

Supporting Information

New Zosteropenillines and Pallidopenillines from the Seagrass-Derived Fungus *Penicillium yezoense* KMM 4679

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DNA Extraction and Amplification

Genomic DNA was isolated from fungal mycelium grown on MEA (malt extract agar) at 25 °C for 7 days using the MagJET Plant Genomic DNA Kit (Thermo Fisher Scientific, Waltham, MA, USA) according to the manufacturer's protocol. PCR was conducted using GoTaq Flexi DNA Polymerase (Promega, Madison, WI, USA). For amplification of the *ITS* region, the standard primer pair, ITS1 and ITS4 was used [1]. The reaction profile was 95 °C for 300 s; 35 cycles of 95 °C for 30 s, 55 °C for 45 s, 72 °C for 90 s; and, finally, 72 °C for 300 s. For amplification of the partial *BenA* gene, the primer pair, tub_P/A_F [2] and Bt-2b [3] was used. The reaction profile was 95 °C for 300 s; 35 cycles of 95 °C for 30 s, 55 °C for 45 s, and 72 °C for 90 s; and, finally, 72 °C for 300 s. For amplification of the partial *CaM* gene, the degenerate primer pair cal_P/A_F and cal_P/A_R was used [2]. The reaction profile was 95 °C for 300 s; 35 cycles of 95 °C for 30 s, 60 °C for 45 s, and 72 °C for 90 s; and, finally, 72 °C for 300 s. For amplification of the partial *RPB2* gene, the standard primer pair, 5Feur and 7CReur was used [4]. The reaction profile used as described in [5]. The amplified *ITS*, *BenA*, *CaM*, and *RPB2* genes were purified with the ExoSAP-IT™ PCR Product Cleanup Reagent (Thermo Fisher Scientific, Waltham, MA, USA). Sequencing was bidirectional performed with the same primers on an Applied Biosystems SeqStudio Genetic Analyzer (Thermo Fisher Scientific, Waltham, MA, USA) using the Big Dye Terminator reagent kit, version 3.1. Gene sequences were deposited in GenBank under accession numbers PP854452 for *ITS*, PP861099 for the partial *BenA*, PP861100 for the partial *CaM*, and PP861101 for the partial *RPB2* gene (Table S1).

1. White, T.J.; Bruns, T.; Lee, S.; Taylor, J. Amplification and direct sequencing of fungal ribosomal RNA genes for phylogenetics. In PCR protocols: a guide to methods and applications, London, 1990; Vol. 38, pp 315-322.
2. Yurchenko, A.N.; Zhuravleva, O.I.; Khmel, O.O.; Oleynikova, G.K.; Antonov, A.S.; Kirichuk, N.N.; Chausova, V.E.; Kalinovsky, A.I.; Berdyshev, D.V.; Kim, N.Y., et al. New cyclopiane diterpenes and polyketide derivatives from marine sediment-derived fungus *Penicillium antarcticum* KMM 4670 and their biological activities. *Mar. Drugs* 2023, 21, 21110584
3. Glass, N.L.; Donaldson, G.C. Development of primer sets designed for use with the PCR to amplify conserved genes from filamentous ascomycetes. *Applied Environmental Microbiology* 1995, 61, 1323-1330. - <https://www.scopus.com/inward/record.uri?eid=2-s2.0-0028969372&doi=10.1128%2faem.61.4.1323-1330.1995&partnerID=40&md5=ee11611f6b61ec23e654da22ba152a0f>
4. J, H.; H, S.; JC, F. *Rasamsonia*, a new genus comprising thermotolerant and thermophilic *Talaromyces* and *Geosmithia* species. *Antonie Van Leeuwenhoek* 2012, 101, 403-421.
5. CM, V.; J, H.; JC, F.; SB, H.; CH, K.; G, P.; KA, S.; J, V.; T, Y.; RA, S. Identification and nomenclature of the genus *Penicillium*. *Stud. Mycol.* 2014, 78, 343-371.

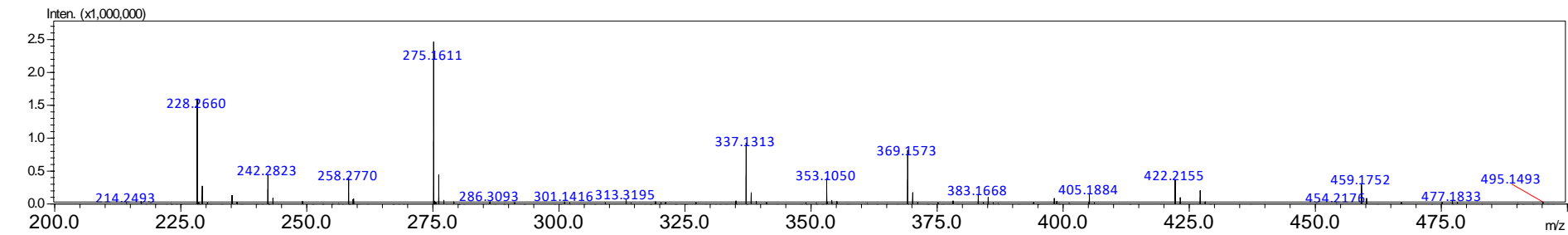
Table S1. The strains of the species used in multi-locus phylogenetic analysis and GenBank accession numbers.

Species	Strain Number	GenBank Accession Number			
		<i>ITS</i>	<i>BenA</i>	<i>CaM</i>	<i>RPB2</i>
<i>Penicillium aurantioviolaceum</i>	CBS 137777 ^T	KM189756	KM089005	KM089392	KM089779

<i>Penicillium austroafricanum</i>	CBS 137773 ^T	KM189610	KM088854	KM089241	KM089628
<i>Penicillium cartierense</i>	CBS 137956 ^T	KM189564	KM088804	KM089189	KM089576
<i>Penicillium contaminatum</i>	CBS 345.52 ^T	KM189554	KM088793	KM089178	KM089565
<i>Penicillium crocicola</i>	CBS 745.70 ^T	KM189581	KJ834445	KM089210	JN406535
<i>Penicillium fusisporum</i>	CBS 137463 ^T	KF769424	KF769400	KF769413	MN969117
<i>Penicillium grevilleicola</i>	CBS 137775 ^T	KM189630	KM088874	KM089261	KM089648
<i>Penicillium jejuense</i>	CBS 138646 ^T	KF818464	KF818461	KF818470	KF818467
<i>Penicillium roseoviride</i>	CBS 267.35 ^T	KM189549	KM088787	KM089172	KM089559
<i>Penicillium thomii</i>	CBS 225.81 ^T	KM189560	KM088799	KM089184	KM089571
<i>Penicillium valentinum</i>	CBS 172.81 ^T	KM189550	KM088788	KM089173	KM089560
<i>Penicillium yezoense</i>	CBS 350.59 ^T	KM189553	KM088792	KM089177	KM089564
<i>Penicillium yezoense</i>	KMM 4679	PP854452	PP861099	PP861100	PP861101
<i>Talaromyces marneffei</i>	CBS 388.87 ^T	JN899344	JX091389	KF741958	KM023283

1. Houbraken J; Kocsubé S; Visagie CM; Yilmaz N; Wang XC; Meijer M; Kraak B; Hubka V; Bensch K; Samson RA, *et al.* Classification of *Aspergillus*, *Penicillium*, *Talaromyces* and related genera (Eurotiales): An overview of families, genera, subgenera, sections, series and species. *Stud. Mycol.* **2020**, 95, 5-169.

Figure S1 HRESIMS for 1



	meas	calc	Δ (ppm)
[M+Na] ⁺	275,1611	275,1617	-2,4

Figure S2. ^1H NMR spectrum of **1** measured at 700 MHz in CDCl_3

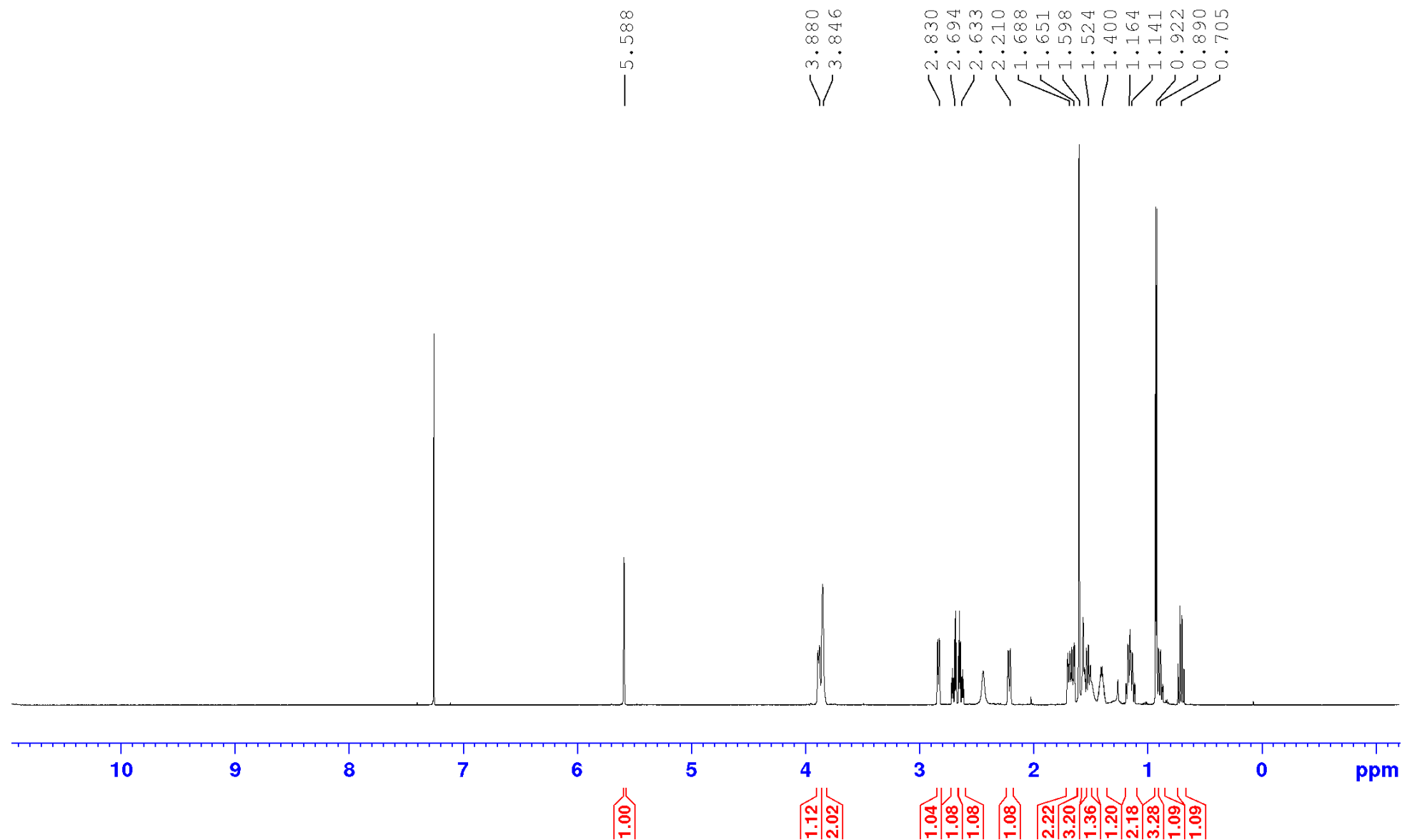


Figure S3. ^{13}C NMR spectrum of **1** measured at 176 MHz in CDCl_3

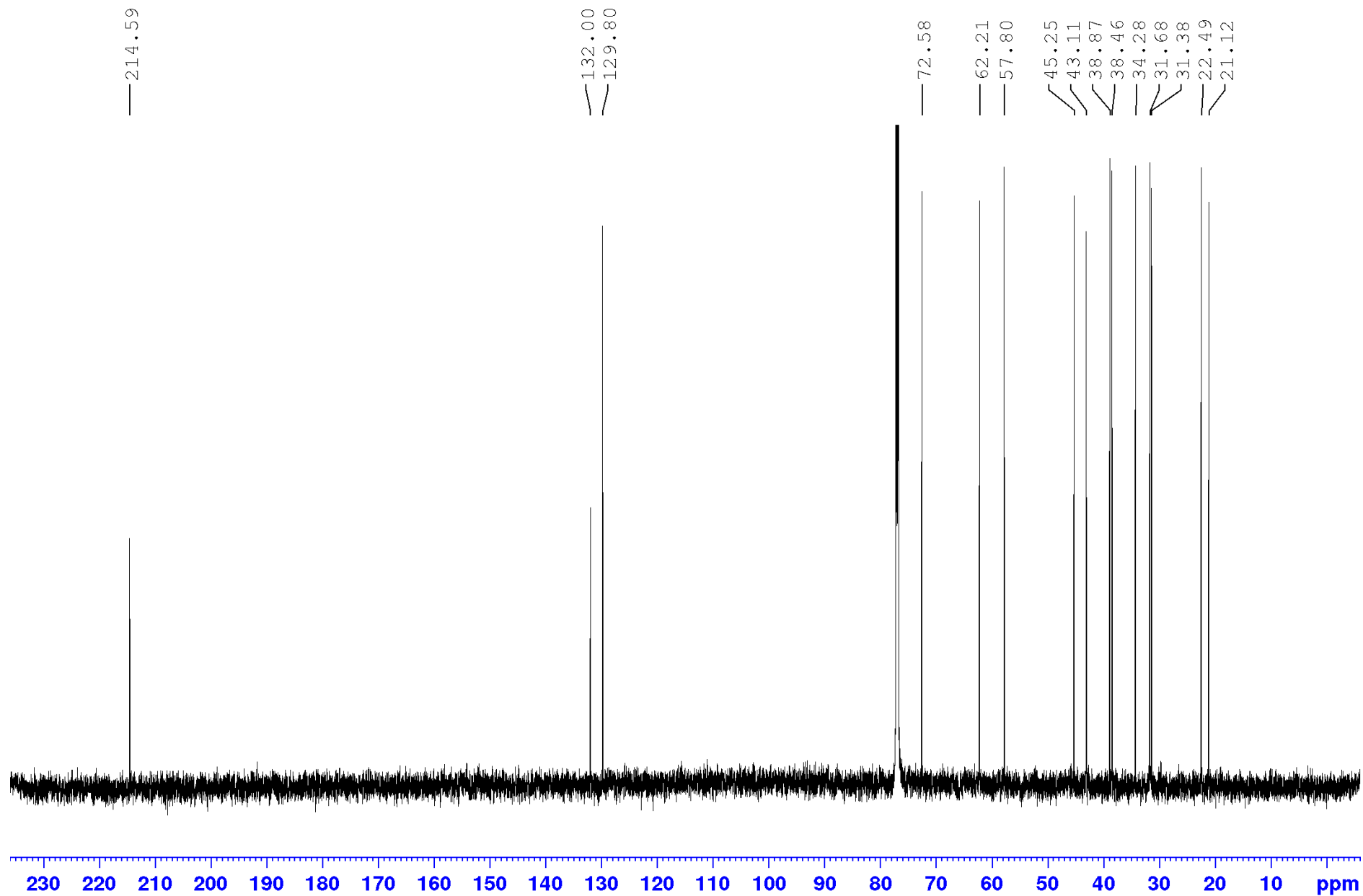


Figure S4. DEPT-135 spectrum of 1 measured at 176 MHz in CDCl₃

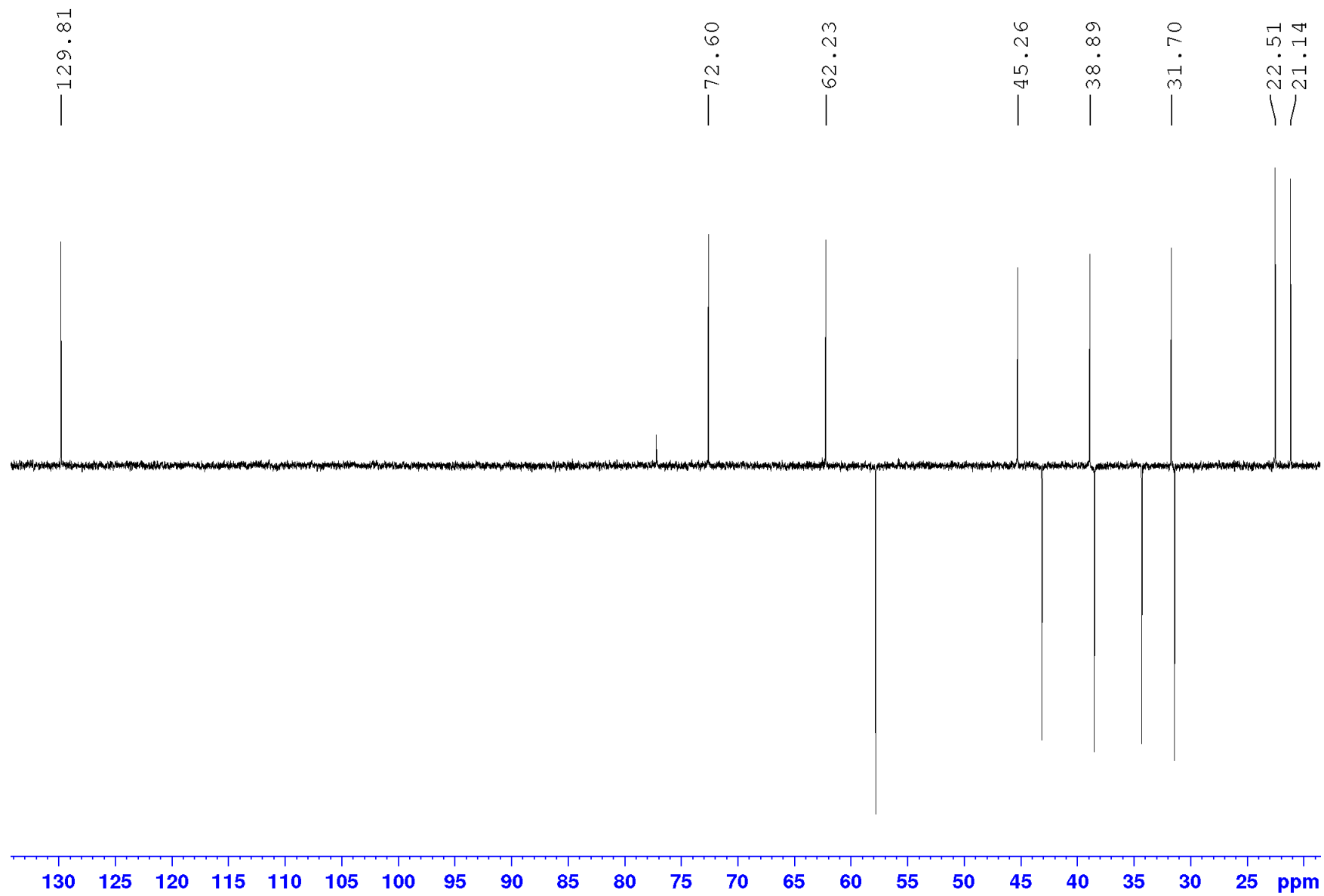


Figure S5. HSQC spectrum of 1 measured in CDCl₃

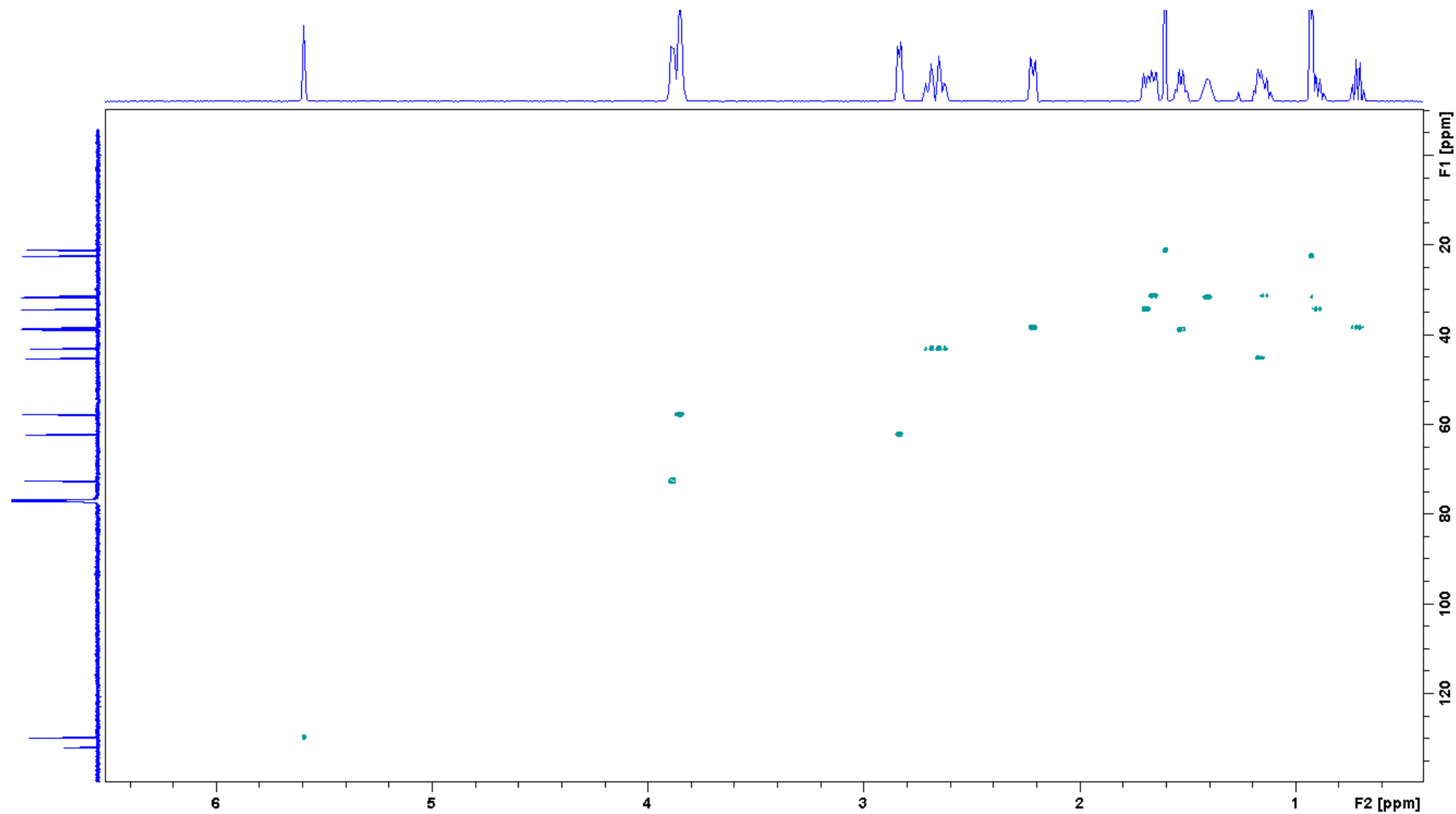


Figure S6. COSY spectrum of 1 measured in CDCl₃

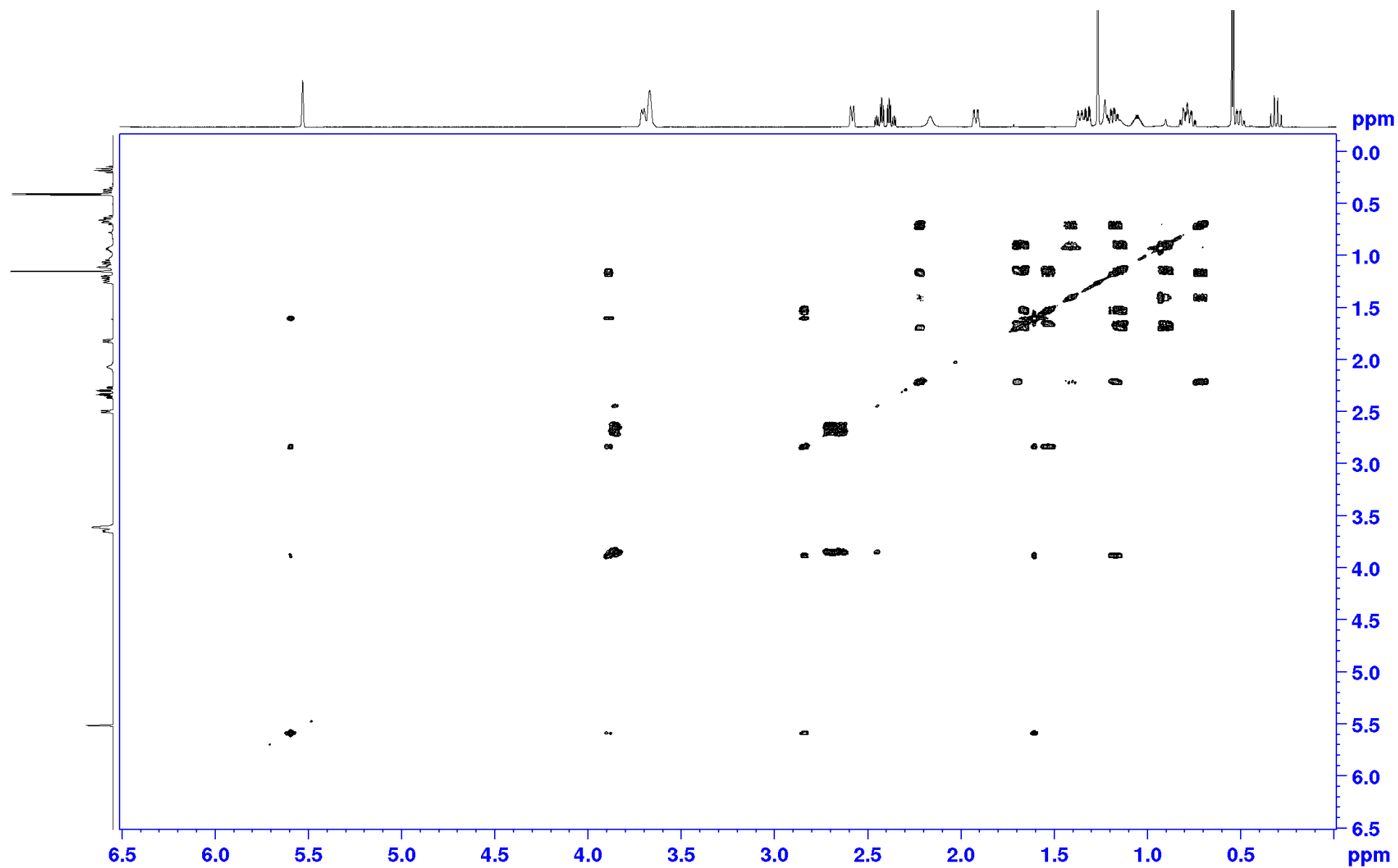


Figure S7. HMBC spectrum of 1 measured in CDCl₃

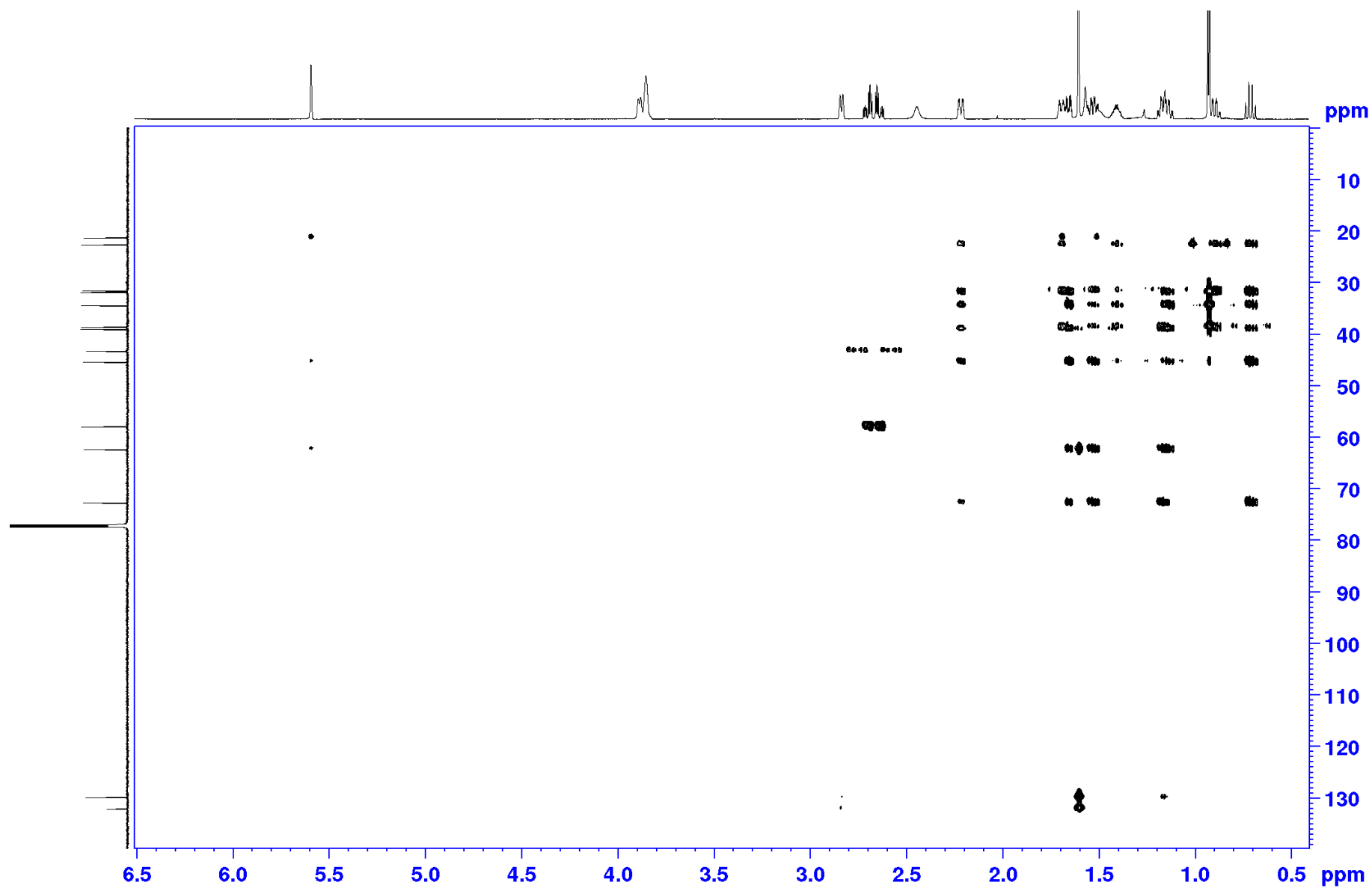


Figure S8. ROESY spectrum of 1 measured in CDCl₃

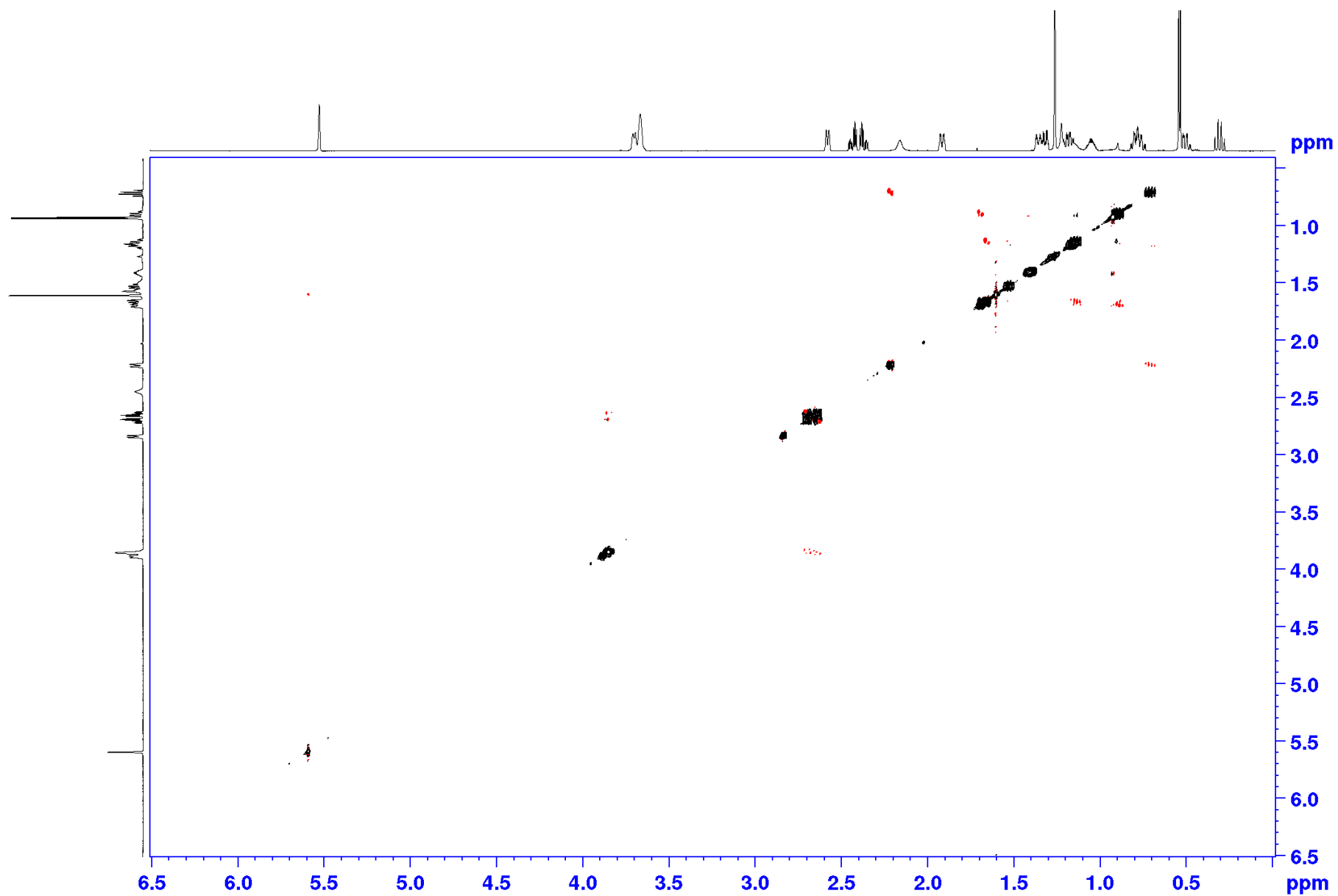
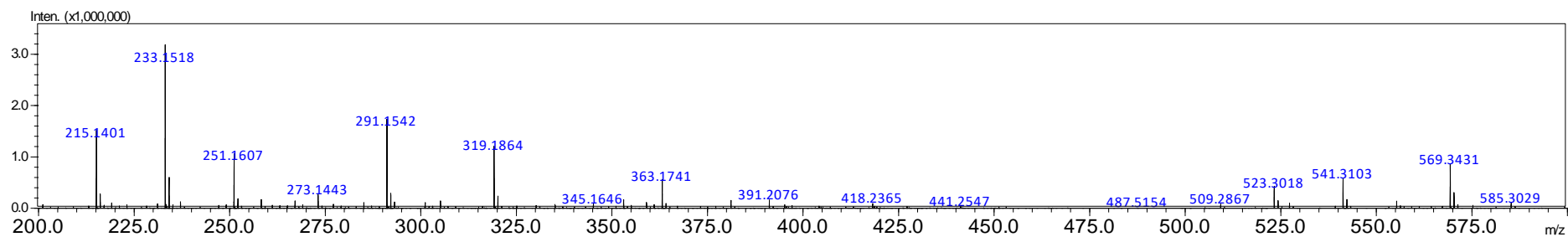


Figure S9. HRESIMS for 2



	meas	calc	Δ (ppm)
$[M+Na]^+$	291,1542	291,1566	-8,5

Figure S10. ^1H NMR spectrum of **2** measured at 700 MHz in CDCl_3

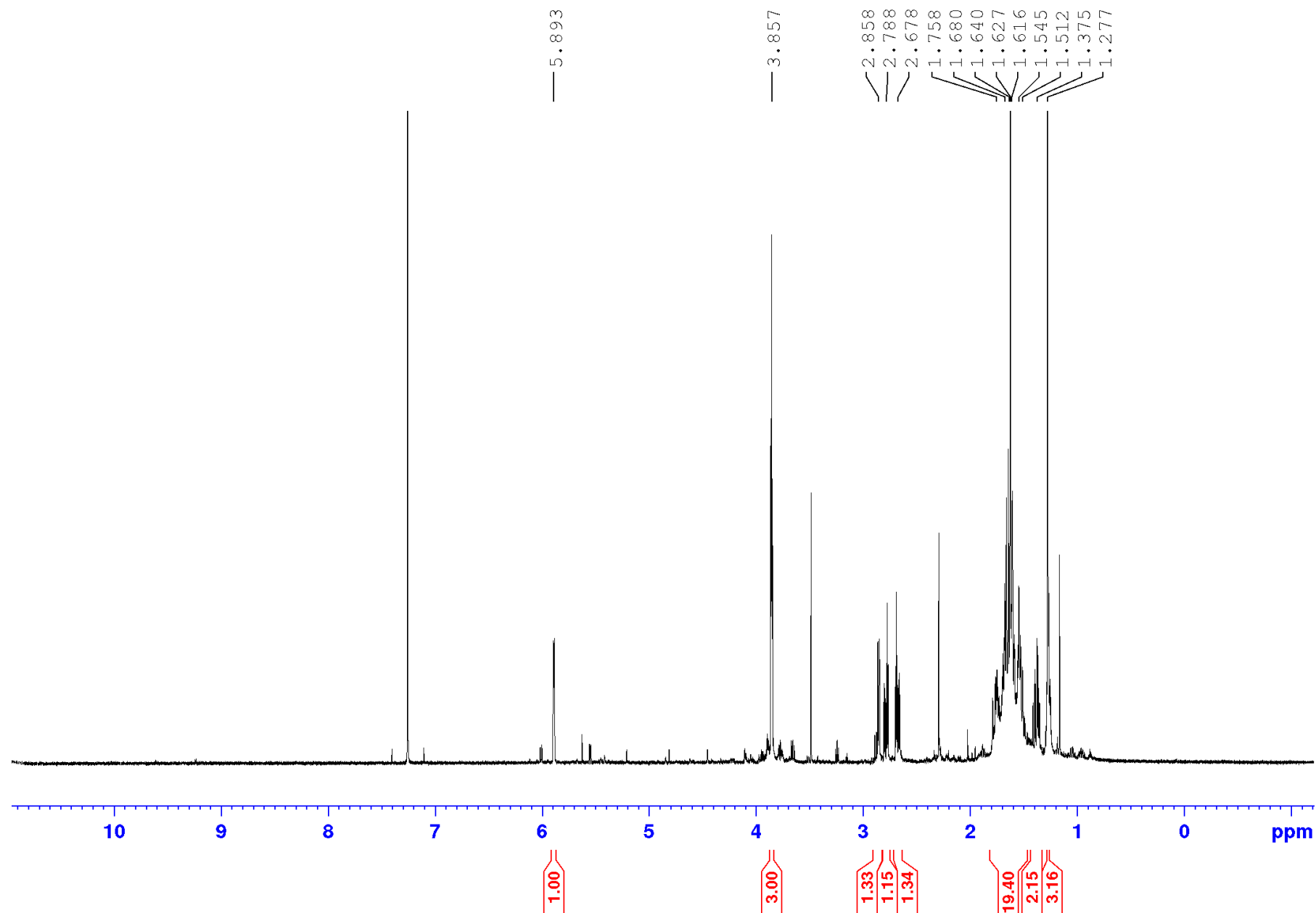


Figure S11. ^{13}C NMR spectrum of **2** measured at 176 MHz in CDCl_3

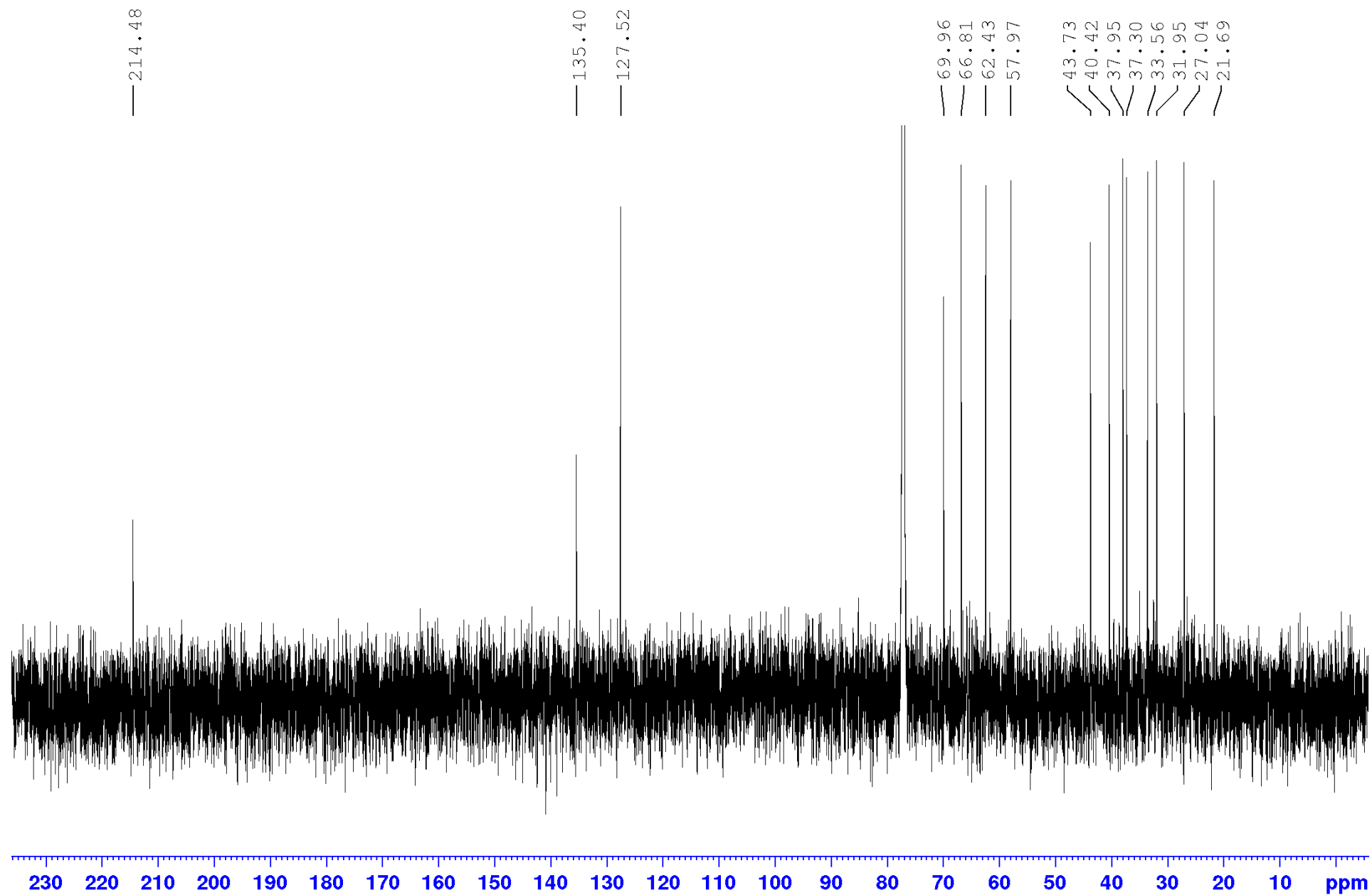


Figure S12. DEPT-135 spectrum of **2** measured at 176 MHz in CDCl₃

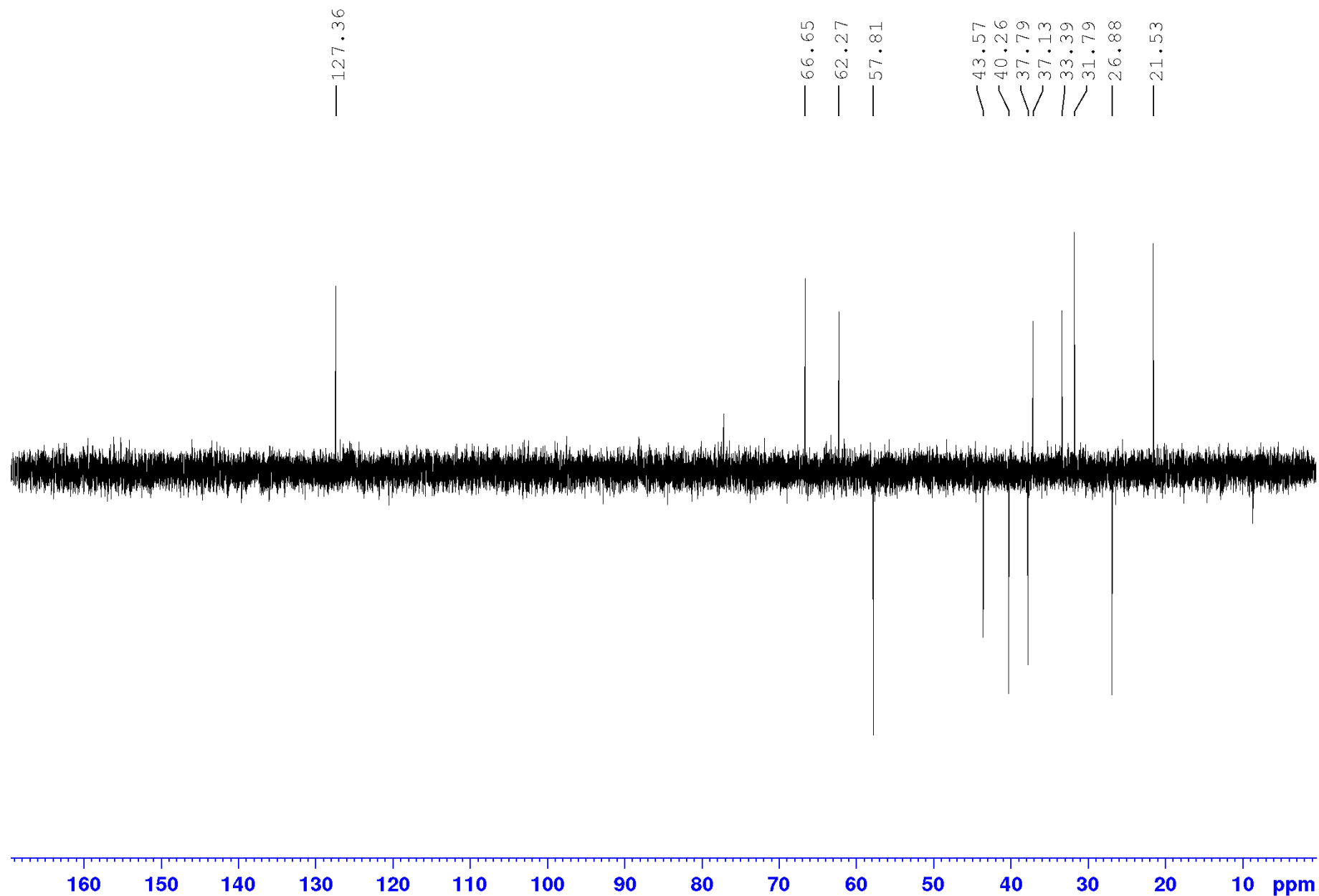


Figure S13. HSQC spectrum of 2 measured in CDCl₃

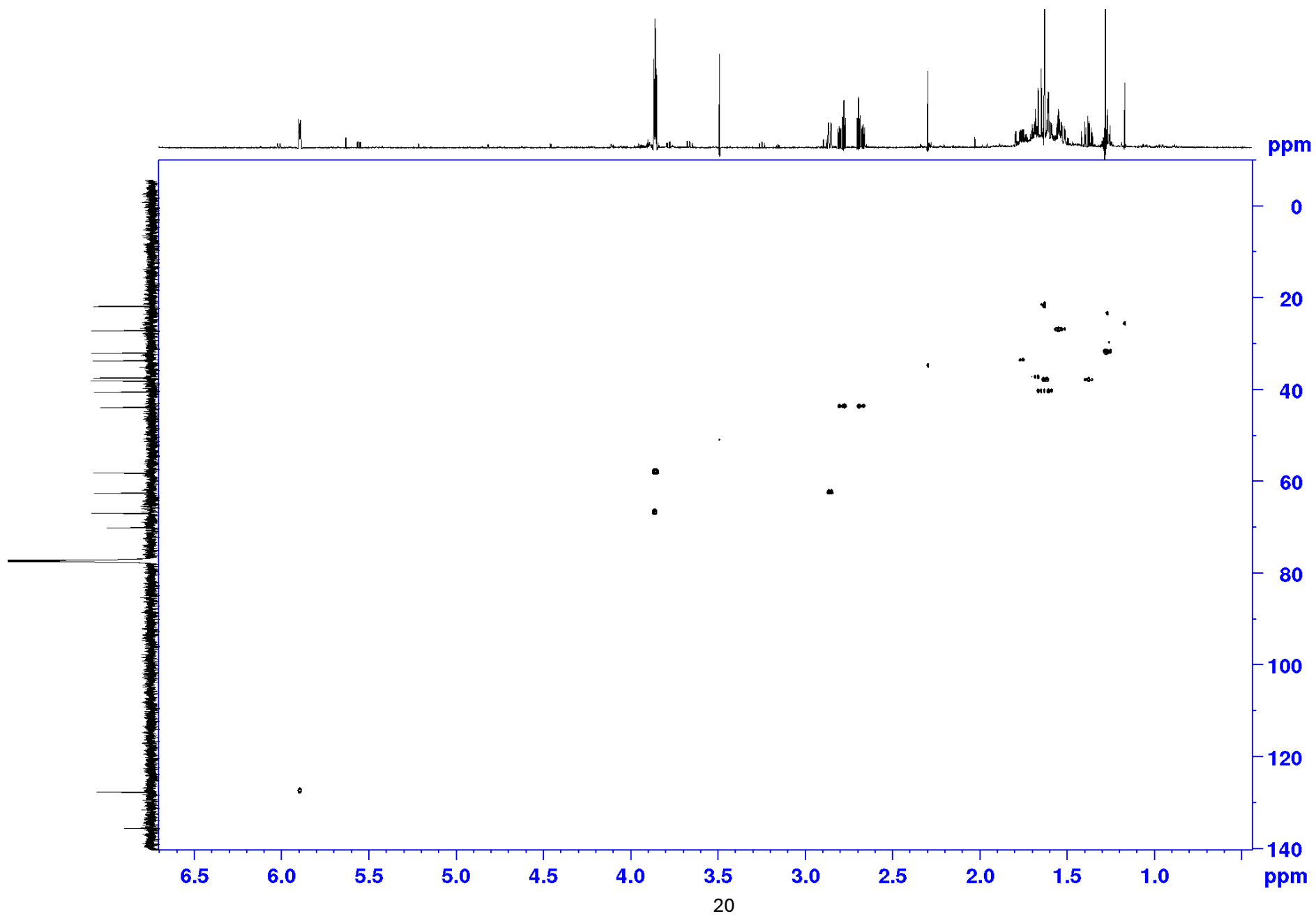


Figure S14. COSY spectrum of 2 measured in CDCl₃

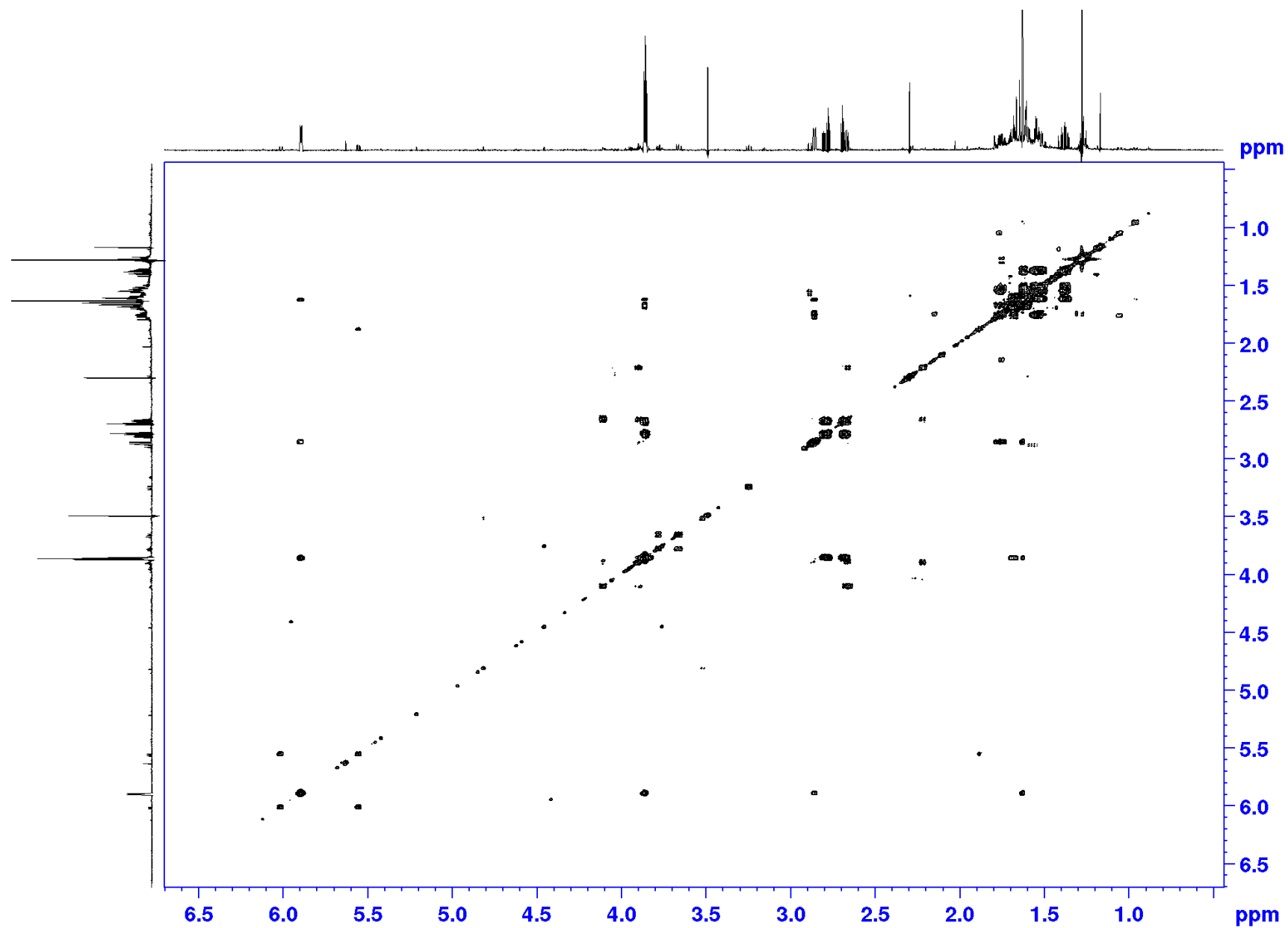


Figure S15. HMBC spectrum of 2 measured in CDCl₃

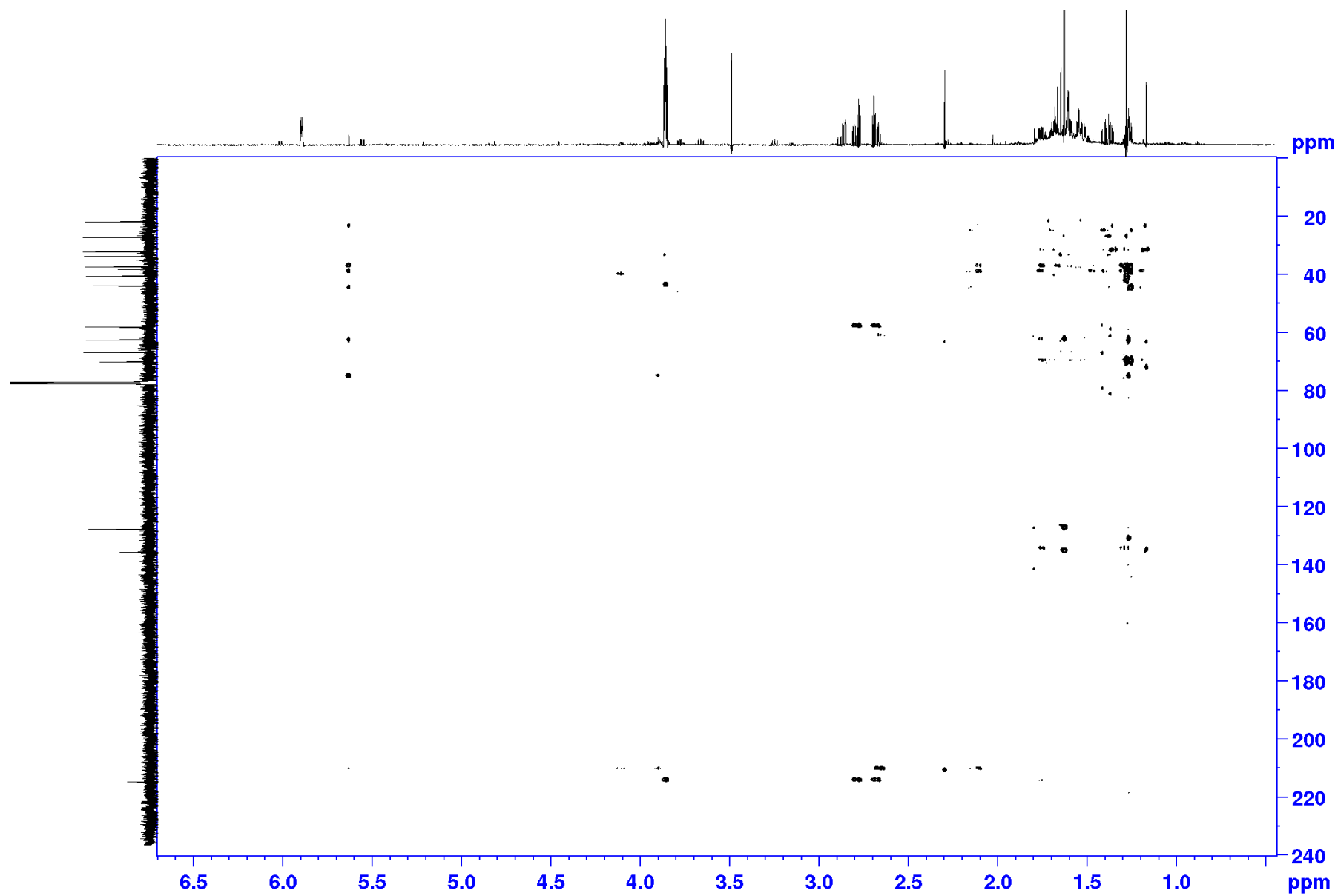


Figure S16. ROESY spectrum of **2** measured in CDCl₃

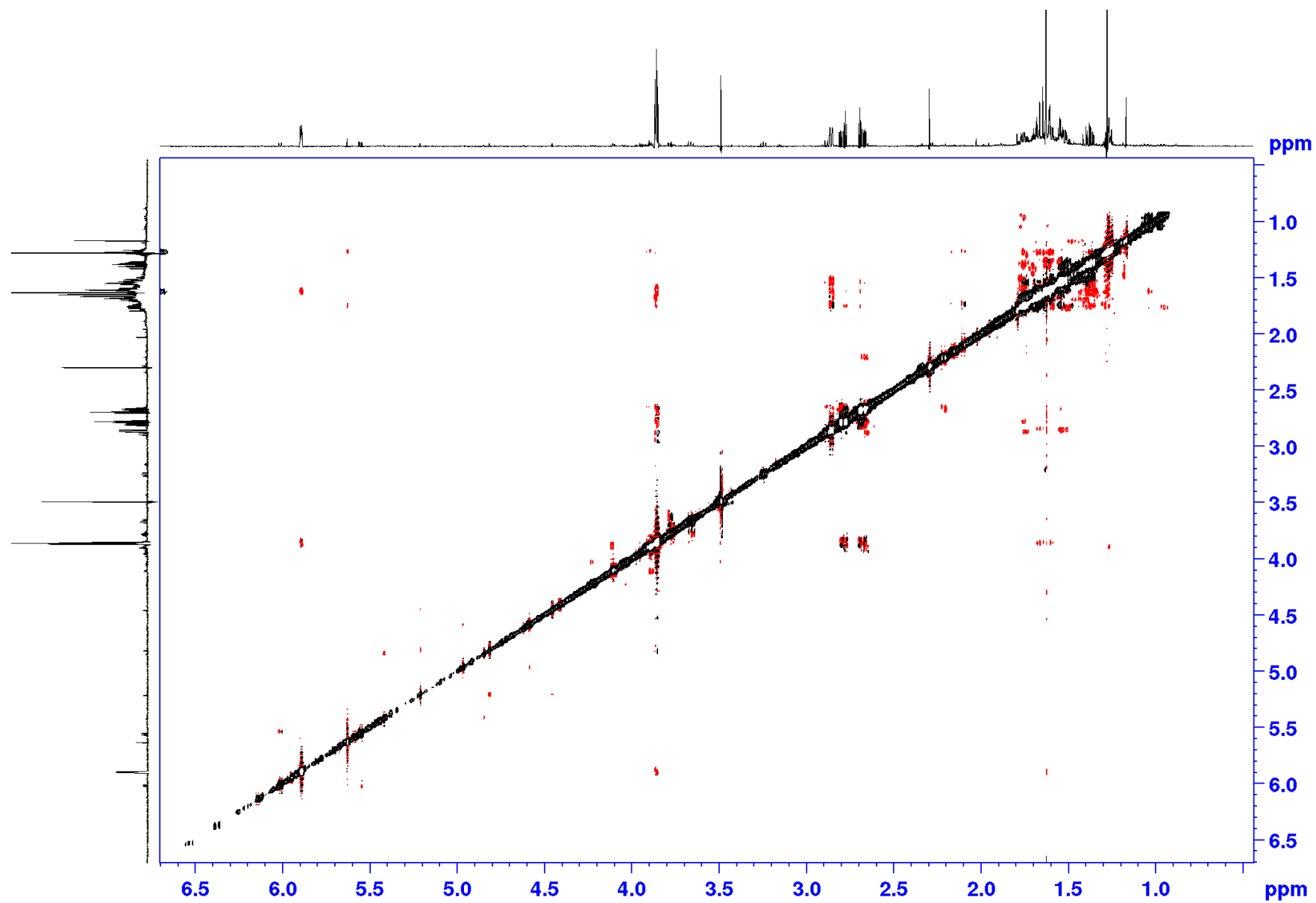
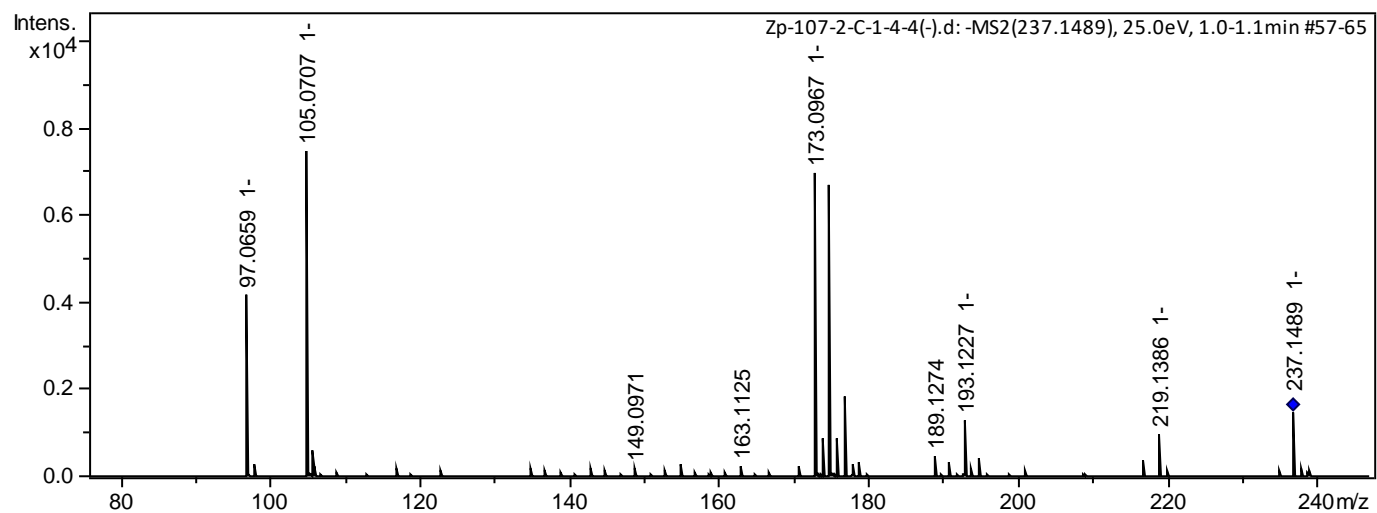
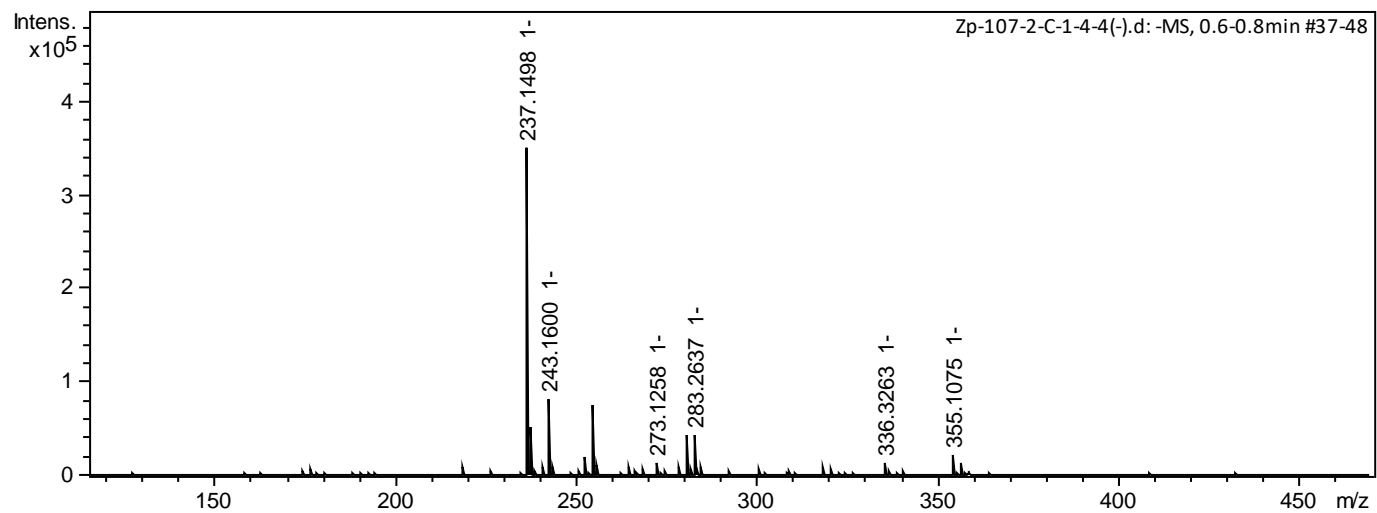
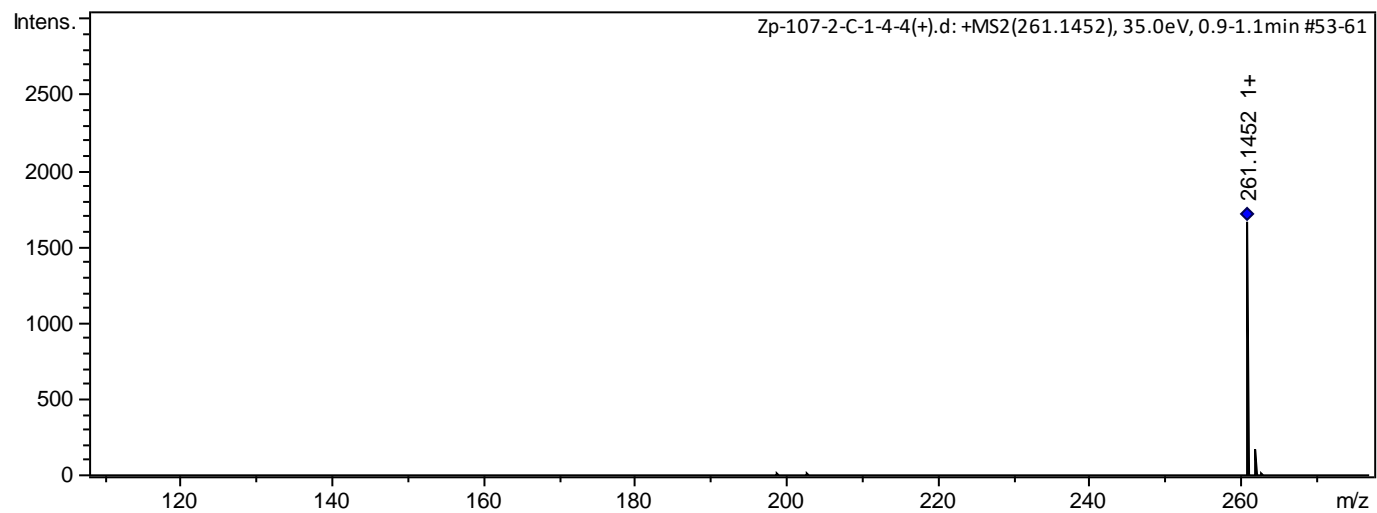
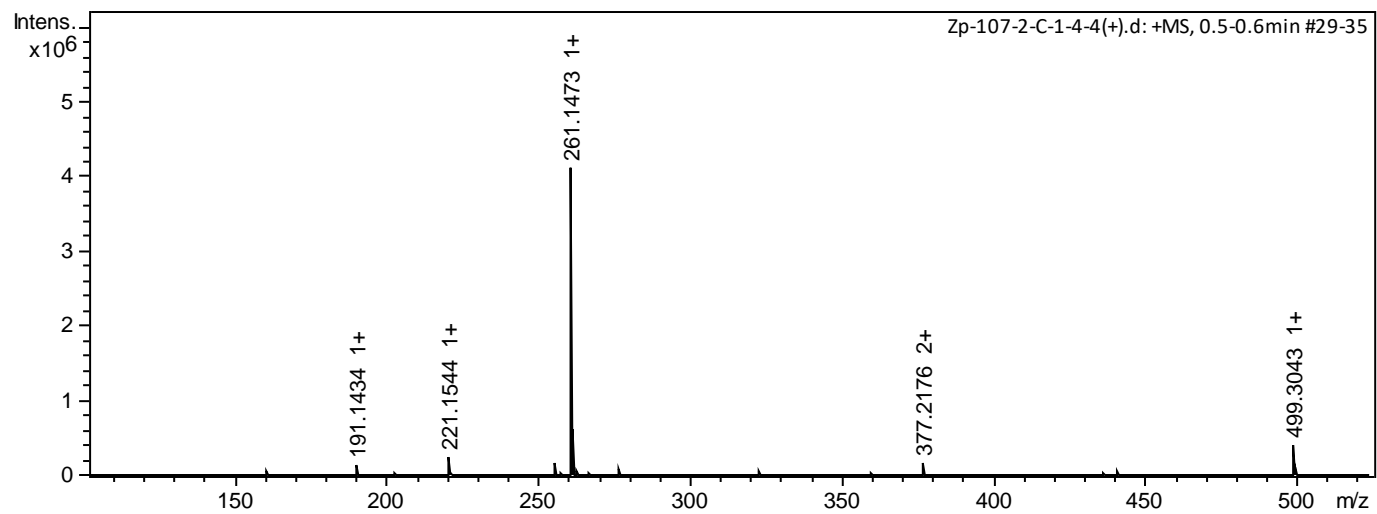


