

Table S1 Intracellular fatty acid composition of *C. reinhardtii* (137c) cells subjected to salt stresses (NaCl, KCl, and LiCl) at days 3, 5, and 7. One-hundred milligrams DCW of each stressed cell were collected, and lipids were extracted as described in the Materials and Methods. Data are expressed as mean \pm SD. ND: not detected. Significant differences between the mean values of control and stress treatments were assessed by a two-tailed Student's t-test. * $p < 0.01$, ** $p < 0.001$ and *** $p < 0.0001$ (n = 3).

Fatty acid (%)	Control	200 mM NaCl			200 mM KCl			120 mM LiCl		
		3 days	5 days	7 days	3 days	5 days	7 days	3 days	5 days	7 days
SATURATED FATTY ACIDS (SFAs)										
Caprylic acid (C8:0)	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND
Capric acid (C10:0)	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND
Lauric acid (C12:0)	ND	0.02 \pm 0.00	0.01 \pm 0.02	0.03 \pm 0.03	ND	0.01 \pm 0.00	ND	0.05 \pm 0.00	0.05 \pm 0.00	0.03 \pm 0.00
Myristic acid (C14:0)	1.93 \pm 0.03	0.57 \pm 0.08	1.78 \pm 1.49	2.24 \pm 1.97	2.80 \pm 0.10	2.98 \pm 0.06	3.28 \pm 0.13	0.66 \pm 0.01	0.67 \pm 0.01	0.07 \pm 0.01
Palmitic acid (C16:0)	31.94 \pm 0.16	43.14 \pm 10.56	51.20 \pm 14.96	59.80 \pm 14.46	36.20 \pm 0.00	36.28 \pm 0.01	39.32 \pm 0.03	35.98 \pm 0.04	36.57 \pm 0.03	37.51 \pm 0.03
Stearic acid (C18:0)	4.70 \pm 0.04	6.47 \pm 2.68	6.78 \pm 3.85	7.75 \pm 4.69	3.70 \pm 0.03	3.80 \pm 0.03	4.16 \pm 0.02	4.30 \pm 0.04	4.39 \pm 0.00	4.64 \pm 0.00
Arachidonic acid (C20:0)	0.22 \pm 0.00	0.28 \pm 0.06	0.27 \pm 0.13	0.37 \pm 0.13	0.22 \pm 0.00	0.26 \pm 0.01	0.29 \pm 0.00	0.21 \pm 0.01	0.22 \pm 0.00	0.21 \pm 0.00
Behenic acid (C22:0)	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND
Lignoceric acid (C24:0)	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND
Total SFA	38.79\pm0.33	50.48\pm6.67***	60.04\pm5.66***	70.19\pm0.00***	42.92\pm0.01***	43.33\pm2.85***	47.05\pm0.00***	41.20\pm0.38**	41.90\pm0.22***	43.09\pm0.00***
MONO-UNSATURATED FATTY ACIDS (MUFAs)										
Palmitoleic acid (C16:1)	0.22 \pm 0.25	0.47 \pm 0.38	0.55 \pm 0.41	0.56 \pm 0.39	0.40 \pm 0.37	0.48 \pm 0.36	0.53 \pm 0.54	0.60 \pm 0.31	0.56 \pm 0.31	0.50 \pm 0.06
Oleic acid (C18:1)	5.69 \pm 1.70	7.20 \pm 3.48	6.20 \pm 2.84	5.22 \pm 2.80	5.05 \pm 0.99	5.91 \pm 0.18	6.55 \pm 3.52	3.09 \pm 2.65	3.38 \pm 2.97	1.58 \pm 0.68
Erucic acid (C22:1)	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND
Total MUFA	5.91\pm0.66	7.67\pm0.00	6.75\pm0.00	5.78\pm0.00	5.45\pm1.29	6.39\pm0.16	7.08\pm0.00	3.69\pm0.00*	3.94\pm0.00	2.08\pm0.00***
POLY-UNSATURATED FATTY ACIDS (PUFAs)										
Linoleic acid (C18:2)	33.64 \pm 0.04	26.65 \pm 7.37	22.14 \pm 8.52	15.93 \pm 12.23	29.51 \pm 2.35	30.16 \pm 5.54	28.48 \pm 18.94	34.19 \pm 2.73	34.42 \pm 3.59	36.97 \pm 7.98
Linolenic acid (C18:3)	22.94 \pm 2.36	13.60 \pm 6.60	11.95 \pm 7.37	7.13 \pm 6.42	22.20 \pm 1.61	18.22 \pm 2.19	17.29 \pm 11.81	20.96 \pm 6.10	19.24 \pm 5.54	17.75 \pm 8.74
Total PUFA	56.58\pm1.03	40.25\pm0.03***	34.09\pm2.11***	23.06\pm0.00***	51.71\pm0.74***	44.47\pm1.55***	45.77\pm0.00***	55.15\pm2.73	53.66\pm3.59***	54.72\pm0.00

