

Review

Genetics and Molecular Pathogenesis of the Chondrosarcoma: A Review of the Literature

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Abstract: The chondrosarcoma, a cartilage-forming bone tumor, presents significant clinical challenges due to its resistance to chemotherapy and radiotherapy. Surgical excision remains the primary treatment, but high-grade chondrosarcomas are prone to recurrence and metastasis, necessitating the identification of reliable biomarkers for diagnosis and prognosis. This review explores the genetic alterations and molecular pathways involved in chondrosarcoma pathogenesis. These markers show promise in distinguishing between benign enchondromas and malignant chondrosarcomas, assessing tumor aggressiveness, and guiding treatment. While these advancements offer hope for more personalized and targeted therapeutic strategies, further clinical validation of these biomarkers is essential to improve prognostic accuracy and patient outcomes in chondrosarcoma management.

Keywords: chondrosarcoma; biomarkers; genetic alterations; molecular pathways; personalized therapy

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1. Introduction

Chondrosarcomas are malignant neoplasms of cartilage origin characterized by diverse morphological features and variable clinical behavior. They represent approximately 20% of all primary bone malignancies [\[1\]](#page-9-0), typically arising in the pelvis or long bones [\[2\]](#page-9-1). Chondrosarcomas are classified as either primary (conventional) or secondary tumors. Primary chondrosarcomas develop in previously normal bone, while secondary chondrosarcomas arise from preexisting lesions, such as enchondromas or osteochondromas [\[1\]](#page-9-0). Conventional chondrosarcomas, comprising 85–90% of cases, are categorized into central, periosteal, and peripheral subtypes [\[3\]](#page-9-2). Non-conventional variants include clear cell, mesenchymal, and dedifferentiated chondrosarcomas [\[3\]](#page-9-2). Radiographic imaging often reveals characteristic features of chondrosarcomas, allowing for a definitive diagnosis based on imaging alone.

Conventional chondrosarcoma (CHS) of the bone is the most common type of primary CHS, typically affecting an older population, with most patients over 50 years old. The peak incidence occurs between the fifth and seventh decades of life, and there is a notable male predilection, with a ratio of 1.5–2:1. CHS can involve any bone, although the incidence of axial and appendicular involvement is similar, with the pelvis, especially the ilium, frequently affected. Long tubular bones, including the proximal femur, the proximal humerus, and the distal femur, are also commonly involved. In contrast, CHS in smaller tubular bones, such as those in the forearm, clavicle, and sesamoids, is extremely rare $(1-4\%$ of all cases) [\[4\]](#page-9-3).

The histological grading of CHS is primarily based on nuclear size, hyperchromasia, cellularity, and mitoses. Low-grade (grade 1) lesions are poorly cellular with small, hyperchromatic nuclei, and they lack mitotic figures. Intermediate (grade 2) tumors are more

cellular, with nuclear enlargement and rare mitotic activity, while high-grade (grade 3) tumors show increased mitotic figures and necrosis. Genetic aberrations become more prevalent as CHS progresses from low- to high-grade, with p53 mutations and loss of INK4A/p16 expression commonly associated with high-grade CHS. For prognosis, the histological grade is the most important predictor of recurrence and metastasis, with higher-grade tumors significantly linked to a higher probability of metastasis [\[4\]](#page-9-3).

Efforts to identify reliable molecular markers and therapeutic targets for CHS have explored collagen subtypes, with types II and X, along with aggrecan, proposed as markers of a mature neoplastic phenotype, while collagen type I is associated with a proliferative, dedifferentiated state [\[5\]](#page-9-4). Cyclooxygenase-2 overexpression has been linked to a higher histologic grade and poorer survival but lacks independent prognostic value [\[6\]](#page-9-5). Attempts to inhibit CHS growth using celecoxib were unsuccessful [\[7\]](#page-10-0), although the Hedgehog signaling pathway remains a potential therapeutic target [\[8\]](#page-10-1). Activation of the IHH/PTHLH pathway and bcl2 reactivation are implicated in CHS progression, with bcl2 serving as a marker to differentiate low-grade CHS from enchondromas [\[9\]](#page-10-2). As CHS progresses from low- to high-grade, genetic aberrations increase, with p53 overexpression and TP53 mutations occurring late in high-grade cases [\[8](#page-10-1)[,10,](#page-10-3)[11\]](#page-10-4). Aberrations, such as 12q13 amplification (involving MDM2) and 9p21 loss (affecting CDKN2A/p16/INK4A), are common, with loss of p16 linked to high-grade CHS [\[12\]](#page-10-5). Prognosis is primarily determined based on the histological grade, with grade 1 tumors having an indolent course and no metastatic risk, while grades 2 and 3 are associated with metastasis and lower survival rates [\[13\]](#page-10-6). Surgical excision remains the mainstay of treatment, with wide excision for high-grade CHS and curettage for grade 1 cases yielding effective long-term control [\[14\]](#page-10-7).

The tumor microenvironment (TME) in chondrosarcomas is characterized by a dense and heterogeneous extracellular matrix (ECM) that supports tumor growth and metastasis through various signaling pathways. Unlike normal chondrocytes, chondrosarcoma cells create a compact matrix rich in type II collagen, hydrophobic proteoglycans, and hyaluronan, contributing to the tumor's structural integrity and invasive potential. ECM remodeling is regulated by matrix metalloproteinases (MMPs), with high levels of MMP-1, MMP-2, and MMP-13 correlating with increased tumor cell invasiveness, while MMP-9 expression has been linked to better survival outcomes. Additionally, cytokines like IL-1β and chemokines like CCL-5 drive tumor vascularization and growth by stimulating vascular endothelial growth factor (VEGF) release [\[15](#page-10-8)[,16\]](#page-10-9). Hypoxia, common in the chondrosarcoma TME, activates hypoxia-inducible factors (HIF-1 α and HIF-2 α), which further promote metastasis by triggering pathways involved in cancer cell dormancy, angiogenesis, and ECM remodeling, as well as contributing to chemoresistance through cancer stem cell survival [\[17\]](#page-10-10).

The immune microenvironment in chondrosarcomas also plays a crucial role in tumor progression, with tumor-associated macrophages (TAMs) representing the predominant immune cell type. Most TAMs exhibit a pro-tumoral M2-like phenotype, supporting angiogenesis, immune suppression, and tumor cell proliferation while suppressing CD8+ cytotoxic T-cell activity through cytokines and reactive metabolites. Additionally, immune cells, such as T-cells and natural killer cells, are often located at the tumor's periphery, possibly due to the dense ECM barrier. However, dedifferentiated chondrosarcomas, with less organized ECM structures, may show immune cells interspersed with tumor cells, suggesting potential responsiveness to immunotherapy in some cases. Recent studies have highlighted markers like CSF1R in TAMs as potential targets for immunomodulation. At the same time, the presence of regulatory T-cells (Tregs) and immunosuppressive factors like PD-L1 further suggests that immune checkpoint inhibitors could provide a therapeutic benefit by reversing immune evasion in chondrosarcomas [\[18,](#page-10-11)[19\]](#page-10-12).

2. Genetic Alterations in Chondrosarcomas

2.1. IDH1/IDH2 Mutations

In recent years, recurrent heterozygous hotspot mutations in isocitrate dehydrogenase 1 and 2 (IDH1 and IDH2) genes, specifically at residues p.R132 and p.R140/p.R172, respectively, have been frequently identified in cartilage tumors, such as enchondromas, central conventional chondrosarcomas, and dedifferentiated chondrosarcomas. These mutations are found in 87% of enchondromas, around 50% of central conventional chondrosarcomas, and over 80% of dedifferentiated chondrosarcomas [\[20\]](#page-10-13). The early occurrence of IDH mutations suggests that they play a pivotal role in tumorigenesis by promoting chondrogenic differentiation while inhibiting osteogenic pathways in mesenchymal stem cells, the likely progenitors of these tumors [\[21,](#page-10-14)[22\]](#page-10-15). Despite their significance in the early stages of tumor formation, the prognostic implications of IDH mutations in chondrosarcomas remain controversial. Some studies suggest that IDH mutations do not affect outcomes [\[23\]](#page-10-16), while others report either worse [\[24\]](#page-10-17) or better [\[25\]](#page-10-18) prognoses in patients with IDH-mutant chondrosarcomas. This variability could be due to differences in chondrosarcoma subtypes, follow-up periods, and detection techniques, such as Sanger sequencing, which might not capture mutations in samples with low variant allele frequency.

IDH1 and IDH2 mutations play a significant role in the progression and molecular characteristics of chondrosarcomas, as identified in recent multi-omics studies. These mutations, often found in cartilage tumors, particularly in central chondrosarcomas, lead to widespread hypermethylation across the genome. This hypermethylation affects key cellular processes, including differentiation and proliferation. Interestingly, different IDH mutations are associated with varying tumor aggressiveness, with IDH2 mutations (R172S/W/T) notably linked to more advanced and dedifferentiated forms of chondrosarcoma. Despite these molecular insights, the presence of IDH mutations does not directly correlate with a worse overall prognosis in all cases. However, tumors with dedifferentiated histology, which often display these mutations, tend to have a poorer outcome, indicating that IDH mutations contribute to more aggressive tumor phenotypes [\[26\]](#page-10-19).

