge, UK, 2004.
- <span id="page-11-13"></span>33. Plavcan, J.M.; van Schaik, C.P. Intrasexual competition and canine dimorphism in anthropoid primates. *Am. J. Phys. Anthropol.* **1992**, *87*, 461–477. [\[CrossRef\]](https://doi.org/10.1002/ajpa.1330870407)
- <span id="page-11-14"></span>34. Pan, R.; Oxnard, C. Dental Variation among Asian Colobines (Nonhuman Primates): Phylogenetic Similarities or Functional Correspondence? *Zool. Stud.* **2003**, *42*, 93–105.
- <span id="page-11-15"></span>35. Martin, R.D.; Genoud, M.; Hemelrijk, C.K. Problems of Allometric Scaling Analysis: Examples from Mammalian Reproductive Biology. *J. Exp. Biol.* **2005**, *208 (Pt 9)*, 1731–1747. [\[CrossRef\]](https://doi.org/10.1242/jeb.01566)
- <span id="page-11-16"></span>36. Pan, R.; Oxnard, C. Metrical dental analysis on golden monkey (*Rhinopithecus roxellana*). *Primates* **2001**, *42*, 75–89. [\[CrossRef\]](https://doi.org/10.1007/BF02640691)
- 37. Smith, R.J. On the Definition of Variables in Studies of Primate Dental Allometry. *Am. J. Phys. Anthropol.* **1981**, *55*, 323–329. [\[CrossRef\]](https://doi.org/10.1002/ajpa.1330550306) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/6791504)
- <span id="page-11-17"></span>38. Pan, L.R. Dental Variation among Asian Colobines, with Specific Reference to the Macaques on the Same Continent. *Zool. Res.* **2007**, *28*, 569–579.
- <span id="page-11-18"></span>39. Warton, D.I.; Wright, I.J.; Falster, D.S.; Westoby, M. Bivariate line-fitting methods for allometry. *Biol. Rev. Camb. Philos. Soc.* **2006**, *81*, 259–291. [\[CrossRef\]](https://doi.org/10.1017/S1464793106007007)
- <span id="page-11-19"></span>40. Smith, R.J. Use and misuse of the reduced major axis for line-fitting. *Am. J. Phys. Anthropol.* **2009**, *140*, 476–486. [\[CrossRef\]](https://doi.org/10.1002/ajpa.21090)
- <span id="page-11-20"></span>41. Brown, M.T.; Tinsley, H.E.A. Discriminant Analysis. *J. Leis. Res.* **2018**, *15*, 290–310. [\[CrossRef\]](https://doi.org/10.1080/00222216.1983.11969564)
- <span id="page-11-21"></span>42. Deutsch, A.R.; Dickinson, E.; Whichard, V.A.; Lagomarsino, G.R.; Perry, J.M.G.; Kupczik, K.; Hartstone-Rose, A. Primate body mass and dietary correlates of tooth root surface area. *Am. J. Biol. Anthropol.* **2022**, *177*, 4–26. [\[CrossRef\]](https://doi.org/10.1002/ajpa.24430) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/36787710)
- <span id="page-11-22"></span>43. Raaum, R.L.; Sterner, K.N.; Noviello, C.M.; Stewart, C.-B.; Disotell, T.R. Catarrhine Primate Divergence Dates Estimated from Complete Mitochondrial Genomes: Concordance with Fossil and Nuclear DNA Evidence. *J. Hum. Evol.* **2005**, *48*, 237–257. [\[CrossRef\]](https://doi.org/10.1016/j.jhevol.2004.11.007) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/15737392)
- <span id="page-11-23"></span>44. Brockelman, W.Y. Ecology and the Social System of Gibbons. In *The Gibbons*; Springer: New York, NY, USA, 2009; pp. 211–239.
- <span id="page-11-24"></span>45. Reichard, U.H. Social Monogamy in Gibbons: The Male Perspective. In *Monogamy*; Reichard, U.H., Boesch, C., Eds.; Cambridge University: Cambridge, UK, 2003; pp. 190–213.
