

SUPPLEMENTARY MATERIAL

Antimicrobial Lavandulylated Flavonoids from a Sponge-Derived *Streptomyces* sp. G248 in East Vietnam Sea

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TD-DFT electronic CD (ECD) calculation of compounds 1 - 3: A conformational search was carried out with Spartan'14 software (Wavefunction Inc., Irvine, CA) for the proposed absolute configuration as shown at the molecular mechanics level (MMFF) [1]. All MMFF minima were re-optimized using DFT calculations at the B3LYP/6-31G(d) level with the Gaussian 09 program [2]. The geometry was optimized starting from various initial conformations with vibrational frequency calculations confirming the presence of minima. The DFT calculations (B3LYP/LanL2DZ) were performed on the lowest-energy conformations using 30 excited states and a polarizable continuum model (PCM) in methanol. The ECD spectra of the conformers were combined using Boltzmann weighting with program SpecDis 1.71 (applying a sigma value of 0.25 eV) [3]. After applying a UV-shift correction, the computed CD spectra were compared with the CD curves experimentally obtained.

[1] Spartan'14. Wavefunction: Irvine, CA, USA, **2013**.

[2] Gaussian 09, Revision D.01, Frisch, M.J.; Trucks, G.W.; Schlegel, H.B.; Scuseria, G.E.; Robb, M.A.; Cheeseman, J.R.; Scalmani, G.; Barone, V.; Mennucci, B.; Petersson, G.A.; Nakatsuji, H.; Caricato, M.; Li, X.; Hratchian, H.P.; Izmaylov, A.F.; Bloino, J.; Zheng, G.; Sonnenberg, J.L.; Hada, M.; Ehara, M.; Toyota, K.; Fukuda, R.; Hasegawa, J.; Ishida, M.; Nakajima, T.; Honda, Y.; Kitao, O.; Nakai, H.; Vreven, T.; Montgomery, J.A., Jr.; Peralta, J.E.; Ogliaro, F.; Bearpark, M.; Heyd, J.J.; Brothers, E.; Kudin, K.N.; Staroverov, V.N.; Kobayashi, R.; Normand, J.; Raghavachari, K.; Rendell, A.; Burant, J.C.; Iyengar, S.S.; Tomasi, J.; Cossi, M.; Rega, N.; Millam, N.J.; Klene, M.; Knox, J.E.; Cross, J.B.; Bakken, V.; Adamo, C.; Jaramillo, J.; Gomperts, R.; Stratmann, R.E.; Yazyev, O.; Austin, A. J.; Cammi, R.; Pomelli, C.; Ochterski, J. W.; Martin, R.L.; Morokuma, K.; Zakrzewski, V.G.; Voth, G.A.; Salvador, P.; Dannenberg, J.J.; Dapprich, S.; Daniels, A.D.; Farkas, Ö.; Foresman, J.B.; Ortiz, J.V.; Cioslowski, J.; Fox, D.J. Gaussian, Inc., Wallingford CT, 2009.

[3] Bruhn, T.; Schaumlöffel, A.; Hemberger, Y.; Bringmann, G. SpecDis: Quantifying the comparison of calculated and experimental electronic circular dichroism spectra. *Chirality* **2013**, *25*, 243–249

***Streptomyces* sp. G248 isolation and characterization**

Bacteria isolation: The sponge *Halichondria panicea* (Pallas, 1766) sample (0.5 g) was crushed by glass chopsticks in a falcon tube, to which sterile sea water (4.5 mL) was added. The mixture was homogenized by vortexing for 1 min, and the suspension was treated using a wet-heat technique (60 °C for 6 min). An amount (0.5 mL) of this suspension was diluted by sterile distilled water (4.5 mL) and vortexed for 1 min, of which an aliquot of 50 µL was spread on A1 medium (soluble starch: 10 g/L, yeast extract: 4 g/L, peptone: 2 g/L, instant ocean: 30 g/L, agar: 15 g/L) supplemented with 50 µg/mL of polymycin B and cycloheximide to inhibit Gram (-) bacterial and fungal contaminations. After 21 days of aerobic incubation at 30 °C, a colony of the actinomycete strain was transferred onto a petri dish of medium A1 for purification.

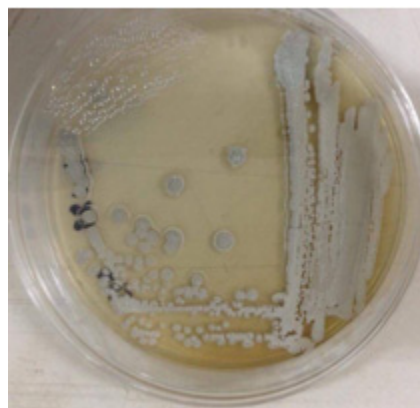


Figure 1S. Morphological appearance of strain G248

Bacteria identification: Genomic DNA was extracted with the Gen Elute Bacterial Genomic DNA kit (Sigma). The sequence of 16S rRNA was used for identification. The gene amplification was performed in a 25.0 µL mixture containing sdH₂O (16.3 µL), 10X PCR buffer (2.5 µL), 25 mM MgCl₂ (1.5 µL), 10 mM dNTP's (0.5 µL), Taq polymerase (0.2 µL), 0.05 mM (1.0 µL) for both primers of 9F (5'-GAGTTTGATCCTGGCTCAG3') and 1541R (5'-AAGGAGGTGATCCAACC3') and genomic DNA (2.0 µL). The reaction tube was then put into MJ Thermalcycler, which had been programmed to preheat at 94 °C for 3 min, followed by 30 cycles of denaturation at 94 °C for 1 min, annealing at 60 °C for 30s and elongation at 72 °C for 45s before a final extension of 72 °C for 10 min. The estimated PCR

product size was about 1500 bp (Figure 2S). PCR products were purified by DNA purification kit (Invitrogen) then sequenced by DNA Analyzer (ABI PRISM 3100, Applied Bioscience). Gene sequences were handled by BioEdit v.2.7.5. and compared with bacterial 16S rRNA sequences in GeneBank database by NCBI Blast program. The results showed that the strain G248 belonged to the genus *Streptomyces*. The sequence of 16S rRNA gene of the strain G248 was registered on GenBank database with the accession number MG917690 (Figure 3S).

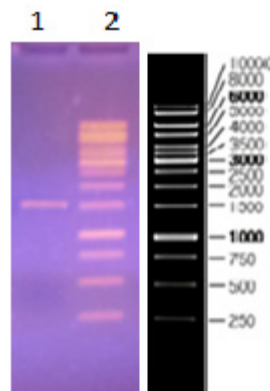


Figure 2S. Gel electrophoresis of PCR product

Lane 1: PCR product; Lane 2: One Kb DNA ladder of Fisher Scientific

Streptomyces sp. strain G248 16S ribosomal RNA gene, partial sequence

GenBank: MG917690.1

[FASTA](#) [Graphics](#) [PopSet](#)

[Go to:](#)

LOCUS MG917690 1368 bp DNA linear BCT 30-JUL-2018
DEFINITION Streptomyces sp. strain G248 16S ribosomal RNA gene, partial
sequence.
ACCESSION MG917690
VERSION MG917690.1
KEYWORDS .
SOURCE Streptomyces sp.
ORGANISM [Streptomyces sp.](#)
Bacteria; Actinobacteria; Streptomycetales; Streptomycetaceae;
Streptomyces.
REFERENCE 1 (bases 1 to 1368)
AUTHORS Le,T.H.M., Vu,T.Q., Nguyen,M.A., Pham,V.C., Doan,T.M.H., Tran,V.H.
and Cao,D.T.
TITLE Partial 16S nRNA of strains isolated from the coastal in Viet Nam
JOURNAL Unpublished
REFERENCE 2 (bases 1 to 1368)
AUTHORS Le,T.H.M., Vu,T.Q., Nguyen,M.A., Pham,V.C., Doan,T.M.H., Tran,V.H.
and Cao,D.T.
TITLE Direct Submission
JOURNAL Submitted (05-FEB-2018) Biotechnology, Institute of Marine
Biochemistry, VAST, 18 Hoang Quoc Viet, Cau Giay, Ha Noi, Ha Noi
10000, Viet Nam
COMMENT ##Assembly-Data-START##
Sequencing Technology :: Sanger dideoxy sequencing
##Assembly-Data-END##
FEATURES Location/Qualifiers
source 1..1368

Figure 3S. 16S rRNA gene sequence of *Streptomyces* sp. G248 on GenBank

Table 1S. Conformational population (%) for the most stable conformers of (2*S*,2''*R*)-1

Conformer	Population (%)
C-1	21.41
C-2	15.78
C-3	19.94
C-4	4.38
C-5	16.57
C-6	6.18
C-7	15.74

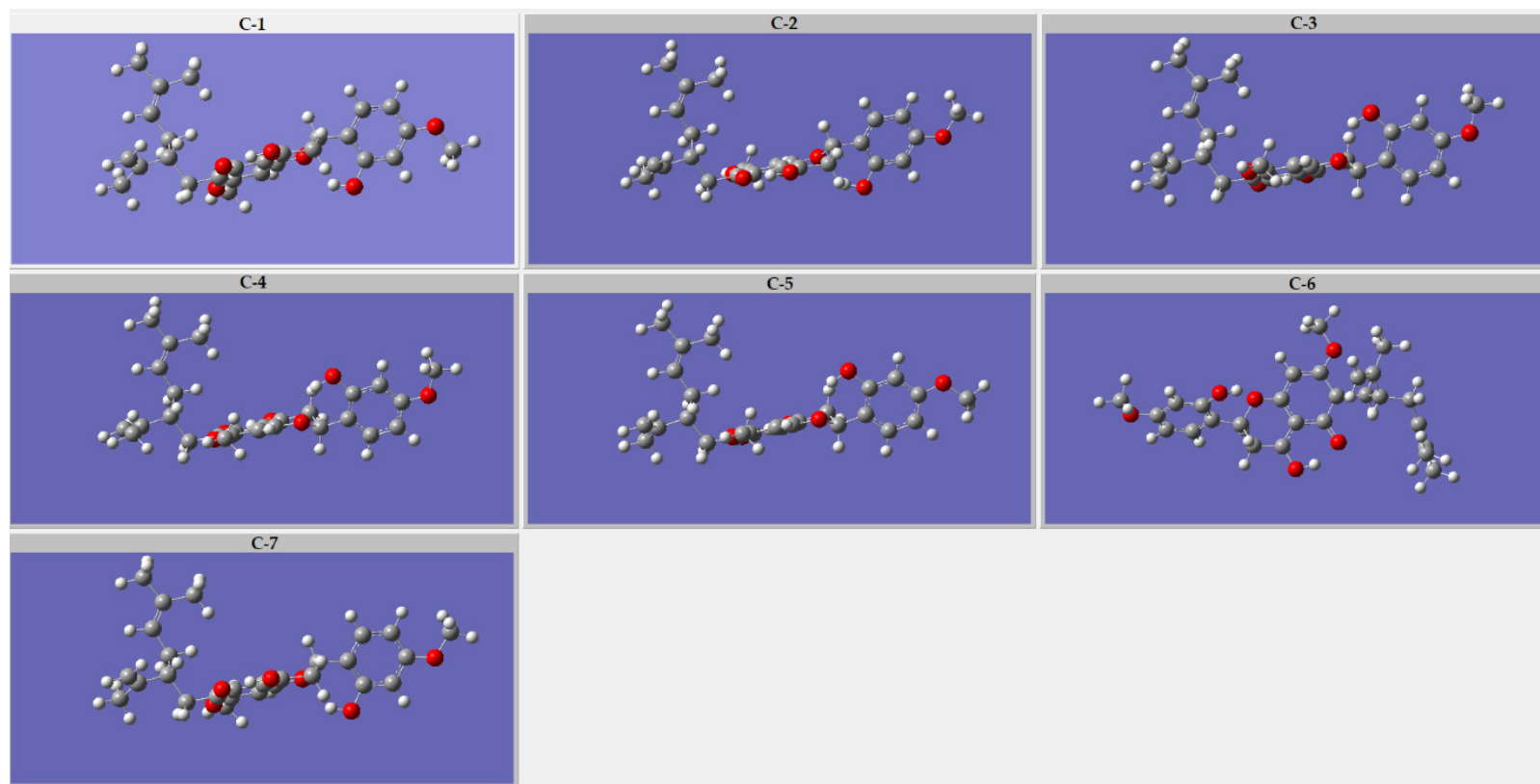


Figure 4S. Optimized conformers of (2*S*,2''*R*)-1

Table 2S. Conformational population (%) for the most stable conformers of (2*S*,2''*S*)-1

Conformer	Population (%)
C-1	13.76
C-2	8.83
C-3	19.86
C-4	12.64
C-5	3.05
C-6	10.71
C-7	6.92
C-8	15.22
C-9	9.02

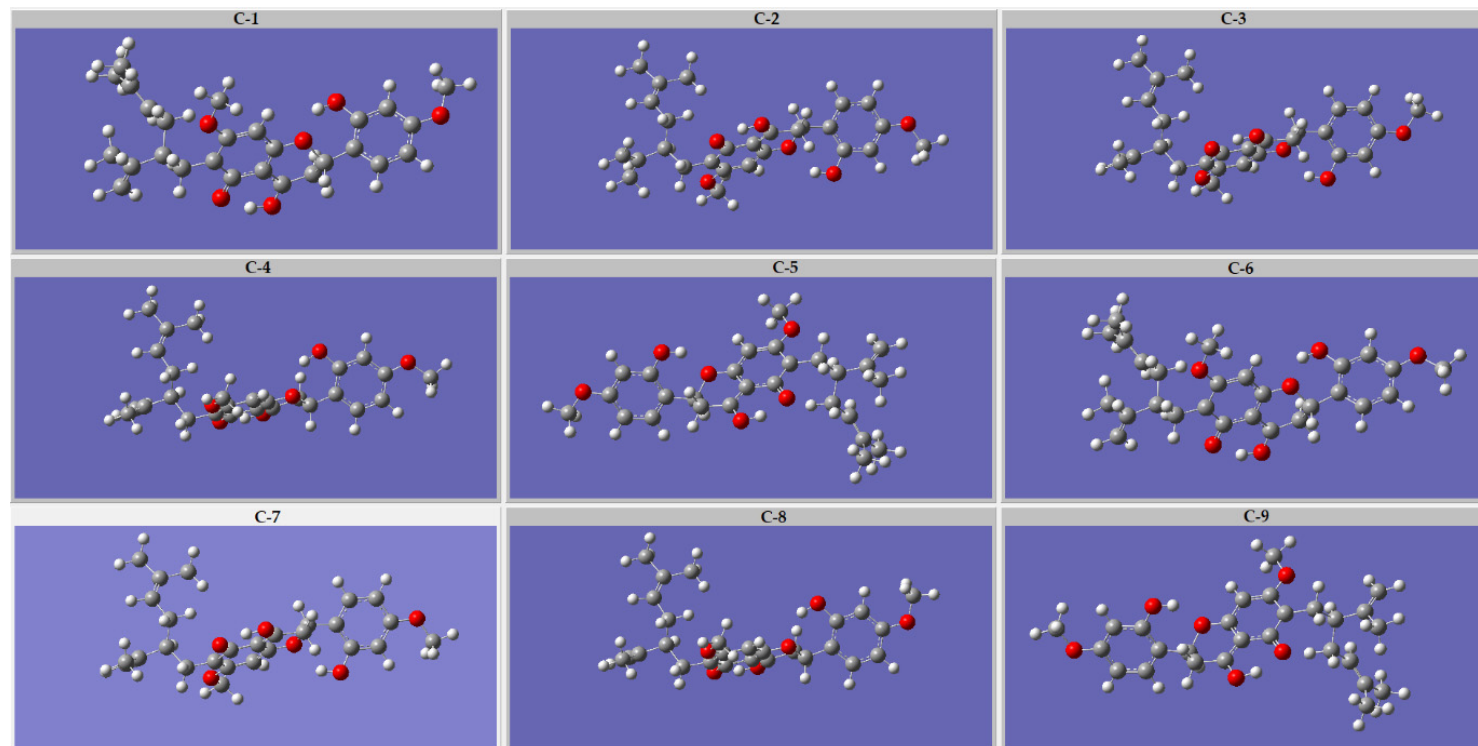


Figure 5S. Optimized conformers of (2*S*,2''*S*)-1

Table 3S. Conformational population (%) for the most stable conformers of (2*S*,2''*R*)-2

Conformer	Population (%)
C-1	8.54
C-2	36.15
C-3	8.85
C-4	8.72
C-5	37.73

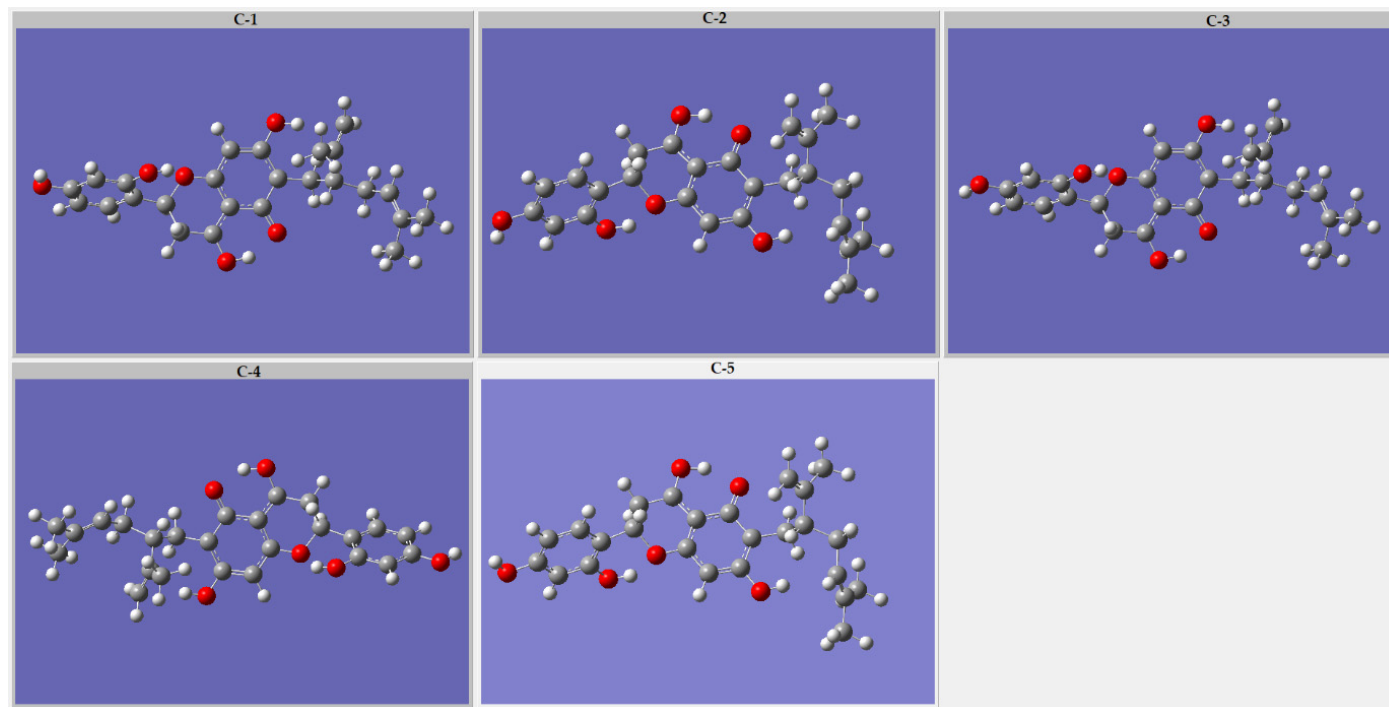


Figure 6S. Optimized conformers of (2*S*,2''*R*)-2

Table 4S. Conformational population (%) for the most stable conformers of (2*S*,2''*S*)-2