Figure S17. HRESIMS for 3





	meas	calc	Δ (ppm)
[M-H] ⁻	237,1498	237,1496	-0,8
[M+Na] ⁺	261,1473	261,1461	-4,6

Figure S18. ^1H NMR spectrum of **3** measured at 700 MHz in CDCl_3

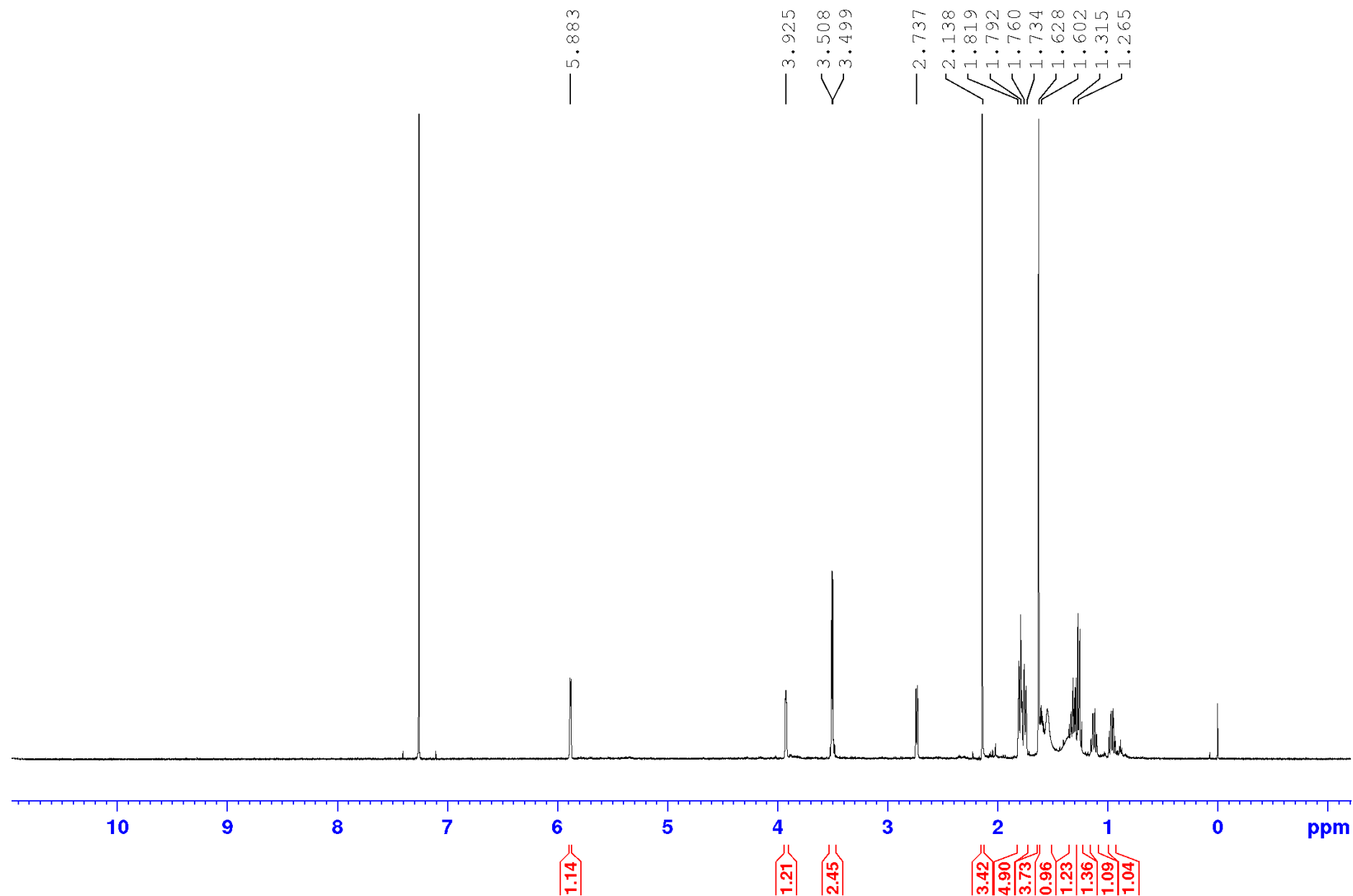


Figure S19. ^{13}C NMR spectrum of **3** measured at 176 MHz in CDCl_3

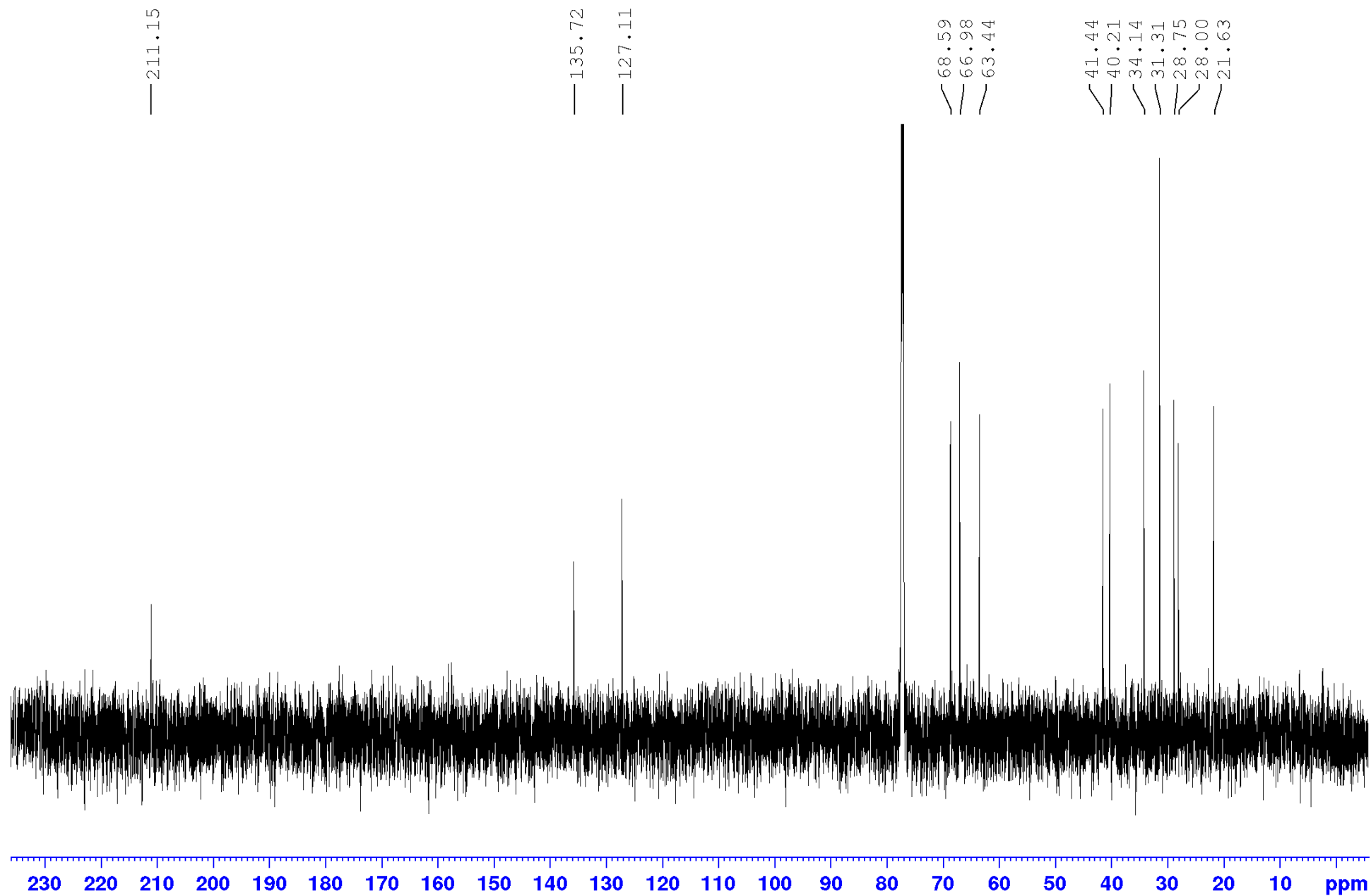


Figure S20. DEPT-135 spectrum of **3** measured at 176 MHz in CDCl₃

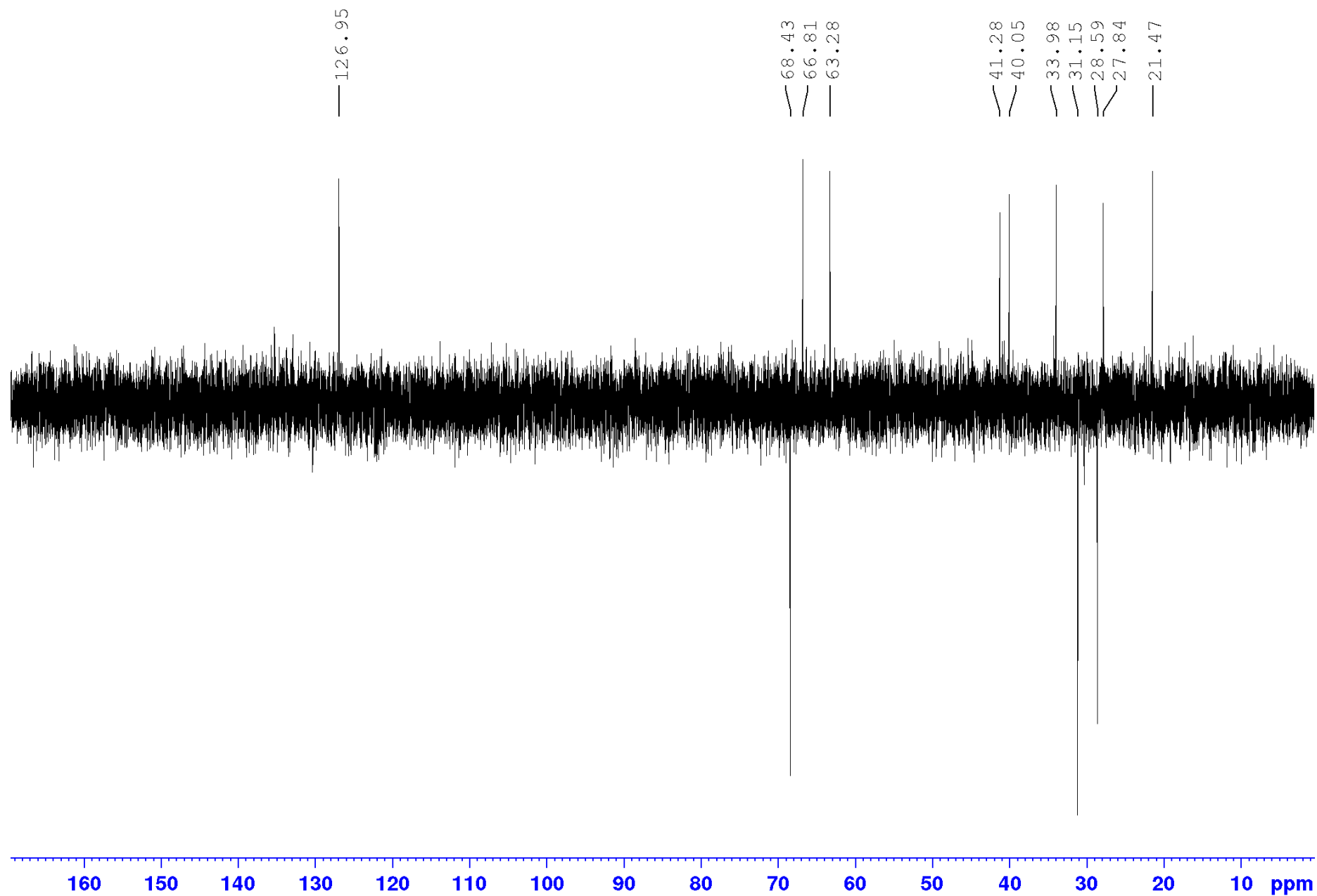


Figure S21. HSQC spectrum of 3 measured in CDCl₃

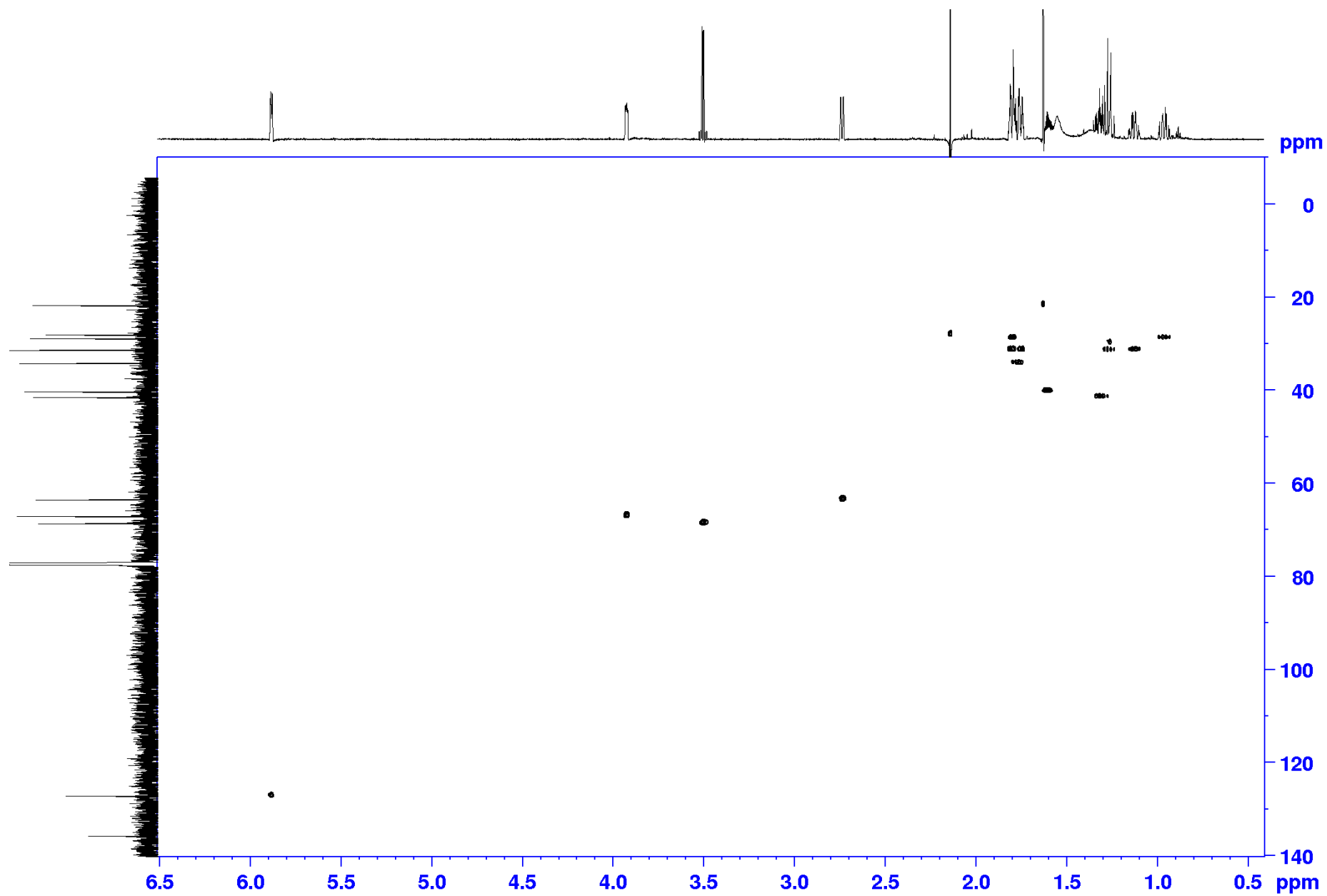


Figure S22. COSY spectrum of 3 measured in CDCl₃

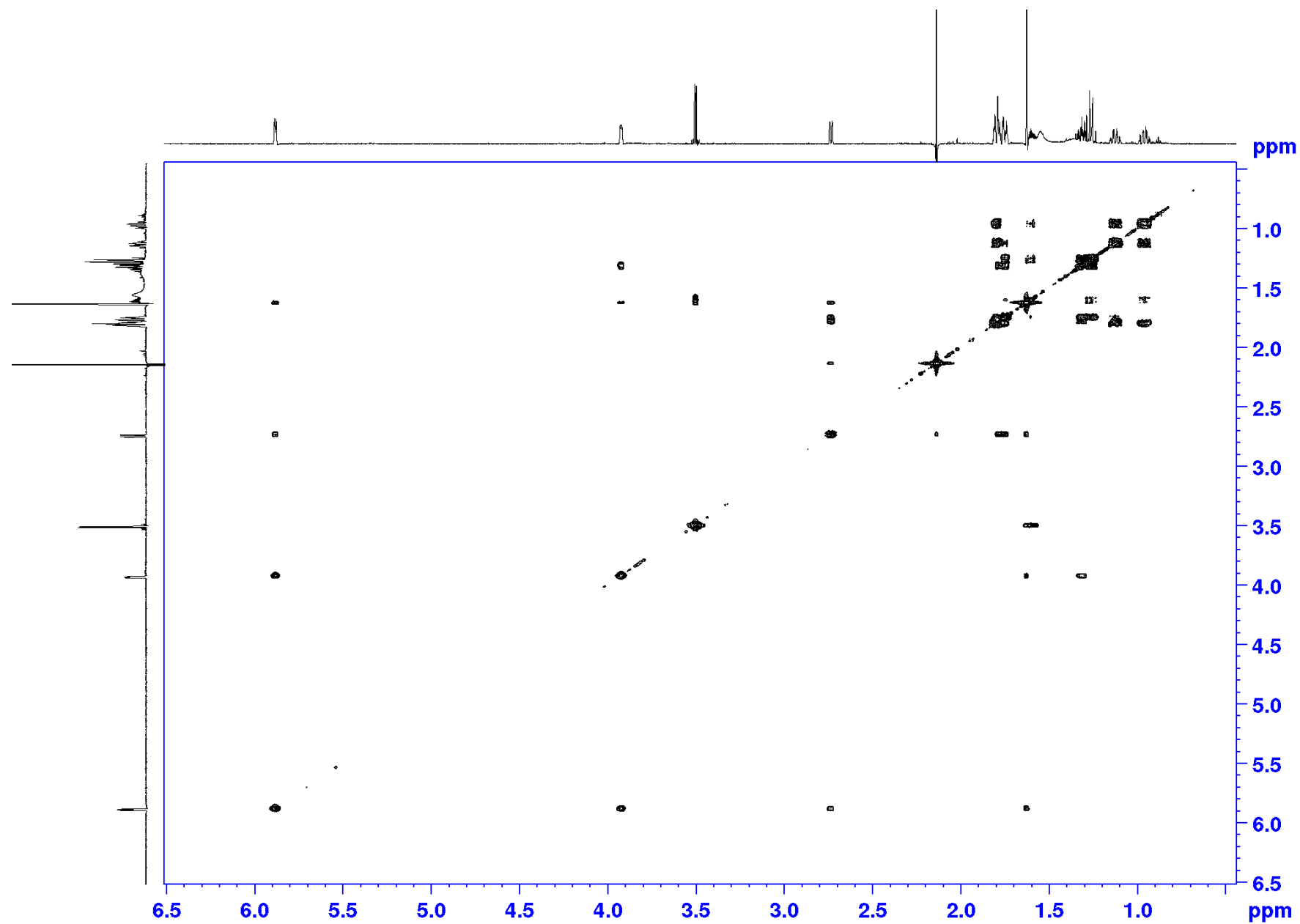


Figure S23. HMBC spectrum of 3 measured in CDCl₃

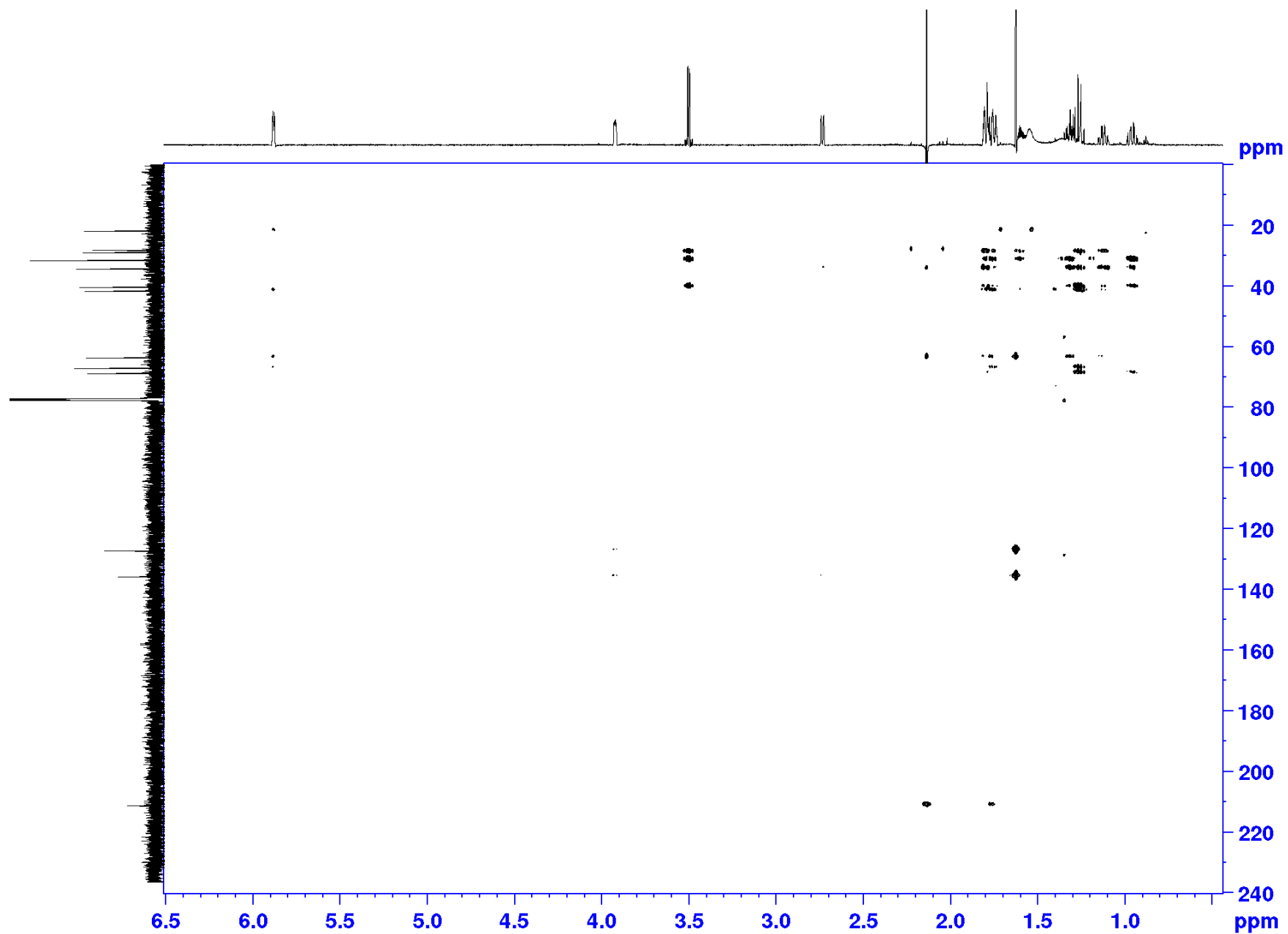


Figure S24. ROESY spectrum of **3** measured in CDCl₃

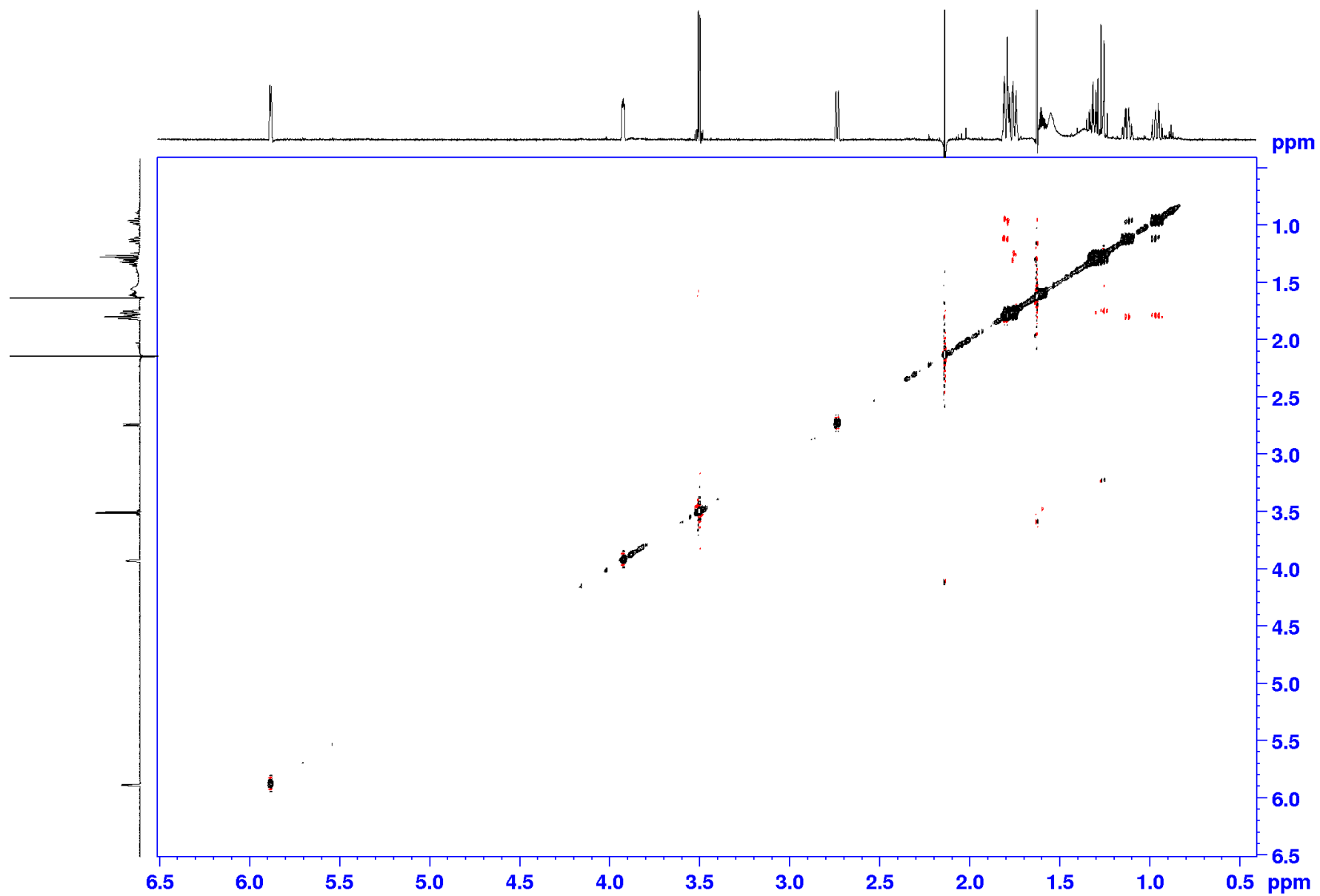
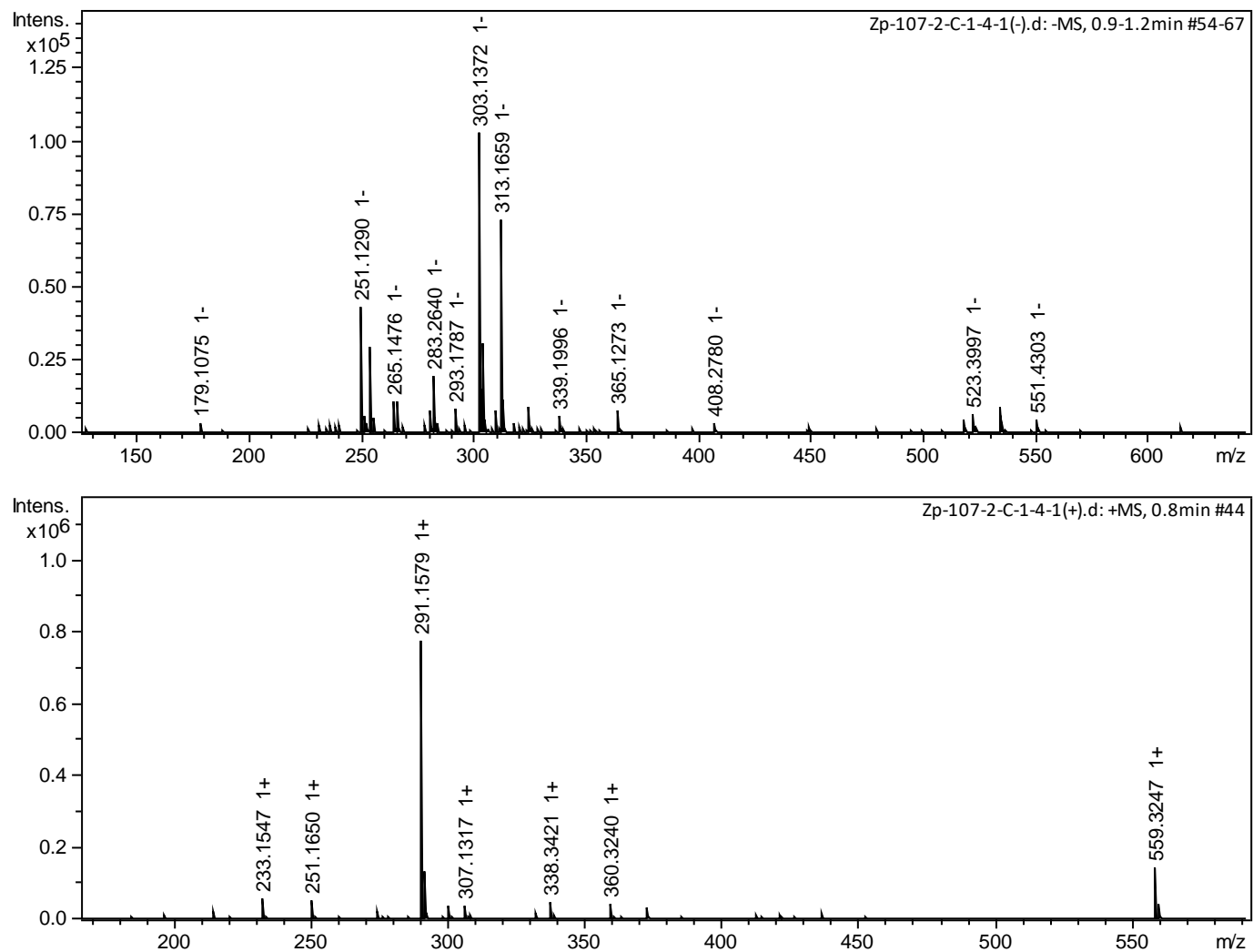
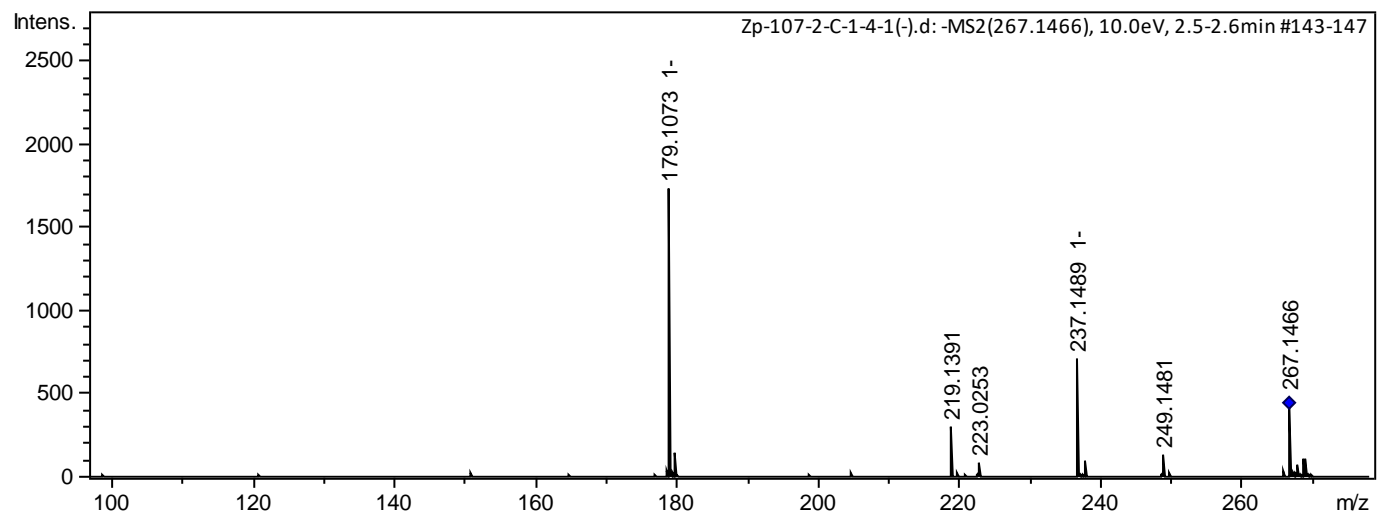
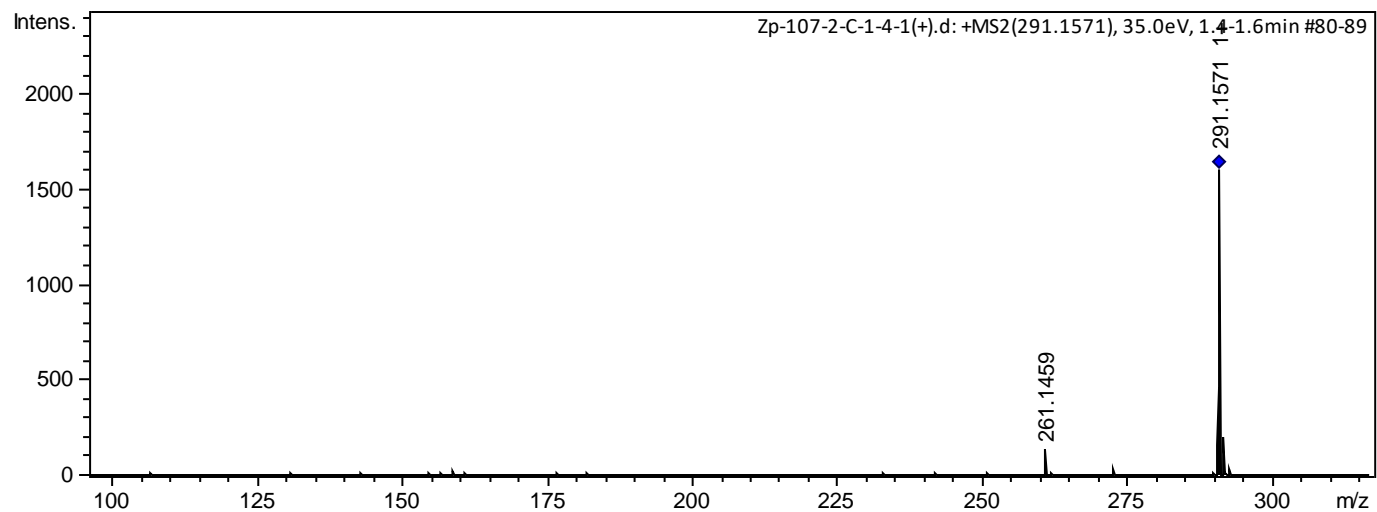


Figure S25. HRESIMS for 4





	meas	calc	Δ (ppm)
[M-H] ⁻	267,1598	267,1602	1,5
[M+Cl] ⁻	303,1372	303,1369	-1,0
[M+Na] ⁺	291,1579	291,1567	-4,1

