

**Integration of the Exogenous Tuning of Thraustochytrid  
Fermentation and Sulfur Polymerization of Single-Cell Oil for Developing Plant-like Oils**

Adarsha Gupta<sup>1</sup>, Max J. H. Worthington<sup>2,3</sup>, Justin M. Chalker<sup>3</sup> and Munish Puri<sup>1,\*</sup>

<sup>1</sup> Medical Biotechnology, College of Medicine and Public Health, and Flinders Health and Medical Research Institute, Flinders University, Bedford Park, Adelaide, SA 5042, Australia

<sup>2</sup> Key Centre for Polymers and Colloids, School of Chemistry, The University of Sydney, Sydney, NSW 2006, Australia

<sup>3</sup> Institute for Nanoscale Science and Technology, College of Science and Engineering, Flinders University, Bedford Park, Adelaide, SA 5042, Australia

\* Correspondence: [munish.puri@flinders.edu.au](mailto:munish.puri@flinders.edu.au); Tel.: +61-8-72218573

**Supplementary figures captions (Fermentation optimization)**

Figure S1: Effect of different Vitamin B12 concentrations on the FA profile of MASA#4 strain

Figure S2: Effect of different Tween 80 concentrations on the FA profile of MASA#4 strain

Figure S3: Effect of time-dependent addition of D-limonene on the FA profile of MASA#4 strain

Figure S4: Effect of different D-limonene concentrations on the FA profile of MASA#4 strain

Figure S1

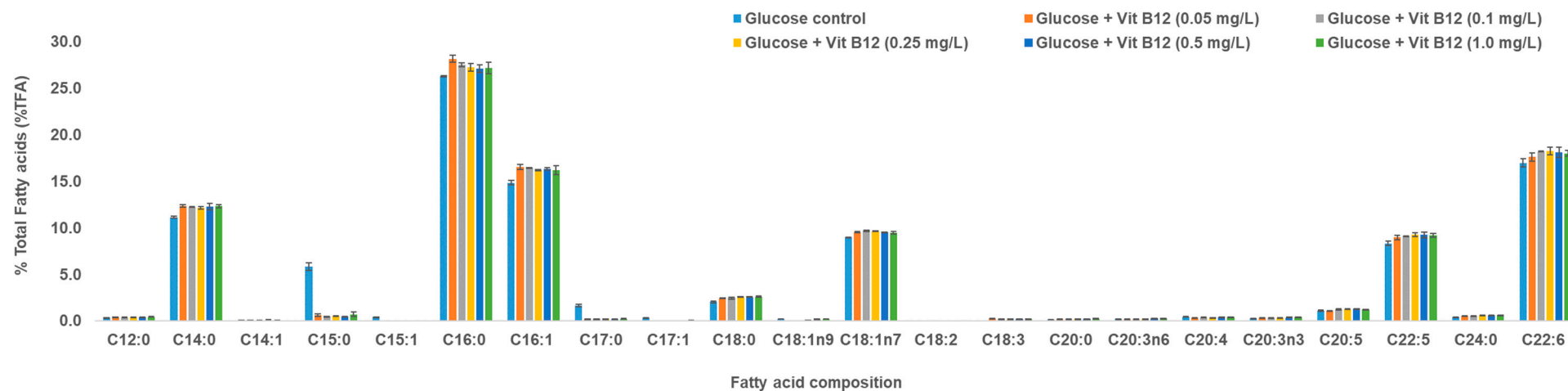


Figure S1: Effect of different Vitamin B12 concentrations on the FA profile of MASA#4 strain