- <span id="page-11-25"></span>46. Elton, S. Environmental Correlates of the Cercopithecoid Radiations. *Folia Primatol.* **2007**, *78*, 344–364. [\[CrossRef\]](https://doi.org/10.1159/000105149)
- <span id="page-12-0"></span>47. Begun, D.R. Catarrhine Origins and Evolution. In *A Companion to Biological Anthropology the Past and the Dead*; Larsen, C.S., Ed.; Wiley Online Books: Hoboken, NJ, USA, 2023; pp. 365–640.
- <span id="page-12-1"></span>48. Brandon-Jones, D.; Eudey, A.A.; Geissmann, T.; Groves, C.P.; Melnick, D.J.; Morales, J.C.; Shekelle, M.; Stewart, C.B. Asian Primate Classification. *Int. J. Primatol.* **2004**, *25*, 97–164. [\[CrossRef\]](https://doi.org/10.1023/B:IJOP.0000014647.18720.32)
- <span id="page-12-2"></span>49. Li, B.G.; He, G.; Guo, S.T.; Hou, R.; Huang, K.; Zhang, P.; Zhang, H.; Pan, R.L.; Chapman, C.A. Macaques in China: Evolutionary Dispersion and Subsequent Development. *Am. J. Primatol.* **2020**, *82*, e23142. [\[CrossRef\]](https://doi.org/10.1002/ajp.23142)
- <span id="page-12-3"></span>50. Maslin, M.A.; Christensen, B. Tectonics, Orbital Forcing, Global Climate Change, and Human Evolution in Africa: Introduction to the African Paleoclimate Special Volume. *J. Hum. Evol.* **2007**, *53*, 443–464. [\[CrossRef\]](https://doi.org/10.1016/j.jhevol.2007.06.005)
- <span id="page-12-4"></span>51. Levin, N.E.; Quade, J.; Simpson, S.W.; Semaw, S.; Rogers, M. Isotopic evidence for Plio–Pleistocene environmental change at Gona, Ethiopia. *Earth Planet. Sci. Lett.* **2004**, *219*, 93–110. [\[CrossRef\]](https://doi.org/10.1016/S0012-821X(03)00707-6)
- <span id="page-12-5"></span>52. deMenocal, P.B. Plio-Pleistocene African Climate. *Science* **1995**, *270*, 53–59. [\[CrossRef\]](https://doi.org/10.1126/science.270.5233.53) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/7569951)
- <span id="page-12-6"></span>53. Williamson, P.G. Evidence for an early Plio-Pleistocene rainforest expansion in East Africa. *Nature* **1985**, *315*, 487–489. [\[CrossRef\]](https://doi.org/10.1038/315487a0)
- <span id="page-12-7"></span>54. Lewis, L.A.; Berry, L. *African Environments and Resources*; Routledge: London, UK; New York, NY, USA, 2011.
- <span id="page-12-8"></span>55. Spicer, R.A.; Su, T.; Valdes, P.J.; Farnsworth, A.; Wu, F.X.; Shi, G.; Spicer, T.E.V.; Zhou, Z. Why 'the Uplift of the Tibetan Plateau' is A Myth. *Natl. Sci. Rev.* **2021**, *8*, nwaa091. [\[CrossRef\]](https://doi.org/10.1093/nsr/nwaa091) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/34691550)
- 56. Zhang, D.F.; Fengquan, L.; Jianmin, B. Eco-environmental Effects of the Qinghai-Tibet Plateau Uplift During the Quaternary in China. *Environ. Geol.* **2000**, *39*, 1352–1359. [\[CrossRef\]](https://doi.org/10.1007/s002540000174)
- <span id="page-12-9"></span>57. Li, J.J. The Environmental Effects of the Uplift of the Qinghai-Xizang Plateau. *Quat. Sci. Rev.* **1991**, *10*, 479–483.
