

Review

## Beta-Adrenergic Agonists

Giovanni Barisione <sup>1,\*</sup>, Michele Baroffio <sup>2</sup>, Emanuele Crimi <sup>2</sup> and Vito Brusasco <sup>2</sup>

<sup>1</sup> Preventive and Occupational Medicine Unit, Respiratory Pathophysiology Laboratory, University Hospital San Martino, Largo R. Benzi, 10 - 16132 Genoa, Italy

<sup>2</sup> Respiratory Pathophysiology Unit, Department of Internal Medicine, University of Genoa, Viale Benedetto XV - 16132 Genoa, Italy; E-Mails: michele.baroffio@unige.it (M.B.); emanuele.crimi@unige.it (E.C.); vito.brusasco@unige.it (V.B.)

\* Author to whom correspondence should be addressed; E-Mail: giovanni.barisione@hsanmartino.it; Tel.: +39-010-5553367; Fax: +39-010-5556667.

Received: 2 February 2010; in revised form: 15 March 2010 / Accepted: 26 March 2010 /

Published: 30 March 2010

---

**Abstract:** Inhaled  $\beta_2$ -adrenoceptor ( $\beta_2$ -AR) agonists are considered essential bronchodilator drugs in the treatment of bronchial asthma, both as symptoms-relievers and, in combination with inhaled corticosteroids, as disease-controllers. In this article, we first review the basic mechanisms by which the  $\beta_2$ -adrenergic system contributes to the control of airway smooth muscle tone. Then, we go on describing the structural characteristics of  $\beta_2$ -AR and the molecular basis of G-protein-coupled receptor signaling and mechanisms of its desensitization/dysfunction. In particular, phosphorylation mediated by protein kinase A and  $\beta$ -adrenergic receptor kinase are examined in detail. Finally, we discuss the pivotal role of inhaled  $\beta_2$ -AR agonists in the treatment of asthma and the concerns about their safety that have been recently raised.

**Keywords:**  $\beta_2$ -adrenoceptors; G-protein-coupled receptor signaling; airway smooth muscle; bronchodilation; desensitization; safety issues

---

## 1. Introduction

In the past,  $\beta$ -adrenoceptor ( $\beta$ -AR) agonists have been utilized in many clinical settings. Nowadays, they are considered as first-line medications in the treatment of airway narrowing, the hallmark feature of bronchial asthma and chronic obstructive pulmonary disease (COPD). Epinephrine was the first bronchodilator used at the beginning of the past century. Ephedrine was introduced into Western medicine in 1924, although it had been used in China for thousands of years [1]. The next major advance in this field was the development in the 1940s of isoproterenol [2], a  $\beta$ -AR selective agonist void of  $\alpha$ -adrenergic activity. More recently, the development of selective  $\beta_2$ -AR agonists has made available drugs with reduced cardiovascular effects [3]. Therefore,  $\beta_2$ -AR agonists represent today a mainstay of the management of COPD [4] and asthma. In the latter condition, they are used both as symptoms-relievers and, in combination with inhaled corticosteroids, as disease-controllers [5,6].

The aim of this article is to review the molecular mechanisms by which  $\beta_2$ -AR contribute to control the tone of airway smooth muscle (ASM) and their implications for practical use of  $\beta_2$ -AR agonists in the treatment of asthma.

## 2. Adrenergic Control of ASM Tone

In humans, unlike in other species, there is no evidence that adrenergic nerves directly supply the ASM [7], the only inhibitory nervous system being the non-adrenergic, non-cholinergic one [8–10]. The sympathetic system affects ASM tone *via* circulating catecholamines [7–9] acting on the  $\beta_2$ -AR present on ASM cell membrane and on parasympathetic nerve endings [11,12].

### 2.1. Circulating Catecholamines

Epinephrine, nor-epinephrine, and dopamine are natural circulating compounds, but only the first has significant physiological effects. Following secretion from adrenal medulla and at plasma concentrations ranging from 0.2 to 0.4 nM/L, epinephrine provides a low-level stimulation of specific receptors [13], while spillover of nor-epinephrine from adjacent structures may also exert some minor effects. The action of epinephrine is rapidly (within about 2 min) terminated in various tissues as a consequence of oxidative deamination and methylation processes catalyzed by monoamine oxidase (MAO) and catechol-*O*-methyltransferase (COMT), respectively [14].

In asthmatic but not healthy subjects,  $\beta$ -AR antagonists ( $\beta$ -blockers) often elicit bronchoconstriction, which may be interpreted as suggesting a protective effect of  $\beta$ -AR stimulation against excessive airway narrowing. However, no evidence of plasmatic epinephrine was provided during acute exacerbations of asthma or airway narrowing induced by a variety of challenges, including intravenously administered propranolol. Therefore, it does not seem that severe bronchoconstriction and activation of ASM elicit a protective epinephrine release [15].

## 2.2. Neural Interactions

Complex interactions between various components of the autonomic nervous system have long been recognized in various species. This means that changes in the function of one neural pathway may affect other pathways. In humans, ASM tone is mainly maintained by acetylcholine (ACh) released from the parasympathetic nervous system [16]. An early study, based on isometric measurement of tension in bronchial rings, suggested that stimulation of  $\beta_2$ -AR could inhibit cholinergic neurotransmission [17]. As tyramine-induced release of nor-epinephrine from sympathetic nerves had no effect, this modulation was probably due to circulating epinephrine [17]. Therefore, apart from the relaxing effect on ASM,  $\beta_2$ -AR may have a modulatory effect on cholinergic neurotransmission [17,18] (*see below*). It has been proposed that an impairment of modulation of ACh release in asthmatic airways may contribute to bronchoconstriction induced by  $\beta$ -blockers [19].

## 3. Adrenergic Receptors

Ahlquist [2] proposed in 1948 that adrenergic receptors, stimulating a variety of physiological responses in various organs, could be classified into two primary types,  $\alpha$  and  $\beta$ . Stimulation of the former causes ASM contraction with the following rank of potency: epinephrine > nor-epinephrine > isoproterenol. By contrast, stimulation of  $\beta$ -AR, by the same agonists, relaxes ASM with a different rank of potency, *i.e.*, isoproterenol > epinephrine > nor-epinephrine.

As a result of studies conducted by Lands *et al.* [20],  $\beta$ -AR have been further classified into  $\beta_1$ - and  $\beta_2$ -subtypes. The former show an almost equal affinity for epinephrine and nor-epinephrine, the latter are considered to be more sensitive to epinephrine than nor-epinephrine. Autoradiographic mapping has demonstrated that  $\beta$ -AR are widely distributed in the lung and are present in several cell types, including ASM from trachea down to the terminal bronchioles [21]. The presence of  $\beta_1$ -AR in different species depends on the density of ASM adrenergic supply and the level of bronchial tree [22]. Consistent with the absence of sympathetic innervation to ASM in humans is the autoradiographic evidence of  $\beta_2$ -AR only in ASM at any airway generation [21]. The amount of  $\beta_2$ -messenger ribonucleic acid (mRNA) in ASM is high relative to the low receptor density, which may indicate a rapid turnover of  $\beta_2$ -AR and may account for the relative resistance to the development of tolerance [23].

Apart from ASM relaxation, other effects of  $\beta_2$ -AR activation have been reported, including increase in ciliary beat-frequency, changes in vascular permeability, decrease in ACh release, and modulation of immune cells function [21,23]. Whether these responses may contribute to the therapeutic efficacy of  $\beta_2$ -AR agonists in asthma treatment remains unclear. More recently, a third subtype (namely,  $\beta_3$ -AR) has been demonstrated in isolated canine [24] but not in humans [25] ASM. Its main action seems to be the enhancement of lipolysis in adipose tissue and thermogenesis (“brown” fat) in skeletal muscle [26].

Functional studies have shown that ASM relaxation, at the level of both central and peripheral human airways, is mediated solely by  $\beta_2$ -AR [27,28]. In asthmatics, selective stimulation of  $\beta_1$ -AR by prenalterol has no bronchodilator action [29]. Most importantly,  $\beta_2$ -AR agonists act as functional antagonists and inhibit or reverse contractile responses, irrespective of constrictor stimuli [30,31]. This is a property that is

of particular interest in asthma, where several physical or chemical spasmogens are likely to be involved. However, in COPD anticholinergics may produce equivalent or even greater bronchodilation than  $\beta_2$ -AR agonists because vagal tone is the major reversible element in such patients [32].

#### 4. G-Protein-Coupled Receptor Signaling

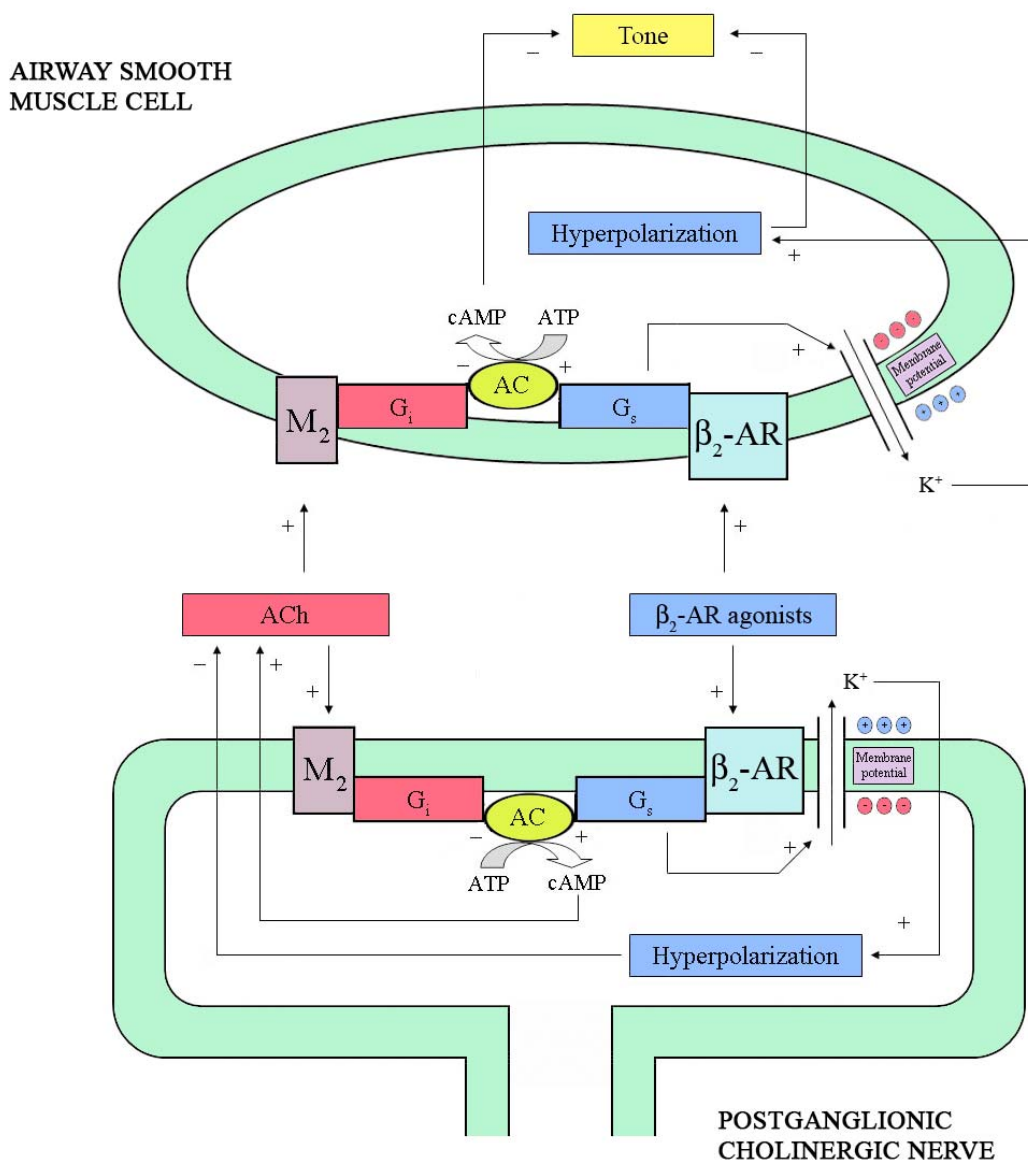
##### 4.1. ASM Relaxation

The  $\beta_2$ -AR belongs to the ubiquitously expressed 7-transmembrane receptors superfamily, which classically signals through heterotrimeric G-proteins [33,34]. They are commonly referred to as G-protein-coupled receptors because accomplish signal transduction to the interior of the cell *via* interactions with guanine nucleotide regulatory binding proteins [35]. The receptor-coupled G-proteins function as “molecular switches” alternating from an inactive guanosine-diphosphate to an active guanosine-triphosphate (GTP) state, which proceeds to regulate downstream cell processes [35]. Signaling *via* many hormones and neurotransmitters, as well as photons and odors, follows the same basic scheme, *i.e.*, by binding of an extracellular ligand to the receptor, which then interacts with the membrane-bound G-protein. This complex, often referred to as the ternary complex, then acts through the activated G-protein to modulate an effector, such as adenylyl cyclase, phospholipase C, or ion channels [35]. For more details on this argument, we refer the reader to the review by Dohlman [36].

