

1 **Transcriptomic-proteomic correlation in the predation-**
2 **evoked venom of the cone snail, *Conus imperialis***

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10 **List of supporting information provided**

11 Supplementary Table 1: 62 sequences common between two *C. imperialis* specimens.

12 Supplementary Table 2: Sequences validated by MS/MS.

13 Supplementary Table 3: Assembly of *C imperialis* venom duct for specimen 1 (S1) and
14 specimen 2 (S2).

15 Supplementary Table 4: Comparison of assembled and raw data from two assemblers.

16 Supplementary Report: AGRF RNA quality measurement (2100 expert_mRNA
17 Pico_DE54700697).

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21 Supplementary Table 1: 62 sequences identified in both *C. imperialis* specimens.

Name by superfamily	Precursor sequence	S1*	S2*
A_1	MGMRMFTVFLVLLVATTVPITLASATDGRNAAADARMSPLISKFKDYCHKYGYTIG	360	1689
A_2	MGMRMFTVFLVLLVATTVPITLASATDGRNAAANARVSPVSKFSKWKCHFNPTVYG	437	1665
A_3_Bn1.3	MGMRMFTVFLVLLVATAVLPVTLDRASDGRNAAANAKTARLIAPPFIRDYCHRGPCMVWCG	245	516
A_4	MGMRMFTVFLVLLVATAVLPVTLDRASDGRNAAANAKTARLIAPPFIRDYCHRGPCMVWCG	2	526
A_5	MGMRMFTVFLVLLVATTVPITLASATDGRNAAADARMSPLISKFKDYCHKYGYTIG	52	47
A_6	MGMRMFTVFLVLLVATTVPITLASATDGRNAAADARMSPLISKFKDYCHKYGYTIG	10	34
A_7	MGMRMFTVFLVLLVATTVPITLASATDGRNAAANARVSPVSKFSKWKCHFNPTVYG	9	28
A_8	MGMRMFTVFLVLLVATTVPITLASATDGRNAAADARMSPLISKFKKGLLS	3	16
A_9	MGMRMFTVFLVLLVATTVPITLASATDGRNAAANARVSPVSKFSKWKCHFNPTVYG	2	14
A_10	MGMRMFTVFLVLLVATTVPITLASATDGRNAAANARVSPVSKFSRNGVIRILTLWKG	2	14
A_11	MGMRMFTVFLVLLVATAVLPVTLDRASDECCSQRNASPDRAIHQGLLS	2	9
A_12	MGMRMFTVFLVLLVATTVPITLASATDGRNAAADARMSPLISKQKGLLS	2	6
A_13	MGMRMFTVFLVLLVATTVPITLASATDGRNAAADARMSPLISKFKDYCHKYGYTIG	3	3
A_14	MGMRMFTVFLVLLVATTVPITLASATDGRNAAADARMSPLISKIKQRIIVISMVTLGK	2	3
A_15	MGMRMFTVFLVLLVATTVPITLASATDGRNAAANARVSPVSKFSRNGVIRILTLWGNKAAAAAGPSEPRPAALCLTCFCLFVPELANNLHI	2	3
D_1_Lt15.5	MPKLEMLLVLILPLCYIDAVGPPPPWMEDEIEHWQKLHCHEISDLTPWILCSPEPLCGGKCAQVDCSGPVCTCPCL	438	1856