- <span id="page-12-10"></span>58. Liu, X.D.; Dong, B.W. Influence of the Tibetan Plateau Uplift on the Asian Monsoon-arid Environment Evolution. *Chin. Sci. Bull.* **2013**, *58*, 4277–4291. [\[CrossRef\]](https://doi.org/10.1007/s11434-013-5987-8)
- 59. Shi, Z.G.; Sha, Y.Y.; Liu, X.D.; Xie, X.N.; Li, X.Z. Effect of Marginal Topography Around the Tibetan Plateau on the Evolution of Central Asian arid Climate: Yunnan–Guizhou and Mongolian Plateaux as Examples. *Clim. Dyn.* **2019**, *53*, 4433–4445. [\[CrossRef\]](https://doi.org/10.1007/s00382-019-04796-z)
- 60. Li, X.R.; Jia, X.H.; Dong, G.R. Influence of desertification on vegetation pattern variations in the cold semi-arid grasslands of Qinghai-Tibet Plateau, North-west China. *J. Arid. Environ.* **2006**, *64*, 505–522. [\[CrossRef\]](https://doi.org/10.1016/j.jaridenv.2005.06.011)
- <span id="page-12-11"></span>61. Favre, A.; Packert, M.; Pauls, S.U.; Jahnig, S.C.; Uhl, D.; Michalak, I.; Muellner-Riehl, A.N. The Role of the Uplift of the Qinghai-Tibetan Plateau for the Evolution of Tibetan Biotas. *Biol. Rev. Camb. Philos. Soc.* **2015**, *90*, 236–253. [\[CrossRef\]](https://doi.org/10.1111/brv.12107) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/24784793)
- <span id="page-12-12"></span>62. Szalay, F.S.; Delson, E. *Evolutionary History of the Primates*; Academic Press: New York, NY, USA, 1979.
- <span id="page-12-13"></span>63. Quade, J.; Cerling, T.E.; Bowman, J.R. Development of Asian monsoon revealed by marked ecological shift during the latest Miocene in northern Pakistan. *Nature* **1989**, *342*, 163–166. [\[CrossRef\]](https://doi.org/10.1038/342163a0)
- 64. Quade, J.; Cerling, T.E. Expansion of C4 grasses in the Late Miocene of Northern Pakistan: Evidence from stable isotopes in paleosols. *Palaeogeogr. Palaeoclimatol. Palaeoecol.* **1995**, *115*, 91–116. [\[CrossRef\]](https://doi.org/10.1016/0031-0182(94)00108-K)
- <span id="page-12-14"></span>65. Singh, S.; Awasthi, A.K.; Parkash, B.; Kumar, S. Tectonics or climate: What drove the Miocene global expansion of C4 grasslands? *Int. J. Earth Sci.* **2013**, *102*, 2019–2031. [\[CrossRef\]](https://doi.org/10.1007/s00531-013-0893-5)
- <span id="page-12-15"></span>66. Kotla, S.S.; Patnaik, R.; Sehgal, R.K.; Kharya, A. Isotopic evidence for ecological and climate change in the richly fossiliferous Plio-Pleistocene Upper Siwalik deposits exposed around Chandigarh, India. *J. Asian Earth Sci.* **2018**, *163*, 32–42. [\[CrossRef\]](https://doi.org/10.1016/j.jseaes.2018.05.019)
- <span id="page-12-16"></span>67. Corvinus, G. Homo erectus in East and Southeast Asia, and the questions of the age of the species and its association with stone artifacts, with special attention to handaxe-like tools. *Quat. Int.* **2004**, *117*, 141–151. [\[CrossRef\]](https://doi.org/10.1016/S1040-6182(03)00124-1)