Figure S26. ^1H NMR spectrum of 4 measured at 500 MHz in CDCl_3

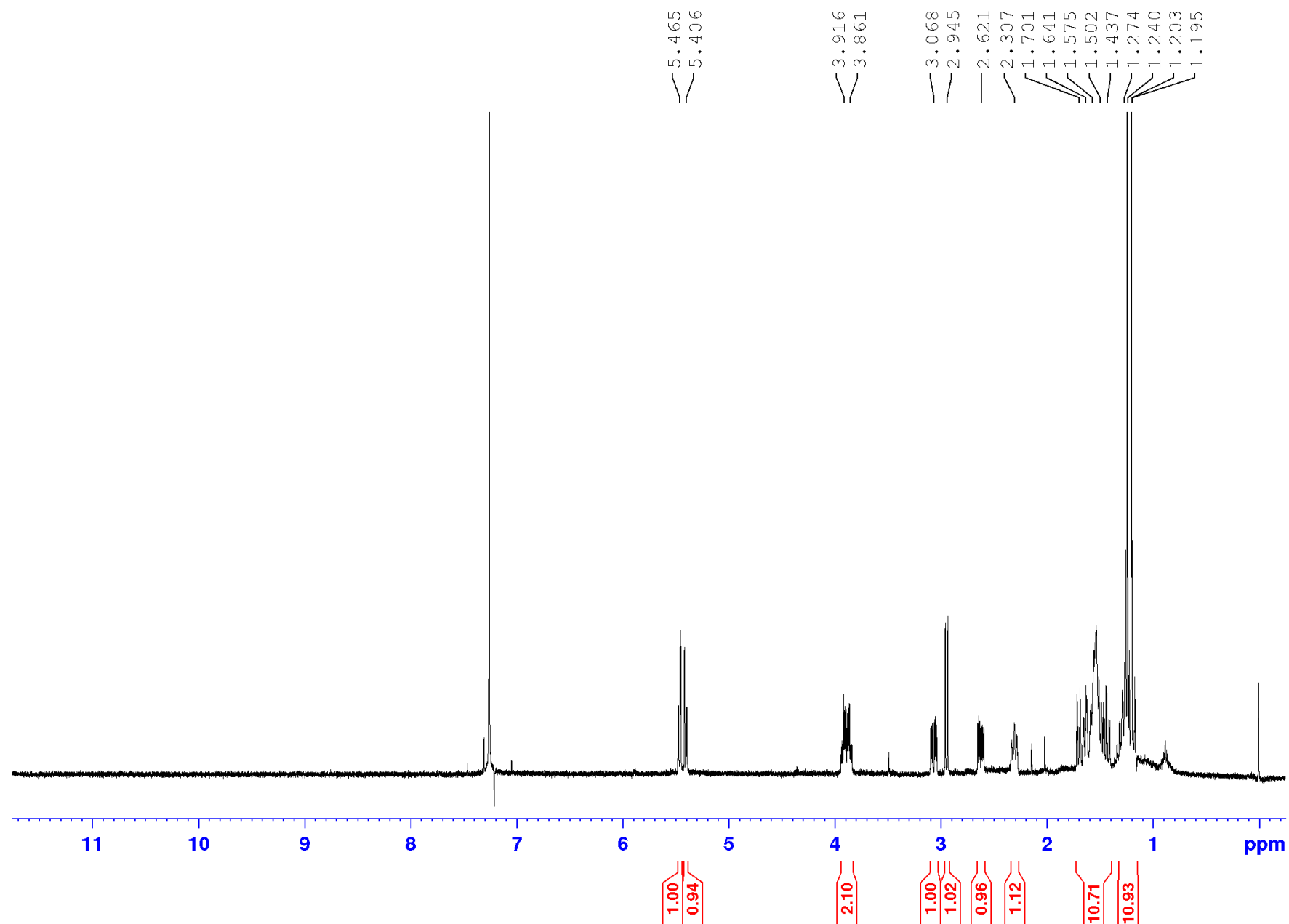


Figure S27. ^{13}C NMR spectrum of **4** measured at 125 MHz in CDCl_3

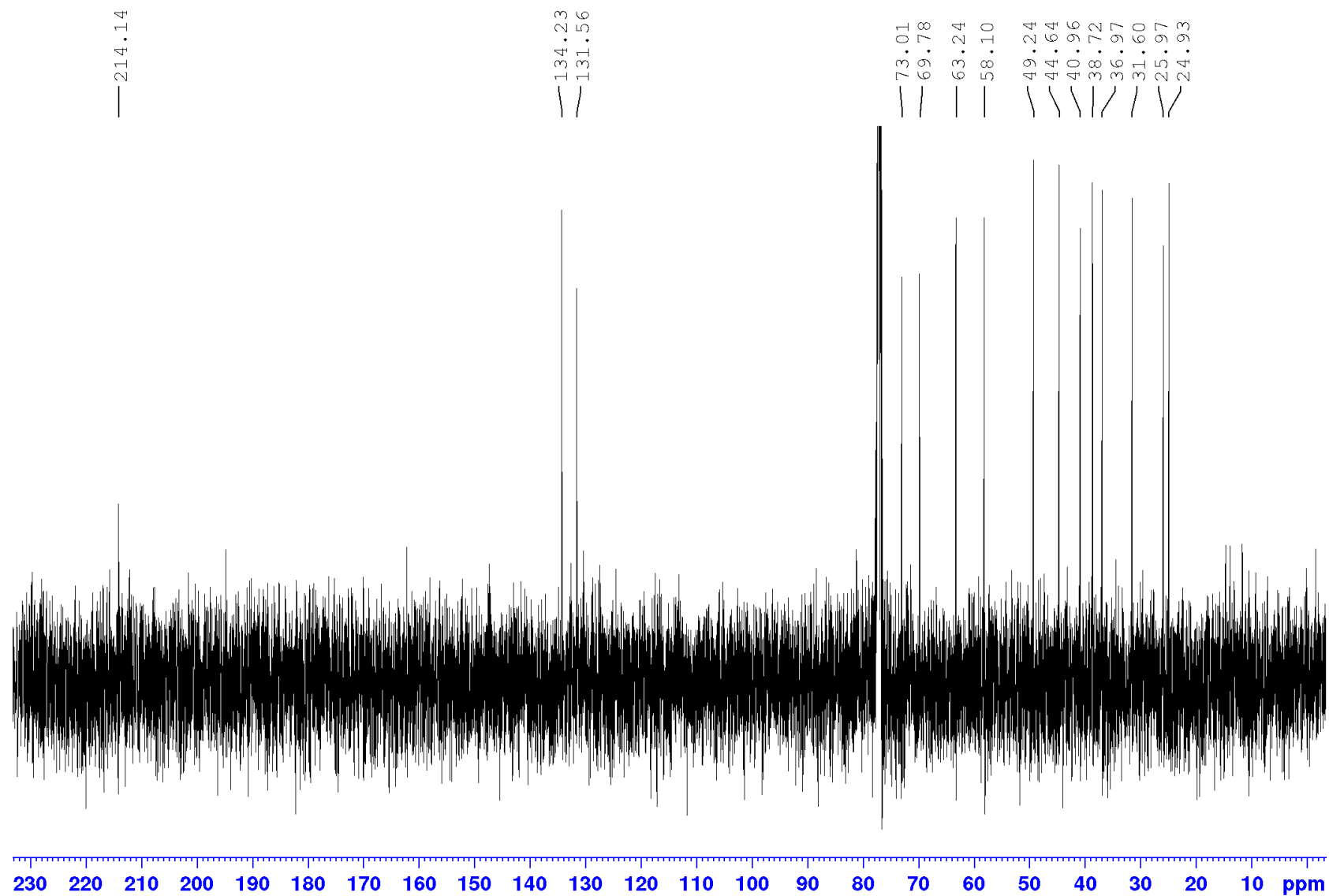


Figure S28. HSQC spectrum of 4 measured in CDCl₃

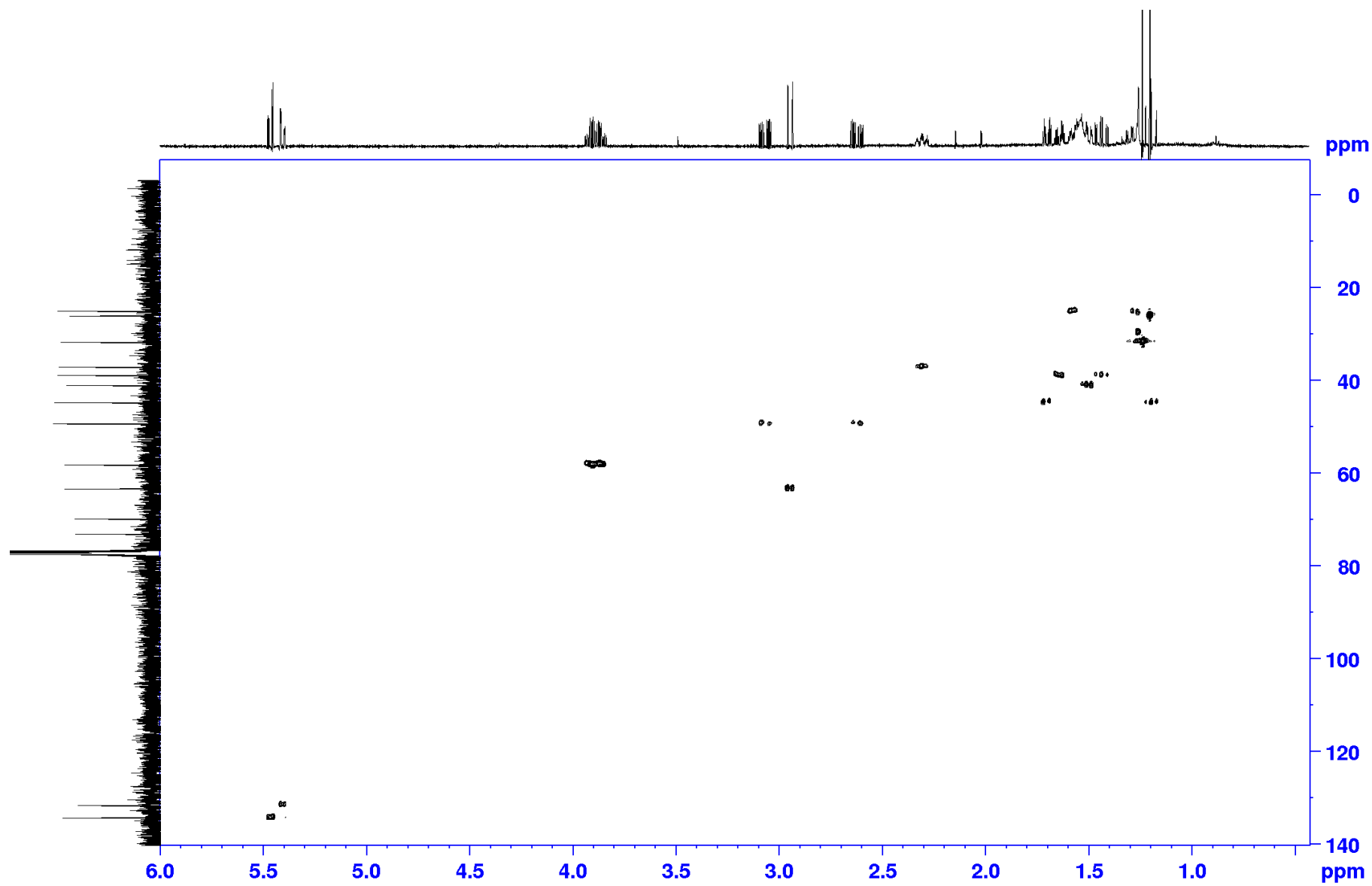


Figure S29. COSY spectrum of 4 measured in CDCl₃

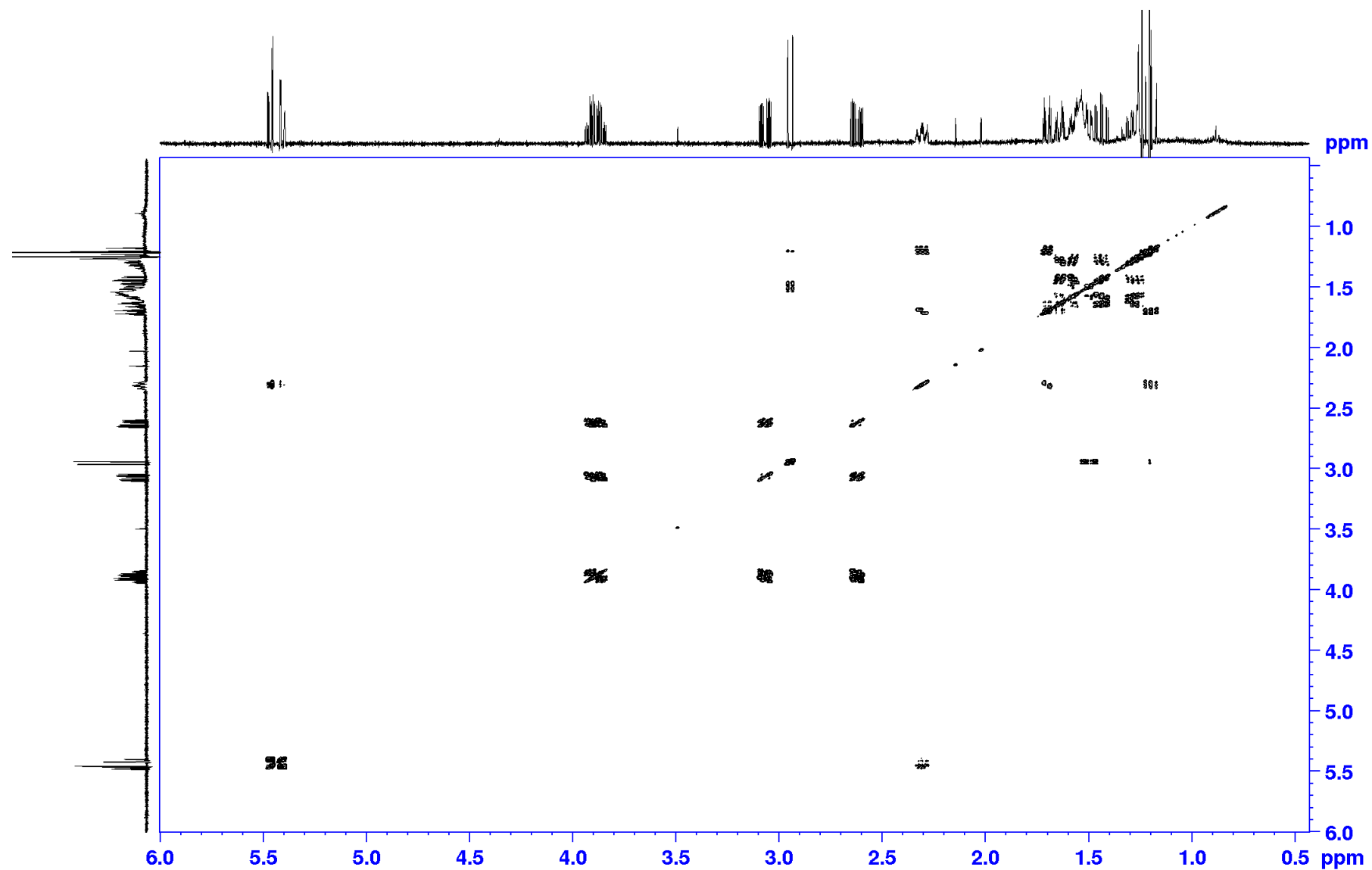


Figure S30. HMBC spectrum of 4 measured in CDCl₃

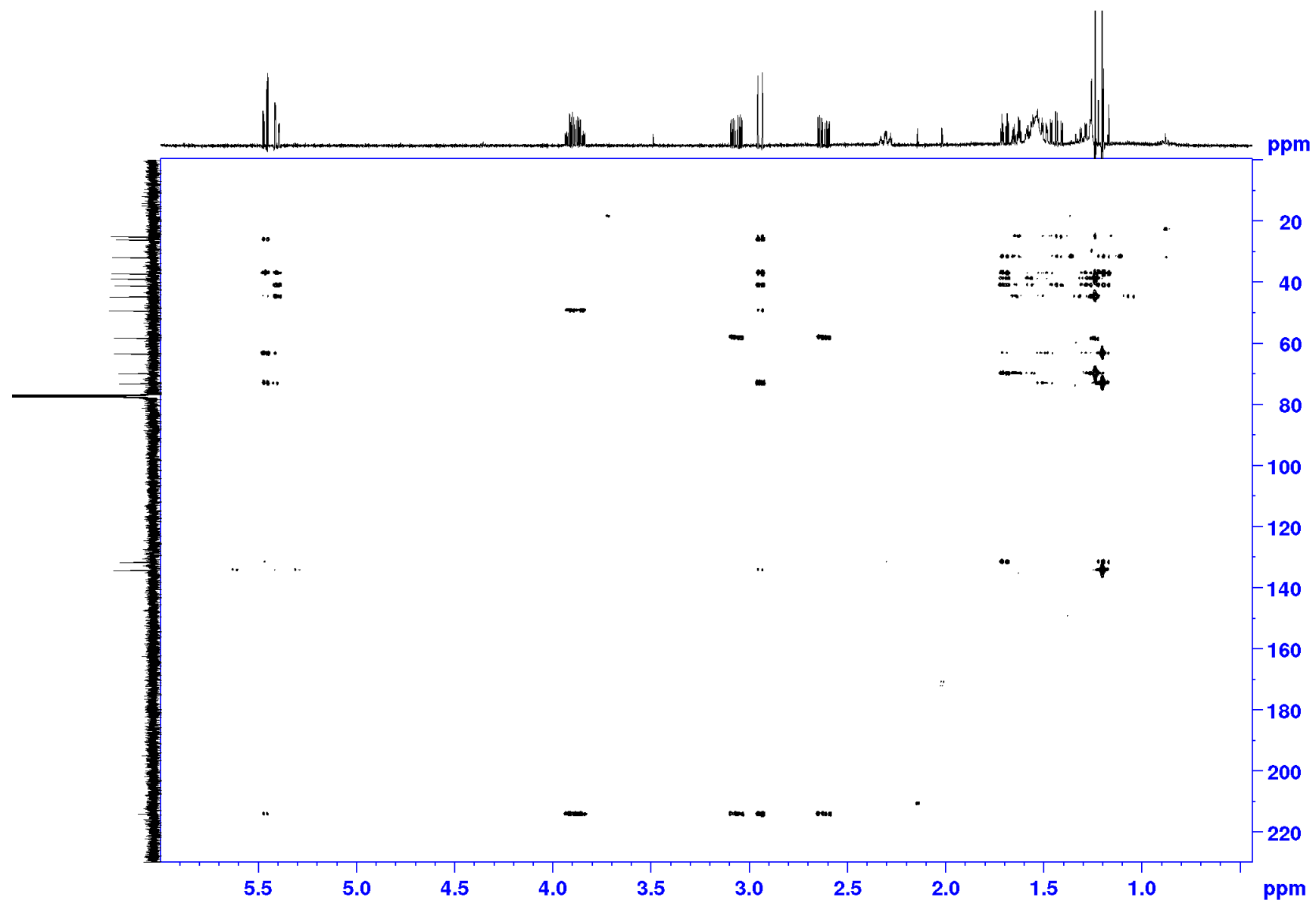


Figure S31. ROESY spectrum of 4 measured in CDCl₃

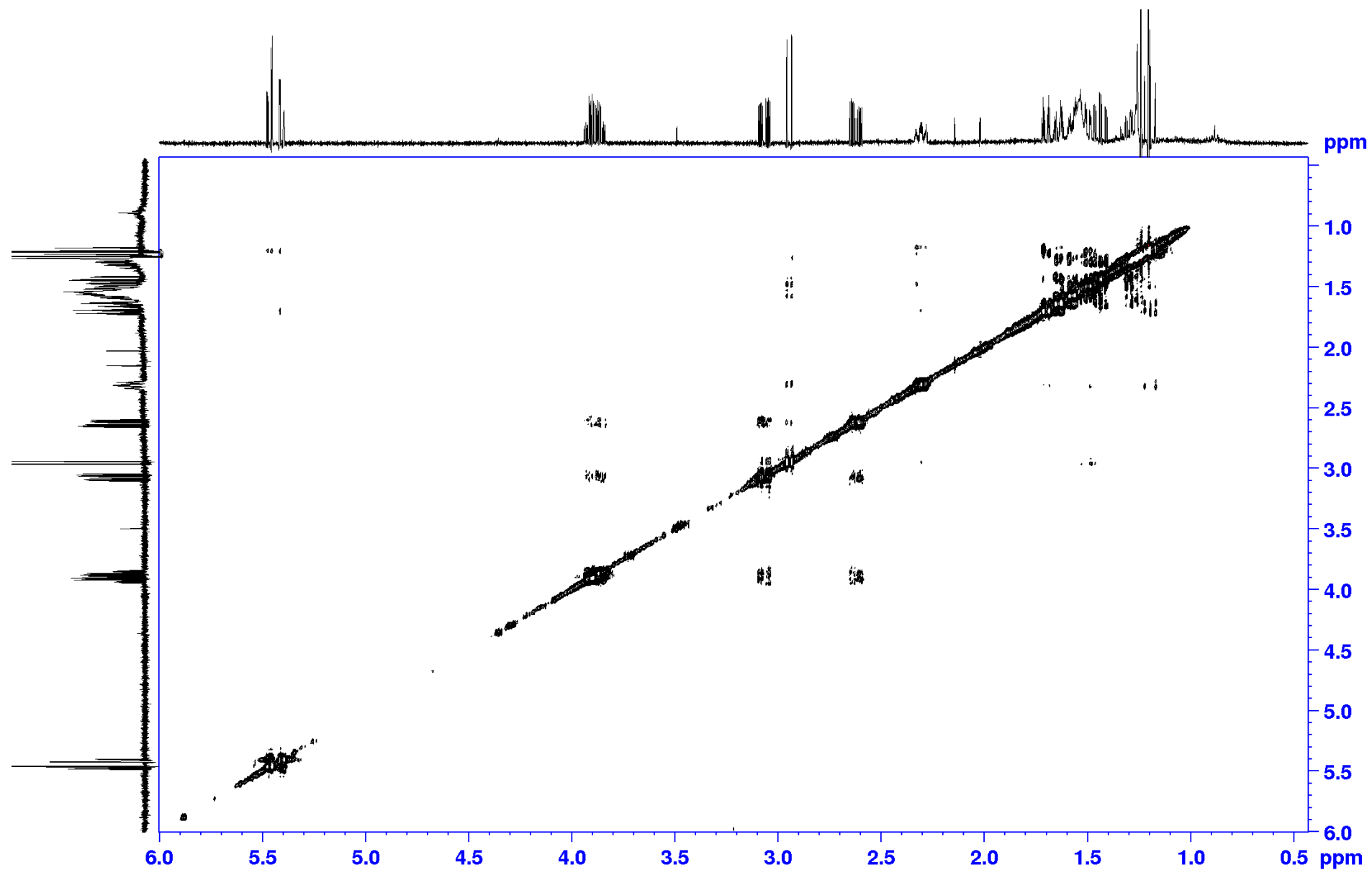
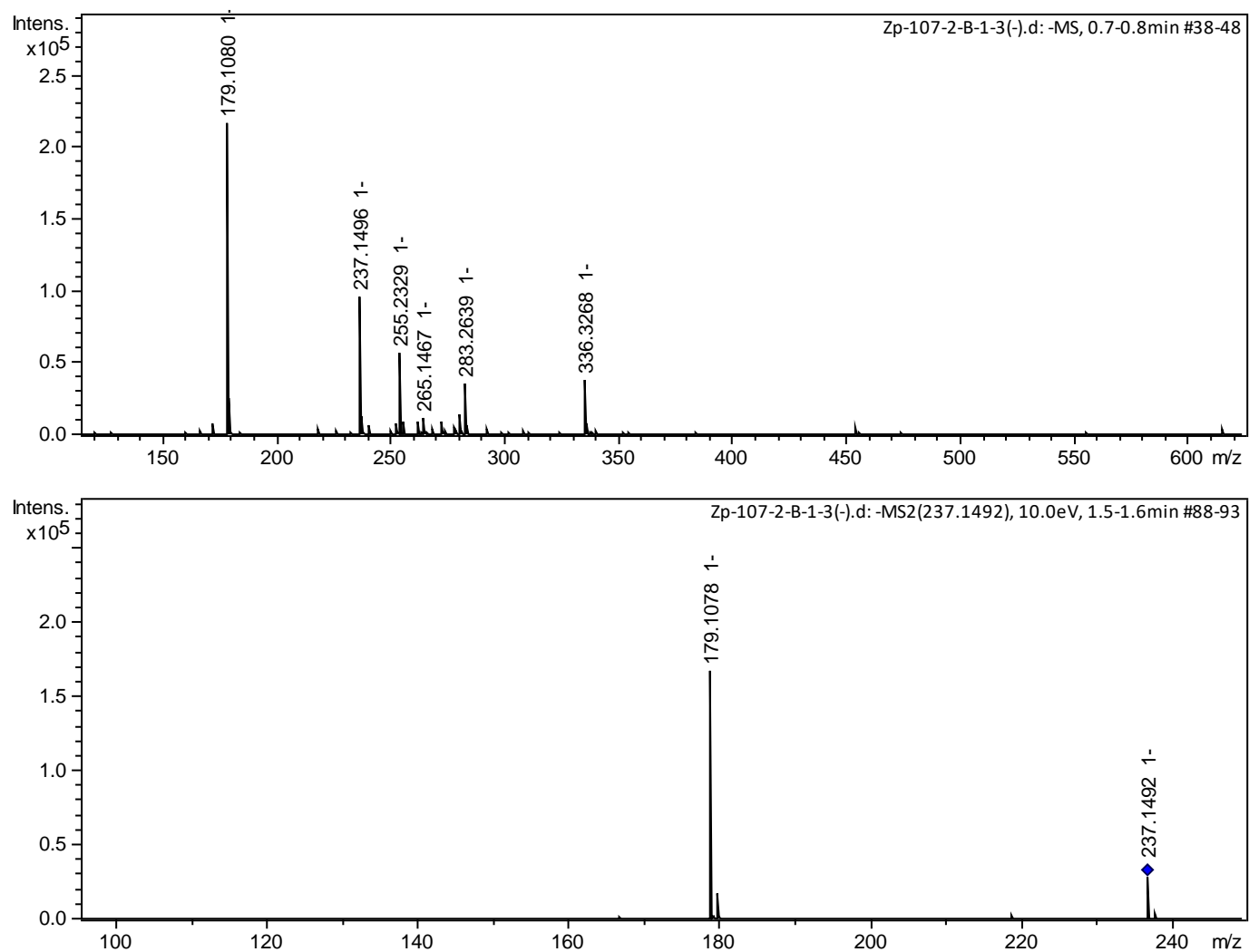
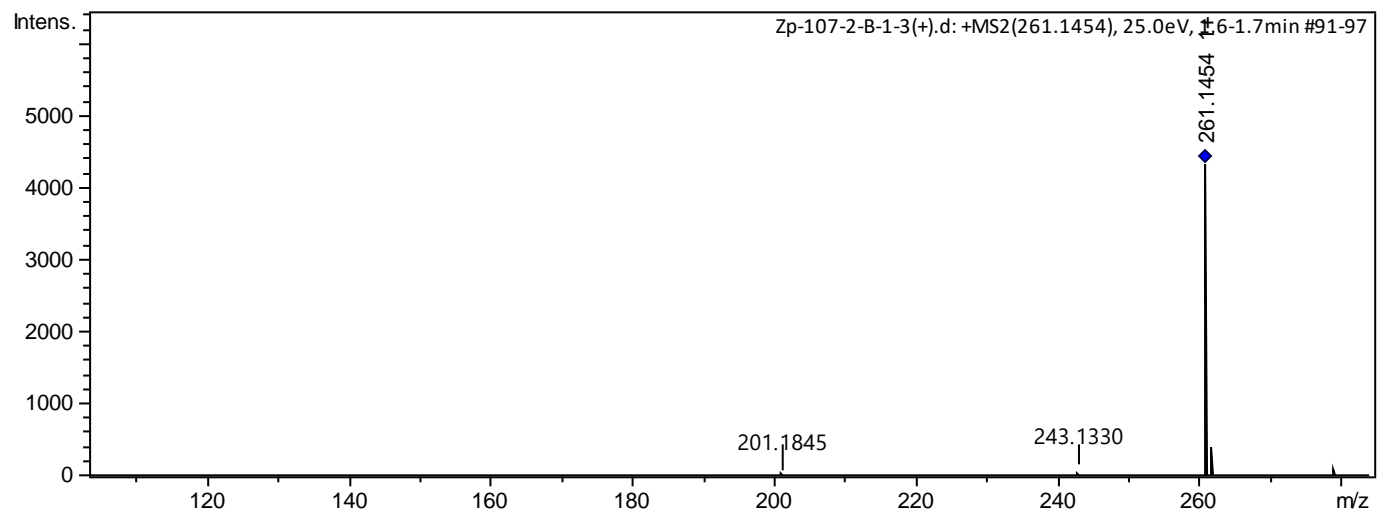
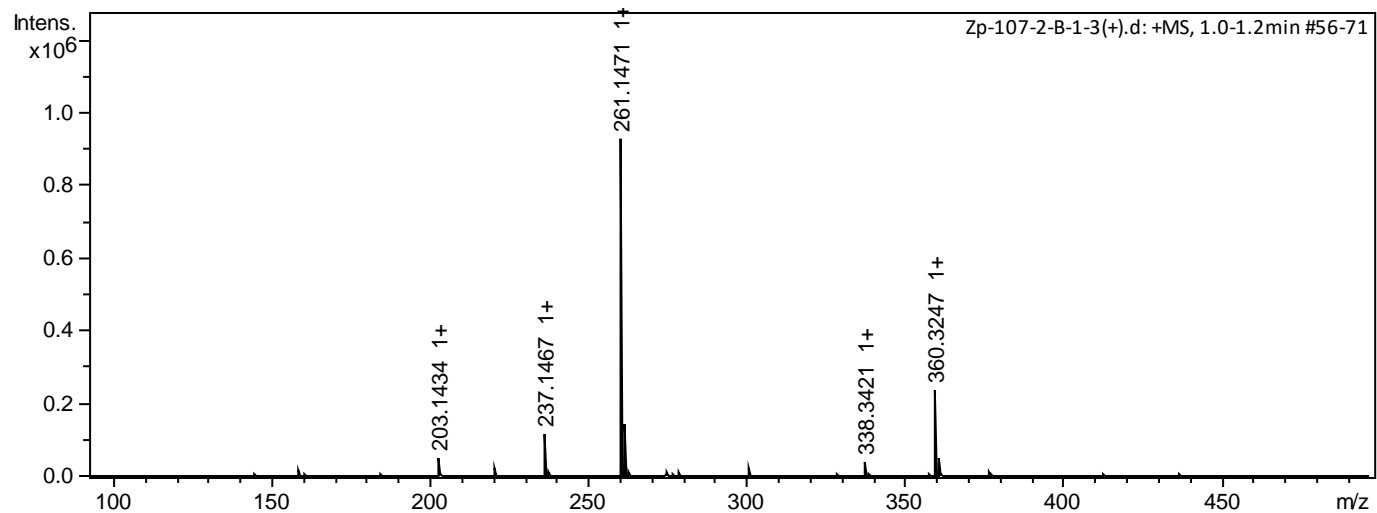


Figure S32. HRESIMS for 5





	meas	calc	Δ (ppm)
[M-H] ⁻	237,1496	237,1496	0,0
[M+Na] ⁺	261,1471	261,1461	-3,8

Figure S33. ^1H NMR spectrum of **5** measured at 700 MHz in CDCl_3

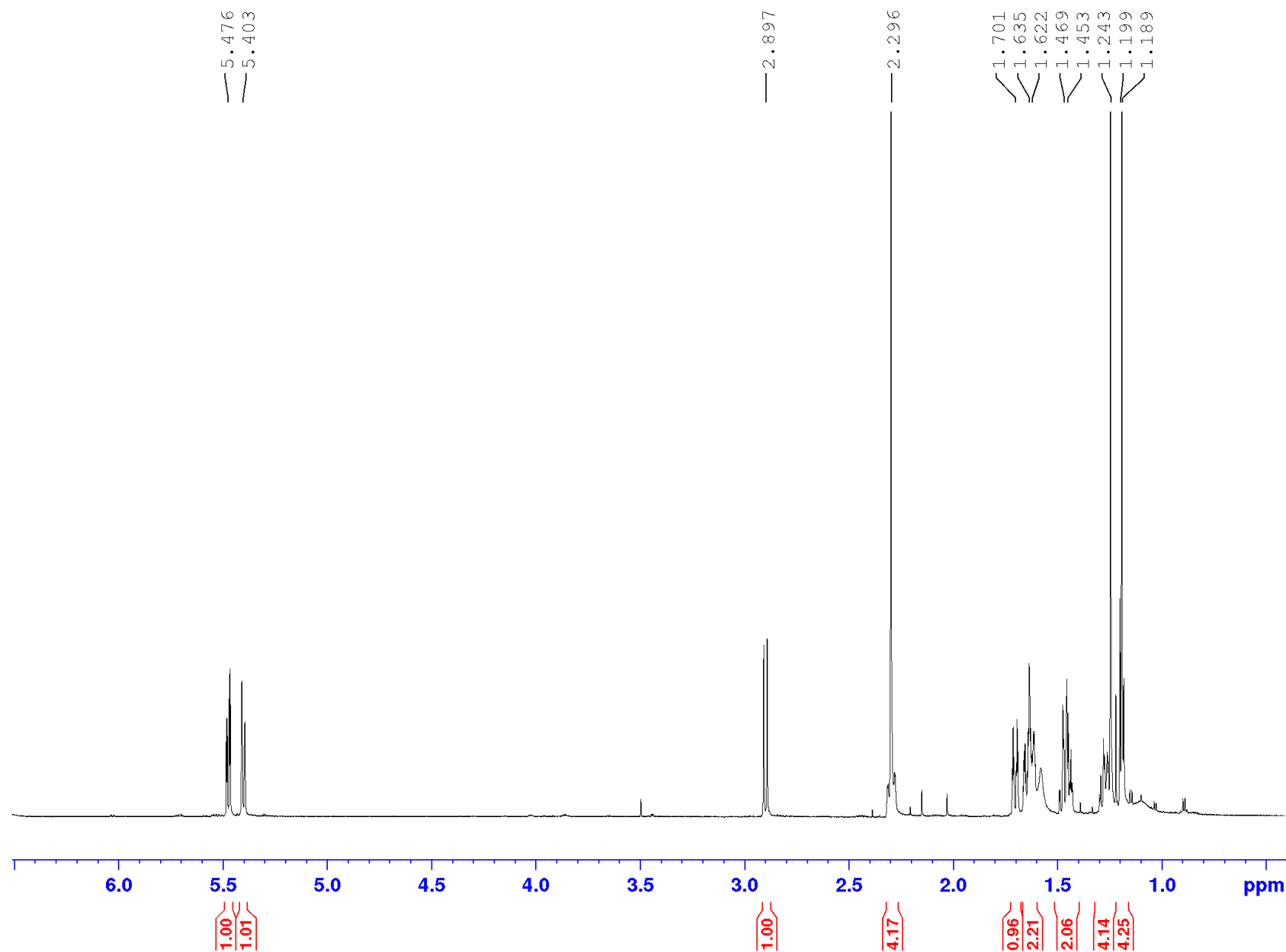


Figure S34. ^{13}C NMR spectrum of 5 measured at 176 MHz in CDCl_3

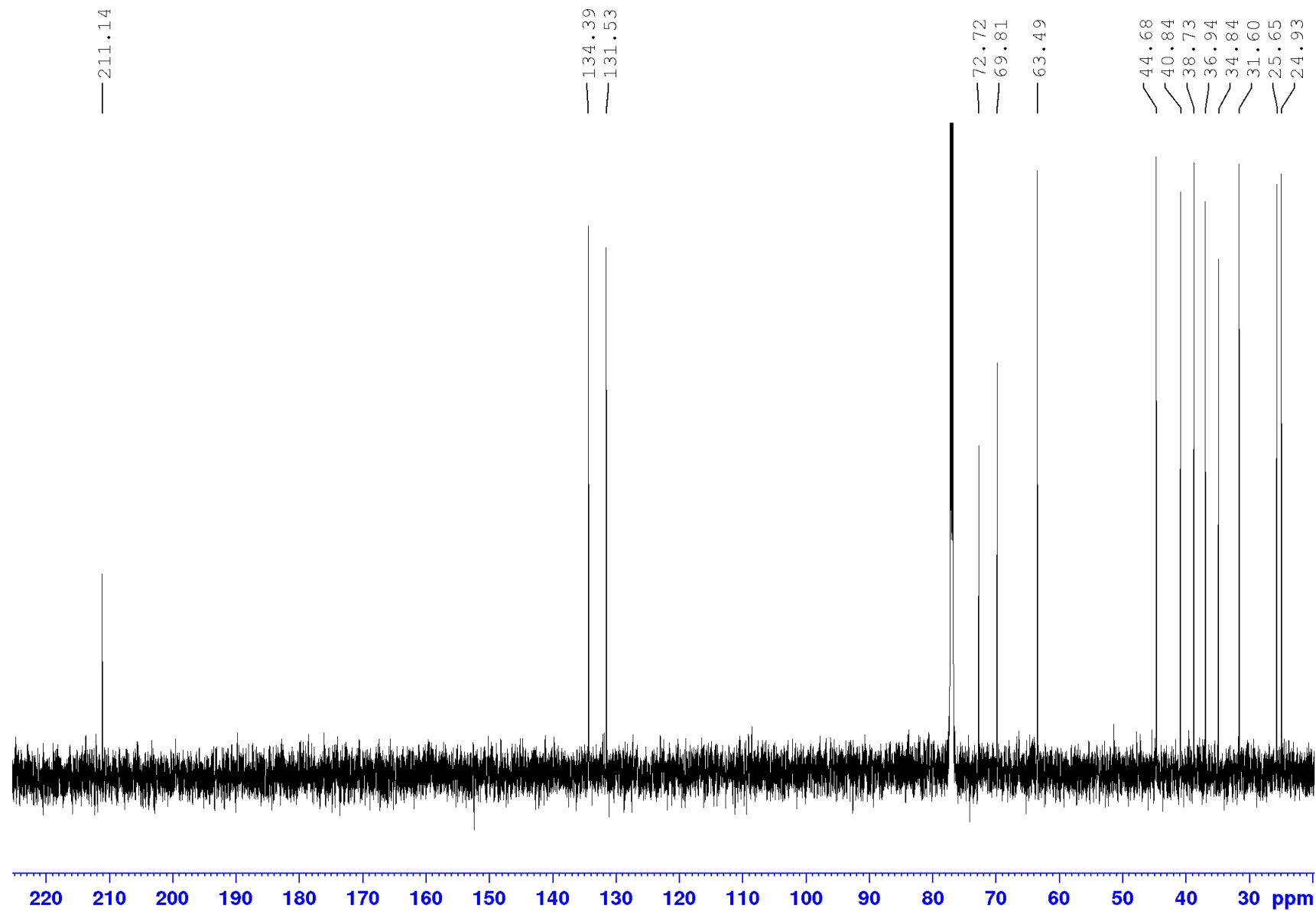


Figure S35. DEPT-135 spectrum of **5** measured at 176 MHz in CDCl₃

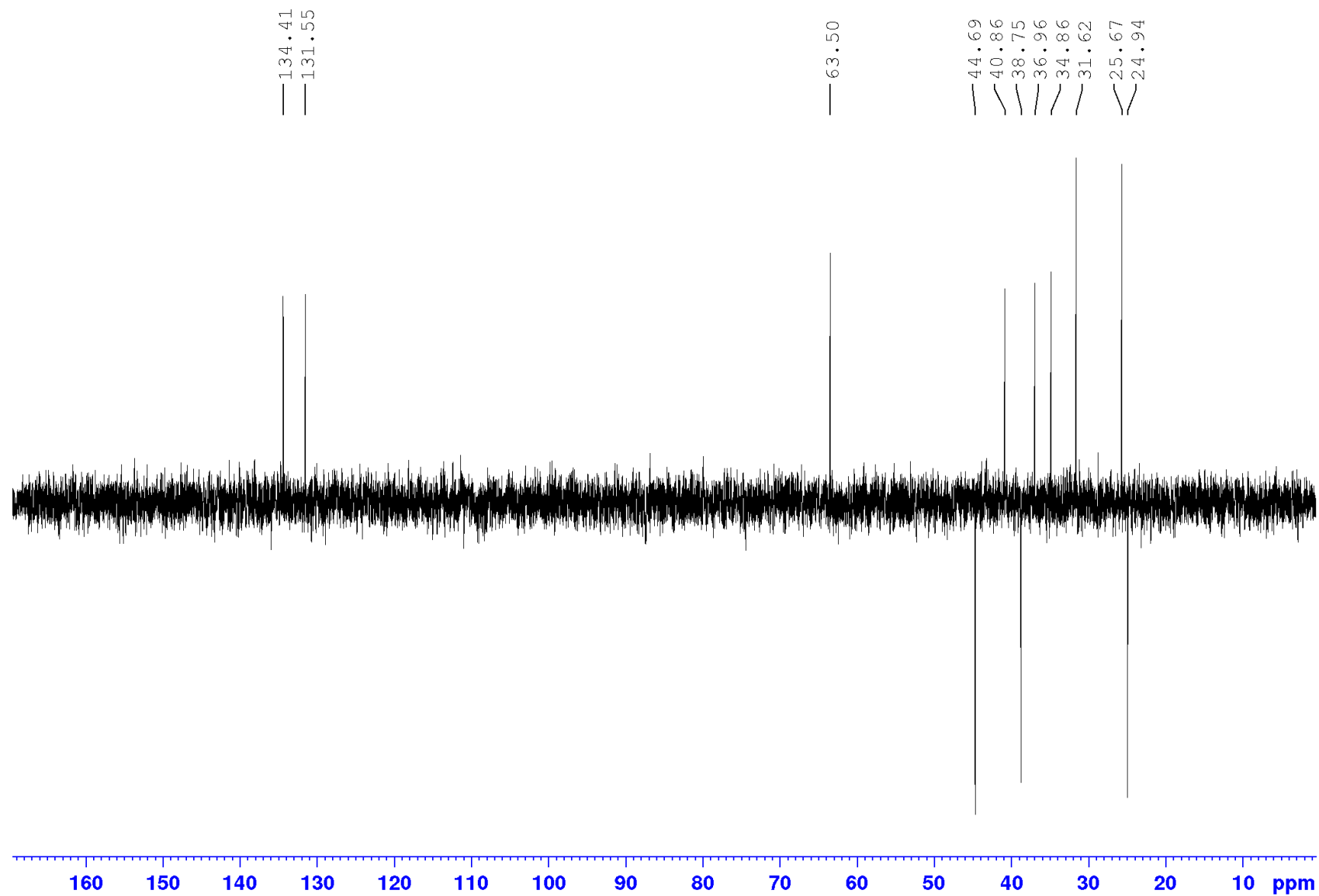


Figure S36. HSQC spectrum of 5 measured in CDCl₃

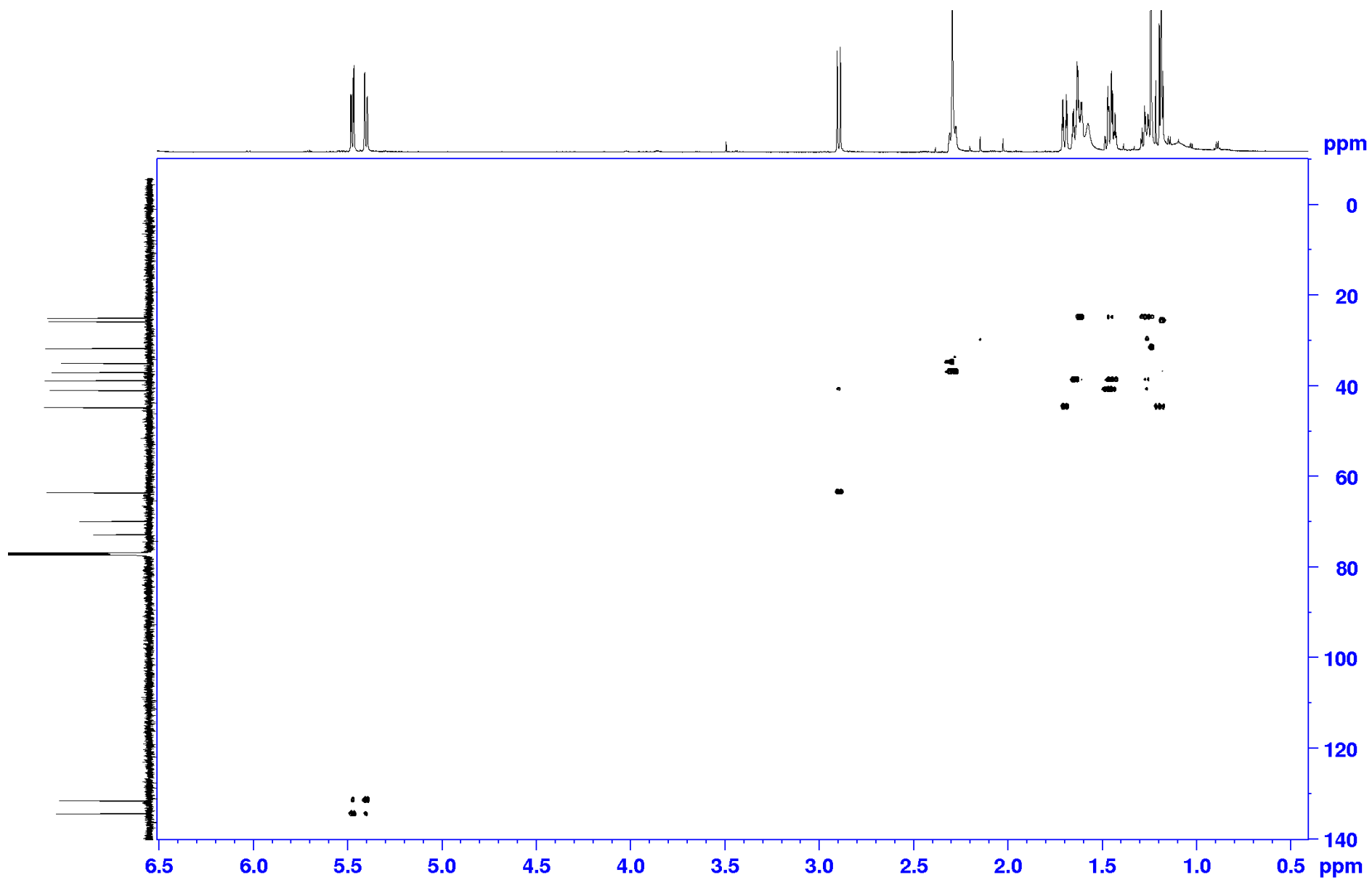


Figure S37. COSY spectrum of 5 measured in CDCl_3

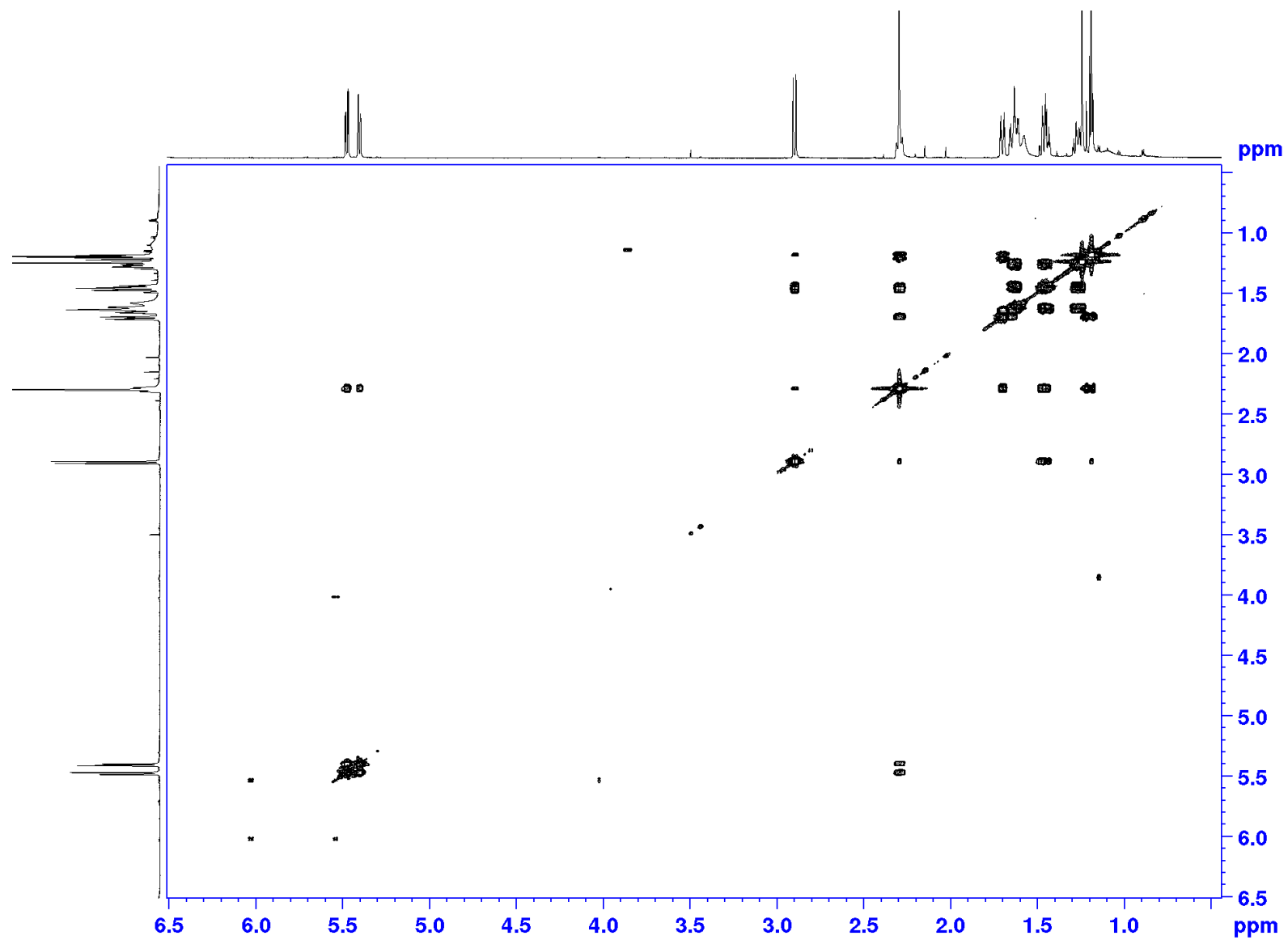


Figure S38. HMBC spectrum of 5 measured in CDCl_3

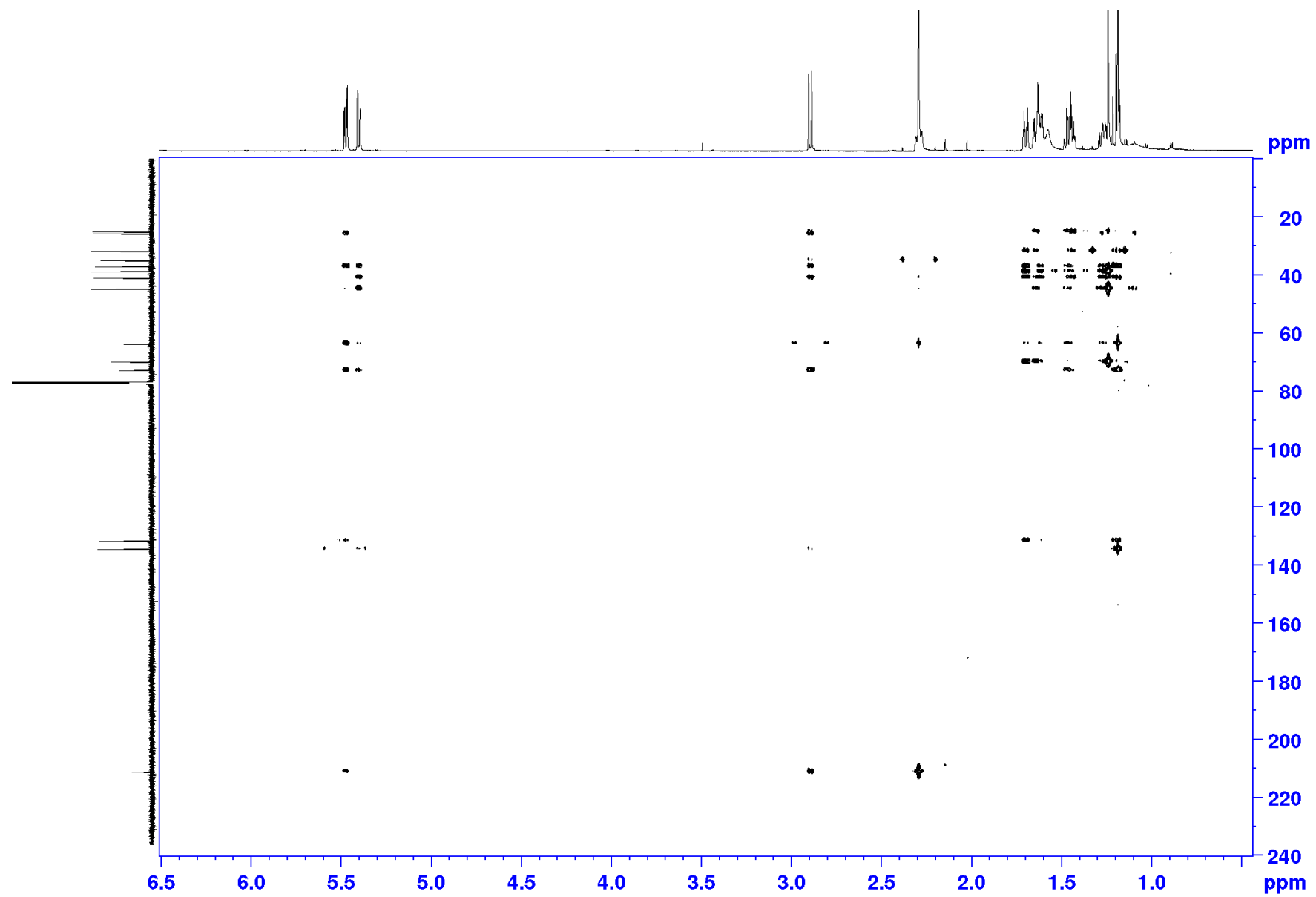


Figure S39. ROESY spectrum of 5 measured in CDCl₃

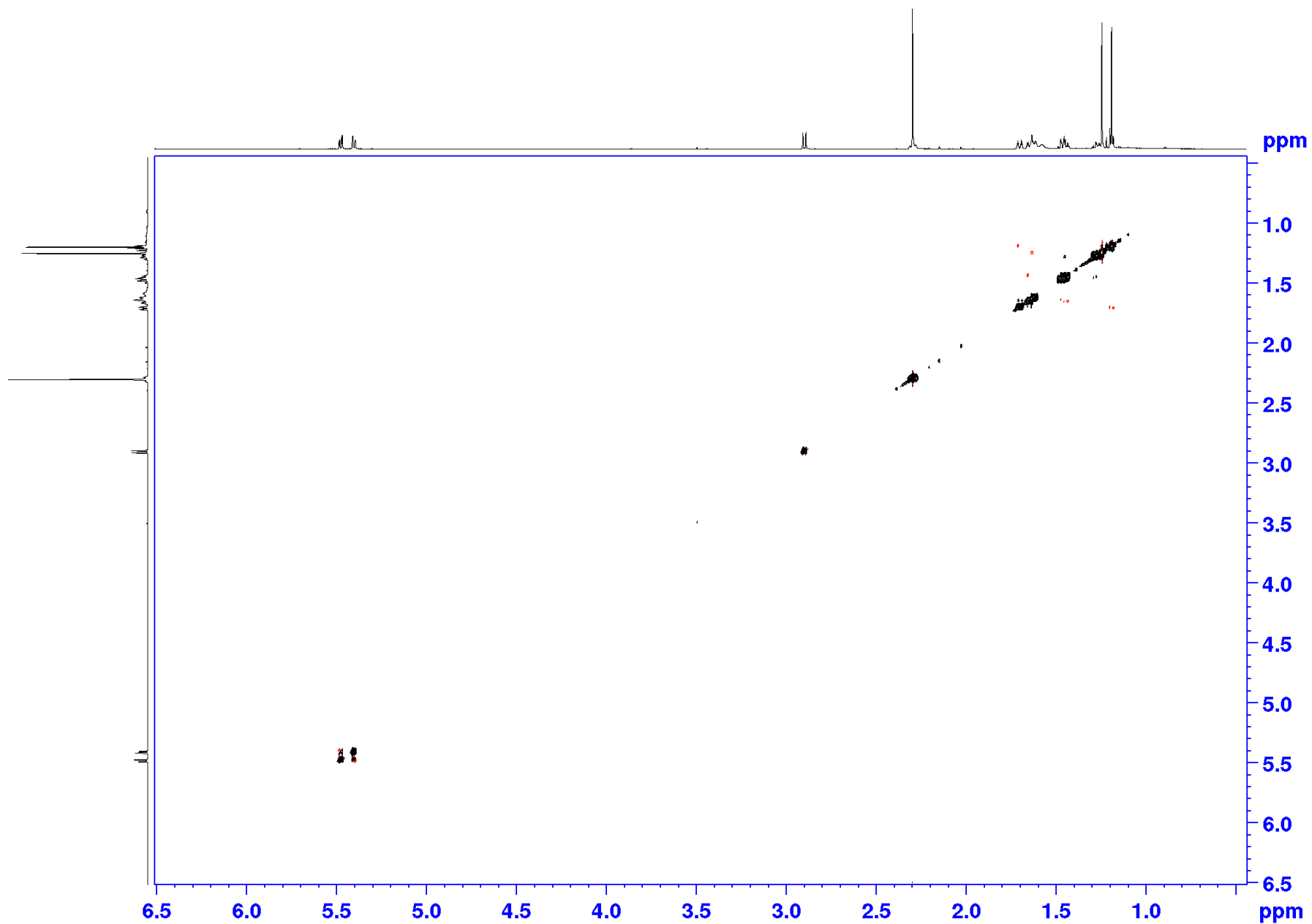
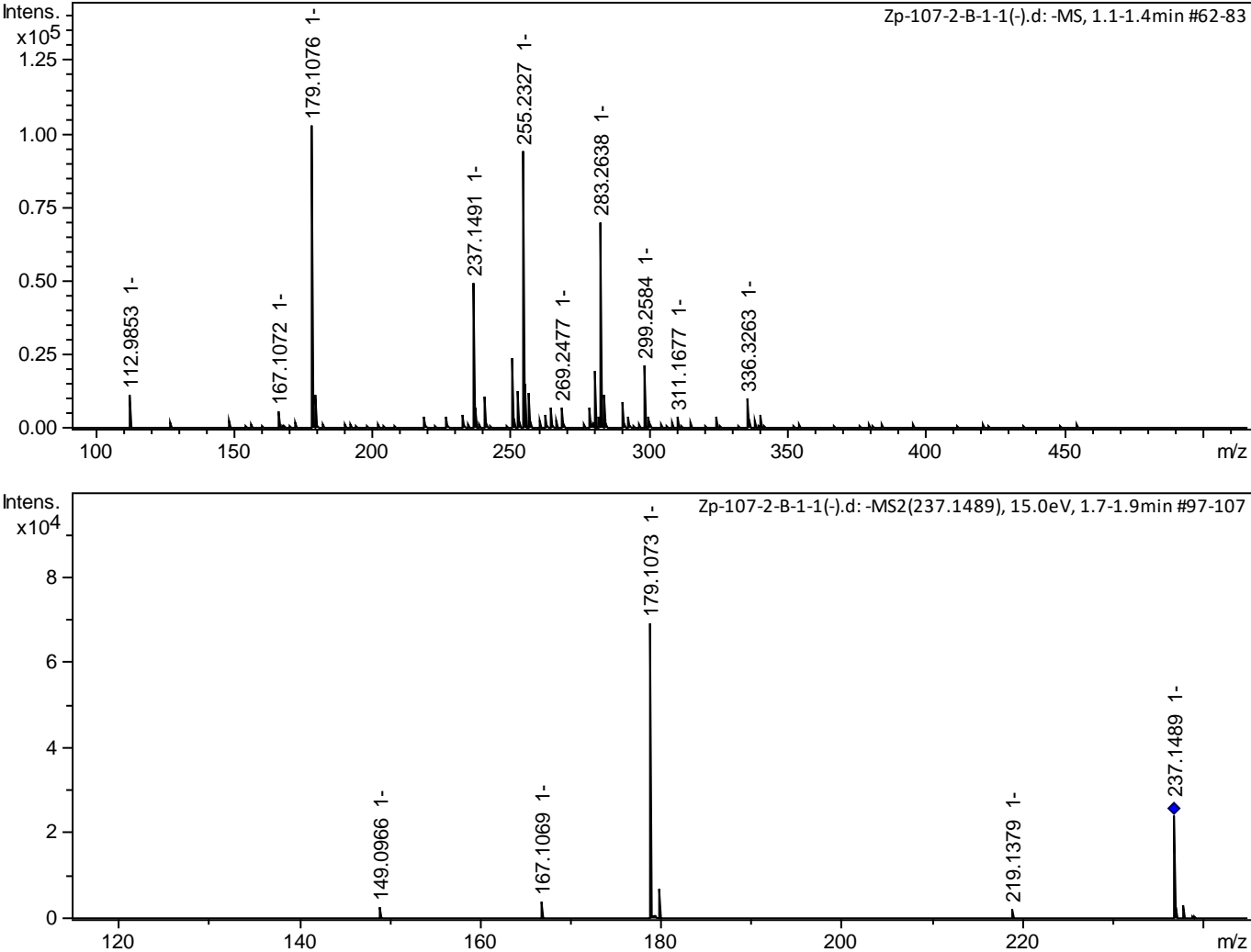
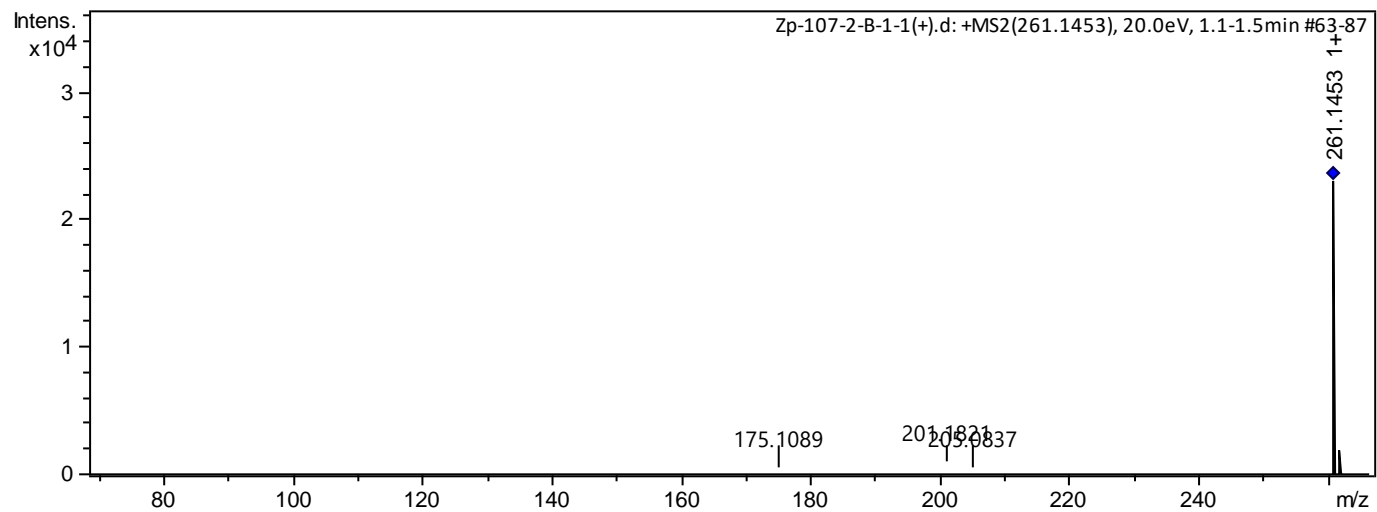
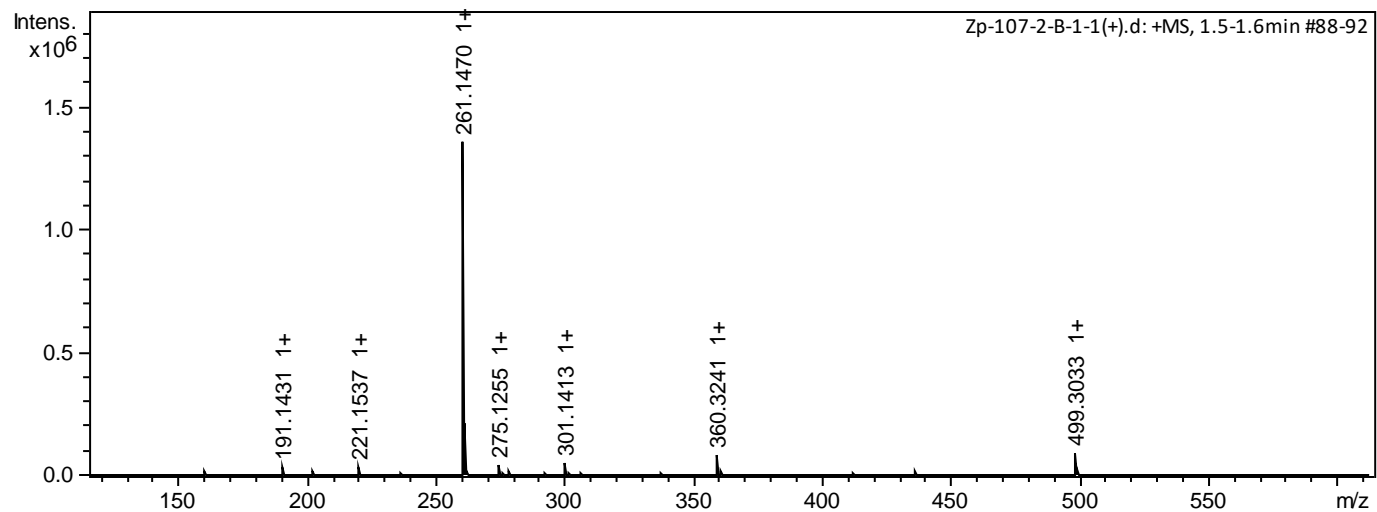