Figure S2

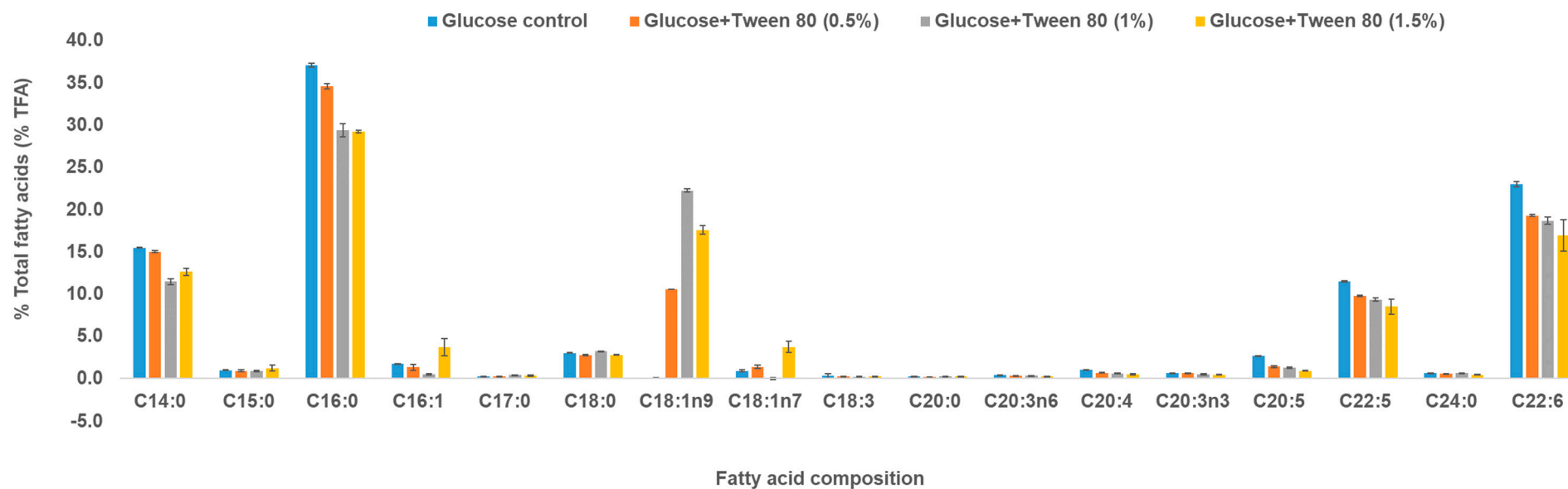


Figure S2: Effect of different Tween 80 concentrations on the FA profile of MASA#4 strain

Figure S3

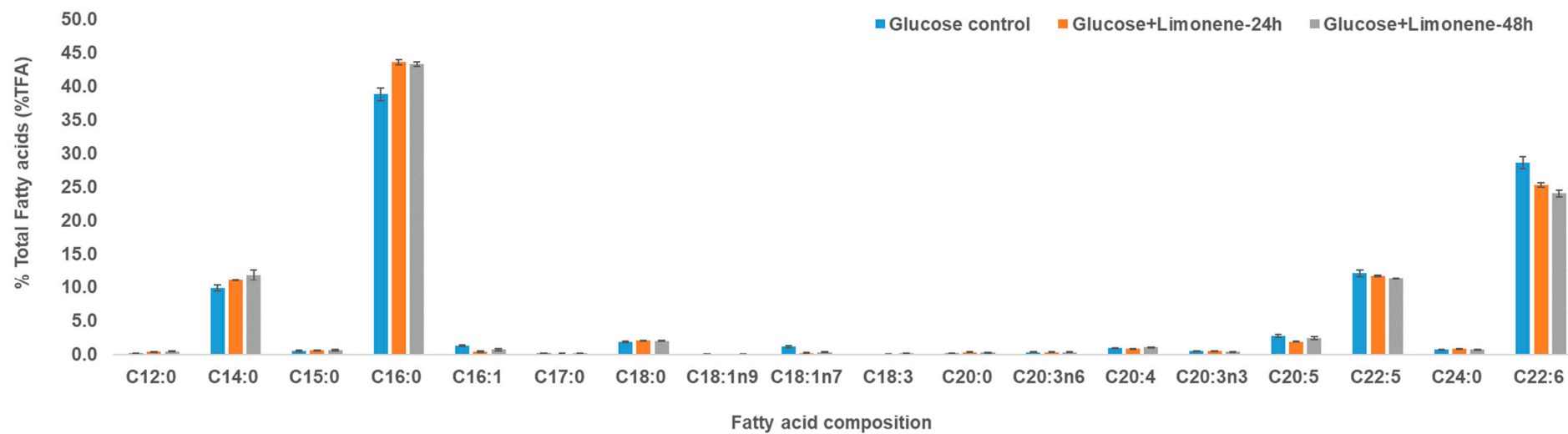
**Figure S3: Effect of time-dependent addition of D-limonene on the FA profile of MASA#4 strain**