- <span id="page-12-17"></span>68. Zhao, C.Z.; Li, X.Q.; Zhou, X.Y.; Zhao, K.L.; Yang, Q. Holocene Vegetation Succession and Response to Climate Change on the South Bank of the Heilongjiang-Amur River, Mohe County, Northeast China. *Adv. Meteorol.* **2016**, *2016*, 2450697. [\[CrossRef\]](https://doi.org/10.1155/2016/2450697)
- <span id="page-12-18"></span>69. Li, Z.-W.; Sun, L.; Li, B.-S.; Wang, F.-N.; Du, D.-D.; Song, Y.-G.; Zhang, H.-J.; Chen, L.-Q.; Xu, D. East Asian summer monsoon changes in subtropical China since late Pleistocene: Evidence from the Ailuropoda-Stegodon fauna. *J. Mt. Sci.* **2022**, *19*, 418–432. [\[CrossRef\]](https://doi.org/10.1007/s11629-021-6985-y)
- <span id="page-12-19"></span>70. Louys, J.; Turner, A. Environment, preferred habitats and potential refugia for Pleistocene Homo in Southeast Asia. *Comptes Rendus Palevol* **2012**, *11*, 203–211. [\[CrossRef\]](https://doi.org/10.1016/j.crpv.2011.03.003)
- <span id="page-12-20"></span>71. Bird, M.I.; Taylor, D.; Hunt, C. Palaeoenvironments of insular Southeast Asia during the Last Glacial Period: A savanna corridor in Sundaland? *Quat. Sci. Rev.* **2005**, *24*, 2228–2242. [\[CrossRef\]](https://doi.org/10.1016/j.quascirev.2005.04.004)
- <span id="page-12-21"></span>72. Wurster, C.M.; Bird, M.I.; Bull, I.D.; Creed, F.; Bryant, C.; Dungait, J.A.; Paz, V. Forest Contraction in North Equatorial Southeast Asia During the Last Glacial Period. *Proc. Natl. Acad. Sci. USA* **2010**, *107*, 15508–15511. [\[CrossRef\]](https://doi.org/10.1073/pnas.1005507107) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/20660748)
- <span id="page-12-22"></span>73. Takai, M.; Soe, A.N.; Maung, M.; Tsubamoto, T.; Egi, N.; Nishimura, T.D.; Nishioka, Y. First Discovery of Colobine Fossils from the Late Miocene/Early Pliocene in Central Myanmar. *J. Hum. Evol.* **2015**, *84*, 1–15. [\[CrossRef\]](https://doi.org/10.1016/j.jhevol.2015.04.003) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/25978976)
- <span id="page-12-23"></span>74. Chang, C.-H.; Takai, M.; Ogino, S. First Discovery of Colobine Fossils from the Early to Middle Pleistocene of Southern Taiwan. *J. Hum. Evol.* **2012**, *63*, 439–451. [\[CrossRef\]](https://doi.org/10.1016/j.jhevol.2012.03.005) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/22749256)
- 75. Takai, M.; Nishioka, Y.; Thaung-Htike; Maung, M.; Khaing, K.; Zin-Maung-Maung-Thein; Tsubamoto, T.; Egi, N. Late Pliocene *Semnopithecus* Fossils from Central Myanmar: Rethinking of the Evolutionary History of Cercopithecid Monkeys in Southeast Asia. *Hist. Biol.* **2016**, *28*, 172–188. [\[CrossRef\]](https://doi.org/10.1080/08912963.2015.1018018)
- 76. Ogino, S.; Egi, N.; Takai, M. New Species of Agriotherium (Mammalia, Carnivora) from the Late Miocene to Early Pliocene of Central Myanmar. *J. Asian Earth Sci.* **2011**, *42*, 408–414. [\[CrossRef\]](https://doi.org/10.1016/j.jseaes.2011.05.017)
- 77. Lopatin, A.; Maschenko, E.; Vislobokova, I.; Serdyuk, N.; Dac, L.X. Pleistocene Mammals from the Lang Trang Cave (Vietnam): New Data. In *Doklady Biological Sciences*; Springer: Berlin/Heidelberg, Germany, 2021.
