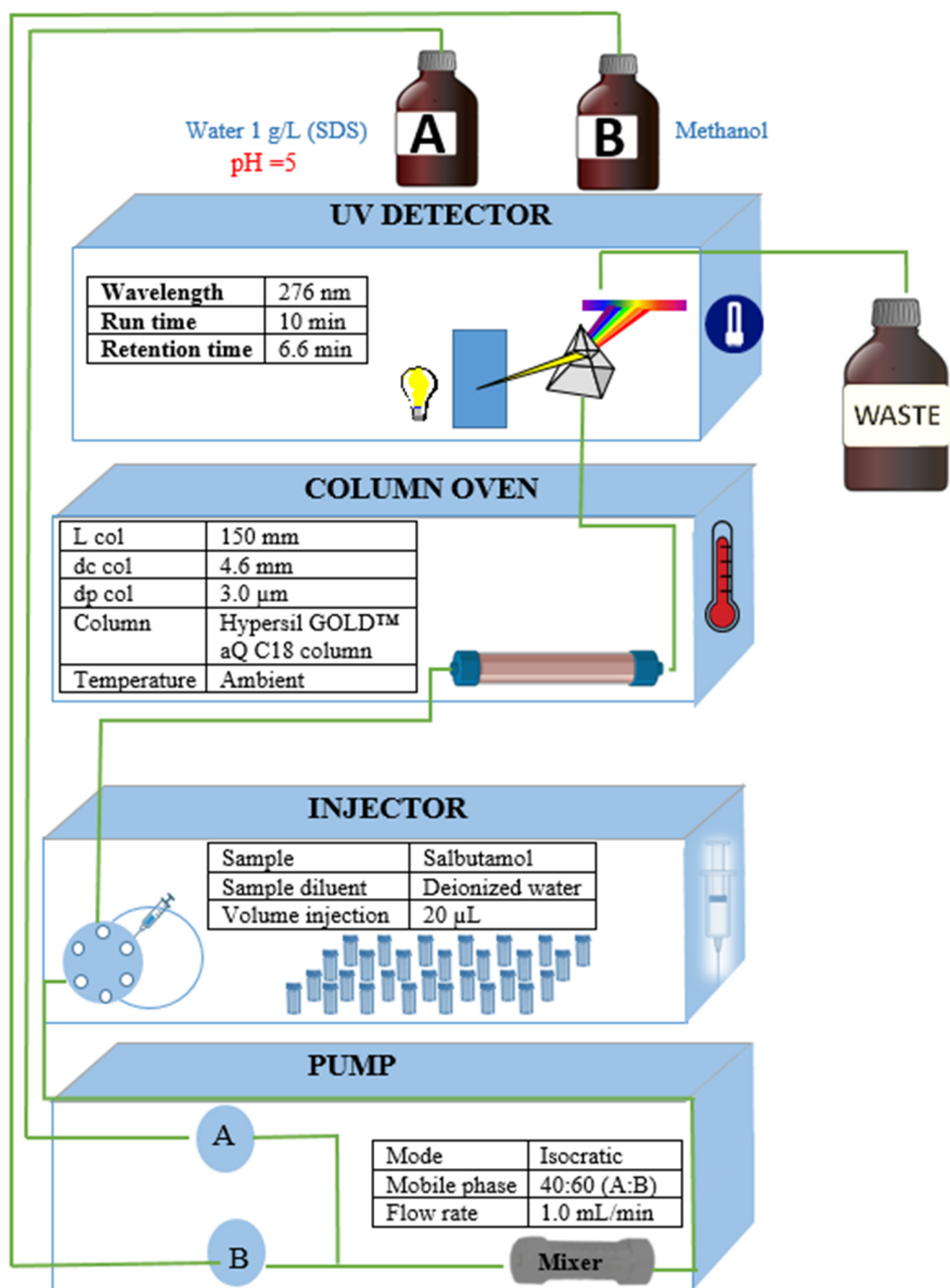
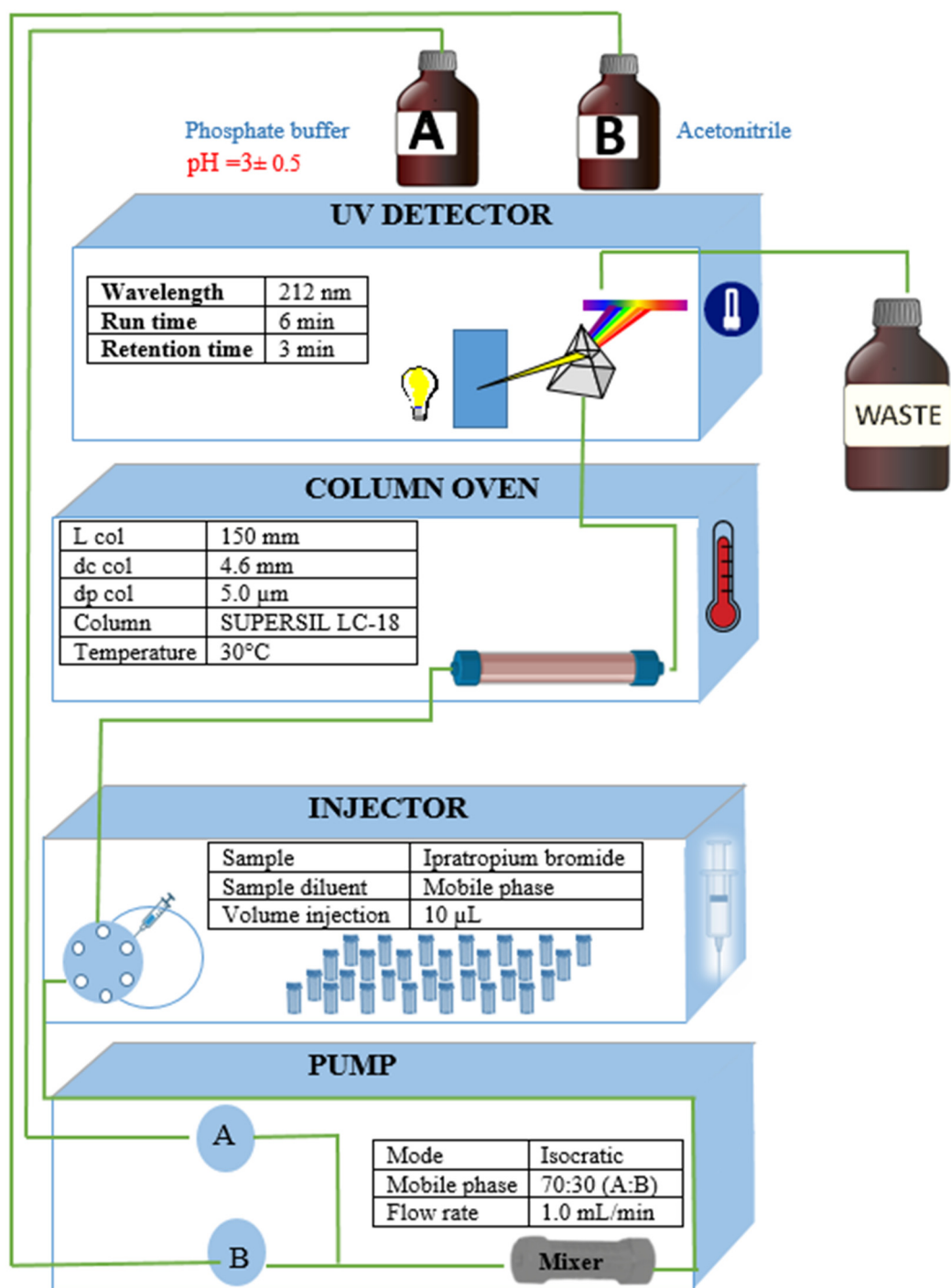