Figure S4

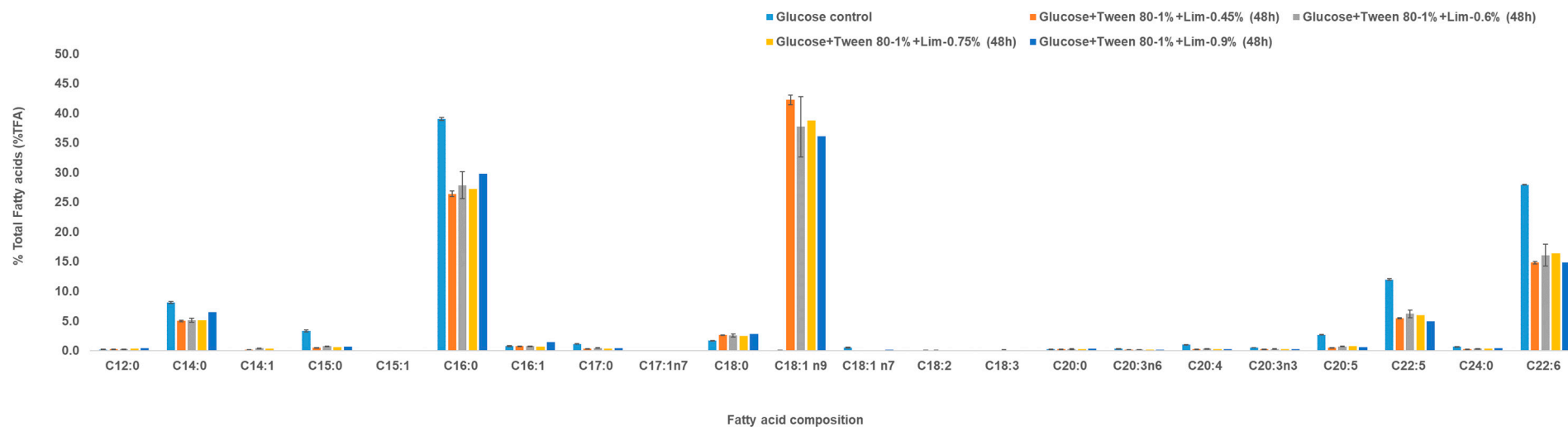


Figure S4: Effect of different D-limonene concentrations on the FA profile of MASA#4 strain

**General experimental considerations (Characterisation of the sulfur polymer)**

**NMR Spectroscopy:** Proton nuclear magnetic resonance ( $^1\text{H}$  NMR) (Bruker) spectra were recorded on a 600 MHz spectrometer or 400 MHz spectrometer where noted. All chemical shifts are quoted on the  $\delta$  scale in ppm using residual solvent as the internal standard ( $^1\text{H}$  NMR:  $\text{CDCl}_3$   $\delta$  = 7.26, methanol- $\text{d}_4$   $\delta$  = 4.87 and pyridine- $\text{d}_5$   $\delta$  = 8.74).

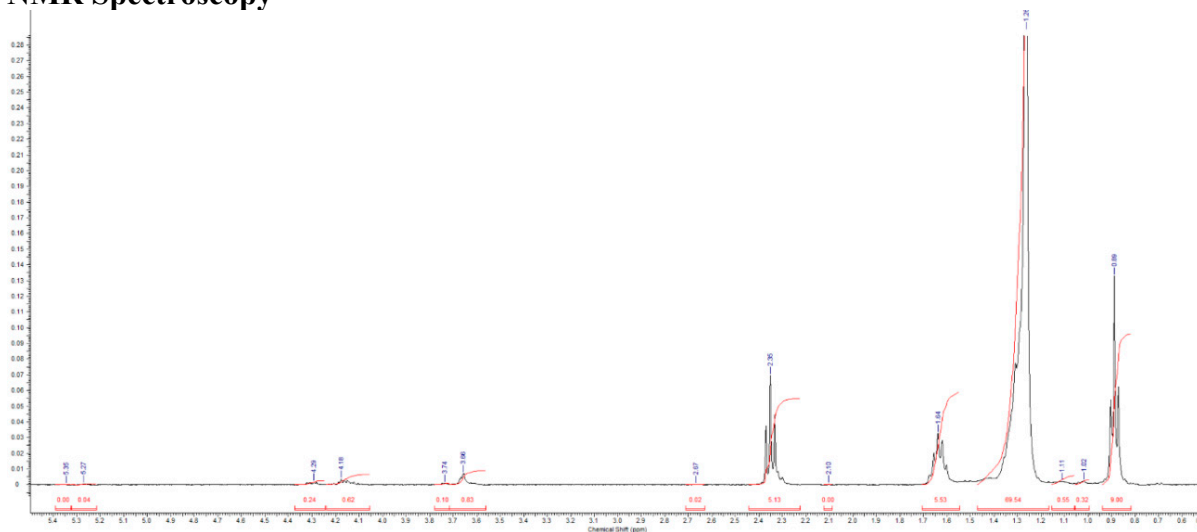
**SEM and EDS:** Scanning Electron Microscopy (SEM) images were obtained using an FEI F50 Inspect system, while corresponding EDS spectra were obtained using an EDAX Octane Pro detector.

**Thermogravimetric Analysis (TGA):** Simultaneous Thermal Analysis (STA) was carried out on a Perkin Elmer STA8000 simultaneous thermal analyzer. A sample size between 10 and 15 mg was used in each run. The furnace was purged at 20 mL/min with nitrogen, and equilibrated for 1 minute at 30 °C before each run. Heating was carried out up to 700 °C using a 20 °C/min heating rate. The temperature was held isothermally at 700 °C at the end of each experiment to oxidize remaining organic matter.