Activated  $\beta_2$ -AR promotes the binding of a heterotrimeric stimulatory G-protein termed  $G_s$  (consisting of  $\alpha$ ,  $\beta$ , and  $\gamma$  subunits) to the receptors [37]. Upon binding of GTP, the  $G_s$ -protein subunits disassociate into  $G_\alpha$  and  $G_{\beta\gamma}$ . Traditionally,  $G_\alpha$  was considered to be the subunit that stimulates adenylyl cyclase (AC), but  $G_{\beta\gamma}$  can also activate certain pathways [37]. The resulting increase in intracellular cyclic 3',5'-adenosine monophosphate (cAMP) concentrations activates protein kinase (PKA), which phosphorylates several proteins causing relaxation [38]. The intrinsic GTPase activity of  $G_\alpha$  subsequently terminates the process, with re-association of heterotrimeric protein [37]. In bronchial smooth muscle, PKA inhibits both myosin-light-chain kinase [39] and phosphoinositide hydrolysis [40]. Moreover, it promotes  $Ca^{2+}/Na^+$  exchange [41], thus resulting in a decrease of intracellular  $[Ca^{2+}]$ , and stimulates  $Na^+/K^+$  ATPase [42]. However, these effects are only observed at relatively high concentrations of  $\beta_2$ -AR agonists, when maximal relaxation responses have been exceeded. In the 1990's, Jones *et al.* [43,44] showed that two potent inhibitors of the opening of the large-conductance  $Ca^{2+}$ -activated  $K^+$  (BKCa) channels, *i.e.*, charybdotoxin and iberiotoxin, may inhibit the bronchodilator responses to  $\beta_2$ -AR agonists and to other agents that can increase cAMP. These effects have been subsequently confirmed by other studies [45,46] and are observed at low concentrations of  $\beta_2$ -AR agonists in isolated human airways [47]. This suggest that  $\beta_2$ -AR may activate BKCa channels in ASM directly *via* the  $\alpha$ -subunit of  $G_s$  [48] (Figure 1).

Studies, based on direct measurement of ACh release from guinea pig [49,50] and horse [51] trachealis, have shown that stimulation of  $\beta_2$ -AR enhances cholinergic neurotransmission in the absence, but not in the presence, of epithelium [52]. Consistent with these results, direct activation of the  $\beta$ -AR-coupled  $G_s$  subunit by cholera toxin, which increases the activity of AC [33], caused an increase of ACh release in epithelium-denuded guinea pig trachealis [49].

**Figure 1.** Pre- and post-junctional intracellular mechanisms modulating cholinergic neurotransmission and airway smooth muscle (ASM) cell tone. At pre-junctional level, stimulation of  $\beta_2$ -adrenoceptor ( $\beta_2$ -AR) by agonists opens  $\text{Ca}^{2+}$ -activated  $\text{K}^+$  (BKCa) channels leading to cell membrane hyperpolarization and reduction of acetylcholine (ACh) release. By contrast, direct activation of adenylyl cyclase (AC) enhances ACh release. In the ASM cell, stimulation of  $\beta_2$ -AR as well as direct stimulation of AC, opens BKCa channels determining cell membrane hyperpolarization and relaxation. The ACh released by postganglionic cholinergic nerves binds  $\text{M}_2$ -muscarinic receptors expressed both at pre- and post-junctional level, thus inhibiting ACh release and increasing ASM cell tone. cAMP: cyclic 3',5'-adenosine monophosphate; ATP: adenosine triphosphate;  $\text{G}_s$  and  $\text{G}_i$ : stimulatory and inhibitory subunits of the receptor-coupled G-protein, respectively.



Moreover, direct stimulation of AC by forskolin [53] or incubation with a cAMP analog, increased ACh release [49]. In ASM cells, however, stimulation of G<sub>s</sub> protein directly opens BKCa channels [48], which has been found to decrease ACh release in guinea pig trachealis [54]. By simultaneously measuring electrically induced ACh release and force development, Bricchetto *et al.* [55] from our laboratory have recently shown that  $\beta_2$ -AR agonists attenuates cholinergic neurotransmission in bovine trachealis by a mechanism involving BKCa channels. Therefore, it can be hypothesized that  $\beta_2$ -AR agonists have the potential to increase or decrease ACh release depending on whether they mainly act through AC or BKCa channels.

Previous studies have shown that  $\beta_2$ -AR can activate PKA-independent mechanisms to elicit functional responses [45,46]. Freund-Michel *et al.* [56], have recently found that the anti-tussive properties of terbutaline were abolished by an inhibitor of protein kinase G (PKG) present in vagal sensory fibres but were unaffected by a PKA inhibitor. As the level of cyclic 3',5'-guanosine monophosphate was not elevated in the vagus nerve, these data suggest that a cAMP-dependent cross-activation of PKG and, subsequently, the opening of the BKCa channels may account for the anti-tussive properties of  $\beta_2$ -AR agonists. If bronchodilator and anti-tussive activity of  $\beta_2$ -AR agonists is mediated by different signaling pathways, then some discrepancies, reported in the clinical literature [57–60] regarding their anti-tussive and bronchodilator efficacies, could be explained.

#### 4.2. Anti-Inflammatory Effects

It has been proposed that actions other than ASM relaxation may play a role in the protective effect of long-acting  $\beta_2$ -AR agonists against allergen-induced late asthmatic reactions [61]. In diseases such as asthma, ASM cells play a synthetic role by secreting inflammatory mediators [62]. In other words, ASM may produce multiple inflammatory mediators (prostanoids, cyto- and chemokines), thus being able to perpetuate and amplify the inflammatory process within the airway wall. Evidence is rapidly accumulating that the asthmatic ASM is abnormal, in that it proliferates faster [63,64], produces more chemokines and cytokines as well as a different profile of extracellular matrix proteins than in its non-asthmatic counterpart [62]. These abnormalities may stem from altered calcium homeostasis leading to increased mitochondrial biogenesis and/or decreased levels of key transcription factors, such as CCAAT enhancer binding protein- $\alpha$  [65]. Recent data indicate that both short- and long-acting  $\beta_2$ -AR agonists are able to reduce the expression of surface intercellular adhesion molecule-1 and the release of granulocyte-macrophage colony-stimulating factor evoked by interleukin-1 $\beta$  in ASM [66]. Interestingly, the repression of these inflammatory indexes was prevented by adenoviral over-expression of PKI- $\alpha$ , a highly selective PKA inhibitor. These data indicate a PKA-dependent mechanism of repression and suggest that agents that elevate intracellular cAMP, and thereby activate PKA, may have an anti-inflammatory effect in ASM cells [66].

### 5. $\beta_2$ -AR Dysfunction

When a cell or tissue is first exposed to a  $\beta_2$ -AR agonist, there is a brisk initial response in the production of cAMP, followed by a decline to nearly basal level despite the continued presence of the drug. The cell

becomes “desensitized” or “tolerant” to further stimulation. Desensitization depends on both the concentration of  $\beta_2$ -AR agonist and time of exposure. In any cell type, the process of desensitization occurs through an acute and a chronic phase [67].

Over the years, there has been some confusion regarding the terminology used to describe receptors dysfunction. More precisely, *desensitization* is defined as a waning of a functional response despite the continuous presence of a stimulus of constant intensity [37]. Typically, a decrease in receptor function over time during exposure of cells or tissues to agonist is one form of desensitization. For  $\beta_2$ -AR, this would entail measurements of cAMP or AC activities. In intact animal models and in clinical setting, agonist-promoted desensitization is referred to as “tachyphylaxis” or “refractoriness” [37].

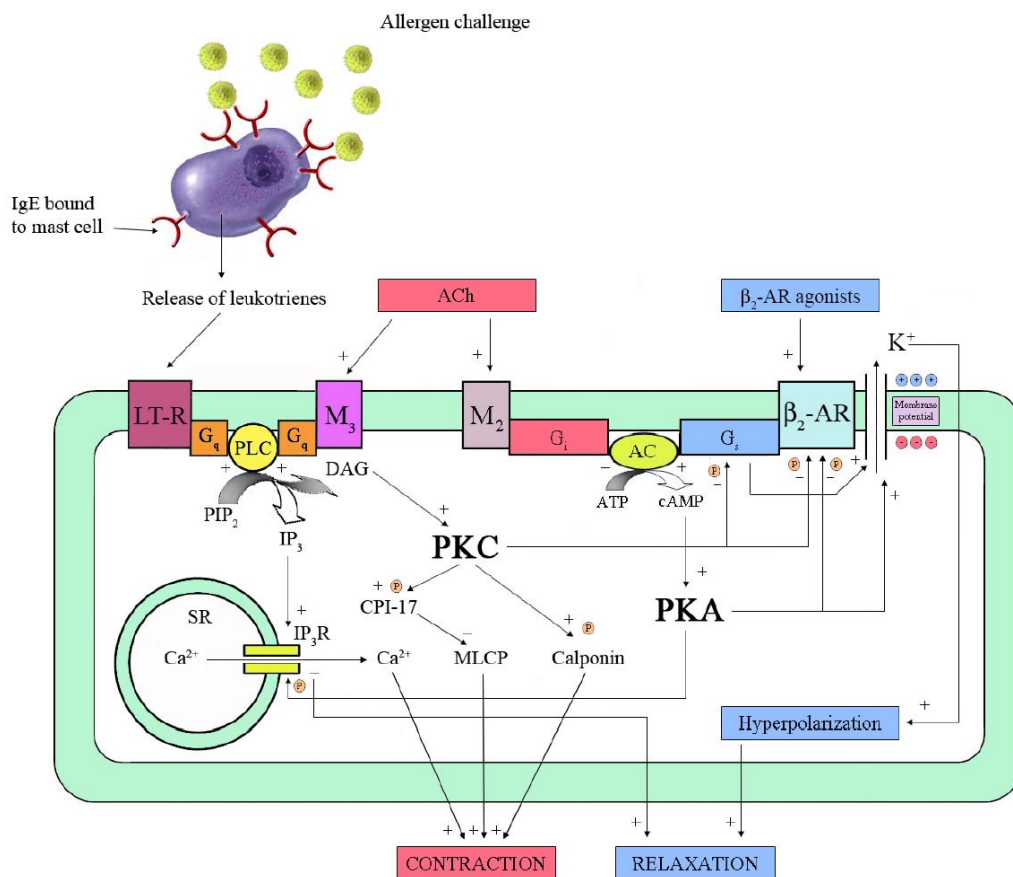
There is both *in vivo* and *in vitro* evidence that  $\beta_2$ -AR are less responsive in asthmatic than healthy individuals [68,69]. It is not completely clear, however, whether this decreased responsiveness is the cause of asthma [70] or a byproduct of therapy with  $\beta_2$ -AR agonists [68]. Trian *et al.* [71] have recently shown that epithelial infection with rhinovirus reduces  $\beta_2$ -AR function in ASM. This may explain the clinical observation of  $\beta_2$ -AR dysfunction during virus-induced asthma exacerbations. Some authors reported that the inhibitory effects of interleukin-1 $\beta$  on the isoproterenol-induced bronchodilation are abolished by COX-2 inhibitors [72]. Thus, it appears more likely that  $\beta_2$ -AR hyporesponsiveness is a consequence of the disease process rather than its cause.

### 5.1. Desensitization by Phosphorylation

The covalent modification of  $\beta$ -AR represents the earliest event in the desensitization process [37]. Over a time frame of less than 30 min of exposure to an agonist *in vitro*, phosphorylation of  $\beta$ -AR takes place by PKA and  $\beta$ -AR kinase ( $\beta$ ARK). The latter belongs to G-protein-coupled receptor kinases family (GRK) [73]. Both kinases cause a complete or partial loss of functional coupling between receptor and  $G_s$  subunit by phosphorylating specific sites [67]. The uncoupling between receptor and  $G_s$  subunit may be a mechanism of allergen-induced  $\beta_2$ -AR dysfunction observed in human isolated passively sensitized bronchi [73] (Figure 2).

PKA is a cAMP-dependent enzyme which is activated by low concentrations of  $\beta_2$ -AR agonist, such as circulating catecholamines or drugs. Any process leading to an increase in intracellular concentrations of cAMP sufficient to activate PKA will result in phosphorylation of  $\beta_2$ -AR, even if unoccupied by agonist [37]. This underlies one of the principal mechanisms of *heterologous desensitization* of the  $\beta_2$ -AR, so termed because dysfunction can be evoked by processes distinct from activation of the receptor as, for instance, those elicited by inflammatory mediators [67]. Data from our laboratory showed, in passively sensitized isolated human bronchial rings, that allergen challenge caused  $\beta_2$ -AR dysfunction without involving cytokines [75] or mechanisms distal to cAMP formation [76]. Moreover, this dysfunction was prevented by a LTD<sub>4</sub>-receptor antagonist or a cell membrane stabilizer, but not by cetirizine or indomethacin, suggesting a central role for leukotrienes released from resident inflammatory cells [77]. The involvement of PKC in this process was subsequently shown on both cultured ASM cell and bronchial rings [78].