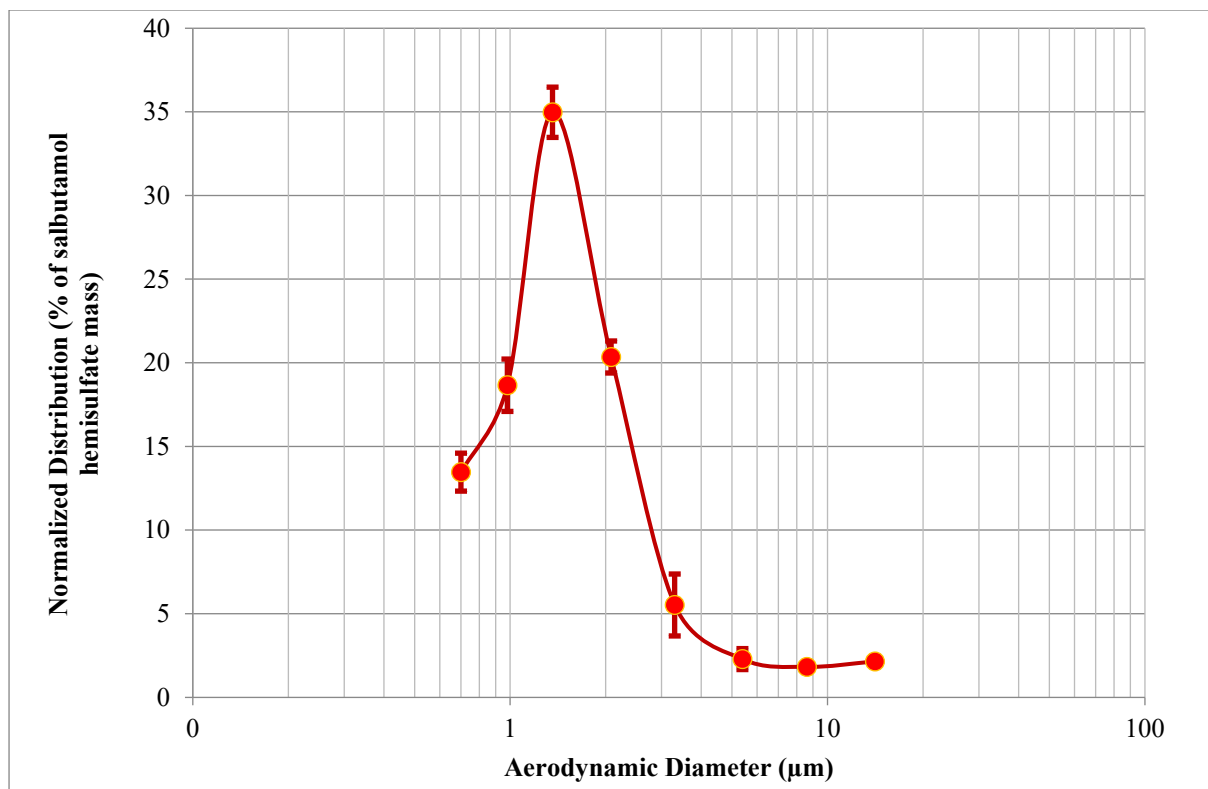
Supplementary material Appendix 1, Figure S1- Chromatographic conditions for Terbutaline quantification.