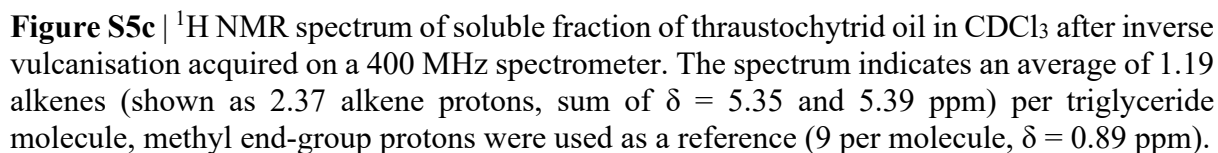
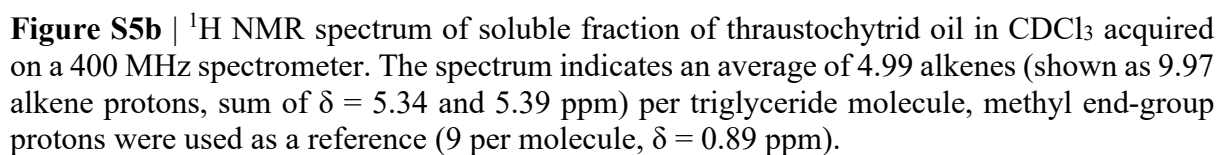
**Differential Scanning Calorimetry (DSC):** In order to determine glass transition temperatures, thermal analysis was performed on a Perkin Elmer DSC8000. A sample size between 5 and 10 mg was used in each run. The furnace was purged at 20 mL/min with nitrogen, and equilibrated for 1 minute at 30 °C before each run. Heating was carried out up to 140 °C using a 10 °C/min heating rate, followed by a cooling step down to -60 °C at the same rate. Heating and cooling from -60 °C to 140 °C and back was repeated twice more to monitor any changes in the DSC profile over multiple cycles.

**X-ray Diffraction:** Powder X-ray diffraction (XRD) patterns were recorded on a Bruker D8 Advance Eco diffractometer (Bragg-Brentano geometry) using  $\text{Co-K}\alpha$  radiation ( $\lambda$  = 1.78897 Å). The Bragg angle ( $2\theta$ ) was varied from 15° to 90° with a step size of 0.019°, measurement time of 0.45 s per step and sample rotation at 15 rpm. The XRD patterns were collected on a silicon low background sample holder, where powder samples were deposited onto the surface of the holder and spread evenly using a drop of acetone.

### Characterisation of Thraustochytrid Oil after Hydrogenation and Sulfur Polymerization

**NMR Spectroscopy**

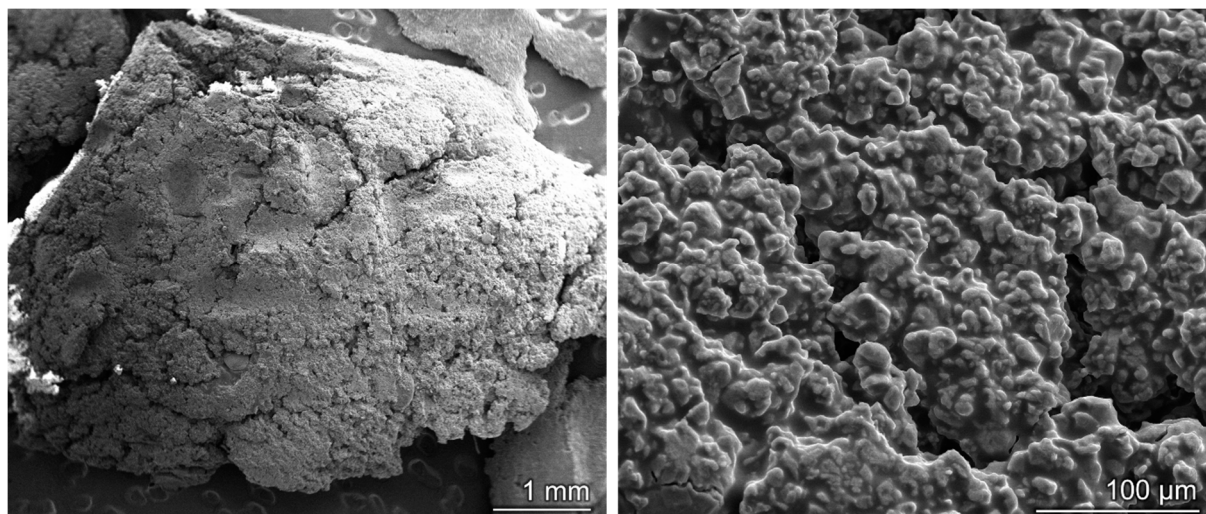
**Figure S5a** |  $^1\text{H}$  NMR spectrum of thraustochytrid oil in methanol- $\text{d}_4$  after hydrogenation acquired on a 400 MHz spectrometer. The spectrum indicates total conversion with an average of zero alkenes remaining (shown as 0.00 alkene protons,  $\delta$  = 5.35) per triglyceride molecule. Methyl end-group protons were used as a reference (9 per molecule,  $\delta$  = 0.92 ppm).