**Figure 2.** Mechanisms of relaxation and  $\beta_2$ -adrenoceptor ( $\beta_2$ -AR) desensitization in airway smooth muscle (ASM). The binding of a specific agonist to  $\beta_2$ -AR stimulates the receptor-coupled  $G_s$ -protein, which activates adenylyl cyclase (AC). The resulting increase of cyclic 3',5'-adenosine monophosphate (cAMP) activates protein kinase A (PKA), which phosphorylates inositol 1,4,5-trisphosphate receptor ( $IP_3R$ ) of sarcoplasmic reticulum (SR) and opens  $Ca^{2+}$ -activated  $K^+$  (BKCa) channels, thus leading to relaxation. However, activated PKA phosphorylates  $\beta_2$ -AR, uncoupling it from  $G_s$ -protein. Exposure of sensitized mast-cells to allergen causes release of leukotrienes (LT). Their interaction with the specific receptor, *i.e.*, LT-R, activates a  $G_q$ -protein, which increases phospholipase C (PLC) activity. PLC catalyzes the hydrolysis of phosphatidylinositol 4,5-bisphosphate ( $PIP_2$ ), which produces  $IP_3$  and diacylglycerol (DAG).  $IP_3$  leads to contraction by increasing release of  $Ca^{2+}$  from SR while DAG activates protein kinase C (PKC). The latter phosphorylates several substrates like calponin and CPI-17, which is an inhibitor of myosin-light-chain phosphatase (MLCP). In addition, PKC phosphorylates both  $\beta_2$ -AR and  $G_s$ -protein. A similar cascade of events seems to occur in response to acetylcholine (ACh) released by cholinergic nerves through  $M_3$ -muscarinic receptors. In addition, activation of  $M_2$ -muscarinic receptors inhibits AC, thus decreasing cAMP level and PKA activity. ATP: adenosine triphosphate; **P** : phosphorylation sites.





If cysteinyl-leukotrienes, released from resident or circulating inflammatory cells or even from the ASM itself, are responsible for  $\beta_2$ -AR desensitization in asthma, then the concurrent administration of cysteinyl-LT<sub>1</sub>-R antagonists might represent a useful strategy to improve the response to  $\beta_2$ -AR agonists in this disease. On the other hand, *homologous desensitization* is mediated by  $\beta$ ARK. It describes processes that are specific to an attenuation of function of activated, *i.e.*, occupied, receptor [79].  $\beta$ ARK requires high ( $\mu$ M/mL) concentrations of the agonist [73]. Other data suggest that a cofactor is required for  $\beta$ ARK function. A protein, inhibiting the signaling function of  $\beta$ ARK-phosphorylated receptors by more than 75%, was found [80] and termed  $\beta$ -arrestin [81]. It is now recognized that  $\beta$ -arrestins act as versatile adapters controlling receptor signaling, desensitization, and trafficking. Most endogenous receptors appear to signal in a balanced fashion using both  $\beta$ -arrestin and G-protein-mediated pathways. Some 7-transmembrane receptors are thought to be non-signaling "decoys" because of their inability to activate typical G-protein signaling pathways. It has been proposed that these receptors act to scavenge ligands or function as co-receptors [82].

Nino *et al.* [83] have recently examined the regulation and role of phosphodiesterase-4 (PDE<sub>4</sub>) activity in mediating the pro-asthmatic changes in ASM responsiveness that accompany its prolonged homologous desensitization. Their findings provide new evidence that prolonged exposure of ASM to  $\beta_2$ -AR agonists results in an upregulated expression of PDE<sub>4</sub>D5 isoform due to PKA-dependent activation of G<sub>i</sub> protein signaling. This is consistent with a potential switch in coupling of the  $\beta_2$ -AR from G<sub>s</sub> to G<sub>i</sub> resulting in G <sub>$\beta\gamma$</sub> -subunit-mediated activation of ERK1/2 which, in turn, leads to a phosphorylating cascade of transcription factors.

During exposure to low levels of agonist, few  $\beta_2$ -AR on the cell are occupied [67]. Such low ( $\leq 10$  nM/mL) concentrations of agonist are present in systemic circulation during exercise-induced activation of the sympathoadrenal system and in stress, such as under haemodynamic instability or respiratory insufficiency. Similar concentrations are also obtained during systemic administration of agonist, such as terbutaline, for the treatment of asthma and various catecholamines for the treatment of hypotension and heart failure [67]. Although only a  $\beta_2$ -AR fraction is occupied, this is sufficient to increase intracellular cAMP, resulting in activation of PKA and phosphorylation of all  $\beta_2$ -AR on the cell. Under these conditions, only the agonist-occupied receptors are phosphorylated by  $\beta$ ARK, which contributes minimally to the process under these circumstances. Other G-protein-coupled receptors with PKA sites may also be phosphorylated, resulting in desensitization by  $\beta_2$ -AR agonists [37].

During exposure to high concentrations of  $\beta_2$ -AR agonists, as occurring in bronchodilator treatment of asthmatic patients, the majority of  $\beta_2$ -AR on the cell may be occupied [67]. Under these circumstances, PKA is activated, as with low agonist exposure, and all  $\beta_2$ -AR are phosphorylated by this kinase. In addition, all occupied receptors are phosphorylated by  $\beta$ ARK. Such phosphorylated receptor binds  $\beta$ -arrestin with subsequent partial uncoupling from G<sub>s</sub> [37].

Phosphorylation by PKA and GRKs leads to desensitization of  $\beta_2$ -AR signaling, and is thought to be a mechanism involved with cell and organ homeostasis and tolerance to agonists. However, there is little direct evidence that these events are relevant to  $\beta_2$ -AR physiological function, such as ASM relaxation. Recently, Wang *et al.* [84] have shed some light on the matter by using a transgenic mouse model

expressing the human wild-type and mutated  $\beta_2$ -AR lacking PKA and/or GRK phosphorylation sites on ASM. They showed that, at low receptor occupancy,  $\beta_2$ -PKA(-) had enhanced agonist-promoted relaxation, while  $\beta_2$ -GRK(-) airways were unaffected. In contrast, at saturating agonist concentrations, the greatest relaxation enhancement was with  $\beta_2$ -GRK(-), with no evidence for additivity when PKA sites were also removed. For the full range of responses, the  $\beta_2$ -PKA(-)/GRK(-) airways had the greatest relaxation efficiency, indicating a graded effect of GRKs as agonist concentration increased. Thus, these two mechanisms impact a relevant  $\beta_2$ -AR physiologic function, by acting as attenuators of the acute response, and represent specific interfaces where adjunct therapy or biased ligands may improve  $\beta_2$ -AR agonist treatment of obstructive lung disease.

### 5.2. Desensitization by Sequestration

After about 30 min of agonist exposure, cell surface  $\beta_2$ -AR undergo a process leading to internalization or sequestration of the receptor to a sub-cellular compartment. The proportion of receptors at the surface that undergo sequestration appears to be highly cell-type dependent. In cells where there is little receptor reserve and sequestration results in ~60% loss of cell surface receptor, desensitization by this mechanism would be expected, as shown in lymphocytes [37]. Recent data indicate that the  $\beta_2$ -AR of human pro-inflammatory cells is exquisitely sensitive to desensitization, whereas  $\beta_2$ -AR-mediated relaxation of human ASM is relatively resistant. An explanation for this discrepancy is that a large  $\beta_2$ -AR reserve exists in ASM cells for sympathomimetic bronchodilators, which protects against desensitization [85]. After agonist removal, sequestered receptors, which have not been shuttled to a degradation pathway, can recycle to the cell surface, with a time course that is about the same as for initiation of the process. Some data suggest that the pool of sequestered  $\beta_2$ -AR undergoes phosphatases-mediated dephosphorylation [86].

### 5.3. Desensitization by Down-Regulation

After 3–6 h of exposure to agonist, a net loss of receptors occurs independent of compartment [37]. This is termed *down-regulation* and reaches a steady state at 18–24 h. The process can result in an loss of as much as 90% of receptors and is presumed to be a major component in long-term agonist-promoted desensitization of  $\beta_2$ -AR [37]. The mechanisms responsible are both a decrease in receptor production and an increase in receptor protein degradation. The decrease in production of new receptors in the presence of agonist is primarily due to a loss of  $\beta_2$ -AR mRNA stability [87], although a change in the rate of transcription may also play a role [88]. Destabilization of  $\beta_2$ -AR mRNA appears to be due to cAMP-promoted binding of a protein to a destabilization sequence (AUUUA) in the 3' non-coding region of the gene [89]. The second mechanism of down-regulation may be related to the degradation of sequestered  $\beta_2$ -AR pool. One study suggests that PKA-mediated phosphorylation is required for full agonist-promoted down-regulation [90]. Also, tyrosine residues in the cytoplasmic tail have been implicated as necessary for agonist-promoted down-regulation [91].

#### 5.4. The $\beta$ -AR Paradox

Long-acting  $\beta_2$ -AR agonists have been introduced as a therapeutic option in asthma [5,6] to improve symptom control. However, there is growing concern about whether control of disease may deteriorate with the regular use of these agents. Several studies have shown a temporary increase in airway responsiveness to histamine upon withdrawal of inhaled  $\beta_2$ -AR agonists [92–94]. In an 8-week study, Cheung *et al.* [95] showed that the bronchodilator effect of salmeterol was maintained, whereas its protective effect against methacholine (MCh)-induced bronchoconstriction was significantly attenuated. This dissociation between bronchodilator and bronchoprotective effect of long-acting  $\beta_2$ -AR agonists was further explored in mice with ablated  $\beta$ -AR genes ( $\beta$ -AR<sup>-/-</sup>) and transgenic mice overexpressing  $\beta_2$ -AR ( $\beta_2$ -AR-OE) [96]. Unexpectedly,  $\beta$ -AR<sup>-/-</sup> mice had markedly decreased bronchoconstrictive responses to MCh and other G<sub>q</sub>-coupled receptor agonists such as thromboxane-A<sub>2</sub> and 5-hydroxytryptamine (5-HT). In contrast,  $\beta_2$ -AR-OE mice had enhanced constrictive responses. Inositol phosphate accumulation by G<sub>q</sub>-coupled M<sub>3</sub>-muscarinic, thromboxane-A<sub>2</sub>, and 5-HT<sub>2</sub> receptors was desensitized in ASM cells from  $\beta$ -AR<sup>-/-</sup> mice and sensitized in cells from  $\beta_2$ -AR-OE mice. Thus,  $\beta$ -AR seem to regulate antithetically constrictive signals, affecting bronchomotor tone/reactivity by additional effects other than direct bronchodilation. Studies of signaling elements in these pathways identified the nodal point of this cross-talk in phospholipase C- $\beta$ 1, whose expression was altered by  $\beta$ -AR in a direction and magnitude consistent with the physiologic and cellular responses. These results establish a possible mechanism of *the  $\beta$ -AR paradox* and identify a potential asthma modifier gene (phospholipase C- $\beta$ 1), which may also be a therapeutic target in asthma when chronic  $\beta$ -AR agonists are required [96]. An alternative explanation is that regular use of  $\beta_2$ -AR agonists may negatively impact the balance of factors contributing to airway inflammation and remodeling in asthma [97].

Two elegant studies [98,99], in ovalbumin-driven murine model of asthma, have shown that  $\beta$ -AR agonists and antagonist, with inverse agonist properties, may exert reciprocating effects on cellular signaling dependent on duration of administration. In particular, acute dosing with  $\beta$ -blockers produced bronchoconstriction, whereas the chronic administration of the same agents did exert a protective effect against MCh-induced airway narrowing [98]. Surprisingly, the bronchoprotective effect of  $\beta$ -blockers was associated with reduced inflammation and mucous metaplasia, up-regulation of  $\beta_2$ -AR, and reduced expression of various spasmogenic proteins [99]. These findings in mice led to the first proof-of-concept open-label study in ten patients with mild steroid-naïve asthma who were given incremental doses of nadolol for nine weeks [100]. In eight out of the 10 subjects tested, chronic treatment with this  $\beta$ -blocker produced a significant, dose-dependent, decrease in airway responsiveness to MCh. However, these findings warrant further testing in future by means of larger trials [101,102].

#### 5.5. Reversal of Desensitization

As detailed by Davies and Lefkowitz [103], corticosteroids have been shown to increase  $\beta_2$ -AR synthesis, reverse down-regulation, and improve coupling, the last by partially restoring the fraction of receptors in the high-affinity state. A study from our laboratory indicates that short-term (3 h) exposure to

beclomethasone dipropionate ablated  $\beta_2$ -AR dysfunction induced by allergen challenge in passively sensitized human bronchi. This effect was mediated by an increase in the activity of the  $\alpha$ -subunit of  $G_s$  protein, suggesting an action independent of gene transcription [104].

## 6. Genetic Polymorphisms of the $\beta_2$ -AR

A series of studies revealed that the gene encoding for  $\beta_2$ -AR is polymorphic within the human population [105,106]. Such polymorphisms may be responsible for heterogeneity in the response to  $\beta_2$ -AR agonists and/or receptor dysfunction in asthma [107]. For example, the B16 Arg/Arg homozygosity is associated with more frequent exacerbations during chronic daily dosing of salbutamol [108] or salmeterol [109]. In contrast, it seems that there is no genotypic risk for exacerbations in patients using inhaled  $\beta_2$ -AR agonists less than once a day [109]. In asthma patients with B16 Arg/Arg and B16 Gly/Gly genotypes, combination treatment with salmeterol and inhaled corticosteroid improved airway function when compared with inhaled corticosteroid therapy alone. These findings suggest that patients should continue to be treated with long-acting  $\beta_2$ -AR agonists plus moderate-dose inhaled corticosteroids irrespective of B16 genotype [110].

