


## Article

# A Novel Fluorescent Sensor Based on Aptamer and qPCR for Determination of Glyphosate in Tap Water

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**Abstract:** Glyphosate (GLYP) is a broad-spectrum, nonselective, organic phosphine postemergence herbicide registered for many food and nonfood fields. Herein, we developed a biosensor (Mbs@dsDNA) based on carboxylated modified magnetic beads incubated with NH<sub>2</sub>-polyA and then hybridized with polyT-glyphosate aptamer and complementary DNA. Afterwards, a quantitative detection method based on qPCR was established. When the glyphosate aptamer on Mbs@dsDNA specifically recognizes glyphosate, complementary DNA is released and then enters the qPCR signal amplification process. The linear range of the method was 0.6 μmol/L–30 mmol/L and the detection limit was set at 0.6 μmol/L. The recoveries in tap water ranged from 103.4 to 104.9% and the relative standard deviations (RSDs) were <1%. The aptamer proposed in this study has good potential for recognizing glyphosate. The detection method combined with qPCR might have good application prospects in detecting and supervising other pesticide residues.

**Keywords:** glyphosate; aptamer; qPCR; SYBR Green I; sensor



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## 1. Introduction

Glyphosate (GLYP), with the chemical name *N*-(phosphoryl-methyl) glycine, is a nonselective, broad-spectrum organic phosphine herbicide used in agriculture to eliminate various annual and perennial weeds [1–4] around the globe. Following application, it is evenly distributed and rapidly translocated to regions of active growth within the plant, causing death. However, the high herbicidal activity led to the abuse of glyphosate [5,6]. Many studies have proven that glyphosate has toxic effects and potentially harms the ecological environment and human health [7–9]. Furthermore, residues have been found in soil, even groundwater, due to extensive usage. Moreover, recent studies have shown that glyphosate accumulation in the environment may cause a certain degree of reproductive toxicity, teratogenicity, mutagenicity and carcinogenicity in humans [10,11]. To reduce the impact of glyphosate residues on food safety and human health, detecting glyphosate residual levels is crucial and essential.

As concerns and studies on the behavior of glyphosate in plants and the environment are growing, several methodologies, including high-performance liquid chromatography (HPLC) [12,13], gas chromatography-mass spectrometry (GC-MS) [14–16], LC-tandem

mass spectrometry (LC-MS/MS) [17,18], liquid chromatography (LC) [19,20], ion chromatography (IC) [21,22], capillary electrophoresis (CE) [23,24], enzyme-linked immunoassays (ELISA) [25,26], fluorescence detection [27,28] and electrochemical luminescence methods [29,30], have been developed to detect its residues. Although the sensitivity and specificity of these methods are relatively high, their shortcomings are also apparent. They usually require expensive instruments, professional operators, time-consuming sample pretreatment and high testing costs, with certain limitations for glyphosate detection. Therefore, developing a rapid, simple, inexpensive, high-sensitivity and high-specificity sensor is crucial for quantitatively detecting pollutants.

Aptamers are nucleic acid molecules synthesized *in vitro* by a process known as Systematic Evolution of Ligands Exponential Enrichment (SELEX), which has unique binding properties to different targets, such as proteins, small molecule metal ions and whole cells [31–34]. As a new type of molecular probe, it can be synthesized with high purity *in vitro*. The cost is lower, which is a point of superiority compared with antibodies. After the aptamer sequence is determined, it can be synthesized with high reproducibility and high purity from commercial sources at a lower cost, which is an advantage compared with antibodies. Antibodies generally take 3–6 months to prepare and synthesize, while aptamers only take a few hours to several days to synthesize. The aptamer has high thermal stability, low immunogenicity, and chemical stability. In addition, the preparation process of antibodies is usually very long, while the synthesis time of aptamers is usually shorter. At the same time, it can meet the experimental design requirements of modification by chemical labels, such as nanoparticles, fluorescent groups and functional groups. It will not affect the affinity between them and the target molecules [35–37]. They are generally more stable under harsh conditions than antibodies [38–40]. After thermal denaturation, the aptamers usually return to their native state. Furthermore, they can be used as effective recognition elements in biosensors and are widely used in the detection field for biosensing. Consequently, some results have been achieved in aptamer-based pesticide detection technology. For instance, what has gained more application in sensing is generally color-based detection, which has more directly detectable results [41]. Aptamer sensors have been reported to detect fipronil, non-nitrothion and diazinon in fruit and vegetable samples. These aptamer sensors have the advantage of being highly sensitive and specific [42–44].

SYBR Green real-time qPCR can monitor the amplification process in real time by detecting the fluorescent signal emitted by a dsDNA-specific dye (SYBR Green I). In real-time fluorescent quantitative PCR amplification, the fluorescent group can monitor the amplification products in each cycle of PCR in real time and the operation is convenient and straightforward. DNA amplification and data analysis can be carried out in a closed system under the same conditions so that samples and products cannot be polluted. Electrophoresis confirmation of standard PCR products can be directly omitted. To the best of our knowledge, there are few studies on the detection of glyphosate based on the combination of aptamers and qPCR. Chen et al. developed a time-resolved luminescence assay for glyphosate based on G-quadruplexes but not in combination with qPCR. Therefore, the aptamer sequence from this article was used. Herein, aptamer-SYBR Green I-based sensing technology was established to rapidly detect glyphosate by qPCR using the single-stranded DNA released from the aptamer after recognizing glyphosate. The recognition process of the aptamer for glyphosate was converted to the qPCR cycle threshold (CT) value.