Conformer	Population (%)
C-1	30.59
C-2	2.06
C-3	1.42
C-4	31.24
C-5	31.05
C-6	2.14
C-7	1.50

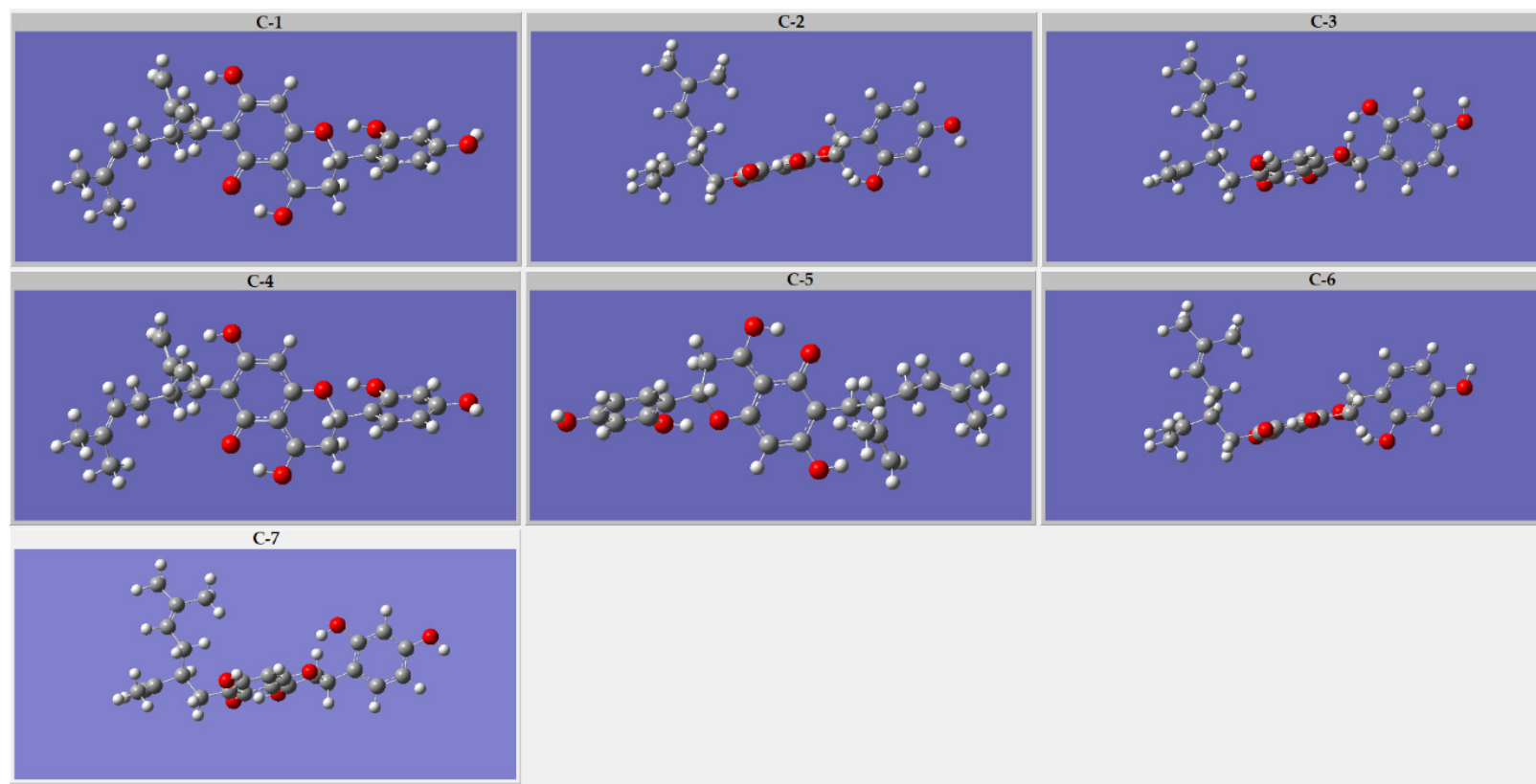


Figure 7S. Optimized conformers of (2*S*,2''*S*)-2

Table 5S. Conformational population (%) for the most stable conformers of (2''*S*)-3

Conformer	Population (%)
C-1	53.55
C-2	39.06
C-3	1.36
C-4	1.71
C-5	3.06
C-6	1.26

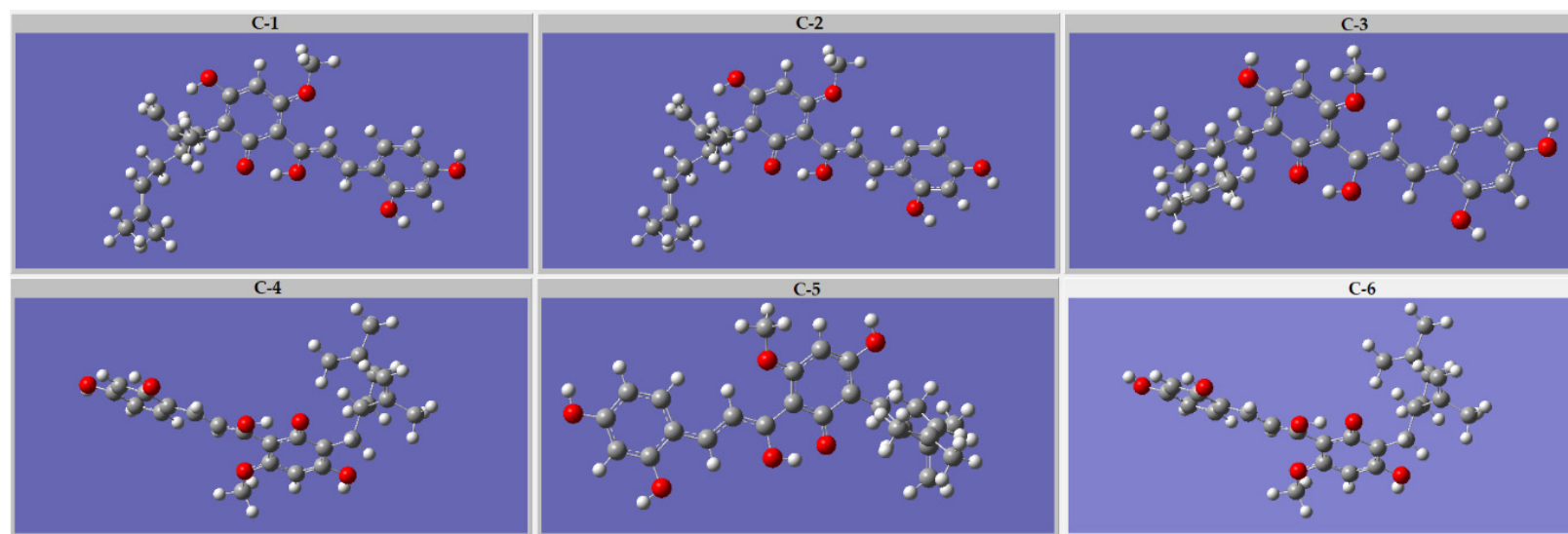
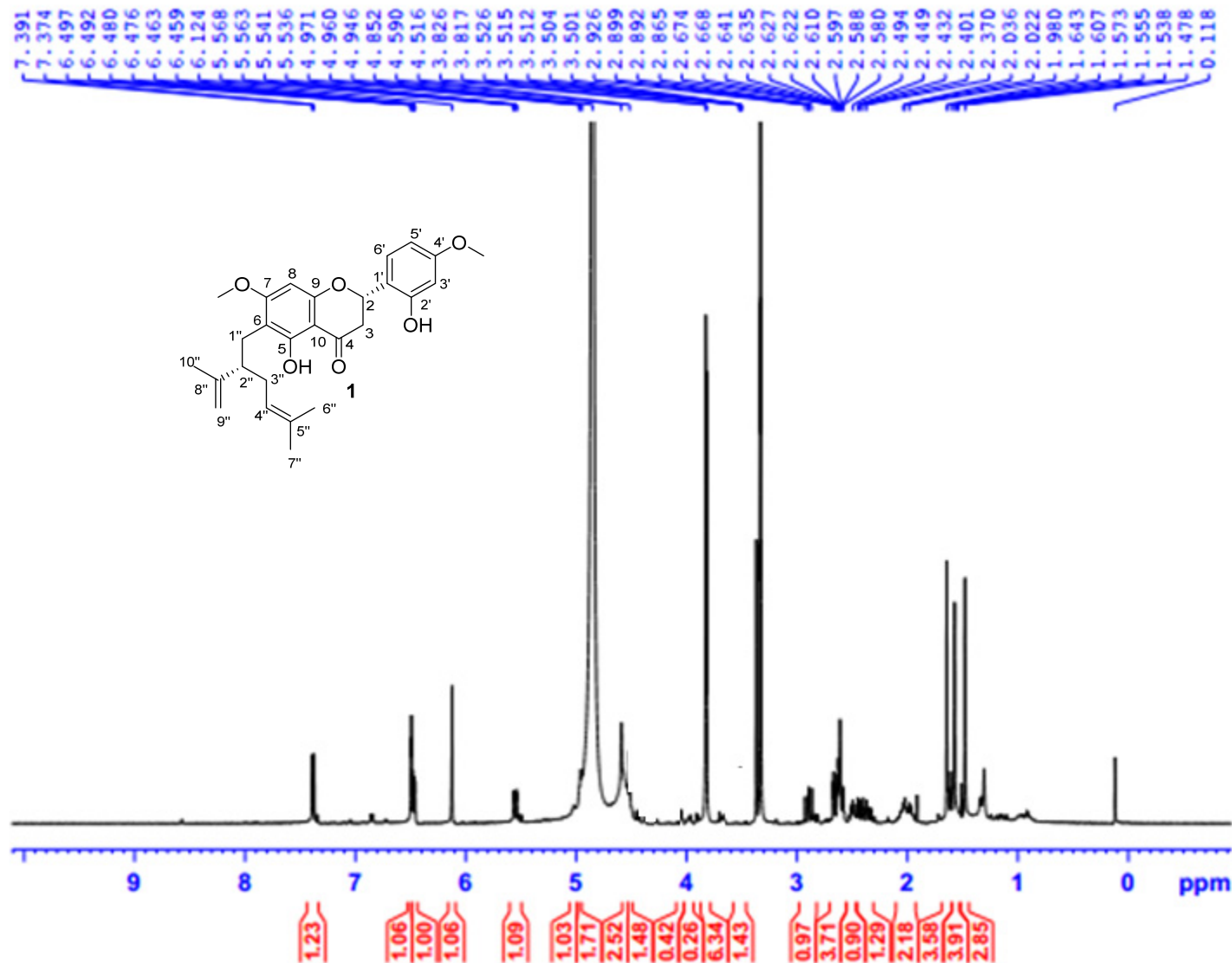


Figure 8S. Optimized conformers of (2''*S*)-3

8N17G248F10.21-MeOD-1H



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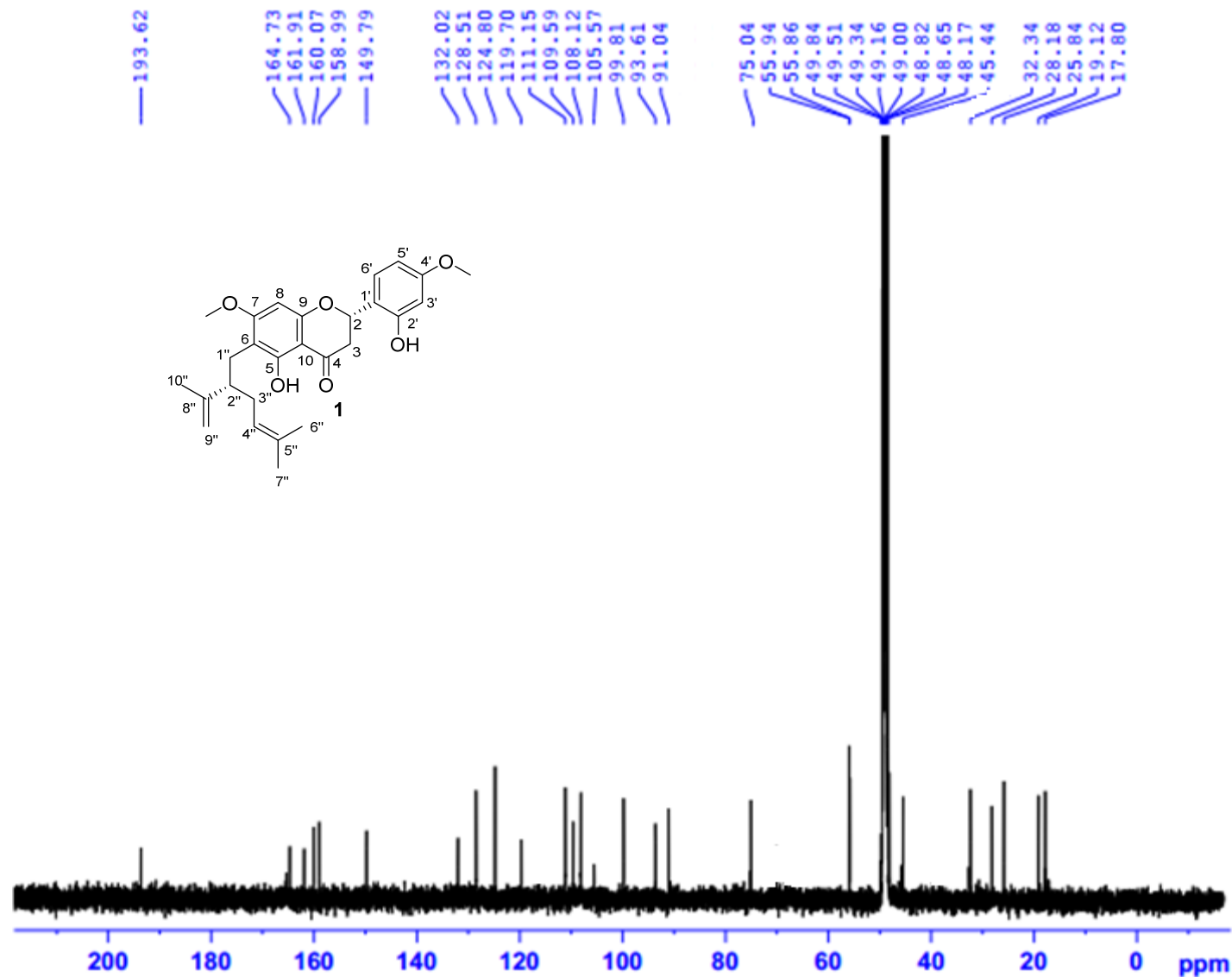
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 NUC1 1H
 P1 10.00 usec
 PLN1 22.00000000 W

F2 - Processing parameters
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 NCM EM
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 LB 0.30 Hz
 GB 0
 PC 1.00

Figure 9S. ¹H NMR spectrum of 1 (500 MHz, CD₃OD)

8N17G248F10.21-MeOD-C13CPD



Current Data Parameters
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 EXPNO 2
 PROCNO 1

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 PULPROG zgpg30
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 SOLVENT MeOD
 NS 4096
 DS 4
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 FIDRES 0.454131 Hz
 AQ 1.1010048 sec
 RG 198.57
 DW 16.800 usec
 DE 6.50 usec
 TE 304.4 K
 D1 2.00000000 sec
 D11 0.03000000 sec
 TDO 1

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 NUC1 13C
 P1 10.00 usec
 PLW1 88.00000000 W

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 SFO2 500.2020008 MHz
 NUC2 1H
 CPDPRG[2] waltz16
 PCPD2 80.00 usec
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 PLW12 0.34375000 W
 PLW13 0.22000000 W

F2 - Processing parameters
 SI 32768
 SF 125.7753900 MHz
 WDW EM
 SSB 0
 LB 1.00 Hz
 GB 0
 PC 1.40

Figure 10S. ¹³C NMR spectrum of 1 (125 MHz, CD₃OD)

8N17G248F10.21-MeOD-COSYGP



Current Data Parameters
 NAME 2VAN_8N17G248F10.21
 EXPNO 7
 PROCNO 1

F2 - Acquisition Parameters
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 PULPROG cosygpppqf
 TD 2048
 SOLVENT MeOD
 NS 4
 DS 8
 SWH 4166.667 Hz
 FIDRES 2.034505 Hz
 AQ 0.2457600 sec
 RG 79.36
 DW 120.000 usec
 DE 6.50 usec
 TE 303.5 K
 D0 0.00000300 sec
 D1 1.90743005 sec
 D11 0.03000000 sec
 D12 0.00002000 sec
 D13 0.00000400 sec
 D16 0.00020000 sec
 IN0 0.00024000 sec

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 NUC1 1H
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 P1 10.00 usec
 P17 2500.00 usec
 PLN1 22.00000000 W
 PLN0 3.25440001 W

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 GP21 10.00 %
 P16 1000.00 usec

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 FIDRES 32.552082 Hz
 SW 8.330 ppm
 FwMOE QF

F2 - Processing parameters
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 SF 500.2000590 MHz
 WDW QSINE
 SSB 0
 LB 0 Hz
 GB 0
 PC 1.40

F1 - Processing parameters
 SI 1024
 MC2 QF
 SF 500.2000590 MHz
 WDW QSINE
 SSB 0
 LB 0 Hz
 GB 0