Figure S40. HRESIMS for 6





	meas	calc	Δ (ppm)
[M-H] ⁻	237,1491	237,1496	2,1
[M+Na] ⁺	261,147	261,1461	-3,4

Figure S41. ^1H NMR spectrum of **6** measured at 700 MHz in CDCl_3

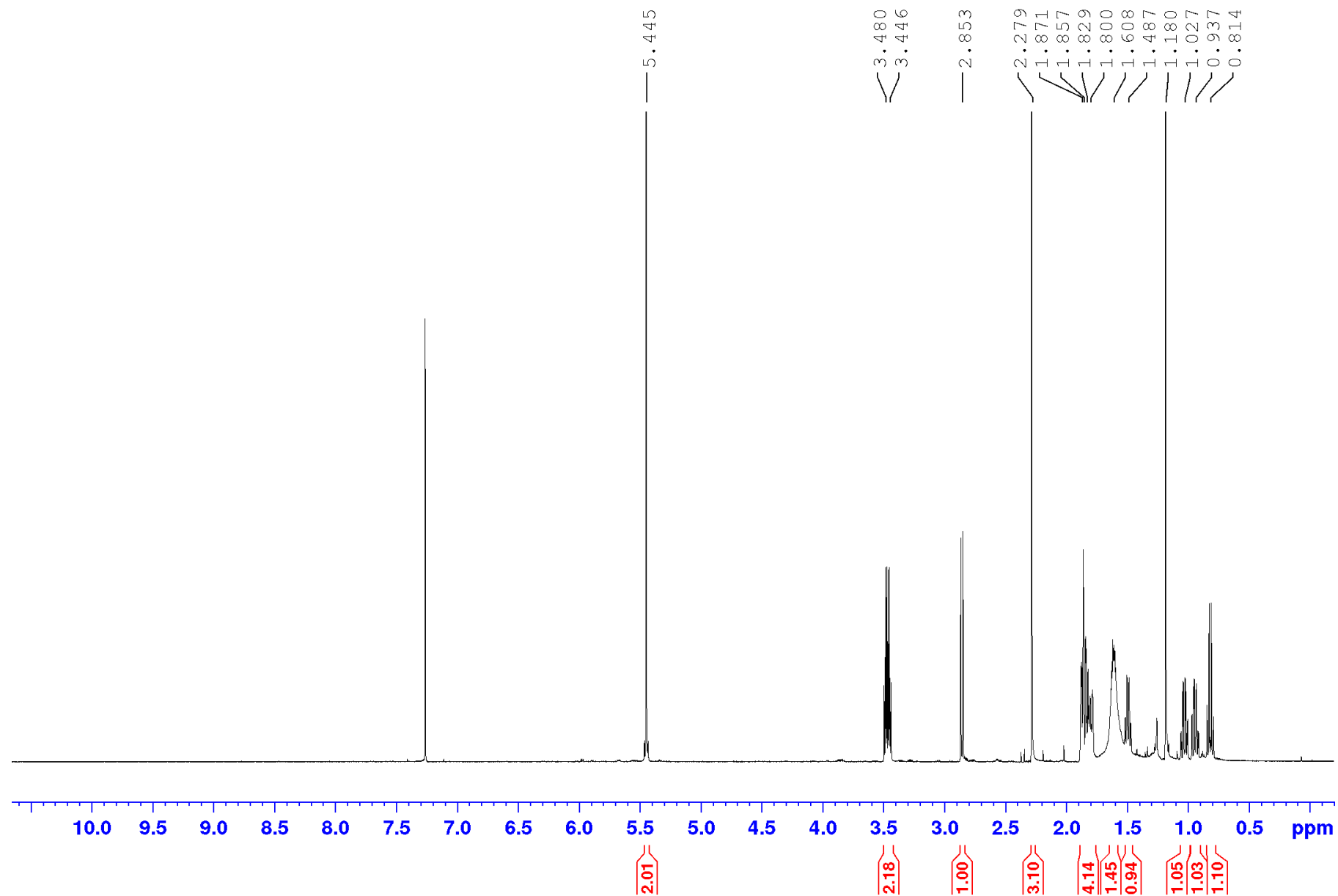


Figure S42. ^{13}C NMR spectrum of 6 measured at 176 MHz in CDCl_3

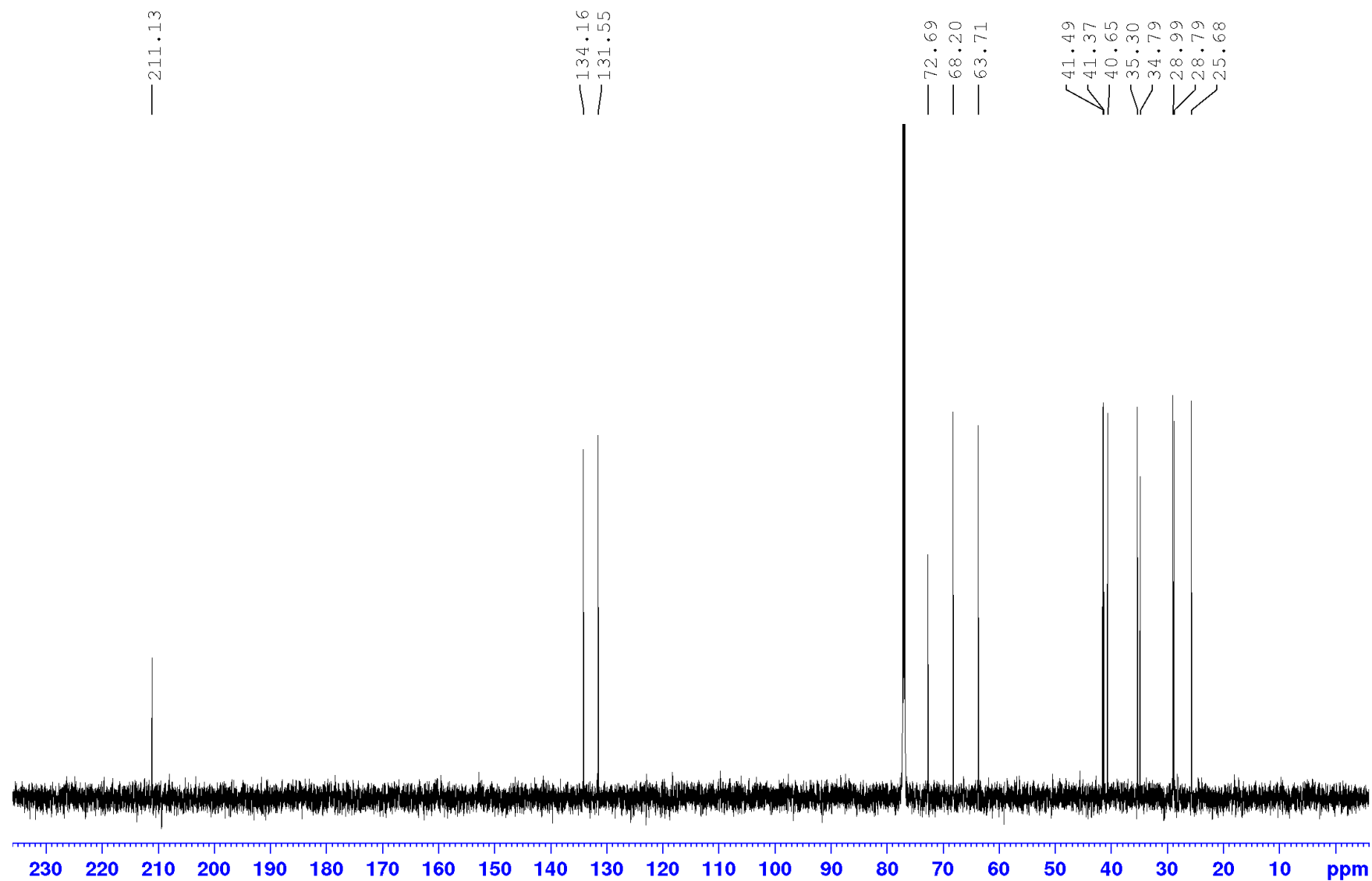


Figure S43. DEPT-135 spectrum of **6** measured at 176 MHz in CDCl₃

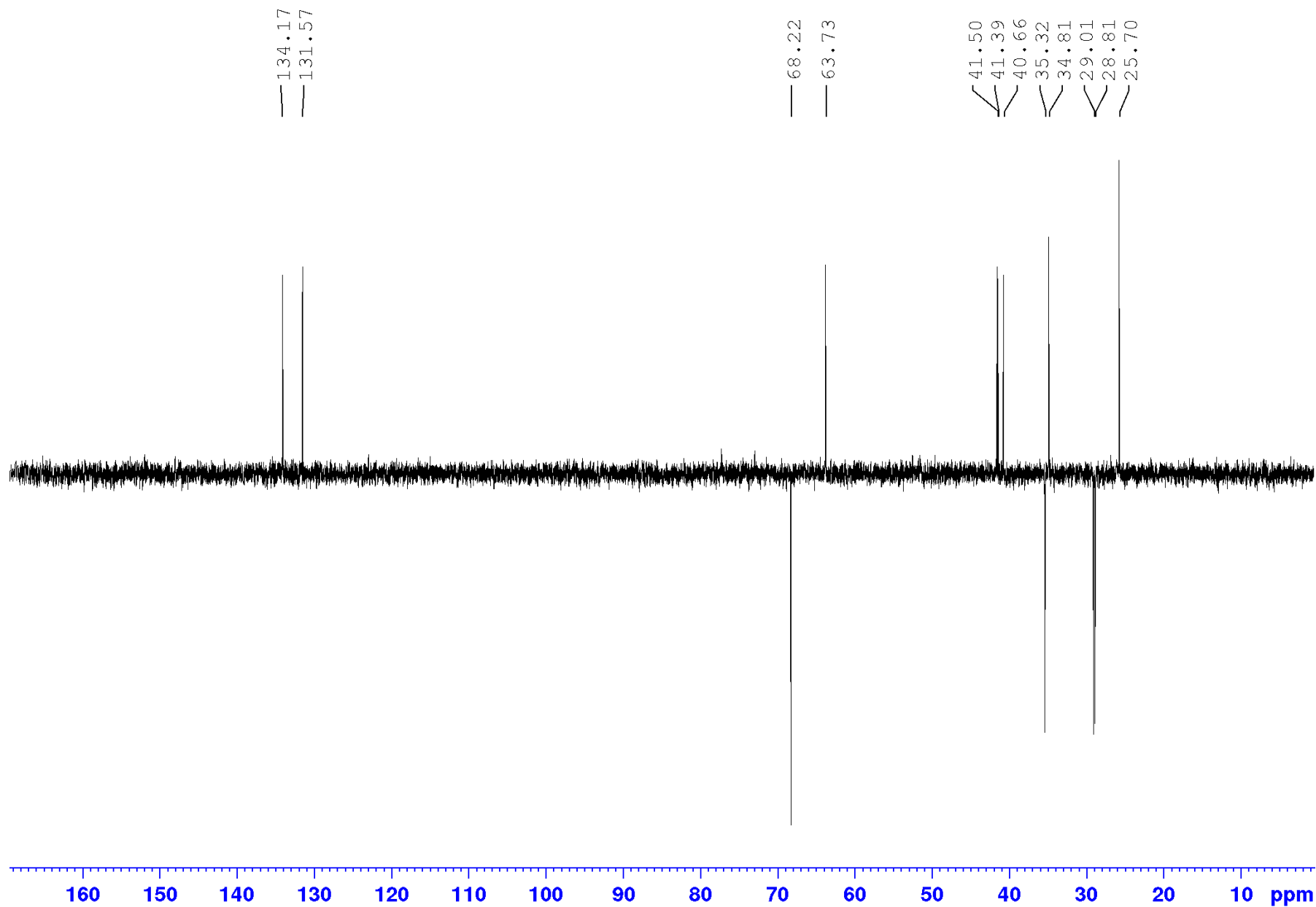


Figure S44. HSQC spectrum of 6 measured in CDCl₃

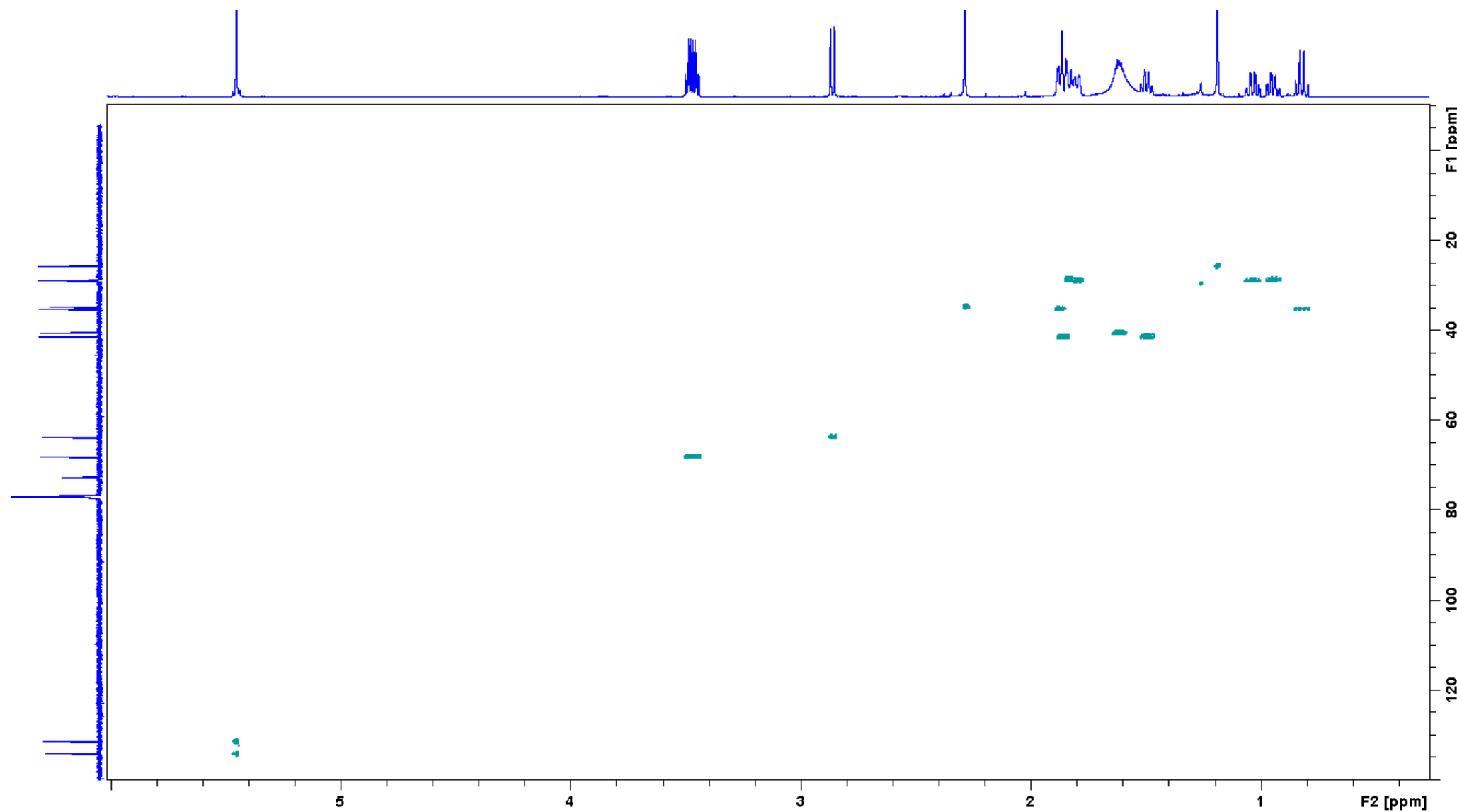


Figure S45. COSY spectrum of 6 measured in CDCl₃

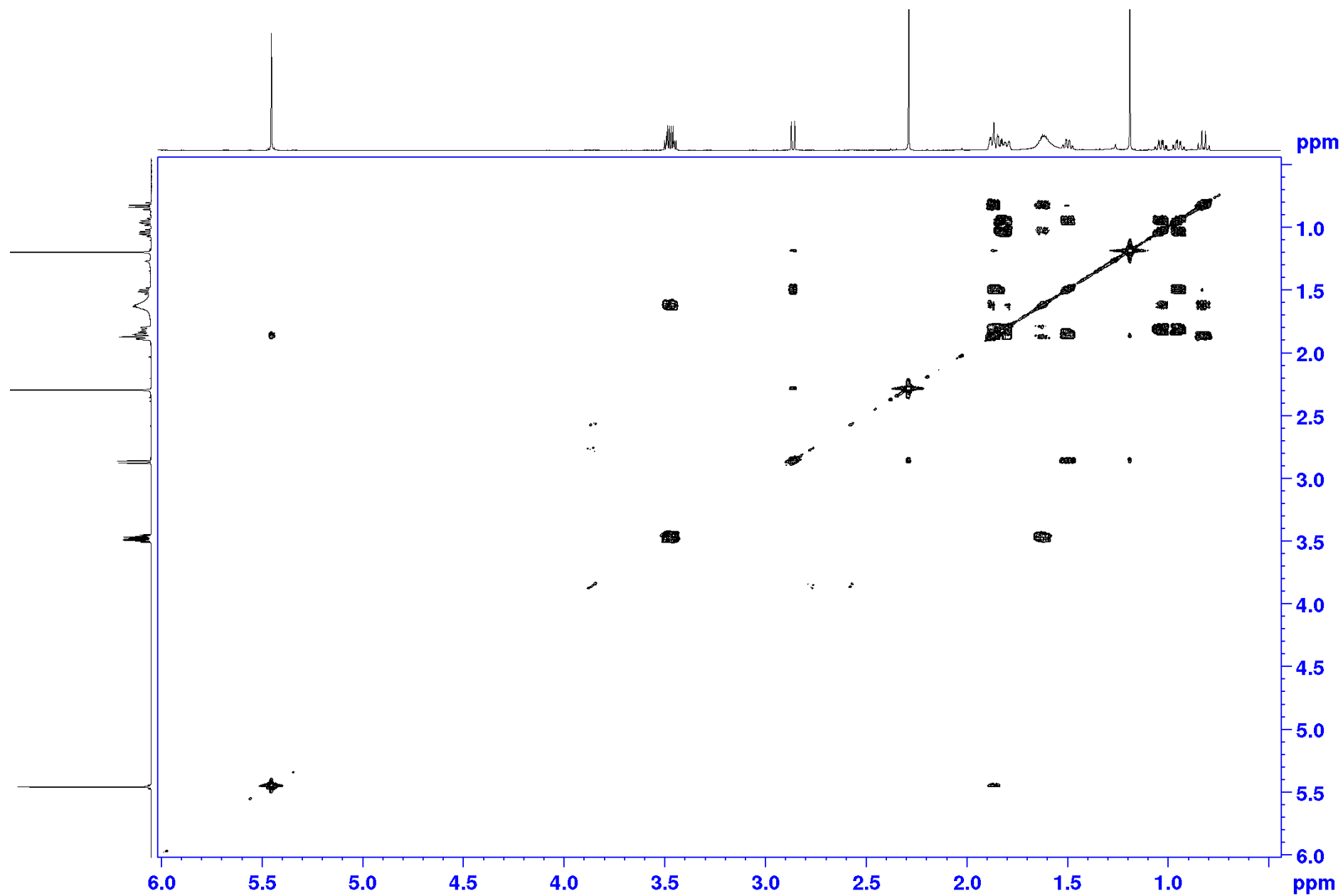


Figure S46. HMBC spectrum of 6 measured in CDCl₃

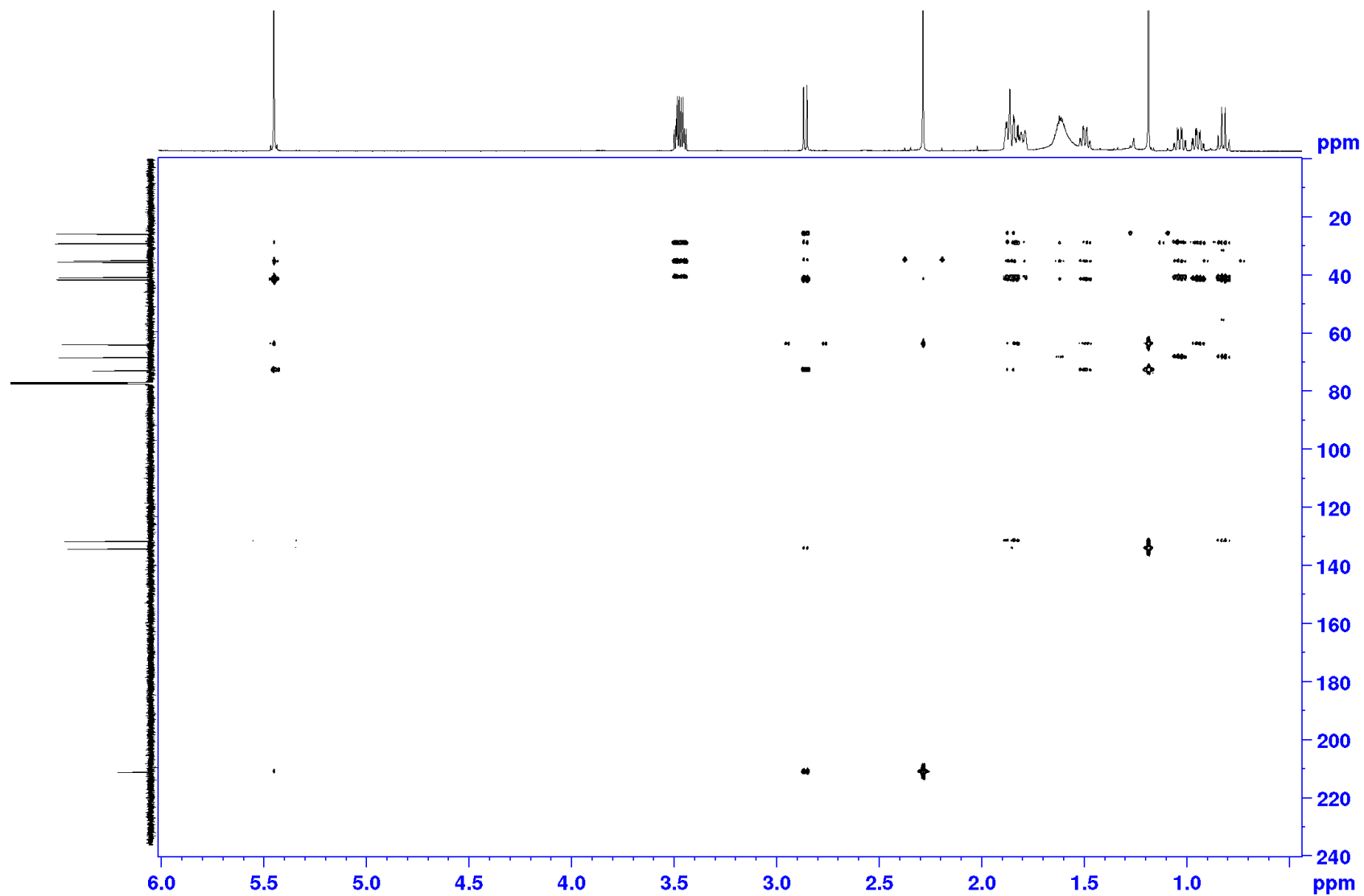


Figure S47. ROESY spectrum of 6 measured in CDCl₃

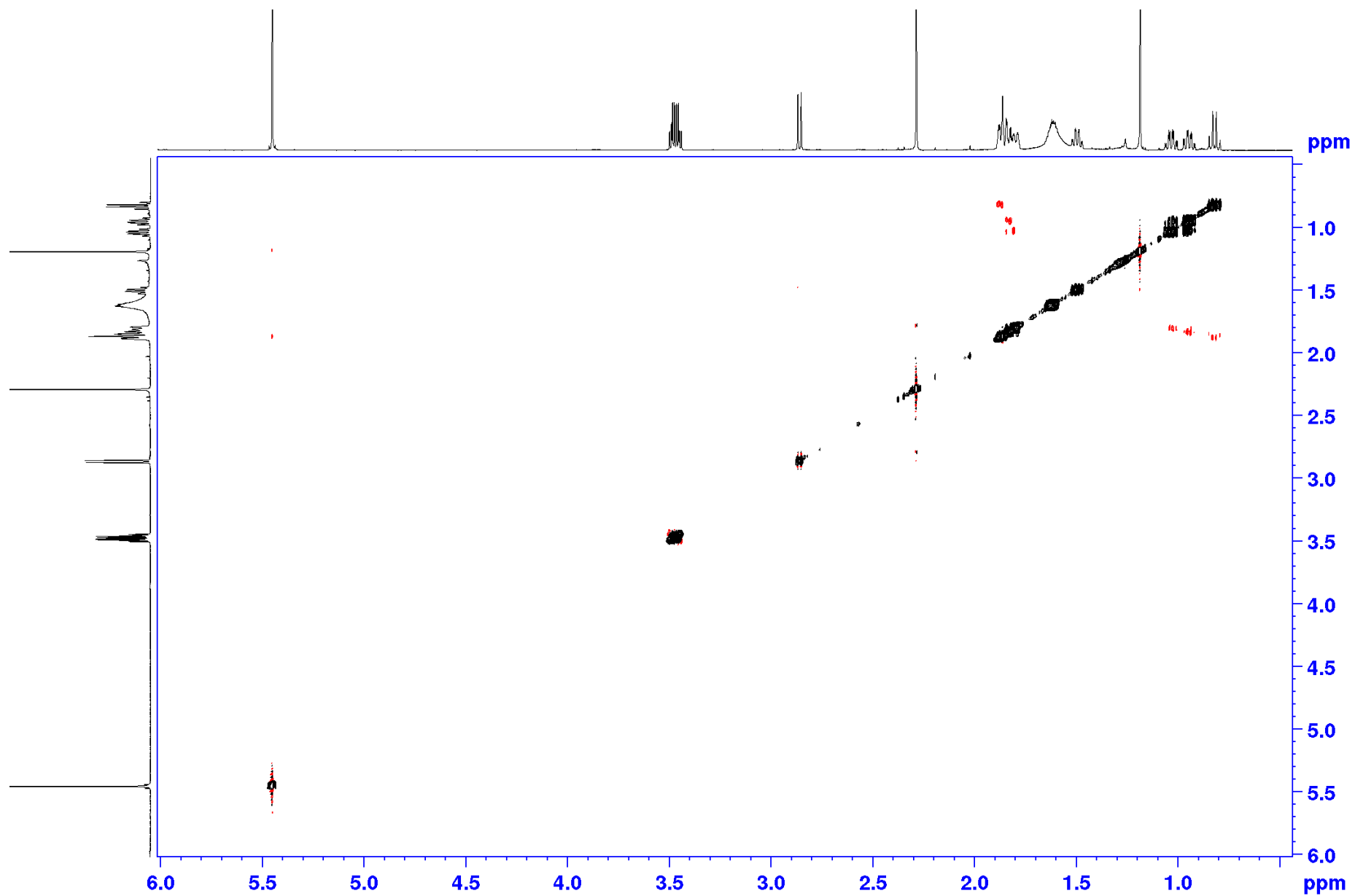
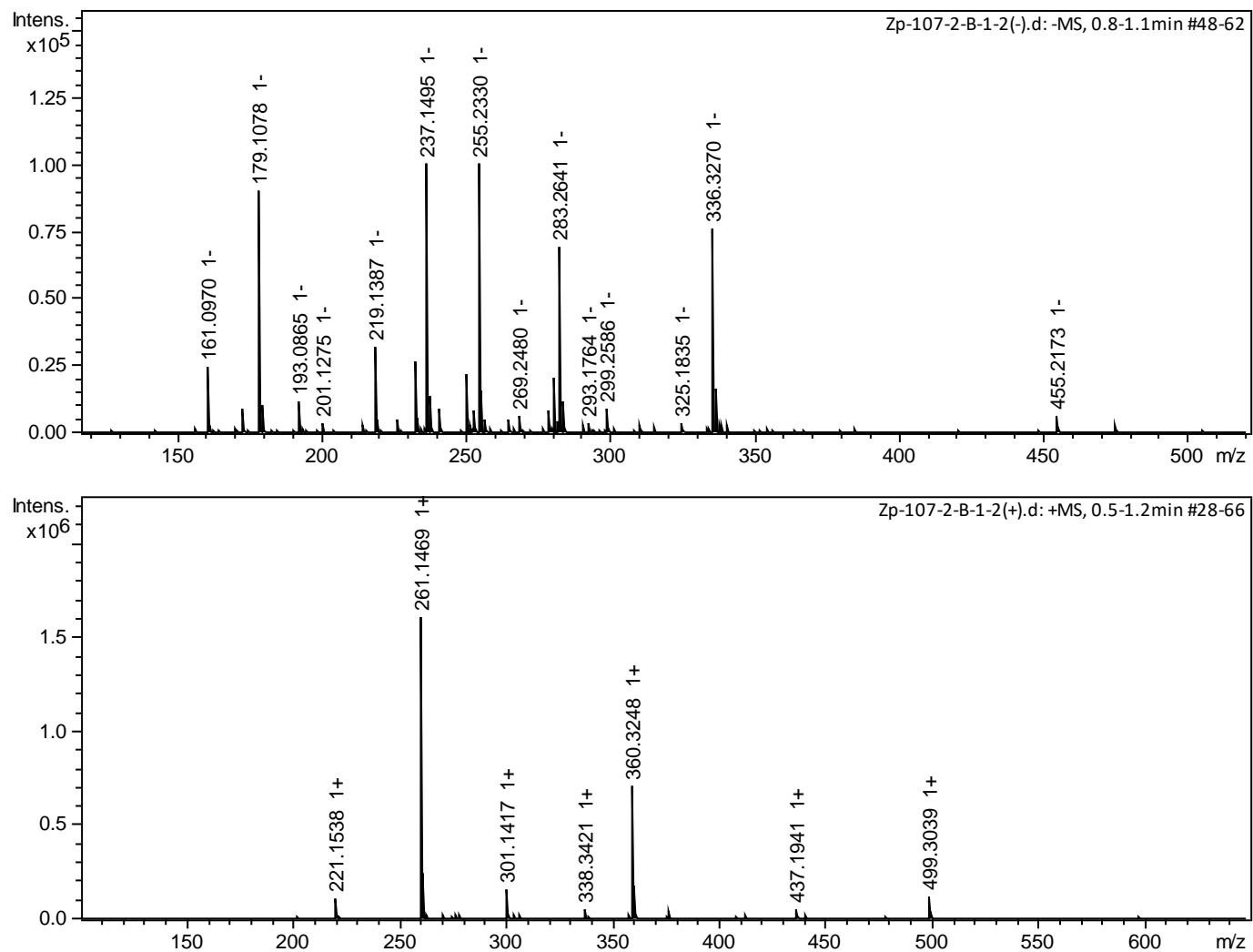
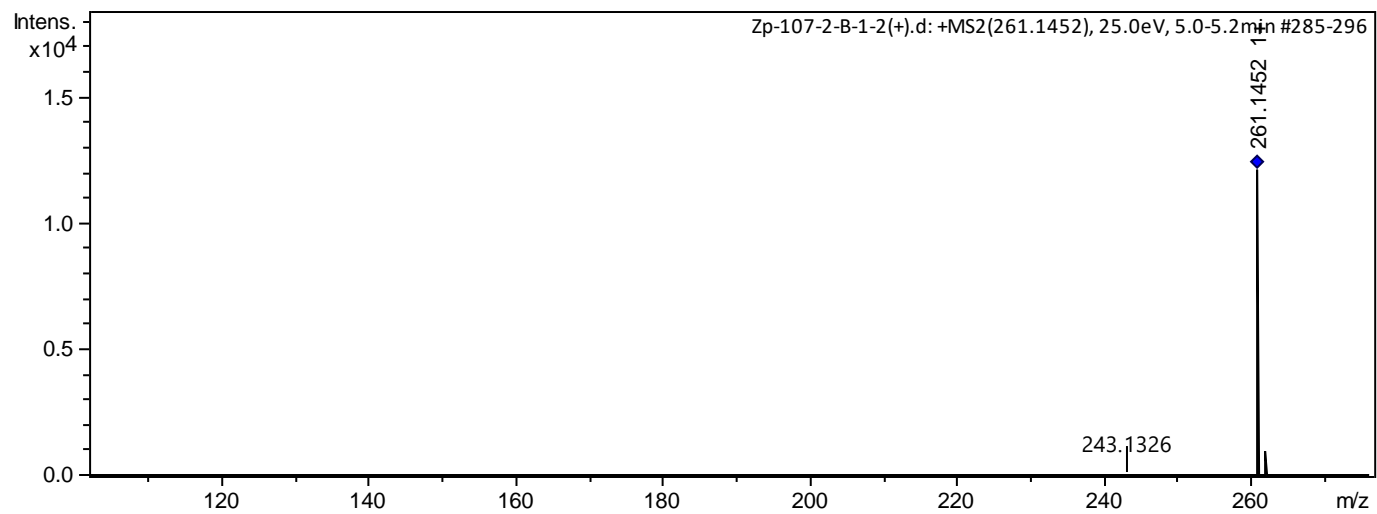