$\beta_2$ -AR function undergoes desensitization during persistent agonist exposure because of receptor phosphorylation by GRKs, which was found to be highly expressed in ASM [111]. The coding region is polymorphic at codon 41, where glutamine (Gln) can be substituted by leucine (Leu) (minor allele), but almost exclusively in those of African descent. So, GRK5-Leu41 represents a gain-of-function polymorphism that evokes enhanced loss-of-function of  $\beta_2$ -AR during persistent agonist exposure, thus possibly contributing to  $\beta_2$ -AR agonist variability in asthma treatment of African-Americans [111].

If one considers  $\beta_2$ -AR expression as being constantly regulated by systemic catecholamines or continuously administered  $\beta_2$ -AR agonists, then the glutamic (Glu)27 allele may be expressed to a higher level in the lung, as it is refractory to down-regulation. More than a decade ago, a study by Hall *et al.* [112] addressed this point and found that asthmatics with the Glu27 polymorphism had approximately fourfold lower airway responsiveness to MCh as compared to those with Gln27 (wild type). Recently, a novel gene (arginine1) associated with acute response to inhaled  $\beta_2$ -AR agonists in both children and adults with asthma was identified [113].

## 7. Role of $\beta_2$ -AR Agonists in the Treatment of Obstructive Lung Diseases

### 7.1. Symptom-Relief

#### 7.1.1. Asthma

Short-acting  $\beta_2$ -AR agonists administered by inhalation are the most effective therapy for rapid reversal of airflow obstruction and prompt relief of asthmatic symptoms. The most widely used are salbutamol (commonly known as albuterol in the US) and terbutaline; their action occurs in 5 min or less, peaks after 30 to 60 min and lasts 4 to 6 h [114]. With regular use of a bronchodilator (four or more times daily), the potency (as measured by the increase in one second forced expiratory volume, FEV<sub>1</sub>) does not decline, but

the duration of action is slightly shortened [115]. A regular schedule of administration four times a day does not improve outcomes as compared with as-needed administration [116]. Moreover, in patients with certain genotypic variants of the  $\beta_2$ -AR it may have a deleterious effect [117,118]. So, the short-acting  $\beta_2$ -AR agonists are recommended for use on request to relieve symptoms or before anticipated exposure to known asthmatic triggers, especially exercise. In mild-to-moderate asthmatic patients, a cumulative bronchodilator effect of salbutamol can be seen up to inhaled doses ranging from 700 to 1,500  $\mu\text{g}$  [119]. Dose-dependent sympathomimetic-type side effects, including tremor, anxiety, heart pounding, and tachycardia (but not hypertension), are common, and a small dose-dependent decrease in serum potassium and magnesium levels is detectable. However, at the usual dose, adverse effects are uncommon [14].

The decision about which of the various short-acting  $\beta_2$ -AR agonists to use is based largely on cost and patient's or physician's preference. Salbutamol has been made available in a metered-dose inhaler free of chlorofluorocarbons (CFCs), and CFC-containing salbutamol inhalers were taken off the market on 31 December 2008. Like CFCs, the alternative propellant, hydrofluoroalkane (HFA), is inert in the human airway [120]. The HFA inhalers are equipotent to the CFC-propelled inhalers [121] and can be used with valved holding chambers (spacers) in patients with poor inhalational technique. They provide bronchodilation comparable to nebulized salbutamol when a sufficient number of puffs is administered and inhalational technique is good [122]. Levalbuterol, the purified D-rotatory isomer of salbutamol, was developed on the purpose to eliminate side effects, which some argue are limited to the S-rotatory isomer [123]. However, when delivered by metered-dose inhaler, levalbuterol has an efficacy and side-effect profile that is indistinguishable from that of the racemic mixture of molecules in salbutamol [124].

The inhaled long-acting  $\beta_2$ -AR agonists salmeterol and formoterol have a bronchodilator potency similar to that of their short-acting counterpart. Features distinguishing the two long-acting  $\beta_2$ -AR agonists are both practical and theoretical. The onset of action of formoterol occurs within 5 min, a period similar to that for short-acting  $\beta_2$ -AR agonists, whereas salmeterol has a slower onset of action (15 to 20 min) [14]. For this reason, a combination formoterol-corticosteroid inhaler is recommended both for quick relief of asthmatic symptoms and, when used regularly, for long-term control [125]. Formoterol is a full agonist in its action at the  $\beta_2$ -AR, whereas salmeterol is a partial agonist (and partial antagonist) [14]. The implication of this pharmacologic distinction, particularly as it might apply to the risk of fatal asthmatic attacks, is uncertain. Both salmeterol and formoterol display a sustained activity (>12 h), and because of their higher degree of  $\beta_2$ -AR specificity, have fewer side effects than short-acting  $\beta_2$ -AR agonists [126]. As much as with short-acting  $\beta_2$ -AR agonists, regular use of long-acting  $\beta_2$ -AR agonists results in only mild tachyphylaxis to the maximal bronchodilator effect and the duration of action [126]. In contrast, the bronchoprotective effect of long-acting  $\beta_2$ -AR agonists (*i.e.*, their inhibition of exercise-induced bronchoconstriction) rapidly wanes with regular use [127], a contrary pharmacologic effect that has not been fully explained. With rare exceptions [128], the quick symptom relief provided by short-acting  $\beta_2$ -AR agonists is not impeded by regular use of long-acting  $\beta_2$ -AR agonists [129].

### 7.1.2. COPD

In COPD, formoterol and salmeterol are more effective bronchodilators than their short-acting counterpart [130] and have an add-on effect to anticholinergics [131]. Both drugs improve FEV<sub>1</sub>, reduce symptoms and use of rescue medication, and improve exercise capacity and health status [132]. There is also evidence that formoterol may be used as a rescue inhaler without significant unwanted effects [133]. Though tolerance in the long-term use of long-acting  $\beta_2$ -AR agonists does not appear to be a problem in clinical practice, concerns have been raised about the side effects, particularly cardiac dysrhythmias, in susceptible elderly populations [132]. Nevertheless, salmeterol and formoterol at usual doses (100  $\mu$ g/day and 24  $\mu$ g/day, respectively) are effective and safe in treating patients with COPD [134]. Higher doses may cause more untoward effects, although serious adverse events are very uncommon [132].

### 7.2. Disease-Control

The fact that long-acting  $\beta_2$ -AR agonists afford sustained improvement in lung function may tempt clinicians to use them as a long-term controller medication, without concomitant use of anti-inflammatory treatment. However, this strategy results in unsuppressed airway inflammation and an unacceptably high rate of asthmatic exacerbations [135–139]. For these reasons, the use of long acting  $\beta_2$ -AR agonists as single treatment for asthma control is not recommended and several formulations combining  $\beta_2$ -AR agonist with an inhaled corticosteroid in a single device have been made available. Combinations of salmeterol with inhaled corticosteroid are generally used for regular treatment only, whereas those of salbutamol or formoterol have been suggested for both as needed and regular treatment [140].

## 8. The Great $\beta_2$ -AR Agonists Controversy

### 8.1. Short-Acting $\beta_2$ -AR Agonists

Epinephrine was the first synthetic  $\beta$ -agonist bronchodilator to be used clinically and was introduced in the early 1900's [141]. As systemic side effects due to potent  $\alpha$ - and non-selective  $\beta_2$ -AR stimulatory effects were reduced but not eliminated by aerosol inhalation, its widespread use for the treatment of asthma was abandoned. Moreover, up to a five-fold increase in mortality among users of inhaled epinephrine was reported in 1948 [142].

Isoproterenol was introduced in 1940 into medicine as short-acting  $\beta_2$ -AR agonist. It gradually replaced epinephrine because of its more potent  $\beta$ -stimulant properties and its virtual absence of  $\alpha$ -agonist activity [143]. However, this epinephrine derivative possesses the same major disadvantage of its parent compound, namely, a relatively short duration of action (due to rapid metabolism mainly by COMT and, to a lesser extent, by MAO) and undesirable cardiac side effects and mortality [143].

The most-often-cited examples of the possible relationship between short-acting  $\beta_2$ -AR agonists and mortality in asthma were the startling rise in asthma deaths in several countries, including the UK, Australia and New Zealand during the early 1960s [144] and in New Zealand in the late

1970s [145,146]. In investigating the earlier epidemic in the UK, Speizer *et al.* [144,147] found that these deaths were most often sudden, unexpected and closely correlated with the increased use of pressurized metered dose inhalers. In the late 1960s, following recognition of the potential danger from the overuse of  $\beta_2$ -AR agonists and restrictions on their over-the-counter sale, the usage of inhaled short-acting  $\beta_2$ -AR agonists declined. Simultaneously, the asthma death rate fell sharply, suggesting that the previously noted relationship might have been a causal one. It is noteworthy, however, that this decline in the asthma death rate occurred concomitantly with a greater reliance on corticosteroid therapy, suggesting that the latter might also have contributed to the drop in mortality due to asthma [148].

Several hypotheses have been advanced incriminating short-acting  $\beta_2$ -AR agonists as contributing to the second epidemic of asthma deaths that occurred in New Zealand in the late 1970s and early 1980s [146]. One suggested factor was a change in prescribing practices, in that inhaled  $\beta_2$ -AR agonists, along with sustained-release theophylline preparations, were prescribed in place of inhaled corticosteroids and cromolyn [149], possibly predisposing to fatal cardiac arrhythmias. Another proposed factor was an increased use of  $\beta_2$ -AR agonists delivered by air-driven home nebulizers for self-treatment of severe asthma. In this case, patients could be exposed to the risk of hypoxic cardiac arrest if large doses of short-acting  $\beta_2$ -AR agonists were delivered in the face of uncorrected hypoxaemia [150]. Two case-control studies involving asthma deaths in patients aged 5–45 years in New Zealand from 1981 to 1983 [151] and from 1977 to 1981 [152] have led to a particularly contentious hypothesis linking some of the excess in asthma mortality to prescribed fenoterol by metered-dose inhaler. In 1992, Spitzer *et al.* [153], by using health insurance data from Saskatchewan (Canada), found an increased risk of death or near-death from asthma in association with regular use of short acting  $\beta_2$ -AR agonists. These last included fenoterol (odds ratio, 5.4 per canister), but also salbutamol (odds ratio, 2.4 per canister). The authors concluded that these findings could be due to adverse effects of the  $\beta_2$ -AR agonists themselves but raised the alternative possibility that their increased use might simply be a marker of more severe disease. A 1 year clinical trial showed increased airway responsiveness and worsened asthma control during regular treatment with fenoterol added to usual therapy compared with short-acting  $\beta_2$ -AR agonists used only as needed for symptom relief [154]. However, subsequent US and UK trials of regular *versus* as-needed salbutamol did not detect sustained adverse effects on asthma control [116,155]. Nevertheless, in 1997 the British Guidelines on Asthma Management increasingly advocated the use of short-acting  $\beta_2$ -AR only as needed for symptom relief [156].

## 8.2. Long-Acting $\beta_2$ -AR Agonists

The introduction of long-acting  $\beta_2$ -AR agonists salmeterol [157] and formoterol [158] in the 1990's was considered a major advance in bronchodilator therapy with evidence that their use led to improved lung function and quality of life [114]. There were also potential safety advantages due to the twice daily, fixed dose usage, which reduced the risk of overuse of  $\beta_2$ -AR agonist therapy in the situation of severe exacerbations. However, concerns about the possible risks associated with long-acting  $\beta_2$ -AR agonists therapy were raised soon after their introduction into clinical practice. This was due to the evidence that their

regular use had the potential to reduce bronchodilator sensitivity to  $\beta_2$ -AR agonists [159,160] and could induce tolerance to their bronchoprotective effects [95], which may not be restored by concurrent use of inhaled corticosteroids [161]. It also became apparent that patients using long-acting  $\beta_2$ -AR agonists may be at risk of severe exacerbations if the symptom control, achieved with long-acting  $\beta_2$ -AR agonists use, led to a discontinuation of inhaled corticosteroid therapy [162].

There were also concerns about a potential risk of mortality from the Salmeterol Nationwide Surveillance Study [163]. In this UK-based study of >25,000 subjects, there was a non-significant three-fold increased risk of asthma death in subjects on salmeterol compared with regular salbutamol. This led to the US-based Salmeterol Multicenter Asthma Research Trial, a placebo-controlled study of the safety of salmeterol in adults with unstable asthma [164]. The trial was stopped after *an interim* analysis showing a statistically significant, four-fold increase in asthma mortality with salmeterol. By contrast, in a large UK-based case-control study there was no positive association between long-acting  $\beta_2$ -AR agonists therapy and asthma death [165]. However, during the period of this study, ~95% of UK asthma patients receiving long-acting  $\beta_2$ -AR agonists therapy were co-prescribed inhaled corticosteroid [166]. Due to conflicting evidence, the Food and Drug Administration (FDA) confirmed the availability of both salmeterol and formoterol, but required black-box warnings on their product labels [167].

Formoterol fumarate is currently the most widely used long-acting  $\beta_2$ -AR worldwide. Three placebo-controlled trials showed that the higher dose (48  $\mu\text{g}/\text{day}$ ) of formoterol tended to be associated with more serious asthma exacerbations than a lower one [168]. However, a large phase IV trial found all doses of formoterol to be associated with fewer exacerbations than placebo, with no indication of any dose-response relationship [169].