Table S2 Primer used in this study.

Primers	Sequences (5' - 3')	Base pairs
ChlmyPDH2_Forward	CCCATTGTGGAGGGC	15
ChlmyPDH2_Reverse	ACAGCAGAACGTGCT	15
ChlamyACCCase_Forward	TCTGCAAGTCGCTCG	15
ChlamyACCCase_Reverse	CGTGTCCACGAAGGT	15
ChlamyMAT_Forward	CCGTGAGTCAGCCCG	15
ChlamyMAT_Reverse	TGGACCGGCTTGTCC	15
ChlamyKAS2_Forward	CGCGGACTACACGAC	15
ChlamyKAS2_Reverse	GGCGAAGGGAATGCA	15
ChlamyGPD1_Forward	CAAAGGCCTTGAGGT	15
ChlamyGPD1_Reverse	CAAGTCCATGCCCTC	15
ChlamyFAT 1_Forward	TCAGGAGTCGCGGTG	15
ChlamyFAT 1_Reverse	GCACTTGCAGGCGTG	15
ChlamyDGTT1_Forward	GATTAAGACCGCCGA	15
ChlamyDGTT1_Reverse	AGCGCCTCAGACGCG	15
ChlamyDGTT2_Forward	TCCTGTCCTTCTGGT	15
ChlamyDGTT2_Reverse	TGAACACCACATTCG	15
ChlamyDGTT3_Forward	TACCTCGCACTTGAC	15
ChlamyDGTT3_Reverse	AAGTAGTGACGCCAG	15
ChlamyDGTT4_Forward	GGCTACTTGTTCCGA	15
ChlamyDGTT4_Reverse	GCTGGAGAGGTAGGC	15
Chlamy18s rRNA_Forward	CGGAACCCGCTGGTC	15
Chlamy18s rRNA_Reverse	TGAAGGTGAGCGGCG	15
ChMAT_NdeI_Forward	ATCATATGGTCGCTGTCC	18
ChMAT_BamHI_Reverse	GGATCCAGCGGTGAT	16
SynMAT_NdeI_Forward	CATATGGCTAAAACGGTGTGGGTGT	25
SynMAT_BamHI_Reverse	GGATCCGACCGTGAGGTCTGC	21
pSyn_psbA	TCTTGCCCTTTACAACCT	18
pSyn_rrnB	CGCTACTGCCGCCAG	15
7942rnpB_Forward	GAGGAAAGTCCGGGCTCCC	19
7942rnpB_Reverse	TAAGCCGGGTTCTGTTCTC	19
7942MAT_Forward	CAGCCCTCTACCCG	15
7942MAT_Reverse	GCCATAAACGGCGAG	15
7942LACS_Forward	TGCTTTCTCACGGCA	15
7942LACS_Reverse	TCGATCAAGCGACGT	15