## 2. Experiment

### 2.1. Materials and Instruments

All DNA sequences were synthesized and purified by Sangon Biotech (Shanghai) Co., Ltd. (Shanghai, China) The sequences of the DNA strands is shown in the supporting information (Table S1). Carboxyl magnetic beads were procured from Sangon Biotech (Shanghai) Co., Ltd. Power Up™ SYBR™ green premix was acquired from Thermo Fisher Scientific (Waltham, MA, USA). Glyphosate, acetamiprid, chlorpyrifos, dimethoate, trichlorfon, methomyl and propoxur (purity greater than 98%) standards were provided by Dr

Ehrenstorfer GmbH (Augsburg, Germany). Other chemical reagents were purchased from Sinopharm Group Co., Ltd. (Beijing, China). All purchased chemicals were used as received without further purification. The water involved in the experiments was purified by Milli-Q (Merck & Co., Rahway, NJ, USA) with a resistivity of 18.2 MΩ cm).

UV-2600 (SHIMADZU (CHINA) Co., Ltd. (Beijing, China)); TECAN Infinite 200 PRO Multifunctional Enzyme Reader (Männedorf, Switzerland); 7500 real-time fluorescence quantitative PCR instrument, Thermo Fisher Scientific (Waltham, MA, USA); and HM100-Pro, Dragon Laboratory Instruments Limited (Beijing, China) were used in this study.

Acetamiprid, chlorpyrifos, dimethoate, trichlorfon, methomyl and propoxur were separately prepared (4.5 mol/L, 0.29 mol/L, 4.4 mol/L, 3.9 mol/L, 0.62 mol/L and 4.8 mol/L) with methanol, diluted to 0.045 mol/L, 0.00029 mol/L, 0.044 mol/L, 0.039 mol/L, 0.00062 mol/L and 0.048 mol/L with deionized water. Glyphosate was prepared at 0.006 mol/L with deionized water and finally diluted to 0.03 mol/L, 0.012, 0.006, 0.003, 0.0012 and 0.0006 mol/L with a deionized water gradient.

## 2.2. Magnetic Bead Activation

The procedure used to activate magnetic beads was performed according to the manufacturer's protocols (Sangon Biotech Co., Ltd., Shanghai, China). After the carboxyl magnetic beads were whirled for 1 min, 100 µL was placed into a 2 mL centrifuge tube. Next, the supernatant was removed after magnetic separation and then 100 µL 25 mM MES solution was added and whirled for 15 s. After magnetic separation, the supernatant was removed and washed three times with deionized water. After that, 500 µL (10 mg/mL) 1-ethyl-3-(3'-dimethylaminopropyl) carbodiimide (EDC) and 100 µL (10 mg/mL) *N*-hydroxysuccinimide (NHS) were quickly introduced into a centrifuge tube. After mixing, the mixture was shaken slowly at room temperature for 30 min and then stored at 4 °C.

## 2.3. Preparation of Mbs@dsDNA and Pre-Experiment

Then, 600 µL of activated magnetic beads was taken, the supernatant was discarded after magnetic response, 600 µL of 10 µM polyA solution was added, thoroughly mixed and incubated in a 37 °C water bath overnight. The following day, the supernatant was removed after magnetic separation, redissolved in 600 µL deionized water and stored at 4 °C.

To obtain purified Mbs@polyA after magnetic separation, that polyA not combined with the magnetic bead is removed from the supernatant. 600 µL of 2 µmol polyT-aptamer solution was denatured at 90 °C for 2 min, then slowly dropped to room temperature and added to Mbs@polyA. After vortex oscillation in solution for 2 min, it is slowly shaken at room temperature for at least 30 min. Unhybridized polyT aptamer in the supernatant was removed after magnetic separation. Then, 600 µL of 2 µmol complementary DNA (C-DNA) solution was denatured at 90 °C for 2 min, slowly dropped to room temperature and added to the above solution. After 2 min of vortexing, the mixture was shaken slowly at room temperature for at least 30 min. Then, magnetic separation was performed to remove the nonhybridized C-DNA from the supernatant. After washing with deionized water three times, Mbs@dsDNA was suspended in deionized water at 4 °C for standby.

Twenty microliters of 0.6 µmol/L–60 mmol/L glyphosate and blank samples were added to the prepared Mbs@dsDNA system and incubated at room temperature for 3 h. After magnetic separation, the supernatant was taken for qPCR and photographed under a UV lamp.

## 2.4. Specificity Detection

A series of centrifugation tubes containing 60 µL of Mbs@dsDNA solution was prepared. After magnetic separation, the supernatant was removed and 60 µL of different solutions, including 0.045 mol/L acetamiprid, 0.00029 mol/L chlorpyrifos, 0.044 mol/L dimethoate, 0.039 mol/L trichlorfon, 0.00062 mol/L methomyl, 0.048 mol/L propoxur and 0.006 mol/L glyphosate, were added to each tube and incubated at room temperature

for 3 h. After incubation, the supernatant was separated by magnetic separation and the released C-DNA in the system was identified by UV spectrophotometry and then detected by qPCR. qPCR was carried out in a total volume of 20  $\mu\text{L}$  in the quantitative study. The conditions of amplification reactions are shown in the supporting information (Table S2).

### 2.5. Determination of Glyphosate

A series of centrifugation tubes with 60  $\mu\text{L}$  of Mbs@dsDNA in each tube were prepared. After magnetic separation, the supernatant was removed and 60  $\mu\text{L}$  glyphosate at different concentrations of 0.03, 0.012, 0.006, 0.003, 0.0012 and 0.0006 mol/L was added into each tube.

After 3 h of incubation, the supernatant was separated by magnetic separation and detected by UV spectrophotometry and qPCR.