D_2	MPKLEMLLVLILPLCYIDAVGPPPPWMEDEIEHWQKLHCHEISDLTPWILCSPEPLCGGKCAQVDCSGPVCTCPCL	6	32
D_3	MPKLEMLLVLILPLCYIDAVGPPPHGTWKMMLLNIGKSFIVMKFQISHHGCAARNHFAVEKAAVPRKSATALDQFVHARHVSCSAHSPFSFPHDLALRKEPIKEQLALRHI	4	12
D_4	MPKLEMLLVLILPLCYIDAVGPPHGTWKMMLLNIGKSFIVMKFQISHHGCAARNHFAVEKAAVPRKSATALDQFVHARHVSCSAHSPFSFPHDLALRKEPIKEQLALRHI	2	4
E_1	MHMVFIAMFLLALVEAGWPRLYDKNCKNLLRTYCSNKICGATKNTNGLQCTMYCRCANGCFRQVQIDWPNQOJTLNFC	65	103
E_2	MHMVFIAMFLLALVEAGWPRLYDKNCKRTYCAPTAQIRYVERQRRIRMGNFSAFCTVGVQAVFAVNTSIGPISRPISSSARFPGFDHCLRRERKWLW	7	12
E_3	MHMVFIAMFLLALVEAGWPRLYDKNCKRTYCAPTAQIRYVERQRRIRMGNFSAFCTVGVQAVFAVNTSIGPISRPISSSARFPGFDHCLRRERKWLW	7	10
I1_1	MKLALFTLLMLPLMTEKTSDDLELGVESLRAIFDRDRCSDNIGATCSDFDCGSMCCIGGQCVVTFEACS	6	28
I2_1	MFRVTSVLLVILVLLVLTNACHMDCSKMTCCSGICCFYCGRPMCPGTRRALLQRLVGHQR	141	740
I2_2	MFRVTSVLLVILVLLVLTNACHMDCSKMTCCSGICCFYCGRPMCPGTRRALLQRLVGHQR	161	257
I2_3	MFRVTSVLLVILVLLVLTNACHMDCSKMTCCSGICCFYCGRPMCPGTRRALLQRLVGHQR	17	85
I2_4	MFRVTSVLLVILVLLVLTNACHMDCSKMTCCSGICCFYCGRPMCPGTRRALLQRLVGHQR	7	4
K_1_Im23b	MIMRMTLFLVLLVMTAASAGDALTEAKRIPYCGQTGAECYSWCIKQDLSKDWCCDFVKTIARLPPAHCISQ	155	348
K_2_Im23a	MIMRMTLFLVLLVMTAASAGDALTEAKRIPYCGQTGAECYSWCIKQDLSKDWCCDFVKTIARLPPAHCISQ	105	174
K_3	MIMRMTLFLVLLVMTAASAGDALTEAKRIPYCGQTGAECYSWCIKQDLSKDWCCDFVKTIARLPPAHCISQ	60	109
K_4	MIMRMTLFLVLLVMTAASAGDALTEAKRIPYCGQTGAECYSWCIKQDLSKDWCCDFVKTIARLPPAHCISQ	12	12
K_5	MIMRMTLFLVLLVMTAASAGDALTEAKRIPYCGQTGAECYSWCIKQDLSKDWCCDFVKTIARLPPAHCISQ	7	11
K_6	MIMRMTLFLVLLVMTAASAGDALTEAKRIPYCGQTGAECYSWCIKQDLSKDWCCDFVKTIARLPPAHCISQ	2	2
M_1	MMSLTVLLVILVLLVLTNACHMDCSKMTCCSGICCFYCGRPMCPGTRRALLQRLVGHQR	238	343
M_2_Eu3.3	MMSLTVLLVILVLLVLTNACHMDCSKMTCCSGICCFYCGRPMCPGTRRALLQRLVGHQR	19	43
M_3_Bt3.1	MMSLTVLLVILVLLVLTNACHMDCSKMTCCSGICCFYCGRPMCPGTRRALLQRLVGHQR	41	25
M_4	MSKVGVPVLLIFLVLISIAALONGDDPRRQDEKQSPQDILRSTLTKYSYNIQRRCWAGGSPHCLSSQVCIAPTGHFPAIMCGRVPILT	2	9
M_5	MSKVGVPVLLIFLVLISIAALONGDDPRRQDEKQSPQDILRSTLTKYSYNIQRRCWAGGSPHCLSSQVCIAPTGHFPAIMCGRVPILT	5	5