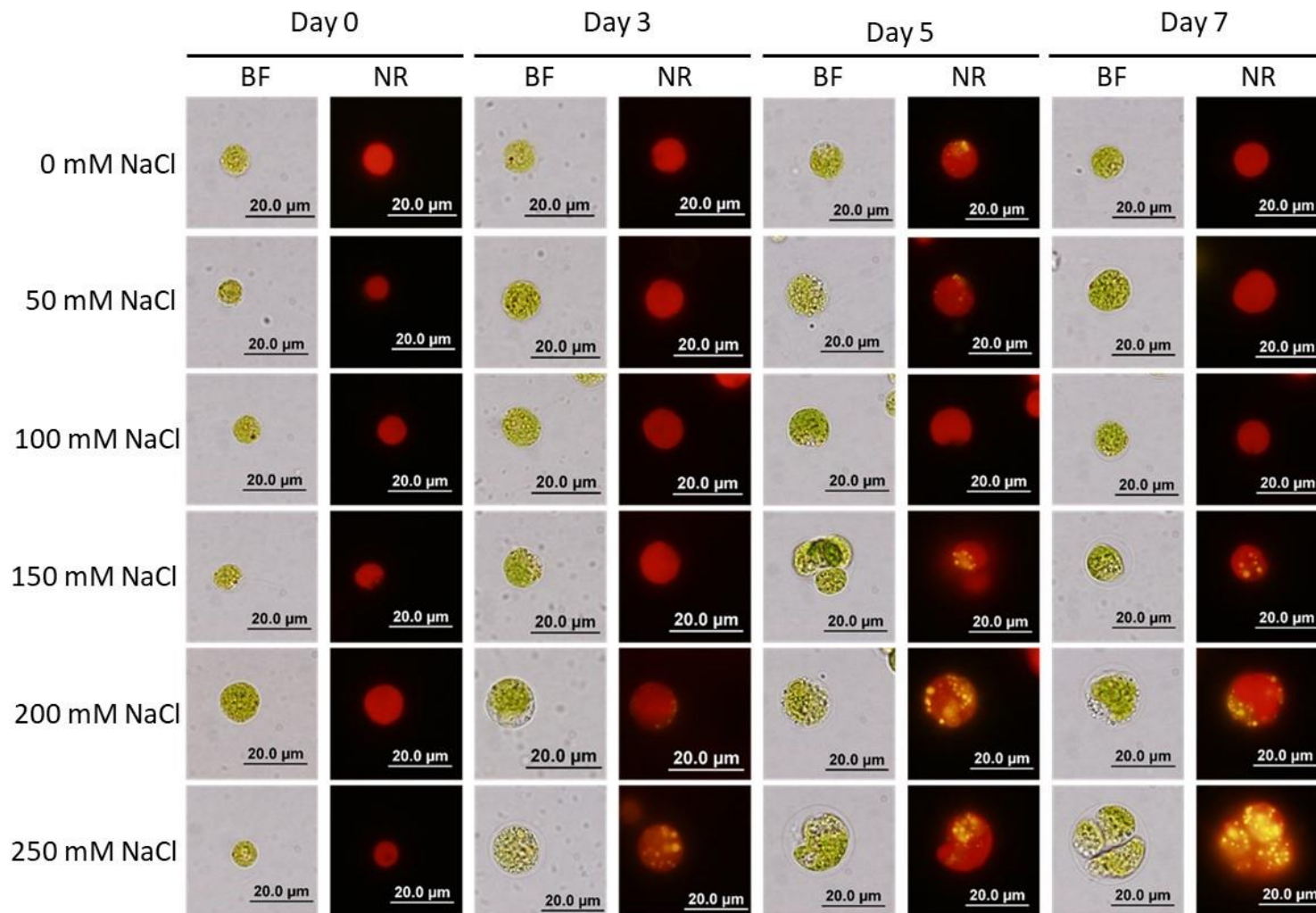


Figure S1 *C. reinhardtii* (137c) cells observed under a bright field light microscope (BF) and fluorescent microscope (NR). Cells at the exponential growth phase were subjected to salt stress (NaCl) at various concentrations, including 0 mM, 50 mM, 100 mM, 150 mM, 200 mM, and 250 mM for 0, 3, 5, and 7 days. Morphology and lipid bodies were observed by Nile red staining.