IDH mutations are identified in 52–59% of central chondrosarcomas (CSs) and 57% of dedifferentiated CSs [\[27\]](#page-10-20). The occurrence of IDH mutations in both benign enchondromas and malignant CSs suggests that these mutations are an early event, indicating that cartilaginous neoplasms may exist on a spectrum of malignant potential. Additionally, IDH mutations are also observed in gliomas, acute myeloid leukemia (AML), and cholangiocarcinomas [\[28\]](#page-10-21). IDH, an enzyme in tricarboxylic acid (Krebs cycle), normally converts isocitrate to α -ketoglutarate (α -KG). However, mutant IDH (mIDH) loses the ability to perform this conversion and acquires a new function that results in the accumulation of δ-2-hydroxyglutarate (D2HG). D2HG is considered an oncometabolite because it inhibits α-KG-dependent dioxygenases, which are critical for DNA and histone demethylation, leading to a hypermethylated state in both DNA and histones [\[29\]](#page-10-22). Nevertheless, IDH mutations alone are insufficient for malignant transformation, similarly to the loss of EXT. Targeting oncogenic mIDH1/2 offers a potential therapeutic approach, with several inhibitors currently being tested in clinical trials for patients with AML or solid tumors, including CS [\[30\]](#page-10-23).

2.2. COL21A

The *COL2A1* gene, which is responsible for encoding the alpha-1 chain of type II collagen, plays a pivotal role in developing chondrosarcomas (ChSs). Mutations in *COL2A1* are commonly observed in ChS, especially in higher-grade tumors, where they cause significant disruptions in the collagen structure and the extracellular matrix (ECM). These alterations in ECM deposition and signaling can drive tumor progression by impairing normal cartilage differentiation processes, potentially facilitating oncogenesis. Notably, these mutations are associated with more aggressive and malignant tumor behavior. However, despite their biological importance, *COL2A1* mutations have yet to be adopted as clinical biomarkers for prognosis prediction [\[26\]](#page-10-19).

The second most common mutations in chondrosarcomas (ChSs) are found in the *COL2A1* gene, which encodes the alpha-1 chain of type II collagen fibers in cartilage. These mutations have been identified in 37% of ChS cases [\[31](#page-11-0)[,32\]](#page-11-1). Studies have revealed truncating, essential splice-site, and missense mutations in high-grade conventional ChS and dedifferentiated ChS. These mutations lead to significant disruption in extracellular matrix (ECM) deposition and signaling, contributing to oncogenesis by interfering with normal differentiation processes in cartilage tissue [\[31\]](#page-11-0). Furthermore, an analysis of a single *COL2A1* gene showed a tumor mutational burden (TMB) ranging from 1 to 115 mutations per case. Among the identified somatic mutations in this gene are missense, nonsense, splice-site, and synonymous mutations, as well as indels and substitutions in microRNAs. Notably, somatic mutations doubled in dedifferentiated ChS and grades 2 and 3 ChS compared to grade 1 ChS. Despite these findings, further detailed research is still required [\[31\]](#page-11-0).

2.3. TP53 and CDKN2A

In chondrosarcomas (ChSs), mutations in the *TP53* gene are observed in approximately 22% of cases, particularly in higher-grade tumors, such as grade 2/3 and dedifferentiated chondrosarcomas. The *TP53* gene is a tumor suppressor that maintains genomic stability by regulating cell cycle progression and apoptosis. Mutations in *TP53* often lead to the loss of its tumor-suppressing function, contributing to the malignant progression of ChS. These alterations are typically absent in well-differentiated tumors, indicating that *TP53* mutations are associated with more aggressive forms of the disease. Despite the clear role of *TP53* in tumor progression, its mutations have limited value in survival prediction or risk stratification [\[32\]](#page-11-1).

Gene aberrations are common in chondrosarcomas (ChSs), with higher grades showing more abnormalities [\[33\]](#page-11-2). Loss of heterozygosity in the 17p1 region, leading to *TP53* loss, occurs in about 25–30% of high-grade ChS [\[34,](#page-11-3)[35\]](#page-11-4), along with mutations in TP53 introns and exons [\[36\]](#page-11-5). High-grade ChS also often amplifies in the 12q13 region involving MDM2 and deletions in the 9p21 region containing *CDKN2A* [\[33\]](#page-11-2). Loss of *CDKN2A*/p16INK4A is crucial for ChS progression [\[37\]](#page-11-6), and p16INK4a hypermethylation is frequently observed in ChS.

Alterations in *CDKN2A* and *TP53* are common in high-grade ChS but have limited prognostic value [\[31,](#page-11-0)[38\]](#page-11-7). Near-haploid karyotypes have been reported, although their significance remains unclear [\[39](#page-11-8)[,40\]](#page-11-9).

In contrast to chondrosarcomas, osteosarcomas exhibit a higher degree of genomic instability, marked by frequent chromosomal rearrangements and elevated mutation rates in key cell cycle regulatory genes, such as *TP53*, *PTEN*, and *CDKN2A*/B. Disruptions in the *TP53* pathway, often through mutations and deletions, lead to uncontrolled cell proliferation. Another defining feature of osteosarcomas is the WNT/ β -catenin signaling pathway, where mutations contribute to abnormal cell differentiation and accelerated tumor growth. The IGF1/IGF2 and IGF1R pathways also play crucial roles, with upregulation fostering aggressive proliferation and tumor survival. Significant differential expression of genes like *BTNL9*, *MMP14*, *ABCA10*, *ACACB*, *COL11A1*, and *PKM2* further highlights osteosarcomas' unique molecular profile, positioning these genes as potential biomarkers. Together with targets in the IGF1R and WNT pathways and TP53-related regulators, these features reveal promising therapeutic strategies distinct from those used for chondrosarcomas [\[41](#page-11-10)[–43\]](#page-11-11).

3. Pathway Dysregulation in Chondrosarcomas

Large-scale transcriptomic and proteomic analyses, including whole-exome sequencing and immunohistochemistry, have identified frequent aberrations in tumor suppressors and cell cycle regulators in ChS cells [\[44](#page-11-12)[,45\]](#page-11-13). Key genes, including *CDK4* and *MDM2*, are implicated in the pRb and p53 pathways [\[44\]](#page-11-12). The degradation of p53 via *MDM2* has been linked to tumor progression in central ChSs, and p53 deficiency can lead to ChS arising from benign lesions like enchondroma [\[46\]](#page-11-14). With *MDM2* upregulated in one-third

of high-grade ChSs and correlating with more aggressive disease, inhibitors targeting the p53–*MDM2* interaction (e.g., RG7112) could be a promising therapy [\[47\]](#page-11-15).

The pRb pathway, disrupted in many cases of high-grade ChS, involves loss of *RB1* gene heterozygosity, reduced *CDKN2A*/p16 expression, or increased *CDK4* or cyclin D1 expression [\[44,](#page-11-12)[45\]](#page-11-13). *CDK4/6* inhibitors (palbociclib, ribociclib, abemaciclib), already approved for metastatic breast cancer, are considered potential treatments for advanced ChS [\[48\]](#page-11-16).

The SRC signaling cascade, crucial in sarcoma survival, migration, and proliferation, regulates PI3K–AKT signaling. Dasatinib (BMS-354825), a tyrosine kinase inhibitor targeting SRC and ABL, has shown efficacy in reducing ChS cell viability, although it did not induce caspase-3-mediated apoptosis [\[49](#page-11-17)[,50\]](#page-11-18). Dasatinib also sensitized mutant p53 ChS cells to doxorubicin, inhibiting migration and inducing apoptosis in vitro [\[50\]](#page-11-18). However, despite the potential of SRC pathway inhibition, a phase II trial of dasatinib in high-grade sarcoma patients showed no clinical benefit [\[51\]](#page-11-19).

The Hedgehog (HH) pathway is essential for embryogenesis and adult tissue maintenance. In mammals, Indian Hedgehog (IHH) binds to Patched (PTCH1), activating Smoothened (SMO). This triggers GLI transcription factors, leading to various cellular responses, like survival and differentiation [\[52\]](#page-11-20).

Chondrogenesis is regulated by the IHH/PTHrP pathway, where IHH promotes chondrocyte division and PTHrP inhibits differentiation in a feedback loop. Given its role in chondrogenesis, targeting HH signaling in chondrosarcomas (CSs) has been explored. SMO inhibitors like HPI-4 and IPI-926 showed efficacy in preclinical models, but clinical trials were unsuccessful [\[53–](#page-11-21)[55\]](#page-12-0). Despite setbacks, more potent inhibitors are in development [\[52\]](#page-11-20).

RTK activation triggers PI3K, leading to Akt activation, which regulates survival and growth. PTEN mutations are rare in CS, but active Akt signaling is present. mTOR kinase, which integrates PI3K/Akt signals, activated 69% of conventional and 44% of dedifferentiated CS samples [\[56\]](#page-12-1). The dual PI3K/mTOR inhibitor BEZ235 induced G1 arrest in CS cell lines without causing apoptosis [\[56\]](#page-12-1). Clinically, sirolimus combined with low-dose cyclophosphamide stabilized disease in 60% of patients with advanced CS for at least 6 months [\[57\]](#page-12-2). Further research into PI3K–Akt–mTOR pathway inhibition in CS is needed.

