



Supplementary Material

Biosensors Platform Based on Chitosan/AuNPs/Phthalocyanine Composite Films for the Electrochemical Detection of Catechol. The Role of the Surface Structure

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Figure S1. Materials used to form the sensing platforms: a) gold nanoparticles (AuNPs) capped with citrate; b) sulfonated copper phthalocyanine (CuPcS); and c) chitosan (CHI).

The AuNPs' size was recalculated following the Mie method, using the following equation:

$$d = \frac{ln(\frac{\lambda_{spr} - \lambda_0}{L1})}{L2}$$

where λ_{spr} is the surface plasmon resonance peak position. and λ_0 =512 nm. L₁=6.53 nm and L₂=0.0216 nm-1, are theoretical parameters when d is known to be d>25 nm. The λ_{spr} value of 533 nm was determined from the absorbance spectrum shown below. The estimated diameter of the obtained AuNPs is 54.nm.



Figure S2. UV-Vis spectrum of AuNP solution.



Figure S3. UV-Vis spectra of a) [(CHI)-(AuNPs)-(CHI)-(CuPcS)]² LbL films with increasing number of layers (n = 4,8,12,16,20); and b) correlation between the absorbance measured at 615 nm and the number of layers.



Figure S4. Comparison of the FTIR spectra of the LbL platform [(CHI)-(AuNPs)-(CHI)-(CuPcS)]² (solid line) and of the biosensor [(CHI)-(AuNPs)-(CHI)-(CuPcS)]²-Lac (dotted line).



Figure S5. a) FTIR spectra of the platform [(CHI)-(AuNPs)-(CHI)-(CuPcS)]² with increasing number of layers (n = 4,8,12,16,20,24) and b) correlation between the transmittance measured at 1033 cm⁻¹ and the number of layers.



Figure S6. CV curves registered in catechol 10⁻⁴ mol·L⁻¹ in 0.01 M phosphate buffer pH 7 at a bare ITO electrode.



Figure S7. CV registered in catechol 10-4 mol·L-1 in 0.01 M phosphate buffer pH 7 at [(CHI)-(AuNPs)-(CHI)-(CuPcS)]2-Lac electrode, where the enzyme was cross-linked by immersion in glutaraldehyde liquid (dotted line) and exposure to glutaraldehyde vapors (solid line).



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