Recently, Sears *et al.* [170] have reported a comprehensive meta-analysis of safety data ( $n = 68,004$  patients) obtained in completed trials. When comparing all formoterol-exposed patients to all patients not exposed to long-acting  $\beta_2$ -AR agonists in the whole data set, no increased risks of cardiac-related deaths or cardiac-related non-fatal serious adverse events were observed. Similarly, when examined by inhaled corticosteroids use there was no increased risk of cardiac serious adverse events with regimens of formoterol with inhaled corticosteroids *vs.* inhaled corticosteroids without long-acting  $\beta_2$ -AR agonists. Noteworthy, there was a significant reduction in asthma-related serious adverse events with formoterol (>90% of which were hospitalizations) speaking against a relationship between formoterol and increased asthma mortality. The main limitation of the study by Sears *et al.* [170] was the lack of sufficient power to form a definitive conclusion regarding the risk of death for patients treated with formoterol.

## 9. Future Prospects and Conclusions

A variety of  $\beta_2$ -AR agonists with longer half-lives are currently undergoing development, with the hope of achieving once-daily dosing. Among them, the development of indacaterol is very advanced and it is likely that this drug will be launched by the end of 2010 [171,172]. Indacaterol provided effective and sustained 24 h bronchodilator effect, with a rapid onset of action (<5 min) and a good tolerability and safety profile in asthma control [173].



Numerous studies have shown the benefit of adding a long-acting  $\beta_2$ -AR instead of doubling or increasing the dose of inhaled corticosteroids [136,174,175] and the greater benefits to most outcomes of adding a long-acting  $\beta_2$ -AR compared with adding a leukotriene antagonist [176]. However, both the International Asthma Treatment Guidelines [5,6] and the FDA [177] now emphasize that long-acting  $\beta_2$ -AR agonists should not be used as monotherapy in asthma but always together with an inhaled corticosteroid.

In conclusion, either as rescue medications and therapy added to or combined with inhaled corticosteroids, short- and long-acting  $\beta_2$ -AR agonists have proved effective in achieving a good control of asthma. In particular, they are essential in reducing daytime and especially nighttime symptoms, improving lung function, reducing the risk of exacerbations, and minimizing the required dose of inhaled corticosteroids.

## References

1. Chen, K.K.; Schmidt, C.F. Ephedrine and related substances. *Medicine (Baltimore)* **1930**, *9*, 1–117.
2. Ahlquist, R.P. A study of adrenotropic receptors. *Am. J. Physiol.* **1948**, *153*, 586–600.
3. Cazzola, M.; Matera, M.G. Emerging inhaled bronchodilators: an update. *Eur. Respir. J.* **2009**, *34*, 757–769.
4. Rabe, K.F.; Hurd, S.; Anzueto, A.; Barnes, P.J.; Buist, S.A.; Calverley, P.; Fukuchi, Y.; Jenkins, C.; Rodriguez-Roisin, R.; van Weel, C.; *et al.* Global Initiative for Chronic Obstructive Lung Disease. Global strategy for the diagnosis, management, and prevention of chronic obstructive pulmonary disease. GOLD executive summary. *Am. J. Respir. Crit. Care Med.* **2007**, *176*, 532–555.
5. National Asthma Education and Prevention Program. Expert Panel Report 3 (EPR-3): Guidelines for the Diagnosis and Management of Asthma – summary report 2007. *J. Allergy Clin. Immunol.* **2007**, *120*, Suppl. S94–S138.
6. Bateman, E.D.; Hurd, S.S.; Barnes, P.J.; Bousquet, J.; Drazen, J.M.; FitzGerald, M.; Gibson, P.; Ohta, K.; O'Byrne, P.; Pedersen, S.E.; *et al.* Global strategy for asthma management and prevention: GINA executive summary. *Eur. Respir. J.* **2008**, *31*, 143–178.
7. Doidge, J.M.; Satchell, D.G. Adrenergic and non-adrenergic inhibitory nerves in mammalian airways. *J. Auton. Nerv. Syst.* **1982**, *5*, 83–99.
8. Barnes, P.J. Neural control of human airways in health and disease. *Am. Rev. Respir. Dis.* **1986**, *134*, 1289–1314.
9. Belvisi, M.G. Overview of the innervation of the lung. *Curr. Opin. Pharmacol.* **2002**, *2*, 211–215.
10. Groneberg, D.A.; Harrison, S.; Dinh, Q.T.; Geppetti, P.; Fischer, A. Tachykinins in the respiratory tract. *Curr. Drug Targets* **2006**, *7*, 1005–1010.
11. Barnes, P.J. Neural control of airway function: new perspectives. *Mol. Aspects Med.* **1990**, *11*, 351–423.
12. Barnes, P.J. Modulation of neurotransmission in airways. *Physiol. Rev.* **1992**, *72*, 699–729.
13. Lees, G.M. A hitch-hiker's guide to the galaxy of adrenoceptors. *Br. J. Med.* **1981**, *283*, 173–178.

14. Hoffman, B.B.; Lefkowitz, R.J. Catecholamines, sympathomimetic drugs, and adrenergic receptor antagonists. In *Goodman & Gilman's the Pharmacological Basis of Therapeutics*, 9th ed.; Hardman, J.G., Limbird, L.E., Molinoff, P.B., Ruddon, R.W., Goodman, L.S., Gilman, A., Eds.; McGraw-Hill: New York, NY, USA, 1996; pp. 199–248.
15. Barnes, P.J. Endogenous catecholamines and asthma. *J. Allergy Clin. Immunol.* **1986**, *77*, 791–795.
16. Nadel, J.A.; Barnes, P.J.; Holtzman, M.J. Autonomic factors in hyperreactivity of airway smooth muscle. In *Handbook of Physiology. The Respiratory System*; Macklem, P.T.; Mead, J., Eds.; American Physiological Society: Bethesda, MD, USA, 1986; Volume 3, Part 2, pp. 693–702.
17. Rhoden, K.J.; Meldrum, L.A.; Barnes, P.J. Inhibition of cholinergic neurotransmission in human airways by  $\beta_2$ -adrenoceptors. *J. Appl. Physiol.* **1988**, *65*, 700–705.
18. Bai, T.R.; Lam, R.; Prasad, F.Y. Effects of adrenergic agonists and adenosine on cholinergic neurotransmission in human tracheal smooth muscle. *Pulmon. Pharmacol.* **1989**, *1*, 193–199.
19. Barnes, P.J. Muscarinic receptor subtype in airways. *Life Sci.* **1993**, *52*, 521–528.
20. Lands, A.M.; Arnold, A.; McAuliff, J.P.; Luduena, F.P.; Brown, T.G., Jr. Differentiation of receptor systems activated by sympathomimetic amines. *Nature* **1967**, *214*, 597–598.
21. Carstairs, J.R.; Nimmo, A.J.; Barnes, P.J. Autoradiographic visualization of beta-adrenoceptor subtype in human lung. *Am. Rev. Respir. Dis.* **1985**, *132*, 541–547.
22. Barnes, P.J.; Nadel, J.A.; Skoogh, B.E.; Roberts, J.M. Characterization of beta adrenoceptor subtypes in canine airway smooth muscle by radioligand binding and physiological responses. *J. Pharmacol. Exp. Ther.* **1983**, *225*, 456–461.
23. Hamid, Q.A.; Mak, J.C.; Sheppard, M.N.; Corrin, B.; Venter, J.C.; Barnes, P.J. Localization of beta<sub>2</sub>-adrenoceptor messenger RNA in human and rat lung using in situ hybridization: correlation with receptor autoradiography. *Eur. J. Pharmacol.* **1991**, *206*, 133–138.
24. Tamaoki, J.; Yamauchi, F.; Chiyotani, A.; Yamawaki, I.; Takeuchi, S.; Konno, K. Atypical  $\beta$ -adrenoceptor ( $\beta_3$ -adrenoceptor) mediated relaxation of canine isolated bronchial smooth muscle. *J. Appl. Physiol.* **1993**, *74*, 297–302.
25. Martin, C.A.; Naline, E.; Bakdach, H.; Advenier, C. Beta 3-adrenoceptor agonists, BRL 37344 and SR 58611A, do not induce relaxation of human, sheep and guinea-pig airway smooth muscle *in vitro*. *Eur. Respir. J.* **1994**, *7*, 1610–1615.
26. Emorine, L.J.; Marullo, S.; Briend-Sutren, M.M.; Patey, G.; Tate, K.; Delavier-Klutchko, C.; Strosberg, A.D. Molecular characterization of the human beta 3-adrenergic receptor. *Science* **1989**, *245*, 1118–1121.
27. Goldie, R.G.; Paterson, J.W.; Spina, D.; Wale, J.L. Classification of  $\beta_2$ -adrenoceptors in human isolated bronchus. *Br. J. Pharmacol.* **1984**, *81*, 611–615.
28. Nials, A.T.; Coleman, R.A.; Johnson, M.; Magnussen, H.; Rabe, K.F.; Vardey C.J. Effects of  $\beta$ -adrenoceptor agonists in human bronchial smooth muscle. *Br. J. Pharmacol.* **1993**, *110*, 1112–1116.
29. Löfdahl, C.G.; Svedmyr, N. Effects of prenalterol in asthmatic patients. *Eur. J. Clin. Pharmacol.* **1982**, *23*, 297–302.

30. Torphy, T.J.; Rinard, G.A.; Rietow, M.G.; Mayer, S.E. Functional antagonism in canine tracheal smooth muscle: inhibition by methacholine of the mechanical and biochemical responses to isoproterenol. *J. Pharmacol. Exp. Ther.* **1983**, *227*, 694–699.
31. Torphy, T.J.; Zheng, C.; Peterson, S.M.; Fiscus, R.R.; Rinard, G.A.; Mayer, S.E. Inhibitory effect of methacholine on drug-induced relaxation, cyclic AMP accumulation, and cyclic AMP-dependent protein kinase activation in canine tracheal smooth muscle. *J. Pharmacol. Exp. Ther.* **1985**, *232*, 409–417.
32. Gross, N.J.; Skorodin, M.S. Anticholinergic, antimuscarinic bronchodilators. *Am. Rev. Respir. Dis.* **1984**, *129*, 856–870.
33. Gilman, A.G. G proteins: Transducers of receptor-generated signals. *Annu. Rev. Biochem.* **1987**, *56*, 615–649.
34. Dohlman, H.G.; Thorner, J.; Caron, M.G.; Lefkowitz, R.J. Model systems for the study of seven-transmembrane-segment receptors. *Annu. Rev. Biochem.* **1991**, *60*, 653–688.
35. Neves, S.R.; Ram, P.T.; Iyengar, R. G protein pathways. *Science* **2002**, *296*, 1636–1639.
36. Dohlman HG. A scaffold makes the switch. *Sci. Signal.* **2008**, *1*, pe46.
37. Liggett, S.B.; Lefkowitz, R.J. Adrenergic receptor-coupled adenylyl cyclase systems: regulation of receptor function by phosphorylation, sequestration and downregulation. In *Regulation of Cellular Signal Transduction Pathways by Desensitization and Amplification*; Sibley, D.; Houslay, M., Eds.; John Wiley & Sons: London, UK, 1993; pp. 71–97.
38. Giembycz, M.A.; Raeburn, D. Putative substrates for cyclic nucleotide-dependent protein kinases and the control of airway smooth muscle tone. *J. Auton. Pharmacol.* **1991**, *11*, 365–398.
39. Gerthoffer, U.T. Calcium dependence of myosin phosphorylation and airway smooth muscle contraction and relaxation. *Am. J. Physiol.* **1986**, *250*, C597–C604.
40. Hall, I.P.; Hill, S.J. Beta-adrenoceptor stimulation inhibits histamine-stimulated inositol phospholipids hydrolysis in bovine tracheal smooth muscle. *Br. J. Pharmacol.* **1988**, *95*, 1204–1212.
41. Twort, C.H.; van Breemen, C. Human airway smooth muscle in cell culture: control of the intracellular calcium store. *Pulm. Pharmacol.* **1989**, *2*, 45–53.
42. Gunst, S.J.; Stropp, J.Q. Effect of Na-K adenosinetriphosphatase activity on relaxation of canine tracheal smooth muscle. *J. Appl. Physiol.* **1988**, *64*, 635–641.
43. Jones, T.R.; Charette, L.; Garcia, M.L.; Kaczorowski, G.J. Selective inhibition of relaxation of guinea-pig trachea by charybdotoxin, a potent Ca<sup>(++)</sup>-activated K<sup>+</sup> channel inhibitor. *J. Pharmacol. Exp. Ther.* **1990**, *255*, 697–706.
44. Jones, T.R.; Charette, L.; Garcia, M.L.; Kaczorowski, G.J. Interaction of iberiotoxin with beta-adrenoceptor agonists and sodium nitroprusside on guinea pig trachea. *J. Appl. Physiol.* **1993**, *74*, 1879–1884.
45. Tanaka, Y.; Yamashita, Y.; Yamaki, F.; Horinouchi, T.; Shigenobu, K.; Koike, K. MaxiK channel mediates beta2-adrenoceptor-activated relaxation to isoprenaline through cAMP-dependent and -independent mechanisms in guinea-pig tracheal smooth muscle. *J. Smooth Muscle Res.* **2003**, *39*, 205–219.