- 78. Ito, T.; Lee, Y.-J.; Nishimura, T.D.; Tanaka, M.; Woo, J.-Y.; Takai, M. Phylogenetic Relationship of a Fossil Macaque (*Macaca* cf. *robusta*) from the Korean Peninsula to Extant Species of Macaques Based on Zygomaxillary Morphology. *J. Hum. Evol.* **2018**, *119*, 1–13. [\[CrossRef\]](https://doi.org/10.1016/j.jhevol.2018.02.002)
- 79. Han, K.S.; So, K.S.; Choe, R.S.; Kang, J.C.; Kang, I.; Ri, C.U. First Record of *Macaca mulatta* (Cercopithecidae: Papionini) from the Taedong River Basin, the Democratic People's Republic of Korea. *Palaeoworld* **2022**, *32*, 573–578. [\[CrossRef\]](https://doi.org/10.1016/j.palwor.2022.09.008)
- 80. Ingicco, T.; Amano, N.; Ochoa, J.; Détroit, F. An allometric study of *Macaca fascicularis* from the Late Pleistocene deposits at the Ille site (Philippines): A possible model for Southeast Asian Dwarf Hominins. *Bull. Mémoires Société Dianthropologie Paris* **2014**. [\[CrossRef\]](https://doi.org/10.1007/s13219-014-0098-x)
- 81. Jouffroy, F. Un Crane Subfossile De Macaque Du Pléistocène Du Viet Nam. *Bull. Mus Nat D'hist Nat. Paris* **1959**, *31*, 309–316.
- 82. Schillaci, M.A.; Klegarth, A.R.; Switzer, W.M.; Shattuck, M.R.; Lee, B.P.; Hollocher, H. Evolutionary Relationships of *Macaca fascicularis fascicularis* (Raffles 1821) (Primates: Cercopithecidae) from Singapore Revealed by Bayesian Analysis of Mitochondrial DNA Sequences. *Raffles Bull. Zool.* **2017**, *65*, 3–19.
- 83. Ogino, S.; Otsuka, H.; Harunari, H. The Middle Pleistocene Matsugae Fauna, Northern Kyushu, West Japan. *Paleontol. Res.* **2009**, *13*, 367–384. [\[CrossRef\]](https://doi.org/10.2517/1342-8144-13.4.367)
- 84. Filoux, A.; Wattanapituksakul, A.; Lespes, C.; Thongcharoenchaikit, C. A Pleistocene Mammal Assemblage Containing Ailuropoda and Pongo from Tham Prakai Phet Cave, Chaiyaphum Province, Thailand. *Geobios* **2015**, *48*, 341–349. [\[CrossRef\]](https://doi.org/10.1016/j.geobios.2015.07.003)
- 85. Bacon, A.-M.; Demeter, F.; Roussé, S.; Long, V.T.; Duringer, P.; Antoine, P.-O.; Thuy, N.K.; Mai, B.T.; Huong, N.T.M.; Dodo, Y.; et al. New Palaeontological Assemblage, Sedimentological and Chronological Data from the Pleistocene Ma U'Oi Cave (Northern Vietnam). *Palaeogeogr. Palaeoclimatol. Palaeoecol.* **2006**, *230*, 280–298. [\[CrossRef\]](https://doi.org/10.1016/j.palaeo.2005.07.023)
- 86. Zeitoun, V.; Lenoble, A.; Laudet, F.; Thompson, J.; Rink, W.J.; Mallye, J.-B.; Chinnawut, W. The Cave of the Monk (Ban Fa Suai, Chiang Dao Wildlife Sanctuary, Northern Thailand). *Quat. Int.* **2010**, *220*, 160–173. [\[CrossRef\]](https://doi.org/10.1016/j.quaint.2009.11.022)
- 87. Pushkina, D.; Bocherens, H.; Chaimanee, Y.; Jaeger, J.-J. Stable Carbon Isotope Reconstructions of Diet and Paleoenvironment from the Late Middle Pleistocene Snake Cave in Northeastern Thailand. *Naturwissenschaften* **2010**, *97*, 299–309. [\[CrossRef\]](https://doi.org/10.1007/s00114-009-0642-6)
- 88. Guangxi-Museum. Museum. Museum Proceedings. In *Guangxi Ethnic Museum*; Guangxi-Museum: Guangxi, China, 2012. (In Chinese)
- 89. Ito, T.; Nishimura, T.D.; Ebbestad, J.O.R.; Takai, M. Computed Tomography Examination of the Face of *Macaca anderssoni* (Early Pleistocene, Henan, northern China): Implications for the Biogeographic History of Asian Macaques. *J. Hum. Evol.* **2014**, *72*, 64–80. [\[CrossRef\]](https://doi.org/10.1016/j.jhevol.2014.04.001)
- 90. Jablonski, N.G. Fossil Old World Monkeys: The Late Neogene Radiation. In *Primate Fossil Record*; Hartwig, W.C., Ed.; Cambridge University: Cambridge, UK, 2008; pp. 255–299.