4. Epigenetic Changes in Chondrosarcomas

DNA methylation, the addition of a methyl group to DNA catalyzed by DNA methyltransferases (DNMTs), plays a significant role in gene regulation [\[58,](#page-12-3)[59\]](#page-12-4). Hypomethylation, the loss of this methyl group, can occur globally or in specific genes and is involved in tumor progression, as seen in repetitive DNA sequences like Satellite 1 and L1 [\[60,](#page-12-5)[61\]](#page-12-6). DNMT inhibitors, such as azacytidine and decitabine (DAC), are FDA-approved for treating hematological conditions like acute myeloid leukemia (AML) and myelodysplastic syndrome (MDS) [\[62\]](#page-12-7). In chondrosarcomas (CSs), DAC treatment has been shown to hypomethylate Satellite 1 and L1 and overexpress genes like Sox-2 and midkine (MDK), leading to increased tumor invasiveness in some models while inhibiting growth in others [\[63](#page-12-8)[,64\]](#page-12-9). DAC have also been associated with decreased methylation at CpG sites, leading to increased expression of epithelial markers like Maspin and 14-3-3σ, proteins involved in cell adhesion, apoptosis, and cell cycle control, suggesting a role in mesenchymal to epithelial transition (MET) [\[65\]](#page-12-10). Additionally, the tumor microenvironment can influence methylation patterns, as studies in SRC cells transplanted into different sites showed variations in gene expression and hypomethylation compared with normal cartilage [\[66\]](#page-12-11). While DAC show promise in some contexts, their role in promoting or inhibiting CS progression remains complex and warrants further research [\[66\]](#page-12-11).

DNA methylation, adding a methyl group to CpG sites in DNA, regulates gene expression and plays a significant role in tumorigenesis, including in cancers like lung, breast, liver, and colon cancer, as well as melanoma and glioma [\[67\]](#page-12-12). Hypermethylation, in particular, is associated with mutations in genes like IDH1 and IDH2, which are frequently altered in

cancers, such as acute myeloid leukemia, glioma, and chondrosarcomas (CSs) [\[68\]](#page-12-13). Mutant IDH leads to the production of the oncometabolite D2HG, which inhibits α -KG-dependent enzymes, causing DNA hypermethylation and histone modification, ultimately contributing to malignancy [\[69\]](#page-12-14). Studies have linked IDH mutations in CS to hypermethylation of genes involved in stem cell maintenance and differentiation, with potential therapeutic targets identified, including TET enzymes, Aurora kinase, and HDAC inhibitors [\[70\]](#page-12-15). Furthermore, hypermethylation of key genes like p16INK4, E-cadherin, and NAMPT has been observed in CS, suggesting potential for NAMPT and NAPRT inhibitors in high-grade CS treatment [\[71\]](#page-12-16). RUNX3, a tumor suppressor gene, is often silenced through hypermethylation in CS, and restoring its expression has been shown to inhibit proliferation and induce apoptosis [\[72\]](#page-12-17). Additionally, hypermethylation of p73, a member of the p53 tumor suppressor family, correlates with CS progression, making it a potential prognostic marker and therapeutic target [\[73\]](#page-12-18).

MicroRNAs (miRNAs) are small, noncoding RNAs (18–24 nucleotides) that regulate gene expression by binding to the 3′ untranslated region (3′ UTR) of target mRNAs. Each miRNA can target multiple mRNAs, with around 600 miRNAs regulating about 60% of human genes. miRNAs are involved in normal chondrogenesis, such as miR-140, which regulates HDAC4 to promote chondrocyte hypertrophy via RUNX2. In chondrosarcomas (CSs), miRNAs also influence oncogenesis. For example, miR-100 suppresses tumors by targeting mTOR but is downregulated in CS, while miR-30a inhibits tumor growth by targeting SOX4. miR-181a, an oncogene, promotes VEGF expression, aiding CS progression [\[32\]](#page-11-1) (Table [1\)](#page-5-0).

Table 1. Overview of the mechanisms and potential therapeutic implications of DNA methylation and miRNAs and their roles in chondrosarcoma progression.

Table 1. *Cont.*

5. Targeted Therapies in Chondrosarcomas

Chemotherapy is generally ineffective for chondrosarcomas, and no standard systemic treatment exists for conventional cases due to factors like slow proliferation, multidrug resistance, high expression of anti-apoptotic proteins, and poor vascularity [\[74\]](#page-12-19). Clinical studies are challenging due to the rarity of the disease. Still, a retrospective study of nivolumab, with or without pazopanib, showed partial responses in 13% of sarcoma patients, including one with a dedifferentiated chondrosarcoma [\[75\]](#page-12-20). IL-8, a chemokine involved in tumor progression, was targeted in a phase 1 trial using BMS-986253, yielding a high disease control rate but no objective response, indicating a need for further research in chondrosarcoma patients [\[76\]](#page-12-21).

Recent studies have identified potential therapeutic targets in chondrosarcomas. PPARγ activation has been shown to inhibit cell proliferation and induce apoptosis, with zaltoprofen demonstrating efficacy in activating PPARγ in chondrosarcoma cells [\[77,](#page-12-22)[78\]](#page-13-0). CDK4 is highly expressed in chondrosarcomas and associated with poor prognosis. The CDK4 inhibitor palbociclib has shown promising results in halting cell cycle progression and inhibiting tumor cell proliferation and invasion [\[79\]](#page-13-1).

Other promising treatments include MLN4924, a NEDD8-activating enzyme inhibitor, which has shown antitumor effects in both cell lines and a xenograft mouse model [\[80\]](#page-13-2), and resveratrol, which inhibited cell viability and suppressed the STAT3 signaling pathway in chondrosarcoma cells [\[81\]](#page-13-3). The mTOR inhibitors rapamycin and everolimus have also demonstrated potential by reducing cell viability and suppressing tumor progression in both cell lines and animal models [\[82,](#page-13-4)[83\]](#page-13-5).

Despite limited advances in chondrosarcoma treatment, recent research has identified promising molecular targets for therapy. Activating peroxisome proliferator-activated receptor gamma (PPARγ) in chondrosarcoma cells—either directly or with drugs like zaltoprofen—has been shown to inhibit cell proliferation, induce apoptosis, and reduce invasion, marking it as a viable therapeutic pathway. Similarly, CDK4, linked to cell cycle regulation and associated with a poor prognosis in chondrosarcoma, can be targeted by palbociclib or CDK4-specific siRNA, which significantly reduces tumor cell proliferation and spread via cell cycle arrest. Inhibiting NEDD8, a ubiquitin-like protein, with MLN4924 also shows antitumor promise by promoting apoptosis and blocking tumor

growth in preclinical models. The phytochemical resveratrol has also demonstrated tumorsuppressive effects by reducing cell viability and inhibiting STAT3 signaling. Targeting the mTOR pathway with inhibitors like rapamycin and everolimus further reduces tumor metabolism and progression, underscoring their potential to enhance chondrosarcoma treatment outcomes [\[84\]](#page-13-6).

Immunotherapy in chondrosarcomas is emerging as a promising avenue of treatment, particularly for aggressive and metastatic forms of the disease, which are often resistant to conventional therapies, such as chemotherapy and radiation. Chondrosarcomas (CSs) exhibit several features, such as poor vascularization and the presence of multidrug-resistant pumps, that contribute to this resistance. However, immune checkpoint inhibitors (ICIs) like pembrolizumab and nivolumab have shown potential in clinical trials by targeting proteins, such as PD-1 and PD-L1, which are involved in tumor immune evasion. Early studies suggest that while response rates to immunotherapy in chondrosarcomas are still low, some patients with high-grade or dedifferentiated tumors have shown significant tumor regression [\[85\]](#page-13-7) (Table [2\)](#page-7-0).

Therapeutic Approach Mechanism/Target Findings Study/Notes Chemotherapy Ineffective Generally ineffective due to slow proliferation, multidrug resistance, and poor vascularity No standard systemic treatment for conventional chondrosarcomas Nivolumab (±Pazopanib) Immune checkpoint inhibition 13% partial response rate in sarcoma patients, including dedifferentiated chondrosarcomas Retrospective study BMS-986253 Targets IL-8 (chemokine involved in tumor progression) High disease control rate, but no objective response

Phase 1 trial

Phase 1 trial Zaltoprofen Activates PPAR_γ Inhibits cell proliferation and induces apoptosis in chondrosarcoma cells Efficacy demonstrated in preclinical studies Palbociclib CDK4 inhibitor Inhibits cell cycle progression, tumor cell proliferation, and invasion CDK4 is highly expressed and associated with poor prognosis in chondrosarcomas MLN4924 NEDD8-activating enzyme inhibitor Demonstrated antitumor effects in cell lines and xenograft mouse models Preclinical study Resveratrol Inhibits STAT3 signaling pathway Reduced cell viability and suppressed tumor progression in chondrosarcoma cells Preclinical study Rapamycin/Everolimus mTOR inhibitors Reduced cell viability and tumor progression in both cell lines and animal models Preclinical studies Immunotherapy (Pembrolizumab, Nivolumab) Targets immune checkpoint proteins (PD-1, PD-L1) Early studies show low response rates but significant tumor regression in some high-grade cases Promising for aggressive/metastatic chondrosarcomas resistant to conventional therapies

Table 2. Potential treatments and outcomes in chondrosarcomas.