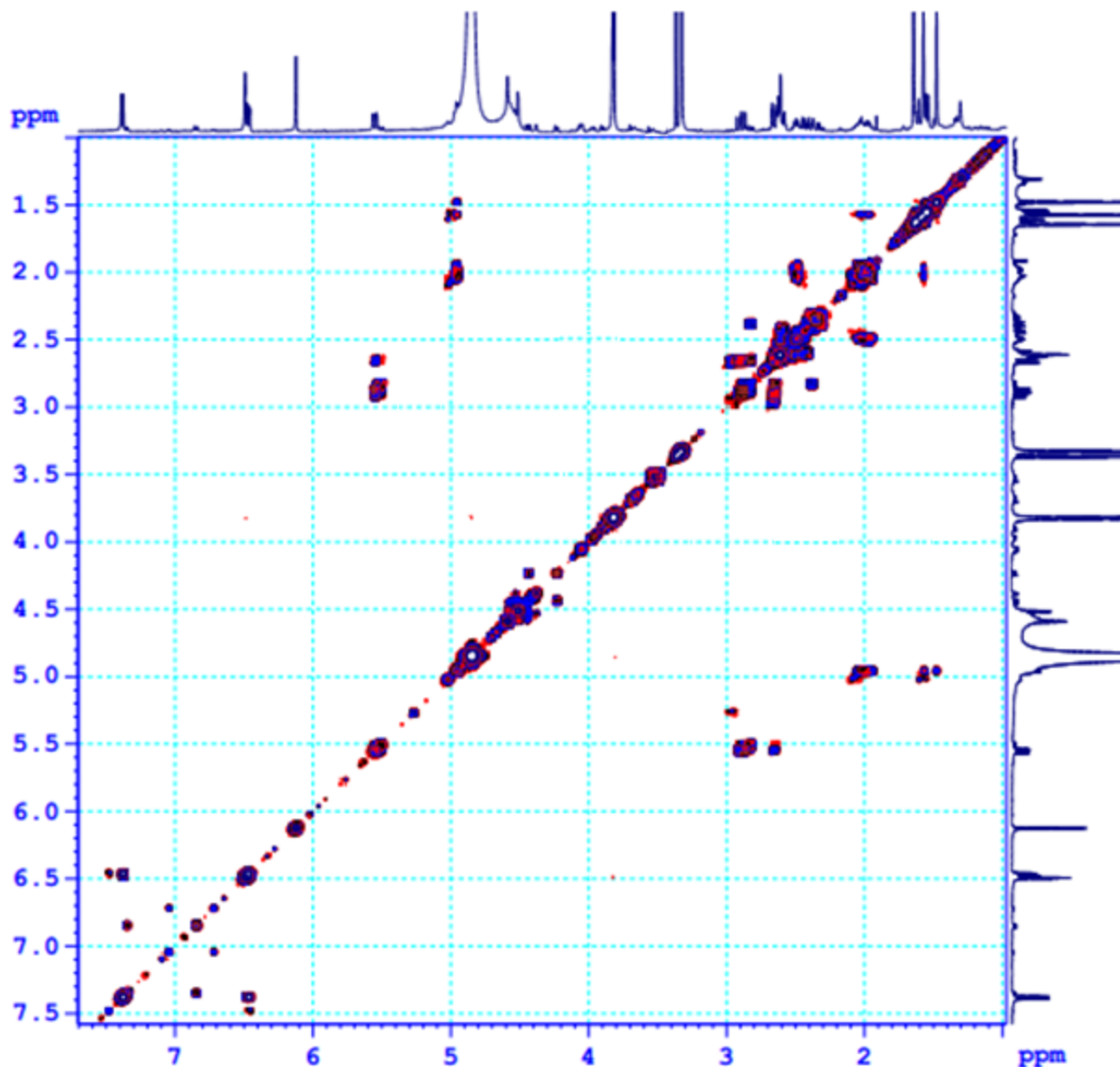


Figure 11S. ¹H-¹H COSY spectrum of 1 (500 MHz, CD₃OD)

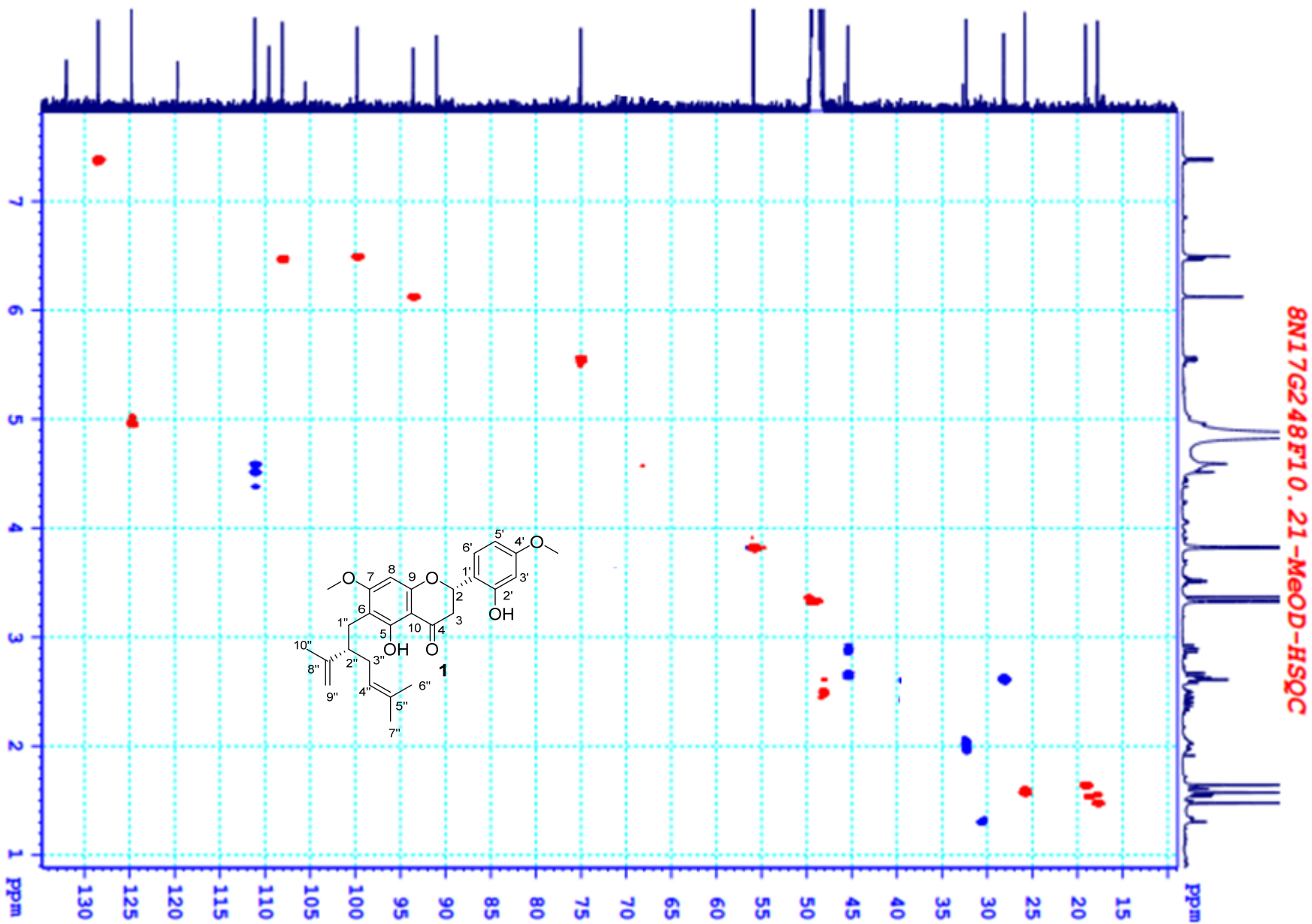


Figure 12S. HSQC spectrum of **1** (^1H : 500 MHz, ^{13}C : 125 MHz, CD_3OD)

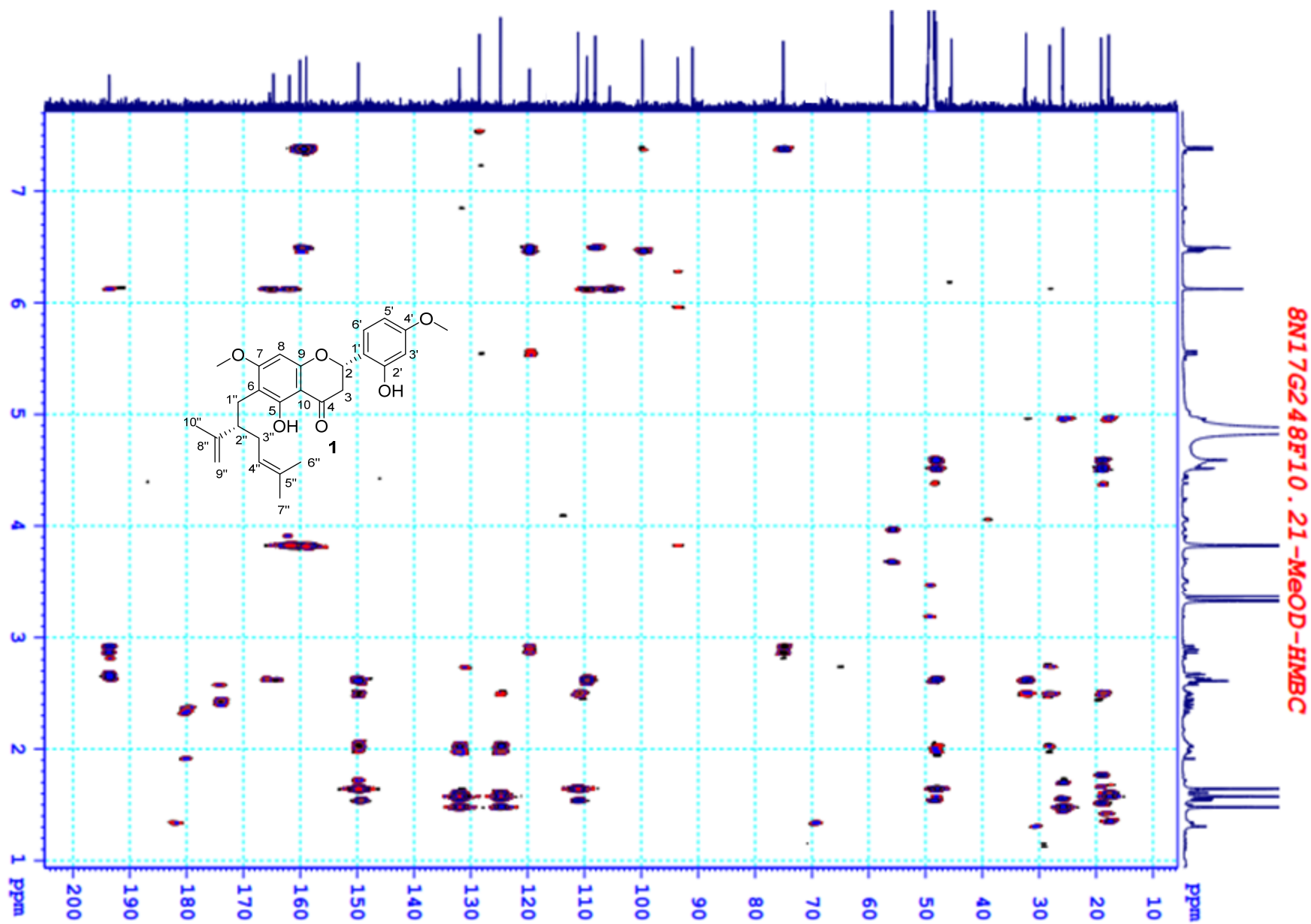
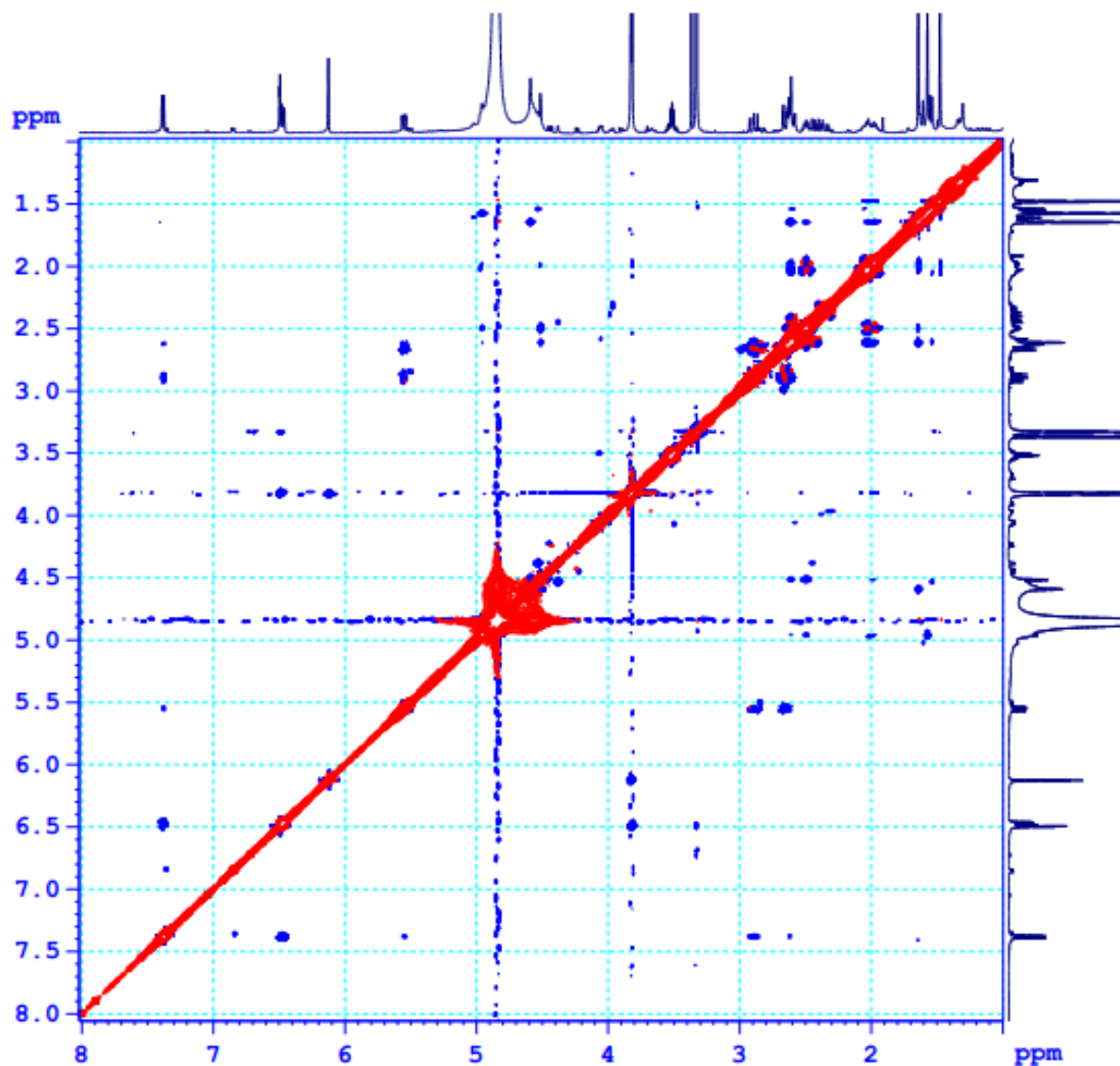


Figure 13S. HMBC spectrum of 1 (^1H : 500 MHz, ^{13}C : 125 MHz, CD_3OD)

8N17G248F10.21-MeOD-ROESY



Current Data Parameters
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 PROCNO 1

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 AQ 0.2457600 sec
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 DE 6.50 usec
 TE 303.7 K
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 L4 1000
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 LB 0 Hz
 GB 0
 PC 1.00

F1 - Processing parameters
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 WDW QSINE
 SSB 2
 LB 0 Hz
 GB 0

Figure 14S. ROESY spectrum of 1 (500 MHz, CD₃OD)