46. Koike, K.; Yamashita, Y.; Horinouchi, T.; Yamaki, F.; Tanaka, Y. cAMP-independent mechanism is significantly involved in beta<sub>2</sub>-adrenoceptor-mediated tracheal relaxation. *Eur. J. Pharmacol.* **2004**, *492*, 65–70.
47. Miura, M.; Belvisi, M.G.; Stretton, C.D.; Yacoub, M.H.; Barnes, P.J. Role of potassium channels in bronchodilator responses in human airways. *Am. Rev. Respir. Dis.* **1992**, *146*, 132–136.
48. Kume, H.; Graziano, M.P.; Kotlikoff, M.I. Stimulatory and inhibitory regulation of calcium-activated potassium channels by guanine nucleotide-binding proteins. *Proc. Natl. Acad. Sci. USA* **1992**, *89*, 11051–11055.
49. Belvisi, M.G.; Patel, H.J.; Takahashi, T.; Barnes, P.J.; Giembycz, M.A. Paradoxical facilitation of acetylcholine release from parasympathetic nerves innervating guinea-pig trachea by isoprenaline. *Br. J. Pharmacol.* **1996**, *117*, 1413–1420.
50. De Haas, J.R.; Terpstra, J.S.; Van der Zwaag, M.; Kockelbergh, P.G.; Roffel, A.D.F.; Zaagsma, J. Facilitatory β<sub>2</sub>-adrenoceptors on cholinergic and adrenergic nerve endings of the guinea pig trachea. *Am. J. Physiol. Lung Cell. Mol. Physiol.* **1999**, *276*, L420–L425.
51. Zhang, X.Y.; Olszewski, M.A.; Robinson, N.E. β<sub>2</sub>-Adrenoceptor activation augments acetylcholine release from tracheal parasympathetic nerves. *Am. J. Physiol. Lung Cell. Mol. Physiol.* **1995**, *268*, L950–L956.
52. Wessler, I.; Reinheimer, T.; Brunn, G.; Anderson, G.P.; Maclagan, J.; Rackè, K. β<sub>2</sub>-adrenoceptors mediate inhibition of [<sup>3</sup>H]-acetylcholine release from the isolated rat and guinea-pig trachea: role of the airway mucosa and prostaglandins. *Br. J. Pharmacol.* **1994**, *113*, 1221–1230.
53. Seamon, K.B.; Padgett, W.; Daly, J.W. Forskolin: Unique diterpene activator of adenylate cyclase in membranes and in intact cells. *Proc. Natl. Acad. Sci. USA* **1981**, *78*, 3363–3367.
54. Patel, H.J.; Giembycz, M.A.; Keeling, J.E.; Barnes, P.J.; Belvisi, M.G. Inhibition of cholinergic neurotransmission in guinea pig trachea by NS1619, a putative activator of large-conductance, calcium-activated potassium channels. *J. Pharmacol. Exp. Ther.* **1998**, *286*, 952–958.
55. Bricchetto, L.; Song, P.; Crimi, E.; Rehder, K.; Brusasco, V. Modulation of cholinergic responsiveness through the β-adrenoceptor signal transmission pathway in bovine trachealis. *J. Appl. Physiol.* **2003**, *95*, 735–741.
56. Freund-Michel, V.C.; Birrell, M.A.; Giembycz, M.A.; Hele, D.J.; Haj-Yahia, S.; Belvisi, M.G. Beta<sub>2</sub>-agonists block tussive responses in guinea pigs via an atypical cAMP-dependent pathway. *Eur. Respir. J.* **2010**, *35*, 647–654.
57. Chong, C.F.; Chen, C.C.; Ma, H.P.; Wu, Y.C.; Chen, Y.C.; Wang, T.L. Comparison of lidocaine and bronchodilator inhalation treatments for cough suppression in patients with chronic obstructive pulmonary disease. *Emerg. Med. J.* **2005**, *22*, 429–432.
58. Mulrennan, S.; Wright, C.; Thompson, R.; Goustas, P.; Morice, A. Effect of salbutamol on smoking related cough. *Pulm. Pharmacol. Ther.* **2004**, *17*, 127–131.
59. Chang, A.B.; Phelan, P.D.; Carlin, J.B.; Sawyer, S.M.; Robertson, C.F. A randomised, placebo controlled trial of inhaled salbutamol and beclomethasone for recurrent cough *Arch. Dis. Child.* **1998**, *79*, 6–11.

60. Smith, C.A.; Adamson, D.L.; Choudry, N.B.; Fuller, R.W. The effect of altering airway tone on the sensitivity of the cough reflex in normal volunteers. *Eur. Respir. J.* **1991**, *4*, 1078–1079.
61. Brusasco, V.; Crimi, E.; Gherson, G.; Nardelli, R.; Oldani, V.; Francucci, B.; Della Cioppa, S.; Senn, S.; Fabbri, L. Actions other than smooth muscle relaxation may play a role in the protective effects of formoterol on the allergen-induced late asthmatic reaction. *Pulm. Pharmacol. Ther.* **2002**, *15*, 399–406.
62. Johnson, S.R.; Knox, A.J. Synthetic functions of airway smooth muscle in asthma. *Trends Pharmacol. Sci.* **1997**, *18*, 288–292.
63. Baroffio, M.; Crimi, E.; Brusasco, V. Airway smooth muscle as a model for new investigative drugs in asthma. *Ther. Adv. Respir. Dis.* **2008**, *2*, 129–139.
64. Baroffio, M.; Barisione, G.; Crimi, E.; Brusasco, V. Noninflammatory mechanisms of airway hyper-responsiveness in bronchial asthma: an overview. *Ther. Adv. Respir. Dis.* **2009**, *3*, 163–174.
65. Black, J.L.; Roth, M. Intrinsic asthma: is it intrinsic to the smooth muscle? *Clin. Exp. Allergy* **2009**, *39*, 962–965.
66. Kaur, M.; Holden, N.S.; Wilson, S.M.; Sukkar, M.B.; Chung, K.F.; Barnes, P.J.; Newton, R.; Giembycz, M.A. Effect of beta<sub>2</sub>-adrenoceptor agonists and other cAMP-elevating agents on inflammatory gene expression in human ASM cells: a role for protein kinase A. *Am. J. Physiol. Lung Cell. Mol. Physiol.* **2008**, *295*, L505–L514.
67. Lefkowitz, R.J.; Hausdorff, W.P.; Caron, M.G. Role of phosphorylation in desensitization of the beta-adrenoceptor. *Trends Pharmacol. Sci.* **1990**, *11*, 190–194.
68. Bai, T.R.; Mak, J.C.W.; Barnes, P.J. A comparison of  $\beta$ -adrenergic receptors and *in vitro* relaxant responses to isoproterenol in asthmatic airway smooth muscle. *Am. J. Respir. Cell. Mol. Biol.* **1992**, *6*, 647–651.
69. Brusasco, V.; Crimi, E.; Baroffio, M. Allergic airway inflammation and beta-adrenoceptor dysfunction. *Cell. Biochem. Biophys.* **2006**, *44*, 129–138.
70. Szentivanyi, A. Beta adrenergic theory of atopic abnormality in bronchial asthma. *J. Allergy* **1968**, *42*, 203–232.
71. Trian, T.; Ge, Q.; Moir, L.M.; Burgess, J.K.; Kuo, C.; King, N.J.; Reddel, H.K.; Black, J.L.; Oliver, B.G.; McParland, B.E. Rhinovirus-induced exacerbations of asthma - How is the beta<sub>2</sub>-adrenoceptor implicated? *Am. J. Respir. Cell. Mol. Biol.* **2009**, Epub ahead of print.
72. Laporte, J.D.; Moore, P.E.; Panettieri, R.A.; Moeller, W.; Heyder, J.; Shore, S.A. Prostanoids mediate IL-1 $\beta$ -induced  $\beta$ -adrenergic hyporesponsiveness in human airway smooth muscle cells. *Am. J. Physiol.* **1998**, *275*, L491–L501.
73. Hausdorff, W.P.; Caron, M.G.; Lefkowitz, R.J. Turning off the signal: desensitization of beta-adrenergic receptor function. *FASEB J.* **1990**, *4*, 2881–2889.
74. Song, P.; Milanese, M.; Crimi, E.; Bruzzone, S.; Zocchi, E.; Rehder, K.; Brusasco, V. G(s) protein dysfunction in allergen-challenged human isolated passively sensitized bronchi. *Am. J. Physiol. Lung Cell. Mol. Physiol.* **2000**, *279*, L209–L215.

75. Milanese, M.; Riccio, A.M.; Gamalero, C.; De Giovanni, B.; Bricchetto, L.; Baroffio, M.; Crimi, E.; Brusasco, V.; Canonica, G.W. A model of allergen-driven human airway contraction: beta<sub>2</sub>- pathway dysfunction without cytokine involvement. *Ann. Allergy Asthma Immunol.* **2005**, *94*, 273–278.
76. Song, P.; Milanese, M.; Crimi, E.; Rehder, K.; Brusasco, V. Allergen challenge of passively sensitized human bronchi alters M<sub>2</sub>- and beta<sub>2</sub>-receptor function. *Am. J. Respir. Crit. Care Med.* **1997**, *155*, 1230–1234.
77. Song, P.; Crimi, E.; Milanese, M.; Duan, J.; Rehder, K.; Brusasco, V. Anti-inflammatory agents and allergen-induced beta<sub>2</sub>-receptor dysfunction in isolated human bronchi. *Am. J. Respir. Crit. Care Med.* **1998**, *158*, 1809–1814.
78. Rovati, G.E.; Baroffio, M.; Citro, S.; Bricchetto, L.; Ravasi, S.; Milanese, M.; Crimi, E.; Brusasco, V. Cysteinyl-leukotrienes in the regulation of beta<sub>2</sub>-adrenoceptor function: an *in vitro* model of asthma. *Respir. Res.* **2006**, *7*, 103–113.
79. Benovic, J.L.; Mayor, F., Jr.; Staniszewski, C.; Lefkowitz, R.J.; Caron, M.G. Purification and characterization of the beta-adrenergic receptor kinase. *J. Biol. Chem.* **1987**, *262*, 9026–9032.
80. Benovic, J.L.; Kühn, H.; Weyand, I.; Codina, J.; Caron, M.G.; Lefkowitz, R.J. Functional desensitization of the isolated beta-adrenergic receptor by the beta-adrenergic receptor kinase: potential role of an analog of the retinal protein arrestin (48-kDa protein). *Proc. Natl. Acad. Sci. USA* **1987**, *84*, 8879–8882.
81. Lohse, M.J.; Benovic, J.L.; Codina, J.; Caron, M.G.; Lefkowitz, R.J. Beta-Arrestin: A protein that regulates beta-adrenergic receptor function. *Science* **1990**, *248*, 1547–1550.
82. Rajagopal, S.; Kim, J.; Ahn, S.; Craig, S.; Lam, C.M.; Gerard, N.P.; Gerard, C.; Lefkowitz, R.J. Beta-arrestin- but not G protein-mediated signaling by the "decoy" receptor CXCR7. *Proc. Natl. Acad. Sci. USA* **2010**, *107*, 628–632.
83. Nino, G.; Hu, A.; Grunstein, J.S.; Grunstein, M. Mechanism regulating proasthmatic effects of prolonged homologous β<sub>2</sub>-adrenergic receptor desensitization in airway smooth muscle. *Am. J. Physiol. Lung Cell. Mol. Physiol.* **2009**, *297*, L746–L757.
84. Wang, W.C.; Mihlbachler, K.A.; Brunnett, A.C.; Liggett, S.B. Targeted transgenesis reveals discrete attenuator functions of GRK and PKA in airway beta<sub>2</sub>-adrenergic receptor physiologic signaling. *Proc. Natl. Acad. Sci. USA* **2009**, *106*, 15007–15012.
85. Giembycz, M.A. An estimation of beta<sub>2</sub>-adrenoceptor reserve on human bronchial smooth muscle for some sympathomimetic bronchodilators. *Br. J. Pharmacol.* **2009**, *158*, 287–299.
86. Yu, S.S.; Lefkowitz, R.J.; Hausdorff, W.P. Beta-adrenergic receptor sequestration. A potential mechanism of receptor resensitization. *J. Biol. Chem.* **1993**, *268*, 337–341.
87. Hadcock, J.R.; Wang, H.Y.; Malbon, C.C. Agonist-induced destabilization of beta-adrenergic receptor mRNA. Attenuation of glucocorticoid-induced up-regulation of beta-adrenergic receptors. *J. Biol. Chem.* **1989**, *264*, 19928–19933.