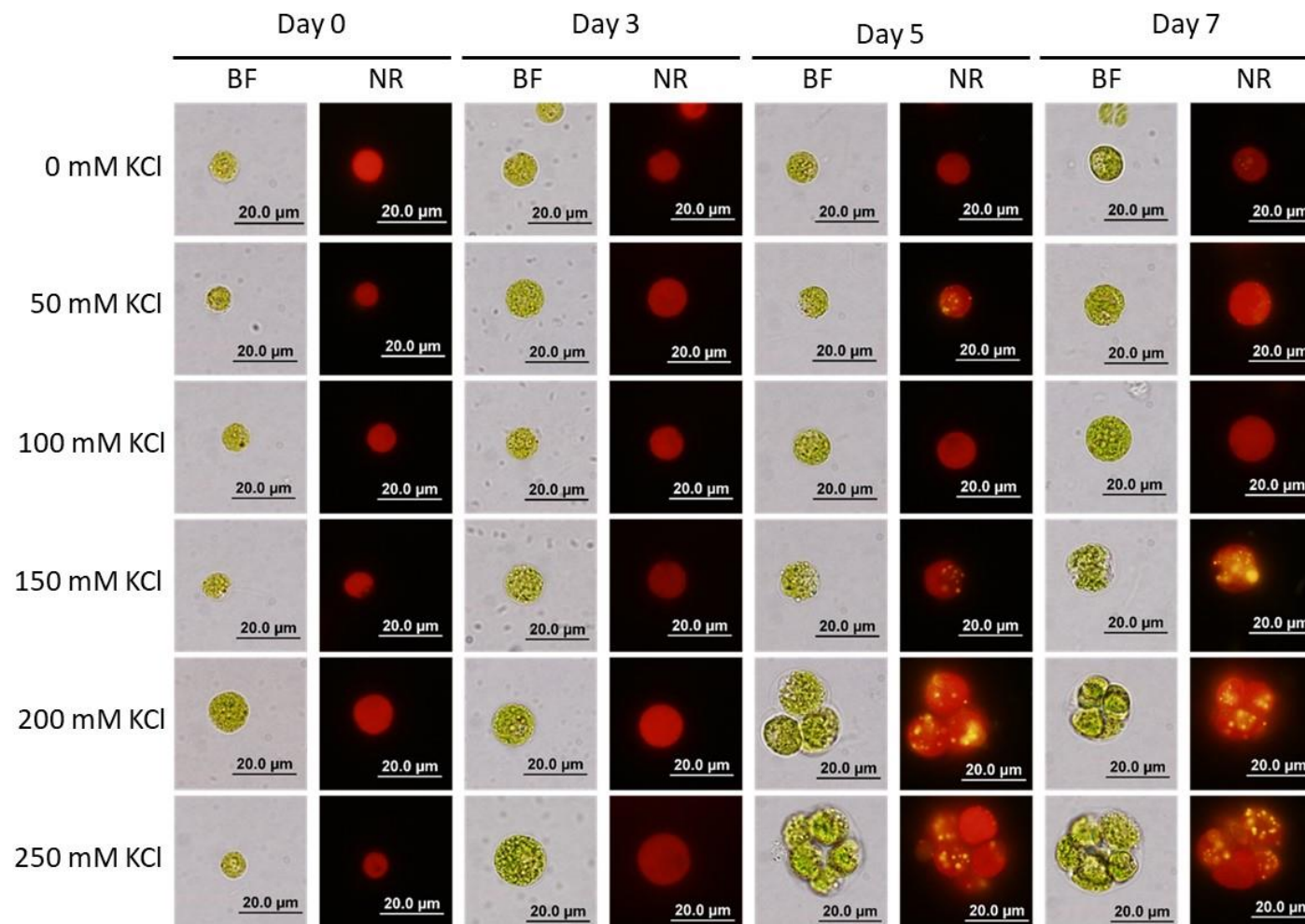


Figure S2 *C. reinhardtii* (137c) cells observed under bright field light microscope (BF) and fluorescent microscope (NR). Cells at exponential growth phase were subjected to salt stress (KCl) at various concentrations; 0 mM, 50 mM, 100 mM, 150 mM, 200 mM, and 250 mM for 0, 3, 5, and 7 days. Morphology and lipid body were observed by Nile red staining.