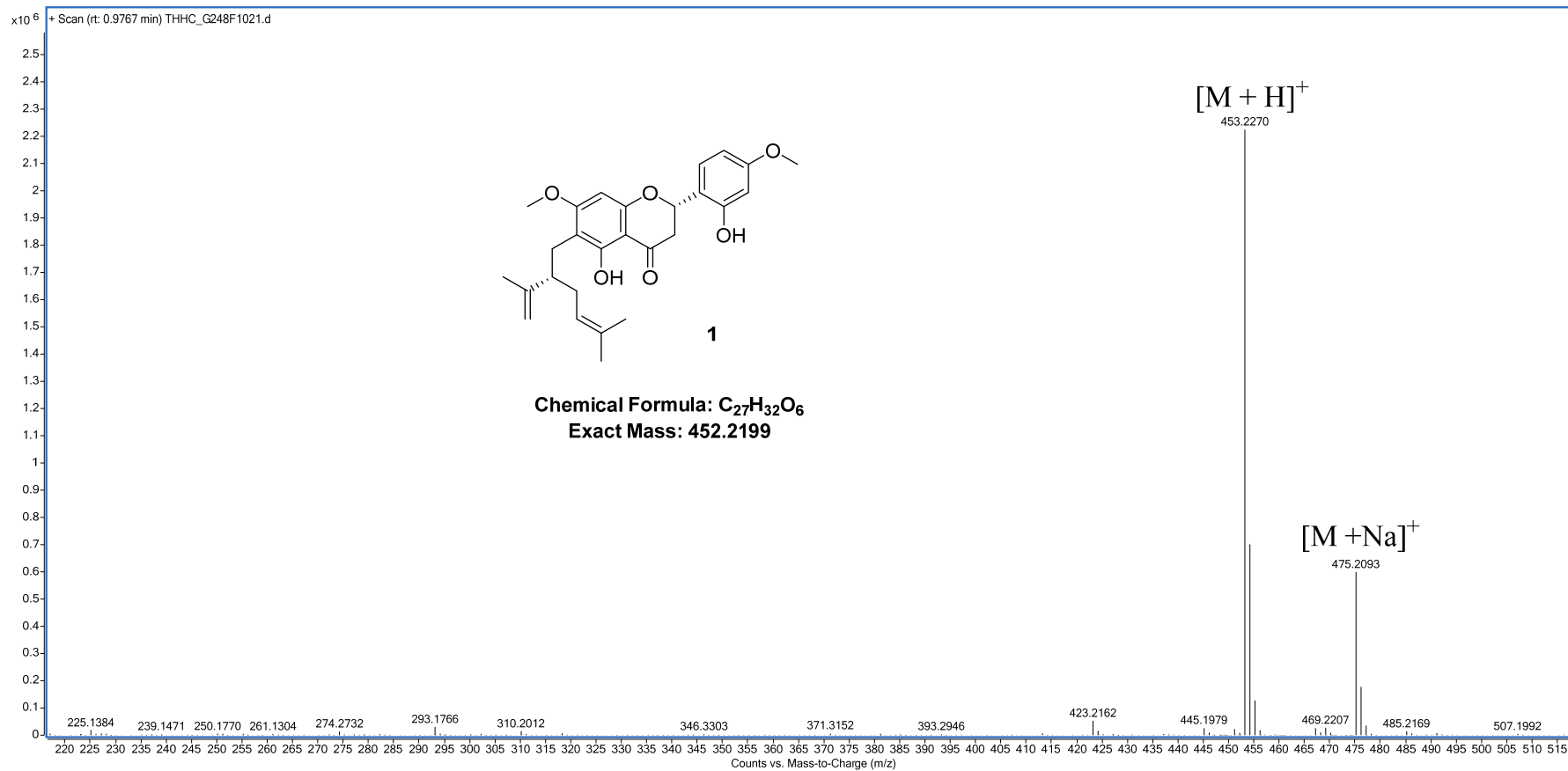


Figure 15S. HR-ESI-MS of **1**

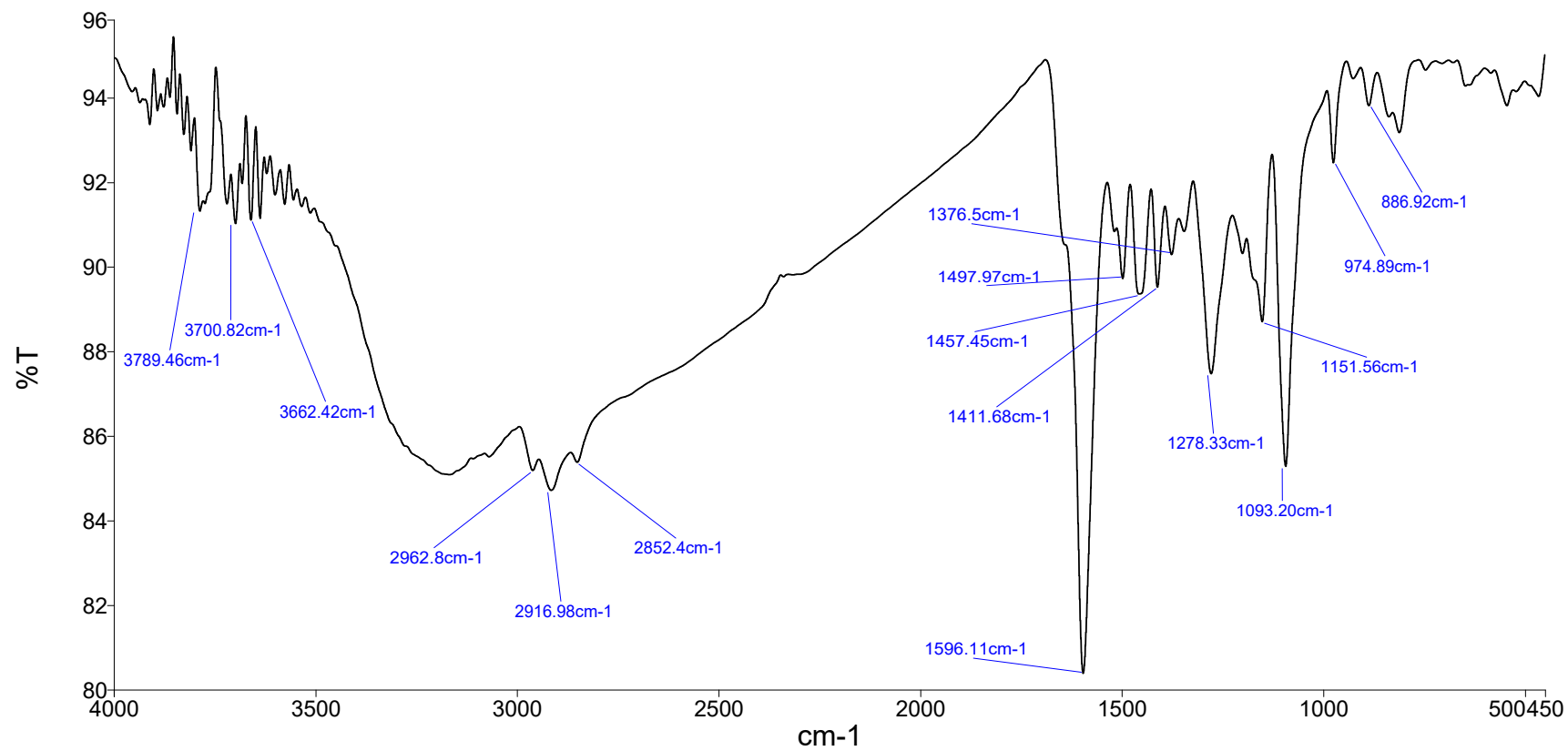


Figure 16S. IR spectrum of **1**

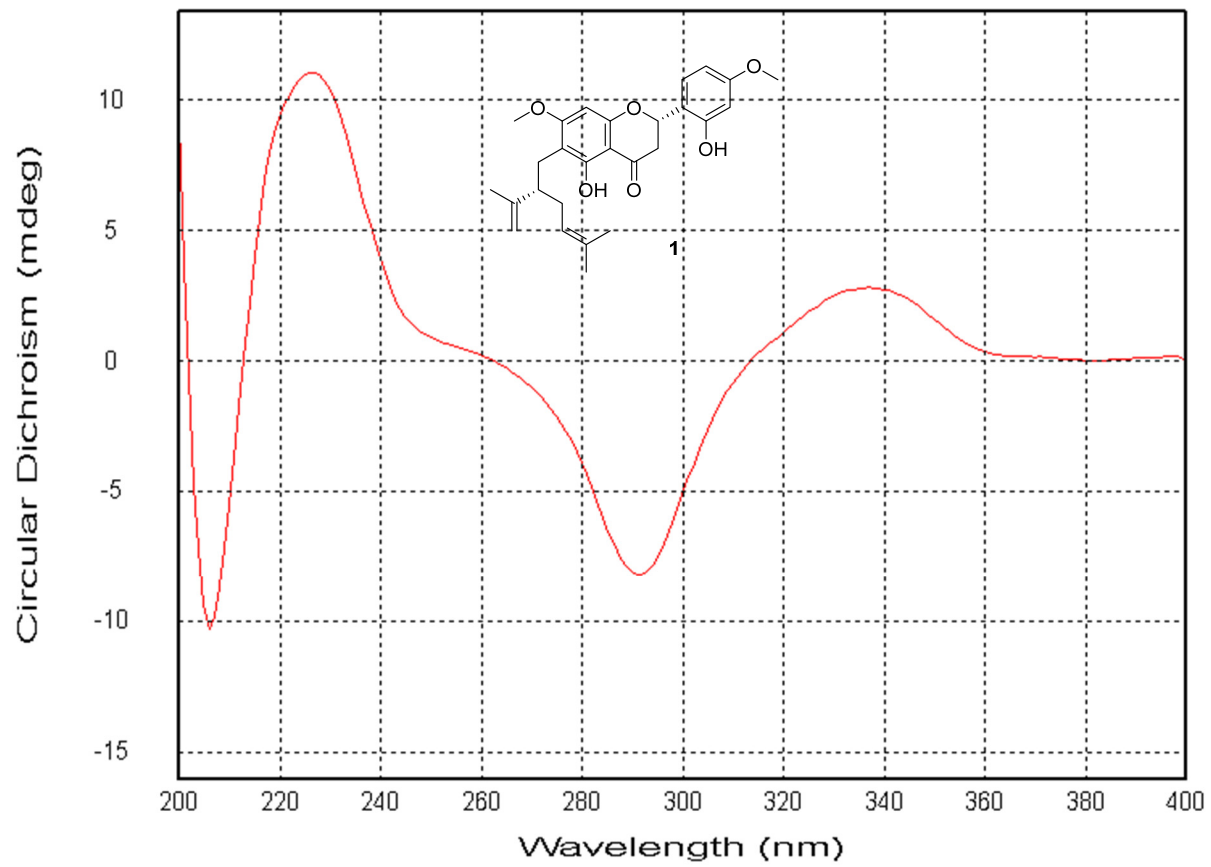
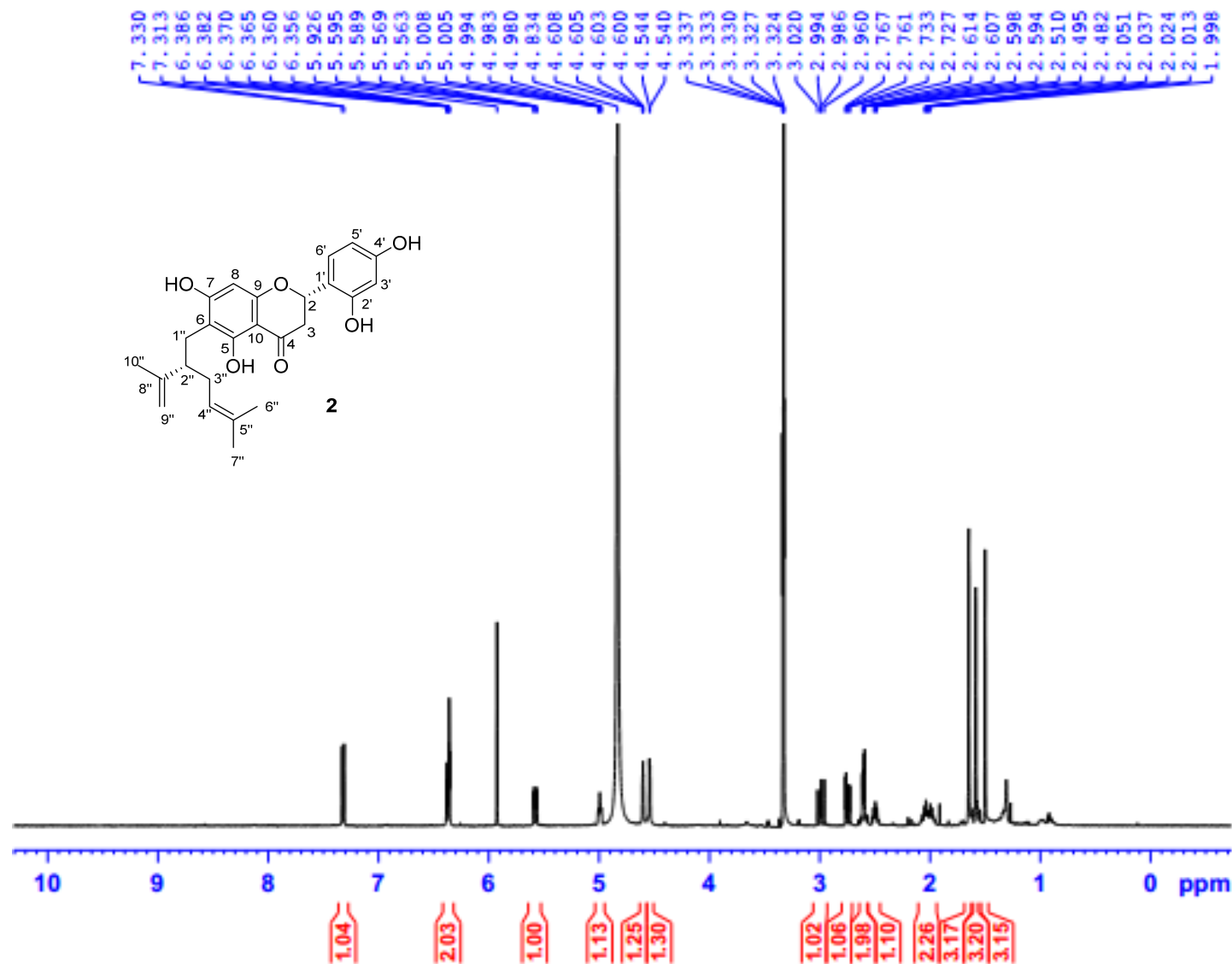


Figure 17S. CD spectrum of **1**

2N17G248F1041-MeOD-1H



Current Data Parameters
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 PROCNO 1

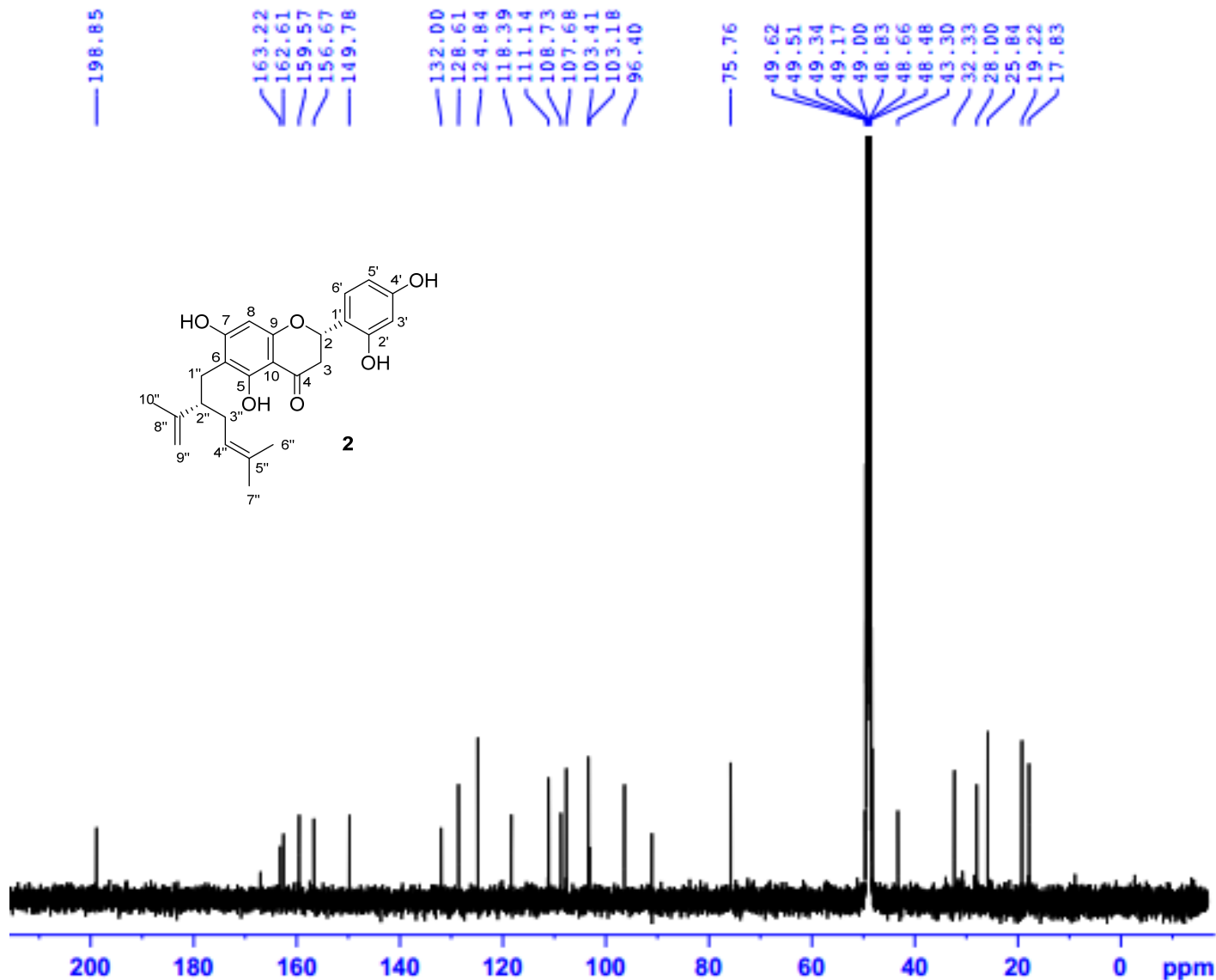
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 RG 157.35
 DW 50.000 usec
 DE 6.50 usec
 TE 303.0 K
 D1 1.00000000 sec
 TD0 1

CHANNEL f1
 SFO1 500.2030889 MHz
 NUC1 1H
 P1 10.00 usec
 PLN1 22.00000000 W

F2 - Processing parameters
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 SF 500.1999996 MHz
 WCW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00

Figure 18S. ¹H NMR spectrum of 2 (500 MHz, CD₃OD)

2N17G248F1041-MeOD-C13CPD



Current Data Parameters
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 PROCNO 1

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 FIDRES 0.454131 Hz
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 D11 0.03000000 sec
 TDO 1

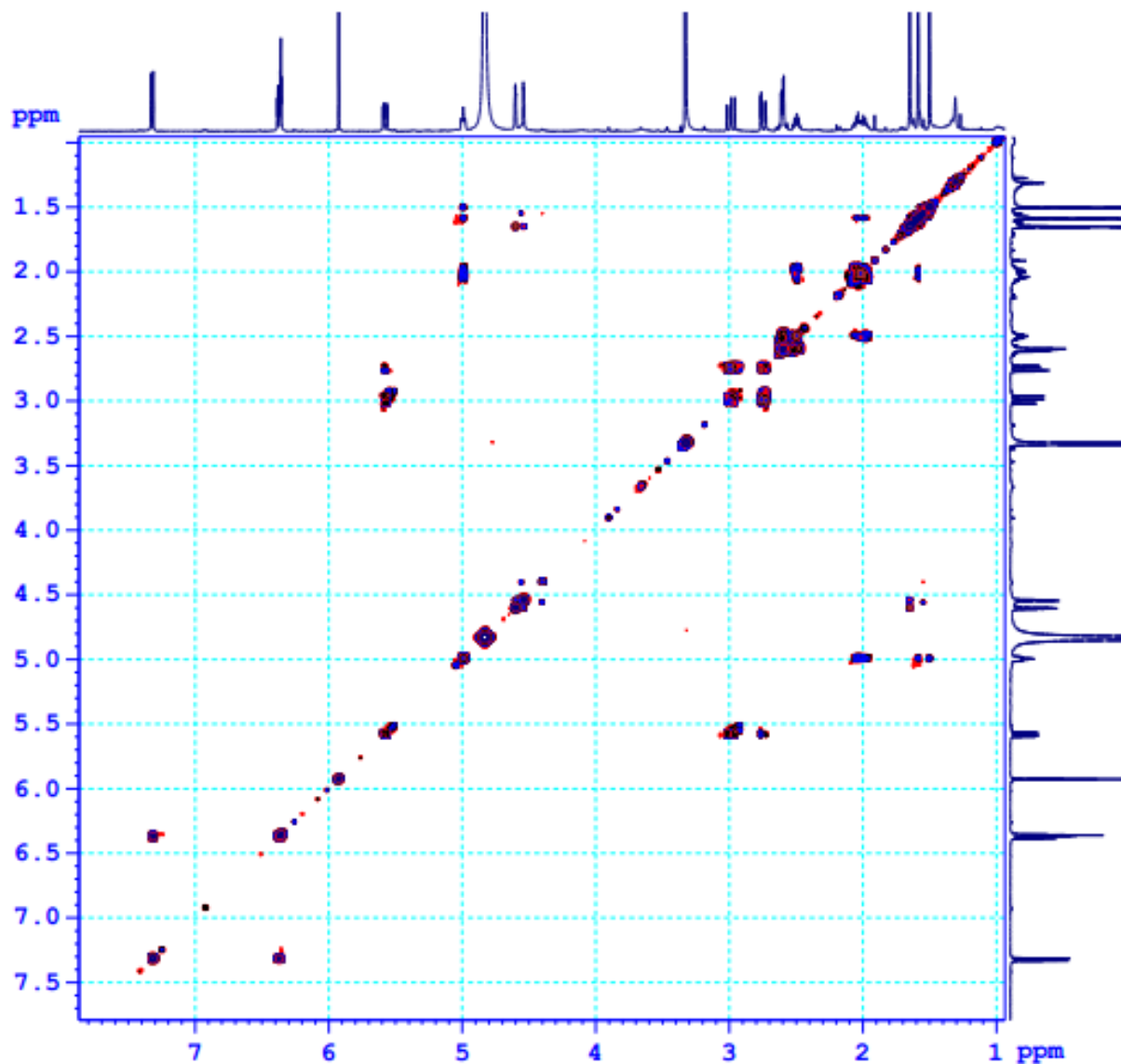
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 PLW1 88.00000000 W