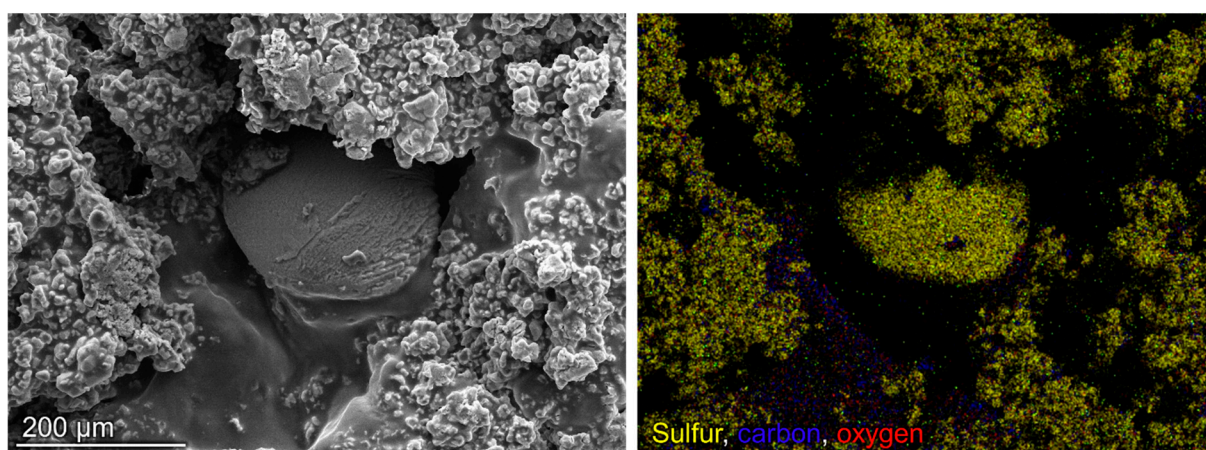


## Scanning Electron Microscopy and Electron Dispersive X-Ray Spectroscopy (SEM and EDS)

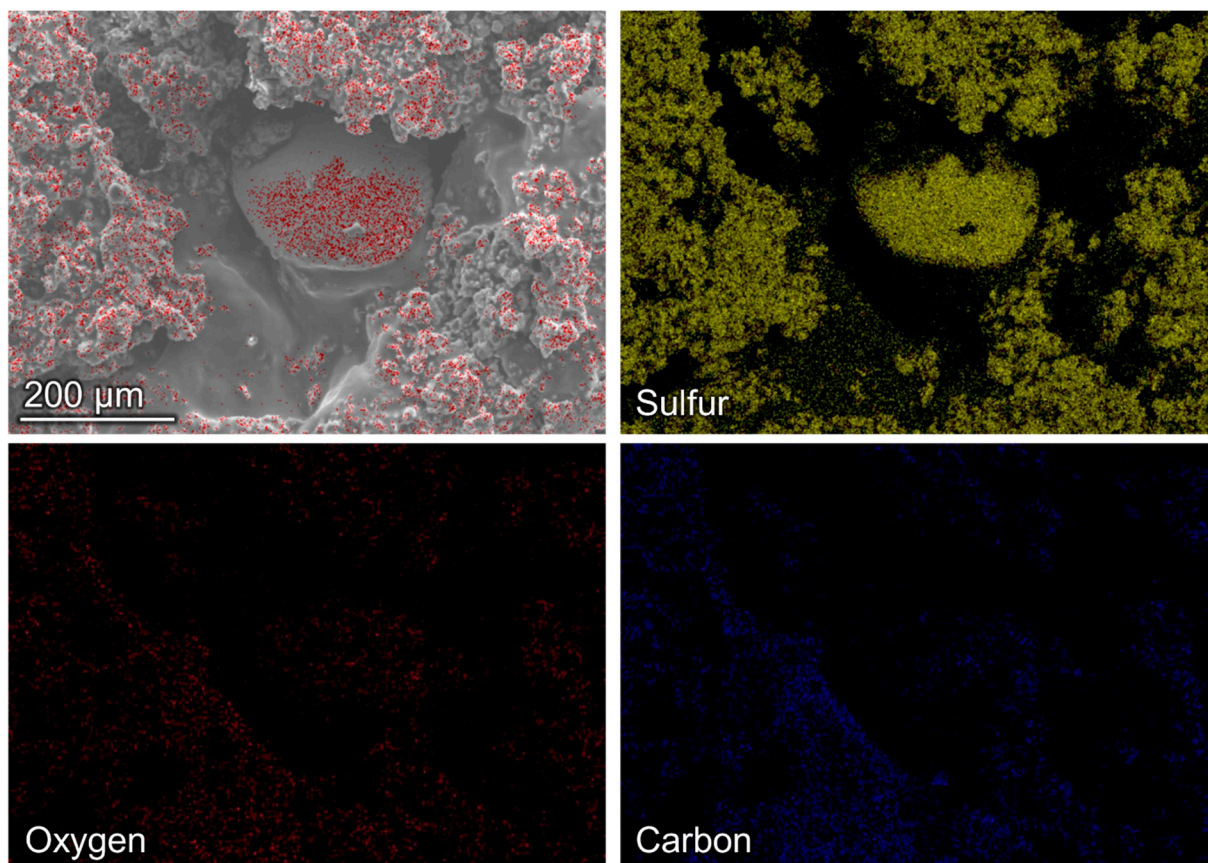
Thraustochytrid oil Polymer



**Figure S6a** | SEM micrograph and corresponding EDX elemental map of thraustochytrid oil polymer. The map has been brightness-adjusted for visual clarity.