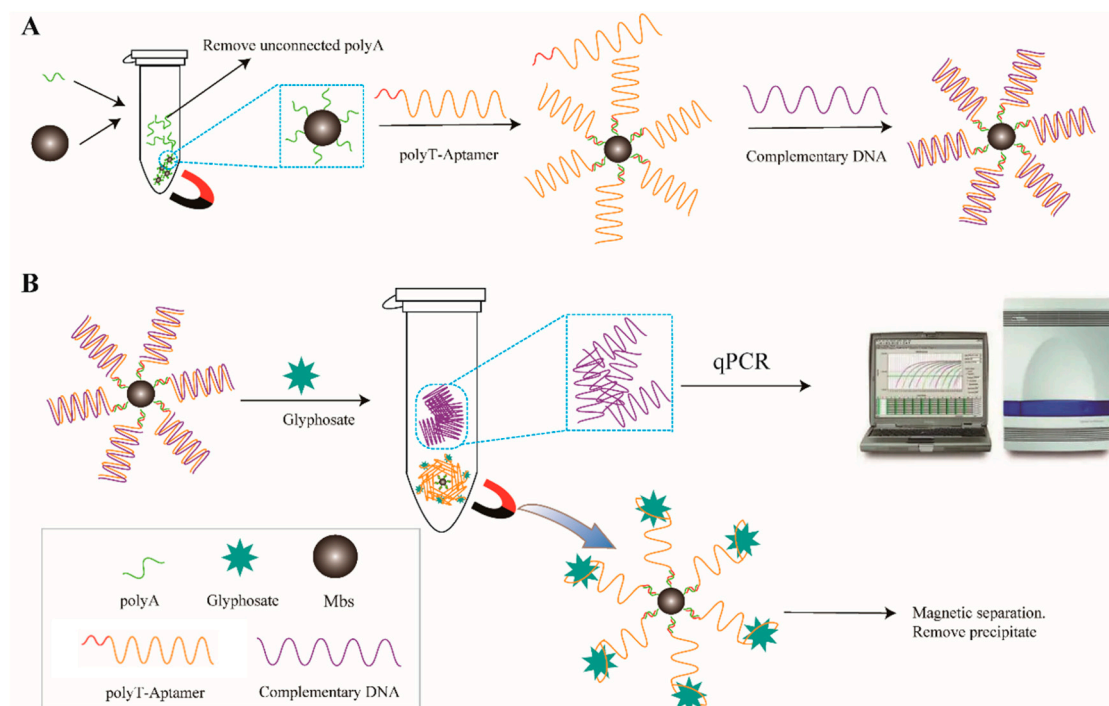
### 2.6. Actual Sample Testing

Tap water from the city of Beijing was taken as the actual sample and glyphosate at concentrations of 8  $\mu\text{mol/L}$ , 4  $\mu\text{mol/L}$  and 2  $\mu\text{mol/L}$  was added according to the national standard of China (GB/T 5749-2006). Each sample was tested 3 times. The relative standard deviation (RSD) of the test results was used to evaluate the method's precision.

## 3. Results and Discussion

### 3.1. Experimental Principle

Figure 1 shows a schematic diagram of the glyphosate detection method based on qPCR signal amplification proposed in this paper. As shown in Figure 1A, polyA is formed after incubation and connection with magnetic beads Mbs@polyA. Subsequently, the aptamer was hybridized with the polyT-modified aptamer (polyT aptamer) according to the principle of complementary base pairing in DNA molecules. The aptamer was assembled on the magnetic bead.



**Figure 1.** (A) Forming dsDNA compound. (B) Detection of glyphosate by qPCR. Aptamer sequence: GCTAGACGATATTCGTCCATCCGAGCCCGTGGCGGGCTTTAG GACTCTGCGGGCTTCGGCGCGCTGTCAGACTGAATATGTC.

Afterwards, the DNA complementary to the aptamer (C-DNA) is assembled on the magnetic bead through hybridization with Mbs@dsDNA. Through magnetic separation, redundant poly T-aptamers and C-DNA without double strands can be eliminated. As Figure 1B shows, when glyphosate is present in the system, the aptamer can specifically recognize glyphosate and form Mbs@Aptamer@GLYP. The binding force between the aptamer and glyphosate is stronger than that between the aptamer and C-DNA. With the help of an external magnetic field, the C-DNA and fMbs@Aptamer@GLYP were separated. The supernatant containing C-DNA was extracted and utilized in qPCR as a template. SYBR Green I was used to record the fluorescence signal obtained by qPCR and the linear equation was established by the logarithm of the CT value and concentration. Then, the content of glyphosate was calculated and the amount of glyphosate was calculated. When other interfering pesticides and blanks are added to the system, C-DNA will not be separated from the Mbs@dsDNA and the signal will not appear in qPCR.

### 3.2. Characterization of Mbs@dsDNA and Aptamer Sensor Feasibility Validation

The prepared Mbs@dsDNA in each step was characterized by ultraviolet and visible light and glyphosate was detected and characterized by fluorescence photography. When the peak value had a significant difference at 260 nm, the Mbs@dsDNA preparation was successful. When glyphosate is detected, the dye SYBR Green I can only be embedded in the double helix structure of DNA, so the fluorescence signal will be sent out when glyphosate is present in the system. As shown in Figure 2D, SYBR I can be embedded in the double helix structure of DNA and emit fluorescence. When there is no glyphosate in the system, SYBR Green I cannot embed into the double helix structure of DNA and will not emit a fluorescence signal because qPCR cannot be successfully carried out to obtain double-stranded DNA. In contrast, when glyphosate is present in the system, the aptamer specifically binds to it and releases C-DNA into the qPCR to successfully amplify double-stranded DNA. SYBR I is embedded in the DNA double helix structure and emits a fluorescence signal under the excitation of ultraviolet light.