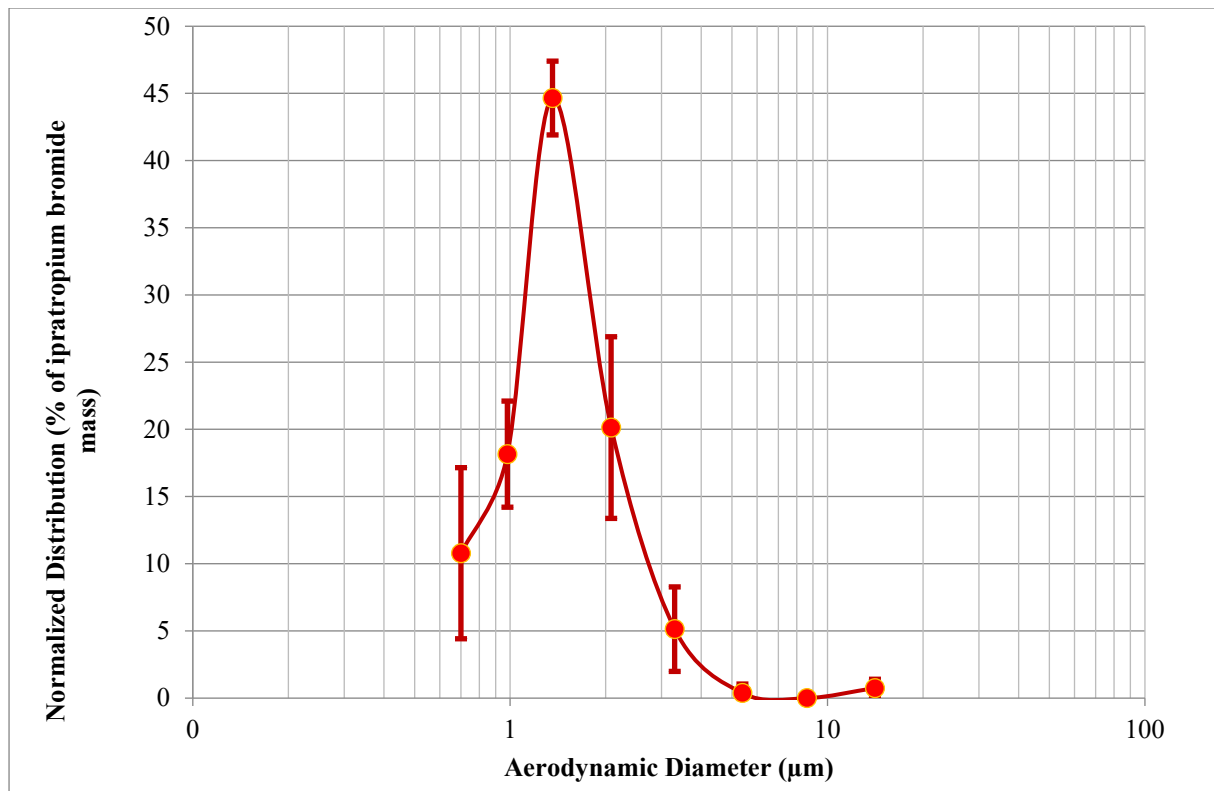
Supplementary material Appendix 1, Figure S2- Chromatographic conditions for Salbutamol hemisulfate quantification.



Supplementary material Appendix 1, Figure S3- Chromatographic conditions for Ipratropium bromide quantification.



Supplementary material Appendix 2, Figure S1. NGI (Next Generation Impactor) Impactor-collected data showing the MMAD (mass median aerodynamic diameter) of a jet nebulizer (cirrus 2 TM). Results are expressed in  $\mu\text{g}$  of salbutamol hemisulfate (mean  $\pm$  SD,  $n = 3$ ).



Supplementary material Appendix 2, Figure S2. NGI (Next Generation Impactor) Impactor-collected data showing the MMAD (mass median aerodynamic diameter) of a jet nebulizer (cirrus 2 TM). Results are expressed in  $\mu\text{g}$  of ipratropium bromide (mean  $\pm$  SD,  $n = 3$ ).