N_1	MSTLGMMLLILVLLVPLATFADDDGPTMRGHSKLLAHTTRDSCPSGTNCPKICCNNGCCSSKSCRETQATKERVCC	16	6
O1_1_Conotoxin3	MKLRMMIVAVLPLTASIFITADNSRNGIENLPRMRHEMKPKASKLNKRGCLPDEYFCGSMIGALLCCSGWGLGICMT	6	18
O2_1_Im6.2	MKLTILLVAALLVLTQARTERRRVSRTSSTYDDEMATFCWYWNFFQYSYPTVYQVCLTLGKACTNDSKCCSKYCNCFMCKINWEG	188	481
O2_2	MKLTILLVAALLVLTQARTERRRVSRTSSTYDDEMATFCWYWNFFQYSYPTVYQVCLTLGKACTNDSKCCSKYCNCFMCKINWEG	3	17
O2_3	MKLTILLVAALLVLTQARTERRRVSRTSSTYDDEMATFCWYWNFFQYSYPTVYQVCLTLGKACTNDSKCCSKYCNCFMCKINWEG	8	9
P_1	MHLSSAALMLLILVLLVLTNACHMDCSKMTCCSGICCFYCGRPMCPGTRRALLQRLVGHQR	38	56
P_2	MHLSTASSVALMFLVLLVAFYGVQPELMTDRVDNQGTLDRNRNLSRVKPTGLFKSRKPSSEDCGKTCETAENPDDCCSSCLSVETTYRCA	39	23
P_3	MHRSLAGSVALMFLVLLVAFYGVQPELMTDRVDNQGTLDRNRNLSRVKPTGLFKSRKPSSEDCGKTCETAENPDDCCSSCLSVETTYRCA	15	20
P_4	MHLSSAALMLLILVLLVLTNACHMDCSKMTCCSGICCFYCGRPMCPGTRRALLQRLVGHQR	2	3
T_1_Im5.4	MRCLPVVVFLLLLLSAAAAPGVGSKTERLPGLTSSGSDSDESLPFLNTICWNSGACCGG	494	1027
T_2	MRCLPVVVFLLLLLSAAAAPGVGSKTERLPGLTSSGSDSDESLPFLNTICWNSGACCGG	42	199
T_3	MRCLPVVVFLLLLLSAAAAPGVGSKTERLPGLTSSGSDSDESLPFLNTICWNSGACCGG	5	31
T_4	MRCLPVVVFLLLLLSAAAAPGVGSKTERLPGLTSSGSDSDESLPFLNTICWNSGACCGG	7	20
T_5	MRCLPVVVFLLLLLSAAAAPGVGSKTERLPGLTSSGSDSDESLPFLNTICWNSGACCGG	16	19
T_6	MRCLPVVVFLLLLLSAAAAPGVGSKTERLPGLTSSGSDSDESLPFLNTICWNSGACCGG	2	7
T_7	MRCLPVVVFLLLLLSAAAAPGVGSKTERLPGLTSSGSDSDESLPFLNTICWNSGACCGG	2	5
T_8	MRCLPVVVFLLLLLSAAAAPGVGSKTERLPGLTSSGSDSDESLPFLNTICWNSGACCGG	4	5
T_9	MRCLPVVVFLLLLLSAAAAPGVGSKTERLPGLTSSGSDSDESLPFLNTICWNSGACCGG	2	3
T_10	MRCLPVVVFLLLLLSAAAAPGVGSKTERLPGLTSSGSDSDESLPFLNTICWNSGACCGG	2	3
SF-1m1_1	MARFLSILLCFAMATGLAAGIRYDPRVLRGCRSTHDLKMEIDTNLDGVSFPHSFCGSGVEYVTAADRNRNHSNYRVYVCGMTEPCTAENPVRDPKGNRWLQCRQYKMYIWRDLVLC	7	25
SF-1m2_1	MRLTTHSVIIMLVAFDNDVGDPEGQTARDVDNRNFMISLRSEKGPVHFLRAIKKRDCTGQACTTGDNCPEVCNEHHFCTGKCCYFLHA	14	31
SF-1m3_1	MSKSGMLLFLVLLVPLAIPELAPAGRSVTHHFRDFGAKRSVPICSVNPSPNQLQSGWQDKCCSKYCNCFMCKINWEG	4	55
SF-1m4_1	MKFTTCLLILVLLVLTNACHMDCSKMTCCSGICCFYCGRPMCPGTRRALLQRLVGHQR	5	27
SF-1m5_1	MKTGMICLLLIAPMDADGSPGDTLVSQKTDATDSGMKRFQKTRKRCVFCPEKPCDGDQCMTAPGTGPFCCG	2	10