Additionally, CDK4, a protein linked to cell cycle regulation, is highly expressed in chondrosarcomas and associated with metastasis and poor prognosis. Targeting CDK4 with palbociclib or CDK4-specific siRNA has demonstrated substantial reductions in chondrosarcoma cell proliferation, migration, and invasion by inducing cell cycle arrest in the G1 phase, mainly through the CDK4/Rb signaling axis. Furthermore, inhibiting the

ubiquitin-like protein NEDD8 with MLN4924 has shown promising antitumor effects by reducing cell proliferation and enhancing apoptosis, even inhibiting tumor growth in xenograft models [\[79\]](#page-13-1). Other therapeutic targets include resveratrol, a phytochemical known for its antitumor effects across various cancers. In chondrosarcomas, resveratrol has been observed to decrease cell viability and proliferation, promote apoptosis, and inhibit STAT3 signaling, contributing to tumor suppression. The mTOR pathway, critical in cell growth and survival, has also been identified as a therapeutic target, with inhibitors like rapamycin and everolimus reducing cell metabolism, Glut1 and HIF1 α expression, and overall tumor progression. These inhibitors have shown effectiveness in preclinical chondrosarcoma models, underscoring the mTOR pathway's role in potentially improving treatment outcomes for this challenging malignancy [\[81\]](#page-13-3).

6. Prognostic Biomarkers and Molecular Diagnostics

Surgery is the preferred treatment for chondrosarcomas, as these tumors are typically resistant to chemotherapy and radiation therapy. Patients with high-grade chondrosarcomas frequently experience local recurrence and distant metastasis following surgical resection. Wide excision is recommended for curative treatment to prevent recurrence and metastasis; however, this approach often results in functional impairments for patients [\[86,](#page-13-8)[87\]](#page-13-9). In contrast, conservative excision is usually sufficient for patients with enchondromas, as these tumors rarely recur or metastasize [\[88\]](#page-13-10). Nevertheless, patients with low-grade chondrosarcomas may still face recurrence and metastasis after conservative excision. Thus, there is a pressing need to identify highly accurate and specific prognostic biomarkers to guide clinical management, assess tumor aggressiveness, and predict disease prognosis in these patients.

Biomarkers are essential in managing chondrosarcomas by facilitating screening, diagnosis, prognosis prediction, and monitoring of tumor progression. IDH1/2 mutations, found in approximately 39% of chondrosarcoma cases, are strongly correlated with poor survival, larger tumor size, higher grade, and increased relapse risk, positioning them as crucial prognostic indicators. Similarly, the overexpression of EphA2, a receptor implicated in angiogenesis and metastasis, has been observed in chondrosarcomas and related bone sarcomas, showing promise as a therapeutic target through its significant response to receptor inhibition [\[89\]](#page-13-11). Proteins involved in post-translational modification, such as SUMO2/3, are also emerging as prognostic biomarkers; their high expression levels are linked with worse overall survival, and inhibition of SUMO pathways has shown antitumor effects in preclinical studies [\[90\]](#page-13-12). Other markers, including esRAGE and HMGB1, correlate with tumor recurrence and poor prognosis in lower-grade chondrosarcomas. At the same time, elevated levels of Aurora kinases and hypoxia-inducible factors (HIF-1 α and HIF-2 α) are associated with higher-grade tumors and worse clinical outcomes. Collectively, these biomarkers enhance diagnostic precision and open new avenues for targeted therapies in chondrosarcoma management [\[84](#page-13-6)[,91\]](#page-13-13).

Several biomarkers show promise for improving chondrosarcoma diagnosis and prognosis. VEGF-A and VEGF-C, key players in angiogenesis and lymphangiogenesis, are upregulated in chondrosarcomas and may be useful for staging. AMACR helps differentiate between benign enchondromas and malignant chondrosarcomas, while periostin is present in low-grade chondrosarcomas but not in enchondromas, aiding diagnosis. miRNAs like miR-27b and miR-624–3p, which regulate VEGF-C, offer insight into tumor progression and potential therapeutic targets. However, these biomarkers need further clinical validation to confirm their effectiveness in chondrosarcoma management [\[92\]](#page-13-14).

Additionally, microRNAs (miRNAs) play a crucial role in chondrosarcoma pathogenesis, with their dysregulation driving tumor growth and metastasis and influencing patient prognosis. Certain miRNAs, like miR-143-3p and miR-145-5p, are downregulated in chondrosarcomas, which leads to the upregulation of target genes, such as Fascin-1, thus promoting cell migration and metastasis. This relationship underscores the value of these miRNAs as biomarkers for disease progression, as their altered levels are associated

with increased tumor aggressiveness. Additionally, low levels of miR-335, which usually suppresses metastasis through targets like SOX4 and Tenascin-C, correlate with poorer survival, further supporting its prognostic significance. MiRNAs, such as miR-21-5p and miR-454-3p, also influence pathways crucial for tumor proliferation and survival, including the STAT3/NF-κB axis, whose activation drives chondrosarcoma progression. By suppressing CCR7, miR-21-5p reduces STAT3 signaling, highlighting a potential therapeutic target to curb tumor growth. Restoring these downregulated miRNAs could help inhibit tumor progression and enhance the effectiveness of therapeutic interventions, making miRNAs

both valuable biomarkers and potential targets for chondrosarcoma treatment [\[92](#page-13-14)[,93\]](#page-13-15). In chondrosarcomas, FFPE-based molecular analysis shows promise in identifying novel biomarkers and therapeutic targets. FFPE analysis can help pinpoint differentially expressed genes linked to tumor progression and metastasis by comparing gene expression profiles between tumor and normal tissues. This approach may also aid in identifying chondrosarcoma-specific markers within its tumor microenvironment or pathways integral to its development. Additionally, FFPE data could facilitate meta-analyses and comparisons with other bone cancers, like osteosarcoma, offering a deeper molecular understanding of chondrosarcomas and supporting the development of targeted therapies [\[94\]](#page-13-16).

Additionally, in a study conducted by Tudor et al., polyethylene glycol-encapsulated iron oxide nanoparticles (IONPDOX) demonstrated potential as hyperspectral biomarkers for assessing radiosensitivity in human chondrosarcoma cells. By combining IONPDOX with carbon ion or photon radiation, researchers observed enhanced cytotoxic effects, including increased DNA damage marked by micronucleus formation and distinct changes in the hyperspectral profiles of cell nuclei. These findings suggest that IONPDOX could serve as a valuable biomarker in evaluating and improving the effectiveness of radiotherapy in highly resistant tumors like chondrosarcomas [\[95\]](#page-13-17).

7. Conclusions

In conclusion, identifying genetic alterations and molecular mechanisms underlying chondrosarcomas has paved the way for developing targeted therapies and prognostic biomarkers. Chondrosarcomas, largely resistant to chemotherapy and radiotherapy, present significant treatment challenges, underscoring the importance of biomarkers like VEGF, leptin, adiponectin, and periostin. These markers help distinguish between benign and malignant lesions and offer insights into tumor progression and metastasis. Although promising, these biomarkers require further clinical validation to ensure their reliability in guiding treatment decisions and improving patient outcomes. Continued research into the molecular pathogenesis of chondrosarcomas is essential for advancing personalized therapeutic strategies and enhancing prognostic accuracy.