## **Supplementary data Appendix 3-Validation protocol for the quantification of salbutamol hemisulfate**

- **Detection method**

The method determines salbutamol hemisulfate (bulk form) dissolved in 0.9% NaCl. Samples are analyzed using a Shimadzu HPLC system (Shimadzu, Tokyo, Japan).

- **Calibration of standard solutions**

Calibration solutions were obtained by dilution of 20mg of salbutamol hemisulfate in 10mL of 0.9% NaCl. Thus, a stock solution of  $[C] = 2000\mu\text{g/mL}$  was obtained. Standard solutions were then obtained by further dilution.

Calibration levels of salbutamol hemisulfate are shown below:

<b>Cal 1 (<math>\mu\text{g/mL}</math>)</b>	<b>Cal 2(<math>\mu\text{g/mL}</math>)</b>	<b>Cal 3(<math>\mu\text{g/mL}</math>)</b>	<b>Cal 4(<math>\mu\text{g/mL}</math>)</b>	<b>Cal 5(<math>\mu\text{g/mL}</math>)</b>	<b>Cal 6(<math>\mu\text{g/mL}</math>)</b>
0	600	800	1000	1200	1400

*Table 1 Calibration levels for salbutamol hemisulfate.*

*N.B: Five calibration points are enough to validate the linearity parameter. A  $[C] = 0\mu\text{g/mL}$  is mainly used for comparison with higher concentrations to predict the exact retention time and peak of the desired molecule.*

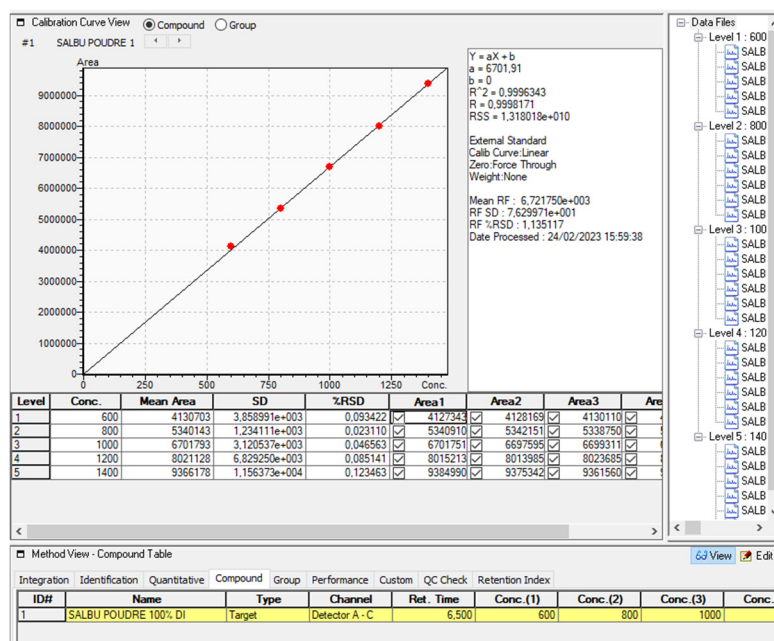
- **Linearity**

Linearity was evaluated in the range of 0 to  $1400\mu\text{g/mL}$  for salbutamol hemisulfate (bulk form).

Linear relationship was observed in the plot of area (mAU.s) as a function of concentration ( $\mu\text{g/mL}$ ).

Before launching any calibration vial, we used a “front” calibration solution in order to ensure the accuracy of the values that will be obtained on the next vials.

Figure S1 shows the calibration curve obtained via this method.



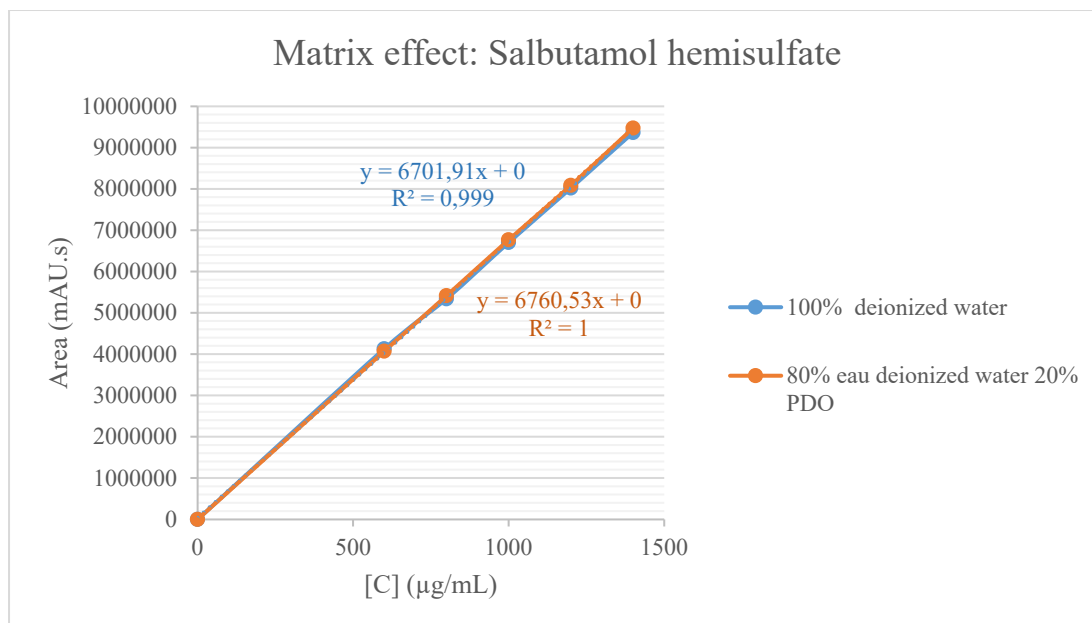
The results obtained for linearity, fully comply with the acceptance criteria set by the ICH guidelines, as the calibration curve has a linear regression coefficient of determination with  $R^2 > 0.999$ .