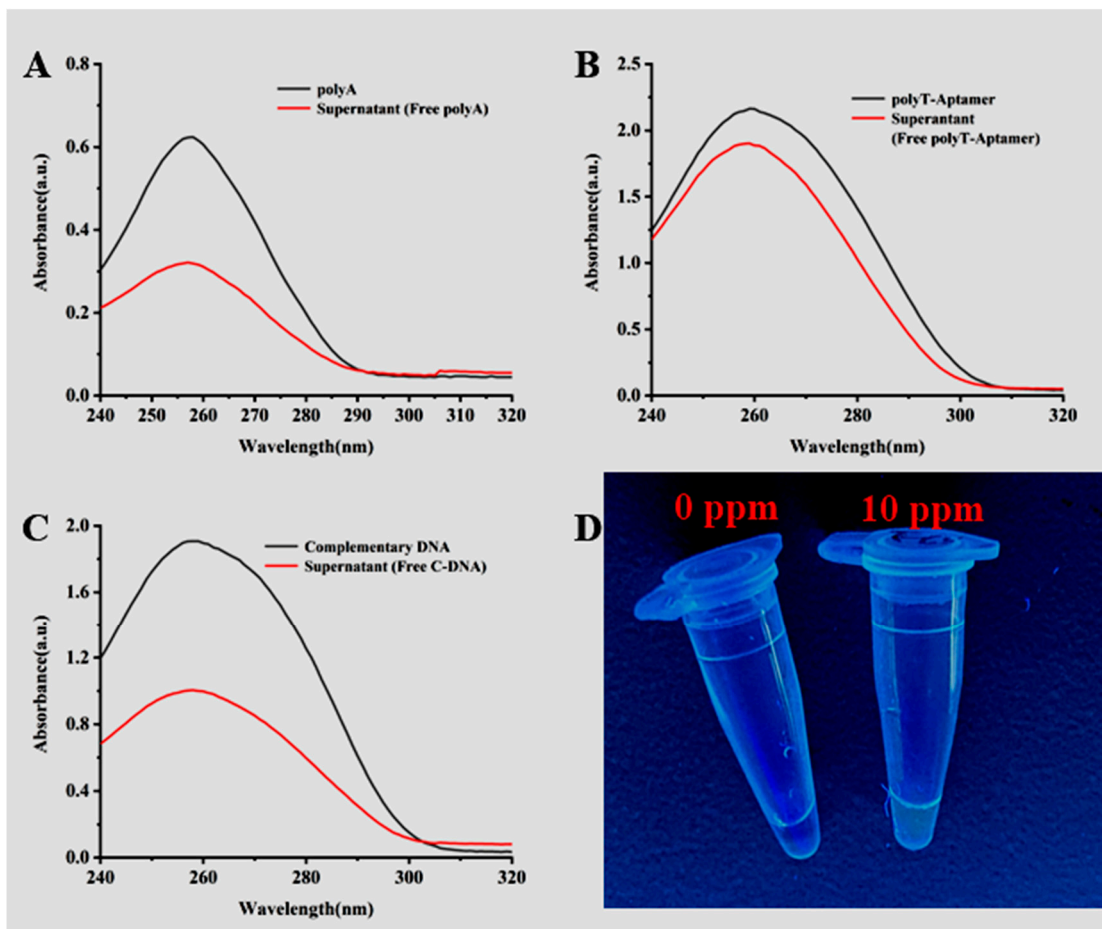
### 3.3. Validation of the Specificity of the Aptamer Sensor

The specificity of the test method was evaluated and the selected pesticides and glyphosate were tested under the same experimental conditions. The specificity was identified by observing the absorbance change of the supernatant and whether the qPCR could obtain the fluorescence signal. As shown in Figure 3A, the addition of other insecticides cannot trigger the binding of the aptamer to them, so the absorbance of the supernatant has not changed, indicating that the detection method has good specificity for glyphosate. As shown in Figure 2B, a strong fluorescence signal can only be generated when glyphosate is detected. As mentioned above, the specificity of the test method is reasonable.

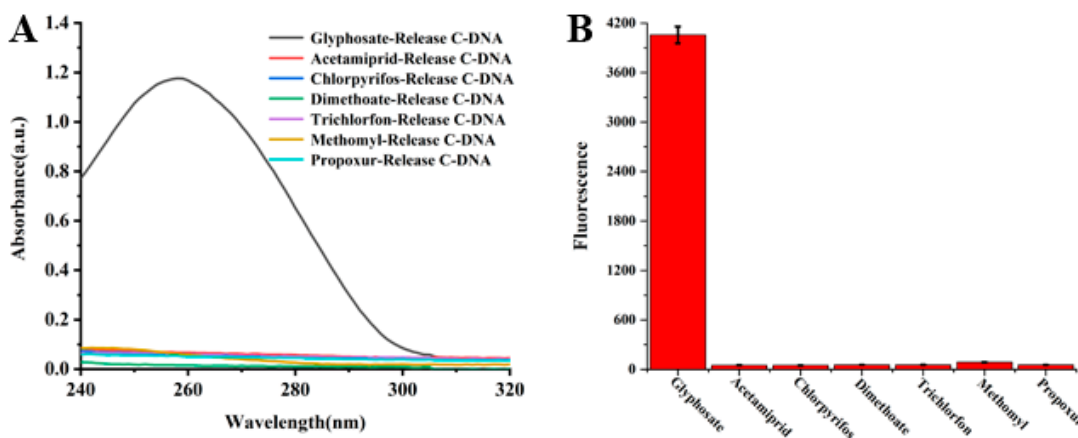
### 3.4. Glyphosate Detection and qPCRs Based on Mbs@dsDNA

In the presence of glyphosate, the C-DNA produced by its specific recognition by Mbs@dsDNA enters the qPCR to obtain a CT value. The CT value is related to the concentration of glyphosate. Even though the difference in absorbance in the supernatant was insignificant, the higher the concentration of glyphosate, the higher the CT value produced. Therefore, CT values are positively correlated with the concentration of glyphosate. Figure 4A,B shows that the absorbance in the supernatant and the CT value increased with increasing glyphosate concentration. There was a good linear relationship between the CT values and the logarithm of the concentration in the range of 0.1–5 ppm glyphosate. The linear equation is  $y = -4.0799x + 26.20509$ . The linear correlation coefficient is  $R^2 = 0.99742$  and the detection limit obtained by the experiment can reach 0.6  $\mu\text{mol/L}$ ; the national standard value is 3  $\mu\text{mol/L}$ , which is far lower than the maximum residue limit of glyphosate in the national standard.