* The values in these columns refer to the read numbers of each paralog found in the transcriptomic analysis for each specimen

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24 Supplementary Table 2: Sequences validated by MSMS (99% confidence)

Name by superfamily	Seq number	MS/MS (> 99% conf)	Fixed modification	Enzyme Digestion	PTMs	Prec MW	z
A_3_Bn1.3	>im003	DYCCR	Ethanolyl	Trypsin		883.34	2
D_1_Lt15.5	>im004	ILCSPEPLCGGK	Ethanolyl	Trypsin		1303.66	2
E_1	>im005	FTWGLQCTMYCR	Ethanolyl	Trypsin		1619.69	2
IL_1	>im006	CSNIGATCSDR	Ethanolyl	Trypsin		1328.54	2
I2_1	>im007	MTCCSGILCCFYCGR	Ethanolyl	Trypsin		1765.69	2
I2_2	>im008	NVCCWNVCESHCR	Ethanolyl	Trypsin		1727.7	3
K_1_Im23b	>im009	LPPAHICSG	Ethanolyl	Trypsin		1008.52	2
K_2_Im23a	>im010	HNPPADKCP	Ethanolyl	Trypsin		1015.47	2
K_3	>im011	VVGMNPPADK	Ethanolyl	Trypsin		1090.54	2
M_1	>im012	CCMRPICMCPCCVN	Ethanolyl	Trypsin	Hyp®10, Amidated®C-term	1853.85	2
M_4	>im015	CWAGSPCHLCSSSQVCIAPTGHPAIMCGR	Ethanolyl	Trypsin	Hyp®7, Hyp®20	2181	3
N_1	>im016	ICCNWNCSSSCR	Ethanolyl	Trypsin		1724.69	2
O2_1_im6.2	>im018	ACTTNSDCCSK	Ethanolyl	Trypsin		1263.49	2
O2_3	>im019	QCSSPAQCCSGNCGAHCK	Ethanolyl	Trypsin	Hyp®5	2018.78	3
P_3	>im022	ADCSYVCFGMGICQSGCYCGPHACMPN	Ethanolyl	Trypsin	M(oxi)®26, Amidated®C-term	3154.23	3
T_1_Im5.4	>im023	PLNTICWNSGACG	Ethanolyl	Trypsin	Amidated®C-term	1651.79	2
SF-im1_1	>im025	VVCGMPTFCTAENPVRDPK	Ethanolyl	Trypsin		2443.33	2
SF-im2_1	>im026	EHFCTGK	Ethanolyl	Trypsin	PyroGlu®N-term	983.45	2
SF-im4_1	>im028	CDRSCTGVVGHGHPSCATCCACFTSA	Ethanolyl	Trypsin	Hyp®12, Amidated®C-term	2689.09	3
SF-im5_1	>im029	CVCFK	Ethanolyl	Trypsin		783.37	2
SF-im6_1	>im031	TGECDEKNPGYVNH	Ethanolyl	Trypsin		1607.68	3

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27 Supplementary Table 3: Assembly of *C imperialis* venom duct for specimen 1 (S1) and
 28 specimen 2 (S2).

IMP S1 (Total De-multiplexed 249,349 and total assembled 198,889)				
	Raw Reads	Newbler Contigs	Newbler Isotigs	Trinity
# Sequences	220,516	703	473	1,594
Length Interval	24-770	1-3,060	58-3,060	101-3,075
Average Length	396.70	436.81	709.57	481.38
N50	459	700	770	502
N75	504	1,197	1,301	818
N90	528	1,690	1,723	1,387
%GC	52.41	47.67	47.04	46.30
%N	0	0	0	0
Total # bases	87,478,648	307,076	335,626	767,323
IMP S2 (Total De-multiplexed 379,998 and total assembled 281,952)				
	Raw Reads	Newbler Contigs	Newbler Isotigs	Trinity
# Sequences	332,390	2,256	1,386	6,021
Length Interval	22-737	1-6,389	26-6,389	101-6,994
Average Length	393.38	463.77	816.63	497.22
N50	459	824	953	521
N75	503	1,382	1,619	868
N90	527	2,218	2,513	1,428
%GC	50.62	46.59	46.36	45.34
%N	0	0	0	0
Total # bases	130,754,372	1,046,266	1,131,845	2,993,735

30 Supplementary Table 4: Comparison of assembled raw data from two assemblers. Detected
 31 sequences (Y) and undetected sequences (N) have been highlighted along with their
 32 subsequent detection in the MSMS spectra.