Figure S48. HRESIMS for 7





	meas	calc	Δ (ppm)
[M-H] ⁻	237,1495	237,1496	0,4
[M+Na] ⁺	261,1469	261,1461	-3,1

Figure S49. ^1H NMR spectrum of 7 measured at 700 MHz in CDCl_3

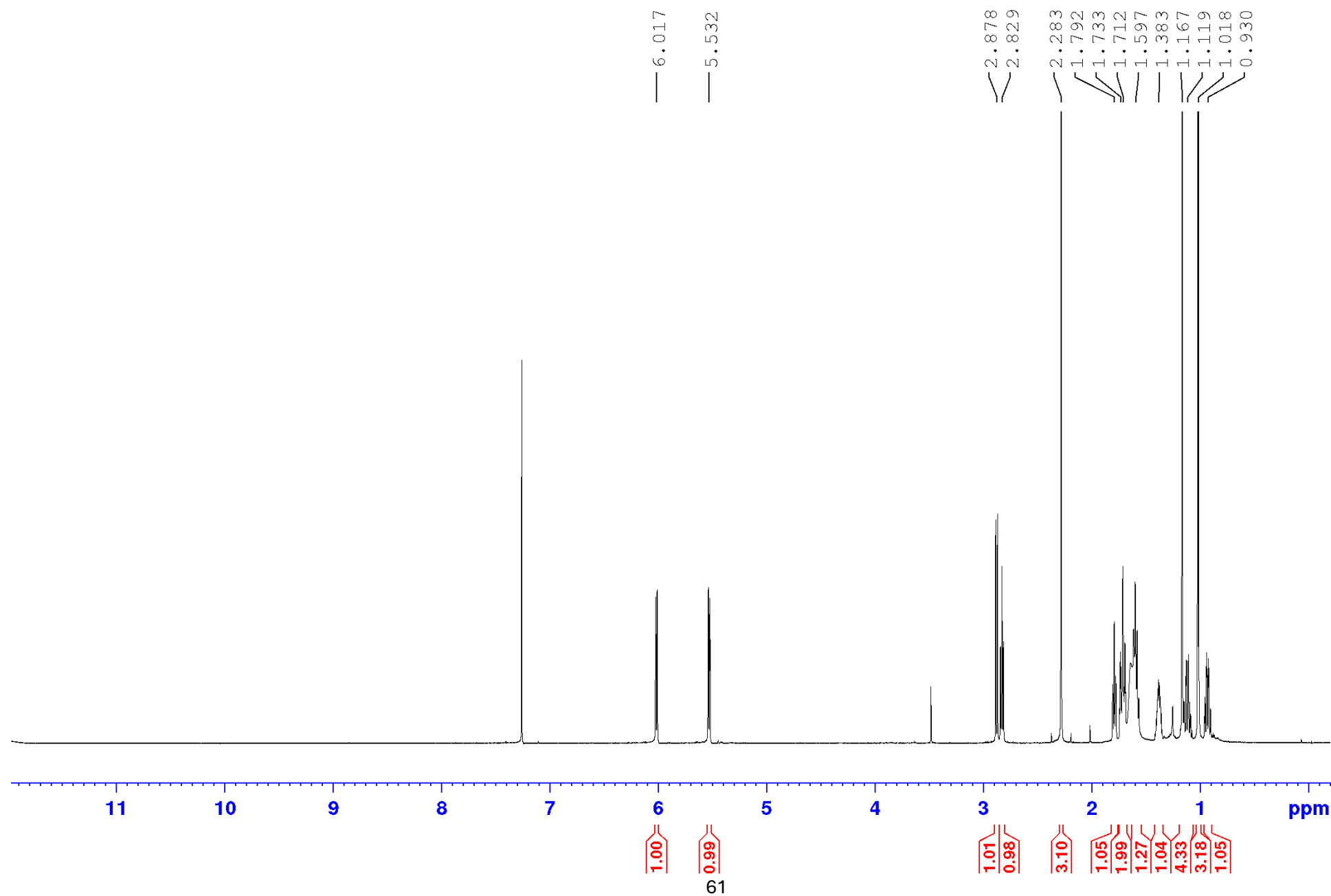


Figure S50. ^{13}C NMR spectrum of 7 measured at 176 MHz in CDCl_3

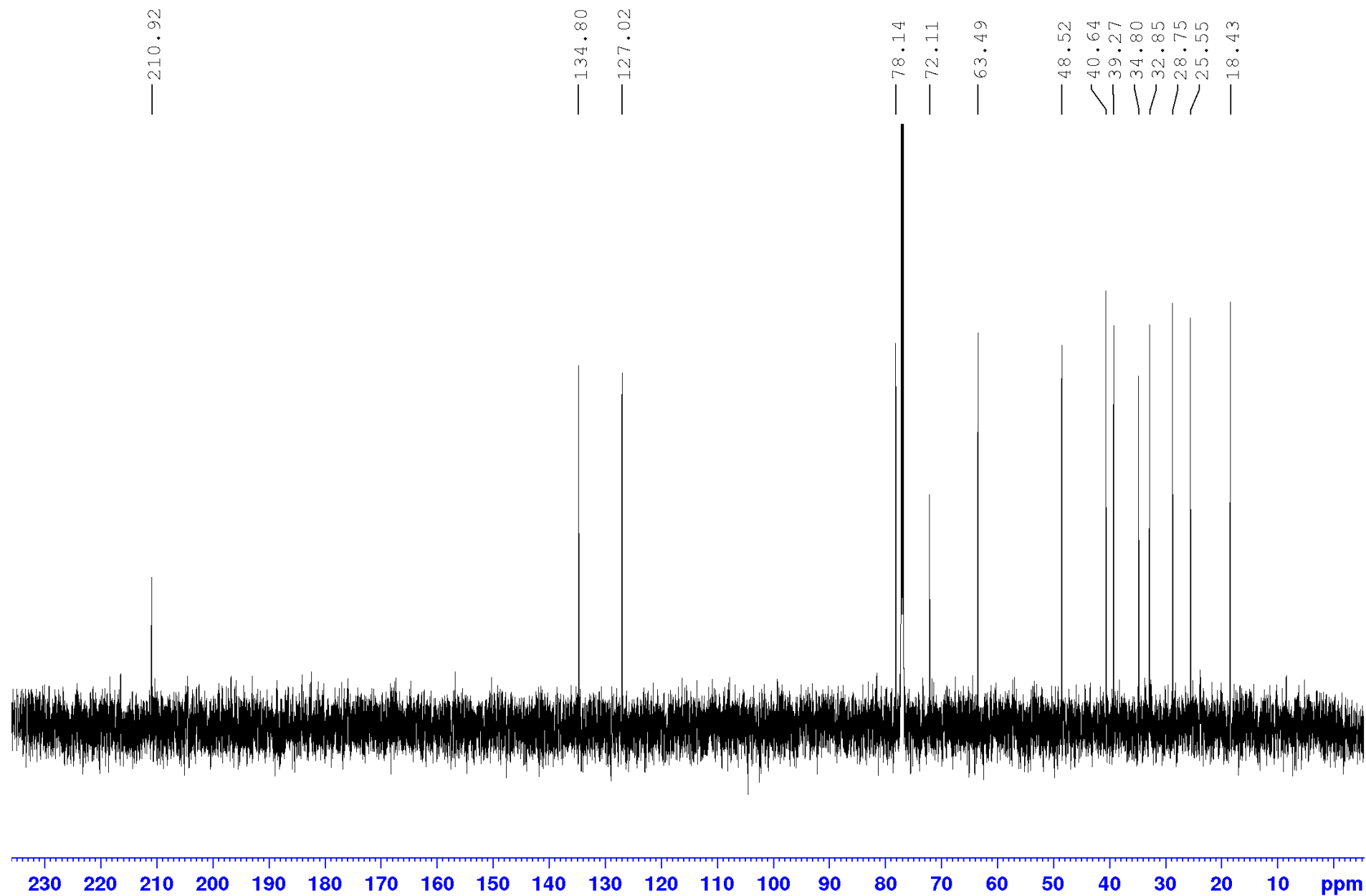


Figure S51. DEPT-135 spectrum of **7** measured at 176 MHz in CDCl₃

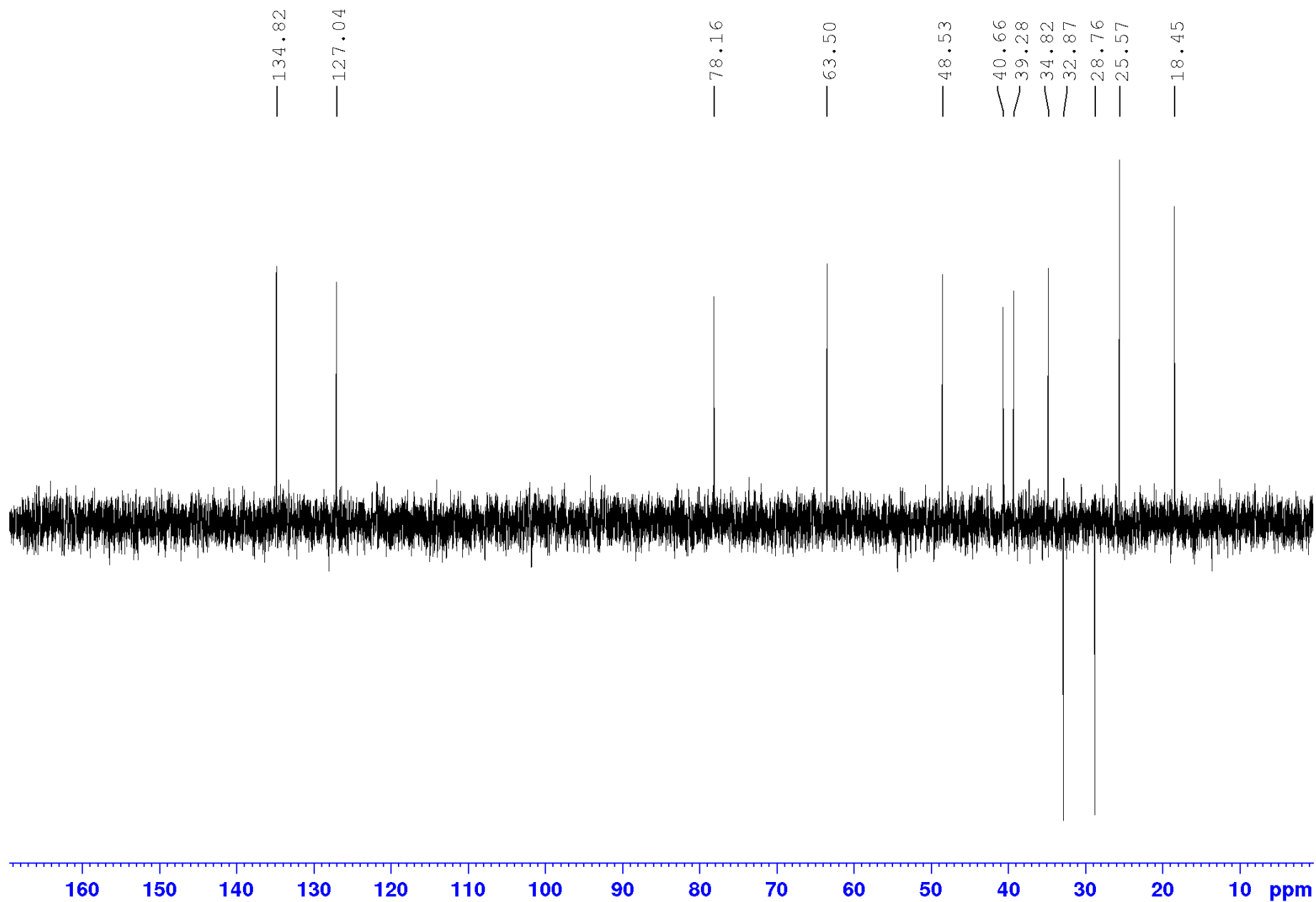


Figure S52. HSQC spectrum of 7 measured in CDCl₃

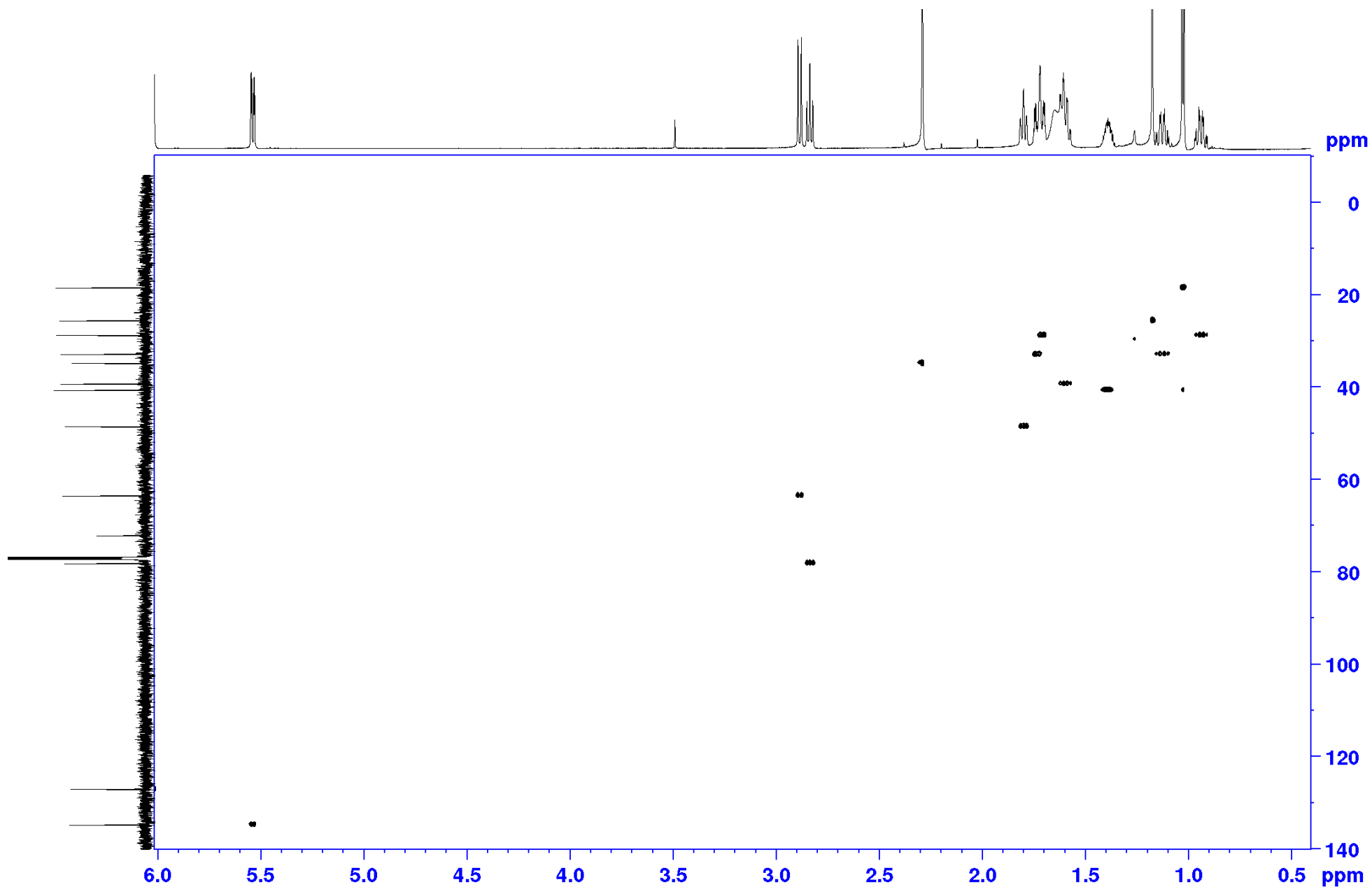


Figure S53. COSY spectrum of 7 measured in CDCl₃

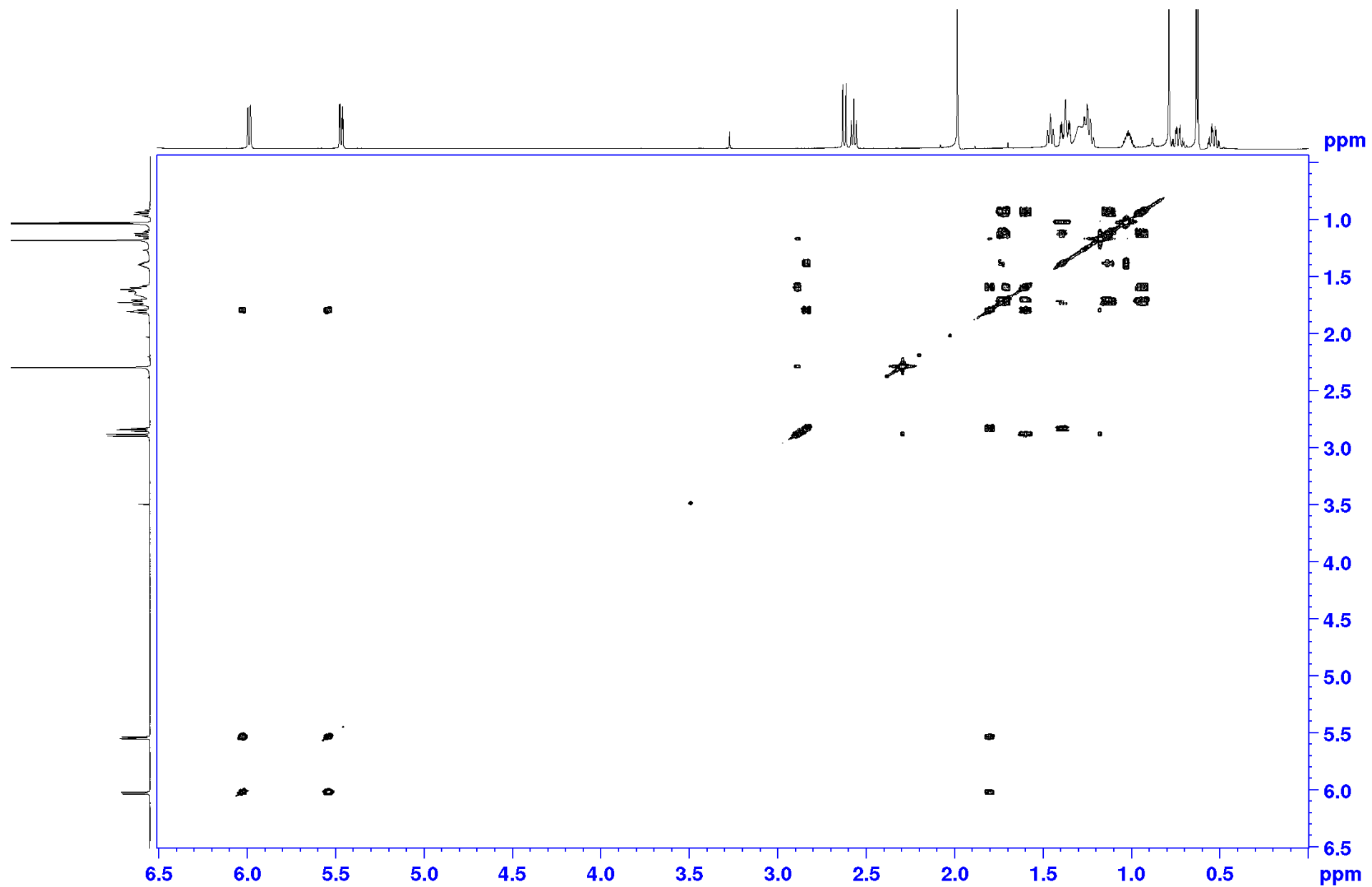


Figure S54. HMBC spectrum of 7 measured in CDCl_3

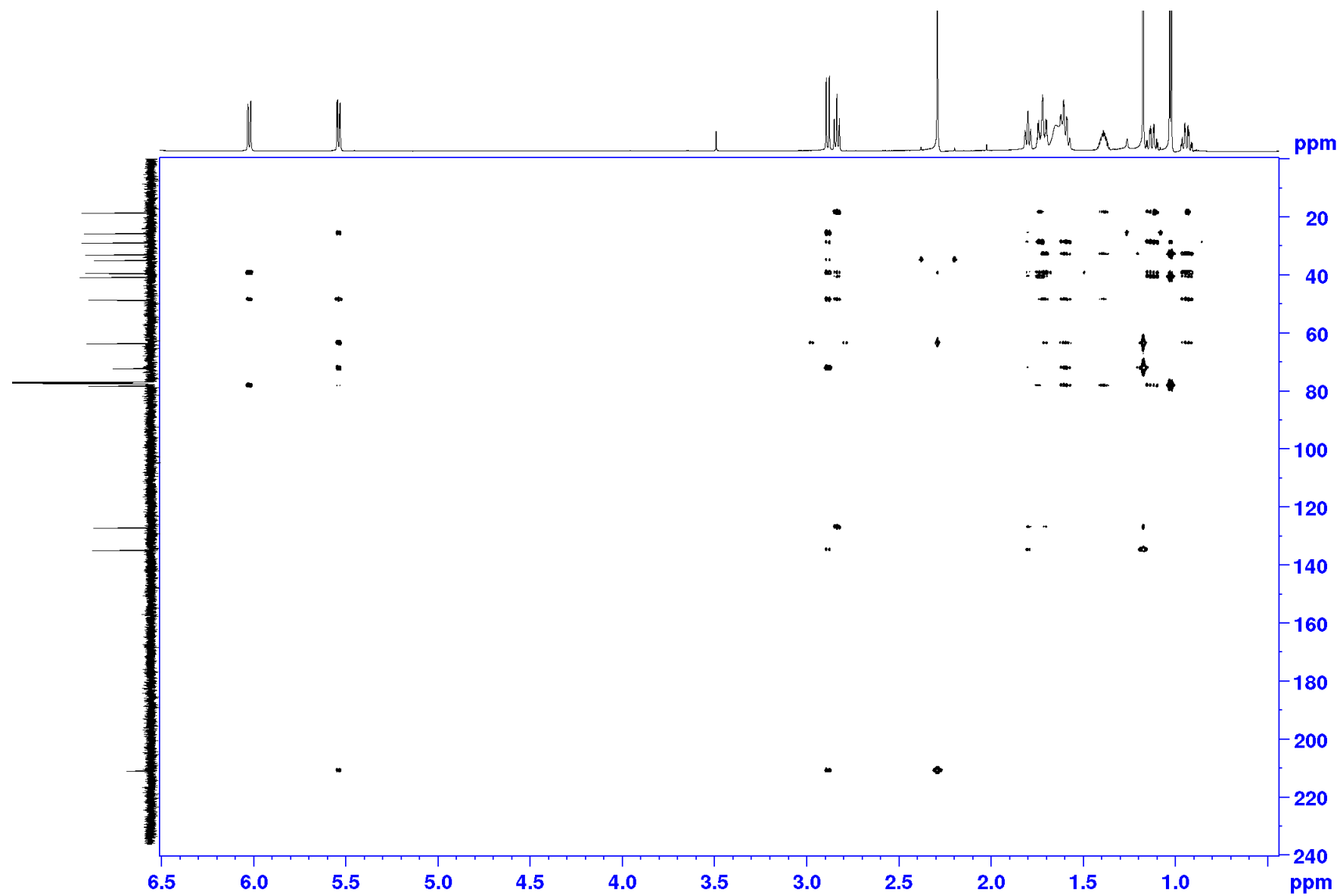


Figure S55. ROESY spectrum of 7 measured in CDCl₃

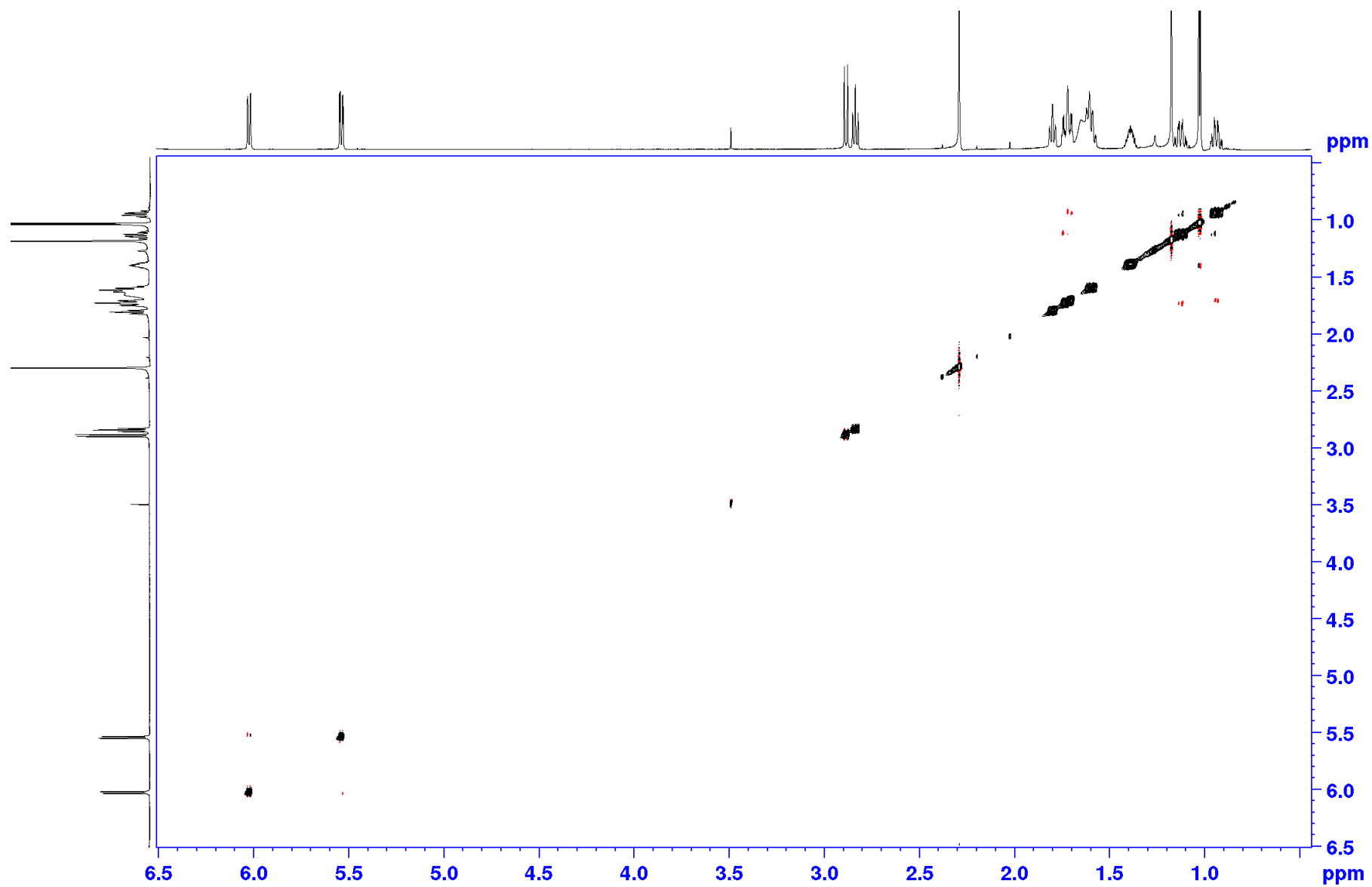


Figure S56. HRESIMS for 8

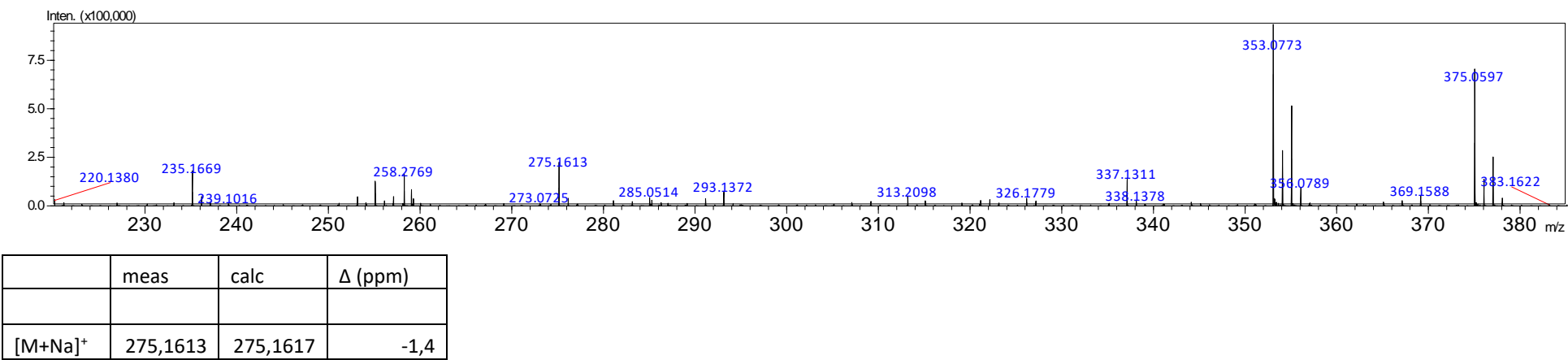


Figure S57. ^1H NMR spectrum of **8** measured at 700 MHz in CDCl_3

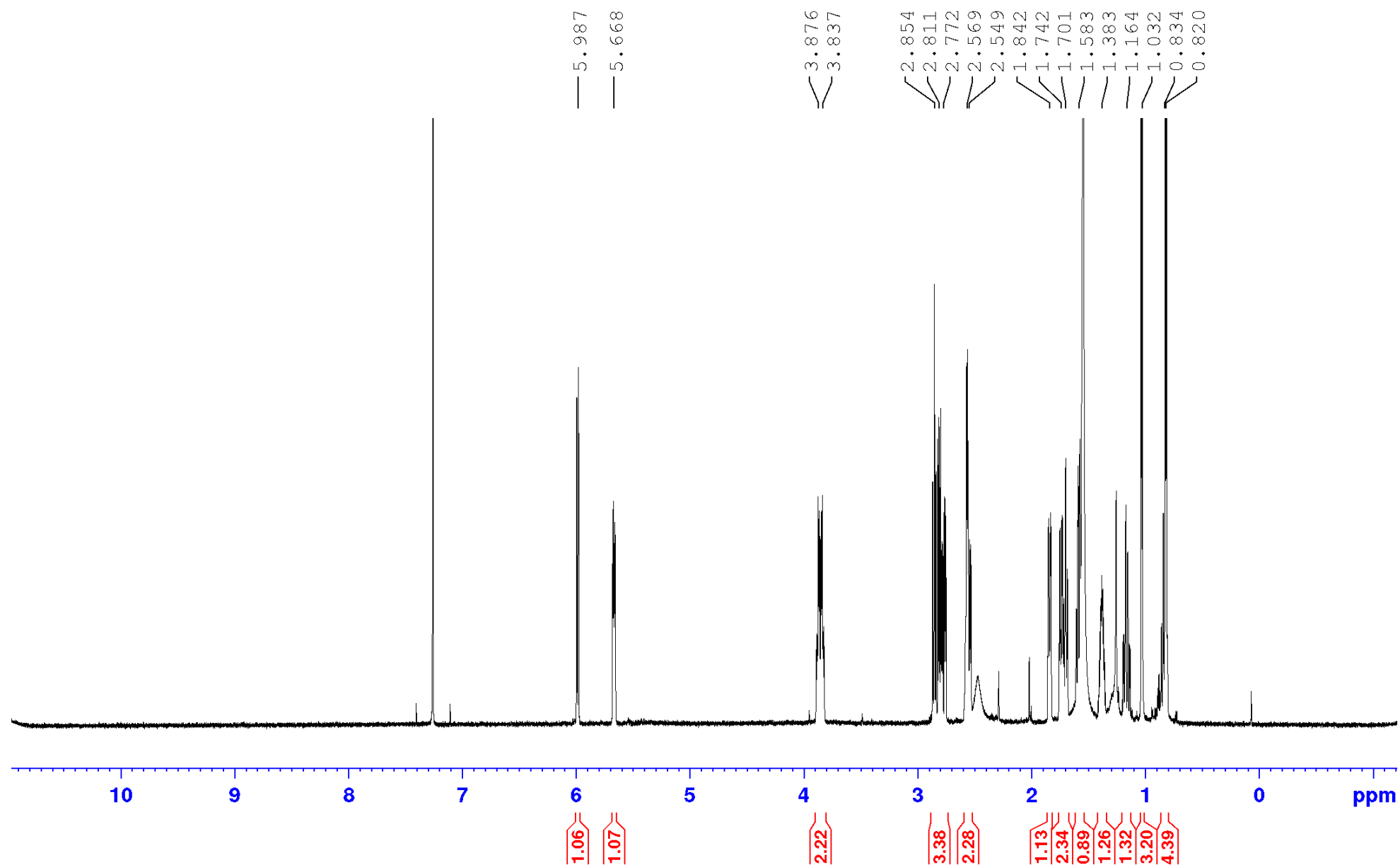


Figure S58. ^{13}C NMR spectrum of 8 measured at 176 MHz in CDCl_3

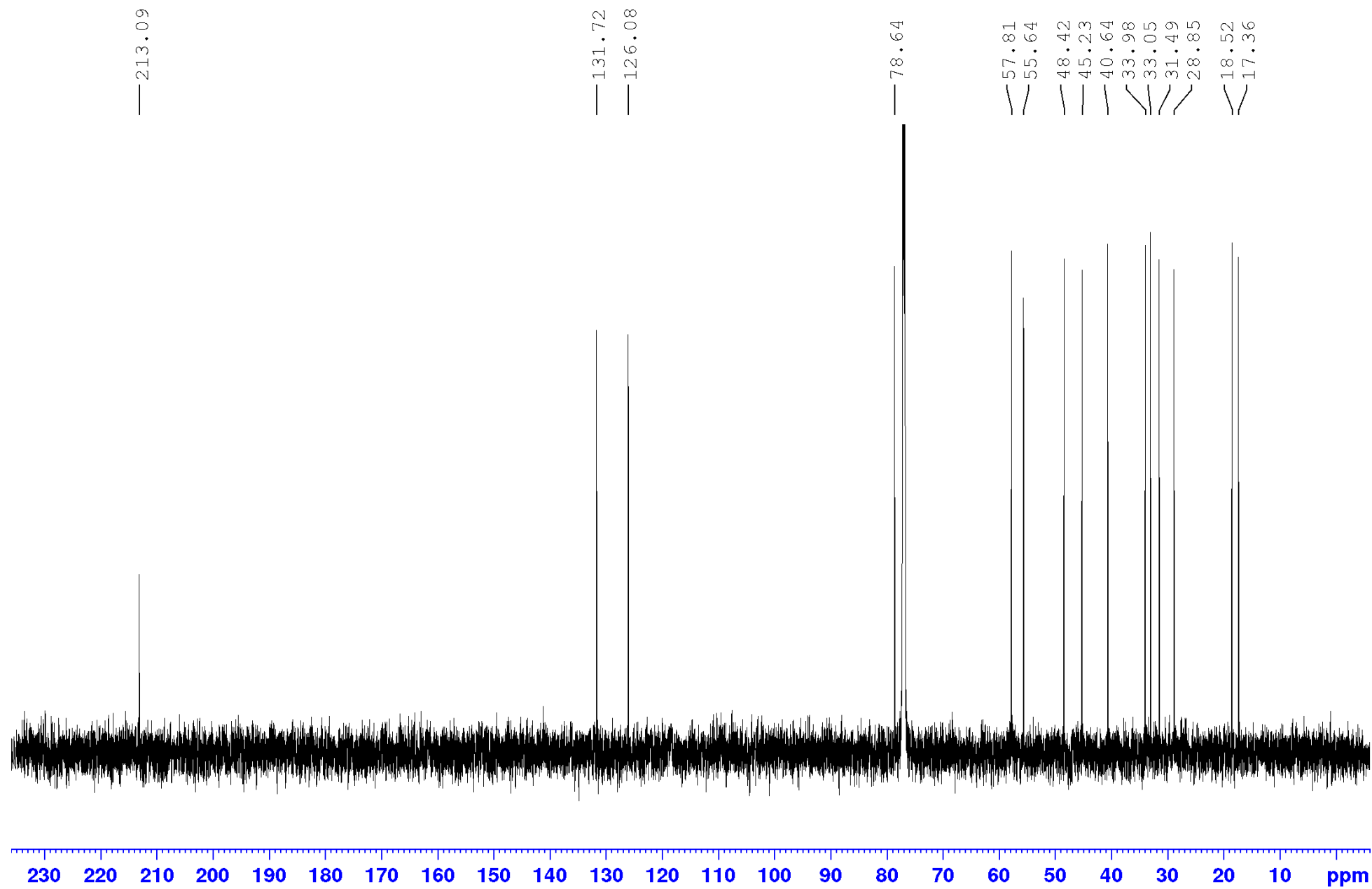


Figure S59. DEPT-135 spectrum of **8** measured at 176 MHz in CDCl₃

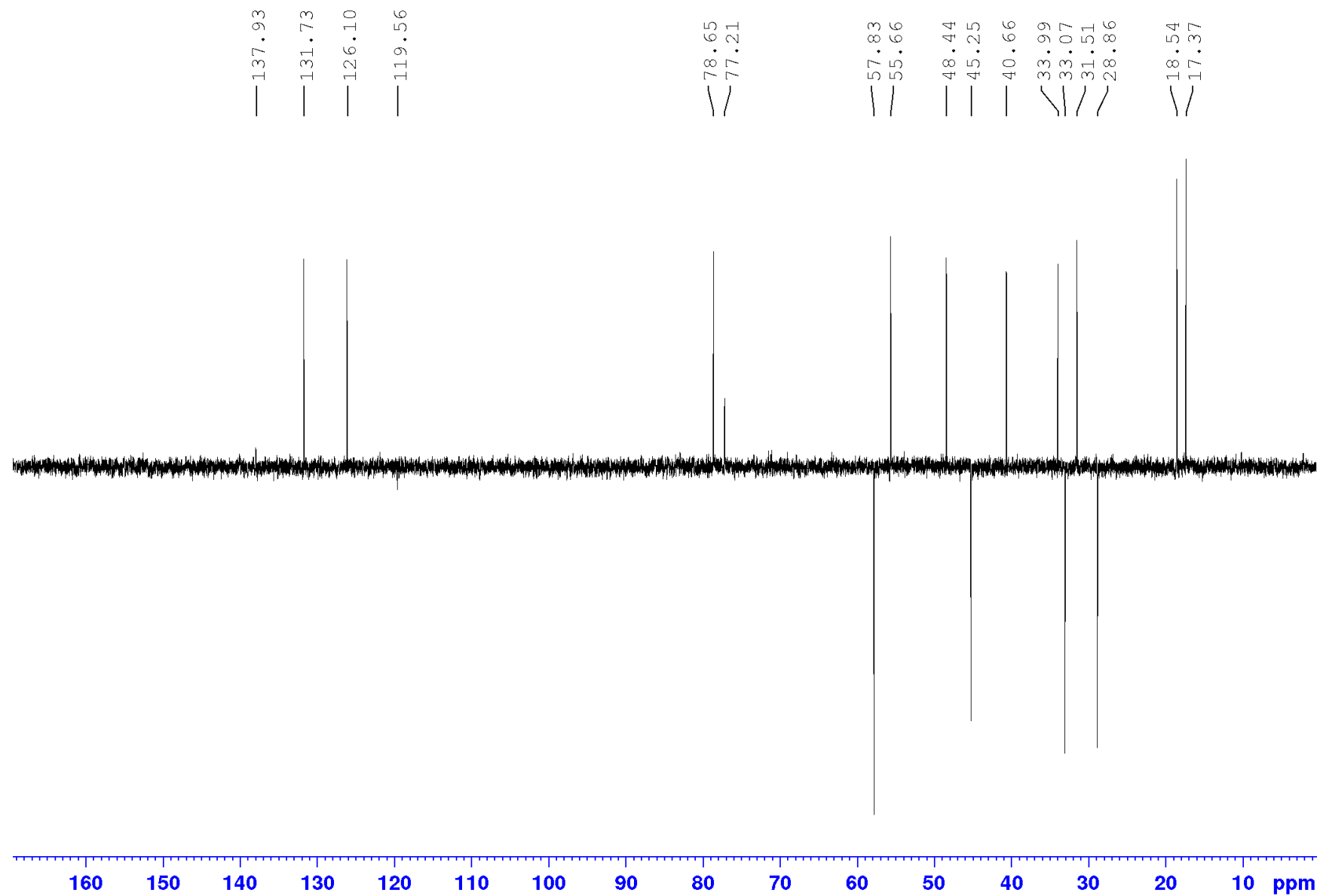


Figure S60. HSQC spectrum of 8 measured in CDCl₃

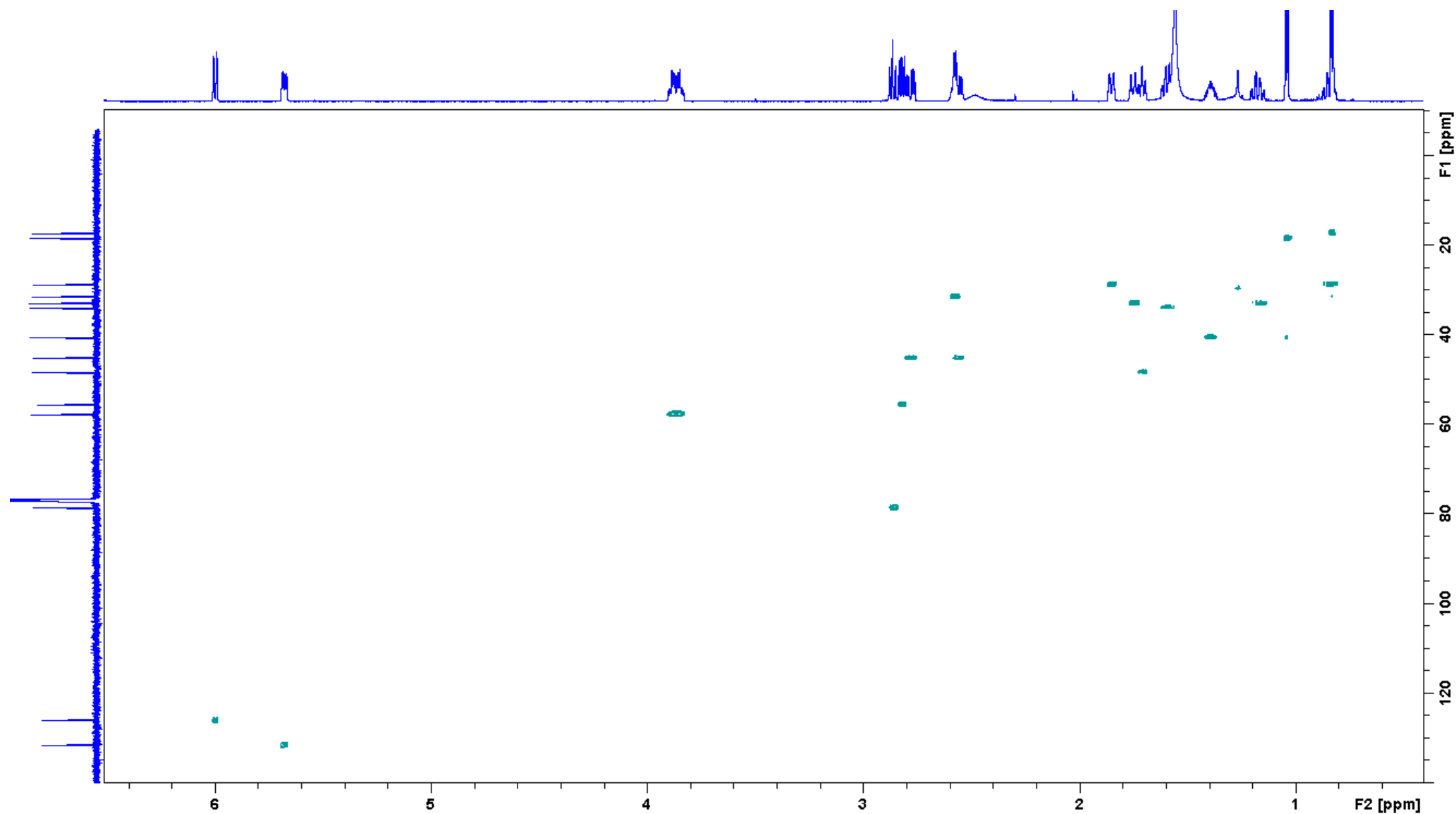


Figure S61. COSY spectrum of 8 measured in CDCl₃

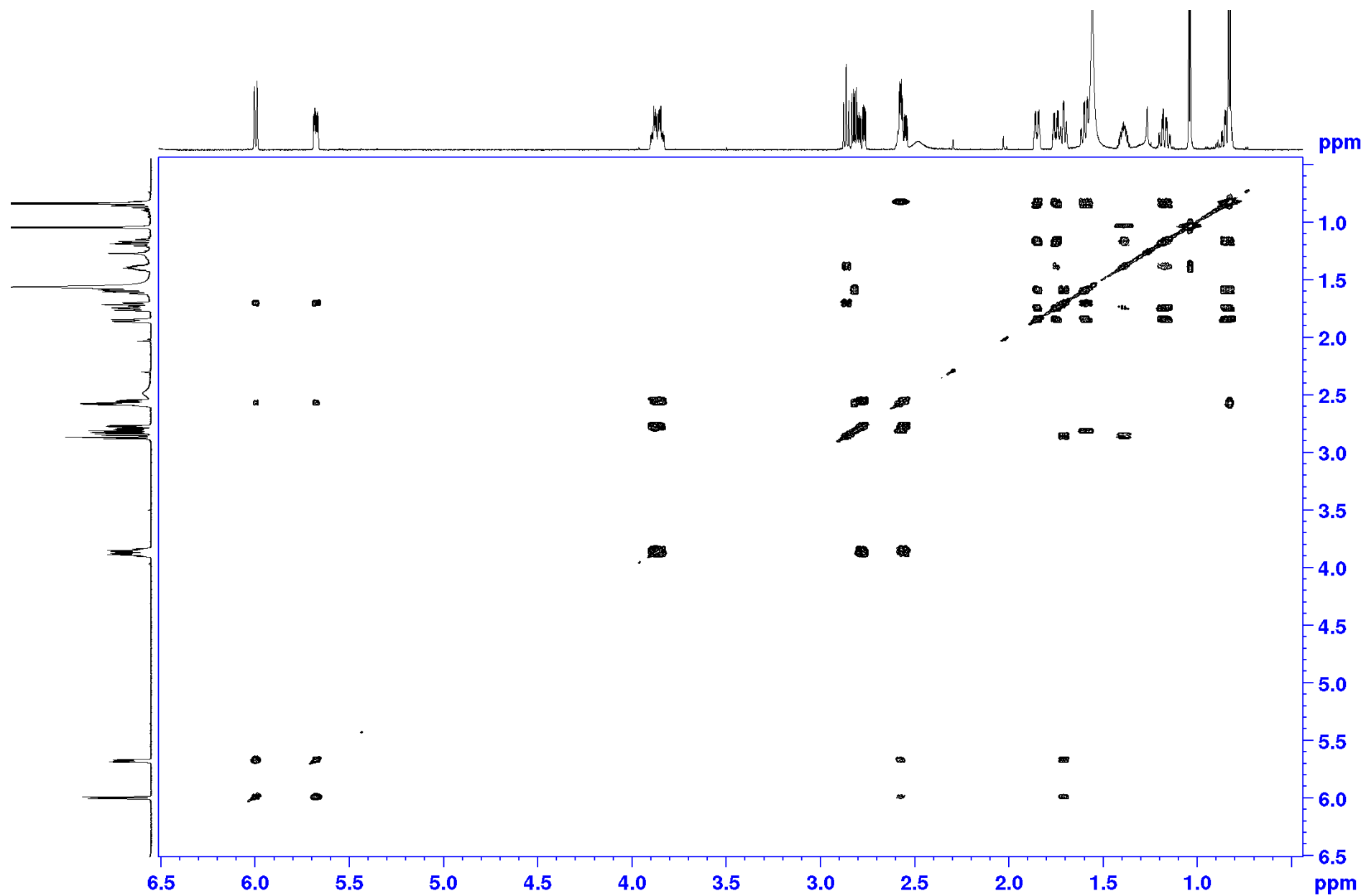


Figure S62. HMBC spectrum of 8 measured in CDCl₃

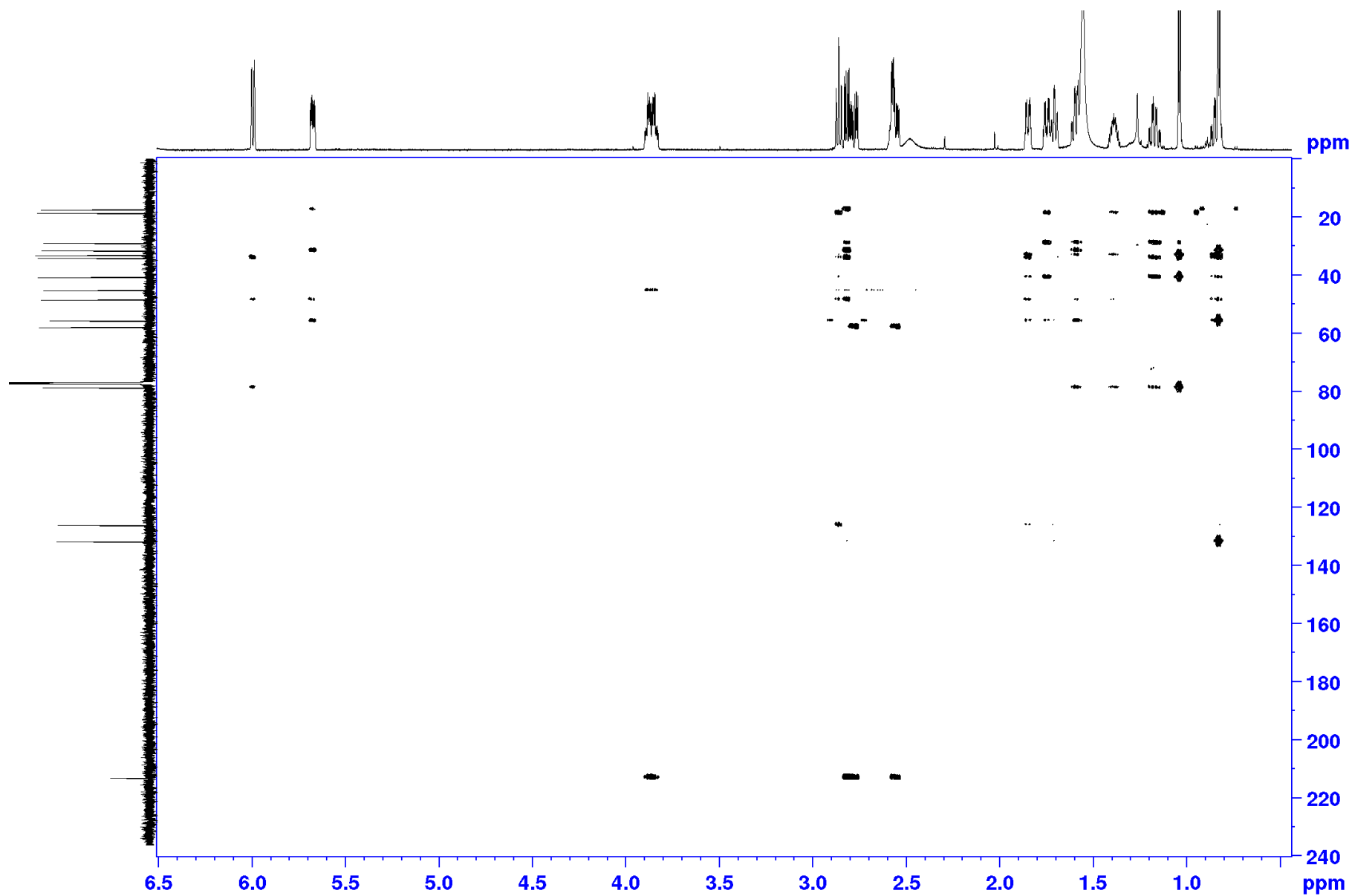


Figure S63. ROESY spectrum of 8 measured in CDCl₃

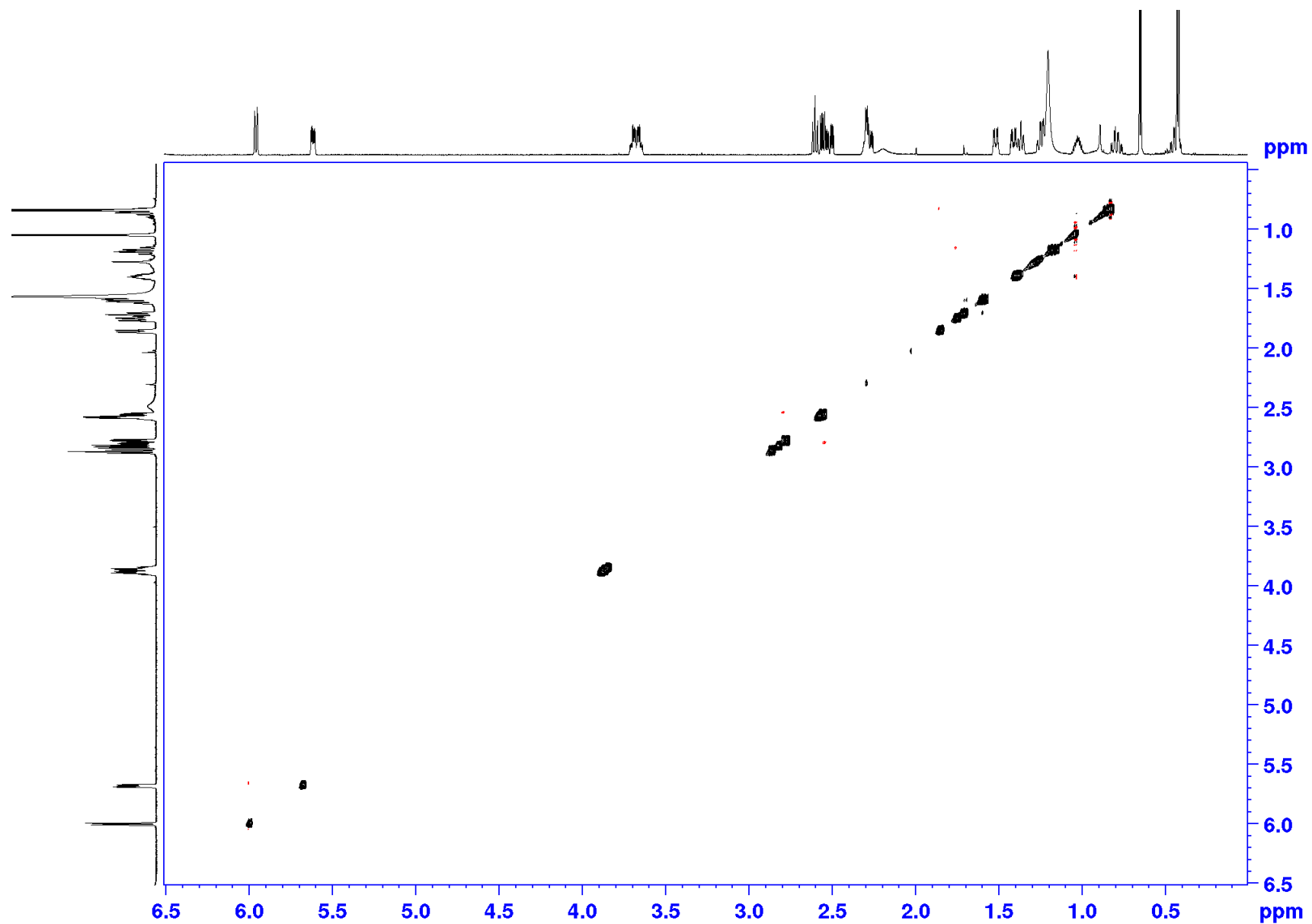
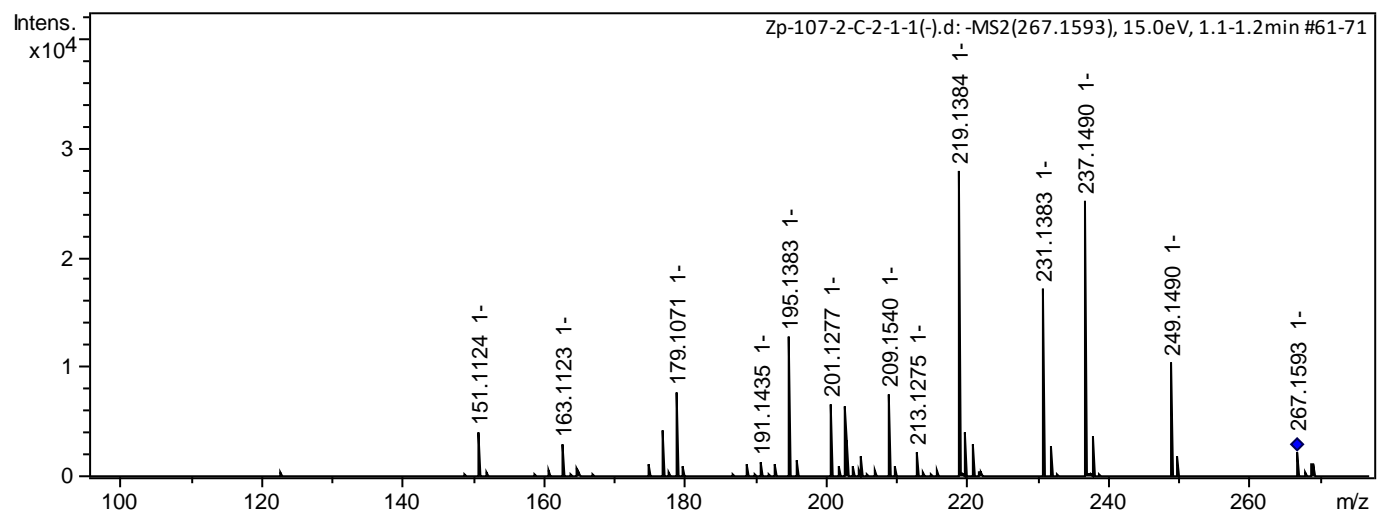
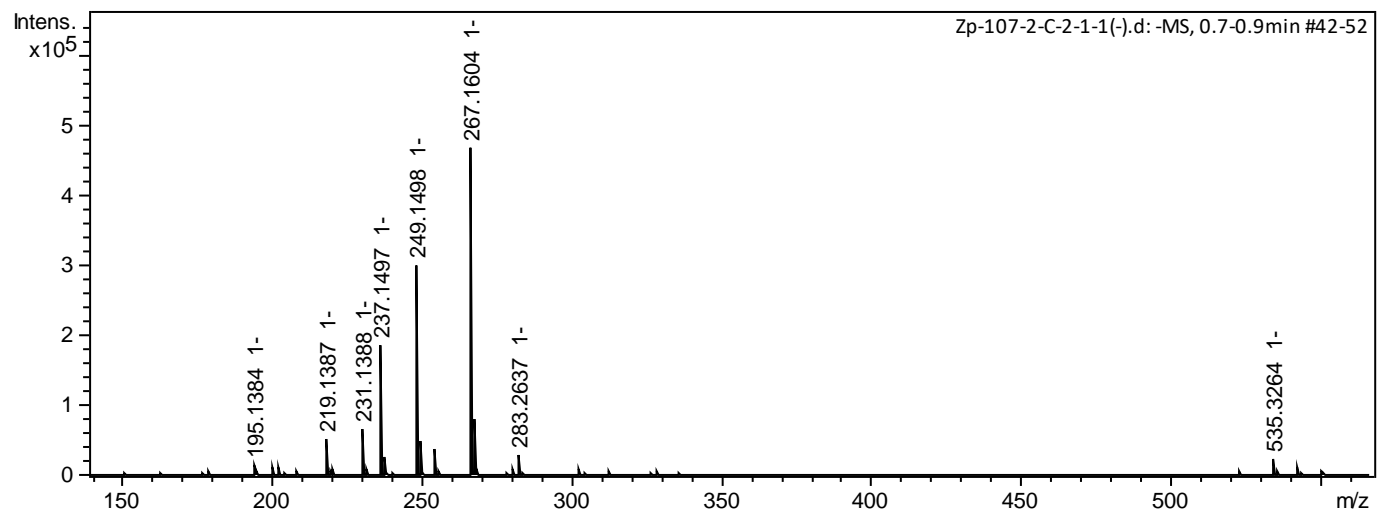
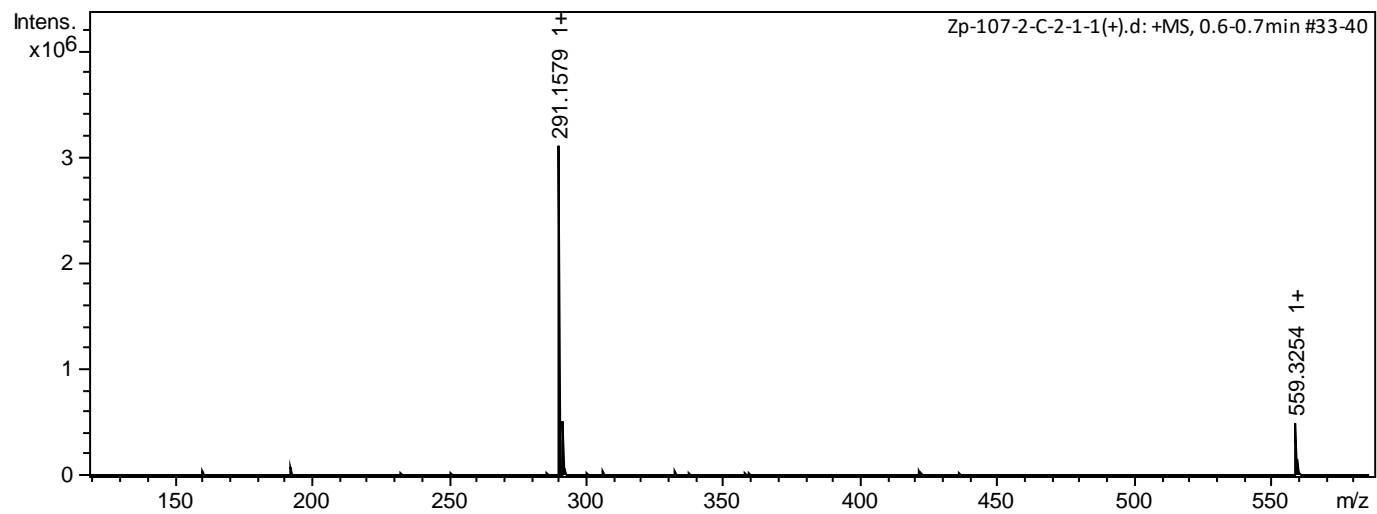


Figure S64. HRESIMS for 9





	meas	calc	Δ (ppm)
[M-H] ⁻	267,1604	267,1602	-0,7
[M+Na] ⁺	291,1579	291,1567	-4,1

Figure S65. ^1H NMR spectrum of **9** measured at 700 MHz in CDCl_3

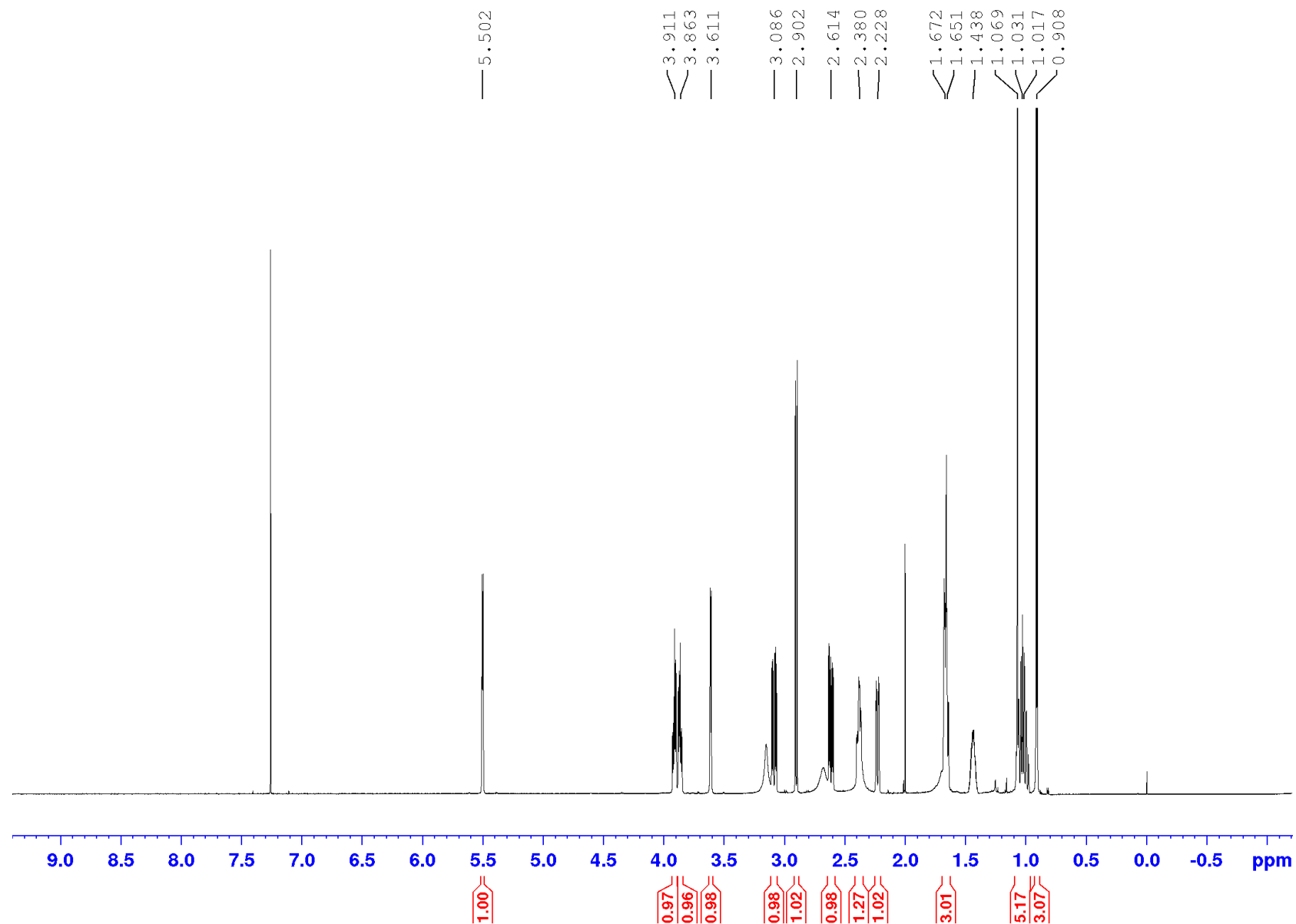


Figure S66. ^{13}C NMR spectrum of **9** measured at 176 MHz in CDCl_3

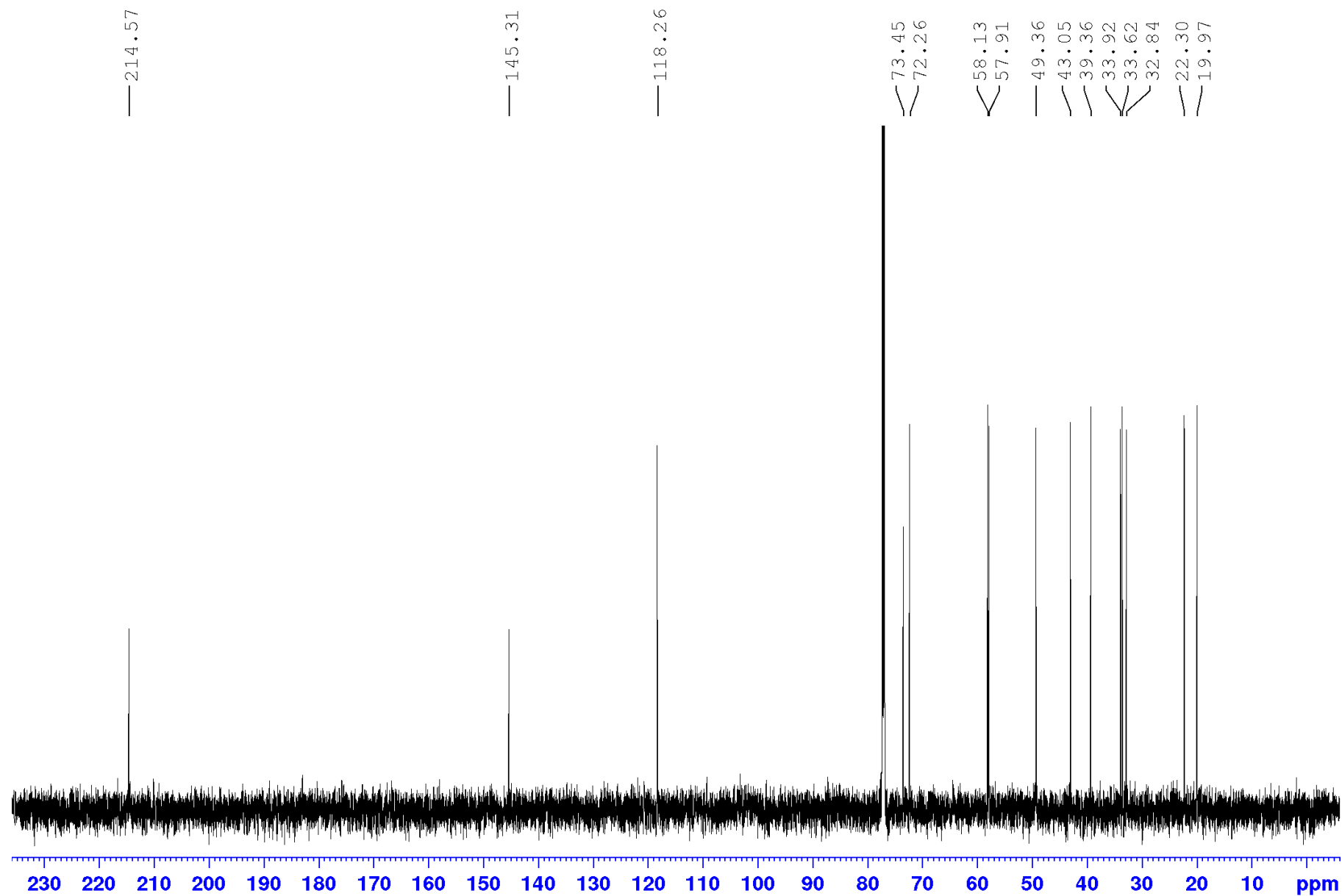


Figure S67. DEPT-135 spectrum of **9** measured at 176 MHz in CDCl₃

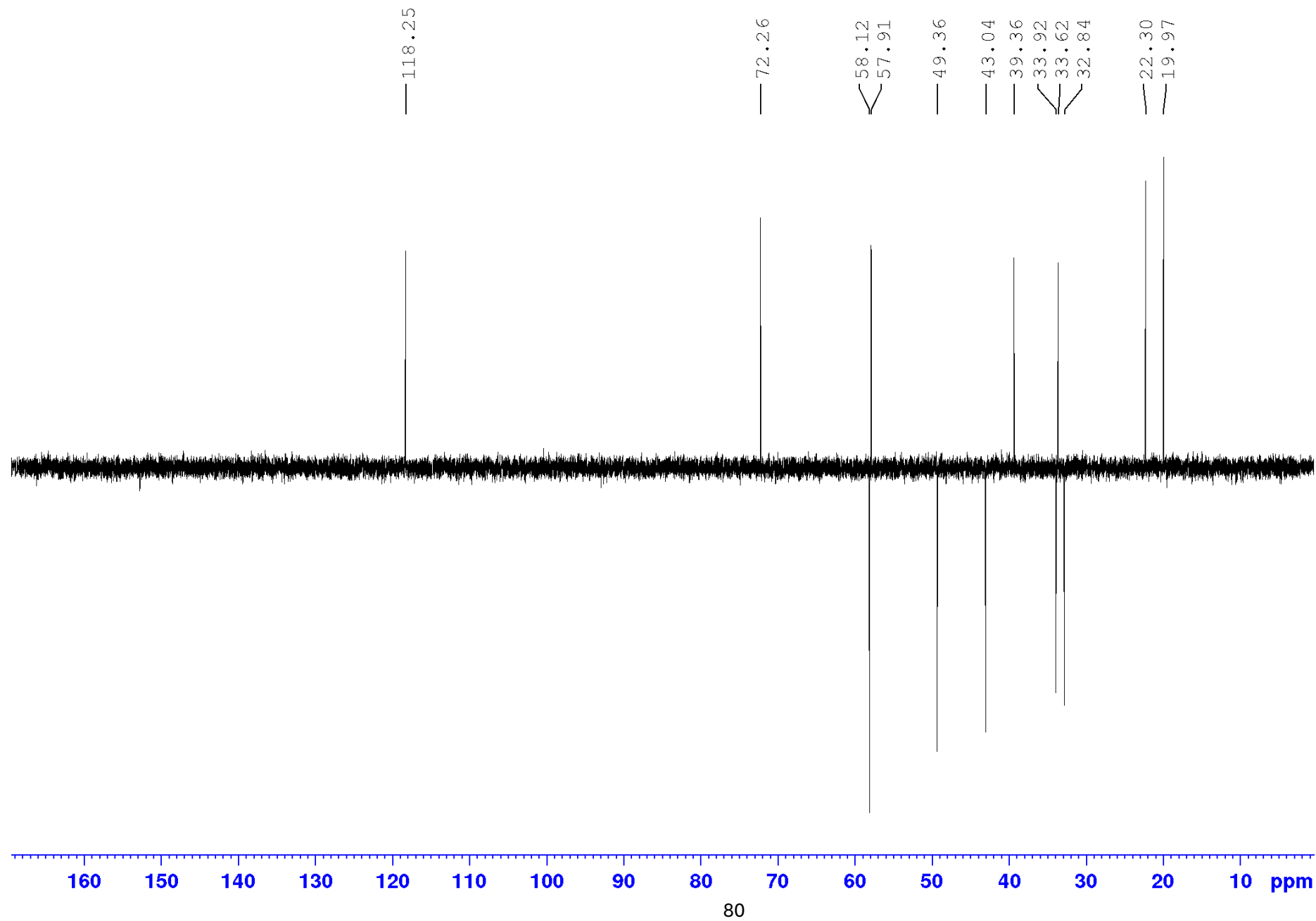


Figure S68. HSQC spectrum of 9 measured in CDCl₃

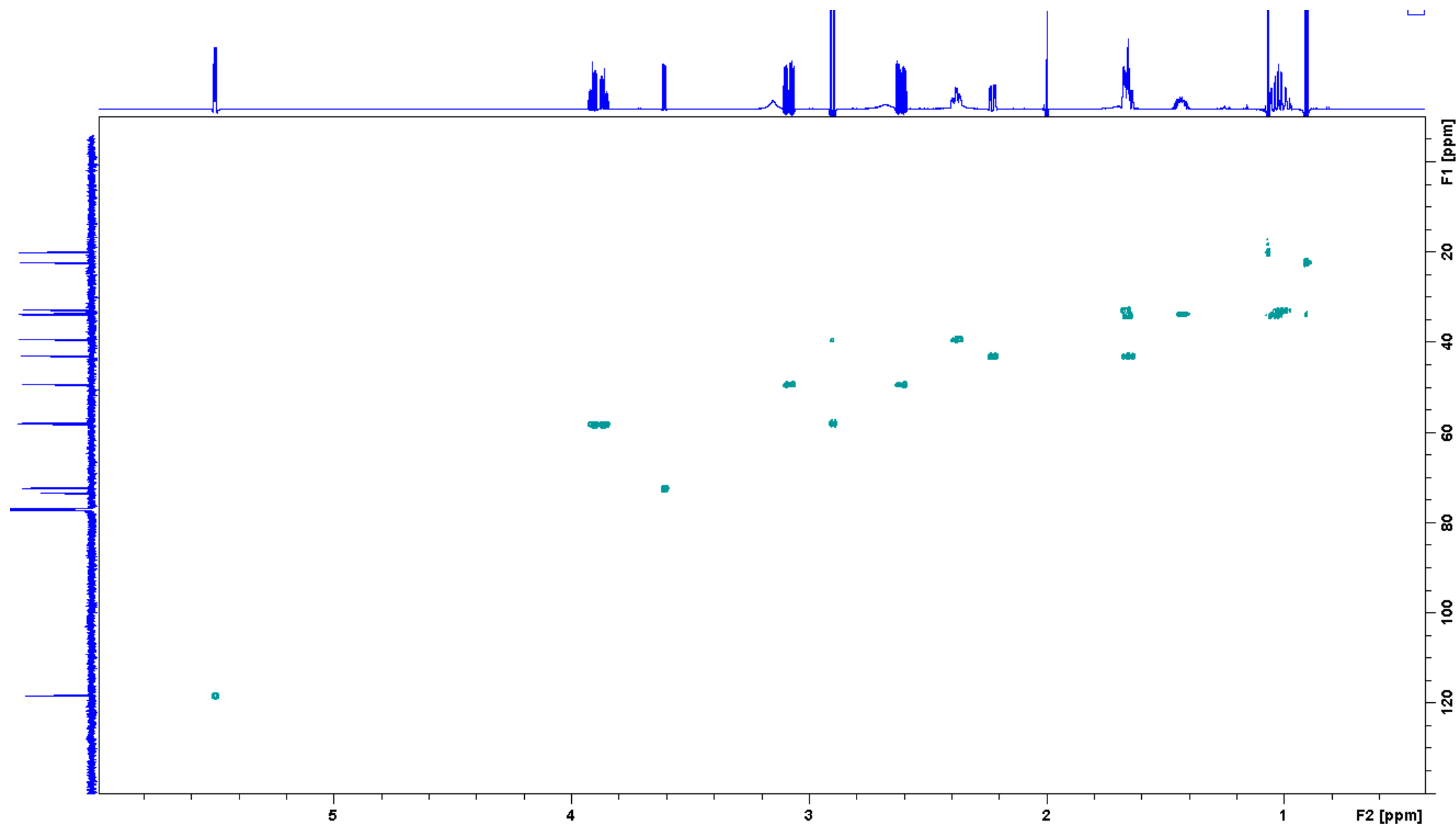


Figure S69. COSY spectrum of **9** measured in CDCl₃

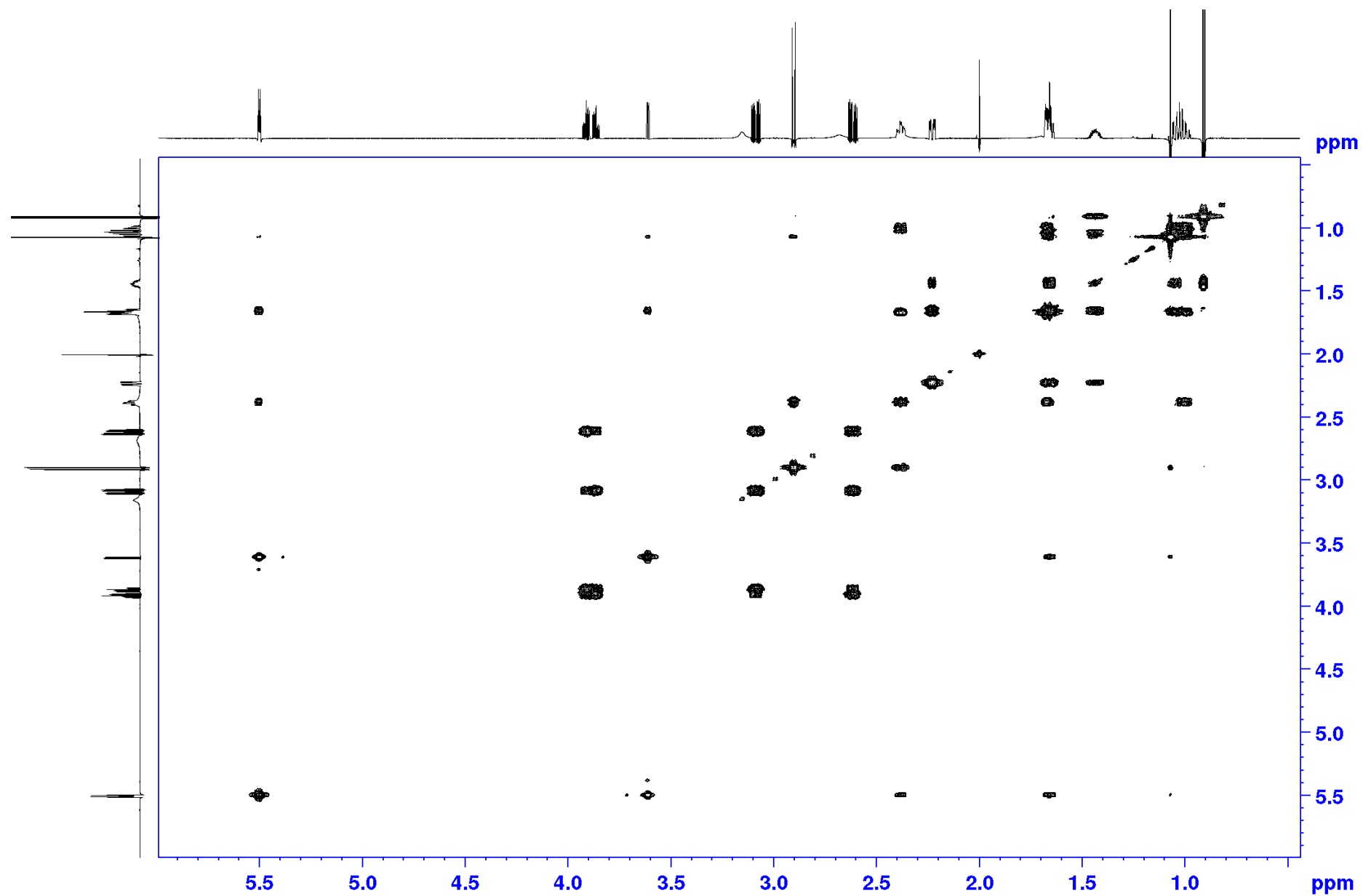


Figure S70. HMBC spectrum of 9 measured in CDCl₃

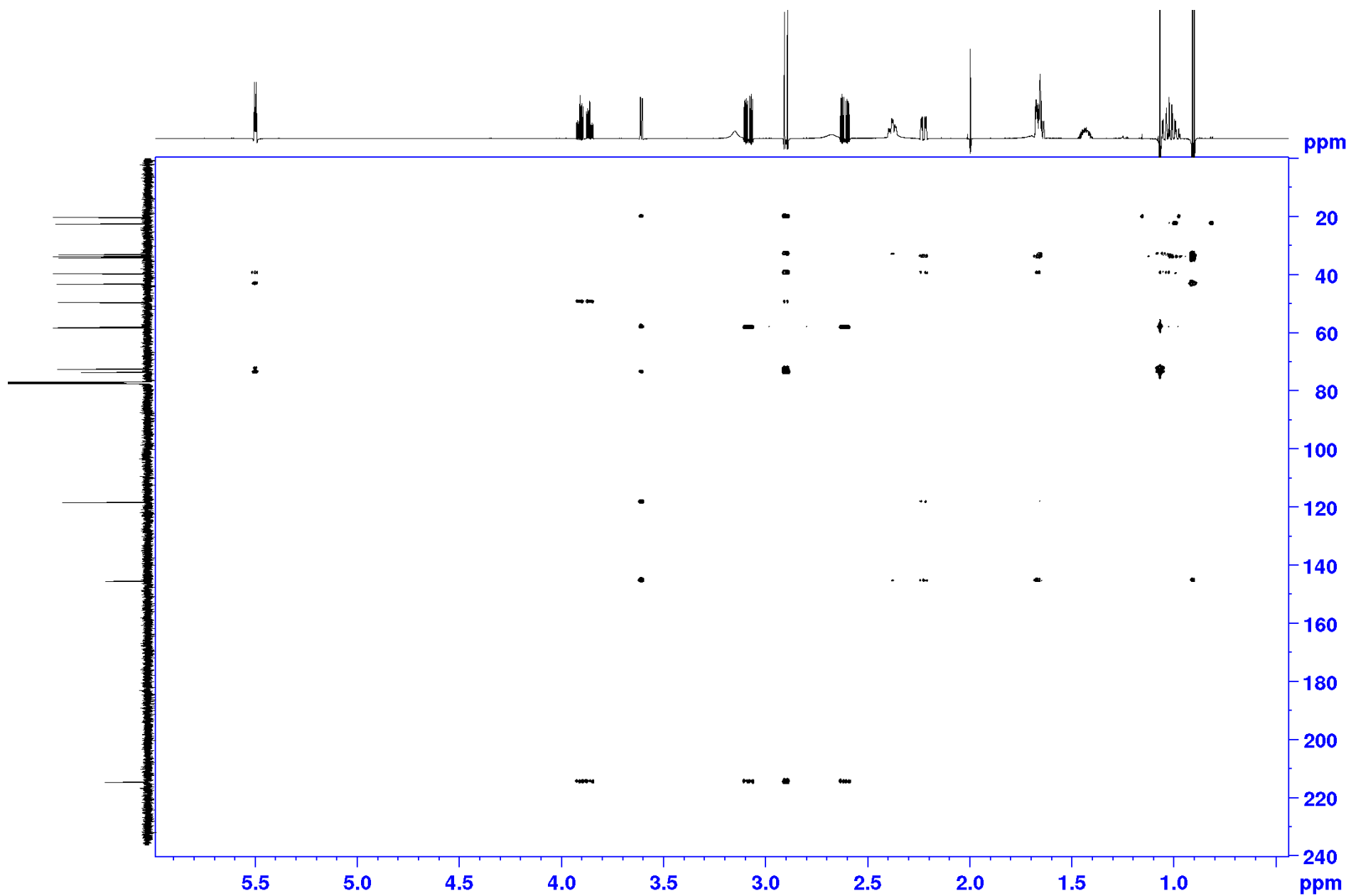


Figure S71. ROESY spectrum of 9 measured in CDCl₃

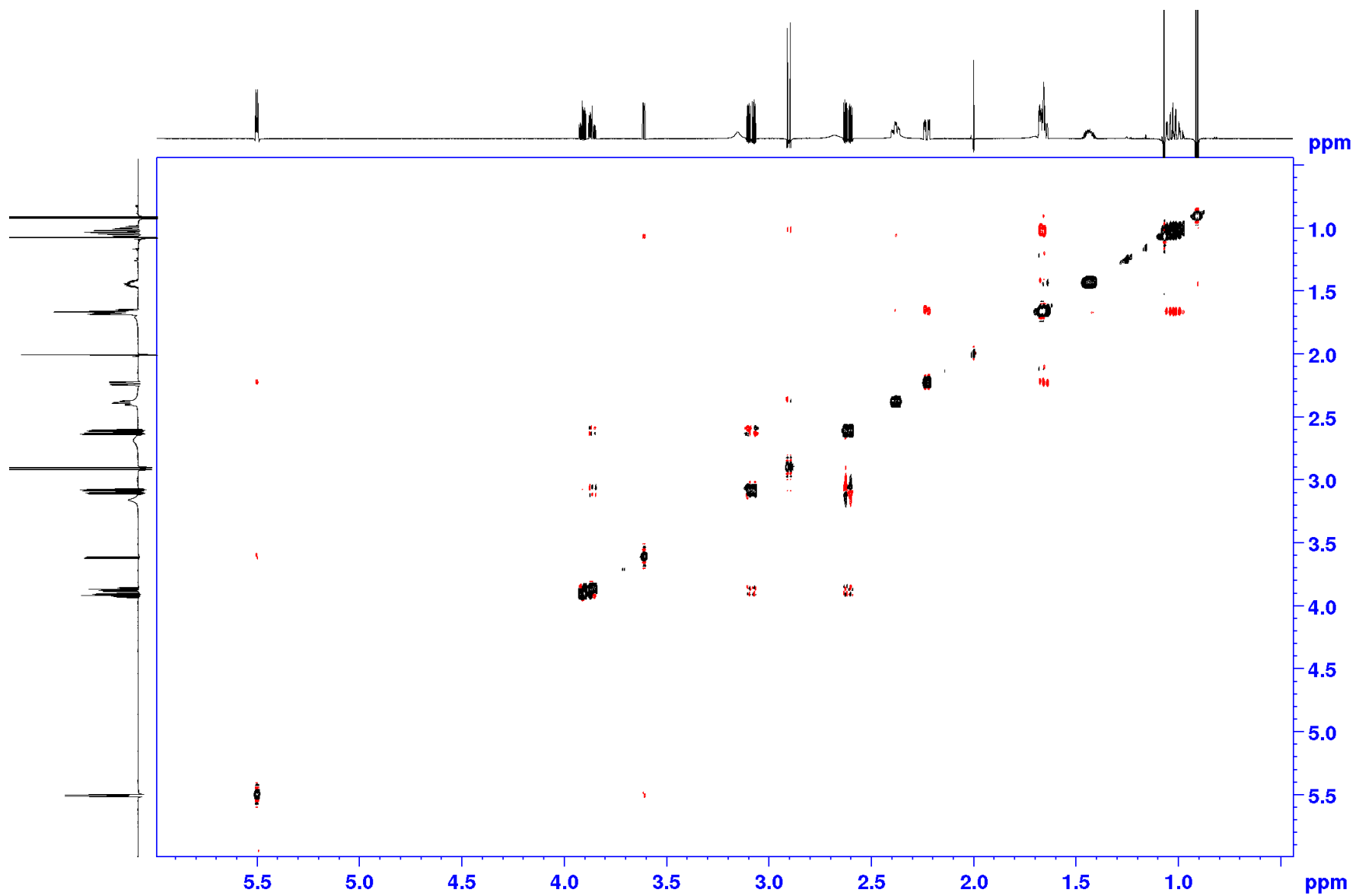
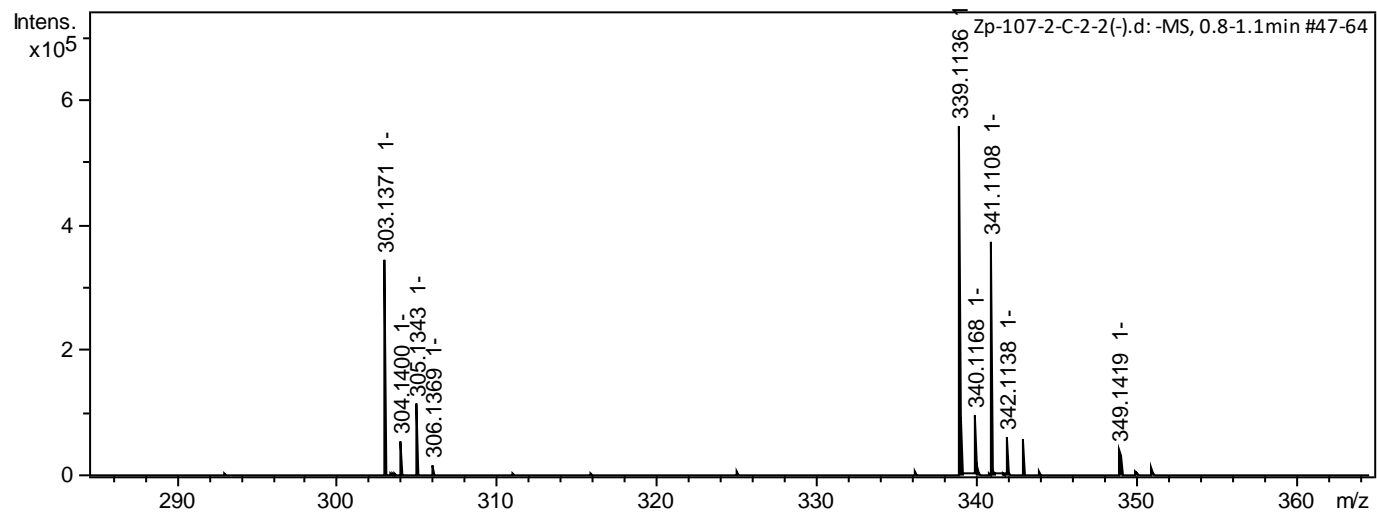
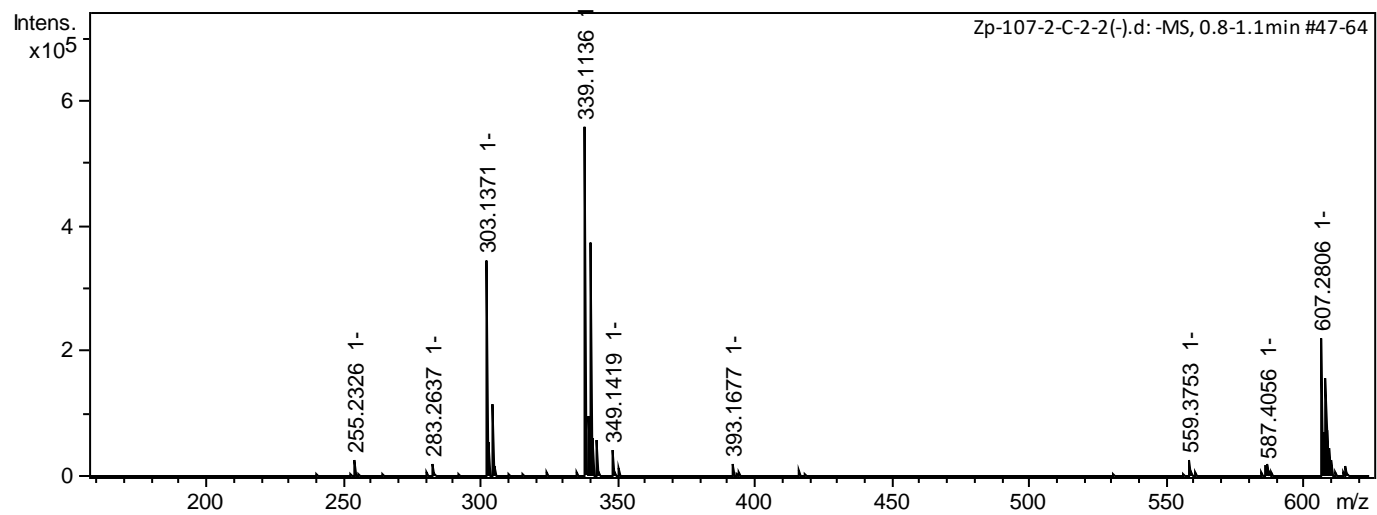
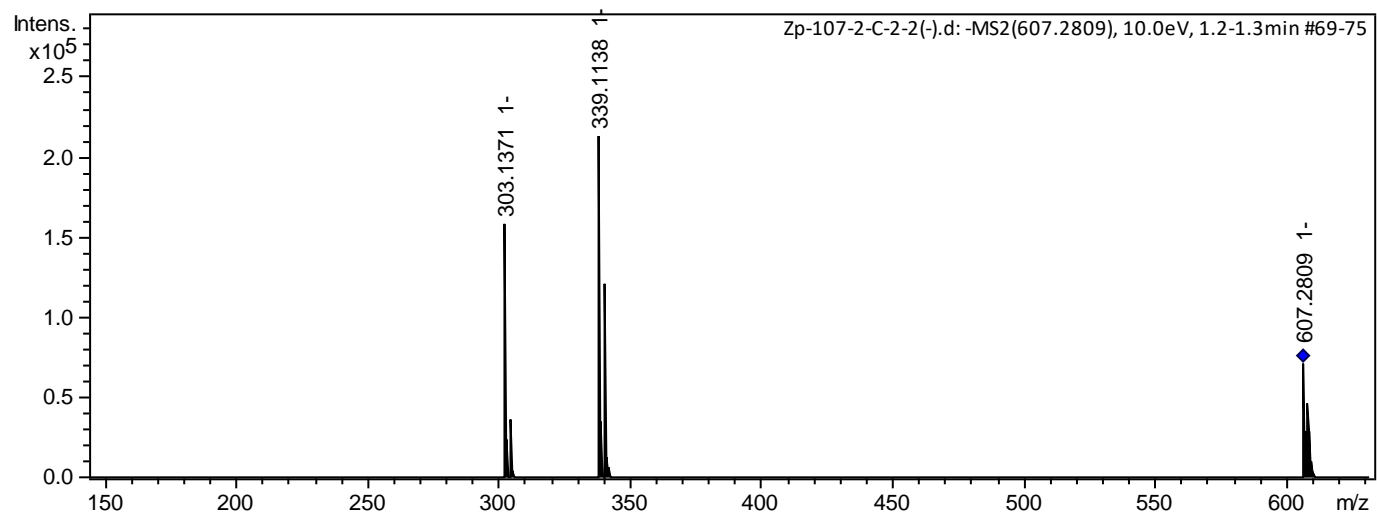
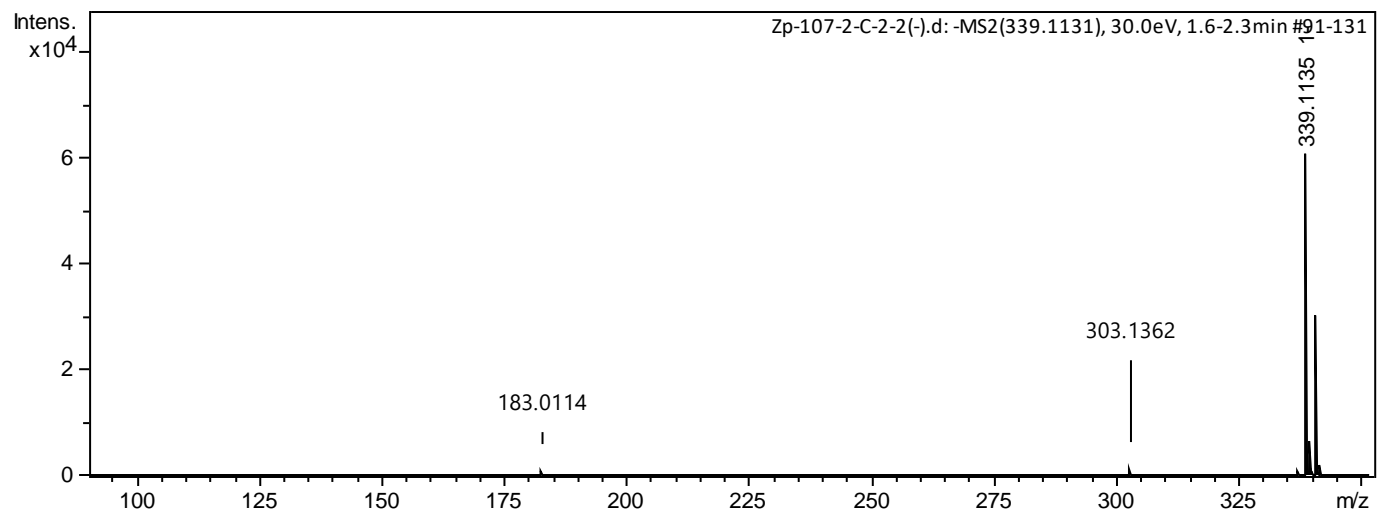
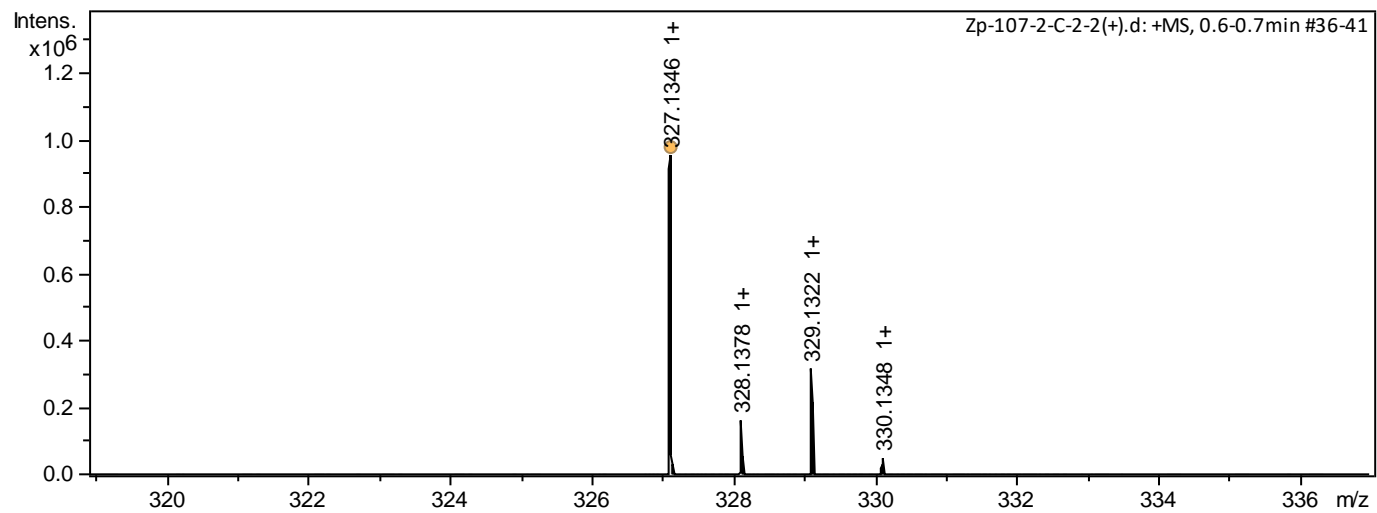
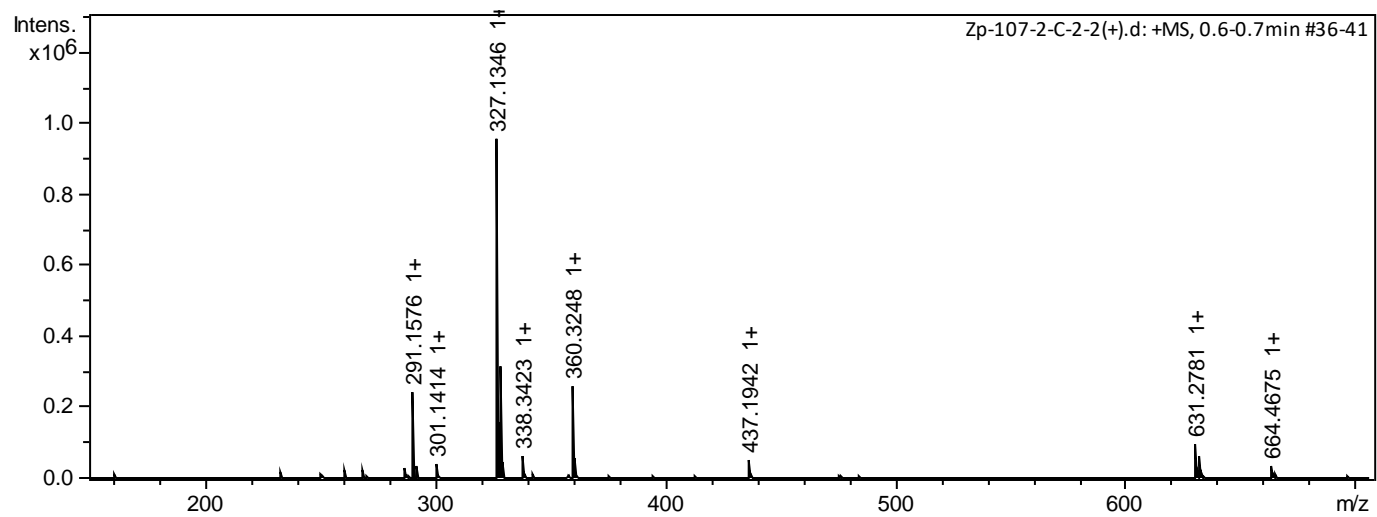
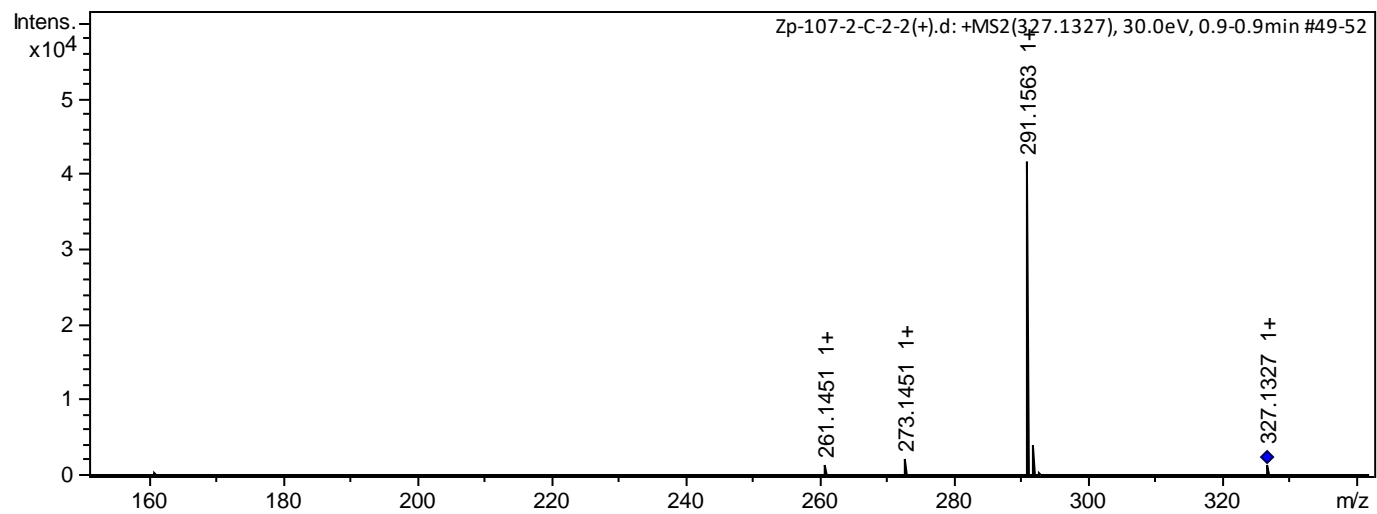