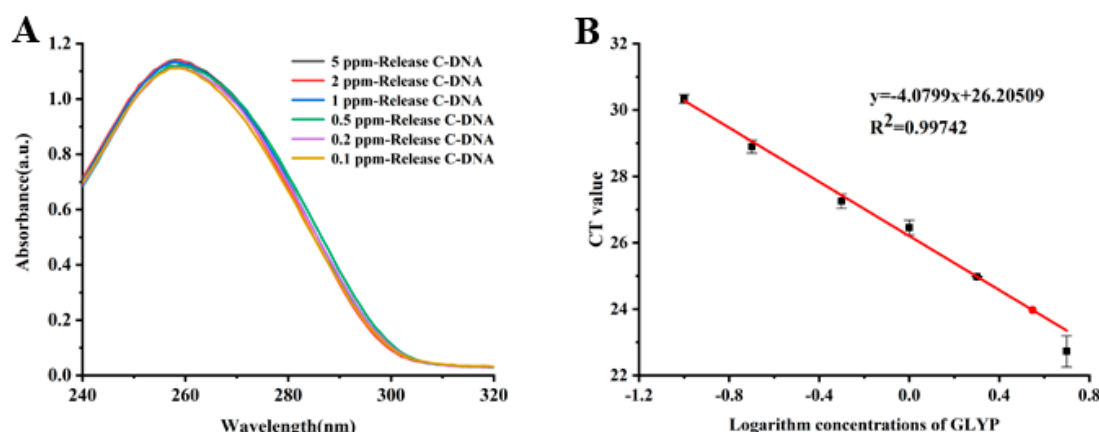




**Figure 2.** (A) UV-Vis absorption spectrum: polyA (black), supernatant (free polyA) (red), (B) UV-Vis absorption spectrum: polyT-Aptamer (black), supernatant (free polyT-Aptamer) (red), (C) UV-Vis absorption spectrum: complementary DNA (black), supernatant (free C-DNA) (red), (D) Fluorescent photos under a 365 nm UV lamp.



**Figure 3.** (A) Different pesticides and Mbs@dsDNA UV-Vis absorption spectrum of C-DNA released after incubation; (B) Fluorescence intensity histogram at 520 nm.



**Figure 4.** (A) Different concentrations of glyphosate and Mbs@dsDNA UV-Vis absorption spectra of C-DNA released after incubation. (B) Standard curve of the CT value and logarithm concentration of glyphosate.

### 3.5. Application of This and Other Methods to the Detection of Glyphosate in Real Samples

To further verify the practicability of the method in the actual scene, tap water spiked with glyphosate was used as a real sample for detection. The validity of the method was confirmed by the reasonable range of recovery (03.4~104.9%) and relative standard deviation (0.4~0.73%) in the supporting information (Table S3). To illustrate the method's ability to detect glyphosate, a crosswise comparison was carried out among all sorts of available technologies in Table 1. Different detection methods detect different kinds of actual samples, so the LOD is different. As shown in Table 1, glyphosate is primarily detected in agricultural products and tap water. Tap water is used as the real sample model in this experiment. The results are consistent with other detection methods, indicating that the developed adaptive sensor has good accuracy and is superior to some detection methods.

**Table 1.** Performance evaluation of glyphosate analysis technology.

Method	LOD	Recovery	Sample	Reference
GC-MS	3 $\mu\text{mol/L}$	96.7–107.7%	Serum and urine	[45]
ELISA	0.047 $\mu\text{mol/L}$	87.4–103.7%	River water, Tea and soil	[46]
HPLC	0.004 $\mu\text{mol/L}$	80.1–109.4%	Natural water	[47]
LC	6 $\mu\text{mol/L}$	80.63–97.11%	Soil	[48]
IC	30 $\mu\text{mol/L}$	80–110%	Honey	[49]
CE	800 $\text{mmol/L}$	89.4–93.7%	Hemp	[50]
LC-MS/MS	6 $\mu\text{mol/L}$	97–110%	Breast milk	[51]
EC	0.16 $\text{mmol/L}$	72.7–98.96%	Cucumber	[52]
qPCR	0.6 $\mu\text{mol/L}$	103.4–104.9%	Tap water	This work

## 4. Conclusions

In conclusion, by utilizing the specific recognition function of the aptamer, a biosensor for the recognition of glyphosate based on the aptamer, magnetic beads and qPCR was invented. The linear range of the method was 30  $\text{mmol/L}$  to 0.6  $\mu\text{mol/L}$  and the lowest detection limit detected by the experiment was 0.6  $\mu\text{mol/L}$ . The detection technology can be applied to detect glyphosate in actual samples (such as tap water) and has good potential for application. However, the method can still be further reduced in terms of detection limit and errors due to sampling pretreatment. Therefore, when these problems are solved in the future, it will be more beneficial for other pesticides.

**Supplementary Materials:** The following supporting information can be downloaded at: <https://www.mdpi.com/article/10.3390/s23020649/s1>, Table S1. Sequences of Oligonucleotides Used in This Work; Table S2. qPCR conditions; Table S3. Estimation of glyphosate recovery based on this method. Reference [53] is cited in the supplementary materials.

**Author Contributions:** M.W. and Y.S. (Yongxin She) conceived and designed the experiments. The experiments were performed by Y.S. (Yong Shao), R.T. and J.D. Data were analyzed by F.J., J.C., G.L. and Z.C. The paper was written by R.T., J.D. and M.W. The language and formal analysis of the paper was revised and polished by A.M.A.E.-A. All authors have read and agreed to the published version of the manuscript.