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References

- 1. Fletcher, C.D.M. (Ed.) Pathology and genetics of tumours of soft tissue and bone. In *World Health Organization Classification of Tumours*; IARC Press: Lyon, France, 2002; No. 4.
- 2. Jemal, A.; Siegel, R.; Ward, E.; Murray, T.; Xu, J.; Smigal, C.; Thun, M.J. Cancer Statistics, 2006. *CA A Cancer J. Clin.* **2006**, *56*, 106–130. [\[CrossRef\]](https://doi.org/10.3322/canjclin.56.2.106) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/16514137)
- 3. Murphey, M.D.; Walker, E.A.; Wilson, A.J.; Kransdorf, M.J.; Temple, H.T.; Gannon, F.H. From the Archives of the AFIP. *Radio-Graphics* **2003**, *23*, 1245–1278. [\[CrossRef\]](https://doi.org/10.1148/rg.235035134) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/12975513)
- 4. Kim, M.-J.; Cho, K.-J.; Ayala, A.G.; Ro, J.Y. Chondrosarcoma: With Updates on Molecular Genetics. *Sarcoma* **2011**, *2011*, 405437. [\[CrossRef\]](https://doi.org/10.1155/2011/405437)
- 5. A Chow, W. Update on chondrosarcomas. *Curr. Opin. Oncol.* **2007**, *19*, 371–376. [\[CrossRef\]](https://doi.org/10.1097/CCO.0b013e32812143d9) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/17545802)
- 6. Endo, M.; Matsumura, T.; Yamaguchi, T.; Yamaguchi, U.; Morimoto, Y.; Nakatani, F.; Kawai, A.; Chuman, H.; Beppu, Y.; Shimoda, T.; et al. Cyclooxygenase-2 overexpression associated with a poor prognosis in chondrosarcomas. *Hum. Pathol.* **2006**, *37*, 471–476. [\[CrossRef\]](https://doi.org/10.1016/j.humpath.2005.12.001)
- 7. Schrage, Y.; Machado, I.; Meijer, D.; Bruijn, I.B.-D.; Akker, B.v.D.; Taminiau, A.; Kalinski, T.; Llombart-Bosch, A.; Bovée, J. COX-2 expression in chondrosarcoma: A role for celecoxib treatment? *Eur. J. Cancer* **2010**, *46*, 616–624. [\[CrossRef\]](https://doi.org/10.1016/j.ejca.2009.11.002)
- 8. Bovée, J.V.M.G.; Hogendoorn, P.C.W.; Wunder, J.S.; Alman, B.A. Cartilage tumours and bone development: Molecular pathology and possible therapeutic targets. *Nat. Rev. Cancer* **2010**, *10*, 481–488. [\[CrossRef\]](https://doi.org/10.1038/nrc2869)
- 9. Rozeman, L.B.; Hameetman, L.; Cleton-Jansen, A.-M.; Taminiau, A.H.; Hogendoorn, P.C.; Bovée, J.V. Absence of IHH and retention of PTHrP signalling in enchondromas and central chondrosarcomas. *J. Pathol.* **2005**, *205*, 476–482. [\[CrossRef\]](https://doi.org/10.1002/path.1723)
- 10. Terek, R.M.; Healey, J.H.; Garin-Chesa, P.; Mak, S.; Albino, A.P.; Huvos, A. p53 Mutations in Chondrosarcoma. *Diagn. Mol. Pathol.* **1998**, *7*, 51–56. [\[CrossRef\]](https://doi.org/10.1097/00019606-199802000-00009)
- 11. Papachristou, D.J.; Goodman, M.A.; Cieply, K.; Hunt, J.L.; Rao, U.N. Comparison of allelic losses in chondroblastoma and primary chondrosarcoma of bone and correlation with fluorescence in situ hybridization analysis. *Hum. Pathol.* **2006**, *37*, 890–898. [\[CrossRef\]](https://doi.org/10.1016/j.humpath.2006.02.014)
- 12. van Beerendonk, H.M.; Rozeman, L.B.; Taminiau, A.H.; Sciot, R.; Bovée, J.V.; Cleton-Jansen, A.; Hogendoorn, P.C. Molecular analysis of the INK4A/INK4A-ARF gene locus in conventional (central) chondrosarcomas and enchondromas: Indication of an important gene for tumour progression. *J. Pathol.* **2004**, *202*, 359–366. [\[CrossRef\]](https://doi.org/10.1002/path.1517) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/14991902)
- 13. Björnsson, J.; McLeod, R.A.; Unni, K.K.; Ilstrup, D.M.; Pritchard, D.J. Primary chondrosarcoma of long bones and limb girdles. *Cancer* **1998**, *83*, 2105–2119. [\[CrossRef\]](https://doi.org/10.1002/(SICI)1097-0142(19981115)83:10%3C2105::AID-CNCR9%3E3.0.CO;2-U)
- 14. Gelderblom, H.; Hogendoorn, P.C.; Dijkstra, S.D.; van Rijswijk, C.S.; Krol, A.D.; Taminiau, A.H.; Bovée, J.V. The Clinical Approach Towards Chondrosarcoma. *Oncol.* **2008**, *13*, 320–329. [\[CrossRef\]](https://doi.org/10.1634/theoncologist.2007-0237) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/18378543)
- 15. Walter, S.G.; Knöll, P.; Eysel, P.; Quaas, A.; Gaisendrees, C.; Nißler, R.; Hieggelke, L. Molecular In-Depth Characterization of Chondrosarcoma for Current and Future Targeted Therapies. *Cancers* **2023**, *15*, 2556. [\[CrossRef\]](https://doi.org/10.3390/cancers15092556)
- 16. Hua, H.; Li, M.; Luo, T.; Yin, Y.; Jiang, Y. Matrix metalloproteinases in tumorigenesis: An evolving paradigm. *Cell. Mol. Life Sci.* **2011**, *68*, 3853–3868. [\[CrossRef\]](https://doi.org/10.1007/s00018-011-0763-x)
- 17. Sun, X.; Lv, X.; Yan, Y.; Zhao, Y.; Ma, R.; He, M.; Wei, M. Hypoxia-mediated cancer stem cell resistance and targeted therapy. *Biomed. Pharmacother.* **2020**, *130*, 110623. [\[CrossRef\]](https://doi.org/10.1016/j.biopha.2020.110623)
- 18. Pan, Y.; Yu, Y.; Wang, X.; Zhang, T. Tumor-Associated Macrophages in Tumor Immunity. *Front. Immunol.* **2020**, *11*, 583084. [\[CrossRef\]](https://doi.org/10.3389/fimmu.2020.583084)
- 19. Simard, F.A.; Richert, I.; Vandermoeten, A.; Decouvelaere, A.-V.; Michot, J.-P.; Caux, C.; Blay, J.-Y.; Dutour, A. Description of the immune microenvironment of chondrosarcoma and contribution to progression. *OncoImmunology* **2017**, *6*, e1265716. [\[CrossRef\]](https://doi.org/10.1080/2162402X.2016.1265716)
- 20. Chen, S.; Fritchie, K.; Wei, S.; Ali, N.; Curless, K.; Shen, T.; Brini, A.T.; Latif, F.; Sumathi, V.; Siegal, G.P.; et al. Diagnostic utility of IDH1/2 mutations to distinguish dedifferentiated chondrosarcoma from undifferentiated pleomorphic sarcoma of bone. *Hum. Pathol.* **2017**, *65*, 239–246. [\[CrossRef\]](https://doi.org/10.1016/j.humpath.2017.05.015)
- 21. Jin, Y.; Elalaf, H.; Watanabe, M.; Tamaki, S.; Hineno, S.; Matsunaga, K.; Woltjen, K.; Kobayashi, Y.; Nagata, S.; Ikeya, M.; et al. Mutant IDH1 Dysregulates the Differentiation of Mesenchymal Stem Cells in Association with Gene-Specific Histone Modifications to Cartilage- and Bone-Related Genes. *PLoS ONE* **2015**, *10*, e0131998. [\[CrossRef\]](https://doi.org/10.1371/journal.pone.0131998)
- 22. Suijker, J.; Baelde, H.J.; Roelofs, H.; Cleton-Jansen, A.-M.; Bovée, J.V. The oncometabolite D-2-hydroxyglutarate induced by mutant IDH1 or -2 blocks osteoblast differentiation in vitro and in vivo. *Oncotarget* **2015**, *6*, 14832–14842. [\[CrossRef\]](https://doi.org/10.18632/oncotarget.4024) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/26046462)
- 23. Cleven, A.H.G.; Suijker, J.; Agrogiannis, G.; Bruijn, I.H.B.-D.; Frizzell, N.; Hoekstra, A.S.; Wijers-Koster, P.M.; Cleton-Jansen, A.-M.; Bovée, J.V.M.G. IDH1 or -2 mutations do not predict outcome and do not cause loss of 5-hydroxymethylcytosine or altered histone modifications in central chondrosarcomas. *Clin. Sarcoma Res.* **2017**, *7*, 8. [\[CrossRef\]](https://doi.org/10.1186/s13569-017-0074-6) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/28484589)
- 24. Lugowska, I.; Teterycz, P.; Mikula, M.; Kulecka, M.; Kluska, A.; Balabas, A.; Piatkowska, M.; Wagrodzki, M.; Pienkowski, A.; Rutkowski, P.; et al. IDH1/2 Mutations Predict Shorter Survival in Chondrosarcoma. *J. Cancer* **2018**, *9*, 998–1005. [\[CrossRef\]](https://doi.org/10.7150/jca.22915)
- 25. Zhu, G.G.; Nafa, K.; Agaram, N.; Zehir, A.; Benayed, R.; Sadowska, J.; Borsu, L.; Kelly, C.; Tap, W.D.; Fabbri, N.; et al. Genomic Profiling Identifies Association of *IDH1/IDH2* Mutation with Longer Relapse-Free and Metastasis-Free Survival in High-Grade Chondrosarcoma. *Clin. Cancer Res.* **2020**, *26*, 419–427. [\[CrossRef\]](https://doi.org/10.1158/1078-0432.CCR-18-4212)