Name by superfamily	Newbler assebly		Trinity assembly		Raw reads		MSMS
	S1	S2	S1	S2	S1	S2	
A_1	N	N	Y	Y	360	1689	
A_2	Y	N	N	N	437	1665	
A_3_Bn1.3	Y	N	N	Y	245	516	Y
A_4	N	N	N	N	2	526	
A_5	Y	N	N	N	52	47	
A_6	N	N	N	N	10	34	
A_7	N	N	N	N	9	28	
A_8	N	N	N	N	3	16	
A_9	N	N	N	N	2	14	
A_10	N	N	N	N	2	14	
A_11	N	N	N	N	2	9	
A_12	N	N	N	N	2	6	
A_13	N	N	N	N	3	3	
A_14	N	N	N	N	2	3	
A_15	N	N	N	N	2	3	
D_1_Lt15.5	Y	Y	Y	Y	438	1856	Y
D_2	N	N	N	N	6	32	
D_3	N	N	N	N	4	12	
D_4	N	N	N	N	2	4	
E_1	Y	Y	Y	N	65	103	Y
E_2	N	N	N	N	7	12	
E_3	N	N	N	N	7	10	
I1_1	Y	Y	Y	Y	6	28	Y
I2_1	Y	Y	Y	Y	141	740	Y
I2_2	N	N	Y	Y	161	257	Y
I2_3	N	N	N	N	17	85	
I2_4	N	N	N	N	7	4	
K_1_Im23b	Y	Y	Y	Y	155	348	Y
K_2_Im23a	Y	Y	N	N	105	174	Y
K_3	Y	Y	N	N	60	109	Y
K_4	N	N	N	N	12	12	
K_5	N	N	N	N	7	11	
K_6	N	N	N	N	2	2	
M_1	Y	Y	Y	Y	238	343	Y
M_2_Eu3.3	N	Y	N	N	19	43	
M_3_Bt3.1	N	Y	N	N	41	25	
M_4	N	N	N	Y	2	9	Y
M_5	N	N	N	N	5	5	
N_1	Y	N	Y	N	16	6	Y
O1_1_Conotoxin3	N	N	N	Y	6	18	
O2_1_im6.2	Y	Y	Y	Y	188	481	Y
O2_2	N	N	N	N	3	17	
O2_3	Y	N	Y	Y	8	9	Y
P_1	Y	Y	Y	Y	38	56	
P_2	N	N	Y	N	39	23	
P_3	Y	Y	Y	Y	15	20	Y
P_4	N	N	N	N	2	3	
T_1_Im5.4	Y	N	Y	Y	494	1027	Y
T_2	N	N	Y	N	42	199	
T_3	N	Y	N	N	5	31	
T_4	N	N	N	N	7	20	
T_5	N	N	N	N	16	19	
T_6	N	N	N	N	2	7	
T_7	N	N	N	N	2	5	
T_8	N	N	N	N	4	5	
T_9	N	N	N	N	2	3	
T_10	N	N	N	N	2	3	
SF-im1_1	Y	N	N	N	7	25	Y
SF-im2_1	N	N	N	Y	14	31	Y
SF-im3_1	N	Y	Y	Y	4	55	
SF-im4_1	N	N	N	N	5	27	Y
SF-im5_1	N	Y	N	Y	2	10	Y