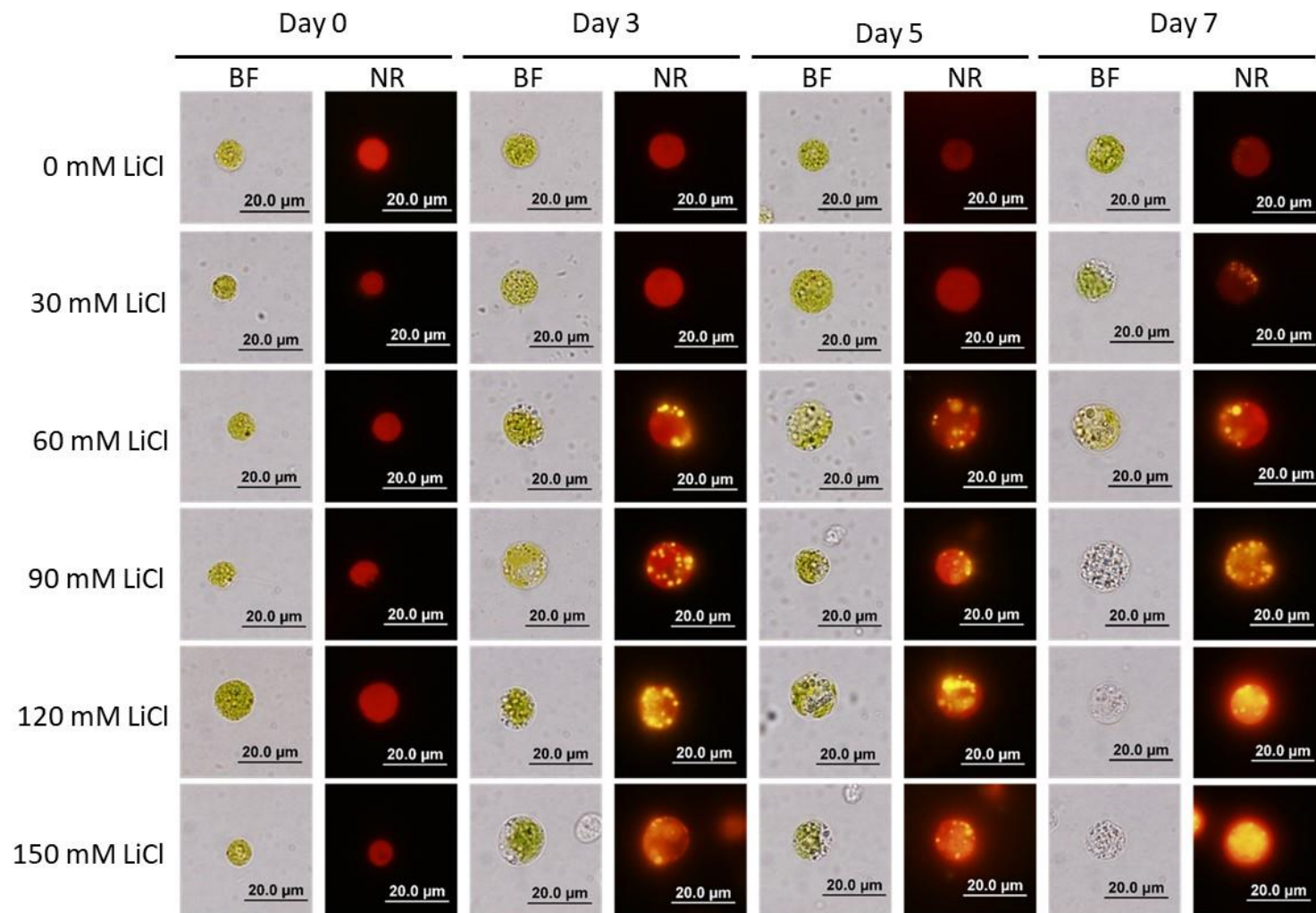


Figure S3 *C. reinhardtii* (137c) cells observed under a bright field light microscope (BF) and fluorescent microscope (NR). Cells at the exponential growth phase were subjected to salt stress (LiCl) at various concentrations, including 0 mM, 30 mM, 60 mM, 90 mM, 120 mM, and 150 mM for 0, 3, 5, and 7 days. Morphology and lipid bodies were observed by Nile red staining.