88. Collins, S.; Bouvier, M.; Bolanowski, M.A.; Caron, M.G.; Lefkowitz, R.J. cAMP stimulates transcription of the beta<sub>2</sub>-adrenergic receptor gene in response to short-term agonist exposure. *Proc. Natl. Acad. Sci. USA* **1989**, *86*, 4853–4857.
89. Huang, L.Y.; Tholanikunnel, B.G.; Vakalopoulou, E.; Malbon, C.C. The M(r) 35,000 beta-adrenergic receptor mRNA-binding protein induced by agonists requires both an AUUUA pentamer and U-rich domains for RNA recognition. *J. Biol. Chem.* **1993**, *268*, 25769–25775.
90. Bouvier, M.; Collins, S.; O'Dowd, B.F.; Campbell, P.T.; de Blasi, A.; Kobilka, B.K.; MacGregor, C.; Irons, G.P.; Caron, M.G.; Lefkowitz, R.J. Two distinct pathways for cAMP-mediated down-regulation of the beta<sub>2</sub>-adrenergic receptor. Phosphorylation of the receptor and regulation of its mRNA level. *J. Biol. Chem.* **1989**, *264*, 16786–16792.
91. Valiquette, M.; Bonin, H.; Hnatowich, M.; Caron, M.G.; Lefkowitz, R.J.; Bouvier, M. Involvement of tyrosine residues located in the carboxyl tail of the human beta<sub>2</sub>-adrenergic receptor in agonist-induced down-regulation of the receptor. *Proc. Natl. Acad. Sci. USA* **1990**, *87*, 5089–5093.
92. Kraan, J.; Koëter, G.H.; vd Mark, T.W.; Sluiter, H.J.; de Vries, K. Changes in bronchial hyperreactivity induced by 4 weeks of treatment with antiasthmatic drugs in patients with allergic asthma: a comparison between budesonide and terbutaline. *J. Allergy Clin. Immunol.* **1985**, *76*, 628–636.
93. Kerrebijn, K.F.; van Essen-Zandvliet, E.E.; Neijens, H.J. Effect of long-term treatment with inhaled corticosteroids and beta-agonists on the bronchial responsiveness in children with asthma. *J. Allergy Clin. Immunol.* **1987**, *79*, 653–659.
94. van Schayck, C.P.; Graafsma, S.J.; Visch, M.B.; Dompeling, E.; van Weel, C.; van Herwaarden, C.L. Increased bronchial hyperresponsiveness after inhaling salbutamol during 1 year is not caused by subsensitization to salbutamol. *J. Allergy Clin. Immunol.* **1990**, *86*, 793–800.
95. Cheung, D.; Timmers, M.C.; Zwinderman, A.H.; Bel, E.H.; Dijkman, J.H.; Sterk, P.J. Long-term effects of a long-acting beta<sub>2</sub>-adrenoceptor agonist, salmeterol, on airway hyperresponsiveness in patients with mild asthma. *N. Engl. J. Med.* **1992**, *327*, 1198–1203.
96. McGraw, D.W.; Almoosa, K.F.; Paul, R.J.; Kobilka, B.K.; Liggett, S.B. Antithetic regulation by β-adrenergic receptors of G<sub>q</sub> receptor signaling *via* phospholipase C underlies the airway β-agonist paradox. *J. Clin. Invest.* **2003**, *112*, 619–626.
97. Barnes, P.J. Scientific rationale for using a single inhaler for asthma control. *Eur. Respir. J.* **2007**, *29*, 587–595.
98. Callaerts-Vegh, Z.; Evans, K.L.; Dudekula, N.; Cuba, D.; Knoll, B.J.; Callaerts, P.F.; Giles, H.; Shardonofsky, F.R.; Bond, R.A. Effects of acute and chronic administration of beta-adrenoceptor ligands on airway function in a murine model of asthma. *Proc. Natl. Acad. Sci. USA* **2004**, *101*, 4948–4953.
99. Nguyen, L.P.; Omoluabi, O.; Parra, S.; Frieske, J.M.; Clement, C.; Ammar-Aouchiche, Z.; Ho, S.B.; Ehre, C.; Kesimer, M.; Knoll, B.J.; *et al.* Chronic exposure to beta-blockers attenuates inflammation and mucin content in a murine asthma model *Am. J. Respir. Cell. Mol. Biol.* **2008**, *38*, 256–262.

100. Hanania, N.A.; Singh, S.; El-Wali, R.; Flashner, M.; Franklin, A.E.; Garner, W.J.; Dickey, B.F.; Parra, S.; Ruoss, S.; Shardonofsky, F.; *et al.* The safety and effects of the beta-blocker, nadolol, in mild asthma: an open-label pilot study. *Pulm. Pharmacol. Ther.* **2008**, *21*, 134–141.
101. Lipworth, B.J.; Williamson, P.A. Beta blockers for asthma: a double-edged sword. *Lancet* **2009**, *373*, 104–105.
102. Hanania, N.A.; Dickey, B.F.; Bond, R.A. Clinical implications of the intrinsic efficacy of beta-adrenoceptor drugs in asthma: full, partial and inverse agonism. *Curr. Opin. Pulm. Med.* **2010**, *16*, 1–5.
103. Davies, A.O.; Lefkowitz, R.J. *In vitro* desensitization of beta-adrenergic receptors in human neutrophils. Attenuation by corticosteroids. *J. Clin. Invest.* **1983**, *71*, 565–571.
104. Bricchetto, L.; Milanese, M.; Song, P.; Patrone, M.; Crimi, E.; Rehder, K.; Brusasco, V. Beclomethasone rapidly ablates allergen-induced beta<sub>2</sub>-adrenoceptor pathway dysfunction in human isolated bronchi. *Am. J. Physiol. Lung Cell. Mol. Physiol.* **2003**, *284*, L133–L139.
105. Green, S.A.; Turki, J.; Hall, I.P.; Liggett, S.B. Implications of genetic variability of human beta<sub>2</sub>-adrenergic receptor structure. *Pulm. Pharmacol.* **1995**, *8*, 1–10.
106. Liggett, S.B. Genetics of beta<sub>2</sub>-adrenergic receptor variants in asthma. *Clin. Exp. Allergy* **1995**, *Suppl. 2*, 89–94.
107. Kazani, S.; Wechsler, M.E.; Israel, E. The role of pharmacogenomics in improving the management of asthma. *J. Allergy Clin. Immunol.* **2010**, *125*, 295–302.
108. Taylor, D.R.; Drazen, J.M.; Herbison, G.P.; Yandava, C.N.; Hancox, R.J.; Town, G.I. Asthma exacerbations during long term beta-agonist use: influence of beta<sub>2</sub>-adrenoceptor polymorphism. *Thorax* **2000**, *55*, 762–767.
109. Basu, K.; Palmer, C.N.; Tavendale, R.; Lipworth, B.J.; Mukhopadhyay, S. Beta<sub>2</sub>-receptor genotype predisposes to exacerbations in steroid-treated asthmatic patients taking frequent albuterol or salmeterol. *J. Allergy Clin. Immunol.* **2009**, *124*, 1188–1194.
110. Wechsler, M.E.; Kunselman, S.J.; Chinchilli, V.M.; Bleecker, E.; Boushey, H.A.; Calhoun, W.J.; Ameredes, B.T.; Castro, M.; Craig, T.J.; Denlinger, L.; *et al.* Effect of beta<sub>2</sub>-adrenergic receptor polymorphism on response to long-acting beta<sub>2</sub>-agonist in asthma (LARGE trial): A genotype-stratified, randomised, placebo-controlled, crossover trial. *Lancet* **2009**, *374*, 1754–1764.
111. Wang, W.C.; Mihlbachler, K.A.; Bleecker, E.R.; Weiss, S.T.; Liggett, S.B. A polymorphism of G-protein coupled receptor kinase 5 alters agonist-promoted desensitization of beta<sub>2</sub>-adrenergic receptors. *Pharmacogenet. Genomics* **2008**, *18*, 729–732.
112. Hall, I.P.; Wheatley, A.; Wilding, P.; Liggett, S.B. Association of Glu 27 beta<sub>2</sub>-adrenoceptor polymorphism with lower airway reactivity in asthmatic subjects. *Lancet* **1995**, *345*, 1213–1214.
113. Litonjua, A.A.; Lasky-Su, J.; Schneiter, K.; Tantisira, K.G.; Lazarus, R.; Klanderman, B.; Lima, J.J.; Irvin, C.G.; Peters, S.P.; Hanrahan, J.P.; *et al.* ARG1 is a novel bronchodilator response gene: screening and replication in four asthma cohorts. *Am. J. Respir. Crit. Care Med.* **2008**, *178*, 688–694.
114. Nelson, H.S. β-Adrenergic bronchodilators. *N. Engl. J. Med.* **1995**, *333*, 499–506.



115. Lipworth, B.J.; Struthers, A.D.; McDevitt, D.G. Tachyphylaxis to systemic but not to airway responses during prolonged therapy with high dose inhaled salbutamol in asthmatics. *Am. Rev. Respir. Dis.* **1989**, *140*, 586–592.
116. Drazen, J.M.; Israel, E.; Boushey, H.A.; Chinchilli, V.M.; Fahy, J.V.; Fish, J.E.; Lazarus, S.C.; Lemanske, R.F.; Martin, R.J.; Peters, S.P.; *et al.* Comparison of regularly scheduled with as-needed use of albuterol in mild asthma. Asthma Clinical Research Network. *N. Engl. J. Med.* **1996**, *335*, 841–847.
117. Israel, E.; Drazen, J.M.; Liggett, S.B.; Boushey, H.A.; Cherniack, R.M.; Chinchilli, V.M.; Cooper, D.M.; Fahy, J.V.; Fish, J.E.; Ford, J.G.; *et al.* The effect of polymorphisms of the beta(2)-adrenergic receptor on the response to regular use of albuterol in asthma. *Am. J. Respir. Crit. Care Med.* **2000**, *162*, 75–80.
118. Israel, E.; Chinchilli, V.M.; Ford, J.G.; Boushey, H.A.; Cherniack, R.; Craig, T.J.; Deykin, A.; Fagan, J.K.; Fahy, J.V.; Fish, J.; *et al.* Use of regularly scheduled albuterol treatment in asthma: genotype-stratified, randomised, placebo-controlled cross-over trial. *Lancet* **2004**, *364*, 1505–1512.
119. Milanese, M.; Saporiti, R.; Bartolini, S.; Pellegrino, R.; Baroffio, M.; Brusasco, V.; Crimi, E. Bronchodilator effects of exercise hyperpnea and albuterol in mild-to-moderate asthma. *J. Appl. Physiol.* **2009**, *107*, 494–499.
120. Hendeles, L.; Colice, G.L.; Meyer, R.J. Withdrawal of albuterol inhalers containing chlorofluorocarbon propellants. *N. Engl. J. Med.* **2007**, *356*, 1344–1351.
121. Ramsdell, J.W.; Colice, G.L.; Ekholm, B.P.; Klinger, N.M. Cumulative dose response study comparing HFA-134a albuterol sulfate and conventional CFC albuterol in patients with asthma. *Ann. Allergy Asthma Immunol.* **1998**, *81*, 593–599.
122. Cates, C.J.; Crilly, J.A.; Rowe, B.H. Holding chambers (spacers) versus nebulizers for beta-agonist treatment of acute asthma. *Cochrane Database Syst. Rev.* **2006**, *2*, CD000052–CD000052.
123. Henderson, W.R., Jr.; Banerjee, E.R.; Chi, E.Y. Differential effects of (S)- and (R)-enantiomers of albuterol in a mouse asthma model. *J. Allergy Clin. Immunol.* **2005**, *116*, 332–340.
124. Berger, W.E.; Milgrom, H.; Skoner, D.P.; Tripp, K.; Parsey, M.V.; Baumgartner, R.A.; Xopenex Pediatric Asthma Group. Evaluation of levalbuterol metered dose inhaler in pediatric patients with asthma: a double-blind, randomized, placebo- and active-controlled trial. *Curr. Med. Res. Opin.* **2006**, *22*, 1217–1226.
125. O'Byrne, P.M.; Bisgaard, H.; Godard, P.P.; Pistolesi, M.; Palmqvist, M.; Zhu, Y.; Ekström, T.; Bateman ED. Budesonide/formoterol combination therapy as both maintenance and reliever medication in asthma. *Am. J. Respir. Crit. Care Med.* **2005**, *171*, 129–136.
126. Pearlman, D.S.; Chervinsky, P.; LaForce, C.; Seltzer, J.M.; Southern, D.L.; Kemp, J.P.; Dockhorn, R.J.; Grossman, J.; Liddle, R.F.; Yancey, S.W.; *et al.* A comparison of salmeterol with albuterol in the treatment of mild-to-moderate asthma. *N. Engl. J. Med.* **1992**, *327*, 1420–1425.
127. Nelson, J.A.; Strauss, L.; Skowronski, M.; Ciufo, R.; Novak, R.; McFadden, E.R., Jr. Effect of long-term salmeterol treatment on exercise-induced asthma. *N. Engl. J. Med.* **1998**, *339*, 141–146.