- Matrix effect

The matrix effect refers to the interference caused by components of a sample matrix on the measurement of the analyte of interest in an analytical method.

Since we are using e-liquid in our formulation (1, 3-propanediol), it is important to understand and characterize matrix effect to ensure the accuracy and precision of the method.

Consequently, to evaluate the matrix effect, it is essential to take into account the equations assigned in each of the calibration curves (graph below).



Through these data, we calculate the relative difference of the slopes for each solvent in order to determine the presence or the absence of the matrix effect:

- Salbutamol hemisulfate in: 100% deionized water:  $6701.91 X + 0$
- Salbutamol hemisulfate in: 20% (1,3-propanediol), 80% deionized water:  $6760.53 X + 0$

The relative difference of the slopes for each solvent is calculated according to the following formula:

Relative difference=  $\left[ \frac{(\text{Solvent slope in matrix} - \text{Solvent slope without matrix})}{\text{Solvent slope in matrix}} \right] * 100$

This relative difference must be less than 10% in order to consider the matrix effect as acceptable.

Relative difference=  $\left[ \frac{(6760.53 - 6701.91)}{6760.53} * 100 \right] = 0.86\% < 10\%$

➤ **Conclusion: absence of matrix effect with the combination of 1,3 propanediol (PDO) and salbutamol hemisulfate.**



- Limit of detection and quantification

Limit of detection (LOD) and limit of quantification (LOQ) are two important concepts in analytical chemistry that relate to the sensitivity of an analytical method.

The limit of detection (LOD) is the lowest amount of an analyte in a sample that can be detected.

The limit of quantification (LOQ) is the lowest amount of an analyte in a sample that can be quantified.

These two parameters are based on the standard deviation of the response ( $\sigma$ ) and the slope ( $S$ ) of the calibration curve