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 NUC2 1H
 CPDPRG[2] waltz16
 PCPD2 80.00 usec
 PLW2 22.00000000 W
 PLW12 0.34375000 W
 PLW13 0.22000000 W

F2 - Processing parameters
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 SF 125.7753900 MHz
 WDW EM
 SSB 0
 LB 1.00 Hz
 GB 0
 PC 1.40

Figure 19S. ¹³C NMR spectrum of 2 (125 MHz, CD₃OD)

2N17G248F1041-MeOD-COSYGP



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 PROCNO 1

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 Time 8.49
 INSTRUM spect
 PROBNM 5 mm PABBO BB/
 PULPROG cosygpppqf
 TD 2048
 SOLVENT MeOD
 NS 4
 DS 8
 SMH 3875.969 Hz
 FIDRES 1.892563 Hz
 AQ 0.2641920 sec
 RG 64.21
 DW 129.000 usec
 DE 6.50 usec
 TE 303.0 K
 D0 0.00000300 sec
 D1 1.88899803 sec
 D11 0.03000000 sec
 D12 0.00002000 sec
 D13 0.00000400 sec
 D16 0.00020000 sec
 INU 0.00025800 sec

----- CHANNEL f1 -----
 SFO1 500.2020264 MHz
 NUC1 1H
 P0 10.00 usec
 P1 10.00 usec
 P17 2500.00 usec
 PLW1 22.00000000 W
 PLW0 3.25440001 W

----- GRADIENT CHANNEL -----
 GPNAM[1] SMSQ10.100
 GPE1 10.00 %
 P16 1000.00 usec

F1 - Acquisition parameters
 TD 128
 SFO1 500.202 MHz
 FIDRES 60.562016 Hz
 SW 7.749 ppm
 FoMODE QF

F2 - Processing parameters
 SI 1024
 SF 500.2000019 MHz
 WDW QSINE
 SSB 0
 LB 0 Hz
 GB 0
 PC 1.40

F1 - Processing parameters
 SI 1024
 MC2 QF
 SF 500.2000027 MHz
 WDW QSINE
 SSB 0
 LB 0 Hz
 GB 0

Figure 20S. ¹H-¹H COSY spectrum of 2 (500 MHz, CD₃OD)

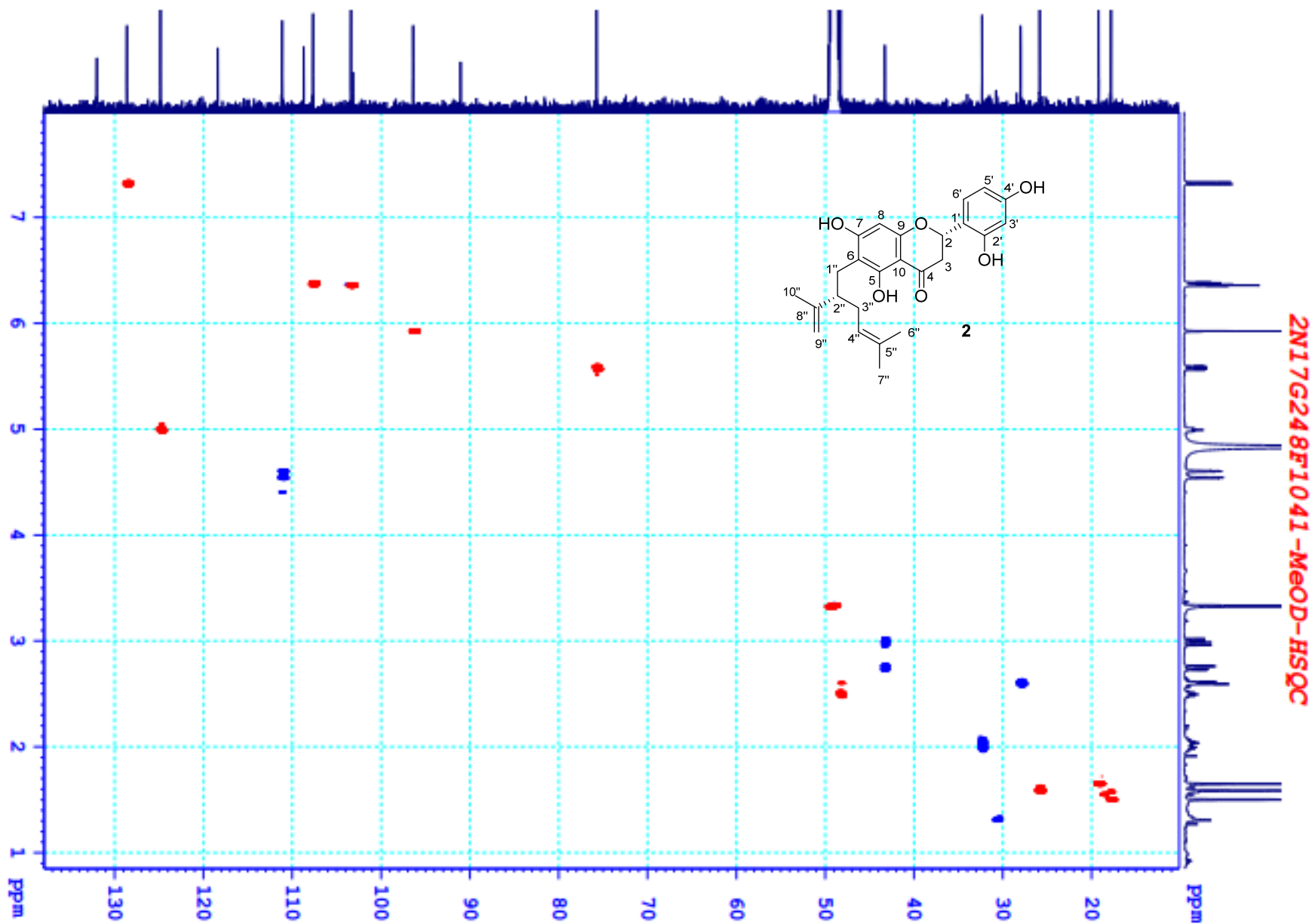


Figure 21S. HSQC spectrum of 2 (^1H : 500 MHz, ^{13}C : 125 MHz, CD_3OD)

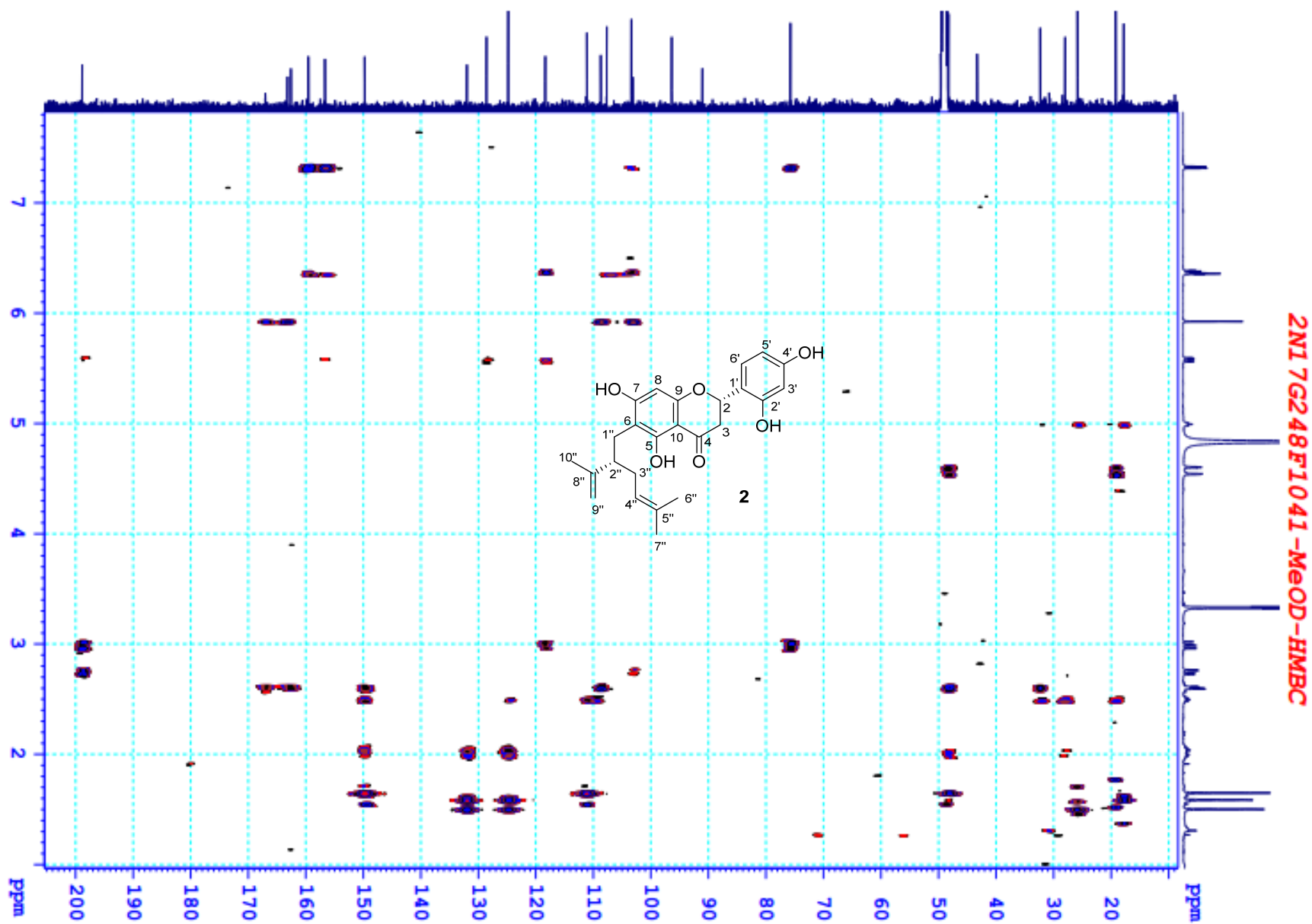
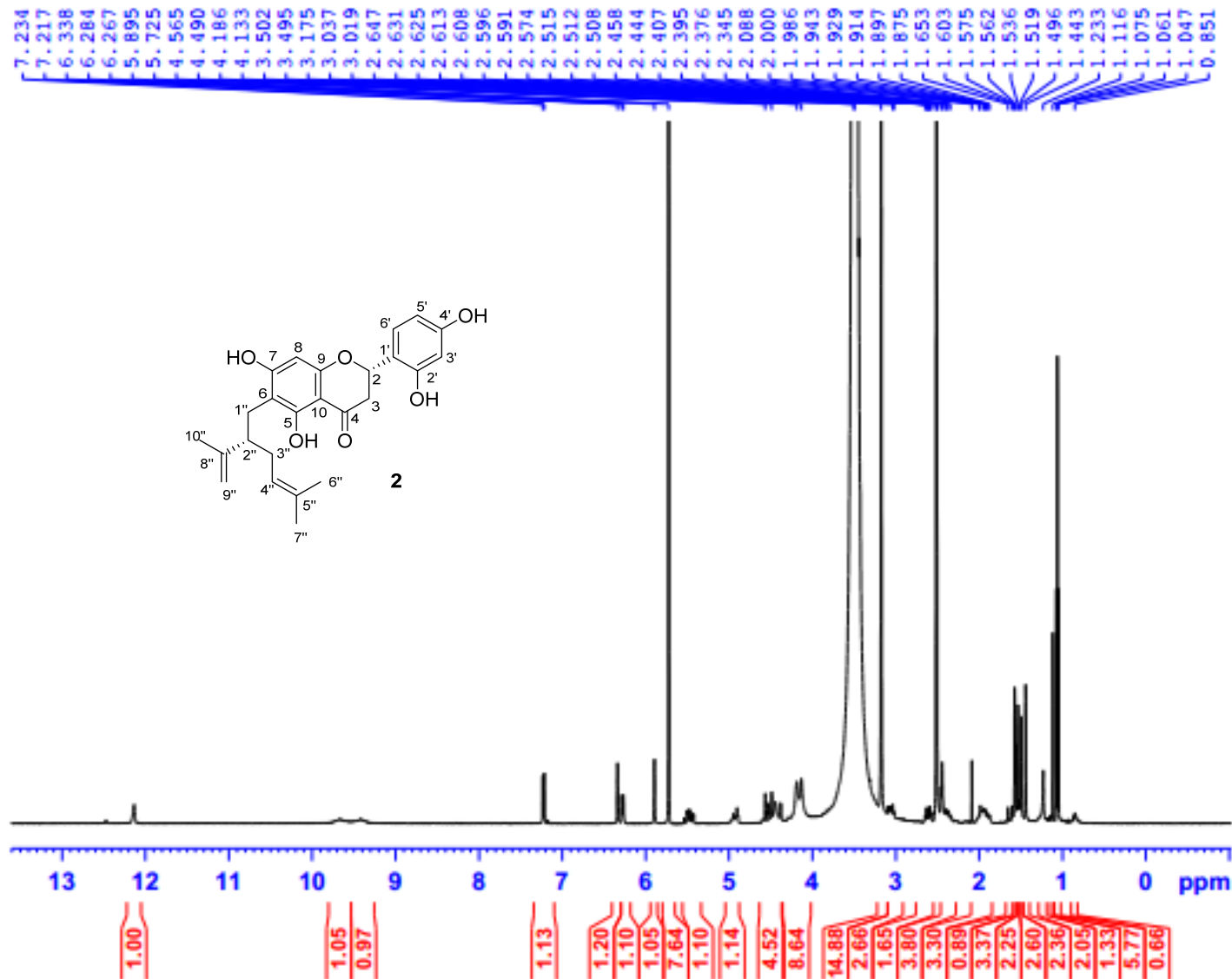


Figure 22S. HMBC spectrum of 2 (^1H : 500 MHz, ^{13}C : 125 MHz, CD_3OD)

2N17G248F10.41.1-DMSO-1H



Current Data Parameters
 NAME 2VAN_2n17g248r10.41.
 EXPNO 1
 PROCNO 1

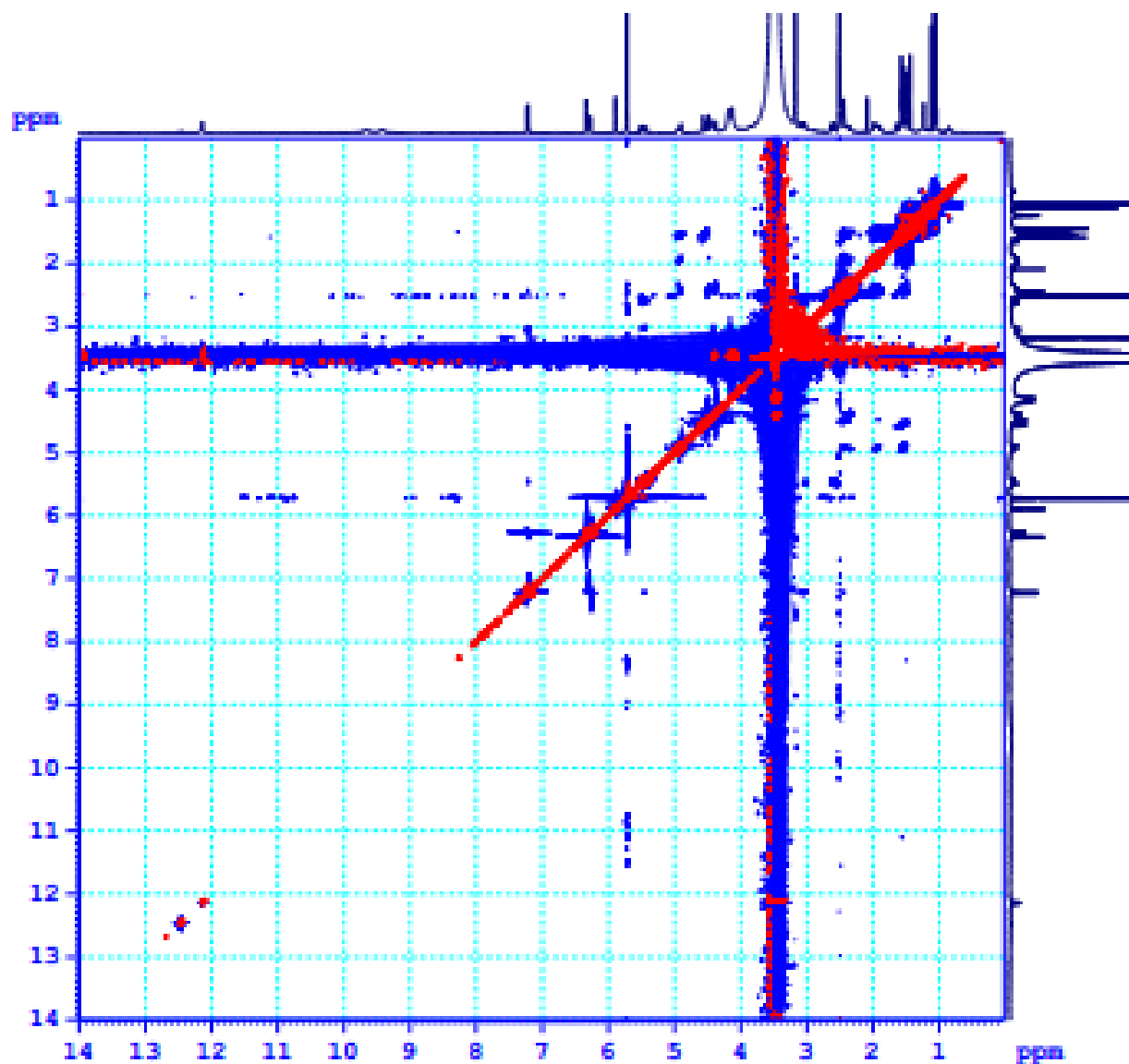
F2 - Acquisition Parameters
 Date_ 20171108
 Time 17.23
 INSTRUM spect
 PROBHD 5 mm PABBO BB/
 PULPROG zg30
 TD 65536
 SOLVENT DMSO
 NS 16
 DS 2
 SSB 10000.000 Hz
 FIDRES 0.152588 Hz
 AQ 3.2767999 sec
 RG 30.85
 DW 50.000 usec
 DE 6.50 usec
 TE 303.0 K
 D1 1.00000000 sec
 TDO 1

----- CHANNEL f1 -----
 SFO1 500.2030889 MHz
 NUC1 1H
 P1 10.00 usec
 PLN1 22.00000000 W

F2 - Processing parameters
 SI 65536
 SF 500.2000000 MHz
 WCN EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00

Figure 23S. ¹H NMR spectrum of **2** (500 MHz, DMSO-*d*₆)