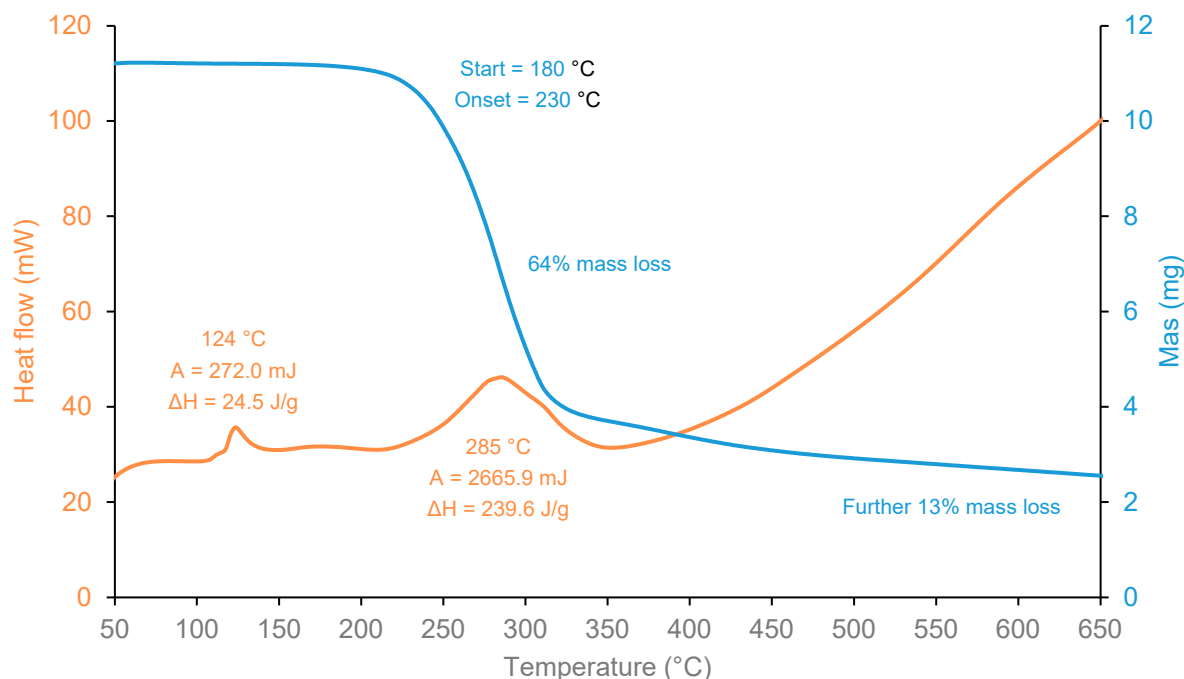


**Figure S6b** | SEM micrograph and corresponding EDX elemental map of thraustochytrid oil polymer. The map has been brightness-adjusted for visual clarity.



**Figure S6c** | Elemental map of thraustochytrid oil polymer by EDX. Top left maps where the signal was concentrated on the sample. Elemental images have had contrast and brightness adjusted for visual clarity.