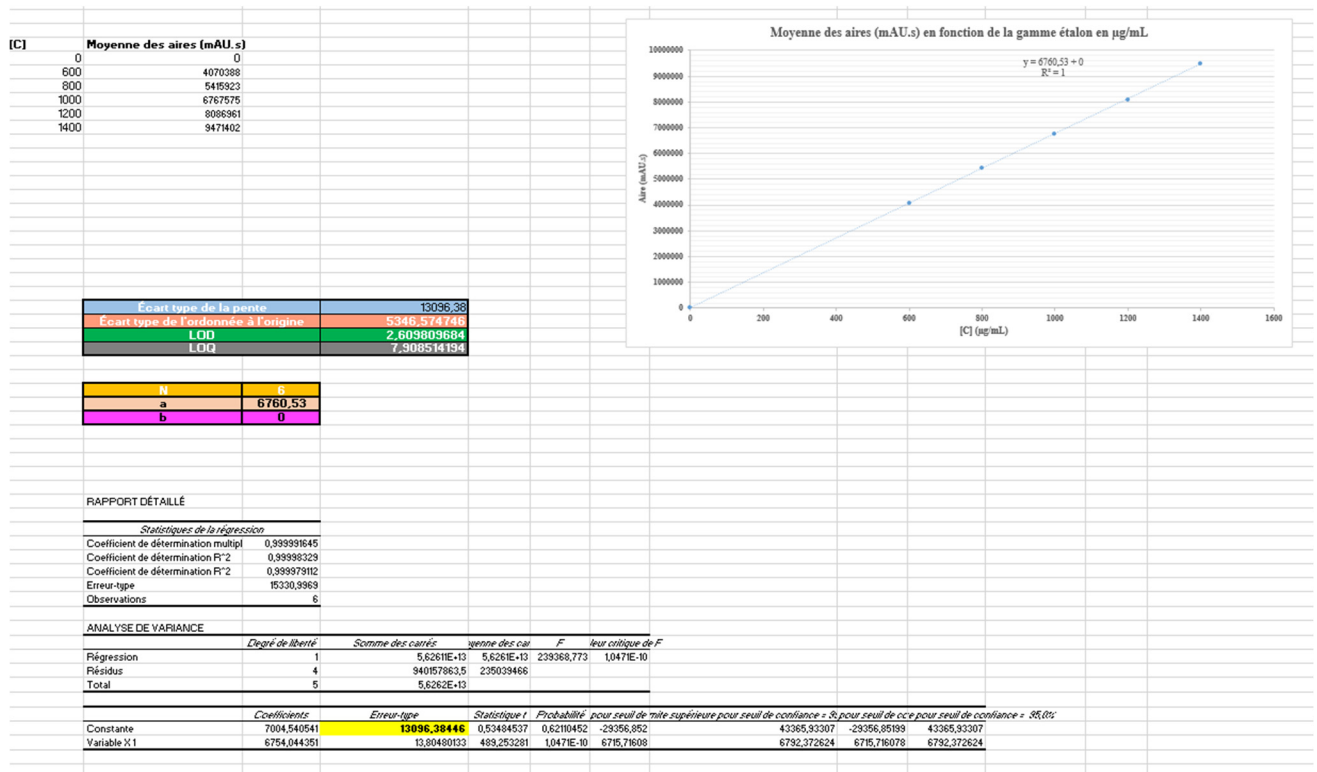
*LOD equation*

$$LOD = 3.3 \times \frac{\sigma}{S}$$

*LOQ equation*

$$LOQ = 10 \times \frac{\sigma}{S}$$

Where  $S$  is the slope of the calibration line,  $\sigma$  is the residual standard deviation of the regression line. The obtained values of: LOD and LOQ were 2.60 and 7.90  $\mu\text{g/mL}$ , respectively.



LOD	2.60 µg/mL
LOQ	7.90 µg/mL

- Repeatability

Repeatability refers to the degree of agreement between multiple measurements of the same sample, under the same conditions, using the same analytical method and equipment. In other words, it measures the precision of the analytical method.

For this specific reason, the quantification was carried out on two different solutions with the same concentration ([C] = 1000 µg/mL).

N.B: the acquisition were launched during the same day, by the same operator and under similar experimental conditions.

Each solution was injected 6 times (as mentioned on the table below):

							Mean (µg/mL)
Solution 1	1004.729	1002.35	996.246	997.6	998.97	995.530	999.2375
Solution 2	998.75	999.79	1001.16	1001.14	1003.80	995.53	1000.0283

The given results make it possible to validate the repeatability parameter, as the obtained

values are equal to 1000µg/mL.

- Recovery

It involves spiking a known amount of an analyte into a sample, and analyzing the sample to determine the amount of analyte that is recovered after the sample preparation.

As a matter of fact, the recovery is calculated by comparing the measured amount of analyte in the spiked sample to the amount that would be expected to be present based on the known amount of analyte that was added.

This parameter is useful for determining the accuracy and precision of a sample preparation method and can help to ensure that the results obtained from an analytical method are reliable and reproducible.

The theoretical dosing concentration of salbutamol hemisulfate (2 mg/mL)

Group	Theoretical concentration (µg/mL )	Concentration found (µg/mL )	Recovery (%)
60%	600	602	100.33
		600	100
		601	100.16
80%	800	801	100.125
		799	99.875
		802	100.25
100%	1000	1002.349	100.2349
		996.246	99.6246
		997.598	99.7598
120%	1200	1200	100
		1191	99.25
		1200	100
140%	1400	1401.447	100.103357
		1400	100
		1400	100
Mean % =99.97			

Table 2: Recovery validation parameter.

**Supplementary data Appendix 4-Validation protocol for the quantification of ipratropium bromide**

- Detection method

The method determines ipratropium bromide (bulk form) dissolved in 0.9% NaCl. Samples are analyzed using a Shimadzu HPLC system (Shimadzu. Tokyo. Japan).

- Calibration of standard solutions

Calibration solutions were obtained by dilution of 10mg of ipratropium bromide in 10mL of 0.9% NaCl. Thus, a stock solution of  $[C] = 1000\mu\text{g/mL}$  was obtained. Standard solutions were then obtained by further dilution.

Calibration levels of ipratropium bromide are shown below:

Cal 1( $\mu\text{g/mL}$ )	Cal 2( $\mu\text{g/mL}$ )	Cal 3( $\mu\text{g/mL}$ )	Cal 4( $\mu\text{g/mL}$ )	Cal 5( $\mu\text{g/mL}$ )
75	100	125	150	175

Table 2 Calibration levels for salbutamol hemisulfate.