2N17G248F10.41.1-DMSO-ROESY



Current Data Parameters
 NAME 2N17G248F10.41.1
 EXPNO 2
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20171108
 Time 18.37
 INSTRUM spect
 PROCNO 2 of 2
 PULPROG zgpg30
 ID 1000
 SOLVENT DMSO
 NS 18
 DS 4
 SWH 7887.171 Hz
 FIDRES 0.112881 Hz
 AQ 0.580000 sec
 RG 327.50
 IN 184.000 MHz
 DE 0.00
 TE 300.2 K
 DD 0.00010000 sec
 DI 1.82187187 sec
 D11 0.00000000 sec
 D12 0.00000000 sec
 D13 0.00010000 sec
 LE 1000
 F1 30000.00 MHz

===== CHANNEL f1 =====
 NUC1 1H
 P1 10.00 MHz
 P1P 1000.00 MHz
 P2 100.00 MHz
 PL1 31.0000000 W
 PL12 1.78100001 W
 PL13 0.00000000 W

F1 - Acquisition parameters
 NS 184
 SFO1 500.136116 MHz
 FREQ1 50.000000 MHz
 NU 1.074 ppm
 PULPROG zgpg30

F2 - Processing parameters
 SI 32768
 SF 500.136116 MHz
 NS 184
 DS 4
 SWH 7887.171 MHz
 IN 184.000 MHz
 FE 0.112881 MHz
 F1 30000.00 MHz

F1 - Processing parameters
 SI 32768
 SF 500.136116 MHz
 NS 184
 DS 4
 SWH 7887.171 MHz
 IN 184.000 MHz
 FE 0.112881 MHz

Figure 24S. ROESY spectrum of 2 (500 MHz, DMSO-*d*₆)

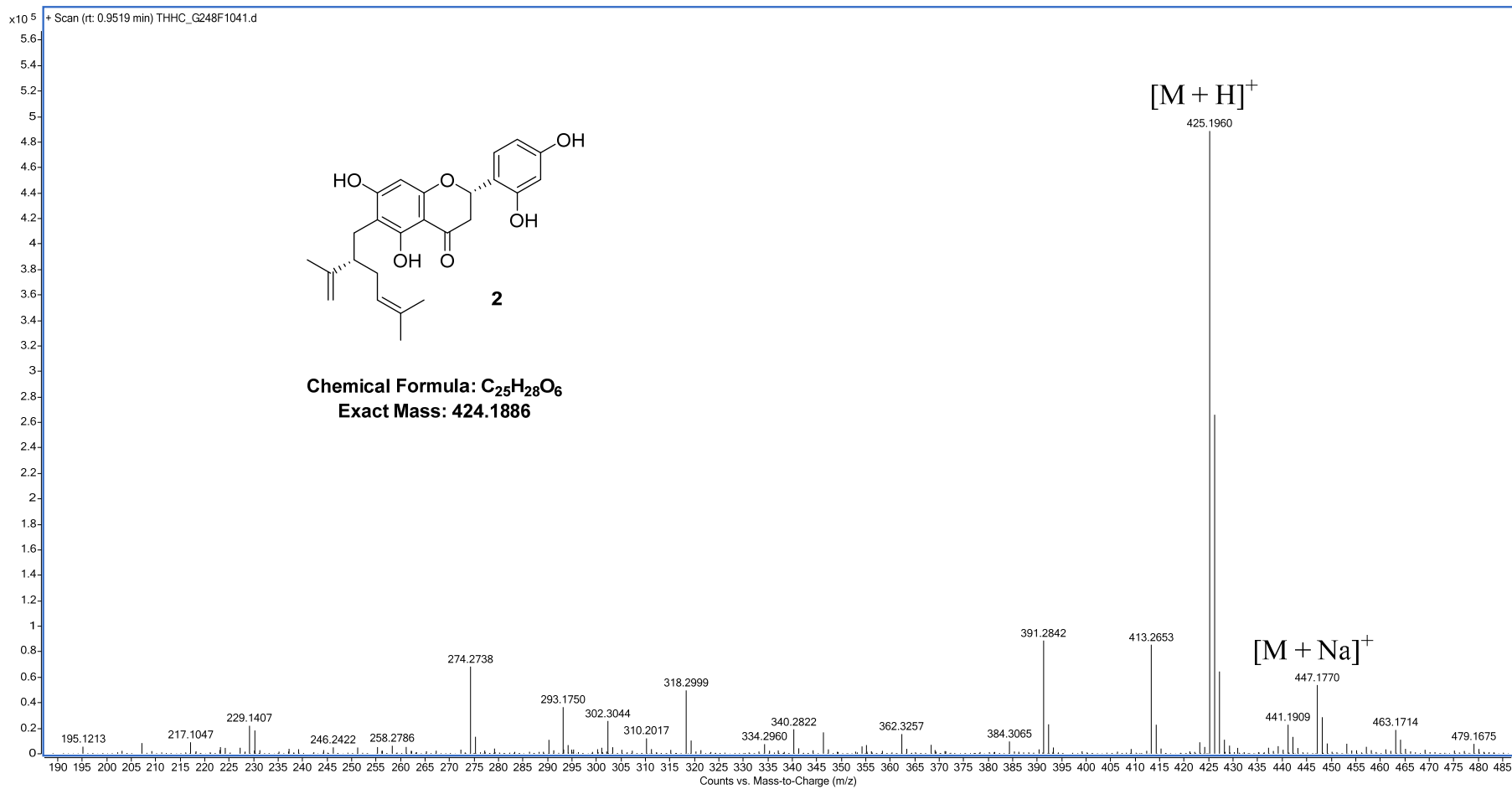


Figure 25S. HR-ESI-MS of **2**

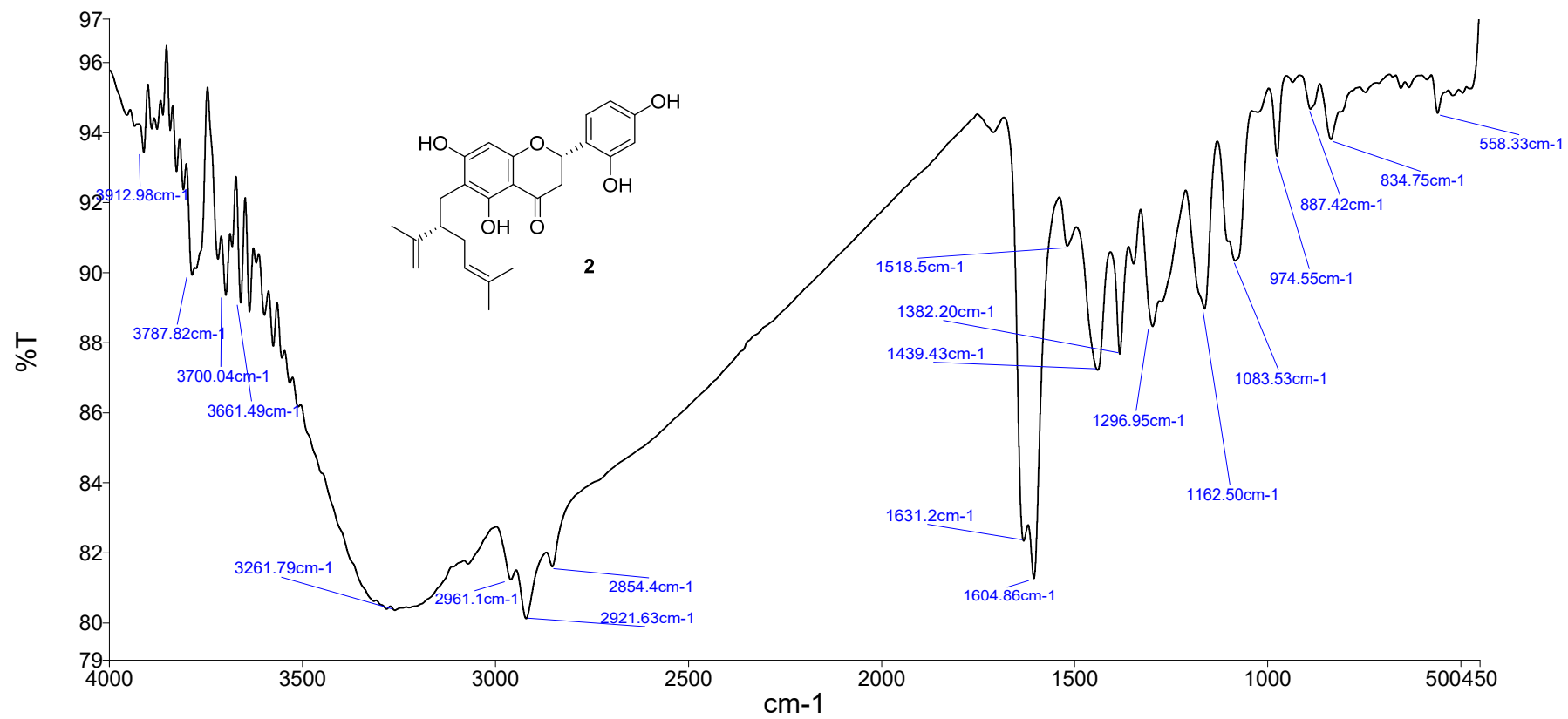


Figure 26S. IR spectrum of 2

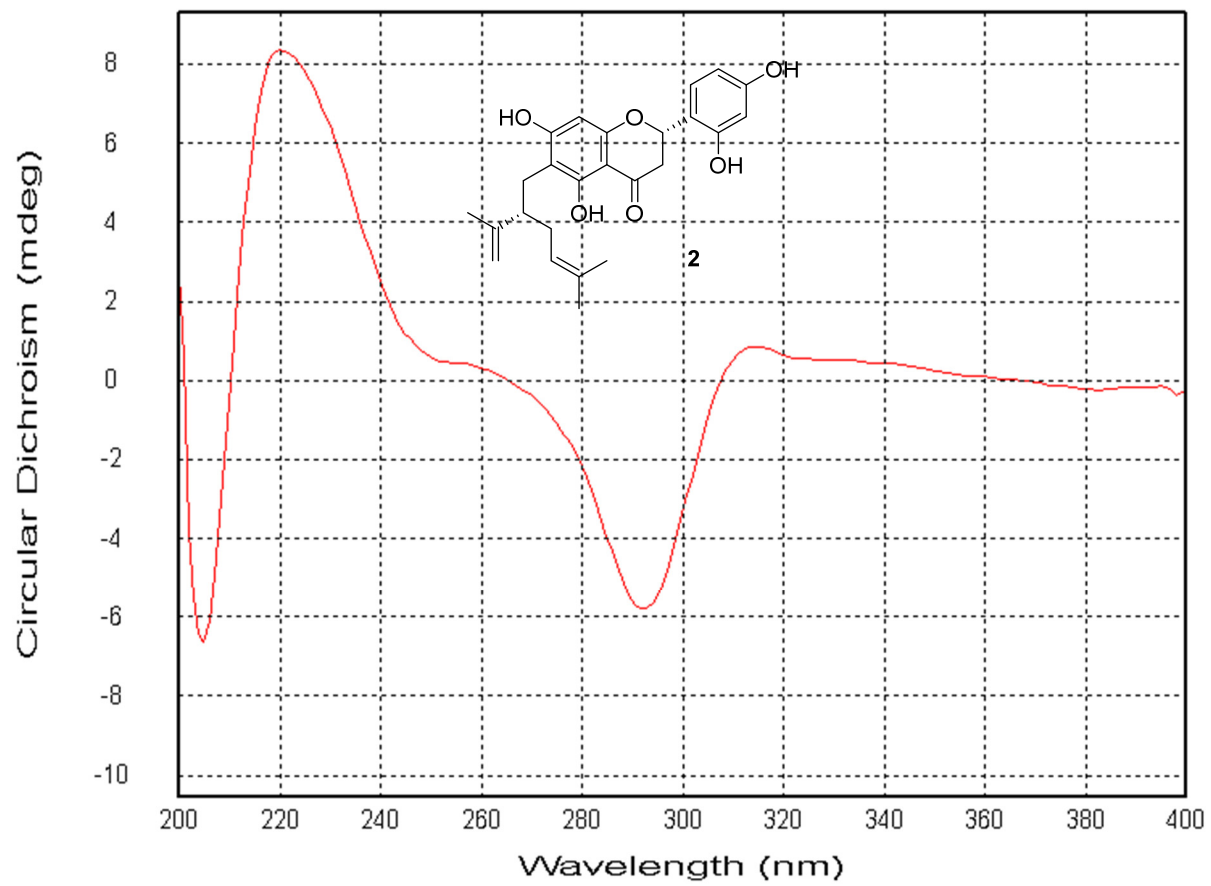


Figure 27S. CD spectrum of **2**

2N17G248F1042-MeOD-1H



Current Data Parameters
 NAME ZVAN_2N17G248F1042
 EXPNO 1
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20171102
 Time 11.22
 INSTRUM spect
 PROBHD 5 mm PABBO BB/
 PULPROG zg30
 TD 65536
 SOLVENT MeOD
 NS 16
 DS 2
 SWH 10000.000 Hz
 FIDRES 0.152588 Hz
 AQ 3.2767999 sec
 RG 142.98
 DW 50.000 usec
 DE 6.50 usec
 TE 303.0 K
 D1 1.00000000 sec
 TDO 1

----- CHANNEL f1 -----
 SF01 500.2030889 MHz
 NUC1 1H
 P1 10.00 usec
 PLW1 22.00000000 W

F2 - Processing parameters
 SI 65536
 SF 500.2000001 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00

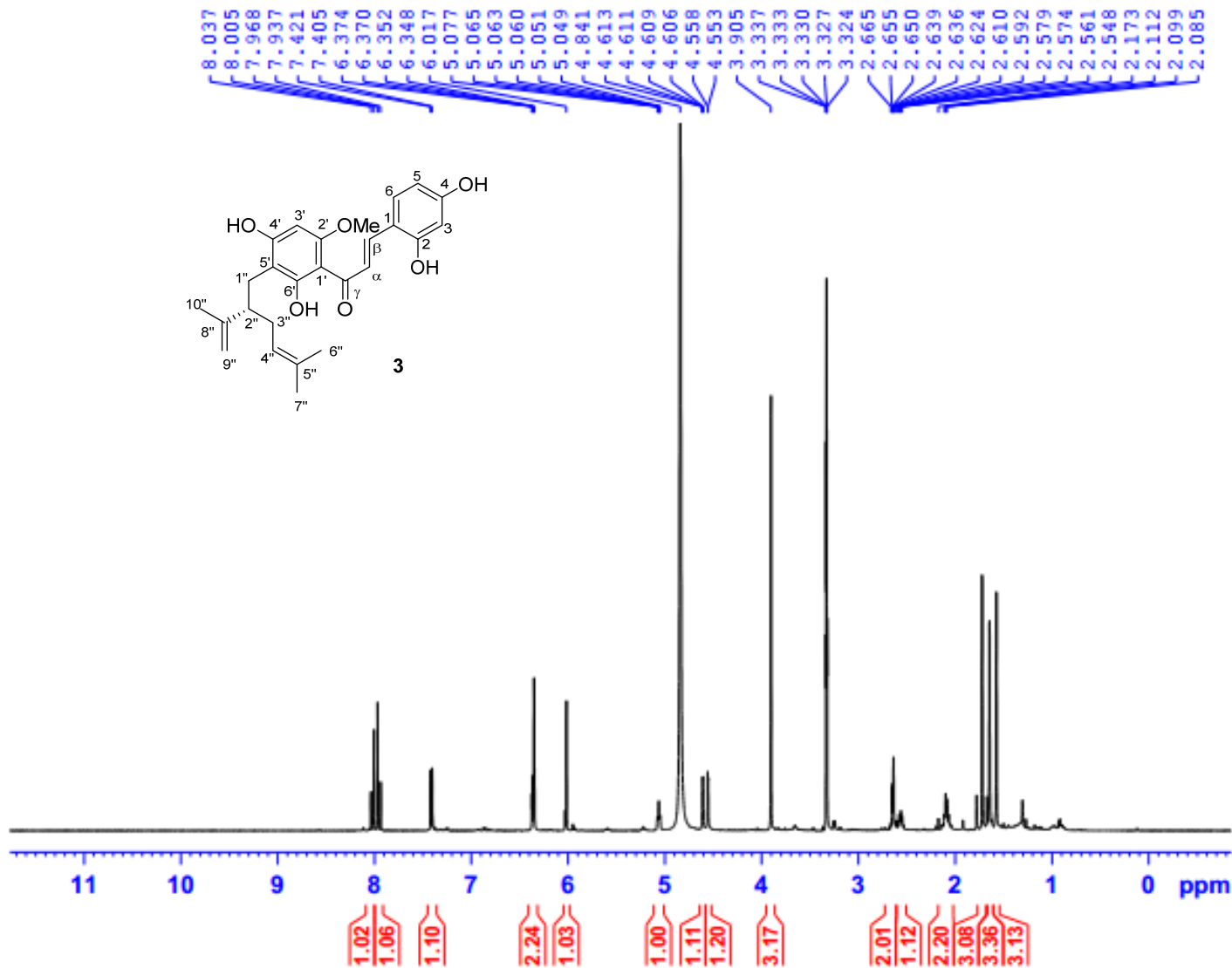
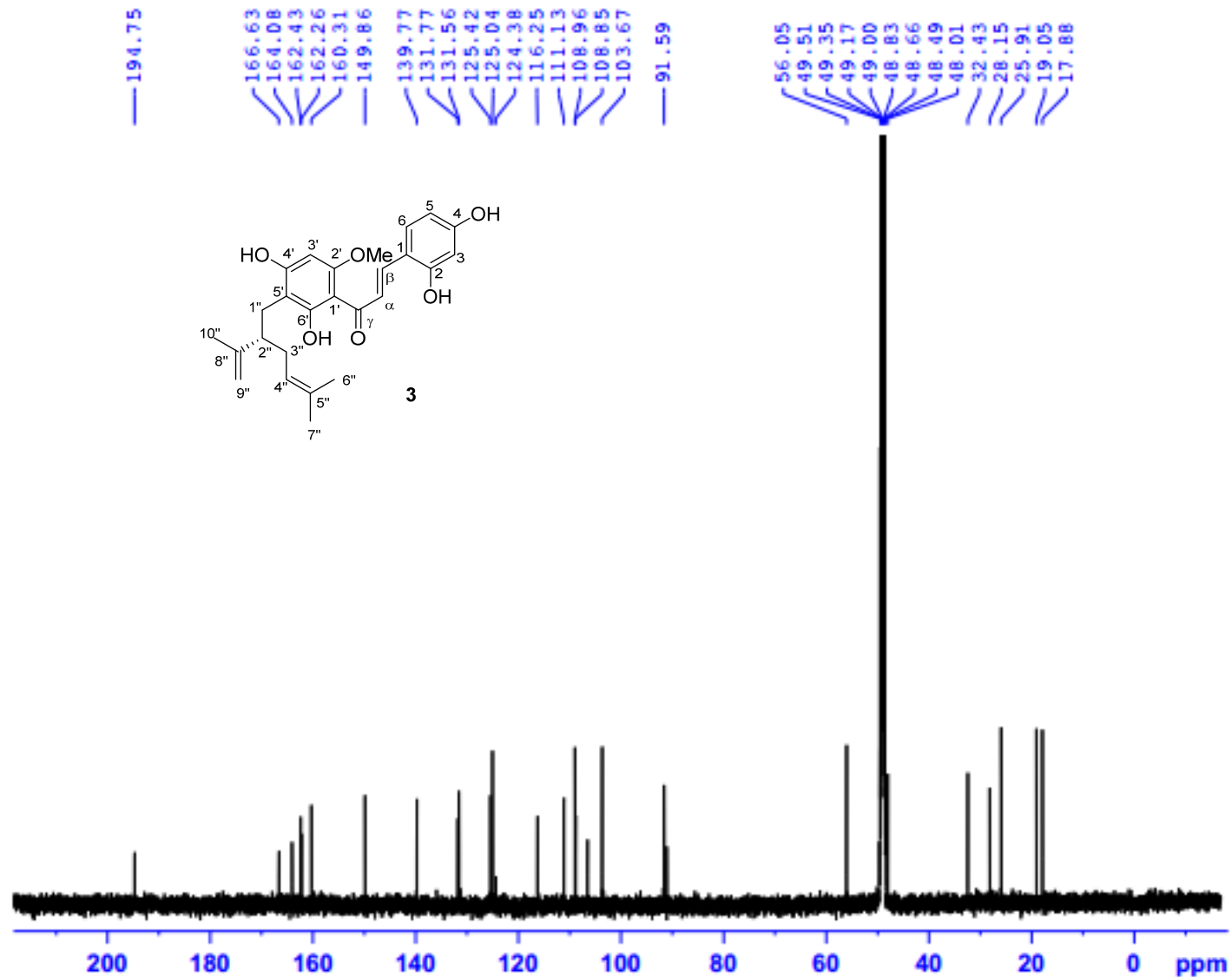