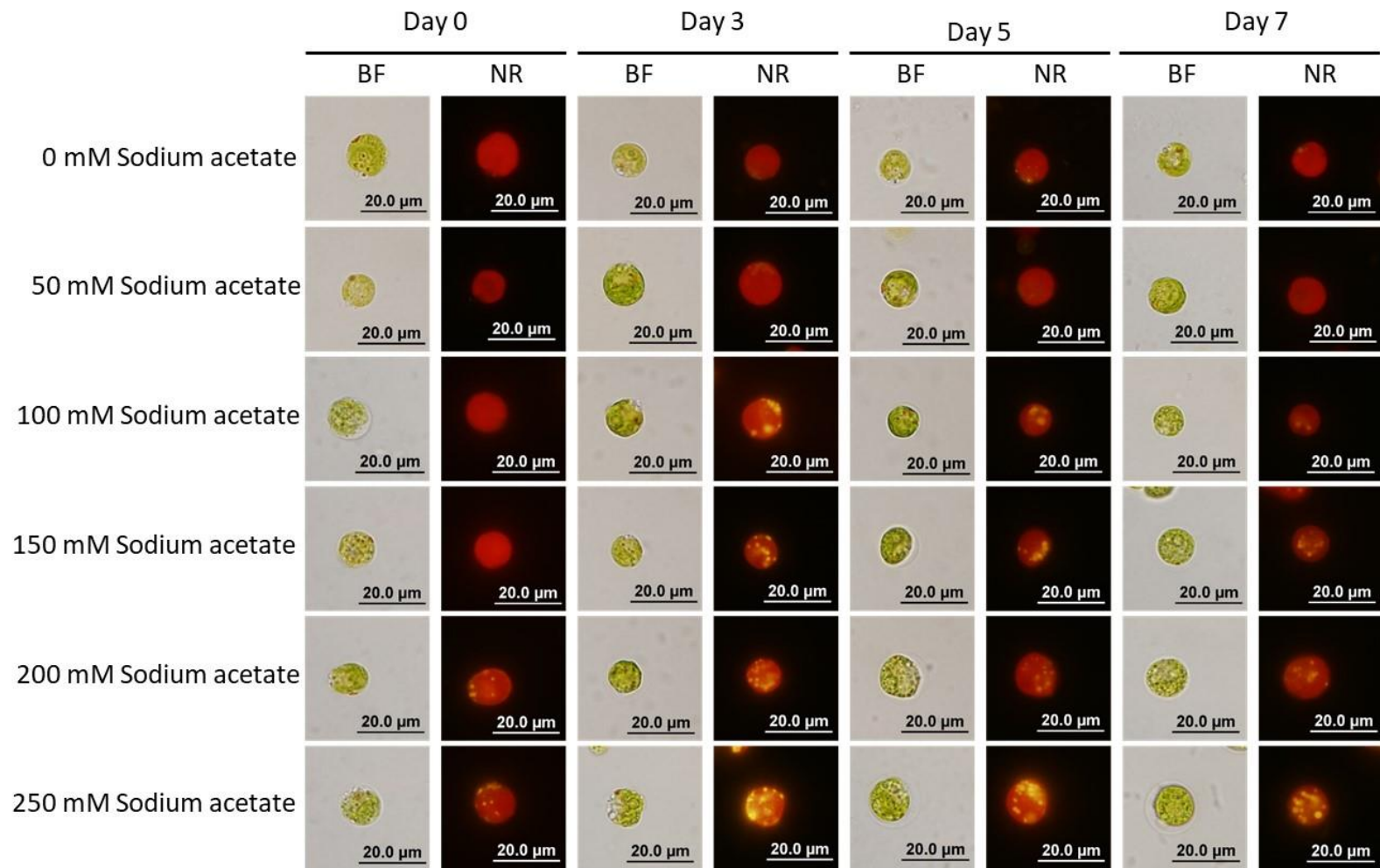


Figure S4 *C. reinhardtii* (137c) cells observed under a bright field light microscope (BF) and fluorescent microscope (NR). Cells at the exponential growth phase were subjected to sodium acetate at various concentrations, including 0 mM, 50 mM, 100 mM, 150 mM, 200 mM, and 250 mM for 0, 3, 5, and 7 days. Morphology and lipid bodies were observed by Nile red staining.

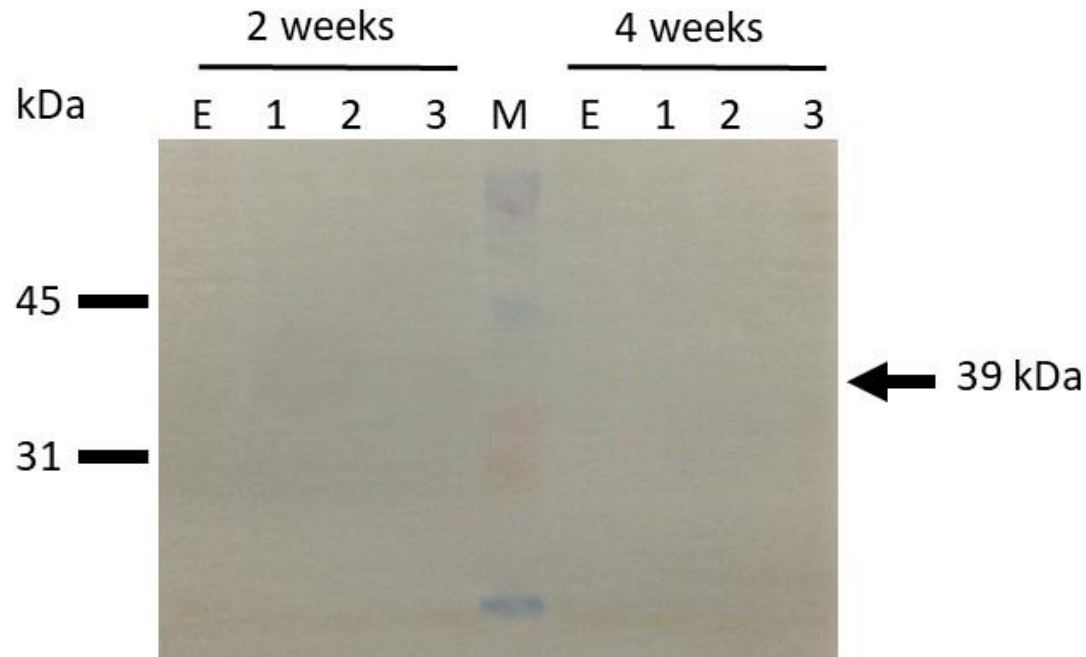


Figure S5 Western blot analysis of empty vector and ChMAT expressing cells. Empty vector (E) and ChMAT (2 and 4 weeks old). An equal amount of crude extract (20 μ g) was loaded in all lanes. Blotting was conducted with a PVDF membrane. Antibody raised against 6x His-tag and HRP-conjugated anti-mouse IgG were used as primary and secondary antibodies, respectively. The membranes were developed using a using Horseradish Peroxidase Conjugate Substrate kit.