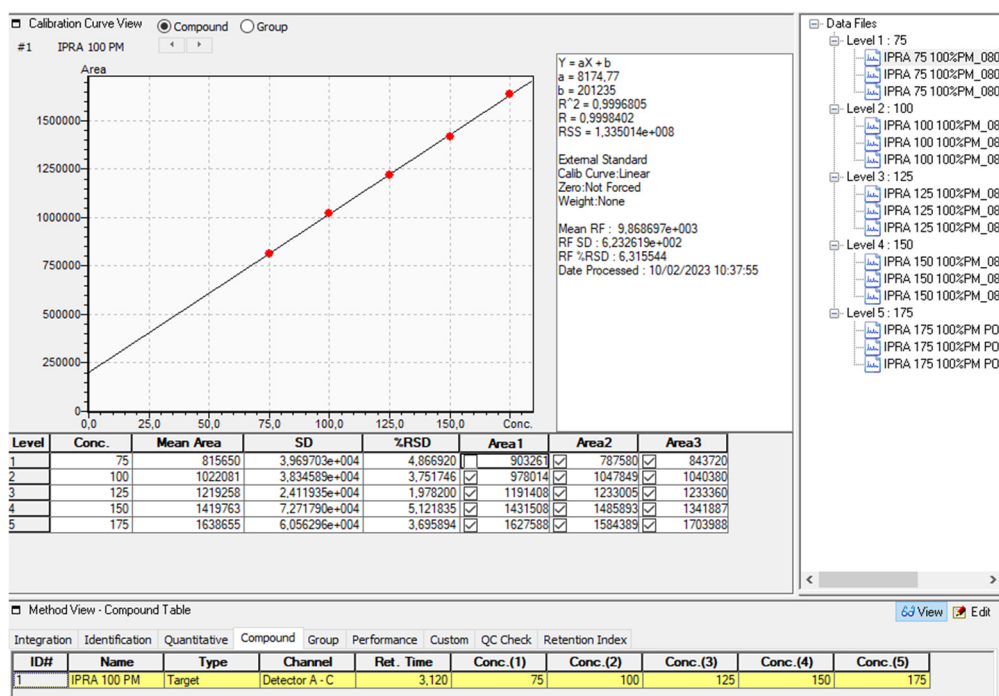
- Linearity

Linearity was evaluated in the range of 75 to 175 $\mu\text{g/mL}$  for ipratropium bromide (bulk form).

Linear relationship was observed in the plot of area (mAU.s) as a function of concentration ( $\mu\text{g/mL}$ ).

Before launching any calibration vial, we used a “front” calibration solution in order to ensure the accuracy of the values that will be obtained on the next vials.

Figure S1 shows the calibration curve obtained via this method.



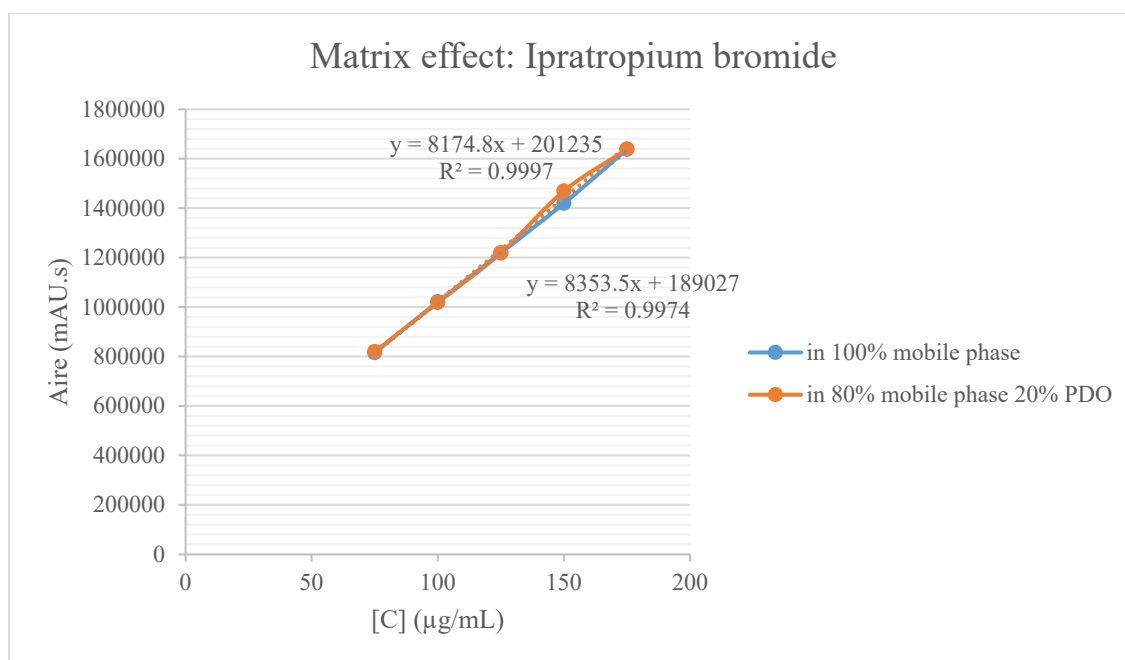
The results obtained for linearity, fully comply with the acceptance criteria set by the ICH guidelines, as the calibration curve has a linear regression coefficient of determination with  $R^2 > 0.999$ .

- Matrix effect

The matrix effect refers to the interference caused by components of a sample matrix on the measurement of the analyte of interest in an analytical method.

Since we are using e-liquid in our formulation (1, 3-propanediol), it is important to understand and characterize matrix effect to ensure the accuracy and precision of the method.

Consequently, to evaluate the matrix effect, it is essential to take into account the equations assigned in each of the calibration curves (graph below).