18 16 15 18

S2 only

S_1	N	N	N	N	0	10	
SF-im6_1	N	Y	N	N	0	6	Y
S1 only							
T-S1	N	N	N	N	41	0	Y

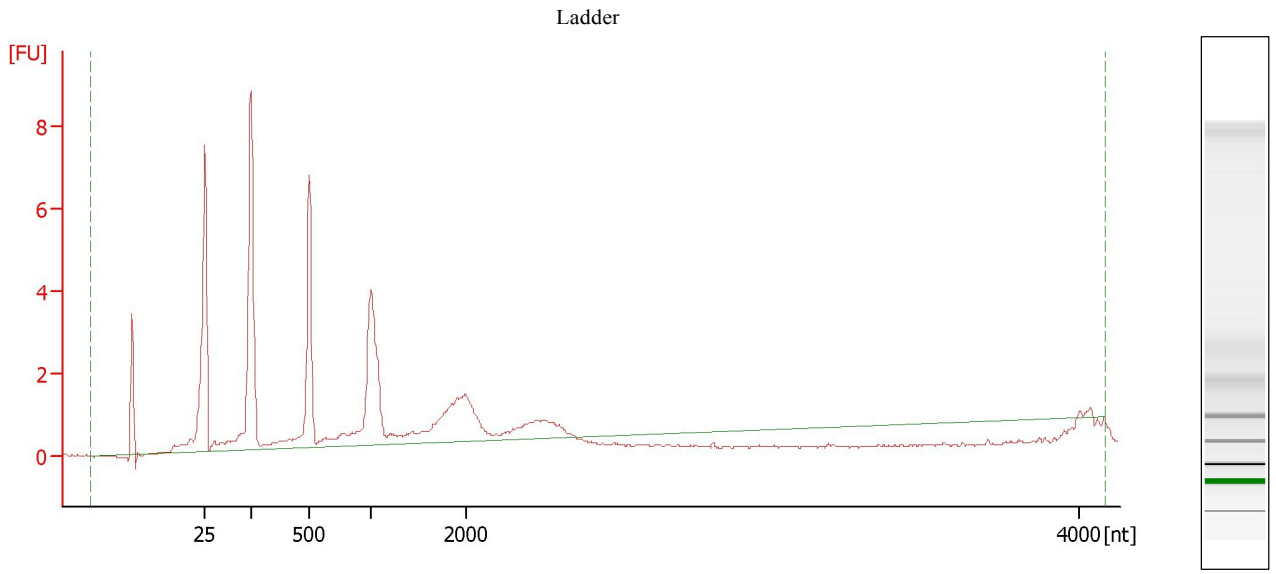
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34 Note: Absent sequences after assembly had been unambiguously confirmed by MSMS, highlighting a
 35 major issue with current assembly algorithms, as this approach can eliminate valid sequences.

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Electropherogram Summary



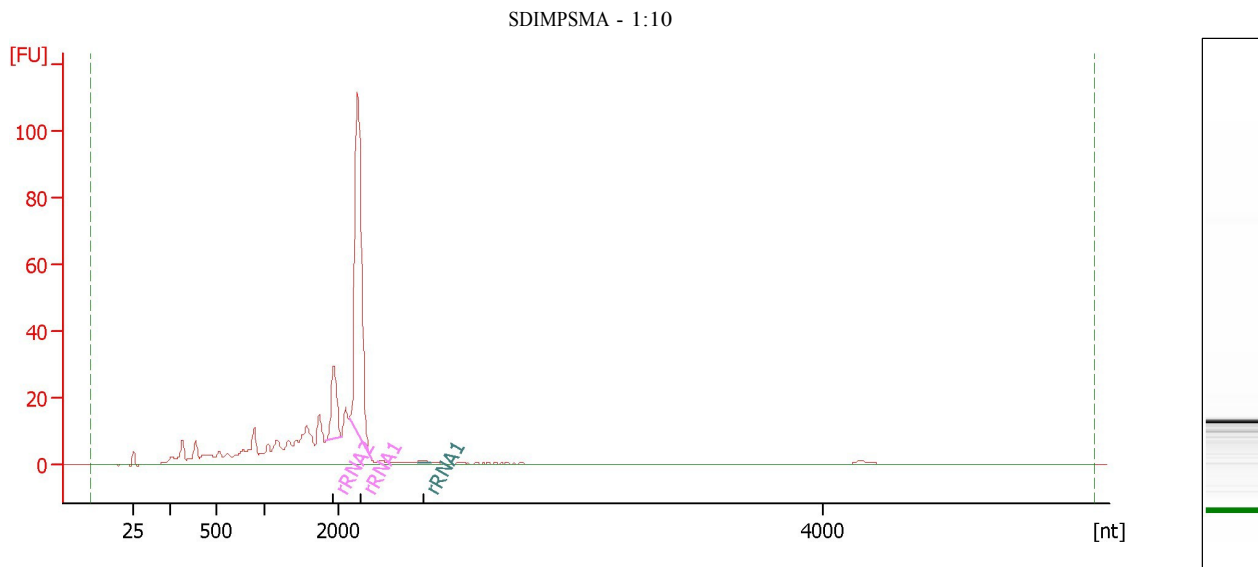
Overall Results for Ladder

RNA Area: 52.0 RNA Concentration: 1,000 pg/μl

Assay Class: mRNA Pico
 Data Path: C:\...16\2100 expert_mRNA Pico_DE54700697_2013-01-16_14-44-32.xad

Created: 1/16/2013 2:44:32 PM
 Modified: 1/16/2013 3:08:43 PM

Electropherogram Summary Continued ...