- 26. Nicolle, R.; Ayadi, M.; Gomez-Brouchet, A.; Armenoult, L.; Banneau, G.; Elarouci, N.; Tallegas, M.; Decouvelaere, A.-V.; Aubert, S.; Rédini, F.; et al. Integrated molecular characterization of chondrosarcoma reveals critical determinants of disease progression. *Nat. Commun.* **2019**, *10*, 4622. [\[CrossRef\]](https://doi.org/10.1038/s41467-019-12525-7)
- 27. Amary, M.F.; Bacsi, K.; Maggiani, F.; Damato, S.; Halai, D.; Berisha, F.; Pollock, R.; O'Donnell, P.; Grigoriadis, A.; Diss, T.; et al. *IDH1* and *IDH2* mutations are frequent events in central chondrosarcoma and central and periosteal chondromas but not in other mesenchymal tumours. *J. Pathol.* **2011**, *224*, 334–343. [\[CrossRef\]](https://doi.org/10.1002/path.2913)
- 28. Schaap, F.G.; French, P.J.; Bovée, J.V.M.G. Mutations in the Isocitrate Dehydrogenase Genes IDH1 and IDH2 in Tumors. *Adv. Anat. Pathol.* **2013**, *20*, 32–38. [\[CrossRef\]](https://doi.org/10.1097/PAP.0b013e31827b654d) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/23232569)
- 29. Dang, L.; White, D.W.; Gross, S.; Bennett, B.D.; Bittinger, M.A.; Driggers, E.M.; Fantin, V.R.; Jang, H.G.; Jin, S.; Keenan, M.C.; et al. Cancer-associated IDH1 mutations produce 2-hydroxyglutarate. *Nature* **2009**, *462*, 739–744, Erratum in *Nature* **2010**, *465*, 966. [\[CrossRef\]](https://doi.org/10.1038/nature08617) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/19935646)
- 30. Urban, D.J.; Martinez, N.J.; Davis, M.I.; Brimacombe, K.R.; Cheff, D.M.; Lee, T.D.; Henderson, M.J.; Titus, S.A.; Pragani, R.; Rohde, J.M.; et al. Assessing inhibitors of mutant isocitrate dehydrogenase using a suite of pre-clinical discovery assays. *Sci. Rep.* **2017**, *7*, 12758. [\[CrossRef\]](https://doi.org/10.1038/s41598-017-12630-x)
- 31. Tarpey, P.S.; Behjati, S.; Cooke, S.L.; Van Loo, P.; Wedge, D.C.; Pillay, N.; Marshall, J.; O'Meara, S.; Davies, H.; Nik-Zainal, S.; et al. Frequent mutation of the major cartilage collagen gene COL2A1 in chondrosarcoma. *Nat. Genet.* **2013**, *45*, 923–926. [\[CrossRef\]](https://doi.org/10.1038/ng.2668)
- 32. Chow, W.A. Chondrosarcoma: Biology, genetics, and epigenetics. *F1000Research* **2018**, *7*, 1826. [\[CrossRef\]](https://doi.org/10.12688/f1000research.15953.1) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/30519452)
- 33. Italiano, A.; Mir, O.; Cioffi, A.; Palmerini, E.; Piperno-Neumann, S.; Perrin, C.; Chaigneau, L.; Penel, N.; Duffaud, F.; Kurtz, J.E.; et al. Advanced chondrosarcomas: Role of chemotherapy and survival. *Ann. Oncol.* **2013**, *24*, 2916–2922. [\[CrossRef\]](https://doi.org/10.1093/annonc/mdt374) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/24099780)
- 34. Tawbi, H.A.; Burgess, M.; Bolejack, V.; Van Tine, B.A.; Schuetze, S.M.; Hu, J.; D'Angelo, S.; Attia, S.; Riedel, R.F.; Priebat, D.A.; et al. Pembrolizumab in advanced soft-tissue sarcoma and bone sarcoma (SARC028): A multicentre, two-cohort, single-arm, open-label, phase 2 trial. *Lancet Oncol.* **2017**, *18*, 1493–1501. [\[CrossRef\]](https://doi.org/10.1016/S1470-2045(17)30624-1)
- 35. Chalmers, Z.R.; Connelly, C.F.; Fabrizio, D.; Gay, L.; Ali, S.M.; Ennis, R.; Schrock, A.; Campbell, B.; Shlien, A.; Chmielecki, J.; et al. Analysis of 100,000 human cancer genomes reveals the landscape of tumor mutational burden. *Genome Med.* **2017**, *9*, 34. [\[CrossRef\]](https://doi.org/10.1186/s13073-017-0424-2) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/28420421)
- 36. Toulmonde, M.; Penel, N.; Adam, J.; Chevreau, C.; Blay, J.-Y.; Le Cesne, A.; Bompas, E.; Piperno-Neumann, S.; Cousin, S.; Grellety, T.; et al. Use of PD-1 Targeting, Macrophage Infiltration, and IDO Pathway Activation in Sarcomas. *JAMA Oncol.* **2018**, *4*, 93–97. [\[CrossRef\]](https://doi.org/10.1001/jamaoncol.2017.1617)
- 37. D'Angelo, S.P.; Mahoney, M.R.; Van Tine, B.A.; Atkins, J.; Milhem, M.M.; Jahagirdar, B.N.; Antonescu, C.R.; Horvath, E.; Tap, W.D.; Schwartz, G.K.; et al. Nivolumab with or without ipilimumab treatment for metastatic sarcoma (Alliance A091401): Two open-label, non-comparative, randomised, phase 2 trials. *Lancet Oncol.* **2018**, *19*, 416–426. [\[CrossRef\]](https://doi.org/10.1016/S1470-2045(18)30006-8)
- 38. Amary, M.F.; Ye, H.; Forbes, G.; Damato, S.; Maggiani, F.; Pollock, R.; Tirabosco, R.; Flanagan, A.M. Isocitrate dehydrogenase 1 mutations (IDH1) and p16/CDKN2A copy number change in conventional chondrosarcomas. *Virchows Arch.* **2015**, *466*, 217–222. [\[CrossRef\]](https://doi.org/10.1007/s00428-014-1685-4)
- 39. Mandahl, N.; Johansson, B.; Mertens, F.; Mitelman, F. Disease-associated patterns of disomic chromosomes in hyperhaploid neoplasms. *Genes, Chromosom. Cancer* **2012**, *51*, 536–544. [\[CrossRef\]](https://doi.org/10.1002/gcc.21947)
- 40. Olsson, L.; Paulsson, K.; Bovée, J.V.M.G.; Nord, K.H. Clonal Evolution through Loss of Chromosomes and Subsequent Polyploidization in Chondrosarcoma. *PLoS ONE* **2011**, *6*, e24977. [\[CrossRef\]](https://doi.org/10.1371/annotation/8f845569-8244-416b-b15e-89562177ce32)
- 41. Reimann, E.; Kõks, S.; Ho, X.D.; Maasalu, K.; Märtson, A. Whole exome sequencing of a single osteosarcoma case—Integrative analysis with whole transcriptome RNA-seq data. *Hum. Genom.* **2014**, *8*, 20. [\[CrossRef\]](https://doi.org/10.1186/s40246-014-0020-0)
- 42. Ho, X.D.; Nguyen, H.G.; Trinh, L.H.; Reimann, E.; Prans, E.; Kõks, G.; Maasalu, K.; Le, V.Q.; Nguyen, V.H.; Le, N.T.N.; et al. Analysis of the Expression of Repetitive DNA Elements in Osteosarcoma. *Front. Genet.* **2017**, *8*, 193. [\[CrossRef\]](https://doi.org/10.3389/fgene.2017.00193) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/29250102)
- 43. Ho, X.D.; Phung, P.; Le, V.Q.; Nguyen, V.H.; Reimann, E.; Prans, E.; Kõks, G.; Maasalu, K.; Le, N.T.; Trinh, L.H.; et al. Whole transcriptome analysis identifies differentially regulated networks between osteosarcoma and normal bone samples. *Exp. Biol. Med.* **2017**, *242*, 1802–1811. [\[CrossRef\]](https://doi.org/10.1177/1535370217736512) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/29050494)
- 44. Schrage, Y.M.; Lam, S.; Jochemsen, A.G.; Cleton-Jansen, A.; Taminiau, A.H.; Hogendoorn, P.C.; Bovée, J.V. Central chondrosarcoma progression is associated with pRb pathway alterations: CDK4 down-regulation and p16 overexpression inhibit cell growth in vitro. *J. Cell. Mol. Med.* **2009**, *13*, 2843–2852. [\[CrossRef\]](https://doi.org/10.1111/j.1582-4934.2008.00406.x)
- 45. Röpke, M.; Boltze, C.; Meyer, B.; Neumann, H.W.; Roessner, A.; Schneider-Stock, R. Rb-loss is associated with high malignancy in chondrosarcoma. *Oncol. Rep.* **2006**, *15*, 89–95. [\[CrossRef\]](https://doi.org/10.3892/or.15.1.89) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/16328039)
- 46. Ho, L.; Stojanovski, A.; Whetstone, H.; Wei, Q.X.; Mau, E.; Wunder, J.S.; Alman, B. Gli2 and p53 Cooperate to Regulate IGFBP-3- Mediated Chondrocyte Apoptosis in the Progression from Benign to Malignant Cartilage Tumors. *Cancer Cell* **2009**, *16*, 126–136. [\[CrossRef\]](https://doi.org/10.1016/j.ccr.2009.05.013)
- 47. Ray-Coquard, I.; Blay, J.-Y.; Italiano, A.; Le Cesne, A.; Penel, N.; Zhi, J.; Heil, F.; Rueger, R.; Graves, B.; Ding, M.; et al. Effect of the MDM2 antagonist RG7112 on the P53 pathway in patients with MDM2-amplified, well-differentiated or dedifferentiated liposarcoma: An exploratory proof-of-mechanism study. *Lancet Oncol.* **2012**, *13*, 1133–1140. [\[CrossRef\]](https://doi.org/10.1016/S1470-2045(12)70474-6)