- 91. Rasmussen, D.T. Early Catarrhines of the African Eocene and Oligocene. In *Primate Fossil Record*; Hartwig, W.C., Ed.; Cambridge University: Cambridge, UK, 2002; pp. 255–299.
- 92. Ji, X.P.; Youlatos, D.; Jablonski, N.G.; Pan, R.; Zhang, C.; Li, P.; Tang, M.; Yu, T.; Li, W.; Deng, C. Oldest Colobine Calcaneus from East Asia (Zhaotong, Yunnan, China). *J. Hum. Evol.* **2020**, *147*, 102866. [\[CrossRef\]](https://doi.org/10.1016/j.jhevol.2020.102866) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/32862123)
- 93. Ji, X.P.; Jablonski, N.G.; Su, D.F.; Deng, C.L.; Flynn, L.J.; You, Y.S.; Kelley, J. Juvenile Hominoid Cranium from the Terminal Miocene of Yunnan, China. *Chin. Sci. Bull.* **2013**, *58*, 3771–3779. [\[CrossRef\]](https://doi.org/10.1007/s11434-013-6021-x)
- 94. Lucas, S.G. *Chinese Fossil Vertebrates*; Columbia University Press: New York, NY, USA, 2001; p. 320.
- 95. Ortiz, A.; Pilbrow, V.; Villamil, C.I.; Korsgaard, J.G.; Bailey, S.E.; Harrison, T. The Taxonomic and Phylogenetic Affinities of Bunopithecus sericus, a Fossil Hylobatid from the Pleistocene of China. *PLoS ONE* **2015**, *10*, e0131206. [\[CrossRef\]](https://doi.org/10.1371/journal.pone.0131206)
- 96. Pan, Y.; Peng, Y.; Zhang, X.; Pan, R. Cercopithecid Fossils Discovered in Yunnan and its Stratigraphical Significance. *Acta Anthropol. Sin.* **1992**, *11*, 303.
- 97. Takai, M.; Maschenko, E.N. Parapresbytis Eohanuman: The Northernmost Colobine Monkey from the Pliocene of Transbaikalia. *Asian Paleoprimatology* **2009**, *5*, 1–14.