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## References

1. Benbrook, C.M. Trends in glyphosate herbicide use in the United States and globally. *Environ. Sci. Eur.* **2016**, *28*, 1–15. [[CrossRef](#)] [[PubMed](#)]
2. Gill, J.P.K.; Sethi, N.; Mohan, A. Analysis of the glyphosate herbicide in water, soil and food using derivatising agents. *Environ. Chem. Lett.* **2017**, *15*, 85–100. [[CrossRef](#)]
3. Jimenez-Lopez, J.; Llorent-Martinez, E.J.; Ortega-Barrales, P.; Ruiz-Medina, A. Graphene quantum dots-silver nanoparticles as a novel sensitive and selective luminescence probe for the detection of glyphosate in food samples. *Talanta* **2020**, *207*, 120344. [[CrossRef](#)] [[PubMed](#)]
4. Liu, Z.; Yang, L.; Sharma, A.S.; Chen, M.; Chen, Q. A system composed of polyethylenimine-capped upconversion nanoparticles, copper (II), hydrogen peroxide and 3,3',5,5'-tetramethylbenzidine for colorimetric and fluorometric determination of glyphosate. *Mikrochim. Acta* **2019**, *186*, 835. [[CrossRef](#)] [[PubMed](#)]
5. Chang, Y.C.; Lin, Y.S.; Xiao, G.T.; Chiu, T.C.; Hu, C.C. A highly selective and sensitive nanosensor for the detection of glyphosate. *Talanta* **2016**, *161*, 94–98. [[CrossRef](#)]
6. Duke, S.O.; Powles, S.B. Glyphosate: A once-in-a-century herbicide. *Pest Manag. Sci.* **2008**, *64*, 319–325. [[CrossRef](#)]
7. Prasad, B.B.; Jauhari, D.; Tiwari, M.P. Doubly imprinted polymer nanofilm-modified electrochemical sensor for ultra-trace simultaneous analysis of glyphosate and glufosinate. *Biosens. Bioelectron.* **2014**, *59*, 81–88. [[CrossRef](#)]
8. Regiart, M.; Kumar, A.; Goncalves, J.M.; Silva, G.J.; Masini, J.C.; Angnes, L.; Bertotti, M. An Electrochemically Synthesized Nanoporous Copper Microsensor for Highly Sensitive and Selective Determination of Glyphosate. *Chemelectrochem* **2020**, *7*, 1558–1566. [[CrossRef](#)]
9. Singh, S.; Kumar, V.; Datta, S.; Wani, A.B.; Dhanjal, D.S.; Romero, R.; Singh, J. Glyphosate uptake, translocation, resistance emergence in crops, analytical monitoring, toxicity and degradation: A review. *Environ. Chem. Lett.* **2020**, *18*, 663–702. [[CrossRef](#)]
10. Bringolf, R.B.; Cope, W.G.; Mosher, S.; Barnhart, M.C.; Shea, D. Acute and chronic toxicity of glyphosate compounds to glochidia and juveniles of *Lampsilis siliquoidea* (Unionidae). *Environ. Toxicol. Chem.* **2007**, *26*, 2094–2100. [[CrossRef](#)]
11. Xu, J.; Smith, S.; Smith, G.; Wang, W.; Li, Y. Glyphosate contamination in grains and foods: An overview. *Food Control* **2019**, *106*, 106710. [[CrossRef](#)]
12. Araujo, A.S.; Monteiro, R.T.; Abarkeli, R.B. Effect of glyphosate on the microbial activity of two Brazilian soils. *Chemosphere* **2003**, *52*, 799–804. [[CrossRef](#)] [[PubMed](#)]
13. Khrolenko, M.V.; Wieczorek, P.P. Determination of glyphosate and its metabolite aminomethylphosphonic acid in fruit juices using supported-liquid membrane preconcentration method with high-performance liquid chromatography and UV detection after derivatization with p-toluenesulphonyl chloride. *J. Chromatogr. A* **2005**, *1093*, 111–117. [[PubMed](#)]
14. Connolly, A.; Koslitz, S.; Bury, D.; Bruning, T.; Conrad, A.; Kolossa-Gehring, M.; Coggins, M.A.; Koch, H.M. Sensitive and selective quantification of glyphosate and aminomethylphosphonic acid (AMPA) in urine of the general population by gas chromatography-tandem mass spectrometry. *J. Chromatogr. B Analyt. Technol. Biomed. Life Sci.* **2020**, *1158*, 122348. [[CrossRef](#)] [[PubMed](#)]
15. Kudzin, Z.H.; Gralak, D.K.; Drabowicz, J.; Luczak, J. Novel approach for the simultaneous analysis of glyphosate and its metabolites. *J. Chromatogr. A* **2002**, *947*, 129–141. [[CrossRef](#)]
16. Morgan, M.A.; Griffith, C.M.; Dinges, M.M.; Lyon, Y.A.; Julian, R.R.; Larive, C.K. Evaluating sub-lethal stress from Roundup® exposure in *Artemia franciscana* using (1)H NMR and GC-MS. *Aquat. Toxicol.* **2019**, *212*, 77–87. [[CrossRef](#)]