Figure S72. HRESIMS for 10









	meas	calc	Δ (ppm)
[M-H] ⁻	303,1371	303,1369	-0,7
[M+Na] ⁺	327,1346	327,1334	-3,7

Figure S73. ^1H NMR spectrum of 10 measured at 700 MHz in CDCl_3

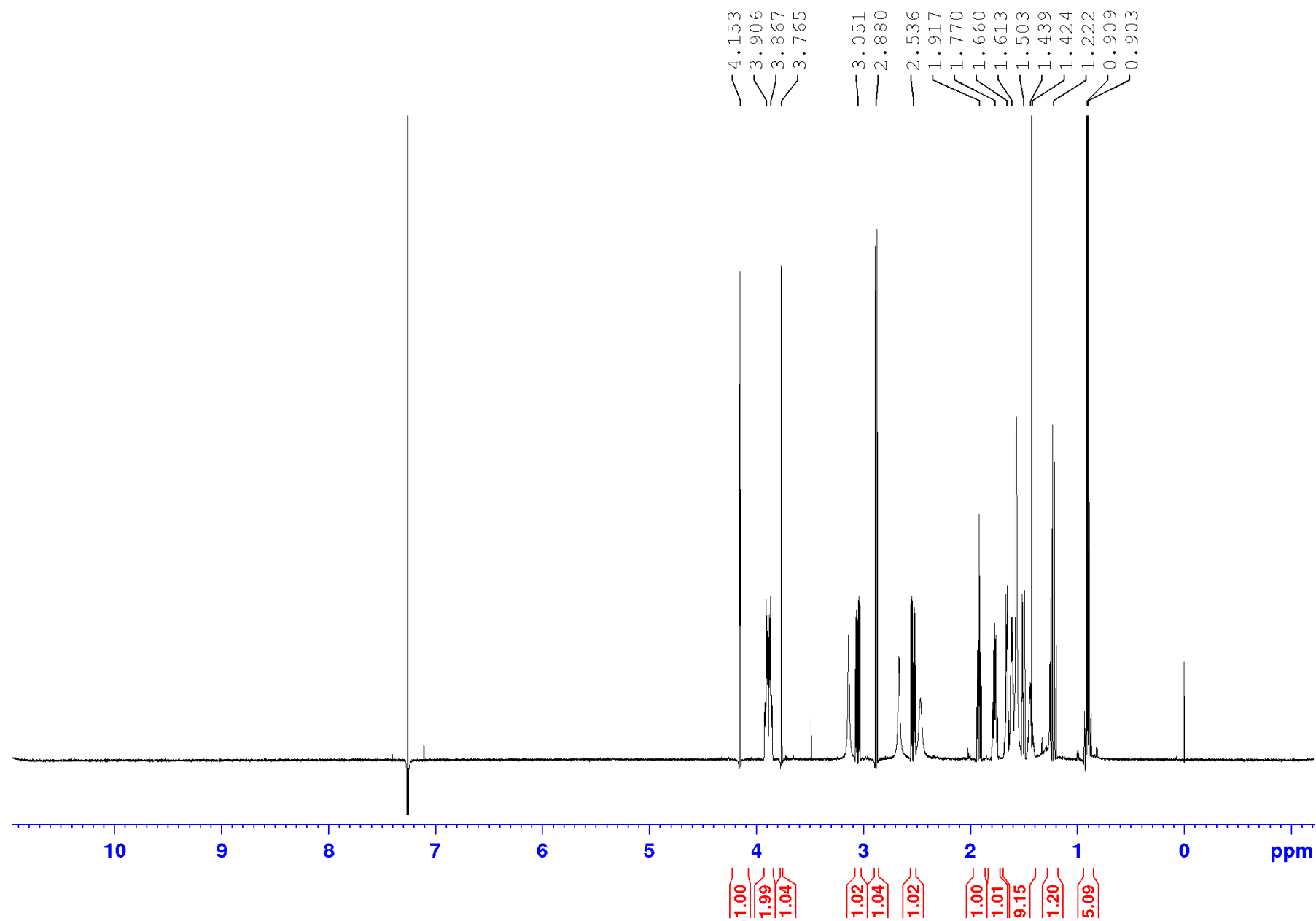


Figure S74. ^{13}C NMR spectrum of 10 measured at 176 MHz in CDCl_3

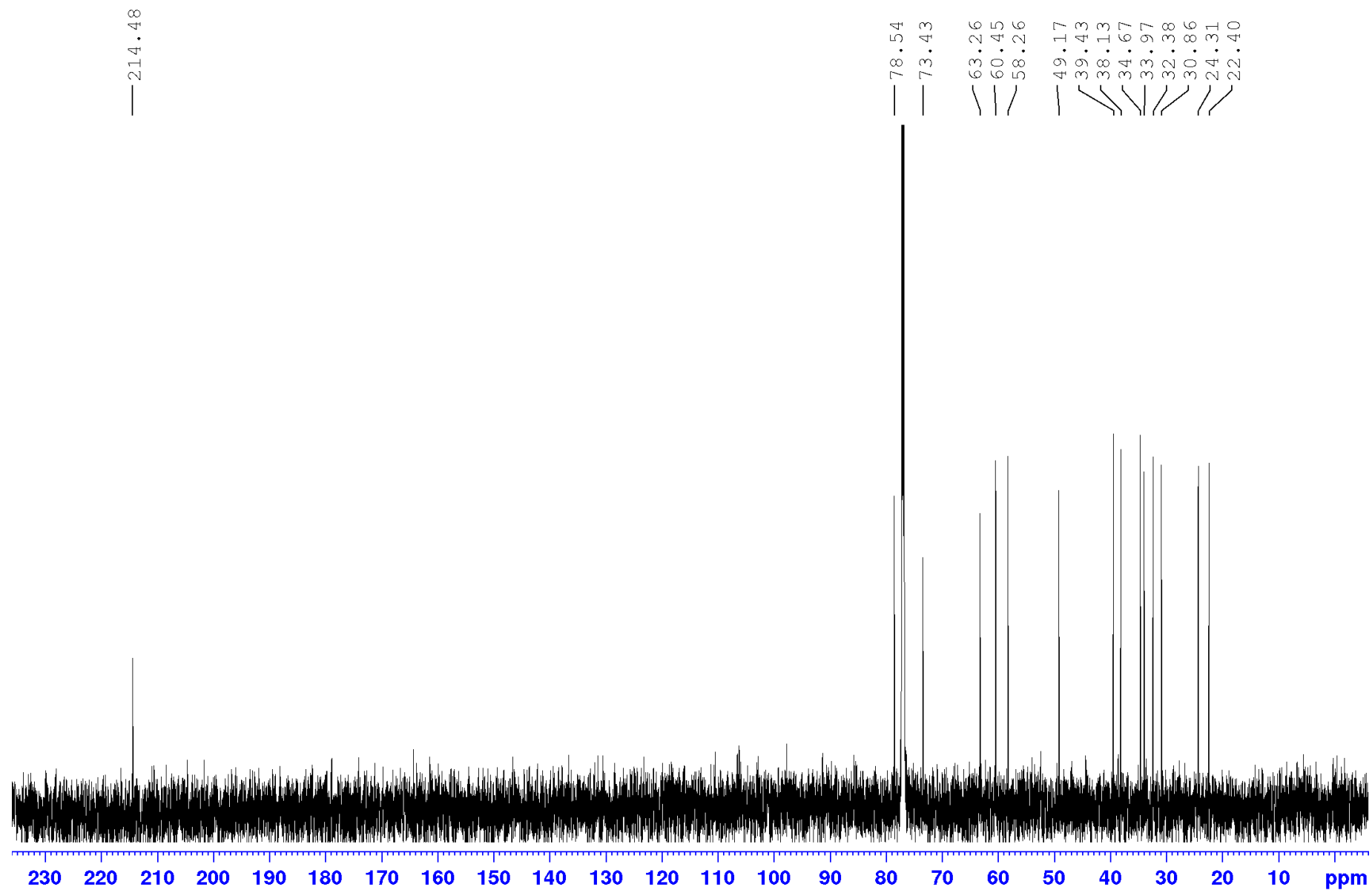


Figure S75. DEPT-135 spectrum of 10 measured at 176 MHz in CDCl₃

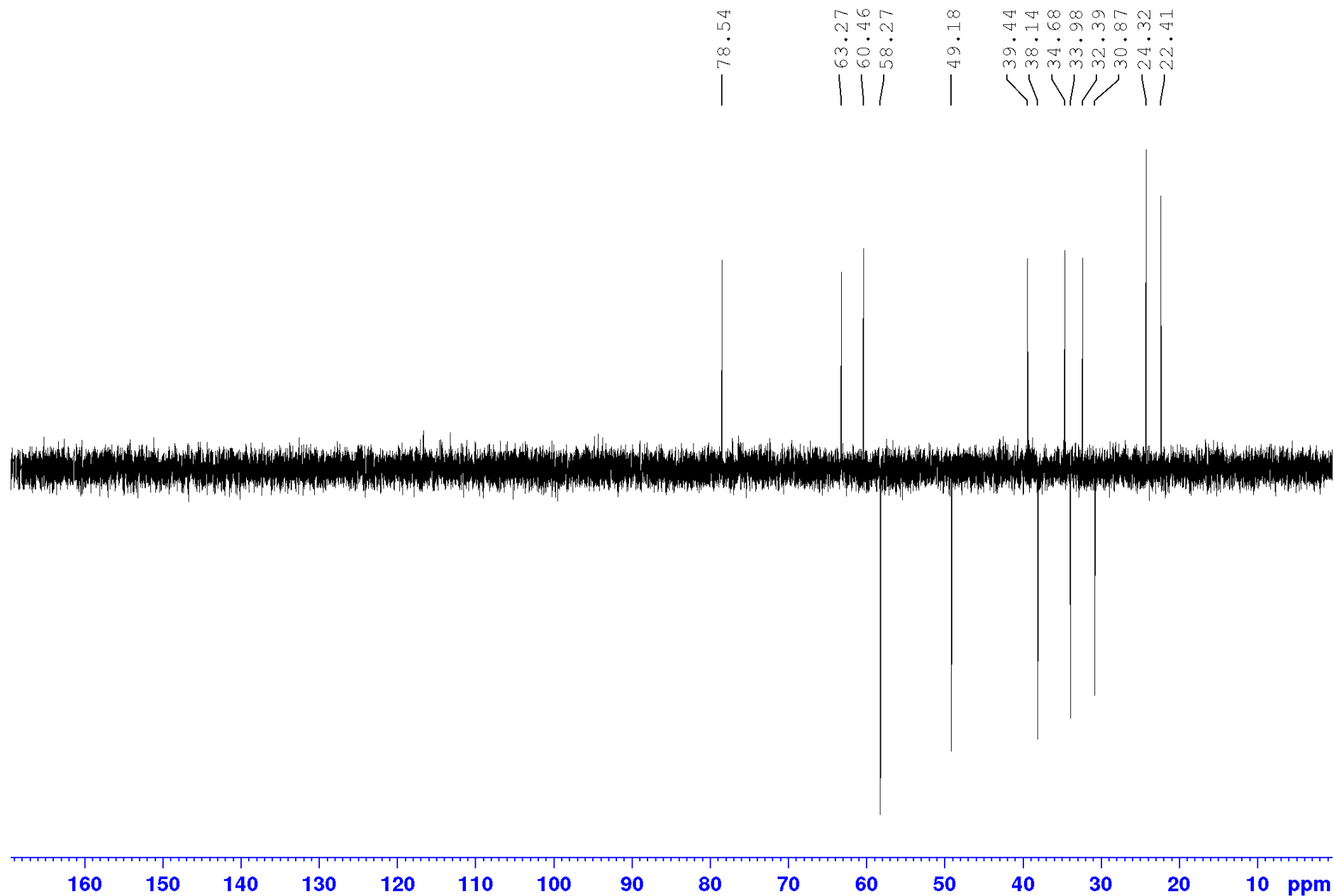


Figure S76. HSQC spectrum of 10 measured in CDCl₃

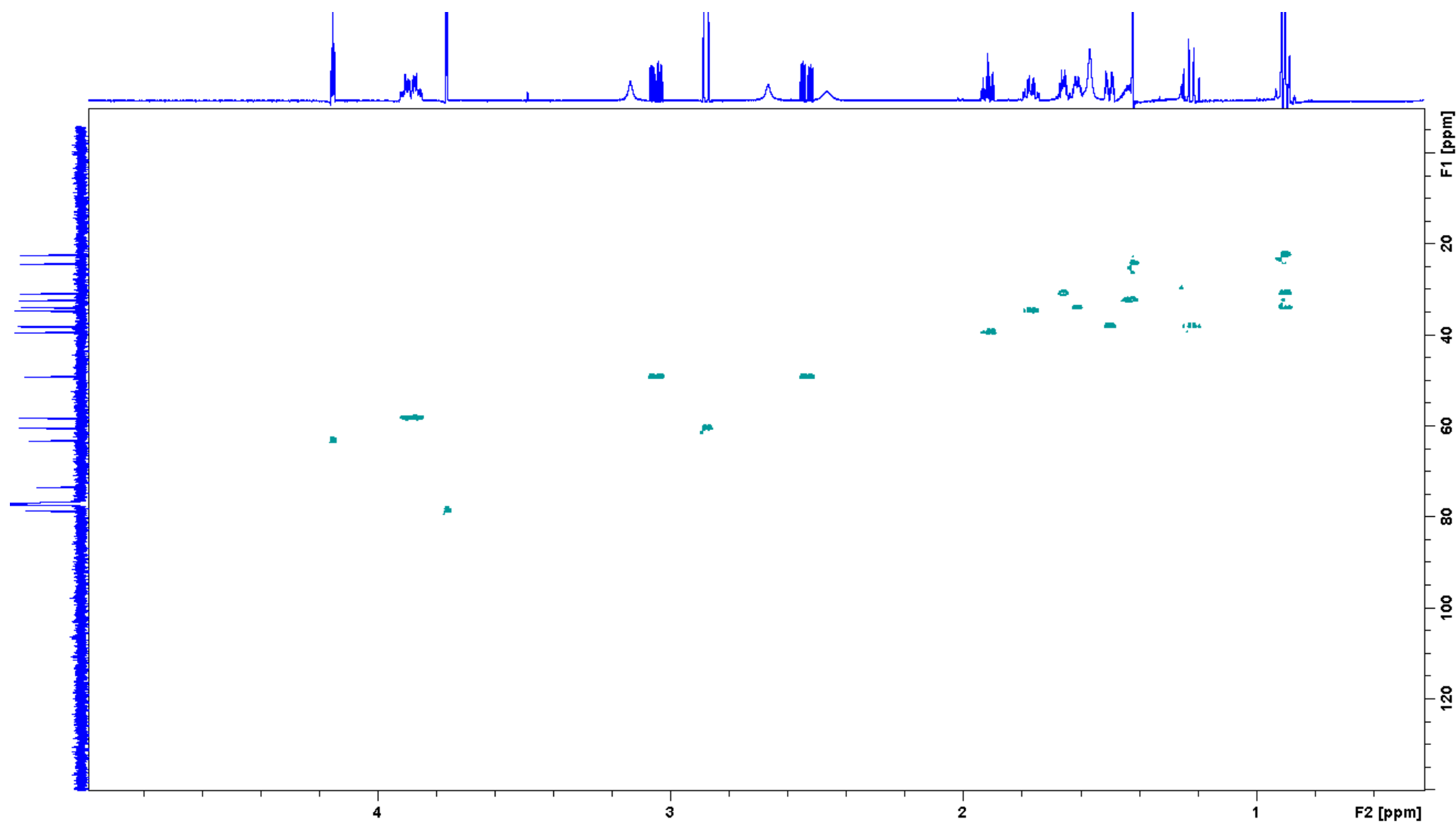


Figure S77. COSY spectrum of 10 measured in CDCl_3

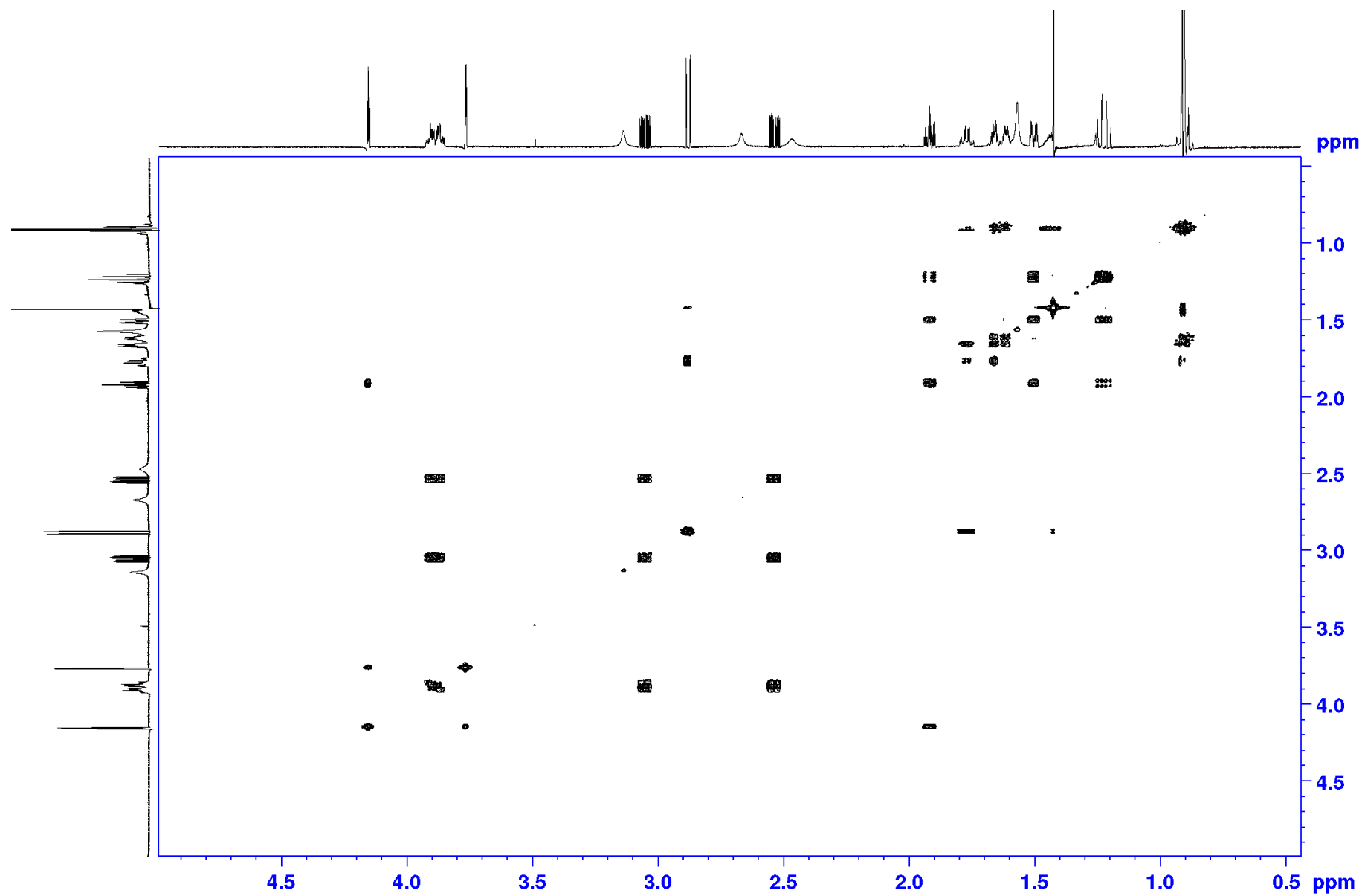


Figure S78. HMBC spectrum of 10 measured in CDCl₃

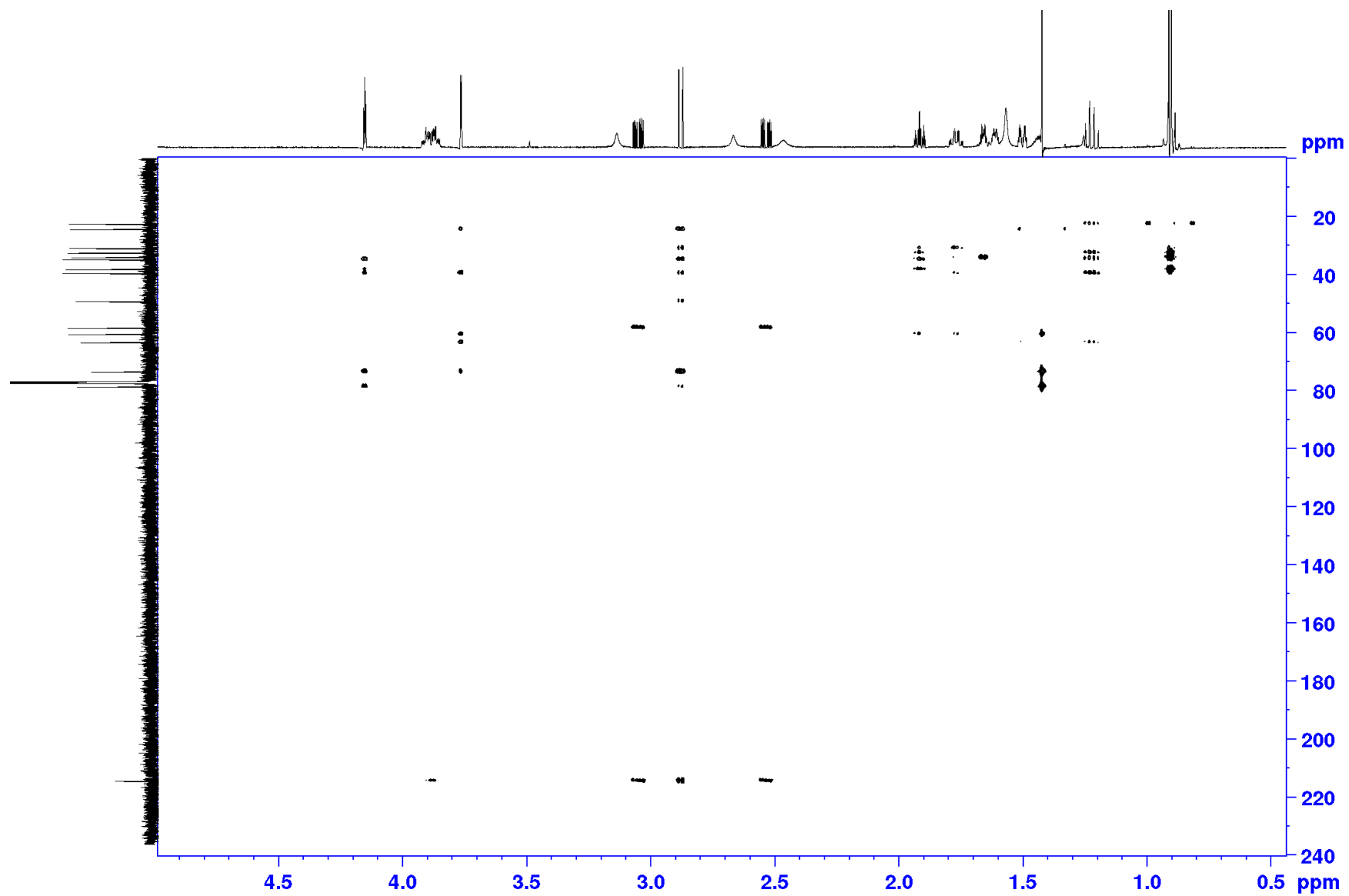


Figure S79. ROESY spectrum of 10 measured in CDCl₃

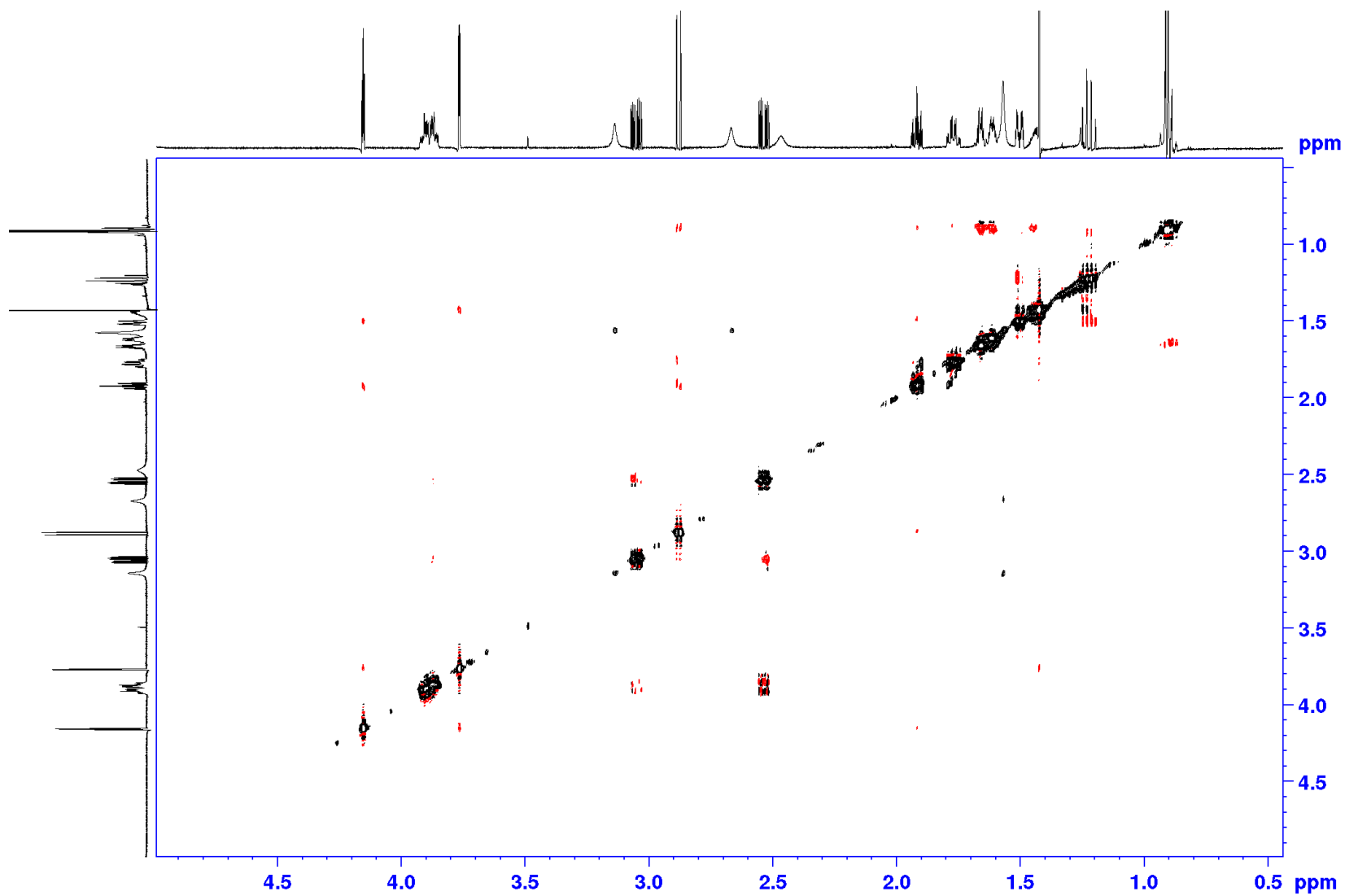


Figure S80. HRESIMS for 11

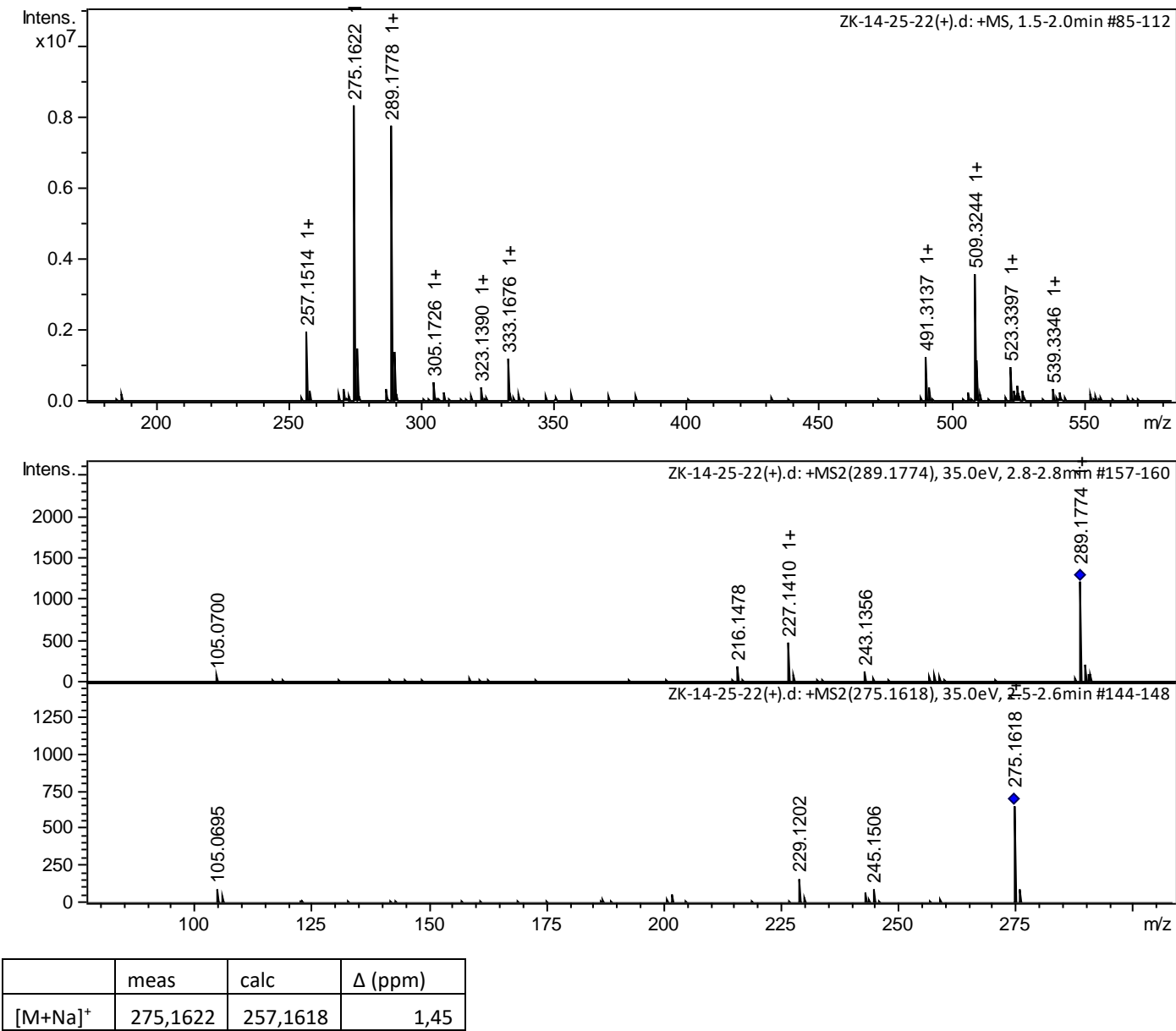


Figure S81. ^1H NMR spectrum of 11 measured at 700 MHz in CDCl_3

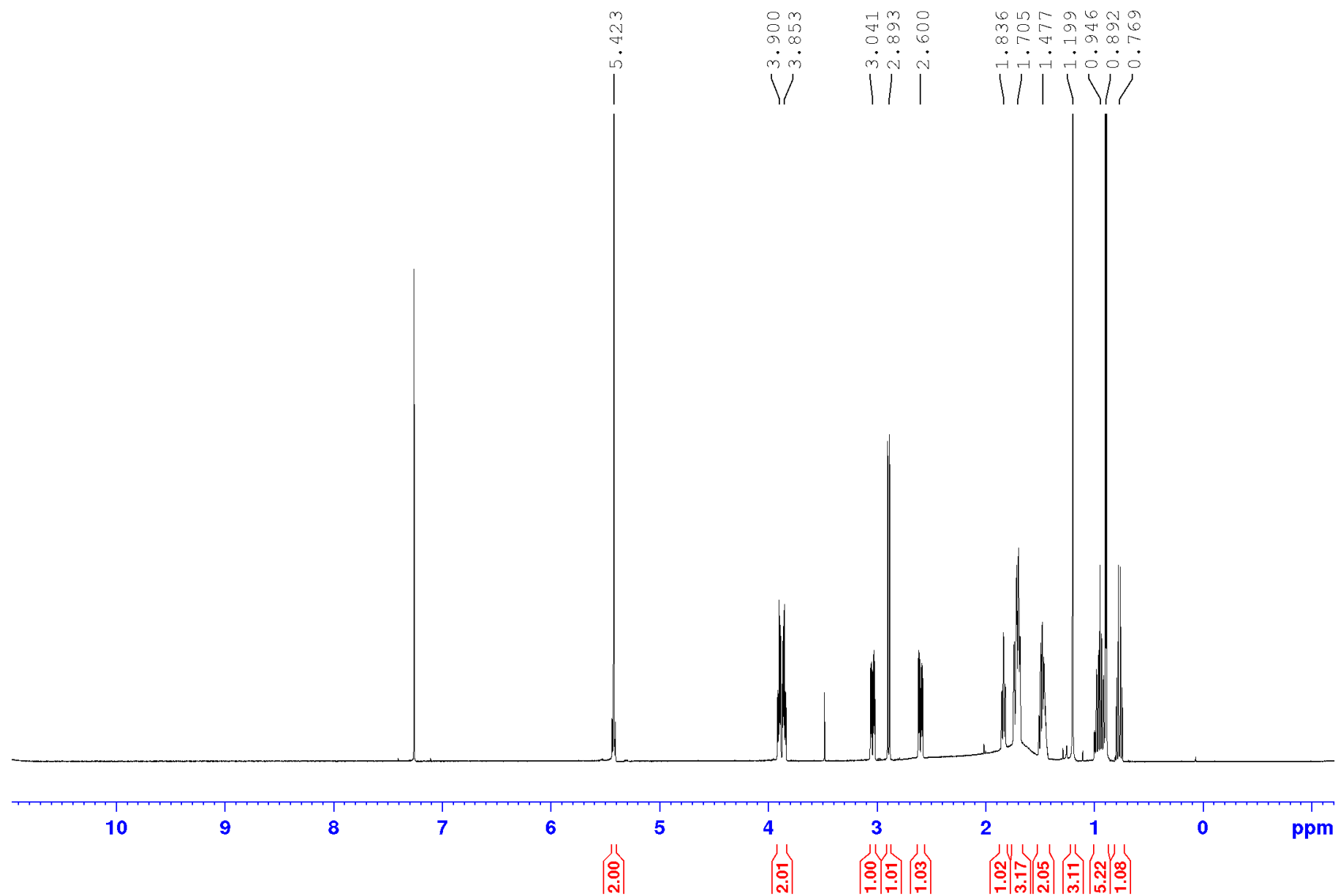


Figure S82. ^{13}C NMR spectrum of 11 measured at 176 MHz in CDCl_3

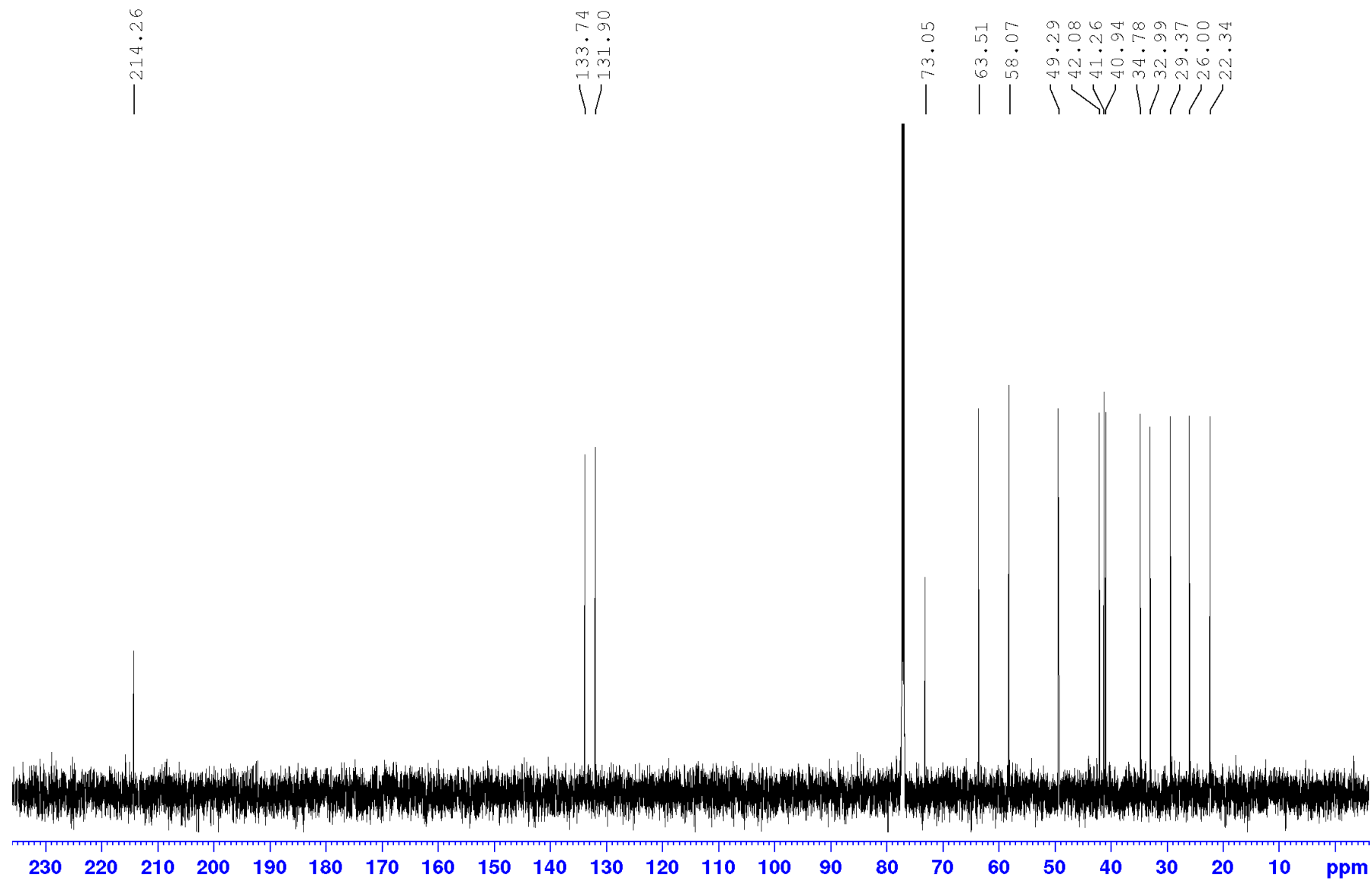
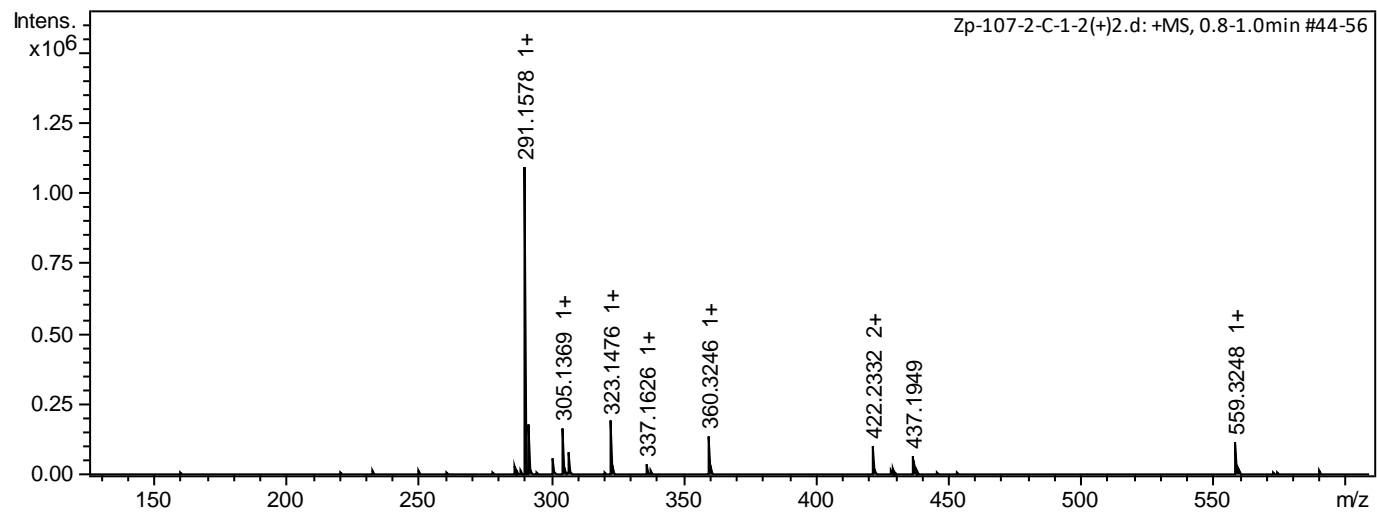
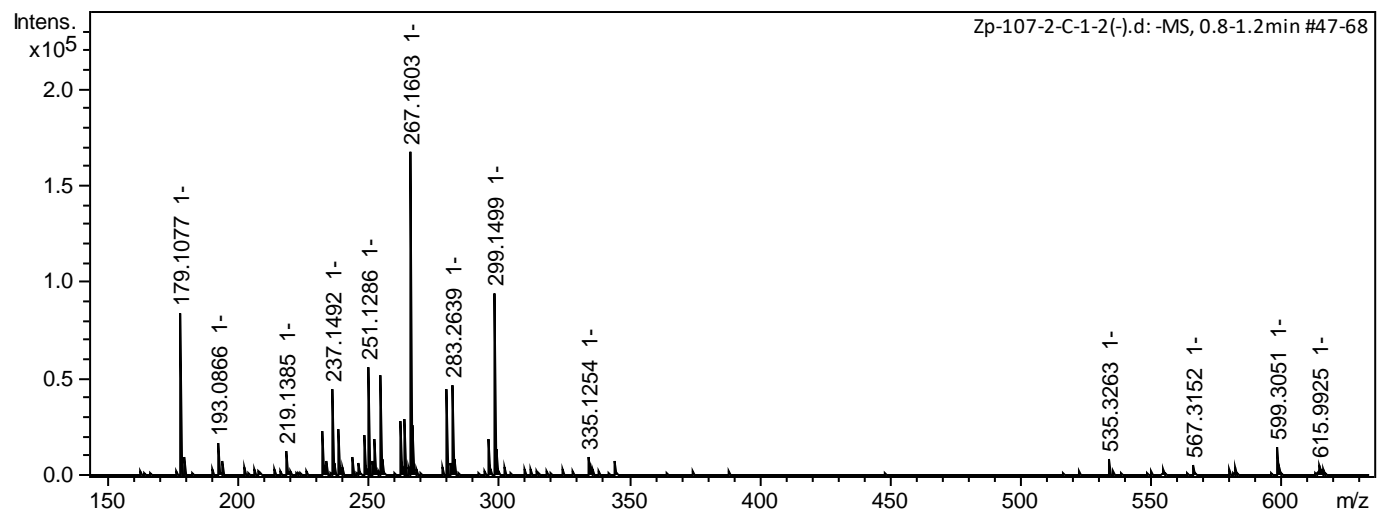
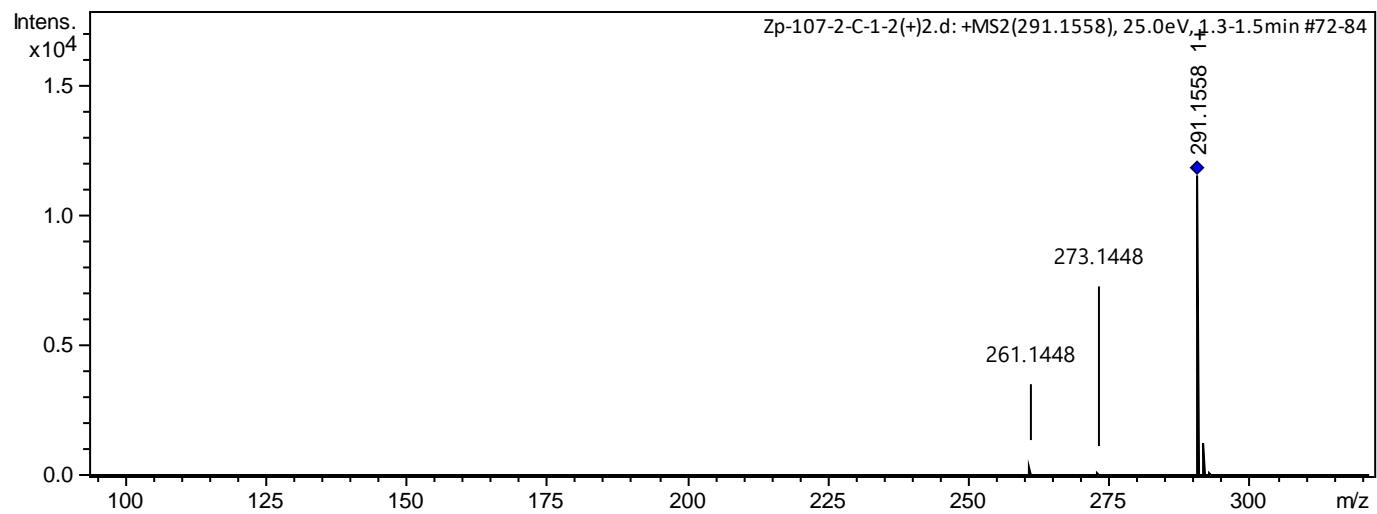


Figure S83. HRESIMS for 12





	meas	calc	Δ (ppm)
[M-H] ⁻	267,1603	267,1602	-0,4
[M+Na] ⁺	291,1578	291,1567	-3,8

Figure S84. ^1H NMR spectrum of 12 measured at 700 MHz in CDCl_3

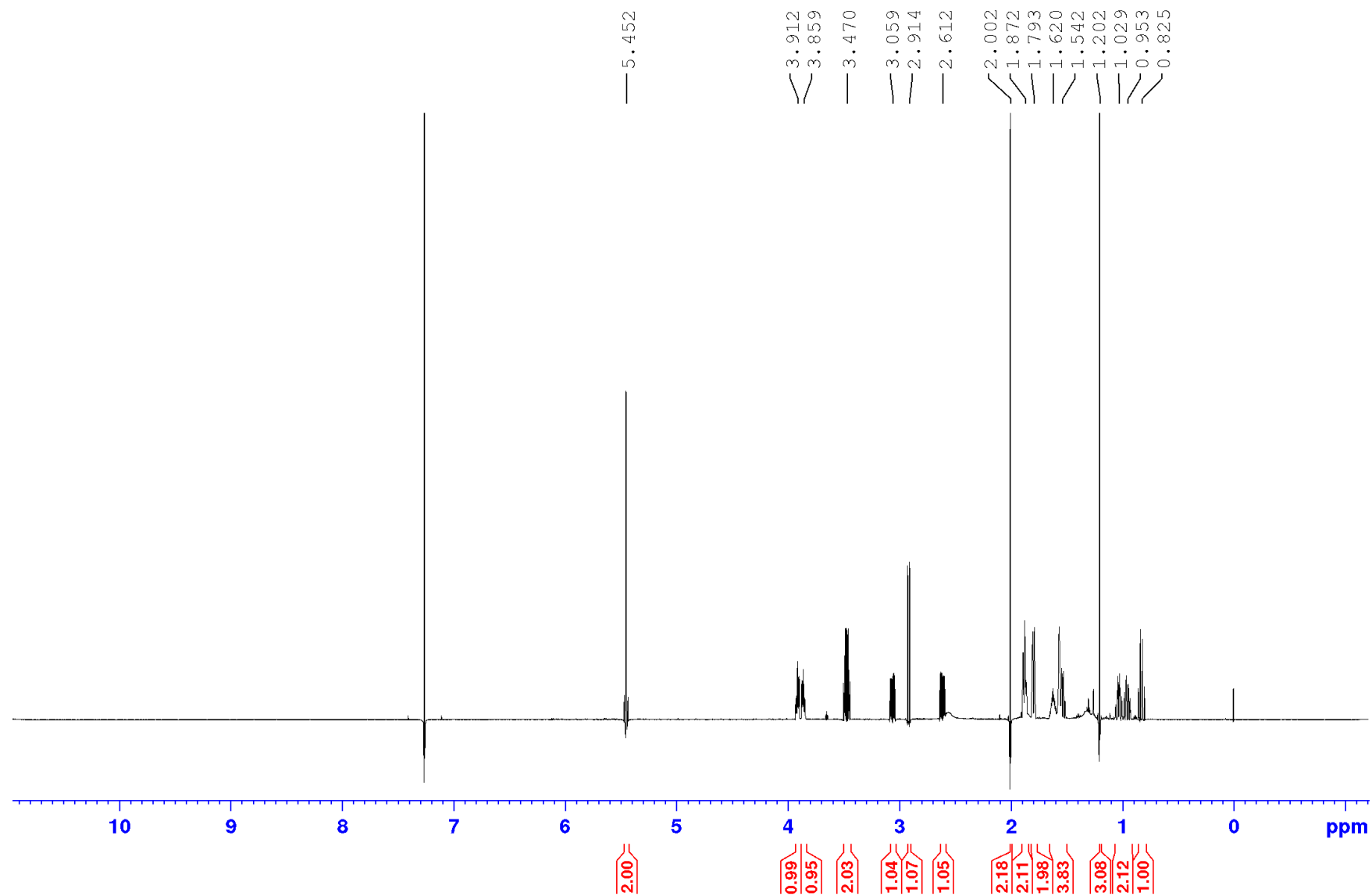


Figure S85. ^{13}C NMR spectrum of 12 measured at 176 MHz in CDCl_3

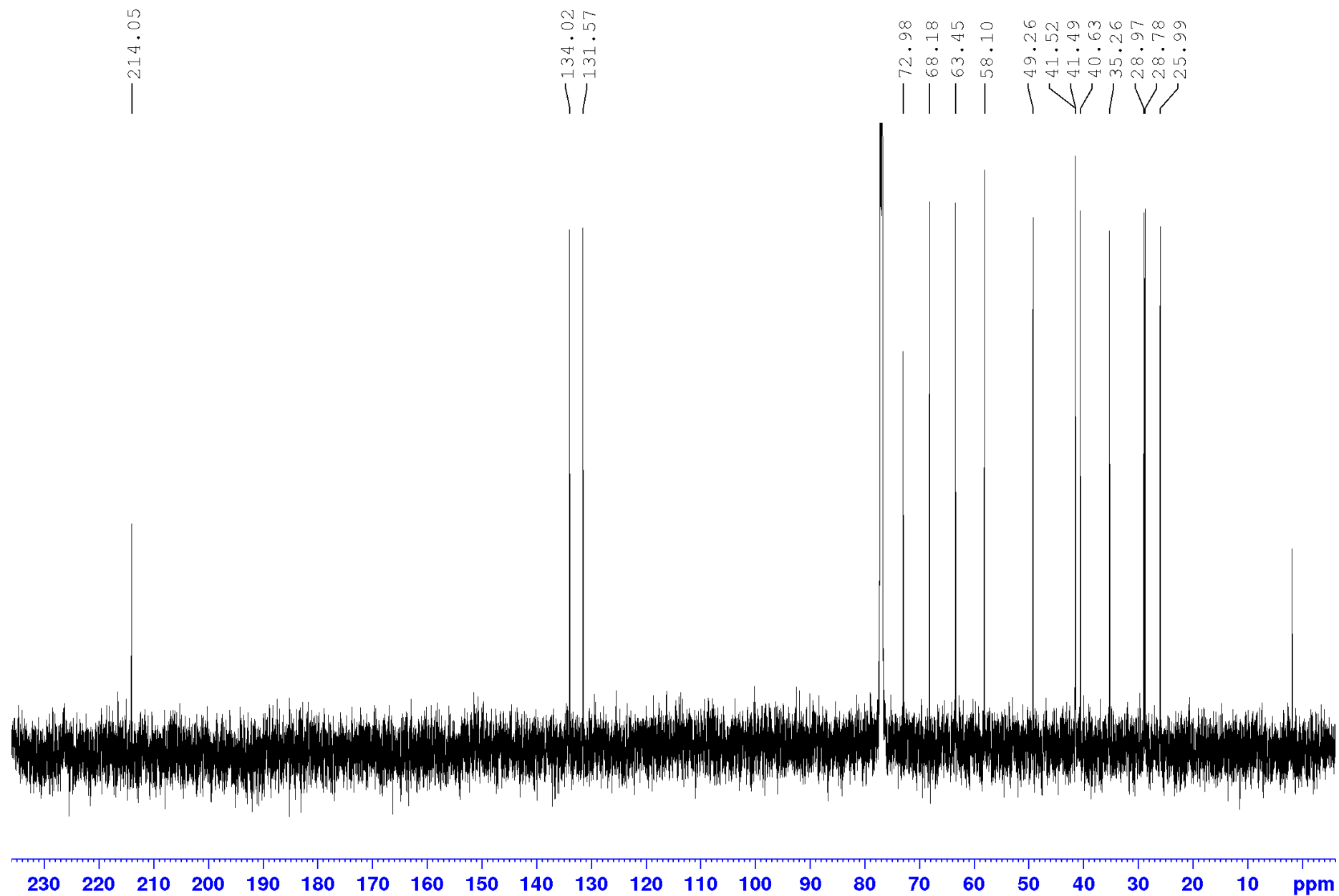
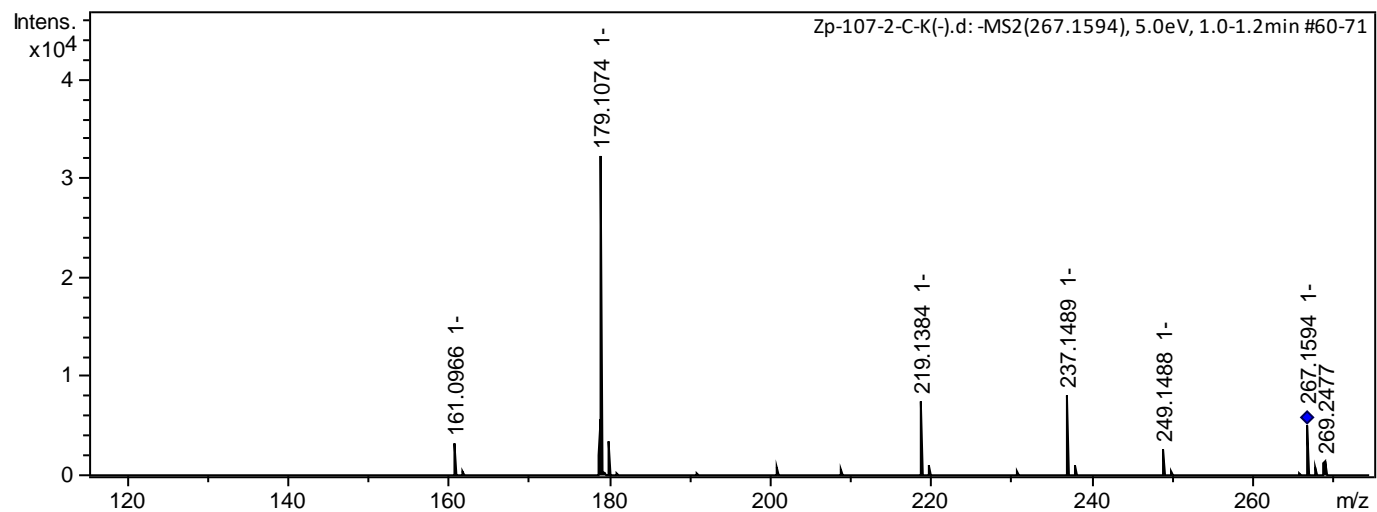
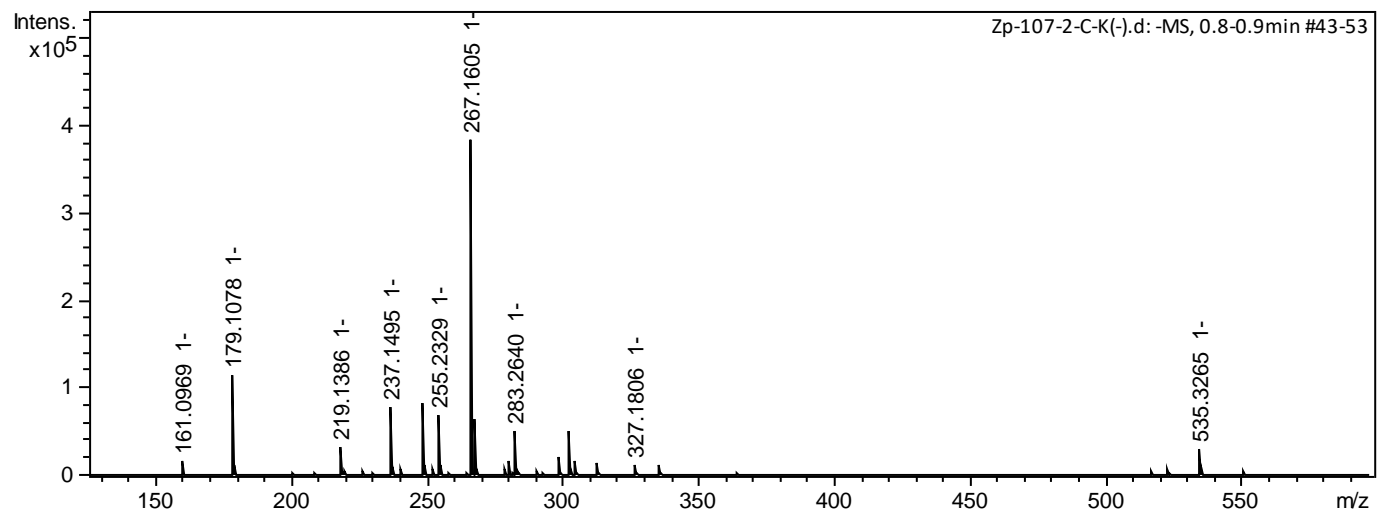
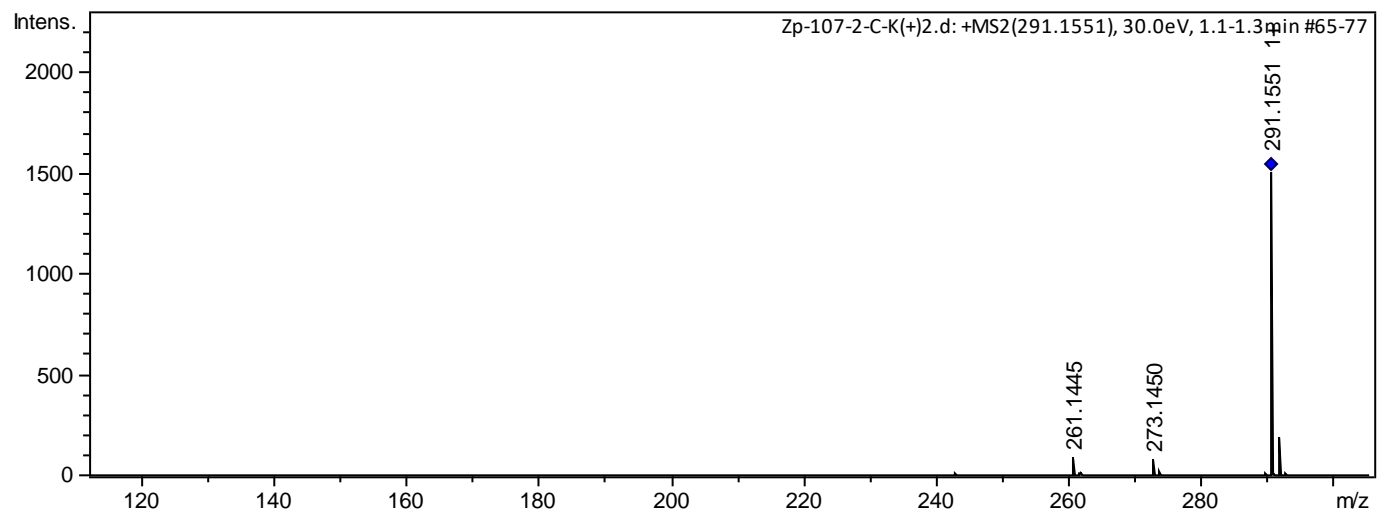
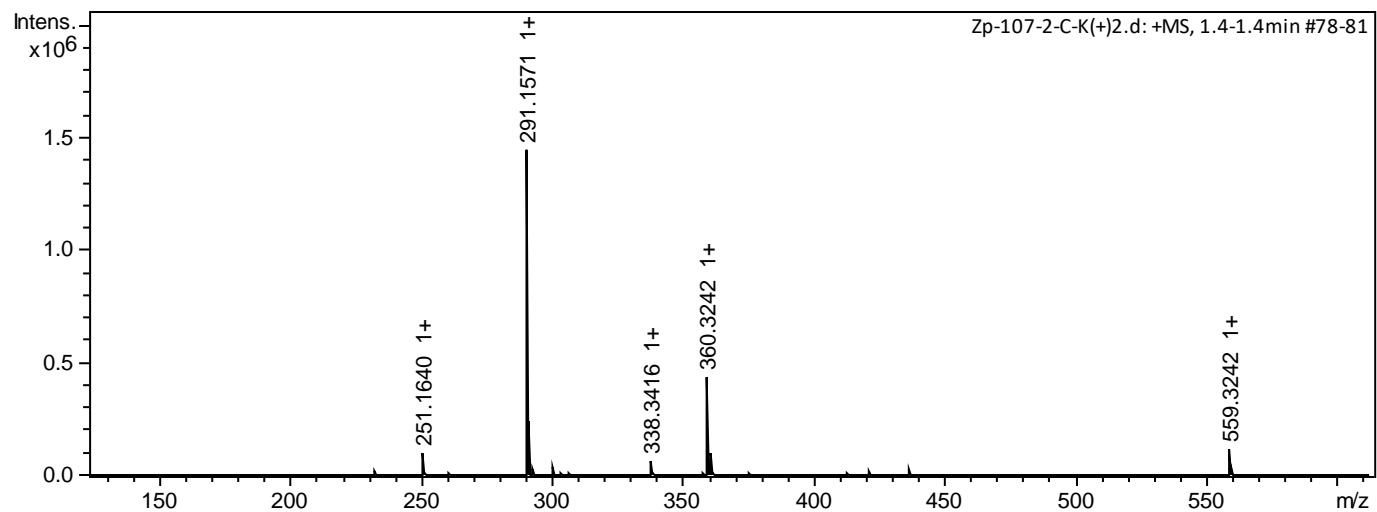