## Simultaneous Thermal Analysis (STA)

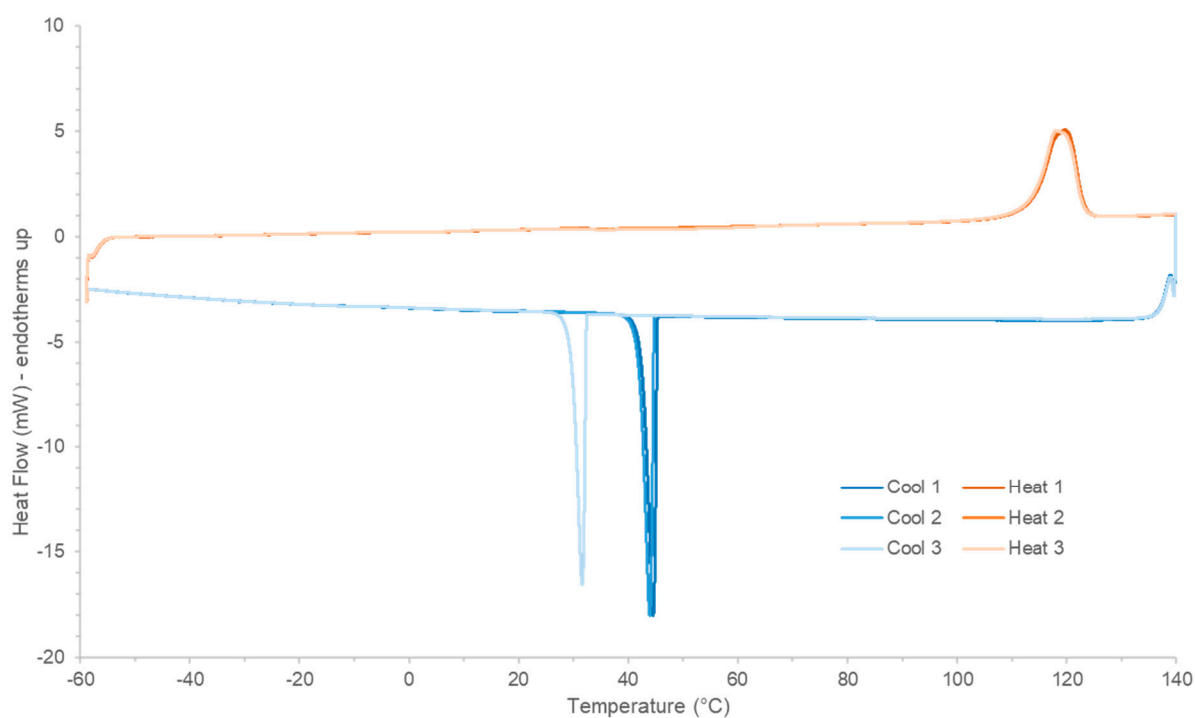


**Figure S7a** | STA analysis the polymer product of inverse vulcanisation of thraustochytrid oil. DSC trace in orange, TGA in blue. The exotherm at 124°C coincides with the melting point of sulfur and indicates the presence of unreacted S<sub>8</sub> within the polymer. Based on a calibration curve of sulfur melting events generated on the same instrument under the same conditions and published in *Laying Waste to Mercury: Inexpensive Sorbents Made from Sulfur and Recycled Cooking Oils, Supporting Information*, this exotherm indicates 51.1% of the mass of the polymer is free sulfur.

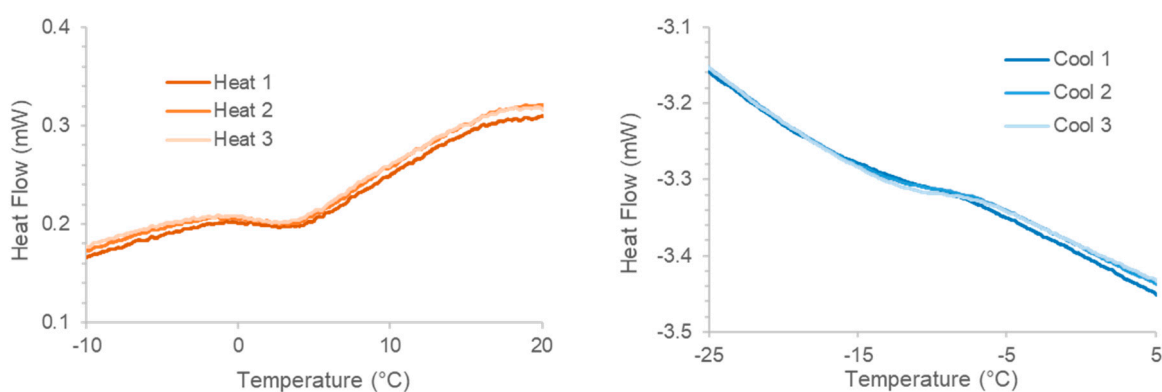
Unreacted sulfur constitutes a roughly half of the mass of the resultant polymer.

## Dynamic Scanning Calorimetry for Glass Transition Temperature

Thraustochytrid oil Polymer



**Figure S7b** | DSC trace of thraustochytrid oil polymer



**Figure S7c** | DSC trace of thraustochytrid oil polymer, focused only on the baseline shift in the cooling and heating phases characteristic of a glass transition.

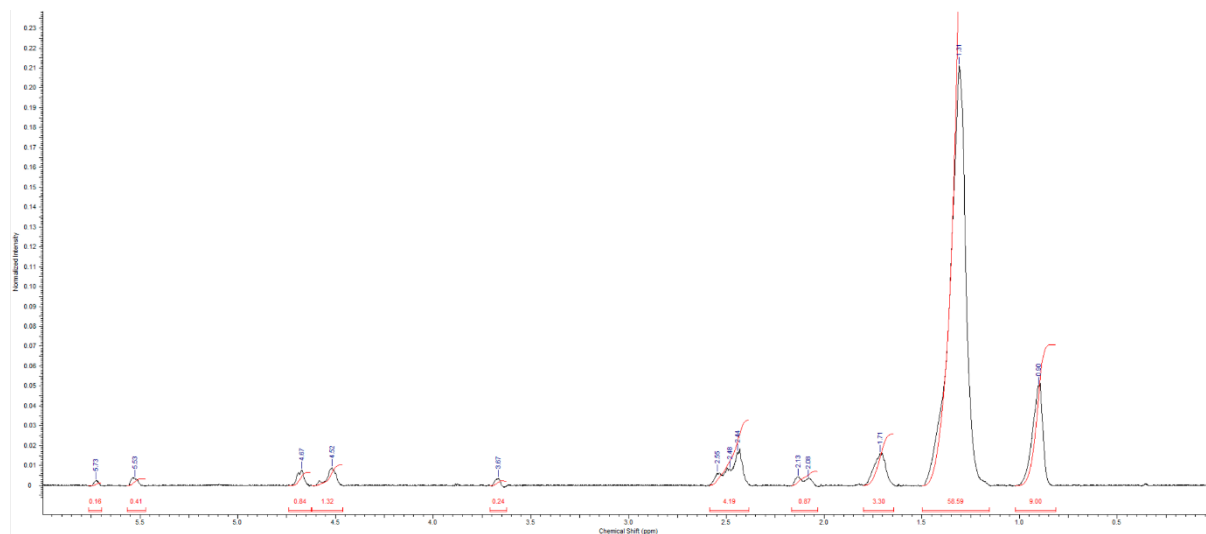
Heating Tg average = 1.1 °C (0.84, 0.98, 1.44)