17. Bressan, I.G.; Llesuy, S.F.; Rodriguez, C.; Ferloni, A.; Dawidowski, A.R.; Figar, S.B.; Gimenez, M.I. Optimization and validation of a liquid chromatography-tandem mass spectrometry method for the determination of glyphosate in human urine after pre-column derivatization with 9-fluorenylmethoxycarbonyl chloride. *J. Chromatogr. B Anal. Technol. Biomed. Life Sci.* **2021**, *1171*, 122616. [[CrossRef](#)]
18. Gormez, E.; Golge, O.; Kabak, B. Quantification of fosetyl-aluminium /phosphonic acid and other highly polar residues in pomegranates using Quick Polar Pesticides method involving liquid chromatography-tandem mass spectrometry measurement. *J. Chromatogr. A* **2021**, *1642*, 462038. [[CrossRef](#)]
19. Jost, U.; Habedank, F. Two-dimensional hydrophilic interaction and reversed phase liquid chromatography easily extracted pesticides and polar pesticides multi-residue method—A concept. *J. Chromatogr. A* **2020**, *1621*, 461040. [[CrossRef](#)]
20. Thompson, T.S.; van den Heever, J.P.; Limanowka, R.E. Determination of glyphosate, AMPA, and glufosinate in honey by online solid-phase extraction-liquid chromatography-tandem mass spectrometry. *Food Addit. Contam. Part A* **2019**, *36*, 434–446. [[CrossRef](#)]
21. Geerdink, R.B.; Hassing, M.; Ayarza, N.; Bruggink, C.; Wielheesen, M.; Claassen, J.; Epema, O.J. Analysis of glyphosate, AMPA, Glufosinate and MPPA with ION chromatography tandem mass spectrometry using A membrane suppressor in the ammonium form application to surface water of low to moderate salinity. *Anal. Chim. Acta* **2020**, *1133*, 66–76. [[CrossRef](#)] [[PubMed](#)]
22. Schutze, A.; Morales-Agudelo, P.; Vidal, M.; Calafat, A.M.; Ospina, M. Quantification of glyphosate and other organophosphorus compounds in human urine via ion chromatography isotope dilution tandem mass spectrometry. *Chemosphere* **2021**, *274*, 129427. [[CrossRef](#)] [[PubMed](#)]
23. Cao, L.W.; Deng, T.; Liang, S.L.; Tan, X.F.; Meng, J.X. Determination of Herbicides and Its Metabolite in Soil and Water Samples by Capillary Electrophoresis-laser Induced Fluorescence Detection Using Microwave-assisted Derivatization. *Anal. Sci.* **2014**, *30*, 759–766. [[CrossRef](#)] [[PubMed](#)]
24. Wimmer, B.; Pattky, M.; Zada, L.G.; Meixner, M.; Haderlein, S.B.; Zimmermann, H.P.; Huhn, C. Capillary electrophoresis-mass spectrometry for the direct analysis of glyphosate: Method development and application to beer beverages and environmental studies. *Anal. Bioanal. Chem.* **2020**, *412*, 4967–4983. [[CrossRef](#)] [[PubMed](#)]
25. Byer, J.D.; Struger, J.; Klawunn, P.; Todd, A.; Sverko, E. Low cost monitoring of glyphosate in surface waters using the ELISA method: An evaluation. *Environ. Sci. Technol.* **2008**, *42*, 6052–6057. [[CrossRef](#)]
26. El-Gendy, K.; Mosallam, E.; Ahmed, N.; Aly, N. Determination of glyphosate residues in Egyptian soil samples. *Anal. Biochem.* **2018**, *557*, 1–6. [[CrossRef](#)]
27. Guan, J.; Yang, J.; Zhang, Y.; Zhang, X.; Deng, H.; Xu, J.; Wang, J.; Yuan, M.S. Employing a fluorescent and colorimetric picolyl-functionalized rhodamine for the detection of glyphosate pesticide. *Talanta* **2021**, *224*, 121834. [[CrossRef](#)]
28. Sasaki, Y.; Asano, K.; Minamiki, T.; Zhang, Z.; Takizawa, S.Y.; Kubota, R.; Minami, T. A Water-Gated Organic Thin-Film Transistor for Glyphosate Detection: A Comparative Study with Fluorescence Sensing. *Chemistry* **2020**, *26*, 14506. [[CrossRef](#)]
29. Habekost, A. Rapid and sensitive spectroelectrochemical and electrochemical detection of glyphosate and AMPA with screen-printed electrodes. *Talanta* **2017**, *162*, 583–588. [[CrossRef](#)]
30. Sahoo, D.; Mandal, A.; Mitra, T.; Chakraborty, K.; Bardhan, M.; Dasgupta, A.K. Nanosensing of Pesticides by Zinc Oxide Quantum Dot: An Optical and Electrochemical Approach for the Detection of Pesticides in Water. *J. Agric. Food Chem.* **2018**, *66*, 414–423. [[CrossRef](#)]
31. Kalra, P.; Dhiman, A.; Cho, W.C.; Bruno, J.G.; Sharma, T.K. Simple Methods and Rational Design for Enhancing Aptamer Sensitivity and Specificity. *Front. Mol. Biosci.* **2018**, *5*, 41. [[CrossRef](#)] [[PubMed](#)]
32. Shaban, S.M.; Kim, D.H. Recent Advances in Aptamer Sensors. *Sensors* **2021**, *21*, 979. [[CrossRef](#)] [[PubMed](#)]
33. Sharma, T.K.; Bruno, J.G.; Dhiman, A. ABCs of DNA aptamer and related assay development. *Biotechnol. Adv.* **2017**, *35*, 275–301. [[CrossRef](#)]
34. Tuerk, C.; Gold, L. Systematic evolution of ligands by exponential enrichment: RNA ligands to bacteriophage T4 DNA polymerase. *Science* **1990**, *249*, 505–510. [[CrossRef](#)] [[PubMed](#)]
35. Li, L.; Jiang, Y.; Cui, C.; Yang, Y.; Zhang, P.; Stewart, K.; Pan, X.; Li, X.; Yang, L.; Qiu, L.; et al. Modulating Aptamer Specificity with pH-Responsive DNA Bonds. *J. Am. Chem. Soc.* **2018**, *140*, 13335–13339. [[CrossRef](#)]
36. Ojha, Y.R.; Giovannucci, D.R.; Cameron, B.D. Selection and characterization of structure-switching DNA aptamers for the salivary peptide histatin 3. *J. Biotechnol.* **2021**, *327*, 9–17. [[CrossRef](#)]
37. Ren, Q.; Mou, J.; Guo, Y.; Wang, H.; Cao, X.; Zhang, F.; Xia, J.; Wang, Z. Simple homogeneous electrochemical target-responsive aptasensor based on aptamer bio-gated and porous carbon nanocontainer derived from ZIF-8. *Biosens. Bioelectron.* **2020**, *166*, 112448. [[CrossRef](#)]
38. Dong, H.; Chen, H.; Jiang, J.; Zhang, H.; Cai, C.; Shen, Q. Highly Sensitive Electrochemical Detection of Tumor Exosomes Based on Aptamer Recognition-Induced Multi-DNA Release and Cyclic Enzymatic Amplification. *Anal. Chem.* **2018**, *90*, 4507–4513. [[CrossRef](#)]
39. Shi, H.; He, X.; Wang, K.; Wu, X.; Ye, X.; Guo, Q.; Tan, W.; Qing, Z.; Yang, X.; Zhou, B. Activatable aptamer probe for contrast-enhanced in vivo cancer imaging based on cell membrane protein-triggered conformation alteration. *Proc. Natl. Acad. Sci. USA* **2011**, *108*, 3900–3905. [[CrossRef](#)]
40. Zhang, J.J.; Cheng, F.F.; Zheng, T.T.; Zhu, J.J. Versatile aptasensor for electrochemical quantification of cell surface glycan and naked-eye tracking glycolytic inhibition in living cells. *Biosens. Bioelectron.* **2017**, *89 Pt 2*, 937–945. [[CrossRef](#)]