- 98. Takai, M.; Zhang, Y.Q.; Kono, R.T.; Jin, C.Z. Changes in the Composition of the Pleistocene Primate Fauna in Southern China. *Quat. Int.* **2014**, *354*, 75–85. [\[CrossRef\]](https://doi.org/10.1016/j.quaint.2014.02.021)
- 99. Wu, X.Z. Fossil Humankind and other Anthropoid Primates of China. *Int. J. Primatol.* **2004**, *25*, 1093–1103. [\[CrossRef\]](https://doi.org/10.1023/B:IJOP.0000043353.24043.19)
- <span id="page-13-0"></span>100. Fa, J.E. The Genus Macaca: A Review of Taxonomy and Evolution. *Mammal Rev.* **1989**, *19*, 45–81. [\[CrossRef\]](https://doi.org/10.1111/j.1365-2907.1989.tb00401.x)
- <span id="page-13-1"></span>101. Selig, K.R.; Silcox, M.T. Measuring Molarization: Change through Time in Premolar Function in an Extinct Stem Primate Lineage. *J. Mamm. Evol.* **2022**, *29*, 947–956. [\[CrossRef\]](https://doi.org/10.1007/s10914-022-09623-7)
- <span id="page-13-2"></span>102. Scott, J.E.; Campbell, R.M.; Baj, L.M.; Burns, M.C.; Price, M.S.; Sykes, J.D.; Vinyard, C.J. Dietary Signals in the Premolar Dentition of Primates. *J. Hum. Evol.* **2018**, *121*, 221–234. [\[CrossRef\]](https://doi.org/10.1016/j.jhevol.2018.04.006) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/29793792)
- <span id="page-13-3"></span>103. Hylander, W.L. The Functional Significance of Primate Mandibular Form. *J. Morphol.* **1979**, *160*, 223–240. [\[CrossRef\]](https://doi.org/10.1002/jmor.1051600208) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/458862)
- <span id="page-13-4"></span>104. Lucas, P.W. *Dental Functional Morphology: How Teeth Work*; Cambridge University Press: Cambridge, UK, 2004.
- <span id="page-13-5"></span>105. Napier, J.R.; Napier, P.H. *A Handbook of Living Primates*; Academic Press: London, UK, 1967.
- 106. Smith, R.J. The Mandibular Corpus of Female Primates: Taxonomic, Dietary, and Allometric Correlates of Interspecific Variations in Size and Shape. *Am. J. Phys. Anthropol.* **1983**, *61*, 315–330. [\[CrossRef\]](https://doi.org/10.1002/ajpa.1330610306)
- <span id="page-13-6"></span>107. Bunn, J.M.; Ungar, P.S. Dental topography and diets of four old world monkey species. *Am. J. Primatol.* **2009**, *71*, 466–477. [\[CrossRef\]](https://doi.org/10.1002/ajp.20676)
- <span id="page-14-0"></span>108. Chapman, C.A.; Lambert, J.E. Habitat alteration and the conservation of African primates: Case study of Kibale National Park, Uganda. *Am. J. Primatol.* **2000**, *50*, 169–185. [\[CrossRef\]](https://doi.org/10.1002/(SICI)1098-2345(200003)50:3%3C169::AID-AJP1%3E3.0.CO;2-P)
- <span id="page-14-1"></span>109. Albert, A.; McConkey, K.; Savini, T.; Huynen, M.-C. The value of disturbance-tolerant cercopithecine monkeys as seed dispersers in degraded habitats. *Biol. Conserv.* **2014**, *170*, 300–310. [\[CrossRef\]](https://doi.org/10.1016/j.biocon.2013.12.016)
- <span id="page-14-2"></span>110. Happel, R. Seed-eating by West African cercopithecines, with reference to the possible evolution of bilophodont molars. *Am. J. Phys. Anthropol.* **1988**, *75*, 303–327. [\[CrossRef\]](https://doi.org/10.1002/ajpa.1330750303) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/3364545)
- <span id="page-14-3"></span>111. Wrangham, R.W.; Conklin-Brittain, N.; Hunt, K.D. Dietary Response of Chimpanzees and Cercopithecines to Seasonal Variation in Fruit Abundance. I. Antifeedants. *Int. J. Primatol.* **1998**, *19*, 949–970. [\[CrossRef\]](https://doi.org/10.1023/A:1020318102257)