Overall Results for sample 3 : SDIMPSMA-1:10

RNA Area: 498.8 rRNA Contamination: 38.4 %
 RNA Concentration: 9,600 pg/μl

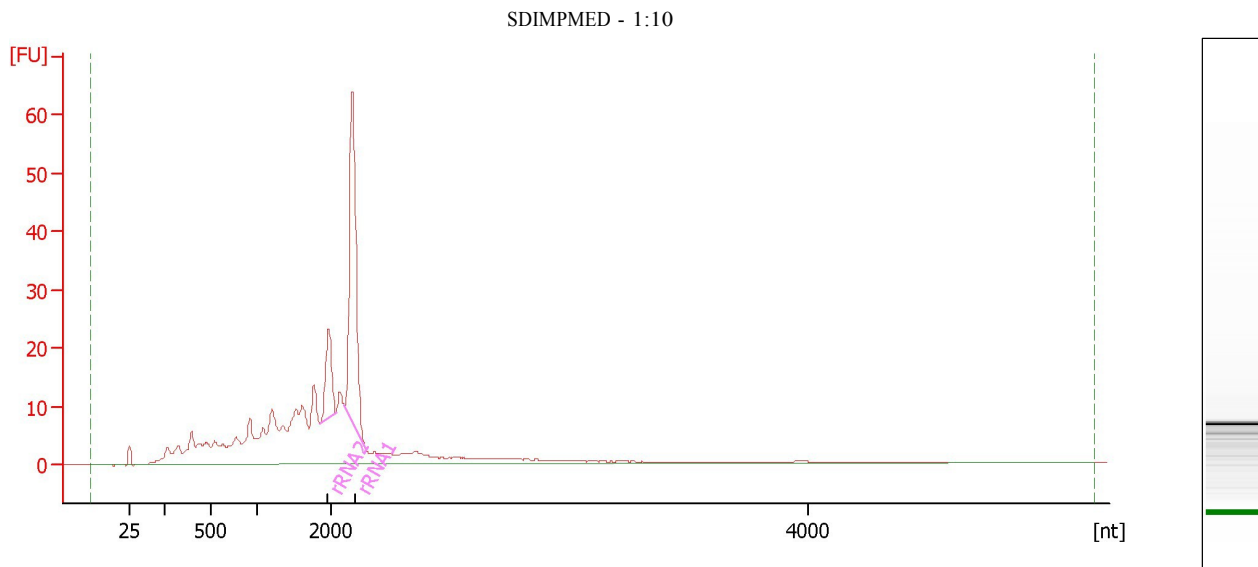
Fragment table for sample 3 : SDIMPSMA-1:10

Name	Start Size [nt]	End Size [nt]	Area	% of total Area
rRNA2	1,830	2,011	29.7	6.0
rRNA1	2,045	2,136	161.0	32.3
rRNA1	2,324	2,379	0.6	0.1

Assay Class: mRNA Pico
 Data Path: C:\...16\2100 expert_mRNA Pico_DE54700697_2013-01-16_14-44-32.xad

Created: 1/16/2013 2:44:32 PM
 Modified: 1/16/2013 3:08:43 PM

Electropherogram Summary Continued ...



Overall Results for sample 4 : SDIMPED-1:10

RNA Area: 435.2 rRNA Contamination: 24.9 %
 RNA Concentration: 8,377 pg/μl

Fragment table for sample 4 : SDIMPED-1:10

Name	Start Size [nt]	End Size [nt]	Area	% of total Area
rRNA2	1,862	2,022	22.3	5.1
rRNA1	2,056	2,146	85.9	19.7