Figure S86. HRESIMS for 13





	meas	calc	Δ (ppm)
[M-H] ⁻	267,1605	267,1602	-1,1
[M+Na] ⁺	291,1571	291,1567	-1,4

Figure S87. ^1H NMR spectrum of 13 measured at 700 MHz in CDCl_3

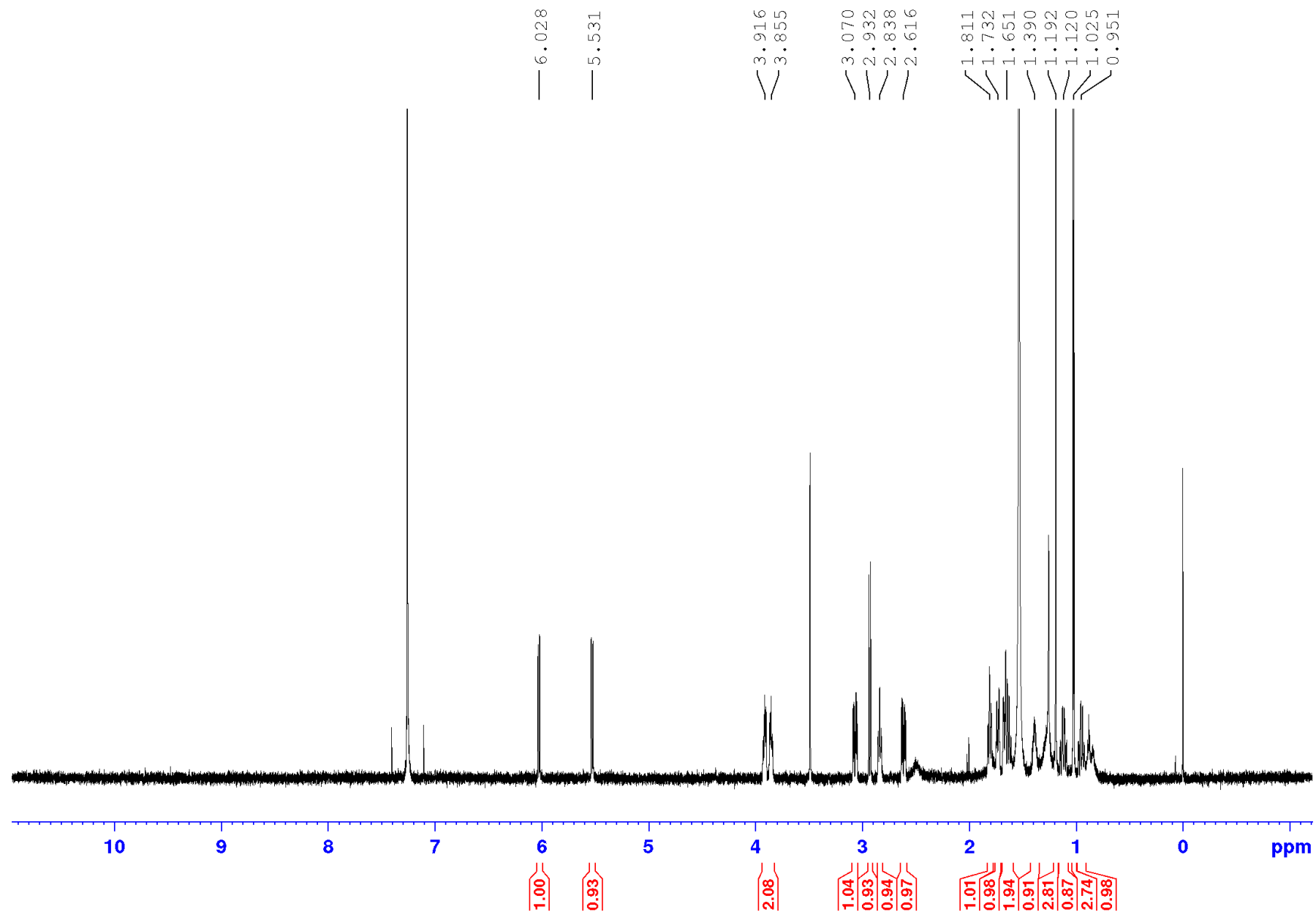


Figure S88. ^{13}C NMR spectrum of 13 measured at 176 MHz in CDCl_3

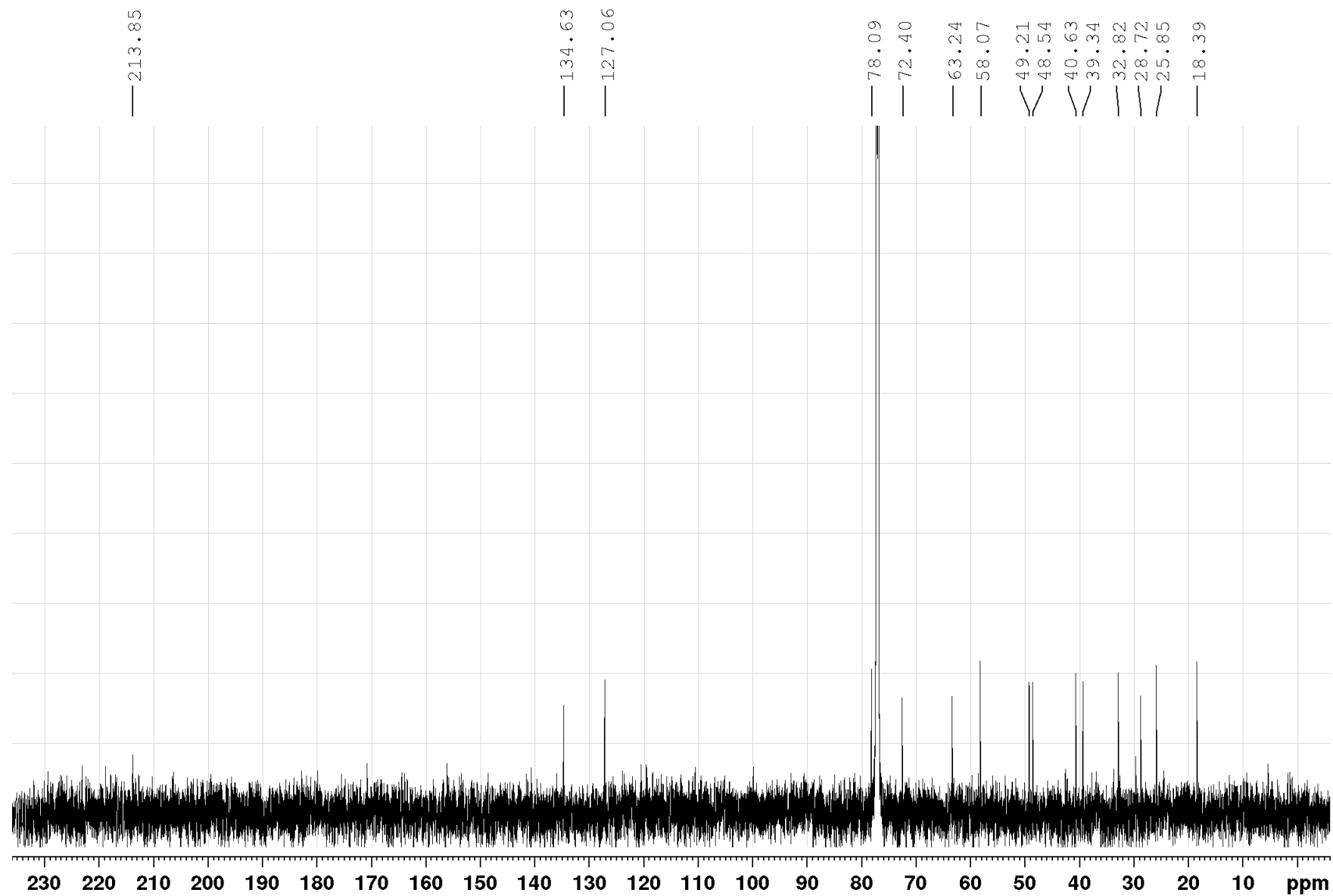
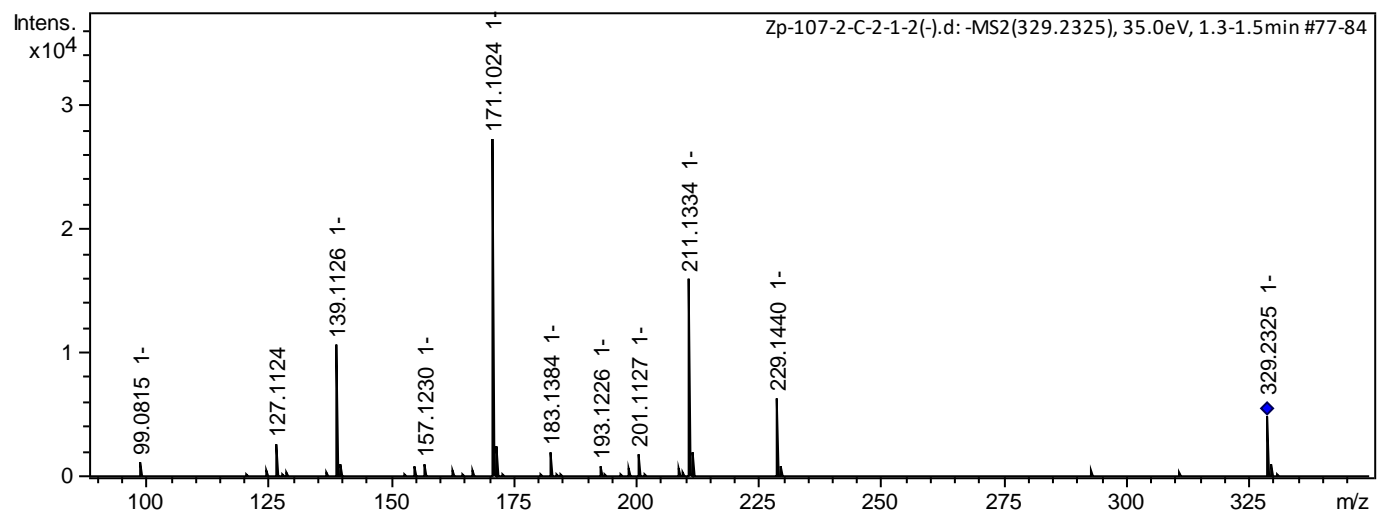
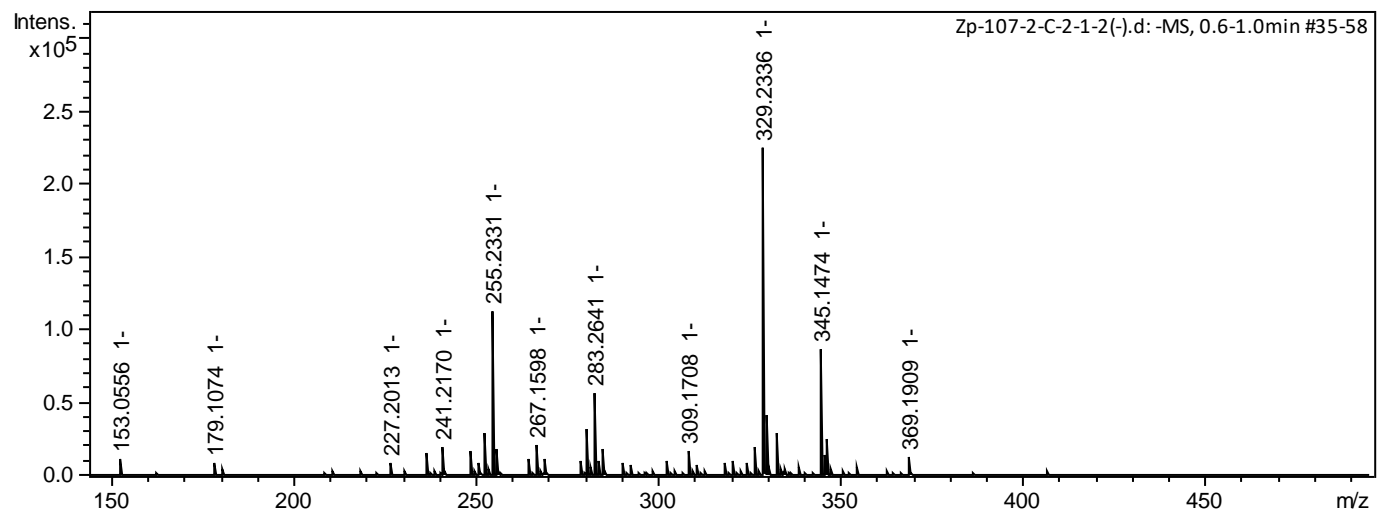
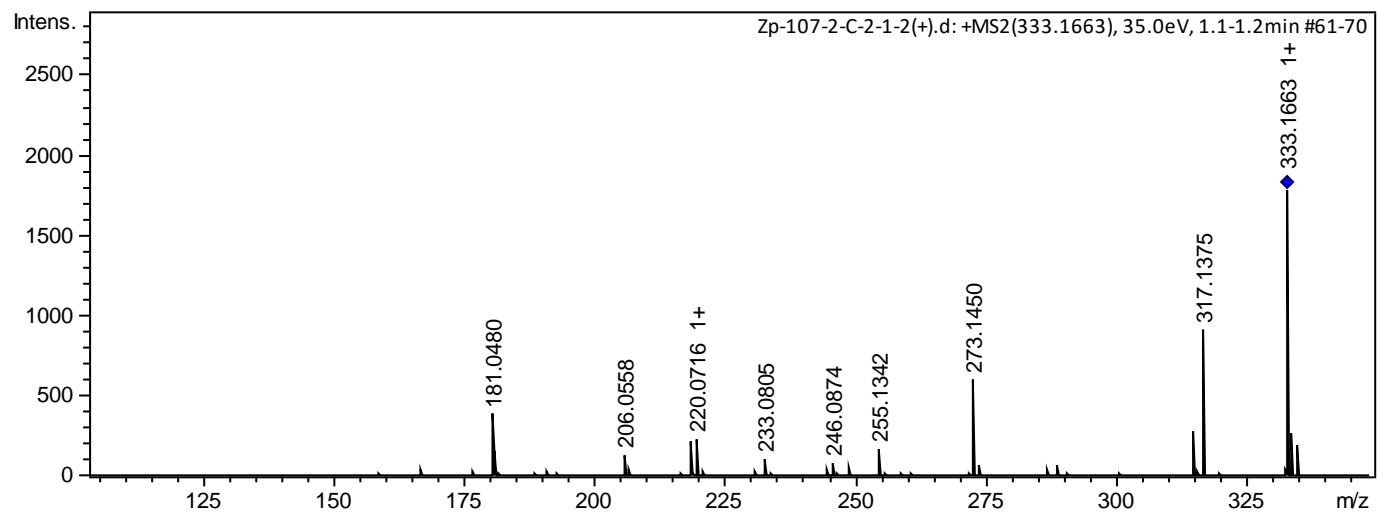
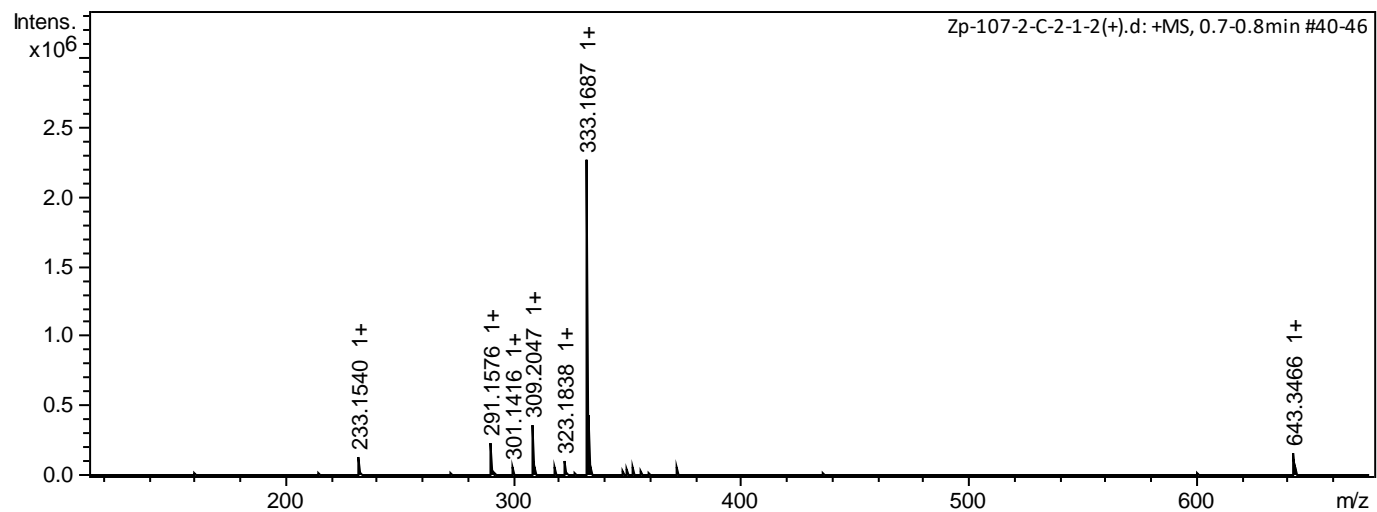


Figure S89. HRESIMS for 14





	meas	calc	Δ (ppm)
[M-H] ⁻	309,1708	309,1707	-0,3
[M+Na] ⁺	333,1687	333,1672	-4,5

Figure S90. ^1H NMR spectrum of 14 measured at 700 MHz in CDCl_3

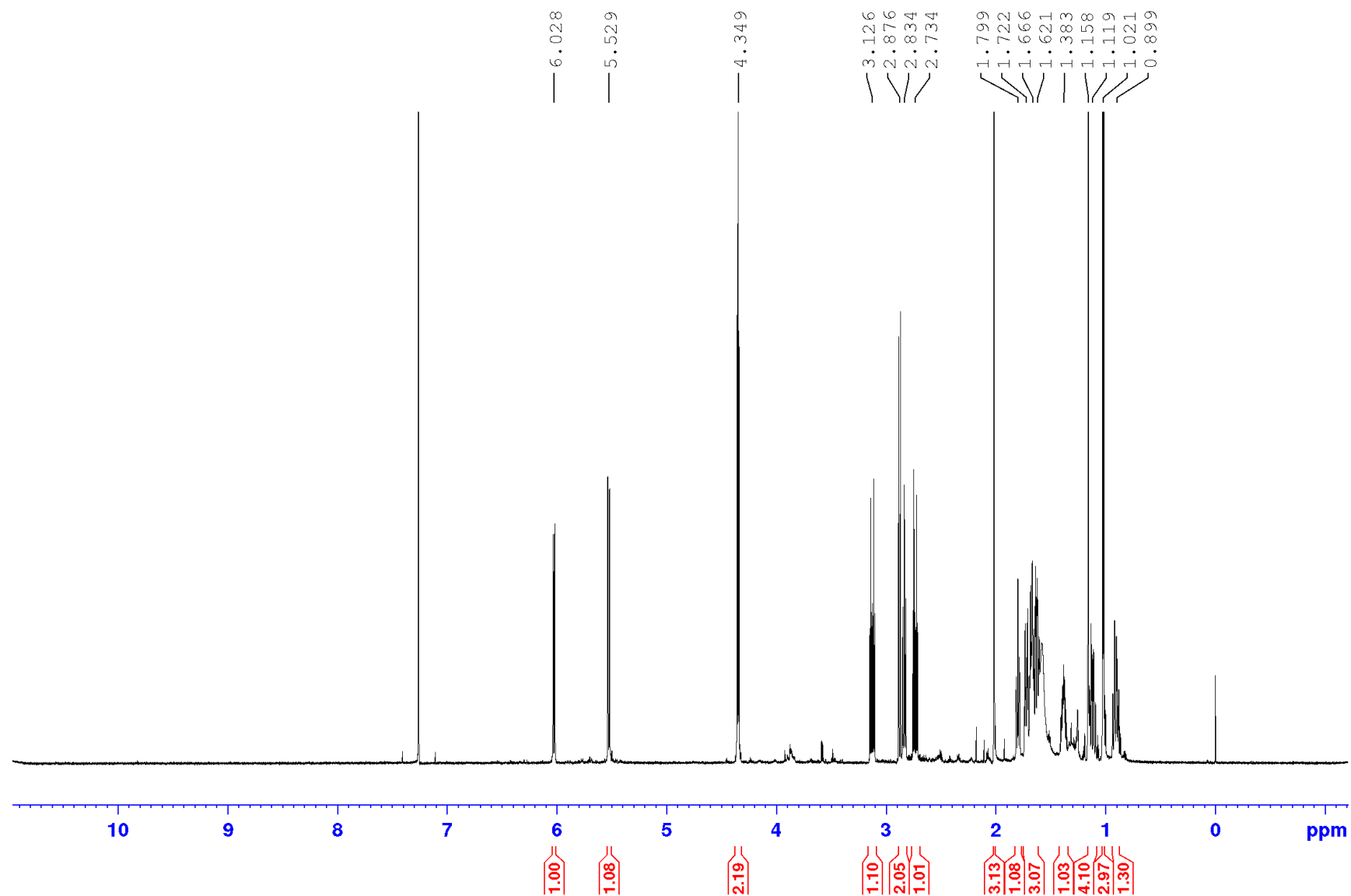


Figure S91. ^{13}C NMR spectrum of 14 measured at 176 MHz in CDCl_3

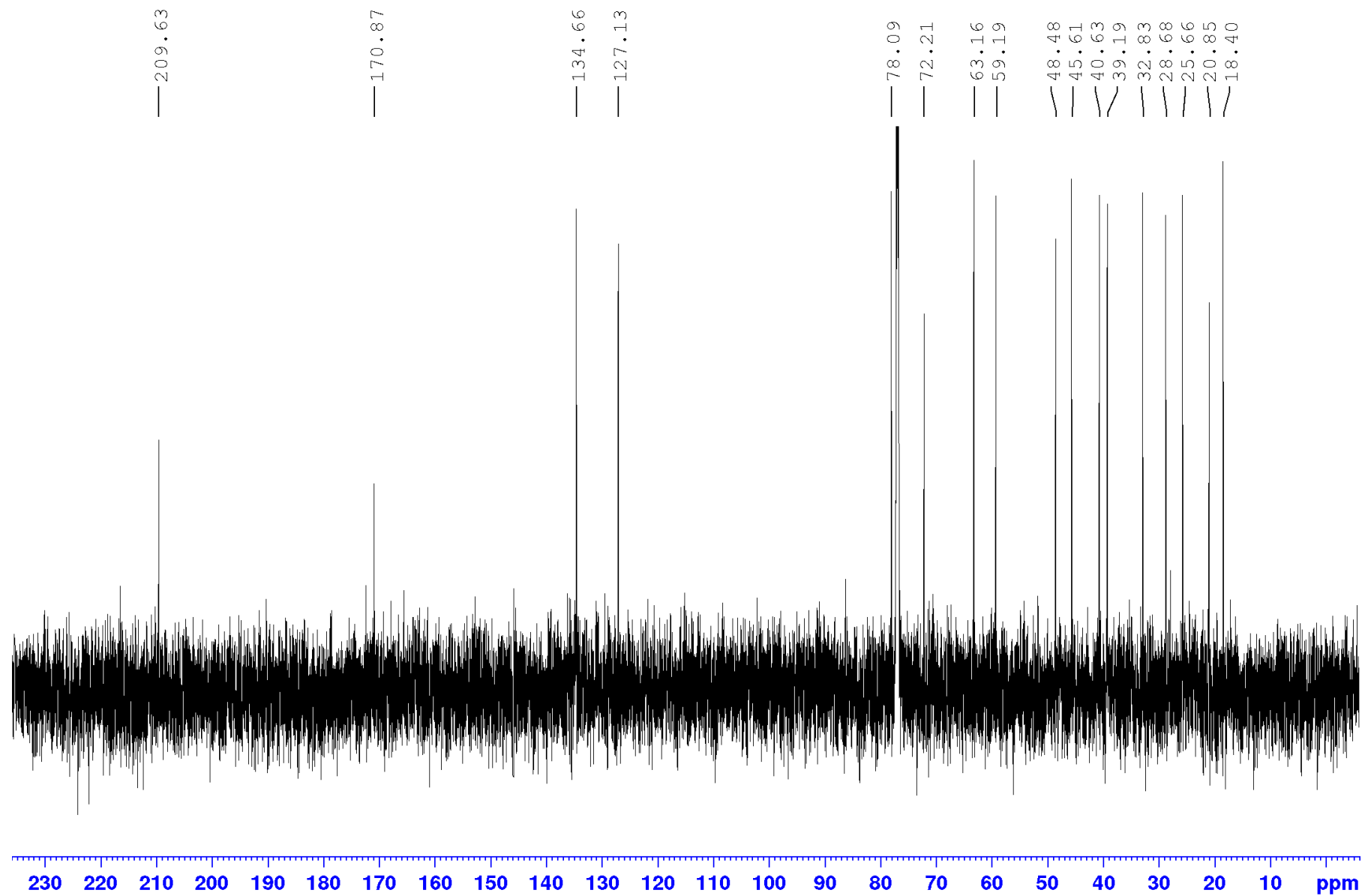
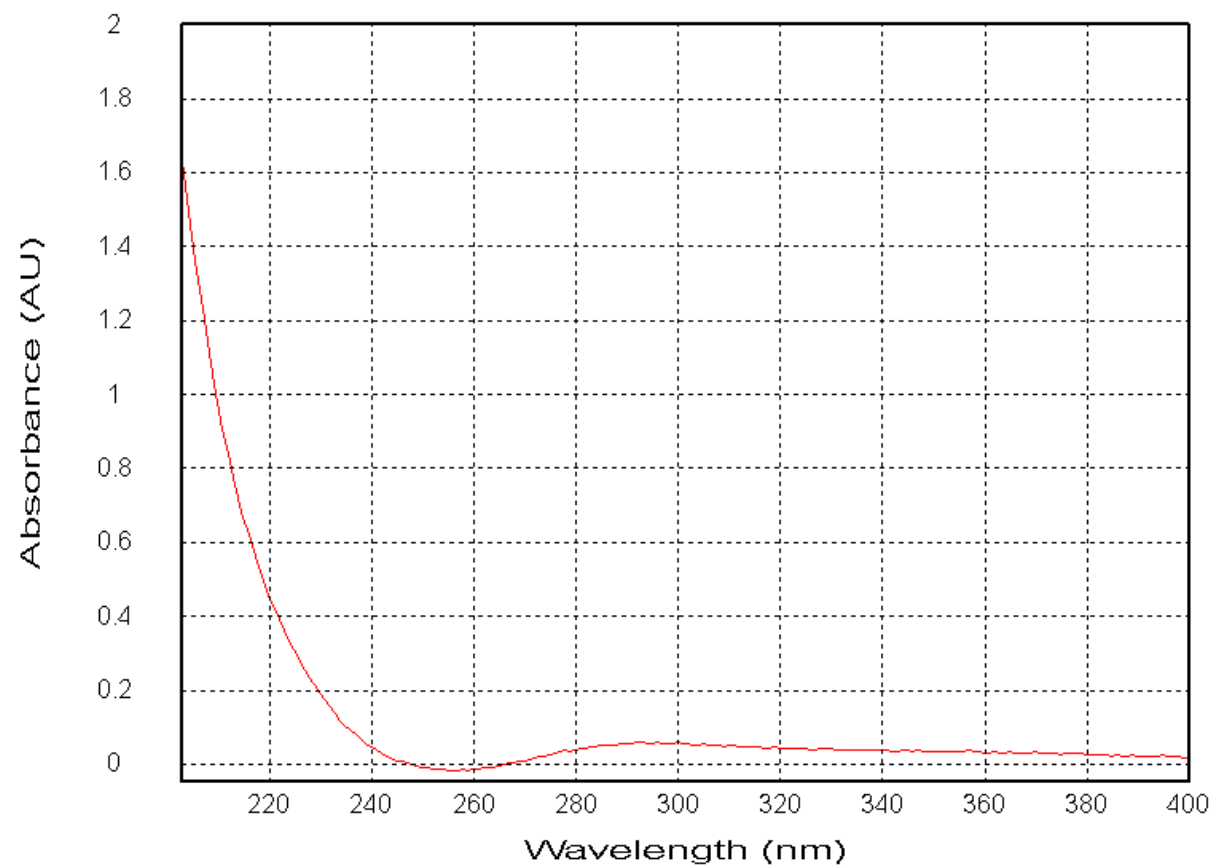


Figure S92. UV and CD data for 1 in MeOH



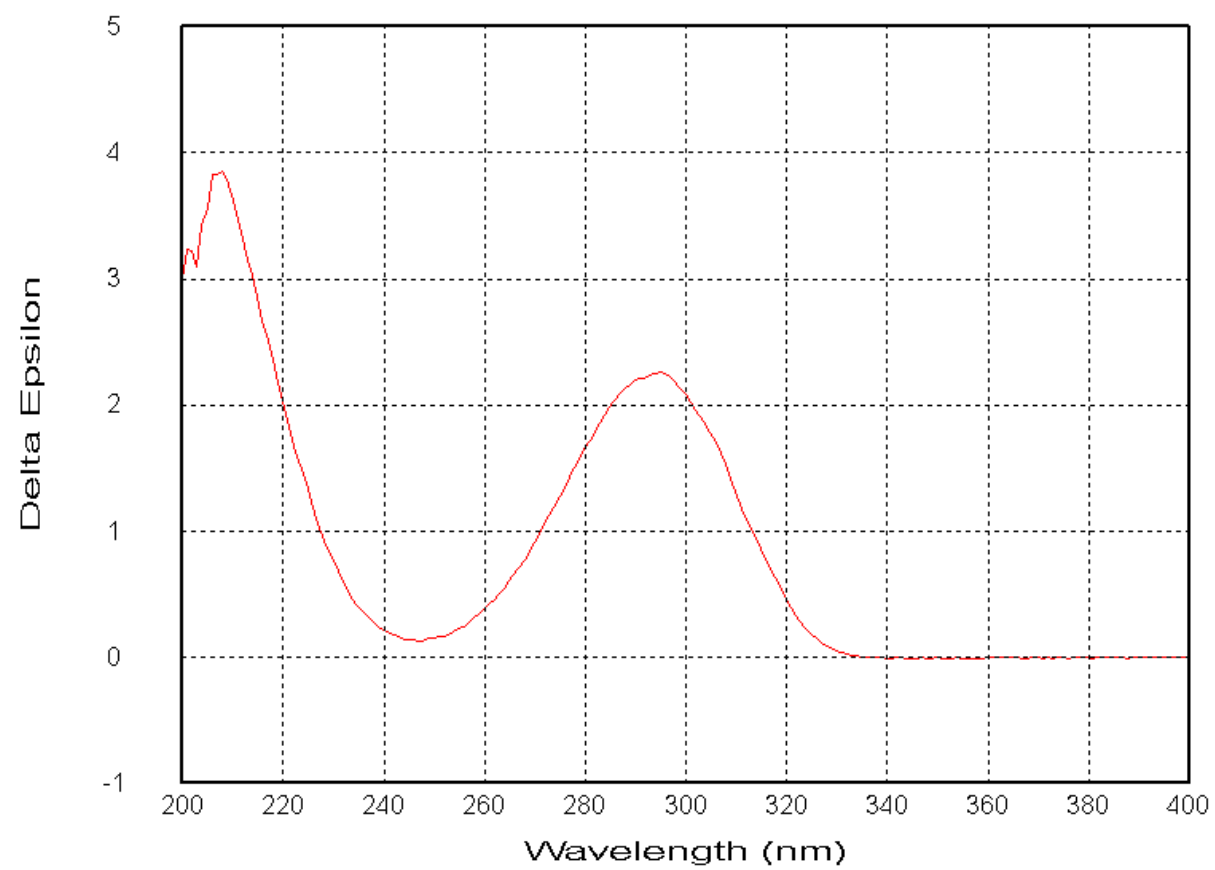
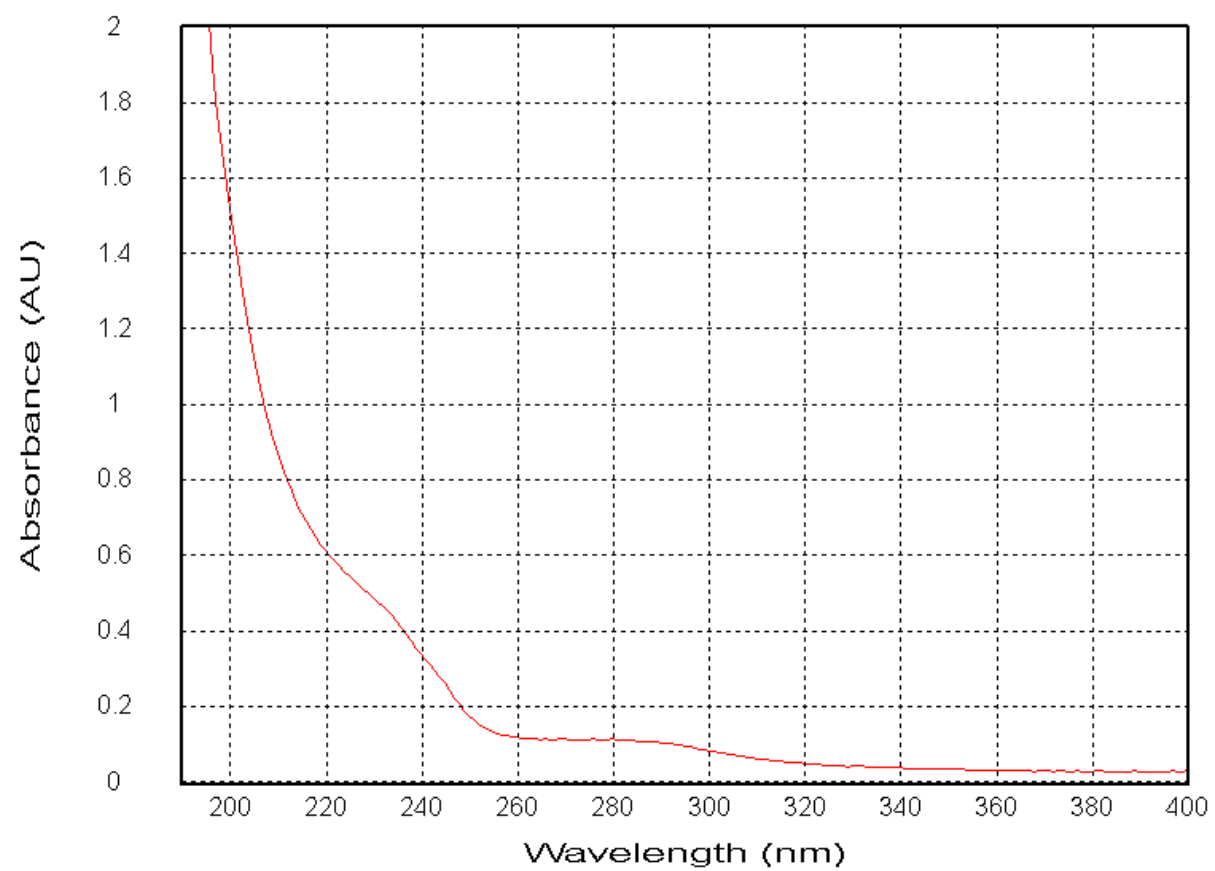


Figure S93. UV and CD data for 2 in MeOH



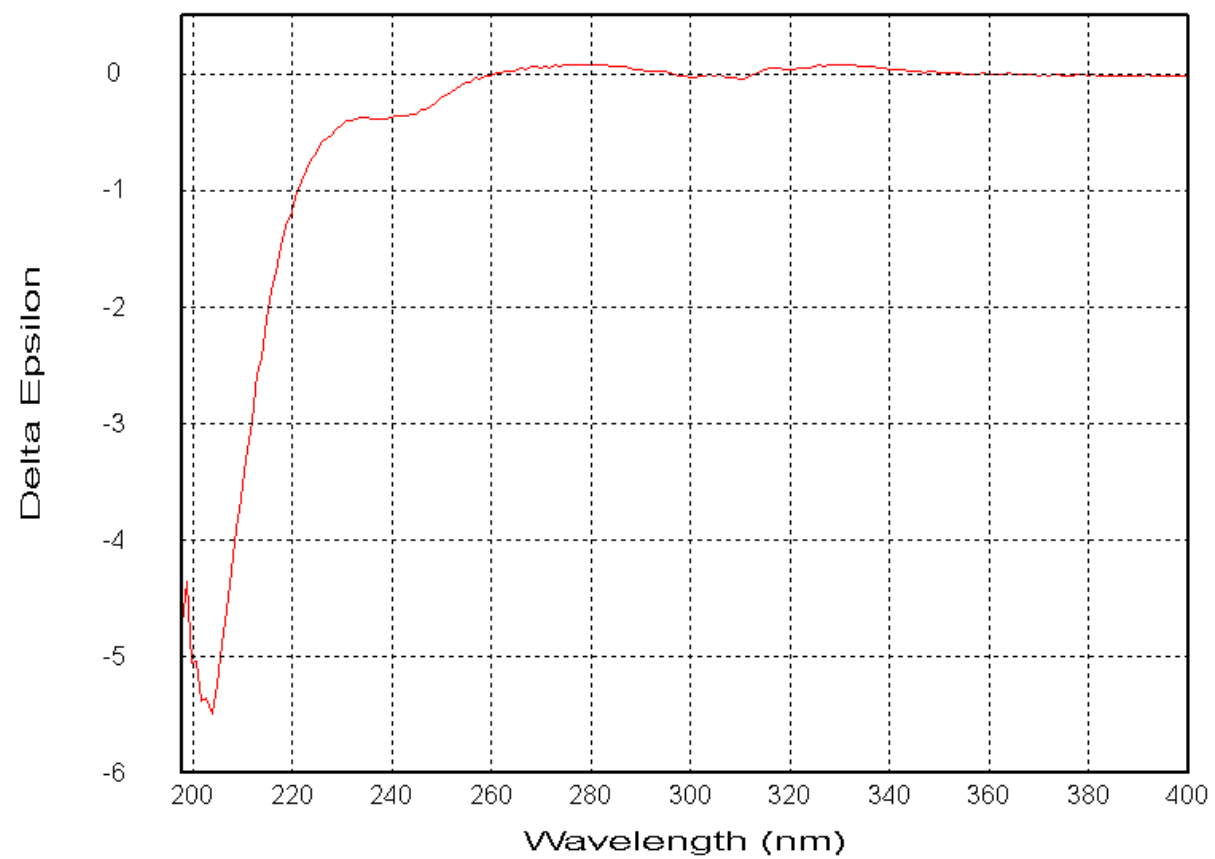
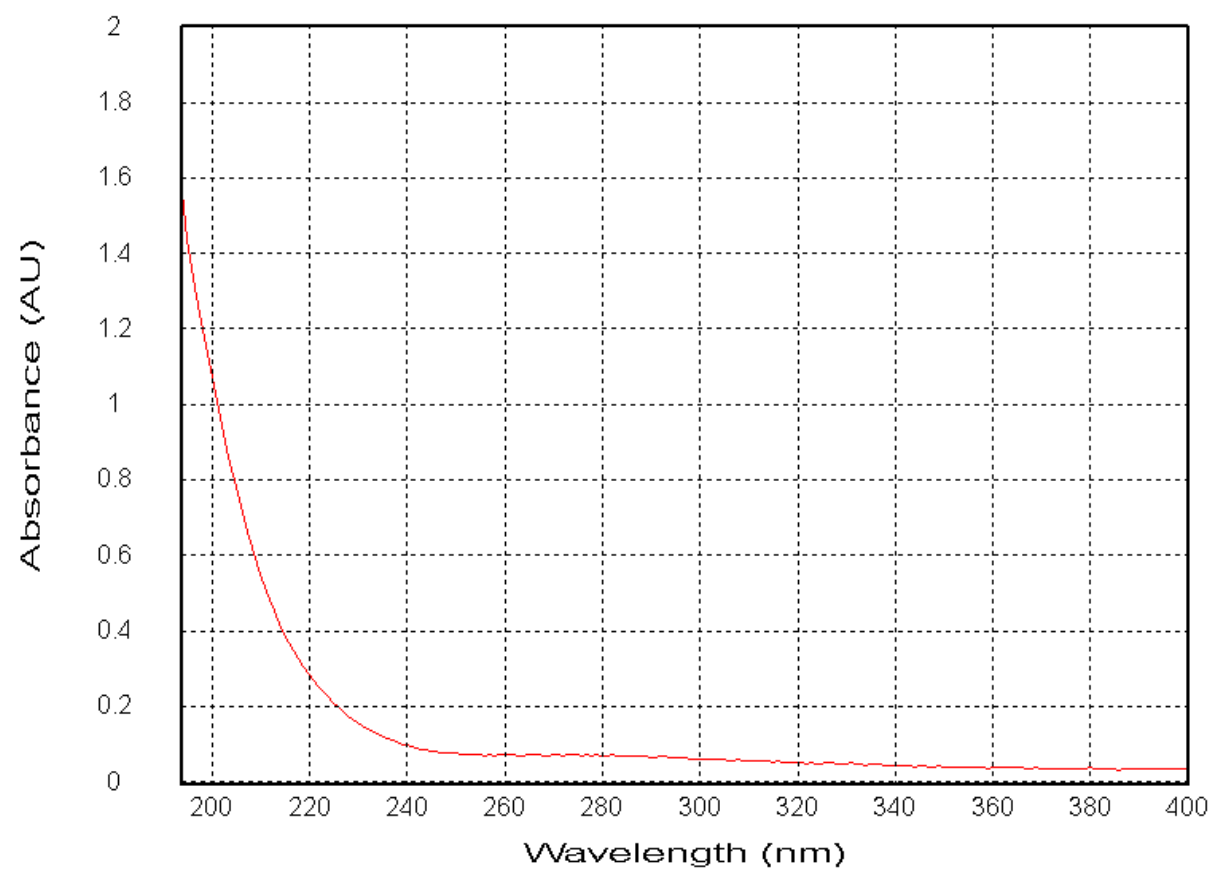


Figure S94. UV and CD data for 3 in MeOH



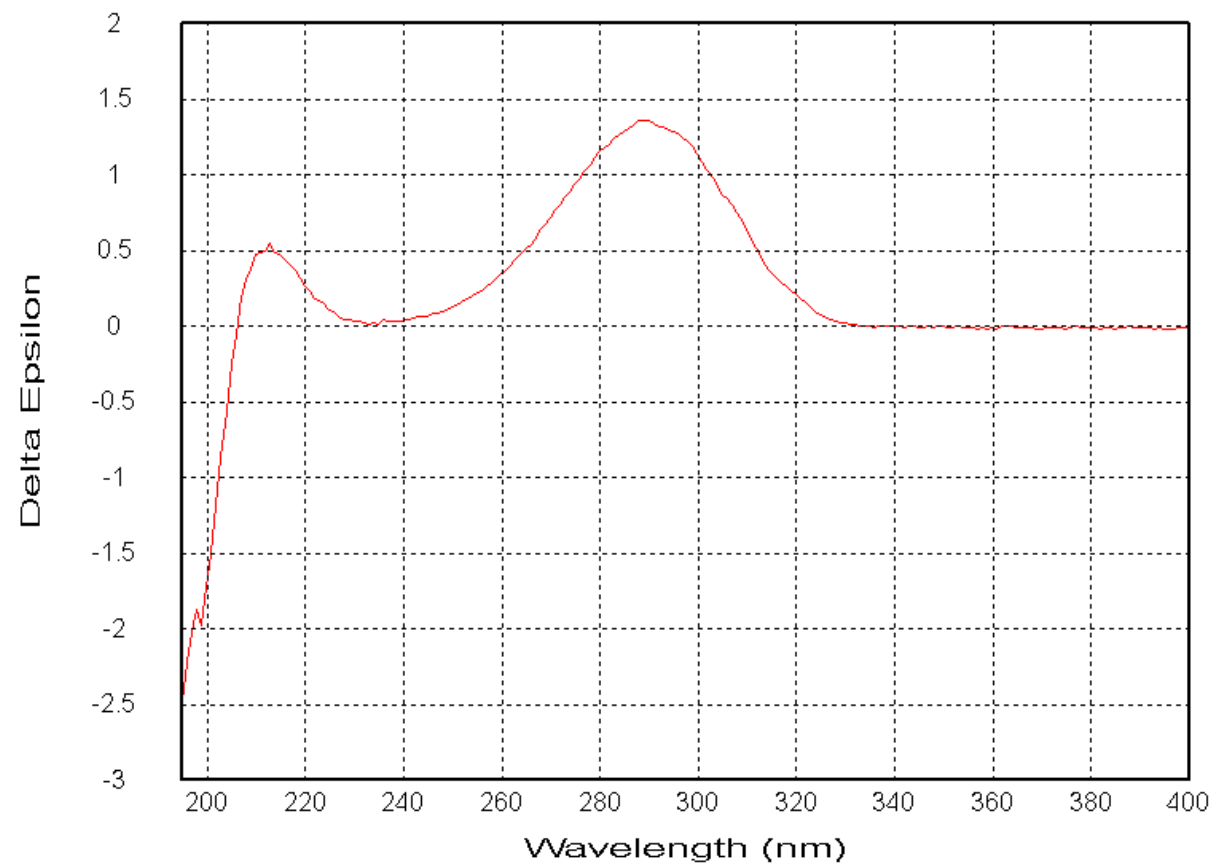
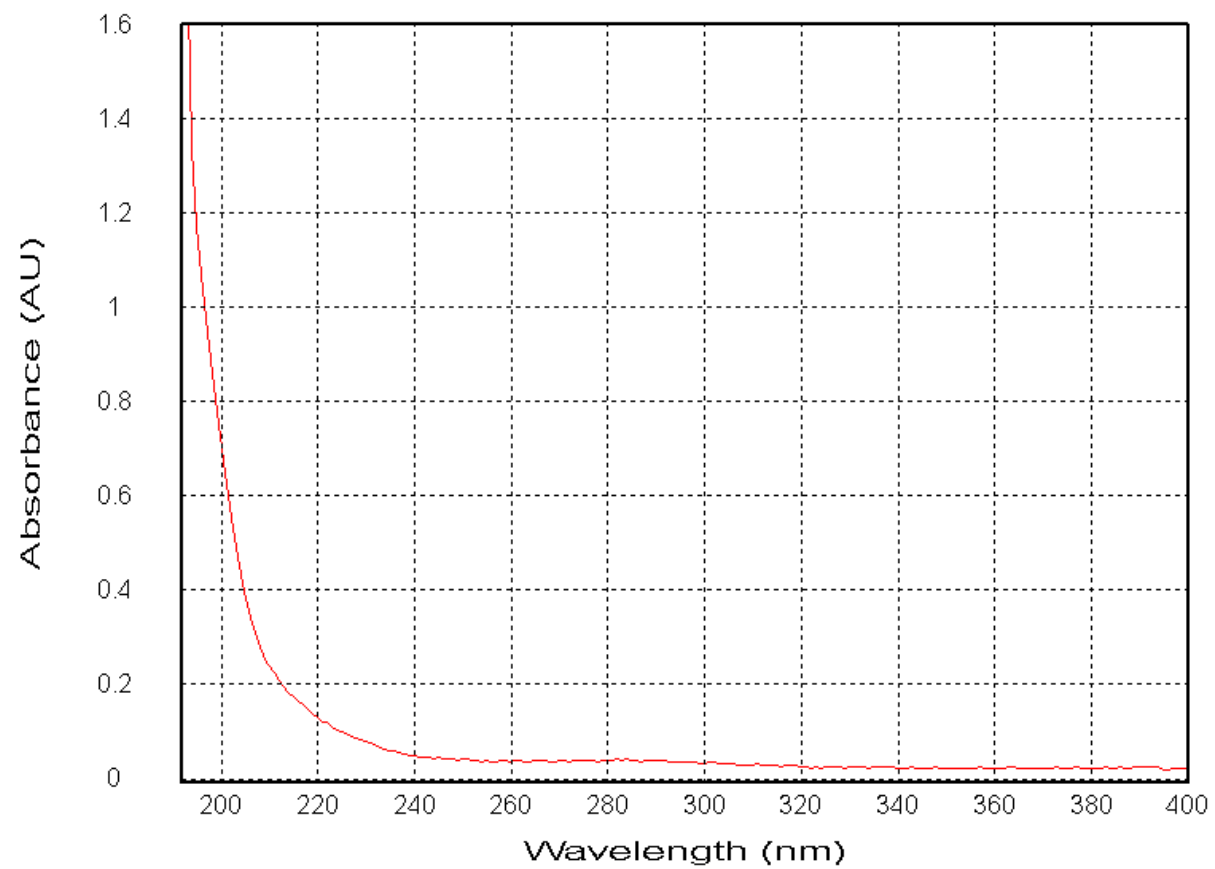


Figure S95. UV and CD data for 4 in MeOH



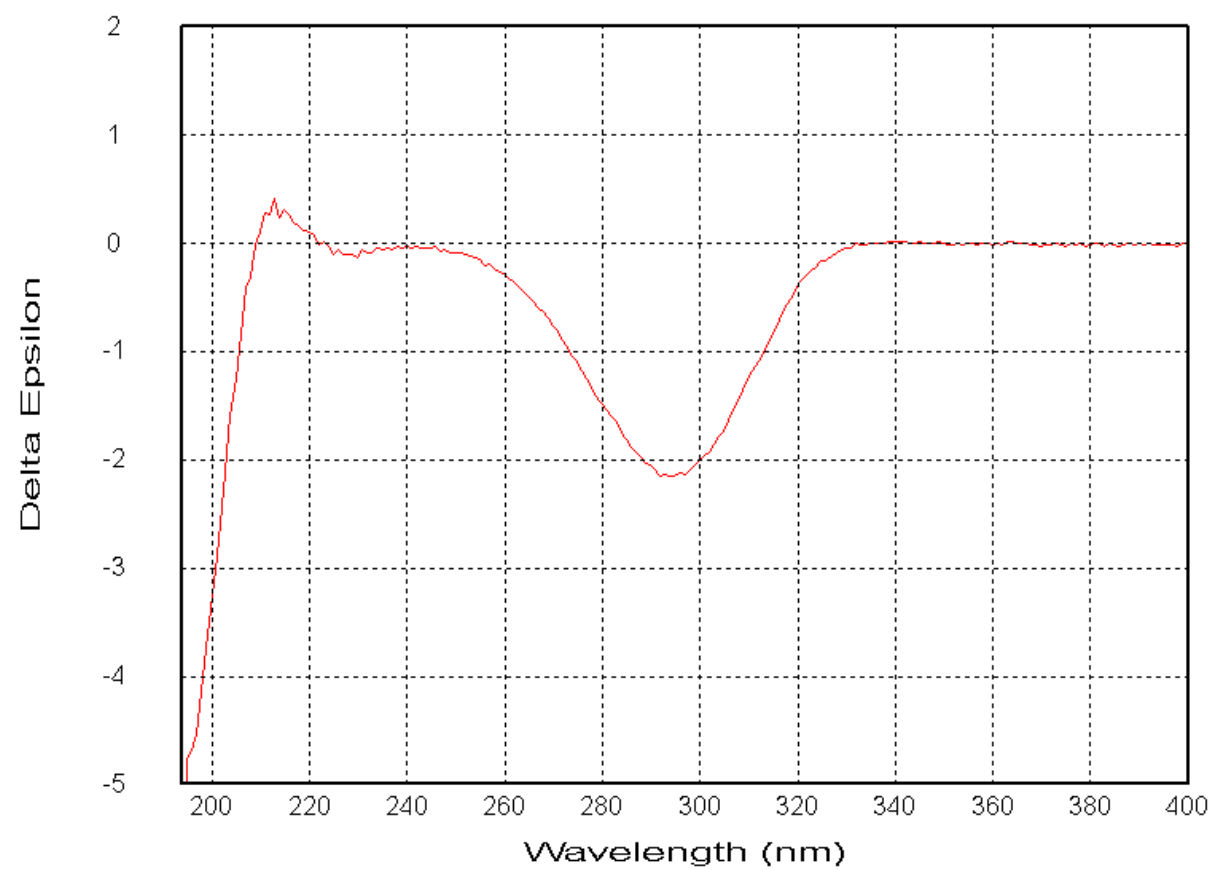
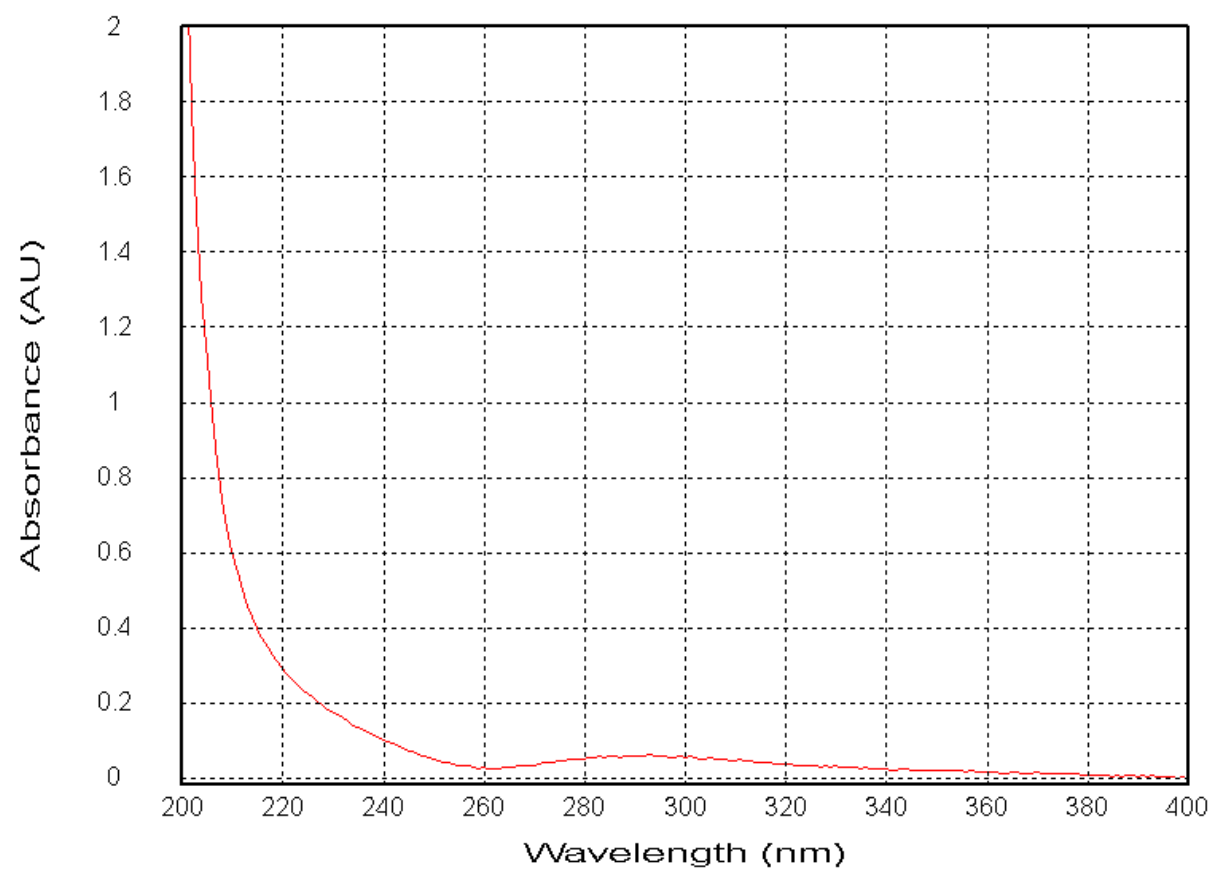


Figure S96. UV and CD data for 5 in MeOH



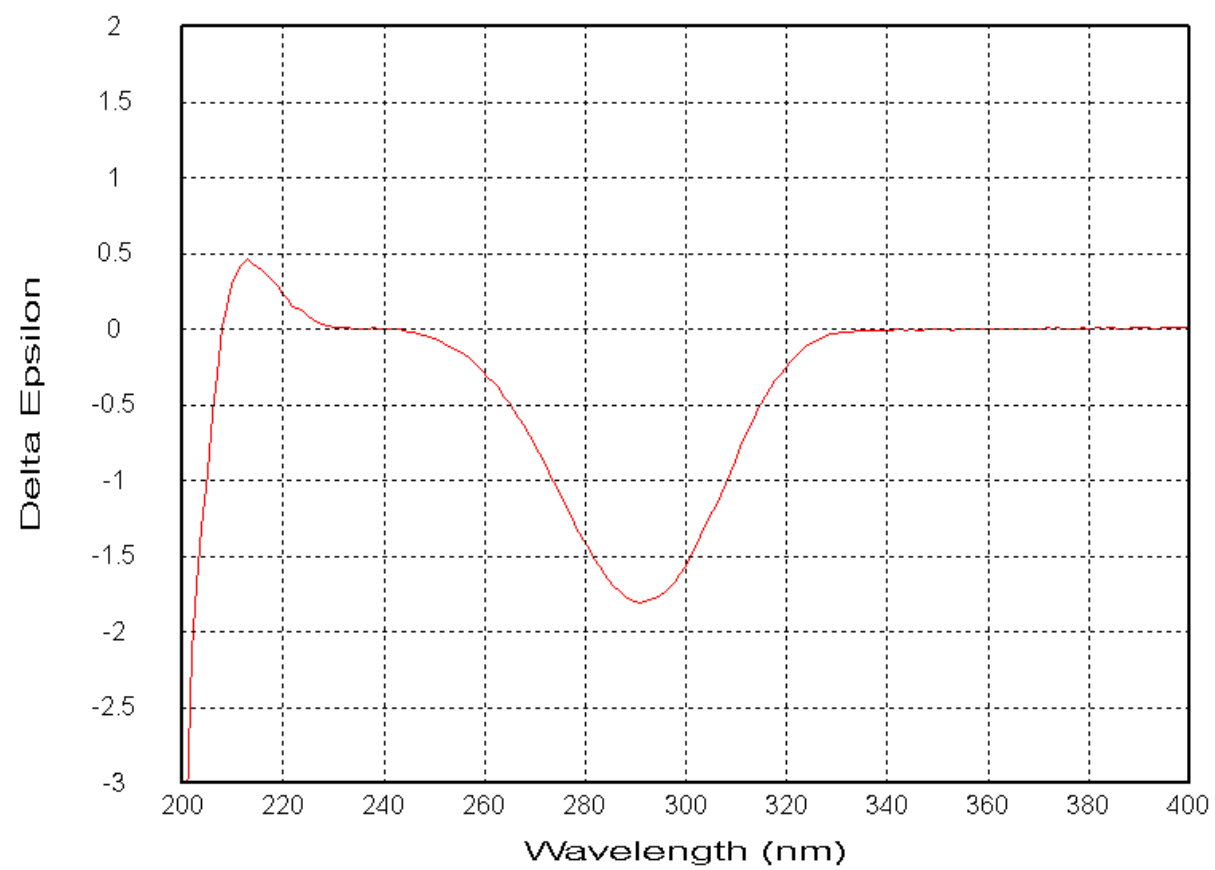
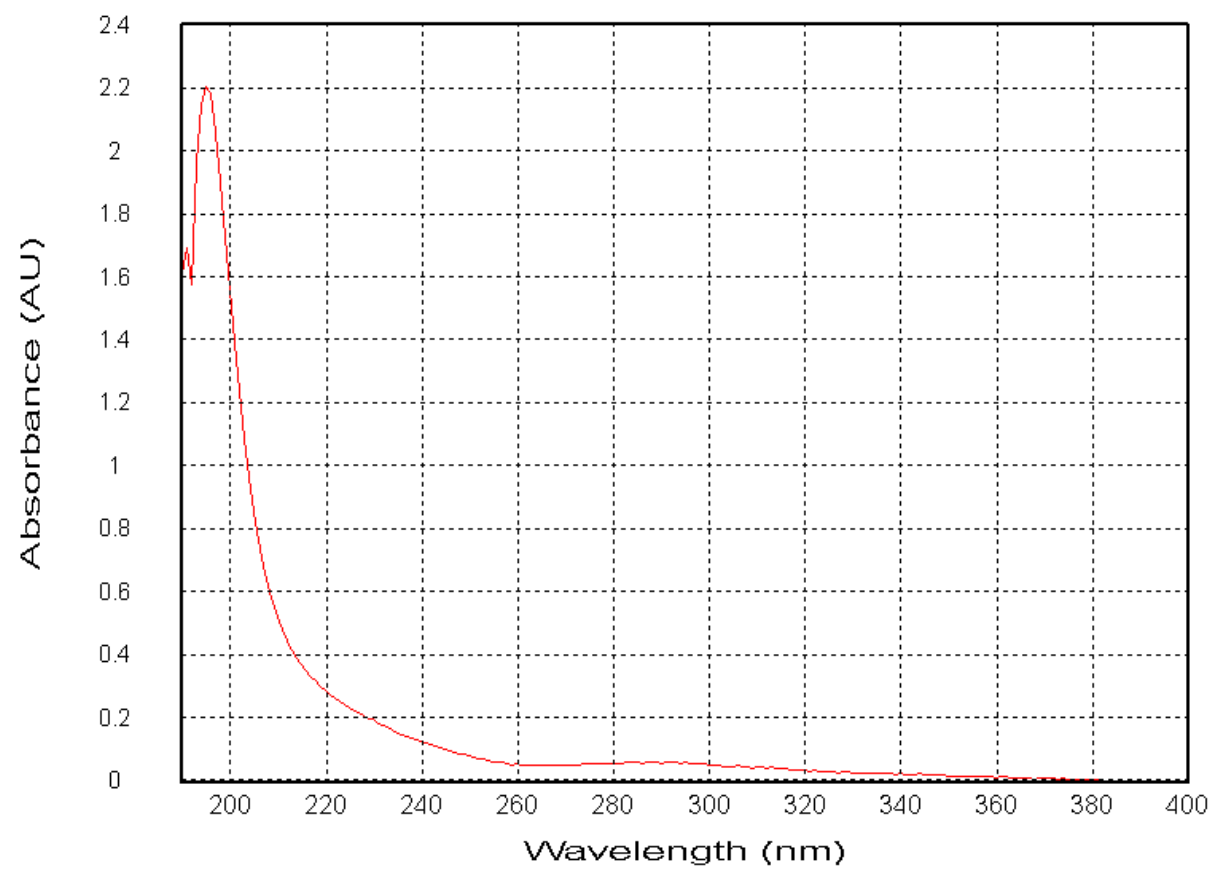


Figure S97. UV and CD data for 6 in MeOH



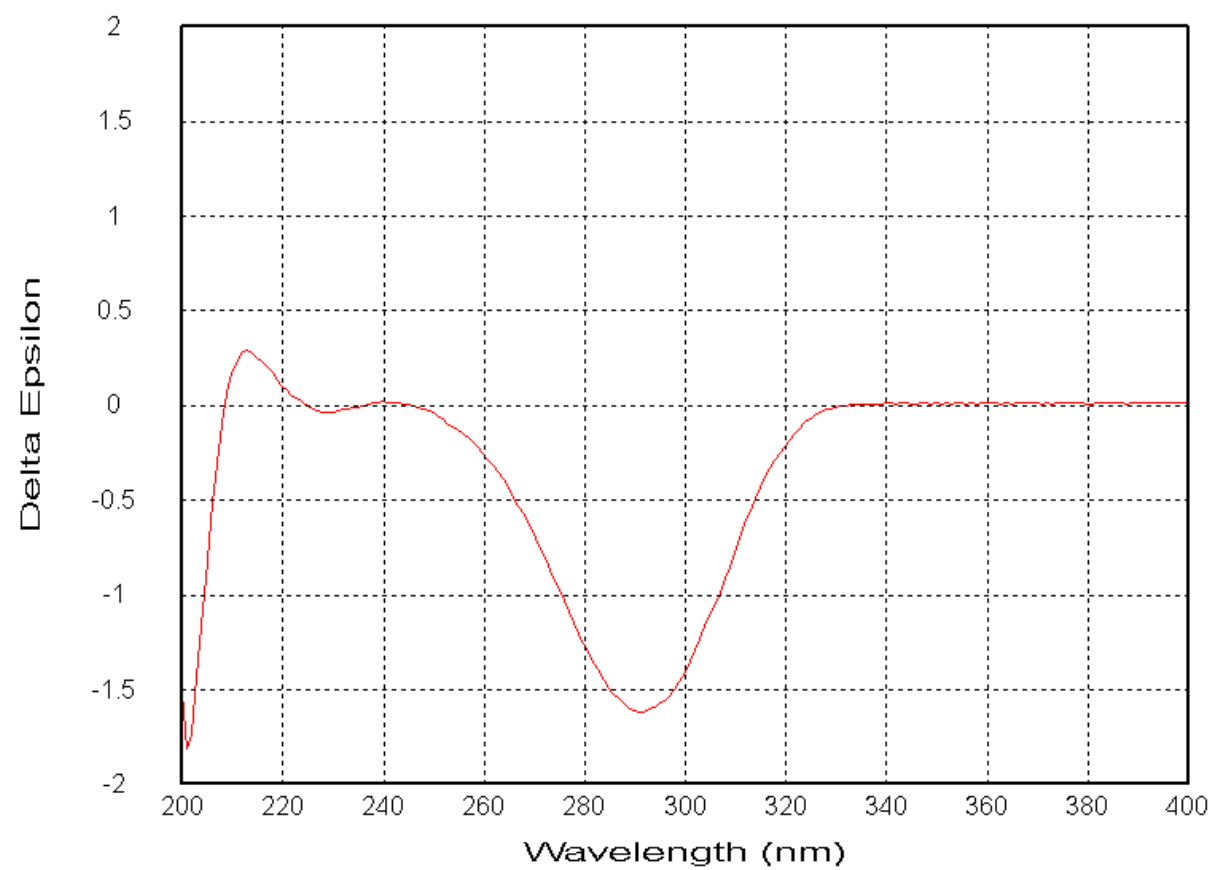
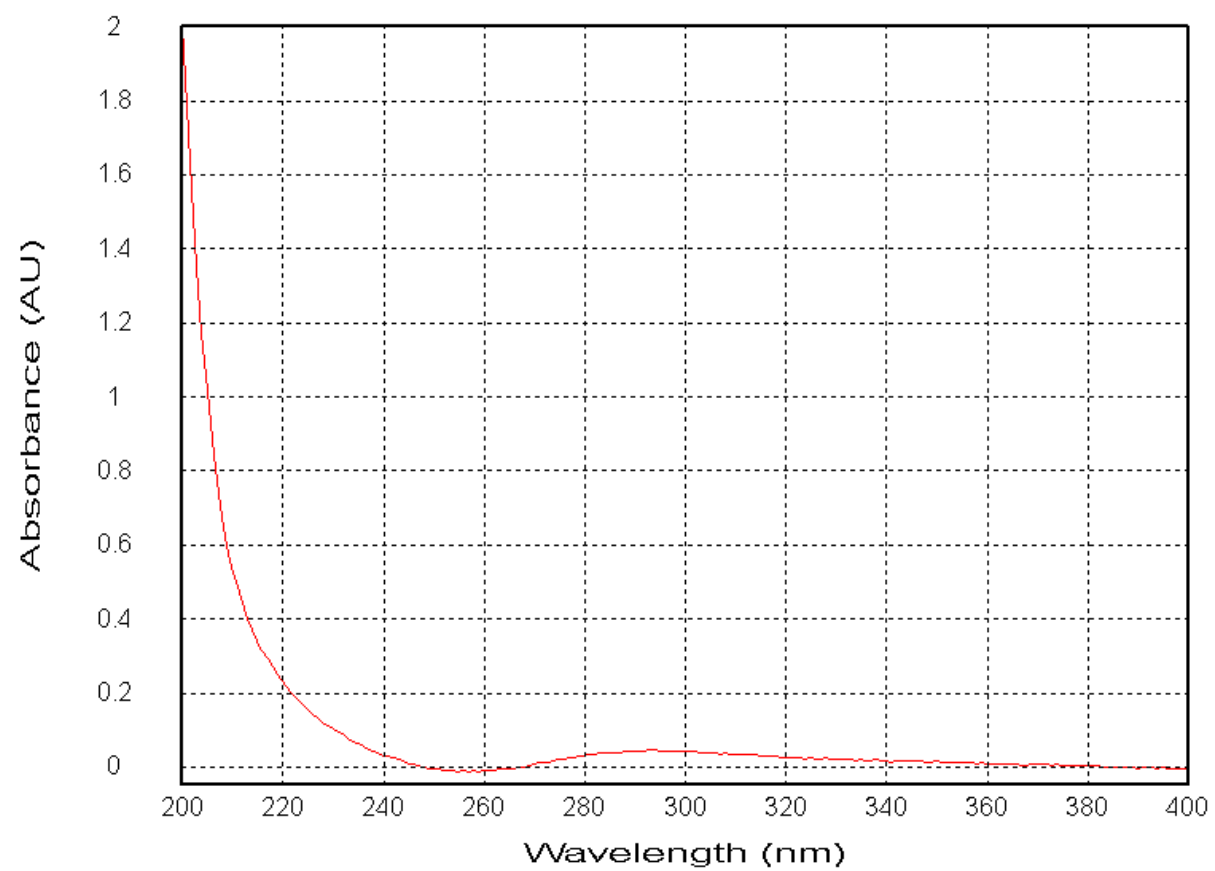