Through these data, we calculate the relative difference of the slopes for each solvent in order to determine the presence or the absence of the matrix effect:

- Ipratropium bromide in: 100% mobile phase (30% acetonitrile: 70% phosphate buffer):

$$8174.8 X + 201235$$

- Ipratropium bromide in: 20% (1,3-propanediol), 80% deionized water:  $8353.5 X + 189027$

The relative difference of the slopes for each solvent is calculated according to the following formula:

$$\text{Relative difference} = \left[ \frac{(\text{Solvent slope in matrix} - \text{Solvent slope without matrix})}{\text{Solvent slope in matrix}} \right] \times 100$$

This relative difference must be less than 10% in order to consider the matrix effect as acceptable.

$$\text{Relative difference} = \left[ \frac{(8353.5 - 8174.8)}{8353.5} \times 100 \right] = 2.13 < 10\%$$

➤ **Conclusion: absence of matrix effect with the combination of 1,3 propanediol (PDO) and ipratropium bromide.**

- Limit of detection and quantification

Limit of detection (LOD) and limit of quantification (LOQ) are two important concepts in analytical chemistry that relate to the sensitivity of an analytical method.

The limit of detection (LOD) is the lowest amount of an analyte in a sample that can be detected.

The limit of quantification (LOQ) is the lowest amount of an analyte in a sample that can be quantified.

These two parameters are based on the standard deviation of the response ( $\sigma$ ) and the slope ( $S$ ) of the calibration curve

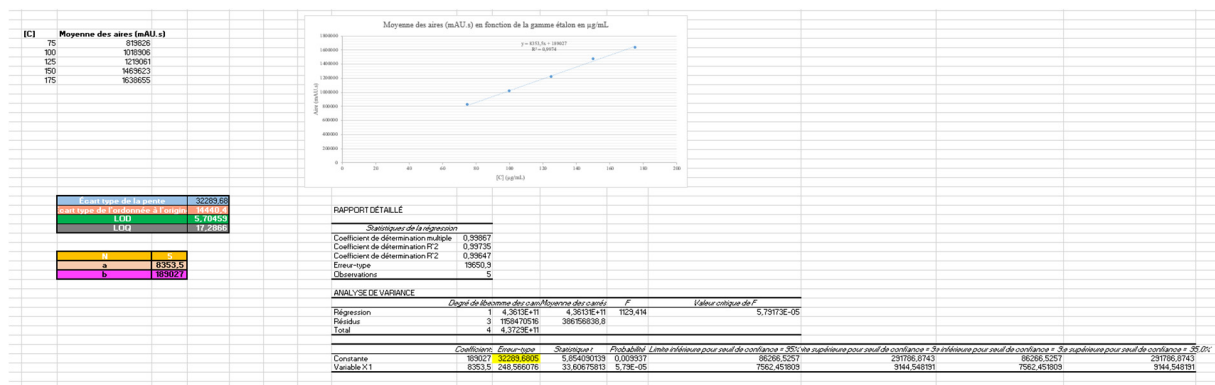
*LOD equation*

$$LOD = 3.3 \times \frac{\sigma}{S}$$

*LOQ equation*

$$LOQ = 10 \times \frac{\sigma}{S}$$

Where  $S$  is the slope of the calibration line,  $\sigma$  is the residual standard deviation of the regression line. The obtained values of: LOD and LOQ were 5.70 and 17.28 $\mu$ g/mL, respectively.



LOD	5.70 µg/mL
LOQ	17.28 µg/mL

- Repeatability

Repeatability refers to the degree of agreement between multiple measurements of the same sample, under the same conditions, using the same analytical method and equipment. In other words, it measures the precision of the analytical method.

For this specific reason, the quantification was carried out on two different solutions with the same concentration ([C] = 125 µg/mL).

N.B: the acquisition were launched during the same day, by the same operator and under similar experimental conditions.

Each solution was injected 6 times (as mentioned on the table below):

							Mean (µg/mL)
Solution 1	125.20	125.00	125.1	125.40	125.06	124.80	125.09
Solution 2	124.95	124.94	125.01	125.13	125.02	124.98	125.009

The given results make it possible to validate the repeatability parameter, as the obtained values are equal to 125 µg/mL.



- Recovery

It involves spiking a known amount of an analyte into a sample, and analyzing the sample to determine the amount of analyte that is recovered after the sample preparation.

As a matter of fact, the recovery is calculated by comparing the measured amount of analyte in the spiked sample to the amount that would be expected to be present based on the known amount of analyte that was added.

This parameter is useful for determining the accuracy and precision of a sample preparation method and can help to ensure that the results obtained from an analytical method are reliable and reproducible.

The theoretical dosing concentration of ipratropium bromide (250 µg/mL)

Group	Concentration theoretical (µg/mL )	Concentration found (µg/mL )	Recovery (%)
60%	75	73.312	97.7493333
	75	77.310	103.08
	75	76.918	102.557333
80%	100	100.503	100.503
	100	98.588	98.588
	100	98.974	98.974
100%	125	119.926	95.9408
	125	120.173	96.1384
	125	129.819	103.8552
120%	150	146.447	97.6313333
	150	156.55	104.366667
	150	156.905	104.603333
140%	175	173.54	99.02857143
	175	170.602	97.48685714
	175	180.1	102.9142857
<b>Moyenne% =100.22</b>			

*Table 2: Recovery validation parameter.*