- <span id="page-14-4"></span>112. Martin, F.; Plastiras, C.-A.; Merceron, G.; Souron, A.; Boisserie, J.-R. Dietary niches of terrestrial cercopithecines from the Plio-Pleistocene Shungura Formation, Ethiopia: Evidence from Dental Microwear Texture Analysis. *Sci. Rep.* **2018**, *8*, 14052. [\[CrossRef\]](https://doi.org/10.1038/s41598-018-32092-z)
- <span id="page-14-5"></span>113. Koyabu, D.B.; Endo, H. Craniofacial Variation and Dietary Adaptations of African Colobines. *J. Hum. Evol.* **2009**, *56*, 525–536. [\[CrossRef\]](https://doi.org/10.1016/j.jhevol.2008.12.009)
- 114. Koyabu, D.B.; Endo, H. Craniodental Mechanics and Diet in Asian Colobines: Morphological Evidence of Mature Seed Predation and Sclerocarpy. *Am. J. Phys. Anthropol.* **2010**, *142*, 137–148. [\[CrossRef\]](https://doi.org/10.1002/ajpa.21213)
- <span id="page-14-6"></span>115. Guatelli-Steinberg, D.; Schwartz, G.T.; O'Hara, M.C.; Gurian, K.; Rychel, J.; Dunham, N.; Cunneyworth, P.M.K.; Donaldson, A.; McGraw, W.S. Aspects of Molar form and Dietary Proclivities of African Colobines. *J. Hum. Evol.* **2023**, *180*, 103384. [\[CrossRef\]](https://doi.org/10.1016/j.jhevol.2023.103384)
- <span id="page-14-7"></span>116. Bouvier, M. A biomechanical analysis of mandibular scaling in old world monkeys. *Am. J. Phys. Anthropol.* **1986**, *69*, 473–482. [\[CrossRef\]](https://doi.org/10.1002/ajpa.1330690406)
- 117. Ravosa, M.J. Functional Assessment of Subfamily Variation in Maxillomandibular Morphology Among Old World Monkeys. *Am. J. Phys. Anthropol.* **1990**, *82*, 199–212. [\[CrossRef\]](https://doi.org/10.1002/ajpa.1330820209)
- <span id="page-14-8"></span>118. Davies, A.G.; Oates, J.F. *Colobine Monkeys: Their Ecology, Behaviour, and Evolution*; Cambridge University Press: Cambridge, UK, 1994.
- <span id="page-14-9"></span>119. Leutenegger, W.; Kelly, J.T. Relationship of Sexual Dimorphism in Canine Size and Body Size to Social, Behavioral, and Ecological Correlates in Anthropoid Primates. *Primates* **1977**, *18*, 117–136. [\[CrossRef\]](https://doi.org/10.1007/BF02382954)
- <span id="page-14-10"></span>120. Petersdorf, M.; Weyher, A.H.; Kamilar, J.M.; Dubuc, C.; Higham, J.P. Sexual Selection in the Kinda Baboon. *J. Hum. Evol.* **2019**, *135*, 102635. [\[CrossRef\]](https://doi.org/10.1016/j.jhevol.2019.06.006) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/31421317)
- <span id="page-14-11"></span>121. Dirks, W.; Lemmers, S.A.M.; Ngoubangoye, B.; Herbert, A.; Setchell, J.M. Odontochronologies in male and female mandrills (Mandrillus sphinx) and the development of dental sexual dimorphism. *Am. J. Phys. Anthropol.* **2020**, *172*, 528–544. [\[CrossRef\]](https://doi.org/10.1002/ajpa.24094) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/32510604)
- 122. Harvey, P.H.; Kavanagh, M.; Clutton-Brock, T.H. Sexual dimorphism in primate teeth. *J. Zool.* **2009**, *186*, 475–485. [\[CrossRef\]](https://doi.org/10.1111/j.1469-7998.1978.tb03934.x)
- <span id="page-14-12"></span>123. Plavcan, J.M. Sexual dimorphism in primate evolution. *Am. J. Phys. Anthropol.* **2001**, *116* (Suppl. S33), 25–53. [\[CrossRef\]](https://doi.org/10.1002/ajpa.10011) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/11786990)

**Disclaimer/Publisher's Note:** The statements, opinions and data contained in all publications are solely those of the individual author(s) and contributor(s) and not of MDPI and/or the editor(s). MDPI and/or the editor(s) disclaim responsibility for any injury to people or property resulting from any ideas, methods, instructions or products referred to in the content.