Figure 28S. ¹H NMR spectrum of 3 (500 MHz, CD₃OD)

2N17G248F1042-MeOD-C13CPD



Current Data Parameters
 NAME 2VAN_2N17G248F1042
 EXPNO 2
 PROCNO 1

F2 = Acquisition Parameters
 Date_ 20171103
 Time 20.17
 INSTRUM spect
 PROBHD 5 mm PABBO BB/
 PULPROG zgpg30
 TD 65536
 SOLVENT MeOD
 NS 2048
 DS 4
 SWH 29761.904 Hz
 FIDRES 0.454131 Hz
 AQ 1.1010048 sec
 RG 198.57
 DM 16.800 usec
 DE 6.50 usec
 TE 303.4 K
 D1 2.00000000 sec
 D11 0.03000000 sec
 TD0 1

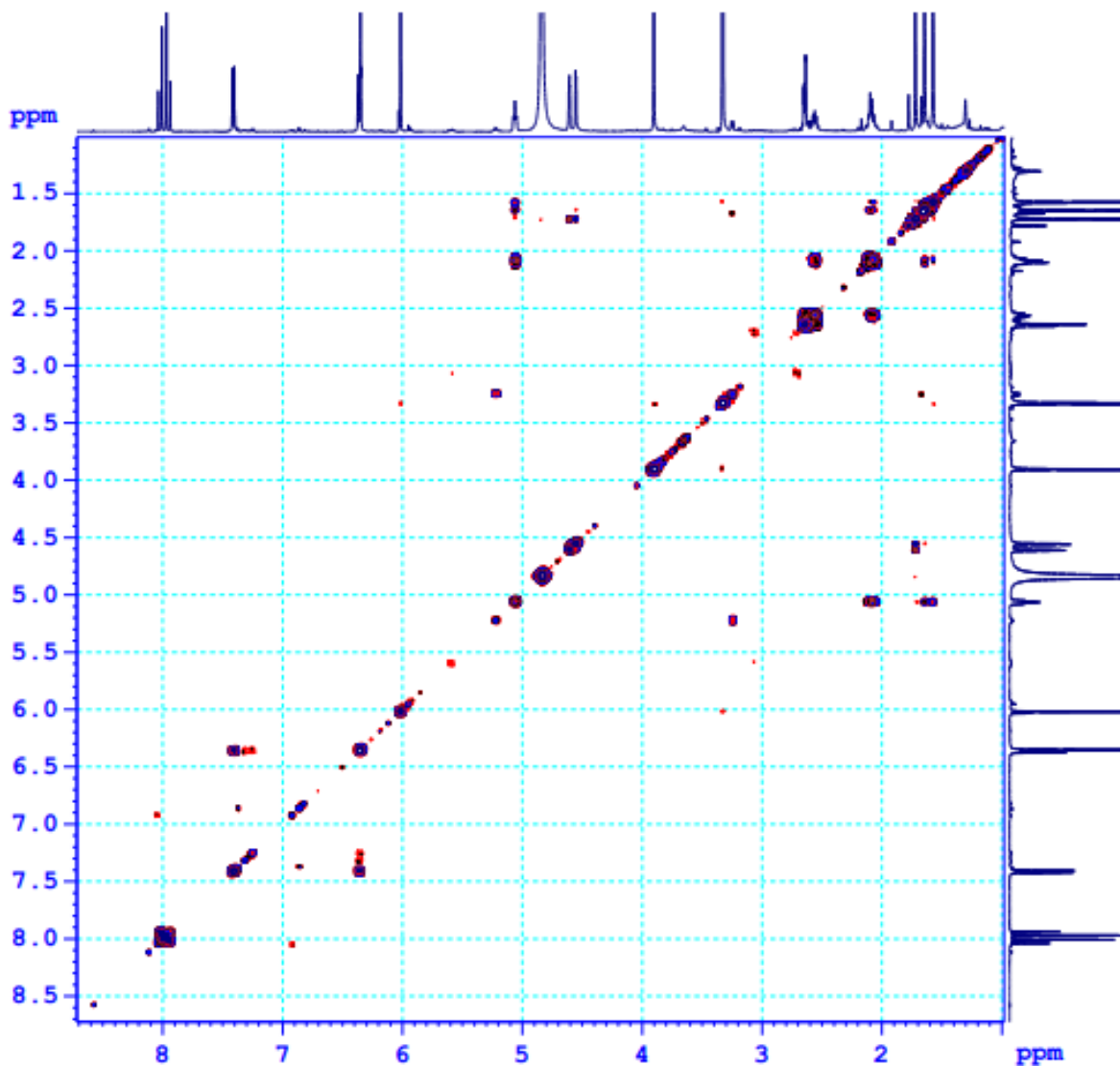
----- CHANNEL f1 -----
 SFO1 125.7879670 MHz
 NUC1 13C
 P1 10.00 usec
 PLM1 88.00000000 W

----- CHANNEL f2 -----
 SFO2 500.2020008 MHz
 NUC2 1H
 CPDPRG[2] waltz16
 PCPD2 80.00 usec
 PLM2 22.00000000 W
 PLM12 0.34375000 W
 PLM13 0.22000000 W

F2 = Processing parameters
 SI 32768
 SF 125.7753900 MHz
 NW EM
 SSB 0
 LB 1.00 Hz
 GB 0
 PC 1.40

Figure 29S. ¹³C NMR spectrum of 3 (125 MHz, CD₃OD)

2N17G248F1042-MeOD-COSYGP



```

Current Data Parameters
NAME      2VAN_2N17G248F1042
EXPNO     7
PROCNO    1

F2 - Acquisition Parameters
Date_     20171103
Time      22.36
INSTRUM   spect
PROBHD    5 mm FAMB0 BM/
PULPROG   cosygpppqf
TD        2048
SOLVENT   MeOD
NS         2
DS         8
SWH        4424.779 Hz
FIDRES     2.160537 Hz
AQ         0.2314240 sec
RG         64.21
DW         113.000 usec
DE         6.50 usec
TE         303.0 K
D0         0.0000300 sec
D1         1.92176604 sec
D11        0.03000000 sec
D12        0.00002000 sec
D13        0.00000400 sec
D16        0.00020000 sec
IN0        0.00022600 sec

----- CHANNEL f1 -----
SFO1      500.2021545 MHz
NUC1       1H
P0         10.00 usec
P1         10.00 usec
P17        2500.00 usec
PLW1       22.00000000 W
PLW10      3.25440001 W

----- GRADIENT CHANNEL -----
GPNAM[1]  SMSQ10.100
GPE1       10.00 %
P16        1000.00 usec

F1 - Acquisition parameters
TD         128
SFO1       500.2022 MHz
FIDRES     34.568584 Hz
SW         8.846 ppm
F0MODE     QF

F2 - Processing parameters
SI         1024
SF         500.2000602 MHz
WDW        QSINE
SSB        0
LB         0 Hz
GB         0
PC         1.40

F1 - Processing parameters
SI         1024
MC2        QF
SF         500.2000602 MHz
WDW        QSINE
SSB        0
LB         0 Hz
GB         0
    
```

Figure 30S. ¹H-¹H COSY spectrum of 3 (500 MHz, CD₃OD)

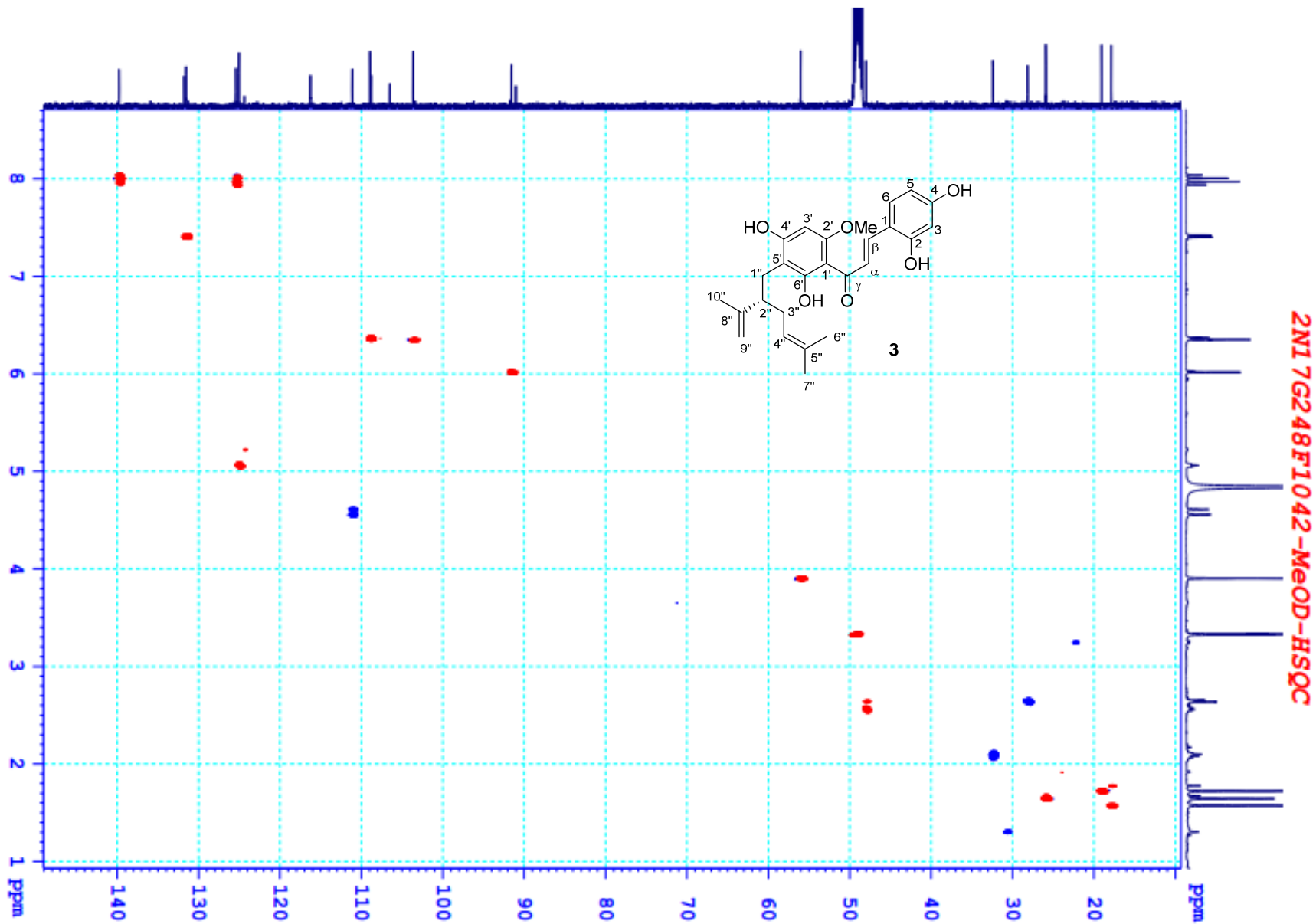


Figure 31S. HSQC spectrum of **3** (^1H : 500 MHz, ^{13}C : 125 MHz, CD_3OD)

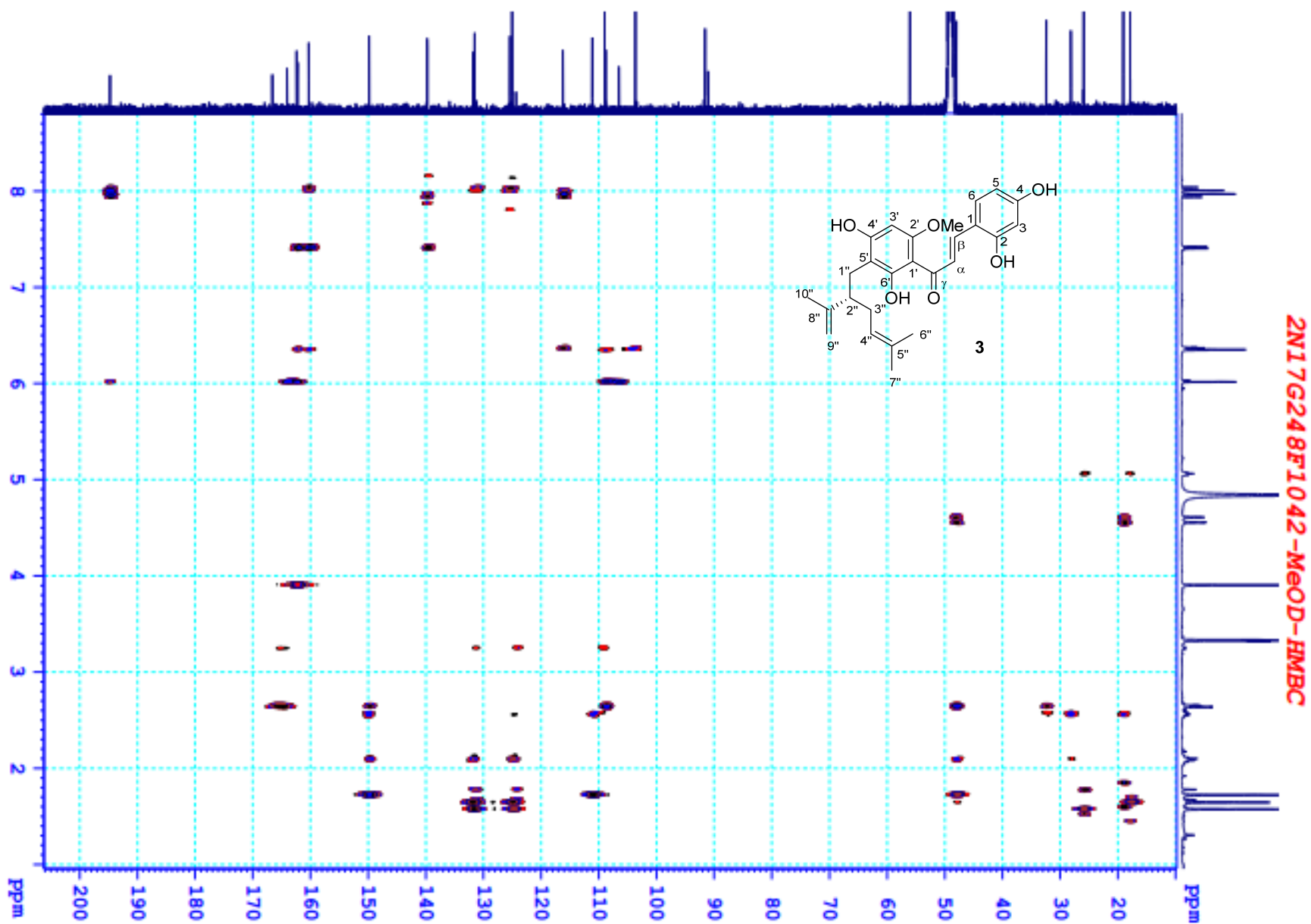
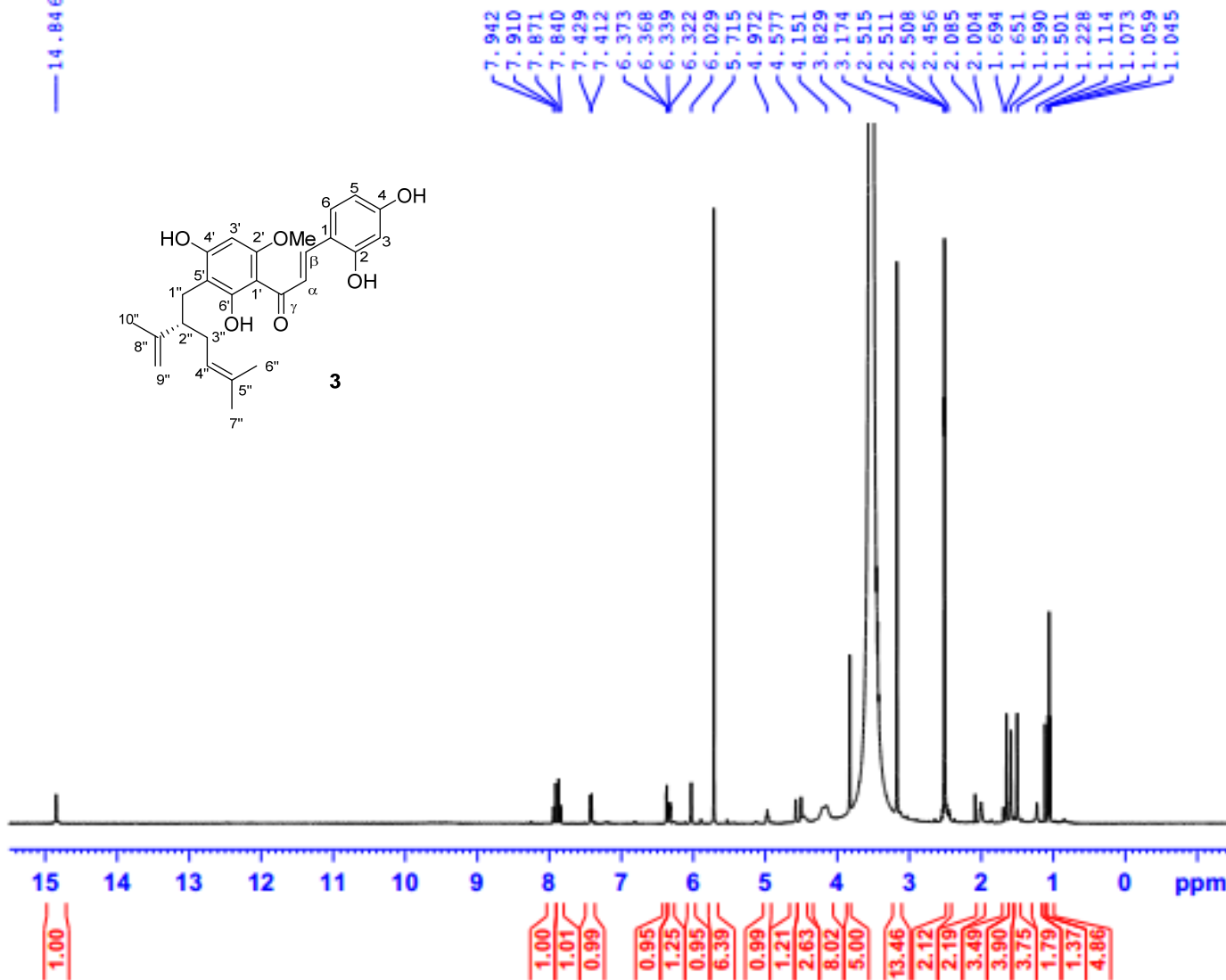
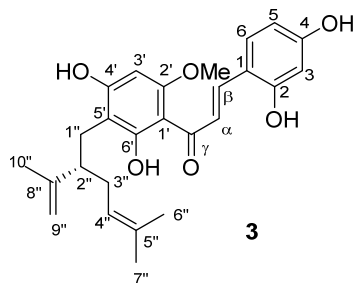