Cooling Tg average = 11.4 °C (-12.41, -11.67, -10.26)

Combined average = **6.3 °C**

## NMR Spectroscopy

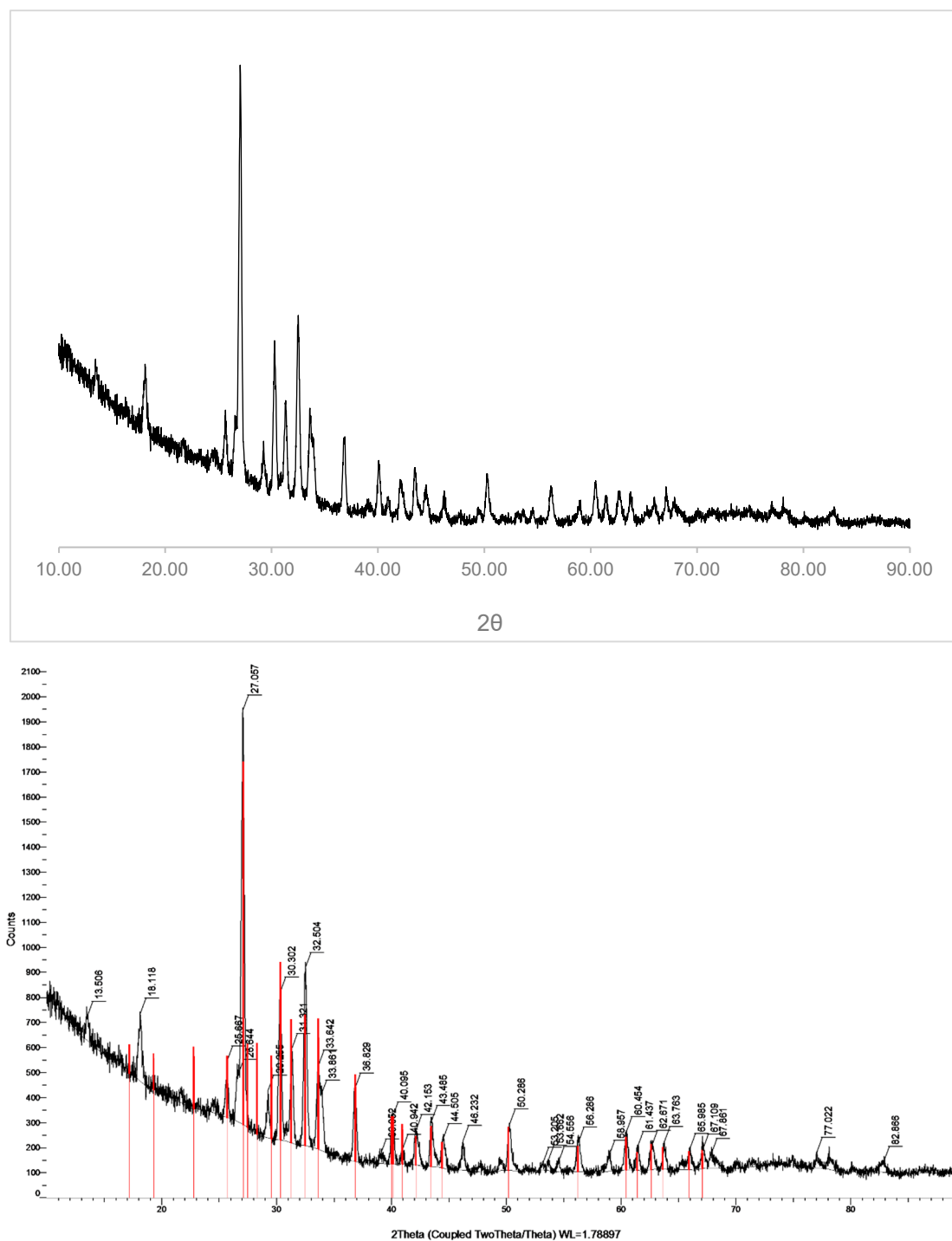
### Thraustochytrid oil Polymer



**Figure S7d** |  $^1\text{H}$  NMR spectrum of thraustochytrid oil polymer in pyridine- $\text{d}_5$ . The spectrum indicates an average of 0.08 alkenes (shown as 0.16 alkene protons,  $\delta = 5.73$ ) per triglyceride molecule. Methyl end-group protons were used as a reference (9 per molecule,  $\delta = 0.90$  ppm).

## X-ray Diffraction

### Thraustochytrid oil Polymer



**Figure S8** | Above: XRD spectrum of thraustochytrid oil polymer Below: sulfur ( $S_8$ ) reference spectra overlaid in red.