128. Weinberger, M.; Abu-Hasan, M. Life-threatening asthma during treatment with salmeterol. *N. Engl. J. Med.* **2006**, *355*, 852–853.
129. Smyth, E.T.; Pavord, I.D.; Wong, C.S.; Wisniewski, A.F.; Williams, J.; Tattersfield, A.E. Interaction and dose equivalence of salbutamol and salmeterol in patients with asthma. *Br. Med. J.* **1993**, *306*, 543–545.
130. Appleton, S.; Poole, P.; Smith, B.; Veale, A.; Bara A. Long-acting  $\beta_2$ -agonists for chronic obstructive pulmonary disease patients with poorly reversible airflow limitation. *Cochrane Database Syst. Rev.* **2002**, *3*, CD001104.
131. D'Urzo, A.D.; De Salvo, M.C.; Ramirez-Rivera, A.; Almeida, J.; Sichletidis, L.; Rapatz, G.; Kottakis, J.; FOR-INT-03 Study Group. In patients with COPD, treatment with a combination of formoterol and ipratropium is more effective than a combination of salbutamol and ipratropium: a 3-week, randomized, double-blind, within-patient, multicenter study. *Chest* **2001**, *119*, 1347–1356.
132. Barnes, P.J.; Stockley, R.A. COPD: current therapeutic interventions and future approaches. *Eur. Respir. J.* **2005**, *25*, 1084–1106.
133. Goldkorn, A.; Diotto, P.; Burgess, C.; Weatherall, M.; Holt, S.; Beasley, R.; Siebers, R. The pulmonary and extra-pulmonary effects of high-dose formoterol in COPD: a comparison with salbutamol. *Respirology* **2004**, *9*, 102–108.
134. Sovrani, M.P.; Whale, C.I.; Tattersfield, A.E. A benefit-risk assessment of inhaled long-acting  $\beta_2$ -agonists in the management of obstructive pulmonary disease. *Drug Safety* **2004**, *27*, 689–715.
135. Lazarus, S.C.; Boushey, H.A.; Fahy, J.V.; Chinchilli, V.M.; Lemanske, R.F., Jr.; Sorkness, C.A.; Kraft, M.; Fish, J.E.; Peters, S.P.; Craig, T.; *et al.* Long-acting beta<sub>2</sub>-agonist monotherapy vs continued therapy with inhaled corticosteroids in patients with persistent asthma: a randomized controlled trial. *JAMA* **2001**, *285*, 2583–2593.
136. Gibson, P.G.; Powell, H.; Ducharme, F.M. Differential effects of maintenance long-acting beta-agonist and inhaled corticosteroid on asthma control and asthma exacerbations. *J. Allergy Clin. Immunol.* **2007**, *119*, 344–350.
137. Woolcock, A.; Lundback, B.; Ringdal, N.; Jacques, L.A. Comparison of addition of salmeterol to inhaled steroids with doubling of the dose of inhaled steroids. *Am. J. Respir. Crit. Care Med.* **1996**, *153*, 1481–1488.
138. Pauwels, R.A.; Löfdahl, C.G.; Postma, D.S.; Tattersfield, A.E.; O'Byrne, P.; Barnes, P.J.; Ullman, A. Effect of inhaled formoterol and budesonide on exacerbations of asthma. *N. Engl. J. Med.* **1997**, *337*, 1405–1411. [Erratum, *N. Engl. J. Med.* **1998**, *338*, 139.]
139. Giembycz, M.A.; Kaur, M.; Leigh, R.; Newton, R. A Holy Grail of asthma management: toward understanding how long-acting  $\beta_2$ -adrenoceptor agonists enhance the clinical efficacy of inhaled corticosteroids. *Br. J. Pharmacol.* **2008**, *153*, 1090–1104.
140. Papi, A.; Canonica, G.W.; Maestrelli, P.; Paggiaro, P.L.; Olivieri, D.; Pozzi, E.; Crimi, N.; Vignola, A.M.; Morelli, P.; Nicolini, G.; *et al.* Rescue use of beclomethasone and albuterol in a single inhaler for mild asthma. *N. Engl. J. Med.* **2007**, *356*, 2040–2052.

141. Barger, G.; Dale, H.H. Chemical structure and sympathomimetic action of amines. *J. Physiol. (Lond.)*, **1910**, *41*, 19–59.
142. Benson, R.L.; Perlman, F. Clinical effects of epinephrine by inhalation. *J. Allergy* **1948**, *19*, 129–140.
143. Davies, D.S. Metabolism of isoprenaline and other bronchodilator drugs in man and dog. *Bull. Physiopathol. Respir.* **1972**, *8*, 679–682.
144. Speizer, F.E.; Doll, R.; Heaf, P. Observations on recent increase in mortality from asthma. *Br. Med. J.* **1968**, *1*, 335–339.
145. Fraser, P.; Doll, R. Geographical variations in the epidemic of asthma deaths. *Br. J. Prev. Soc. Med.* **1971**, *25*, 34–36.
146. Jackson, R.T.; Beaglehole, R.; Rea, H.H.; Sutherland, D.C. Mortality from asthma : a new epidemic in New Zealand. *Br. Med. J.* **1982**, *285*, 771–774.
147. Speizer, F.E.; Doll, R.; Heaf, P.; Strang, L.B. Investigation into use of drugs preceding death from asthma. *Br. Med. J.* **1968**, *1*, 339–343.
148. Conolly, M.E.; Davies, D.S.; Dollery, C.T.; George, C.F. Resistance to beta-adrenoceptor stimulants (a possible explanation for the rise in ashtma deaths). *Br. J. Pharmacol.* **1971**, *43*, 389–402.
149. Wilson, J.D.; Sutherland, D.C.; Thomas, A.C. Has the change to beta-agonists combined with oral theophylline increased cases of fatal asthma? *Lancet* **1981**, *1*, 1235–1237.
150. Grant, I.W.B. Asthma in New Zealand. *Br. Med. J.* **1983**, *286*, 374–377.
151. Crane, J.; Flatt, A.; Jackson, R.; Ball, M.; Pearce, N.; Burgess, C.; Kwong, T.; Beasley, R. Prescribed fenoterol and death from asthma in New Zealand, 1981–83; Case-control study. *Lancet* **1989**, *1*, 917–922.
152. Pearce, N.; Grainger, J.; Atkinson, M.; Crane, J.; Burgess, C.; Culling, C.; Windom, H.; Beasley, R. Case-control study of prescribed fenoterol and death from asthma in New Zealand, 1977–81. *Thorax* **1990**, *45*, 170–175.
153. Spitzer, W.O.; Suissa, S.; Ernst, P.; Horwitz, R.I.; Habbick, B.; Cockcroft, D.; Boivin, J.F.; McNutt, M.; Buist, A.S.; Rebusck, A.S. The use of beta-agonists and the risk of death and near death from asthma. *N. Engl. J. Med.* **1992**, *326*, 501–506.
154. Taylor, D.R.; Sears, M.R.; Herbison, G.P.; Flannery, E.M.; Print, C.G.; Lake, D.C. Yates, D.M.; Lucas, M.K.; Li, Q. Regular inhaled beta agonist in asthma: effect on exacerbations and lung function. *Thorax* **1993**, *48*, 134–138.
155. Dennis, S.M.; Sharp, S.J.; Vickers, M.R.; Frost, C.D.; Crompton, G.K.; Barnes, P.J.; Lee, T.H. Regular inhaled salbutamol and asthma control: the TRUST randomized trial. *Lancet* **2000**, *355*, 1675–1679.
156. British Thoracic Society, National Asthma Campaign, Royal College of Physicians of London, General Practitioner in Asthma Group. The British Guidelines on Asthma Management 1995 Review and Position Statement. *Thorax* **1997**, *52*, Suppl. 1, S1–S20.
157. Ullman, A.; Svedmyr, N. Salmeterol, a new long-acting inhaled beta<sub>2</sub>-adrenoceptor agonist: comparison with salbutamol in adult asthmatic patients. *Thorax* **1988**, *43*, 674–678.

158. Faulds, D.; Hollingshead, L.M.; Goa, K.L. Formoterol. A review of its pharmacologic properties and therapeutic efficacy in reversible obstructive airways disease. *Drugs* **1991**, *42*, 115–137.
159. Grove, A.; Lipworth, B.J. Bronchodilator subsensitivity to salbutamol after twice daily salmeterol in asthmatic patients. *Lancet* **1995**, *346*, 201–206.
160. Newnham, D.M.; McDevitt, D.G.; Lipworth, B.J. Bronchodilator subsensitivity after chronic dosing with eformoterol in patients with asthma. *Am. J. Med.* **1994**, *97*, 29–37.
161. Yates, D.H.; Kharitonov, S.A.; Barnes, P.J. An inhaled glucocorticoid does not prevent tolerance to the bronchoprotective effect of a long-acting  $\beta_2$ -agonist. *Am. J. Respir. Crit. Care Med.* **1996**, *154*, 1603–1607.
162. Arvidsson, P.; Larsson, S.; Löfdahl, C-G.; Melander, B.; Svedmyr, N.; Wåhlander, L. Inhaled formoterol during one year in asthma: a comparison with salbutamol. *Eur. Respir. J.* **1991**, *4*, 1168-1173.
163. Castle, W.; Fuller, R.; Hall, J.; Palmer, J. Serevent nationwide surveillance study: comparison of salmeterol with salbutamol in asthmatic patients who require regular bronchodilator treatment. *Br. Med. J.* **1993**, *306*, 1034–1037.
164. Nelson, H.S.; Weiss, S.T.; Bleecker, E.R.; Yancey, S.W.; Dorinsky, P.M. The Salmeterol Multicenter Asthma Research Trial. A comparison of usual pharmacotherapy for asthma or usual pharmacotherapy plus salmeterol. *Chest* **2006**, *129*, 15–26.
165. Anderson, H.R.; Ayres, J.G.; Sturdy, P.M.; Bland, J.M.; Butland, B.K.; Peckitt, C.; Taylor, J.C.; Victor, C.R. Bronchodilator treatment and deaths from asthma: case-control study. *Br. Med. J.* **2005**, *330*, 117–123.
166. Maringe, C.; Rickard, K.; DiSantostefano, R.; Davis, K.; Kiri, V. Concomitant use of long-acting  $\beta$ -agonists with inhaled corticosteroids among asthma patients in the UK primary care. *Eur. Respir. J.* **2007**, *30*, Suppl. 51, P3698.
167. Food and Drug Administration Center for Drug Evaluation and Research. Summary Minutes of the Pulmonary-Allergy Drugs Advisory Committee. [www.fda.gov/ohrms/dockets/ac/05/minutes/2005-4148M1\\_Final.pdf/](http://www.fda.gov/ohrms/dockets/ac/05/minutes/2005-4148M1_Final.pdf/). Last accessed: July 2008. Last updated: June 13, 2005.
168. Mann, M.; Chowdhury, B.; Sullivan, E.; Nicklas, R.; Anthracite, R.; Meyer, R.J. Serious asthma exacerbations in asthmatics treated with high-dose formoterol. *Chest* **2003**, *124*, 70–74.
169. Wolfe, J.; LaForce, C.; Friedman, B.; Sokol, W.; Till, D.; Della Cioppa, G.; van As, A. Formoterol, 24  $\mu$ g bid, and serious asthma exacerbations: similar rates compared with formoterol, 12  $\mu$ g bid, with and without extra doses taken on demand, and placebo. *Chest* **2006**, *129*, 27–38.
170. Sears, M.R.; Ottosson, A.; Radner, F.; Suissa, S. Long-acting  $\beta$ -agonists: a review of formoterol safety data from asthma clinical trials. *Eur. Respir. J.* **2009**, *33*, 21–32.
171. Battram, C.; Charlton, S.J.; Cuenoud, B.; Dowling, M.R.; Fairhurst, R.A.; Farr, D.; Fozard, J.R.; Leighton-Davies, J.R.; Lewis, C.A.; McEvoy, L.; et al. *In vitro* and *in vivo* pharmacological characterization of 5-[(R)-2-(5,6-diethyl-indan-2-ylamino)-1-hydroxy-ethyl]-8-hydroxy-1H-quinolin-2-one (indacaterol), a novel inhaled  $\beta_2$  adrenoceptor agonist with a 24-h duration of action. *J. Pharmacol. Exp. Ther.* **2006**, *317*, 762–770.

172. Sturton, R.G.; Trifilieff, A.; Nicholson, A.G.; Barnes, P.J. Pharmacological characterization of indacaterol, a novel once daily inhaled  $\beta_2$ -adrenoceptor agonist, on small airways in human and rat precision-cut lung slices. *J. Pharmacol. Exp. Ther.* **2008**, *324*, 270–275.
173. Beeh, K.M.; Derom, E.; Kanniess, F.; Cameron, R.; Higgins, M.; van As, A. Indacaterol, a novel inhaled  $\beta_2$ -agonist, provides sustained 24 h bronchodilation in asthma. *Eur. Respir. J.* **2007**, *29*, 871–878.
174. Pauwels, R.A.; Löfdahl, C.G.; Postma, D.S.; Tattersfield, A.E.; O’Byrne, P.; Barnes, P.J.; Ullman, A. Effect of inhaled formoterol and budesonide on exacerbations of asthma. Formoterol and Corticosteroids Establishing Therapy (FACET) International Study Group. *N. Engl. J. Med.* **1997**, *337*, 1405–1411.
175. Price, D.; Dutchman, D.; Mawson, A.; Bodalia, B.; Duggan, S.; Todd, P. Early asthma control and maintenance with eformoterol following reduction of inhaled corticosteroid dose. *Thorax* **2002**, *57*, 791–798.
176. Currie, G.P.; Lee, D.K.; Srivastava, P. Long-acting bronchodilator or leukotriene modifier as add-on therapy to inhaled corticosteroids in persistent asthma? *Chest* **2005**, *128*, 2954–2962.
177. FDA Public Health Advisory. Serevent Diskus (salmeterol xinafoate inhalation powder), Advair Diskus (fluticasone propionate & salmeterol inhalation powder), Foradil Aerolizer (formoterol fumarate inhalation powder). [www.fda.gov/cder/drug/advisory/LABA.htm](http://www.fda.gov/cder/drug/advisory/LABA.htm) Date last updated: May 15, 2006. Date last accessed: October 30, 2008.

© 2010 by the authors; licensee Molecular Diversity Preservation International, Basel, Switzerland. This article is an open-access article distributed under the terms and conditions of the Creative Commons Attribution license (<http://creativecommons.org/licenses/by/3.0/>).