- 48. Roberts, P.J.; Kumarasamy, V.; Witkiewicz, A.K.; Knudsen, E.S. Chemotherapy and CDK4/6 Inhibitors: Unexpected Bedfellows. *Mol. Cancer Ther.* **2020**, *19*, 1575–1588. [\[CrossRef\]](https://doi.org/10.1158/1535-7163.MCT-18-1161) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/32546660)
- 49. Schrage, Y.M.; Bruijn, I.H.B.-D.; de Miranda, N.F.; van Oosterwijk, J.; Taminiau, A.H.; van Wezel, T.; Hogendoorn, P.C.; Bovée, J.V. Kinome Profiling of Chondrosarcoma Reveals Src-Pathway Activity and Dasatinib as Option for Treatment. *Cancer Res.* **2009**, *69*, 6216–6222. [\[CrossRef\]](https://doi.org/10.1158/0008-5472.CAN-08-4801) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/19602594)
- 50. van Oosterwijk, J.G.; van Ruler, M.A.J.H.; Bruijn, I.H.B.-D.; Herpers, B.; Gelderblom, H.; Van De Water, B.; Bovee, J.V.M.G. Src kinases in chondrosarcoma chemoresistance and migration: Dasatinib sensitises to doxorubicin in TP53 mutant cells. *Br. J. Cancer* **2013**, *109*, 1214–1222. [\[CrossRef\]](https://doi.org/10.1038/bjc.2013.451)
- 51. Schuetze, S.M.; Wathen, J.K.; Lucas, D.R.; Choy, E.; Samuels, B.L.; Staddon, A.P.; Ganjoo, K.N.; von Mehren, M.; Chow, W.A.; Loeb, D.M.; et al. SARC009: Phase 2 study of dasatinib in patients with previously treated, high-grade, advanced sarcoma. *Cancer* **2016**, *122*, 868–874. [\[CrossRef\]](https://doi.org/10.1002/cncr.29858)
- 52. Wu, F.; Zhang, Y.; Sun, B.; McMahon, A.P.; Wang, Y. Hedgehog Signaling: From Basic Biology to Cancer Therapy. *Cell Chem. Biol.* **2017**, *24*, 252–280. [\[CrossRef\]](https://doi.org/10.1016/j.chembiol.2017.02.010) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/28286127)
- 53. Italiano, A.; Le Cesne, A.; Bellera, C.; Piperno-Neumann, S.; Duffaud, F.; Penel, N.; Cassier, P.; Domont, J.; Takebe, N.; Kind, M.; et al. GDC-0449 in patients with advanced chondrosarcomas: A French Sarcoma Group/US and French National Cancer Institute Single-Arm Phase II Collaborative Study. *Ann. Oncol.* **2013**, *24*, 2922–2926. [\[CrossRef\]](https://doi.org/10.1093/annonc/mdt391) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/24170610)
- 54. Campbell, V.T.; Nadesan, P.; Ali, S.A.; Wang, C.Y.Y.; Whetstone, H.; Poon, R.; Wei, Q.; Keilty, J.; Proctor, J.; Wang, L.W.; et al. Hedgehog Pathway Inhibition in Chondrosarcoma Using the Smoothened Inhibitor IPI-926 Directly Inhibits Sarcoma Cell Growth. *Mol. Cancer Ther.* **2014**, *13*, 1259–1269. [\[CrossRef\]](https://doi.org/10.1158/1535-7163.MCT-13-0731) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/24634412)
- 55. Xiang, W.; Jiang, T.; Guo, F.; Gong, C.; Yang, K.; Wu, Y.; Huang, X.; Cheng, W.; Xu, K. Hedgehog pathway inhibitor-4 suppresses malignant properties of chondrosarcoma cells by disturbing tumor ciliogenesis. *Oncol. Rep.* **2014**, *32*, 1622–1630. [\[CrossRef\]](https://doi.org/10.3892/or.2014.3372)
- 56. Zhang, Y.-X.; van Oosterwijk, J.G.; Sicinska, E.; Moss, S.; Remillard, S.P.; van Wezel, T.; Bühnemann, C.; Hassan, A.B.; Demetri, G.D.; Bovée, J.V.; et al. Functional Profiling of Receptor Tyrosine Kinases and Downstream Signaling in Human Chondrosarcomas Identifies Pathways for Rational Targeted Therapy. *Clin. Cancer Res.* **2013**, *19*, 3796–3807. [\[CrossRef\]](https://doi.org/10.1158/1078-0432.CCR-12-3647)
- 57. Bernstein-Molho, R.; Kollender, Y.; Issakov, J.; Bickels, J.; Dadia, S.; Flusser, G.; Meller, I.; Sagi-Eisenberg, R.; Merimsky, O. Clinical activity of mTOR inhibition in combination with cyclophosphamide in the treatment of recurrent unresectable chondrosarcomas. *Cancer Chemother. Pharmacol.* **2012**, *70*, 855–860. [\[CrossRef\]](https://doi.org/10.1007/s00280-012-1968-x)
- 58. Liu, P.; Shen, J.K.; Xu, J.; A Trahan, C.; Hornicek, F.J.; Duan, Z. Aberrant DNA Methylations in Chondrosarcoma. *Epigenomics* **2016**, *8*, 1519–1525. [\[CrossRef\]](https://doi.org/10.2217/epi-2016-0071)
- 59. Ouadid-Ahidouch, H.; Rodat-Despoix, L.; Matifat, F.; Morin, G.; Ahidouch, A. DNA methylation of channel-related genes in cancers. *Biochim. et Biophys. Acta (BBA) Biomembr.* **2015**, *1848*, 2621–2628. [\[CrossRef\]](https://doi.org/10.1016/j.bbamem.2015.02.015) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/25703813)
- 60. Xiao-Jie, L.; Hui-Ying, X.; Qi, X.; Jiang, X.; Shi-Jie, M. LINE-1 in cancer: Multifaceted functions and potential clinical implications. *Anesthesia Analg.* **2016**, *18*, 431–439. [\[CrossRef\]](https://doi.org/10.1038/gim.2015.119)
- 61. Adega, F.; Guedes-Pinto, H.; Chaves, R. Satellite DNA in the Karyotype Evolution of Domestic Animals—Clinical Considerations. *Cytogenet. Genome Res.* **2009**, *126*, 12–20. [\[CrossRef\]](https://doi.org/10.1159/000245903)
- 62. Sorrentino, V.G.; Thota, S.; Gonzalez, E.A.; Rameshwar, P.; Chang, V.T.; Etchegaray, J.-P. Hypomethylating Chemotherapeutic Agents as Therapy for Myelodysplastic Syndromes and Prevention of Acute Myeloid Leukemia. *Pharmaceuticals* **2021**, *14*, 641. [\[CrossRef\]](https://doi.org/10.3390/ph14070641) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/34358067)
- 63. Hamm, C.A.; Xie, H.; Costa, F.F.; Vanin, E.F.; Seftor, E.A.; Sredni, S.T.; Bischof, J.; Wang, D.; Bonaldo, M.F.; Hendrix, M.J.C.; et al. Global Demethylation of Rat Chondrosarcoma Cells after Treatment with 5-Aza-2′ -Deoxycytidine Results in Increased Tumorigenicity. *PLoS ONE* **2009**, *4*, e8340. [\[CrossRef\]](https://doi.org/10.1371/journal.pone.0008340) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/20019818)
- 64. Bui, C.; Ouzzine, M.; Talhaoui, I.; Sharp, S.; Prydz, K.; Coughtrie, M.W.H.; Fournel-Gigleux, S. Epigenetics: Methylationassociated repression of heparan sulfate 3-*O*-sulfotransferase gene expression contributes to the invasive phenotype of H-EMC-SS chondrosarcoma cells. *FASEB J.* **2010**, *24*, 436–450. [\[CrossRef\]](https://doi.org/10.1096/fj.09-136291)
- 65. Sen, S.; Chauhan, S.S.; Pushker, N.; Tandon, R.; Kashyap, S.; Vanathi, M.; Bajaj, M.S. Stratifin in ocular surface squamous neoplasia and its association with p53. *Acta Ophthalmol.* **2021**, *99*, E1483–E1491. [\[CrossRef\]](https://doi.org/10.1111/aos.14844)
- 66. A Hamm, C.; Stevens, J.W.; Xie, H.; Vanin, E.F.; A Morcuende, J.; Abdulkawy, H.; A Seftor, E.; Sredni, S.T.; Bischof, J.M.; Wang, D.; et al. Microenvironment alters epigenetic and gene expression profiles in Swarm rat chondrosarcoma tumors. *BMC Cancer* **2010**, *10*, 471–516. [\[CrossRef\]](https://doi.org/10.1186/1471-2407-10-471)
- 67. Pan, Y.; Liu, G.; Zhou, F.; Su, B.; Li, Y. DNA methylation profiles in cancer diagnosis and therapeutics. *Clin. Exp. Med.* **2018**, *18*, 1–14. [\[CrossRef\]](https://doi.org/10.1007/s10238-017-0467-0)
- 68. Cohen, A.L.; Holmen, S.L.; Colman, H. IDH1 and IDH2 Mutations in Gliomas. *Curr. Neurol. Neurosci. Rep.* **2013**, *13*, 345. [\[CrossRef\]](https://doi.org/10.1007/s11910-013-0345-4)
- 69. Losman, J.-A.; Kaelin, W.G. What a difference a hydroxyl makes: Mutant IDH, (*R*)-2-hydroxyglutarate, and cancer. *Genes Dev.* **2013**, *27*, 836–852. [\[CrossRef\]](https://doi.org/10.1101/gad.217406.113)
- 70. Venneker, S.; Kruisselbrink, A.B.; Baranski, Z.; Palubeckaite, I.; Bruijn, I.H.B.-D.; Oosting, J.; French, P.J.; Danen, E.H.J.; Bovée, J.V.M.G. Beyond the Influence of *IDH* Mutations: Exploring Epigenetic Vulnerabilities in Chondrosarcoma. *Cancers* **2020**, *12*, 3589. [\[CrossRef\]](https://doi.org/10.3390/cancers12123589)