Figure 32S. HMBC spectrum of 3 (^1H : 500 MHz, ^{13}C : 125 MHz, CD_3OD)

2N17G248F10.42.1-DMSO-1H



14.846



Current Data Parameters
 NAME 2VAN_2N17G248F10.42.1
 EXPNO 1
 PROCNO 1

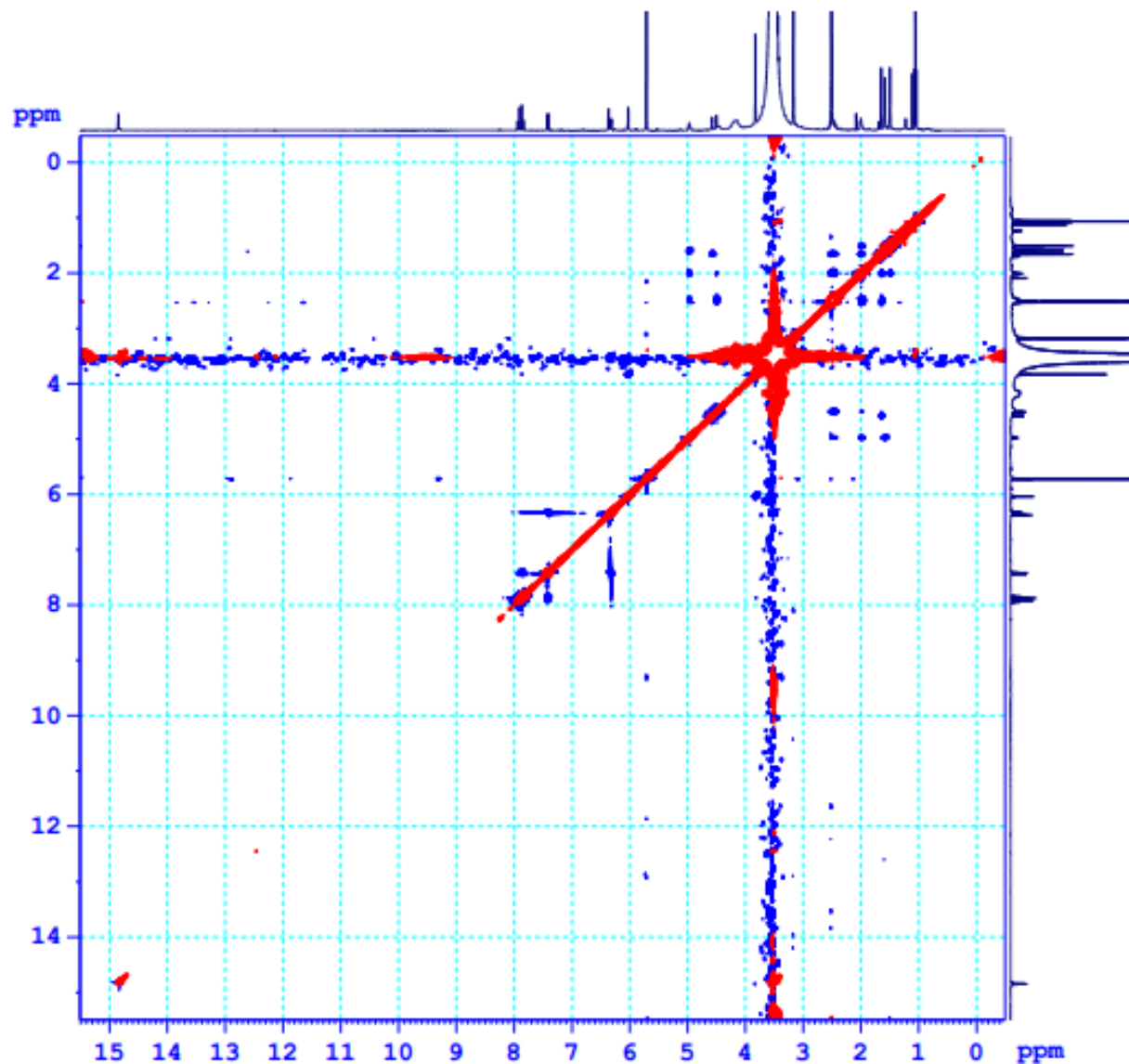
F2 = Acquisition Parameters
 Date_ 20171108
 Time 17.27
 INSTRUM spect
 PROBHD 5 mm PABBO BB/
 PULPROG zg30
 TD 65536
 SOLVENT DMSO
 NS 16
 DS 2
 SMI 10000.000 Hz
 FIDRES 0.152588 Hz
 AQ 3.2767999 sec
 RG 30.85
 DW 50.000 usec
 DE 6.50 usec
 TE 303.0 K
 D1 1.00000000 sec
 TDO 1

----- CHANNEL f1 -----
 SFO1 500.2030889 MHz
 NUC1 1H
 P1 10.00 usec
 PLW1 22.00000000 W

F2 = Processing parameters
 SI 65536
 SF 500.2000000 MHz
 NEM EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00

Figure 33S. ¹H-NMR spectrum of **3** (500 MHz, DMSO-*d*₆)

2N17G248F10.42.1-DMSO-ROESY



Current Data Parameters
 NAME 2VAN_2N17G248F10.42.1
 EXPNO 4
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20171109
 Time 16.17
 INSTRUM spect
 PROBNM 5 mm F4000 BB/
 PULPROG roesyppp.2
 TD 2048
 SOLVENT DMSO
 NS 16
 DS 32
 SMH 8012.820 Hz
 FIDRES 3.912510 Hz
 AQ 0.1277952 sec
 RG 17.93
 CW 62.400 usec
 DE 6.50 usec
 TE 303.9 K
 D0 0.00005213 sec
 D1 1.85663998 sec
 D11 0.03000000 sec
 D12 0.00002000 sec
 IM0 0.00012500 sec
 L4 1000
 P15 200000.00 usec

----- CHANNEL f1 -----
 SFO1 500.2037515 MHz
 NUC1 1H
 P1 10.00 usec
 P17 2500.00 usec
 P25 100.00 usec
 PLW1 22.00000000 W
 PLW10 3.25440001 W
 PLW27 0.88000000 W

F1 - Acquisition parameters
 TD 256
 SFO1 500.2038 MHz
 FIDRES 62.500000 Hz
 SW 15.993 ppm
 FMODE States=TPPI

F2 - Processing parameters
 SI 1024
 SF 500.2000000 MHz
 WDW QSINE
 SSB 2
 LB 0 Hz
 GB 0
 PC 1.00

F1 - Processing parameters
 SI 1024
 MC2 States=TPPI
 SF 500.2000000 MHz
 WDW QSINE
 SSB 2
 LB 0 Hz
 GB 0

Figure 34S. ROESY spectrum of 3 (500 MHz, DMSO-*d*₆)

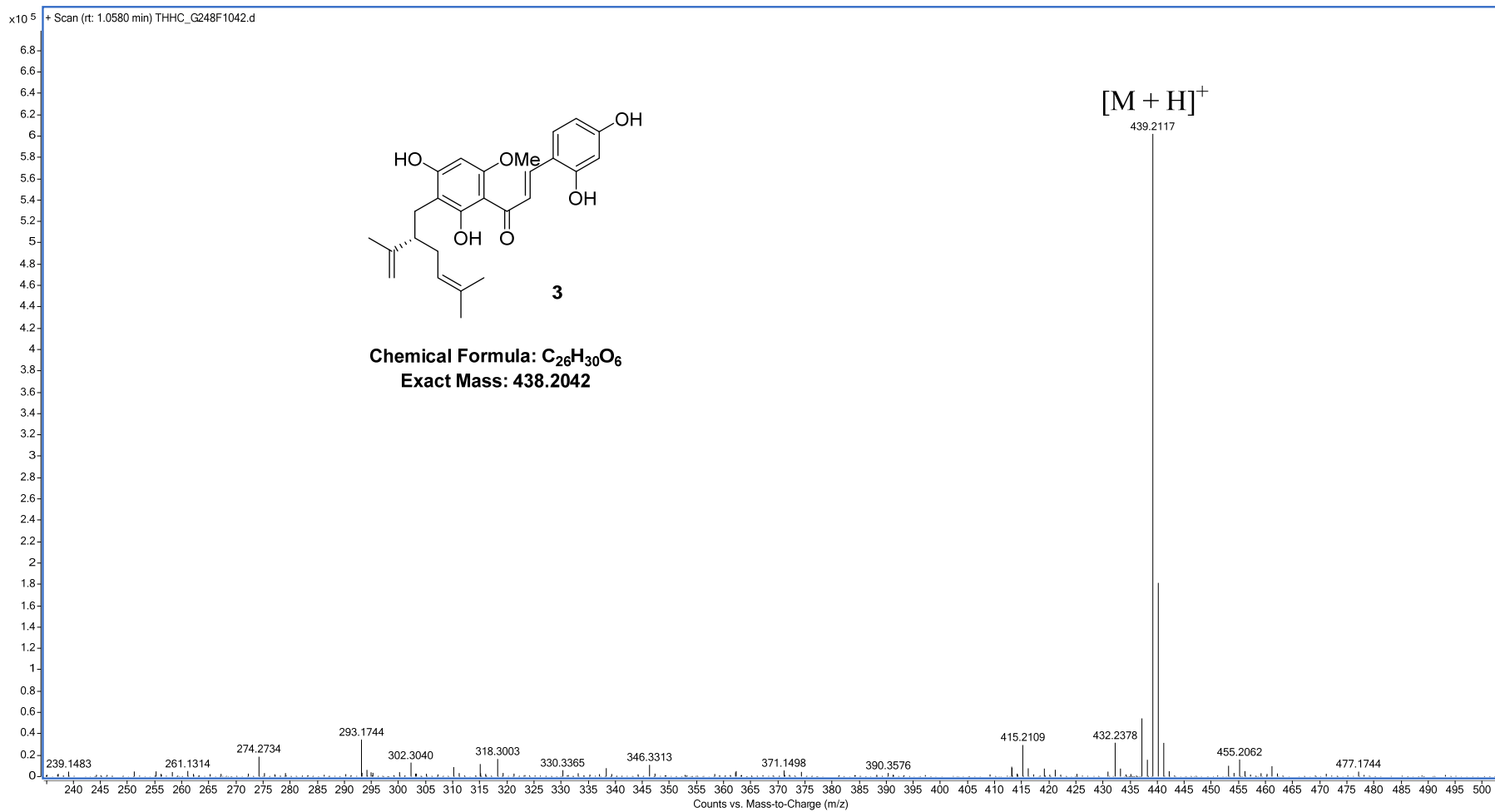


Figure 35S. HR-ESI-MS of **3**

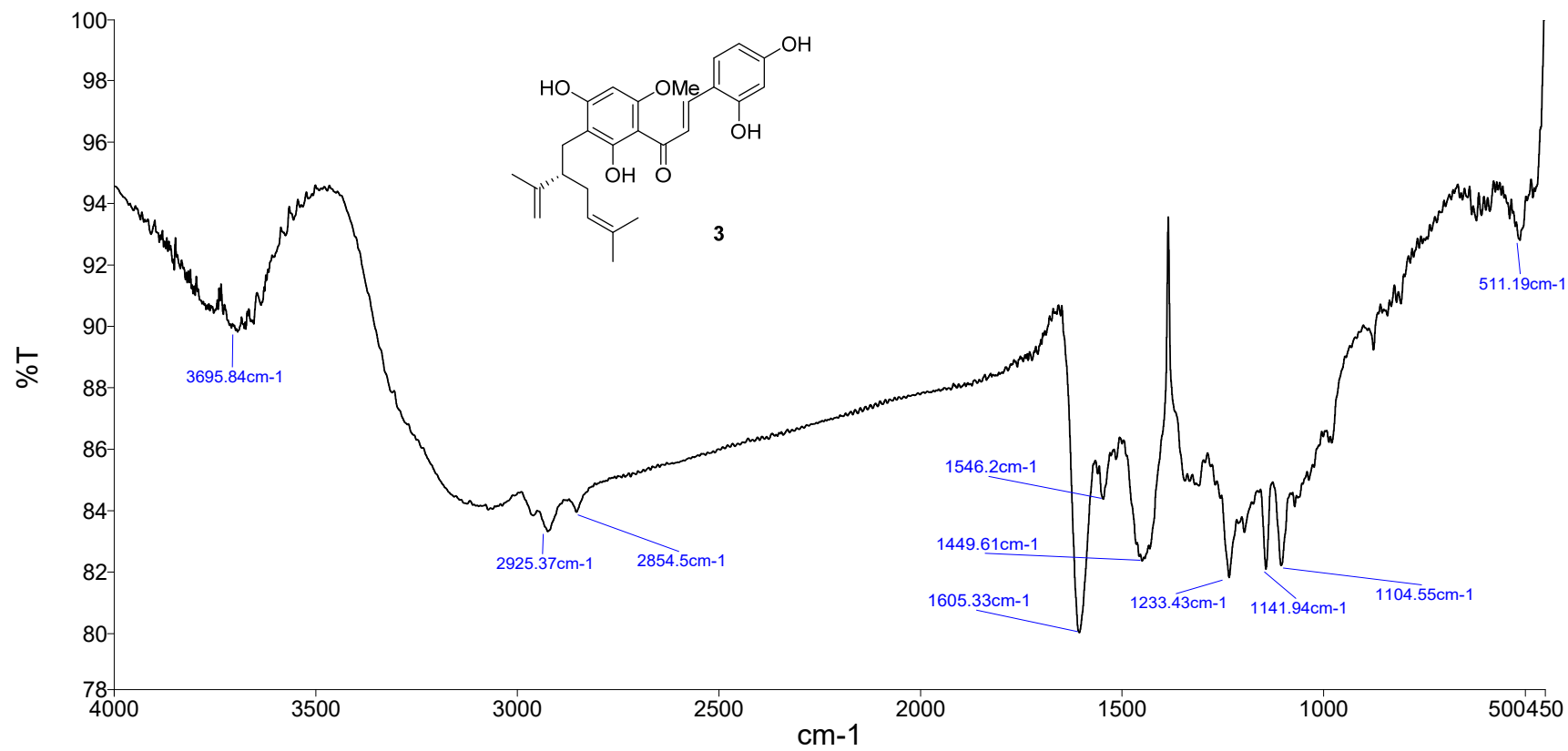


Figure 36S. IR spectrum of **3**

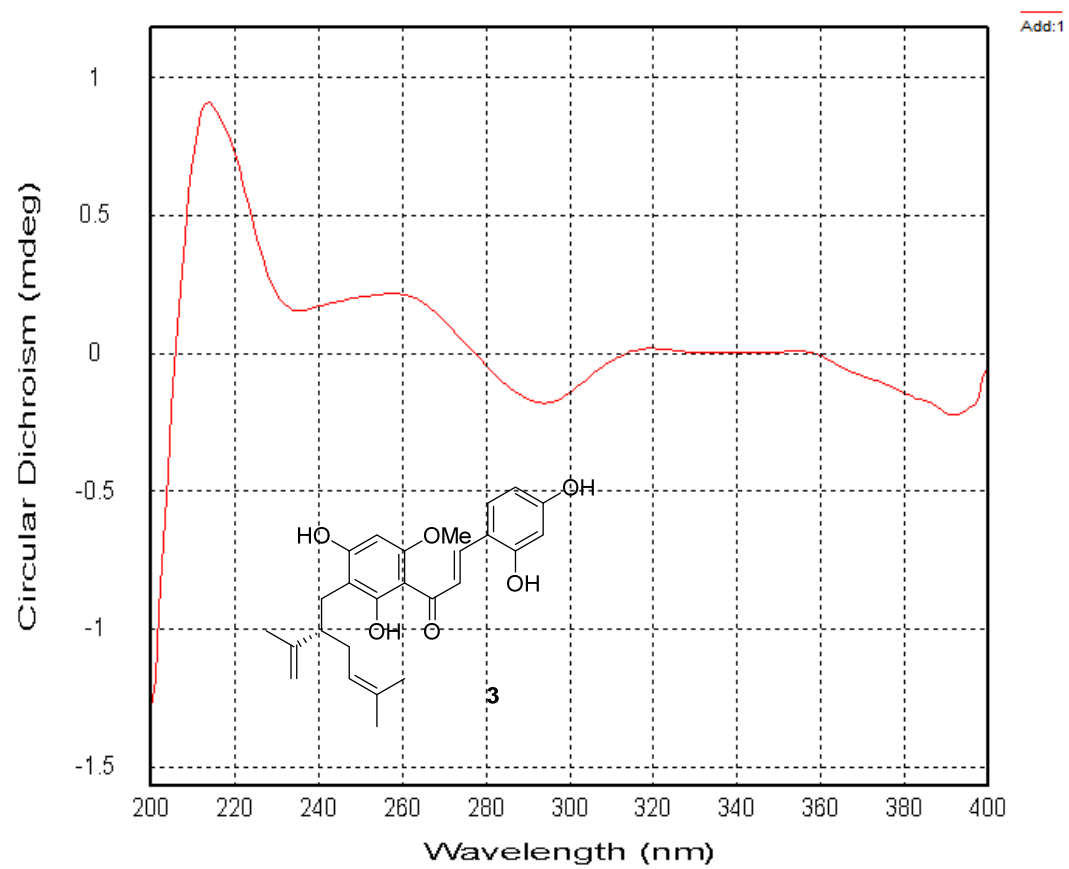


Figure 37S. CD spectrum of **3**