41. Selvolini, G.; Bajan, I.; Hosu, O.; Cristea, C.; Sandulescu, R.; Marrazza, G. DNA-Based Sensor for the Detection of an Organophosphorus Pesticide: Profenofos. *Sensors* **2018**, *18*, 2035. [[CrossRef](#)] [[PubMed](#)]
42. Hong, C.; Zhang, X.; Ye, S.; Yang, H.; Huang, Z.; Yang, D.; Cai, R.; Tan, W. Aptamer-Pendant DNA Tetrahedron Nanostructure Probe for Ultrasensitive Detection of Tetracycline by Coupling Target-Triggered Rolling Circle Amplification. *ACS Appl. Mater. Interfaces* **2021**, *13*, 19695–19700. [[CrossRef](#)] [[PubMed](#)]
43. Trinh, K.H.; Kadam, U.S.; Rampogu, S.; Cho, Y.; Yang, K.A.; Kang, C.H.; Lee, K.W.; Lee, K.O.; Chung, W.S.; Hong, J.C. Development of novel fluorescence-based and label-free noncanonical G4-quadruplex-like DNA biosensor for facile, specific, and ultrasensitive detection of fipronil. *J. Hazard Mater.* **2022**, *427*, 127939. [[CrossRef](#)] [[PubMed](#)]
44. Trinh, K.H.; Kadam, U.S.; Song, J.; Cho, Y.; Kang, C.H.; Lee, K.O.; Lim, C.O.; Chung, W.S.; Hong, J.C. Novel DNA Aptameric Sensors to Detect the Toxic Insecticide Fenitrothion. *Int. J. Mol. Sci.* **2021**, *22*, 10846. [[CrossRef](#)]
45. Saito, T.; Aoki, H.; Namera, A.; Oikawa, H.; Miyazaki, S.; Nakamoto, A.; Inokuchi, S. Mix-mode TiO-C18 monolith spin column extraction and GC-MS for the simultaneous assay of organophosphorus compounds and glufosinate, and glyphosate in human serum and urine. *Anal. Sci.* **2011**, *27*, 999–1005. [[CrossRef](#)]
46. Wang, D.; Lin, B.; Cao, Y.; Guo, M.; Yu, Y. A Highly Selective and Sensitive Fluorescence Detection Method of Glyphosate Based on an Immune Reaction Strategy of Carbon Dot Labeled Antibody and Antigen Magnetic Beads. *J. Agric. Food Chem.* **2016**, *64*, 6042–6050. [[CrossRef](#)]
47. Vu, C.T.; Le, P.T.; Chu, D.B.; Bui, V.H.; Phung, T.L.A.; Nguyen Le, H.Y.; Labanowski, J.; Mondamert, L.; Herrmann, M.; Behra, P. One-step purification/extraction method to access glyphosate, glufosinate, and their metabolites in natural waters. *J. Chromatogr. A* **2021**, *1649*, 462188. [[CrossRef](#)]
48. Druart, C.; Delhomme, O.; de Vaufleury, A.; Ntcho, E.; Millet, M. Optimization of extraction procedure and chromatographic separation of glyphosate, glufosinate and aminomethylphosphonic acid in soil. *Anal. Bioanal. Chem.* **2011**, *399*, 1725–1732. [[CrossRef](#)]
49. Pareja, L.; Jesús, F.; Heinzen, H.; Hernando, M.D.; Rajski, Ł.; Fernández-Alba, A.R. Evaluation of glyphosate and AMPA in honey by water extraction followed by ion chromatography mass spectrometry. A pilot monitoring study. *Anal. Methods* **2019**, *11*, 2123–2128. [[CrossRef](#)]
50. Lanaro, R.; Costa, J.L.; Cazenave, S.O.; Zanolli-Filho, L.A.; Tavares, M.F.; Chasin, A.A. Determination of herbicides paraquat, glyphosate, and aminomethylphosphonic acid in marijuana samples by capillary electrophoresis. *J. Forensic. Sci.* **2015**, *60* (Suppl. 1), S241–S247. [[CrossRef](#)]
51. Steinborn, A.; Alder, L.; Michalski, B.; Zomer, P.; Bendig, P.; Martinez, S.A.; Mol, H.G.; Class, T.J.; Pinheiro, N.C. Determination of Glyphosate Levels in Breast Milk Samples from Germany by LC-MS/MS and GC-MS/MS. *J. Agric. Food Chem.* **2016**, *64*, 1414–1421. [[CrossRef](#)] [[PubMed](#)]
52. Zhang, C.; She, Y.; Li, T.; Zhao, F.; Jin, M.; Guo, Y.; Zheng, L.; Wang, S.; Jin, F.; Shao, H.; et al. A highly selective electrochemical sensor based on molecularly imprinted polypyrrole-modified gold electrode for the determination of glyphosate in cucumber and tap water. *Anal. Bioanal. Chem.* **2017**, *409*, 7133–7144. [[CrossRef](#)] [[PubMed](#)]
53. Chen, F.; Li, G.; Liu, H.; Leung, C.-H.; Ma, D.-L. G-quadruplex-based detection of glyphosate in complex biological systems by a time-resolved luminescent assay. *Sens. Actuators B Chem.* **2020**, *320*, 128393. [[CrossRef](#)]

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