- 71. Peterse, E.F.P.; van den Akker, B.E.W.M.; Niessen, B.; Oosting, J.; Suijker, J.; De Jong, Y.; Danen, E.H.J.; Cleton-Jansen, A.-M.; Bovée, J.V.M.G. NAD Synthesis Pathway Interference Is a Viable Therapeutic Strategy for Chondrosarcoma. *Mol. Cancer Res.* **2017**, *15*, 1714–1721. [\[CrossRef\]](https://doi.org/10.1158/1541-7786.MCR-17-0293)
- 72. Jin, Z.; Han, Y.-X.; Han, X.-R. Loss of RUNX3 expression may contribute to poor prognosis in patients with chondrosarcoma. *Histochem. J.* **2013**, *44*, 645–652. [\[CrossRef\]](https://doi.org/10.1007/s10735-013-9511-x) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/23666463)
- 73. Liu, P.; Garbutt, C.; Hornicek, F.J.; Liu, F.; Duan, Z. Aberration of p73 Promoter Methylation in Chondrosarcoma. *Anticancer. Res.* **2017**, *37*, 2939–2945. [\[CrossRef\]](https://doi.org/10.21873/anticanres.11647) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/28551631)
- 74. Monga, V.; Mani, H.; Hirbe, A.; Milhem, M. Non-Conventional Treatments for Conventional Chondrosarcoma. *Cancers* **2020**, *12*, 1962. [\[CrossRef\]](https://doi.org/10.3390/cancers12071962)
- 75. Paoluzzi, L.; Cacavio, A.; Ghesani, M.; Karambelkar, A.; Rapkiewicz, A.; Weber, J.; Rosen, G. Response to anti-PD1 therapy with nivolumab in metastatic sarcomas. *Clin. Sarcoma Res.* **2016**, *6*, 24. [\[CrossRef\]](https://doi.org/10.1186/s13569-016-0064-0)
- 76. Bilusic, M.; Heery, C.R.; Collins, J.M.; Donahue, R.N.; Palena, C.; Madan, R.A.; Karzai, F.; Marté, J.L.; Strauss, J.; Gatti-Mays, M.E.; et al. Phase I trial of HuMax-IL8 (BMS-986253), an anti-IL-8 monoclonal antibody, in patients with metastatic or unresectable solid tumors. *J. Immunother. Cancer* **2019**, *7*, 240. [\[CrossRef\]](https://doi.org/10.1186/s40425-019-0706-x)
- 77. Nishida, K.; Kunisada, T.; Shen, Z.N.; Kadota, Y.; Hashizume, K.; Ozaki, T. Chondrosarcoma and Peroxisome Proliferator-Activated Receptor. *PPAR Res.* **2008**, *2008*, 250568. [\[CrossRef\]](https://doi.org/10.1155/2008/250568)
- 78. Higuchi, T.; Takeuchi, A.; Munesue, S.; Yamamoto, N.; Hayashi, K.; Kimura, H.; Miwa, S.; Inatani, H.; Shimozaki, S.; Kato, T.; et al. Anti-tumor effects of a nonsteroidal anti-inflammatory drug zaltoprofen on chondrosarcoma via activating peroxisome proliferator-activated receptor gamma and suppressing matrix metalloproteinase-2 expression. *Cancer Med.* **2018**, *7*, 1944–1954. [\[CrossRef\]](https://doi.org/10.1002/cam4.1438)
- 79. Ouyang, Z.; Wang, S.; Zeng, M.; Li, Z.; Zhang, Q.; Wang, W.; Liu, T. Therapeutic effect of palbociclib in chondrosarcoma: Implication of cyclin-dependent kinase 4 as a potential target. *Cell Commun. Signal.* **2019**, *17*, 17. [\[CrossRef\]](https://doi.org/10.1186/s12964-019-0327-5)
- 80. Wu, M.-H.; Lee, C.-Y.; Huang, T.-J.; Huang, K.-Y.; Tang, C.-H.; Liu, S.-H.; Kuo, K.-L.; Kuan, F.-C.; Lin, W.-C.; Shi, C.-S. MLN4924, a Protein Neddylation Inhibitor, Suppresses the Growth of Human Chondrosarcoma through Inhibiting Cell Proliferation and Inducing Endoplasmic Reticulum Stress-Related Apoptosis. *Int. J. Mol. Sci.* **2018**, *20*, 72. [\[CrossRef\]](https://doi.org/10.3390/ijms20010072)
- 81. Fu, X.; Li, M.; Tang, C.; Huang, Z.; Najafi, M. Targeting of cancer cell death mechanisms by resveratrol: A review. *Apoptosis* **2021**, *26*, 561–573. [\[CrossRef\]](https://doi.org/10.1007/s10495-021-01689-7) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/34561763)
- 82. Zou, Z.; Tao, T.; Li, H.; Zhu, X. mTOR signaling pathway and mTOR inhibitors in cancer: Progress and challenges. *Cell Biosci.* **2020**, *10*, 31. [\[CrossRef\]](https://doi.org/10.1186/s13578-020-00396-1)
- 83. Addie, R.D.; de Jong, Y.; Alberti, G.; Kruisselbrink, A.B.; Que, I.; Baelde, H.; Bovée, J.V. Exploration of the chondrosarcoma metabolome; the mTOR pathway as an important pro-survival pathway. *J. Bone Oncol.* **2019**, *15*, 100222. [\[CrossRef\]](https://doi.org/10.1016/j.jbo.2019.100222)
- 84. Miwa, S.; Yamamoto, N.; Hayashi, K.; Takeuchi, A.; Igarashi, K.; Tsuchiya, H. Therapeutic Targets and Emerging Treatments in Advanced Chondrosarcoma. *Int. J. Mol. Sci.* **2022**, *23*, 1096. [\[CrossRef\]](https://doi.org/10.3390/ijms23031096) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/35163019)
- 85. Cohen-Nowak, A.J.; Dressler, D.B.; Rock, A.; Mojica, K.; Woo, D.; Zuckerman, L.M.; Chow, W.; Agulnik, M. Role of immunotherapy in chondrosarcoma: A case report and review of the literature. *Ther. Adv. Med Oncol.* **2023**, *15*, 17588359231199877. [\[CrossRef\]](https://doi.org/10.1177/17588359231199877)
- 86. Jamil, N.; Howie, S.; Salter, D.M. Therapeutic molecular targets in human chondrosarcoma. *Int. J. Exp. Pathol.* **2010**, *91*, 387–393. [\[CrossRef\]](https://doi.org/10.1111/j.1365-2613.2010.00749.x)
- 87. Dahlin, D.C.; Henderson, E.D. Chondrosarcoma, a surgical and pathological problem; review of 212 cases. *J. Bone Joint Surg. Am.* **1956**, *38*, 1025–1038. [\[CrossRef\]](https://doi.org/10.2106/00004623-195638050-00007) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/13367080)
- 88. Nota, S.P.F.T.; Braun, Y.; Schwab, J.H.; van Dijk, C.N.; Bramer, J.A.M. The Identification of Prognostic Factors and Survival Statistics of Conventional Central Chondrosarcoma. *Sarcoma* **2015**, *2015*, 623746. [\[CrossRef\]](https://doi.org/10.1155/2015/623746)
- 89. Giordano, G.; Merlini, A.; Ferrero, G.; Mesiano, G.; Fiorino, E.; Brusco, S.; Centomo, M.L.; Leuci, V.; D'ambrosio, L.; Aglietta, M.; et al. EphA2 Expression in Bone Sarcomas: Bioinformatic Analyses and Preclinical Characterization in Patient-Derived Models of Osteosarcoma, Ewing's Sarcoma and Chondrosarcoma. *Cells* **2021**, *10*, 2893. [\[CrossRef\]](https://doi.org/10.3390/cells10112893)
- 90. Kroonen, J.S.; Kruisselbrink, A.B.; Bruijn, I.H.B.-D.; Olaofe, O.O.; Bovée, J.V.M.G.; Vertegaal, A.C.O. SUMOylation Is Associated with Aggressive Behavior in Chondrosarcoma of Bone. *Cancers* **2021**, *13*, 3823. [\[CrossRef\]](https://doi.org/10.3390/cancers13153823)
- 91. Liang, X.; Wang, D.; Wang, Y.; Zhou, Z.; Zhang, J.; Li, J. Expression of Aurora Kinase A and B in chondrosarcoma and its relationship with the prognosis. *Diagn. Pathol.* **2012**, *7*, 84. [\[CrossRef\]](https://doi.org/10.1186/1746-1596-7-84) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/22809428)
- 92. Jeong, W.; Kim, H.-J. Biomarkers of chondrosarcoma. *J. Clin. Pathol.* **2018**, *71*, 579–583. [\[CrossRef\]](https://doi.org/10.1136/jclinpath-2018-205071) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/29593061)
- 93. Skipar, P.; Dey, M.; Piątkowski, J.; Sulejczak, D.; Rutkowski, P.; Czarnecka, A.M. MicroRNAs as Prognostic Biomarkers and Therapeutic Targets in Chondrosarcoma. *Int. J. Mol. Sci.* **2024**, *25*, 3176. [\[CrossRef\]](https://doi.org/10.3390/ijms25063176) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/38542153)
- 94. Poudel, B.H.; Koks, S. The whole transcriptome analysis using FFPE and fresh tissue samples identifies the molecular fingerprint of osteosarcoma. *Exp. Biol. Med.* **2024**, *249*, 10161. [\[CrossRef\]](https://doi.org/10.3389/ebm.2024.10161)
- 95. Tudor, M.; Popescu, R.C.; Negoita, R.D.; Gilbert, A.; Ilisanu, M.A.; Temelie, M.; Dinischiotu, A.; Chevalier, F.; Mihailescu, M.; Savu, D.I. In vitro hyperspectral biomarkers of human chondrosarcoma cells in nanoparticle-mediated radiosensitization using carbon ions. *Sci. Rep.* **2023**, *13*, 14878. [\[CrossRef\]](https://doi.org/10.1038/s41598-023-41991-9)

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