Figure S98. UV and CD data for 7 in MeOH



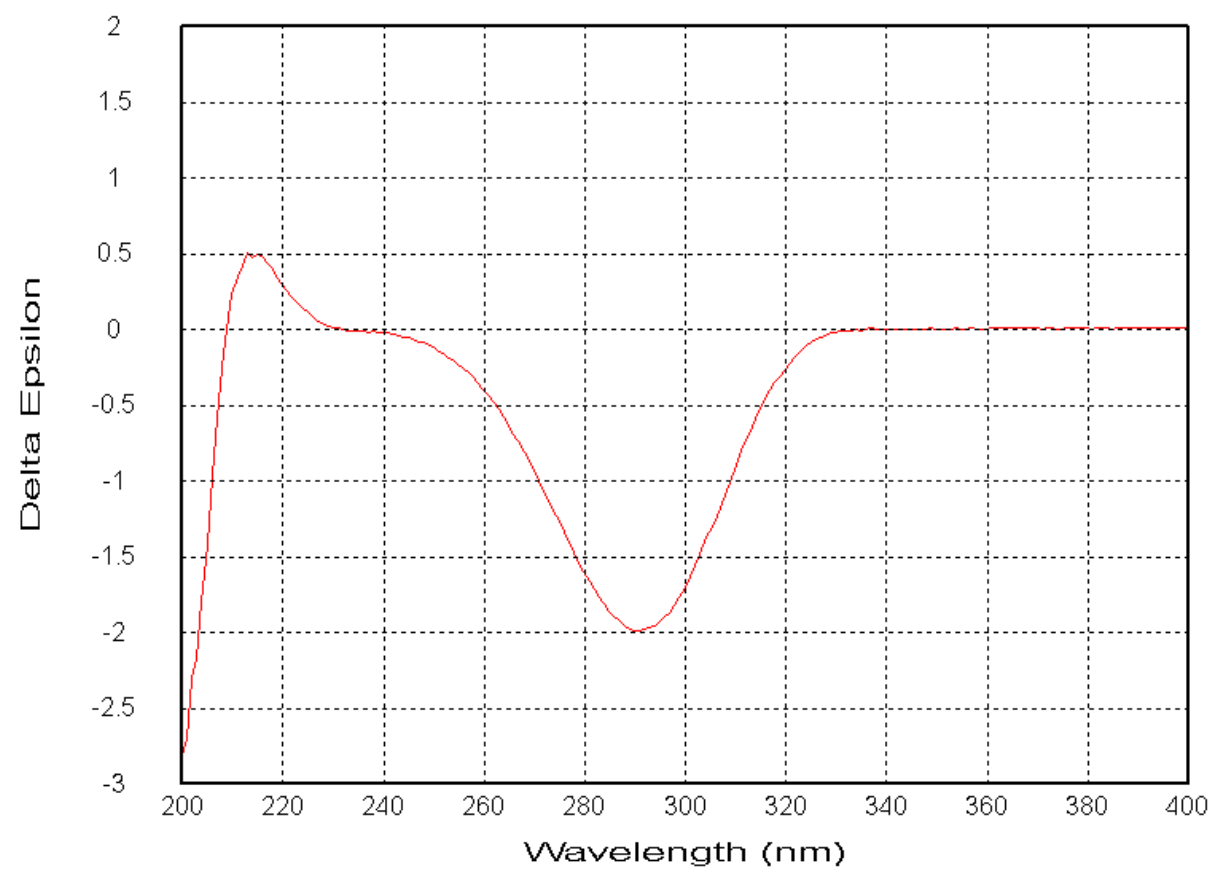
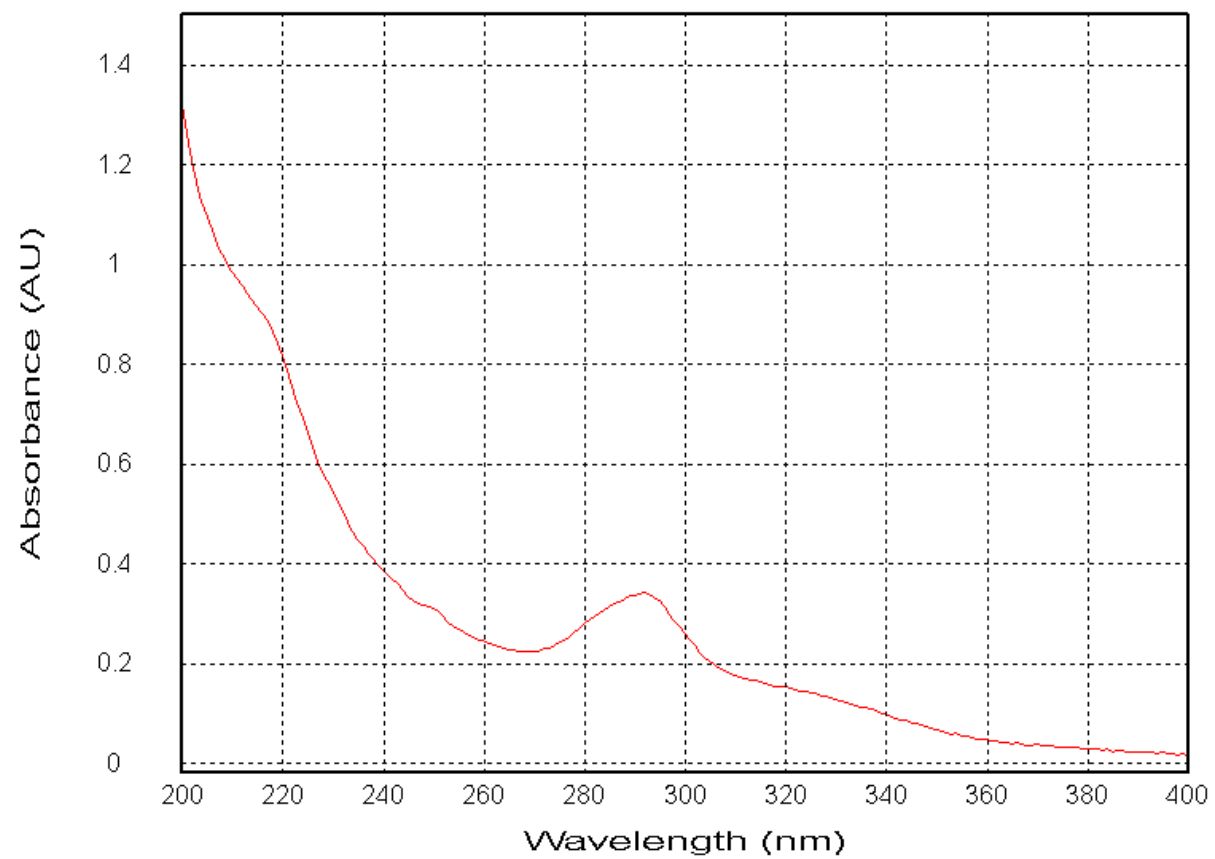


Figure S99. UV and CD data for 8 in MeOH



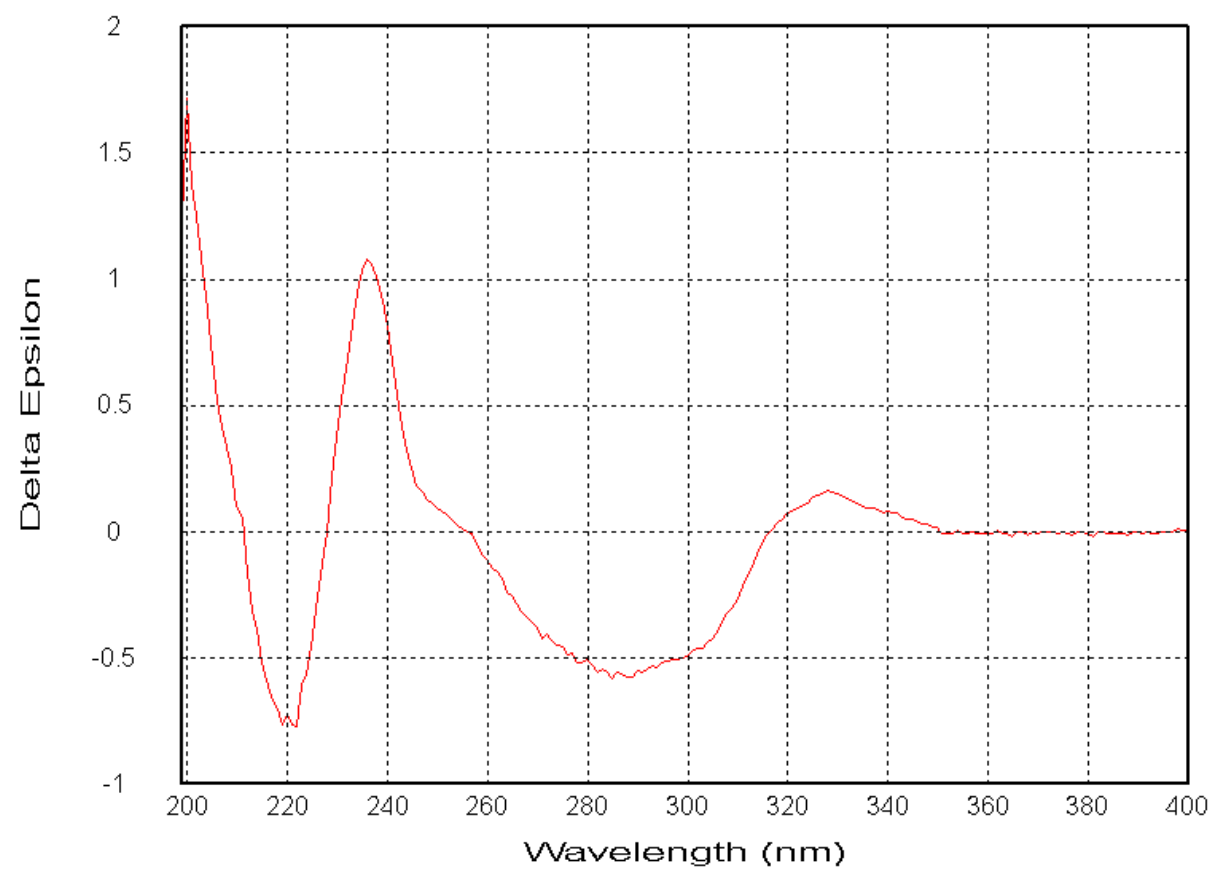
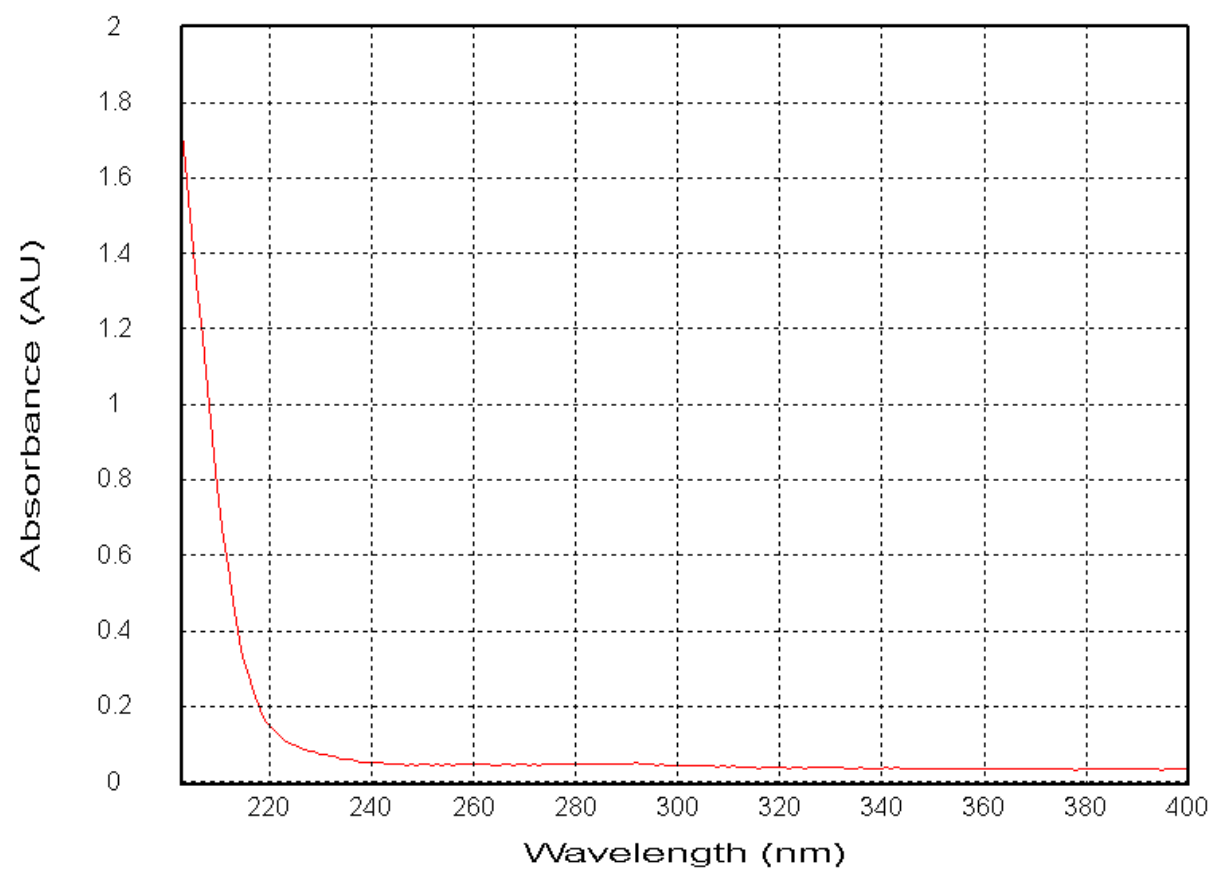


Figure S100. UV and CD data for 9 in MeOH



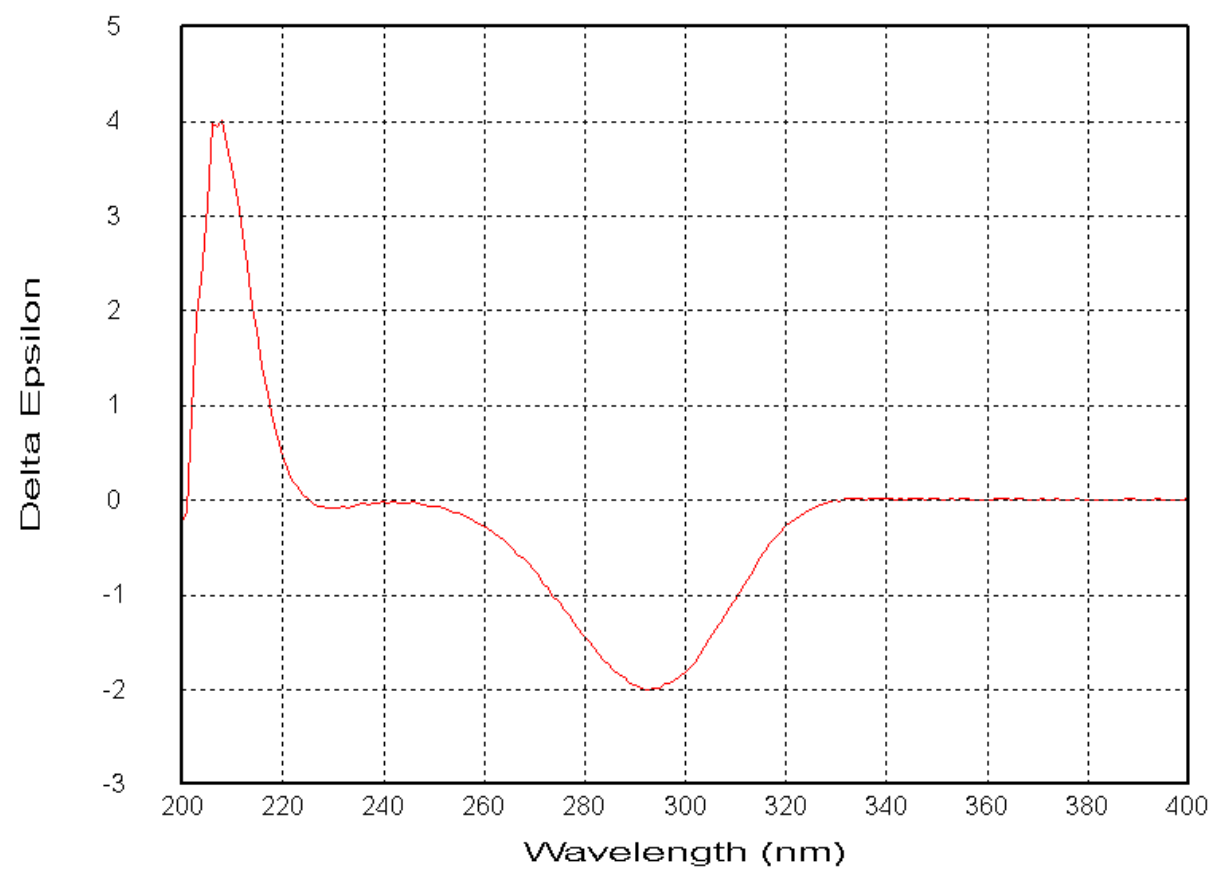
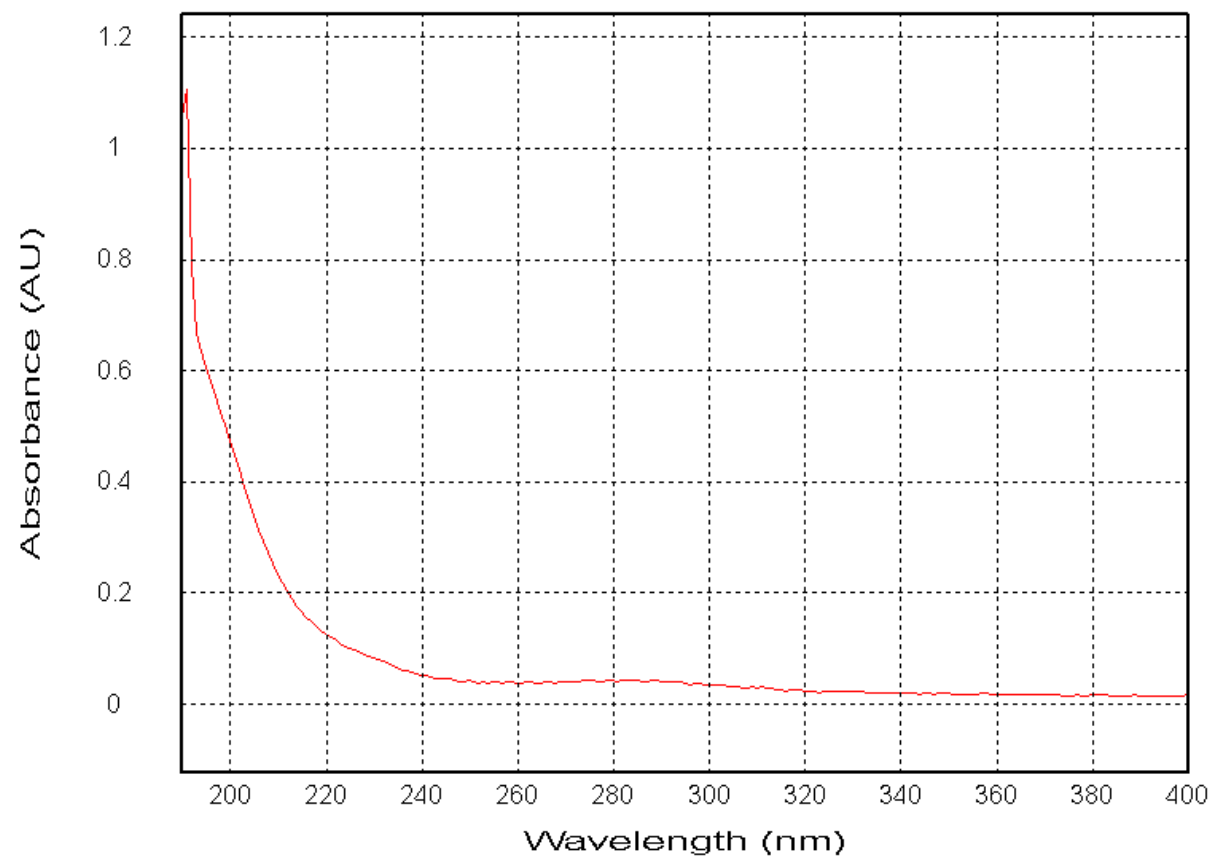


Figure S101. UV and CD data for 10 in MeOH



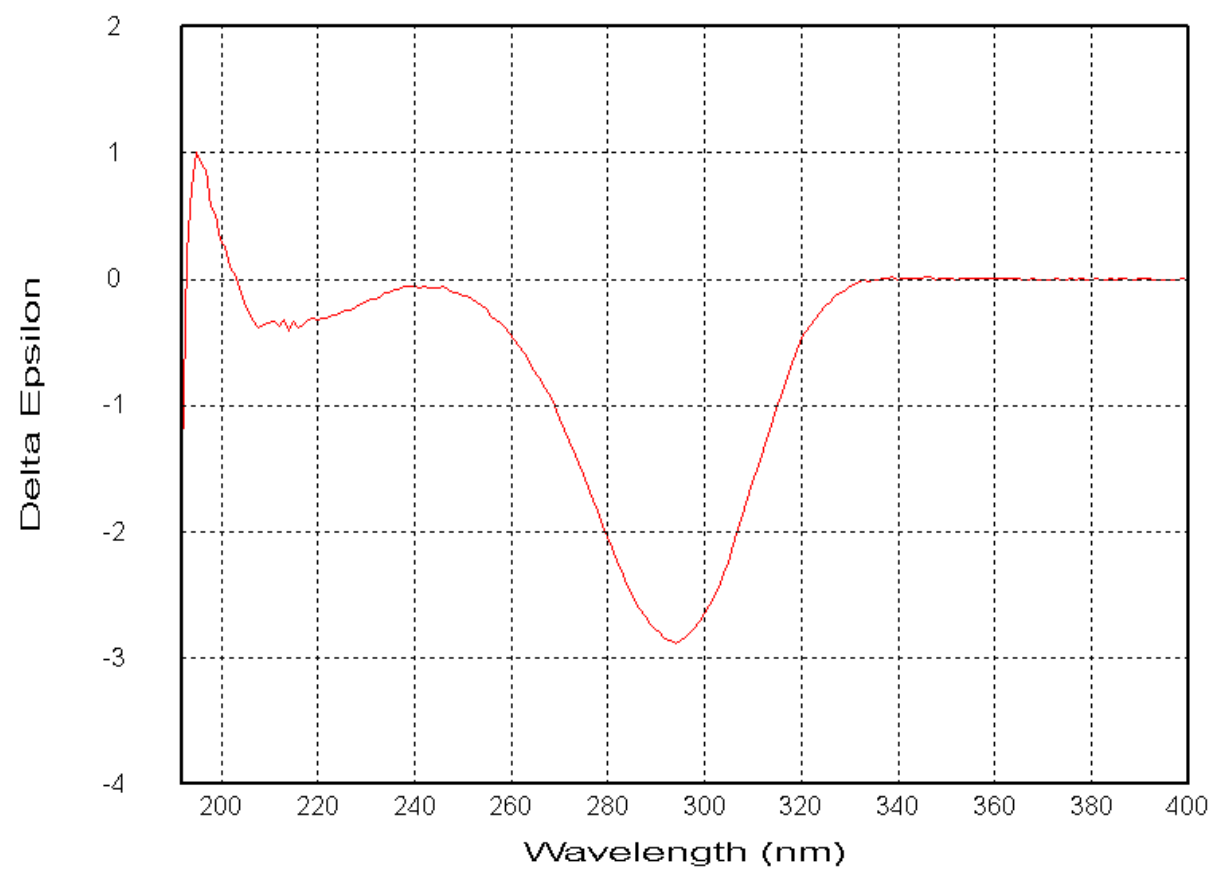


Figure S102. ¹H NMR spectrum of 7b ((S)-MTPA ester) measured at 700 MHz in CDCl₃

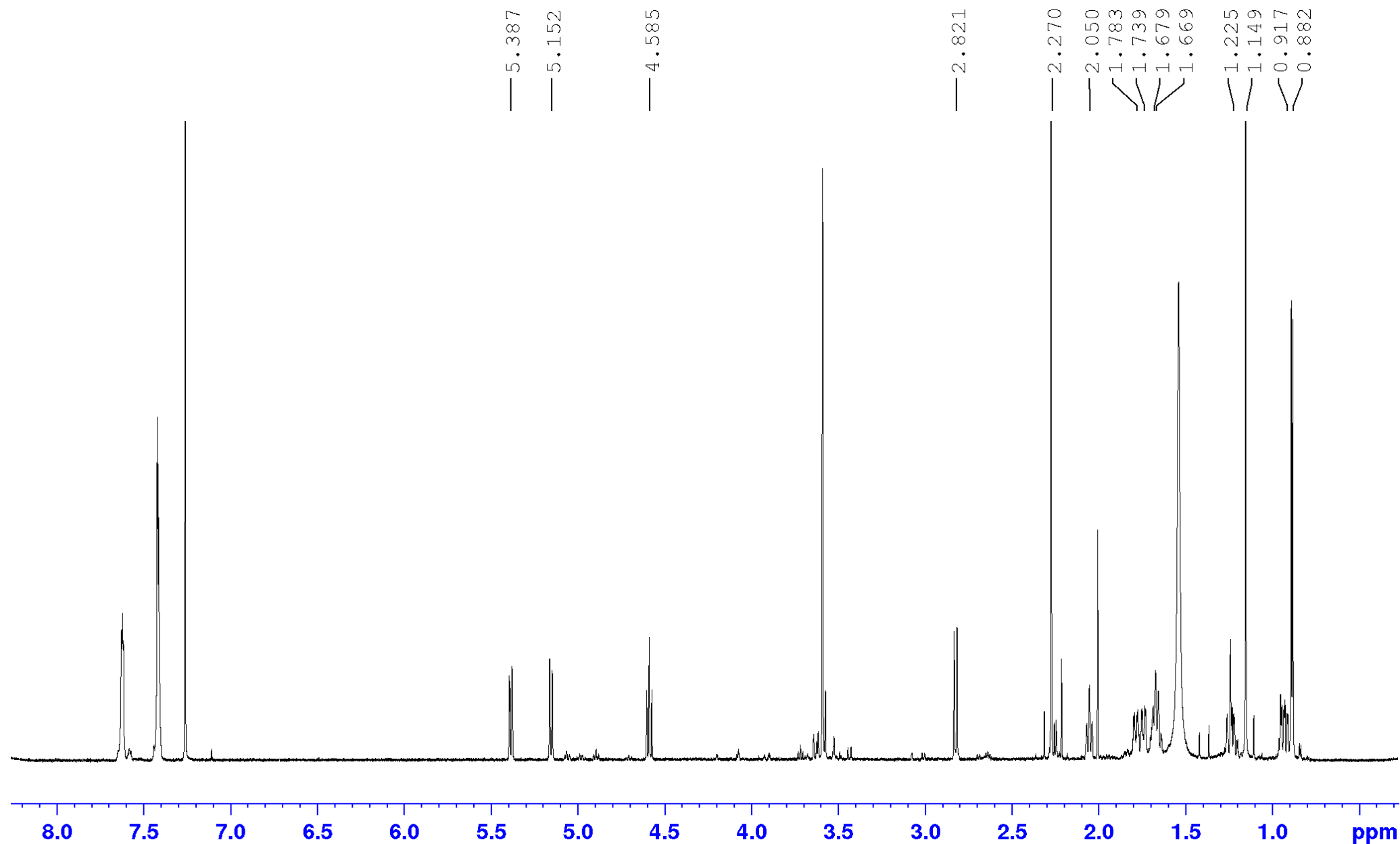


Figure S103. ¹H NMR spectrum of 7a ((R)-MTPA ester) measured at 700 MHz in CDCl₃

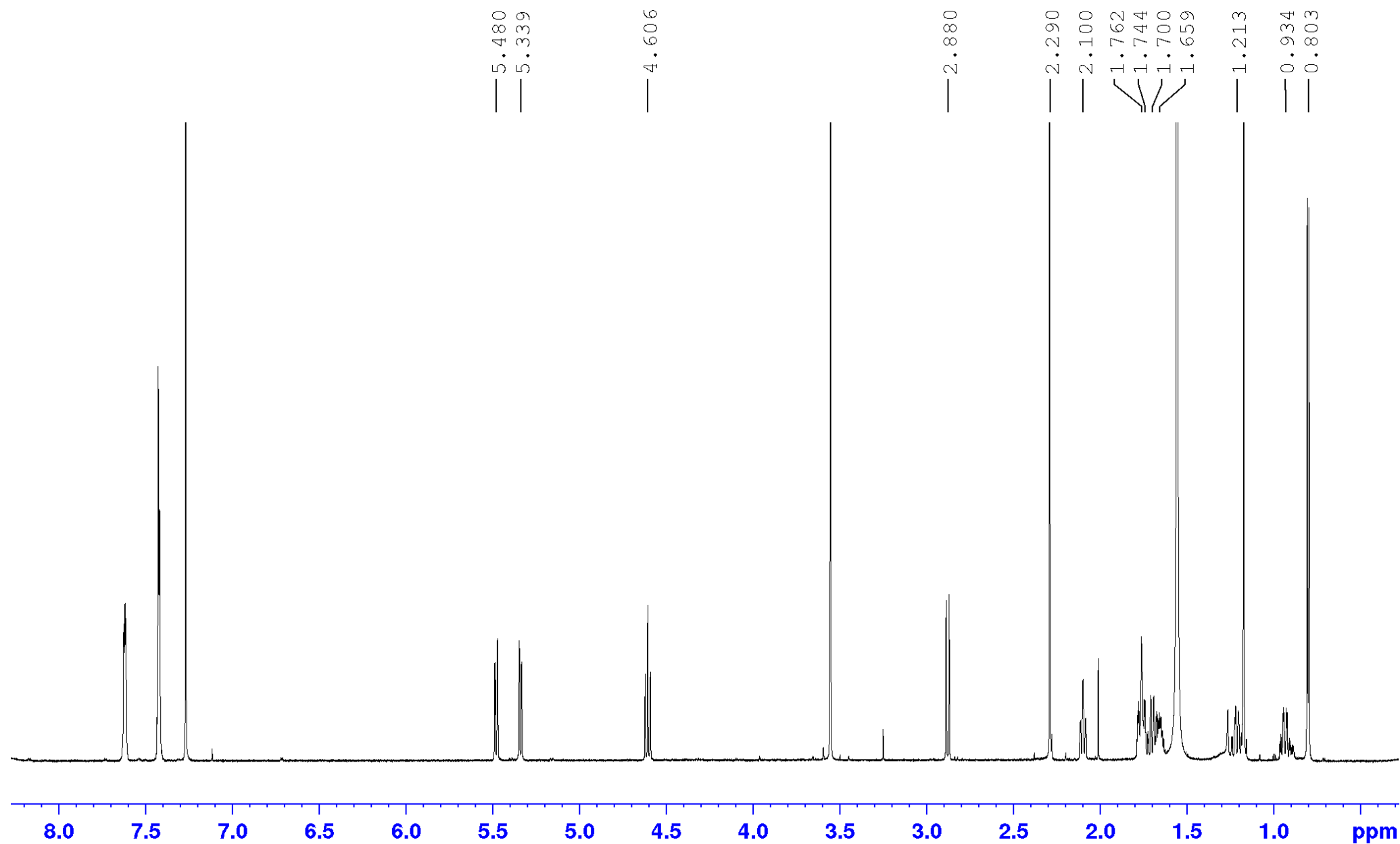
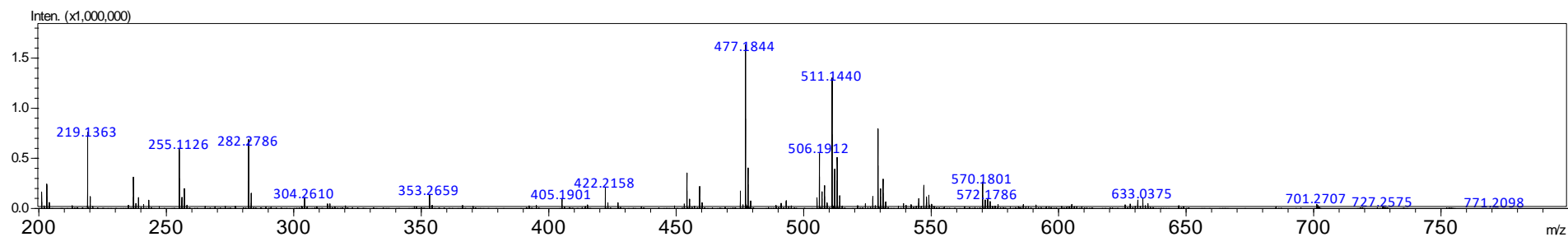
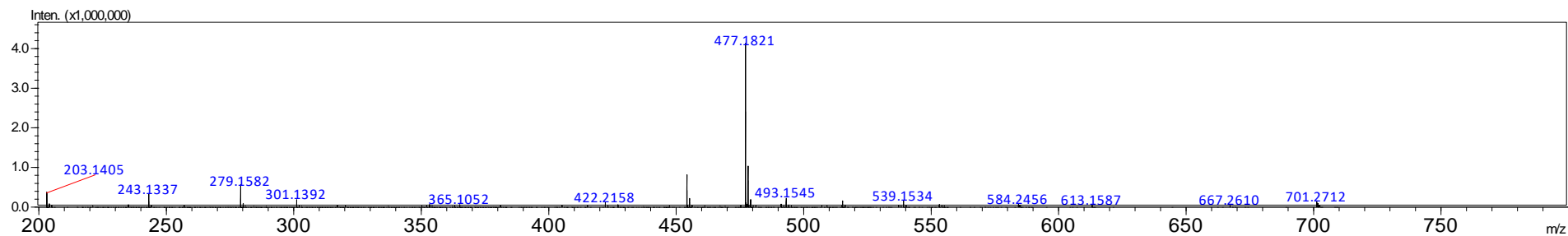


Figure S104. HRESIMS for 7b ((S)-MTPA ester)



	meas	calc	Δ (ppm)
[M+Na] ⁺	477,1844	477,1859	-3,14

Figure S105. HRESIMS for 7a ((R)-MTPA ester)



	meas	calc	Δ (ppm)
[M+Na] ⁺	477,1821	477,1859	-7,96

Figure S106. ^1H NMR spectrum of 9b ((S)-MTPA ester) measured at 700 MHz in CDCl_3

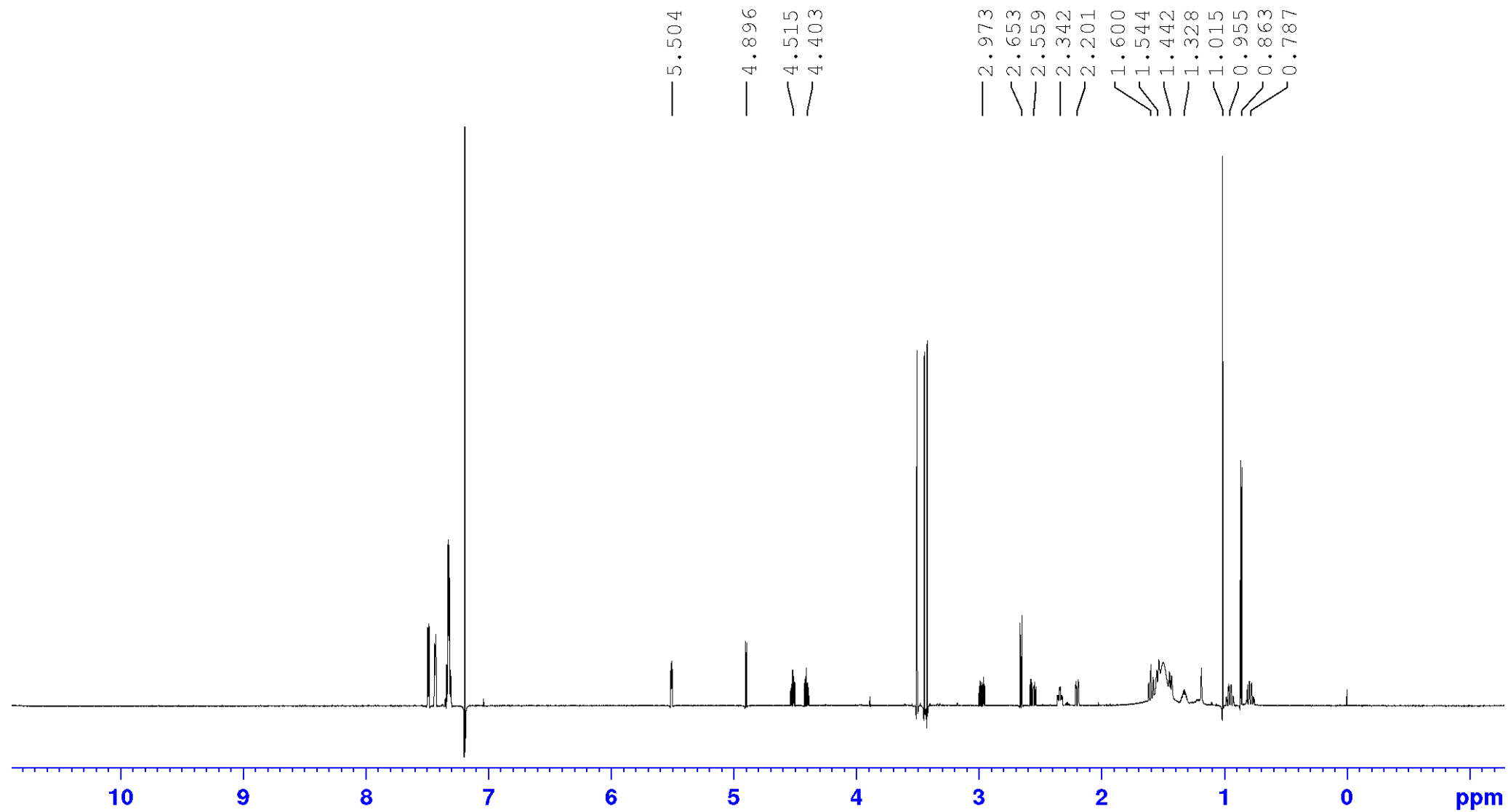


Figure S107. ^1H NMR spectrum of 9a ((R)-MTPA ester) measured at 700 MHz in CDCl_3

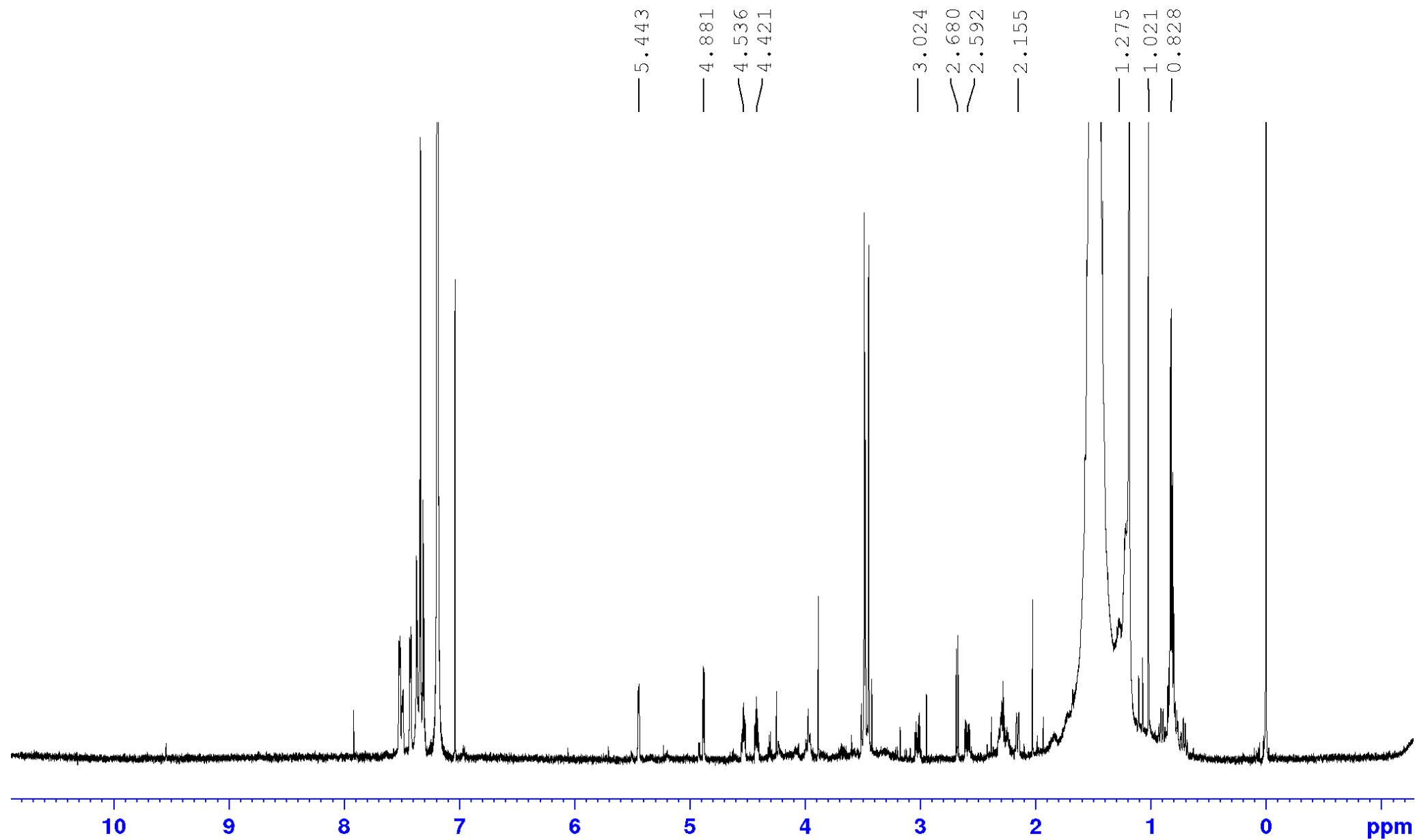


Figure S108. ^1H NMR spectrum of 10b ((S)-MTPA ester) measured at 700 MHz in CDCl_3

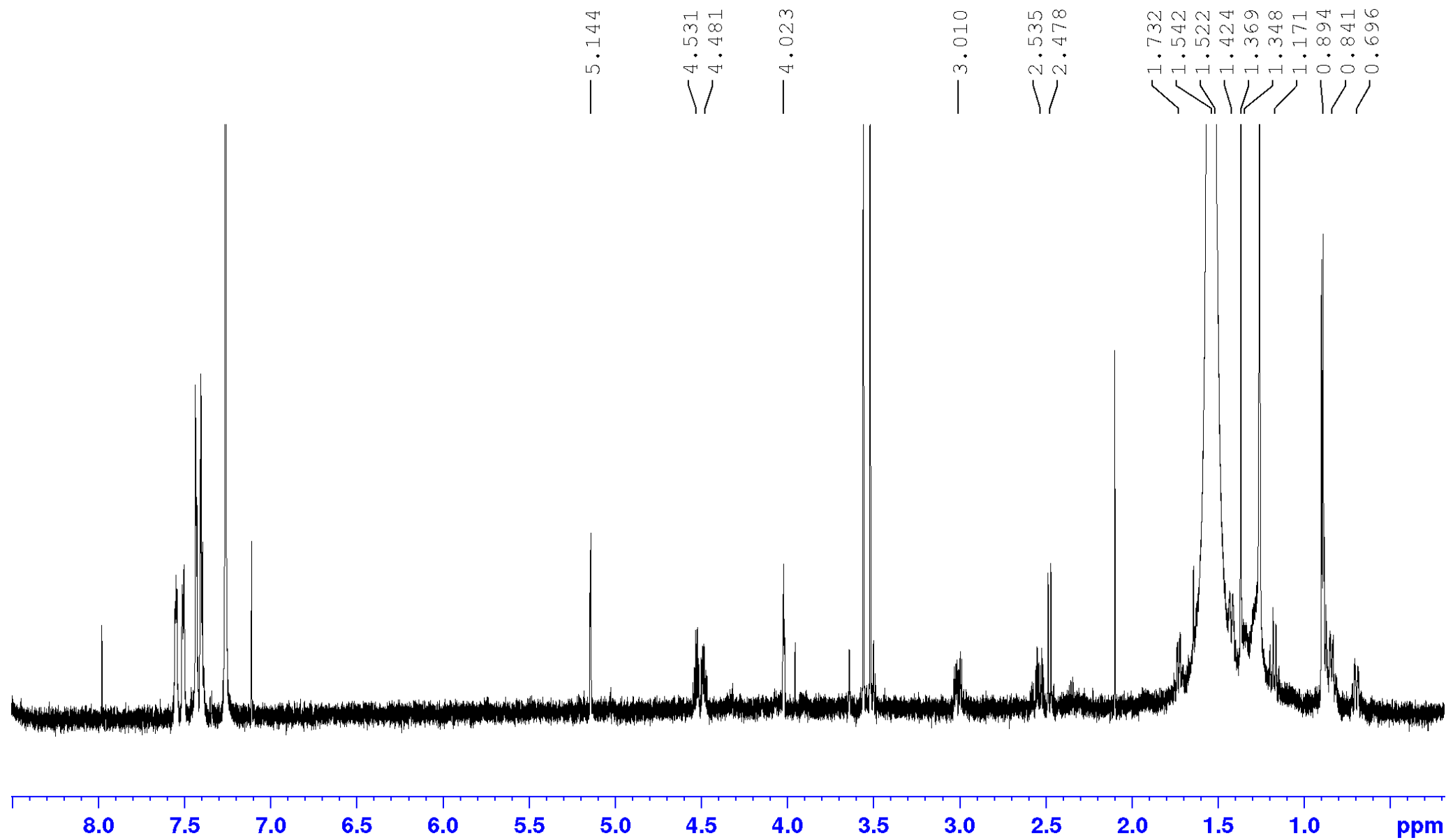


Figure S109. HSQC spectrum of 10b ((S)-MTPA ester) measured at 700 MHz in CDCl₃

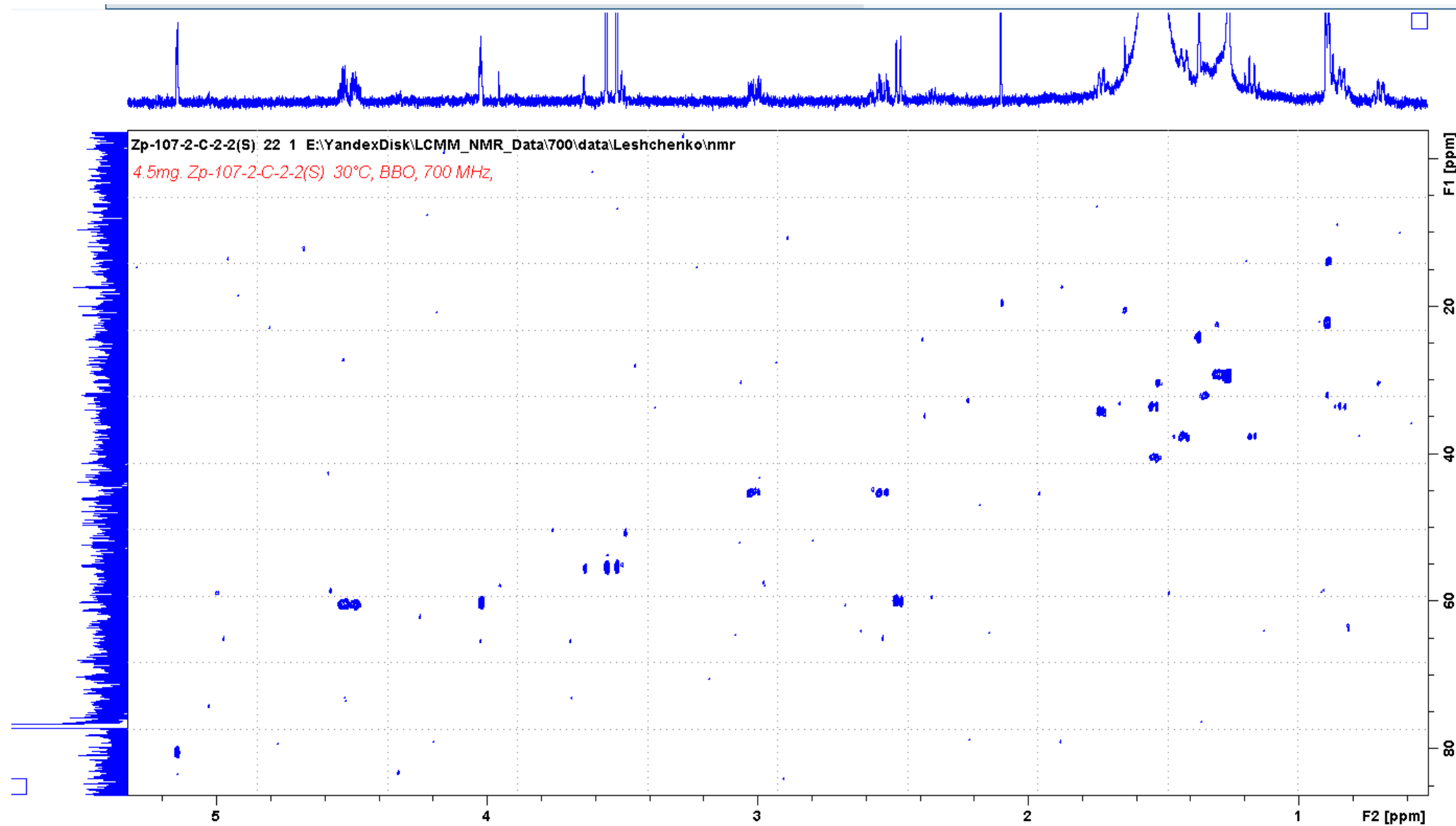


Figure S110. ^1H NMR spectrum of 10a ((R)-MTPA ester) measured at 700 MHz in CDCl_3

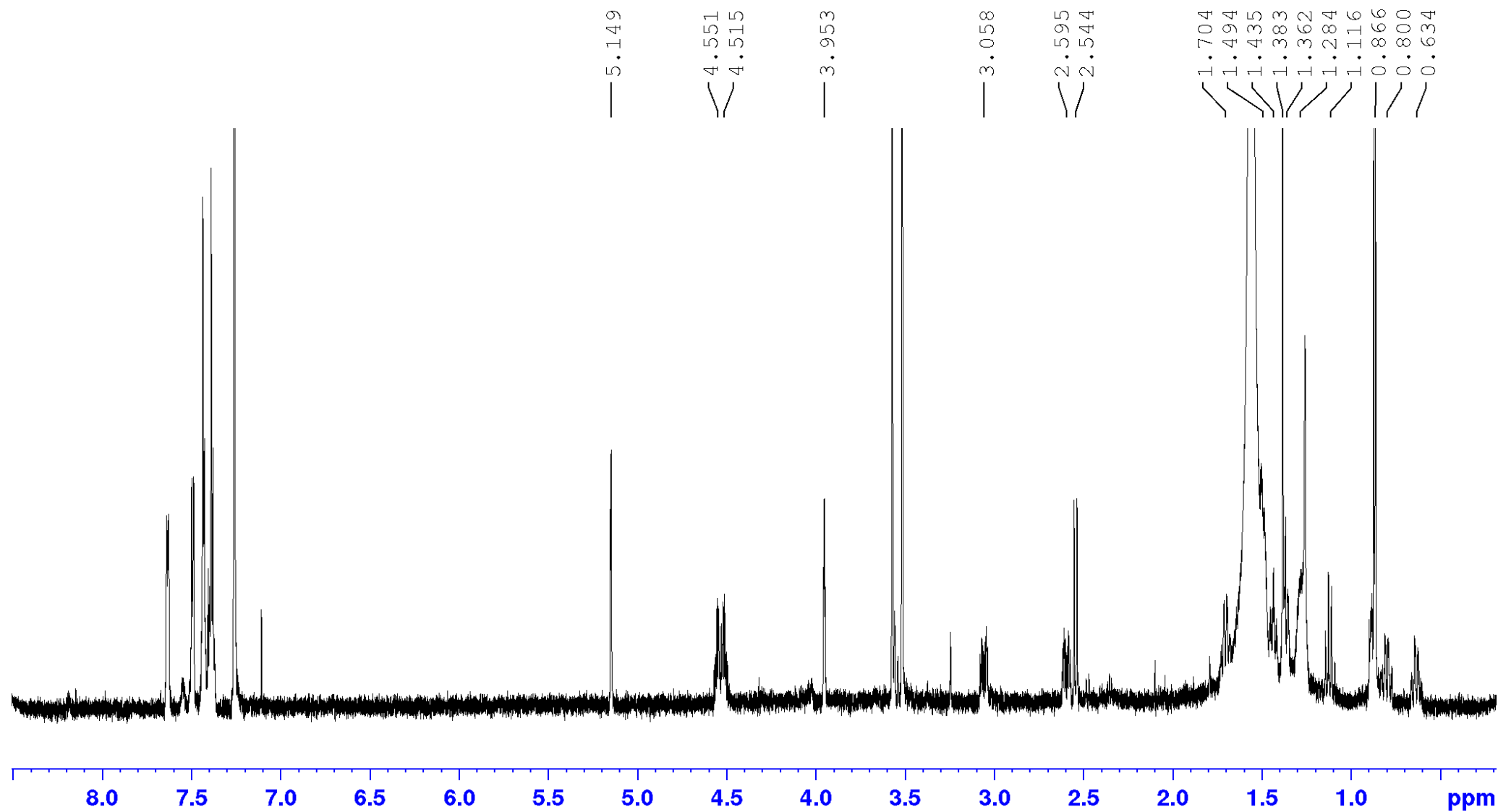


Figure S111. HSQC spectrum of 10a ((R)-MTPA ester) measured at 700 MHz in CDCl₃

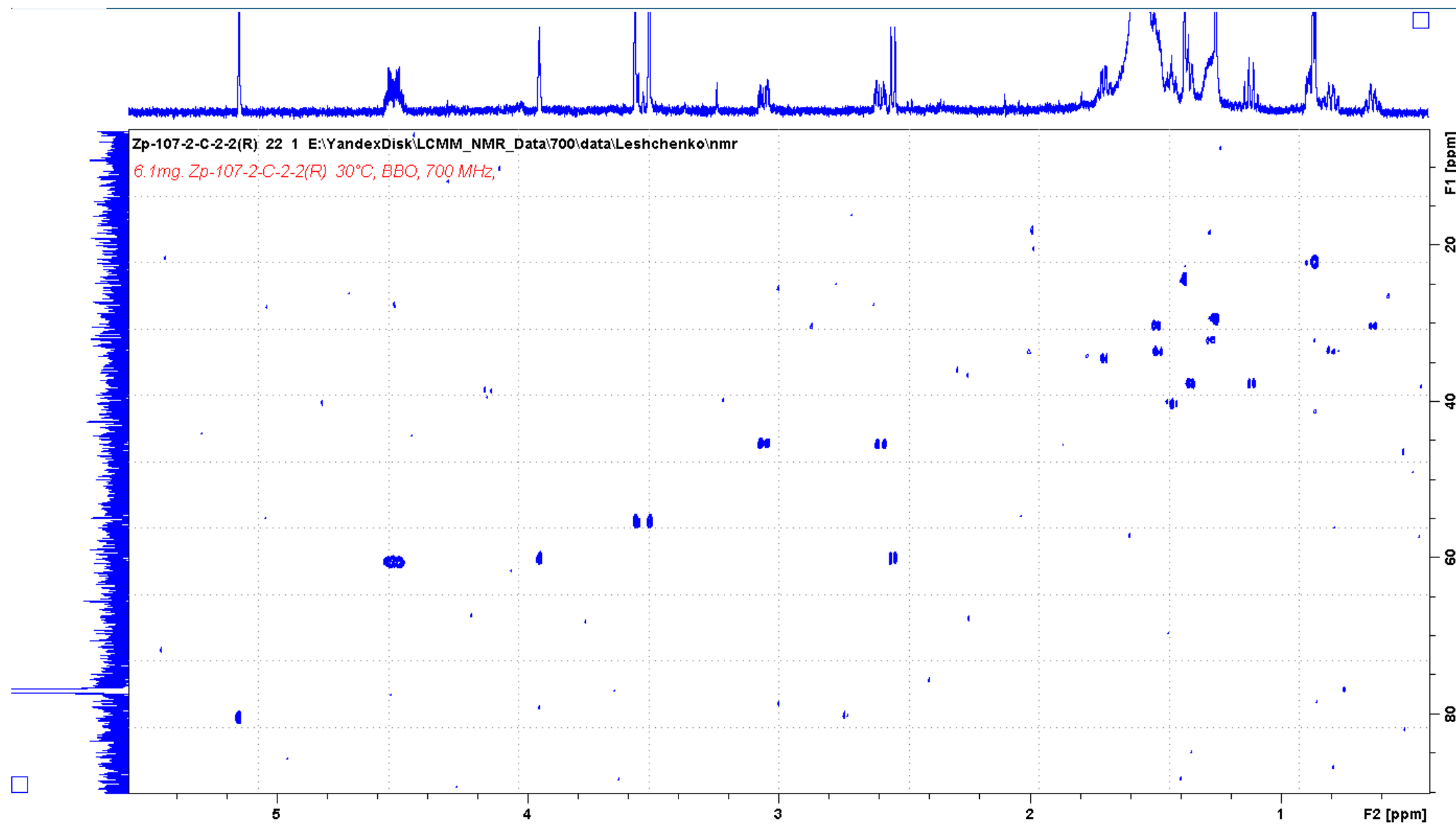
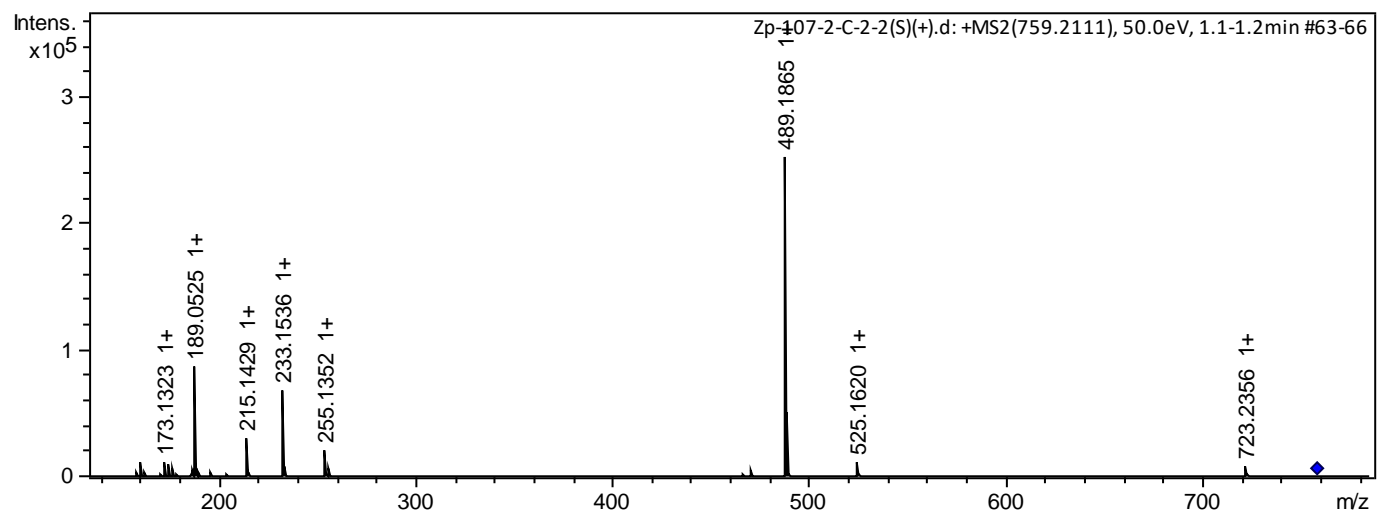
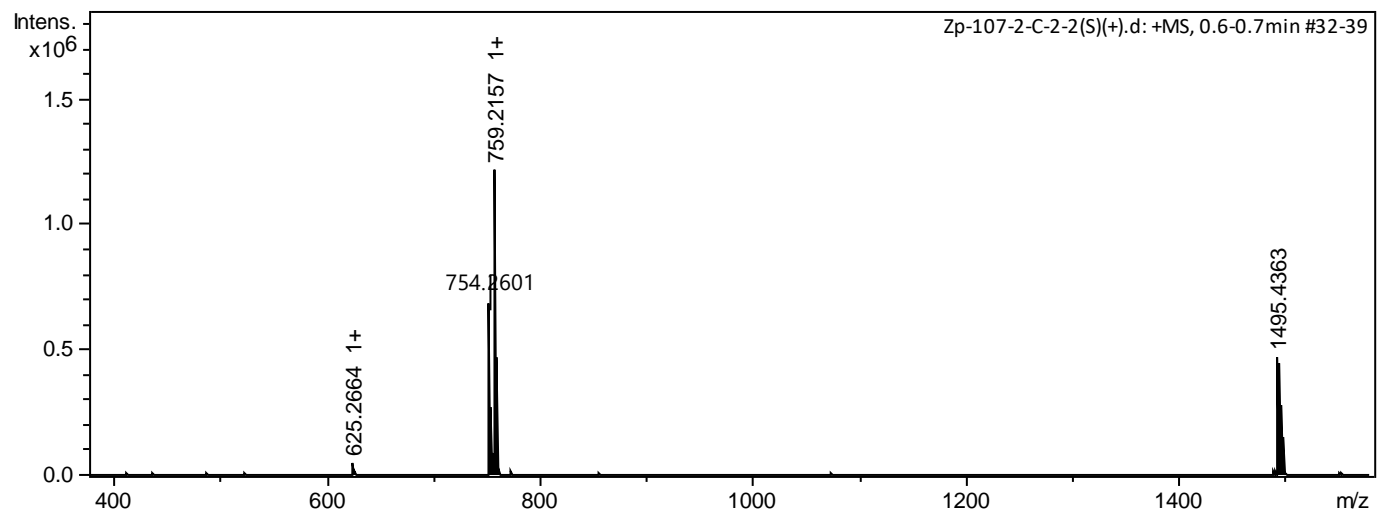
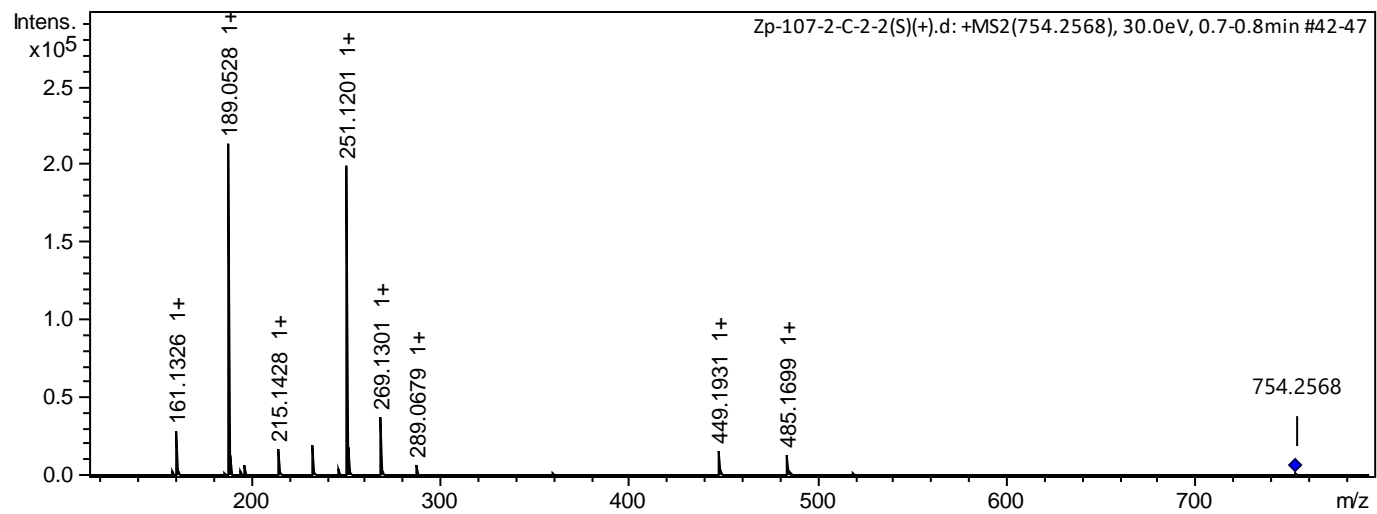


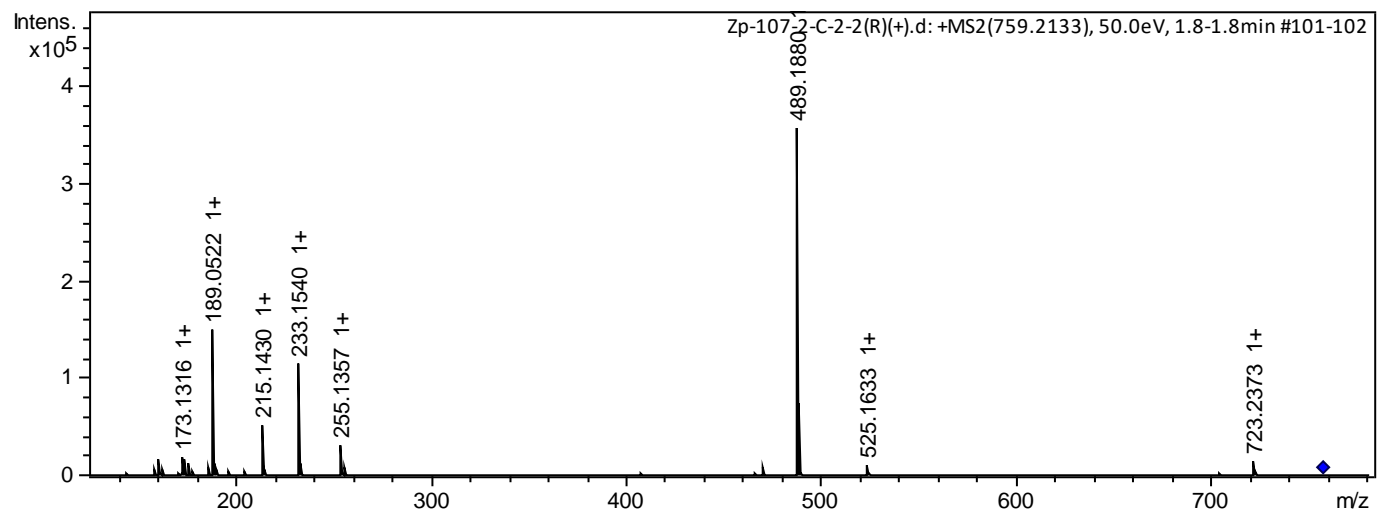
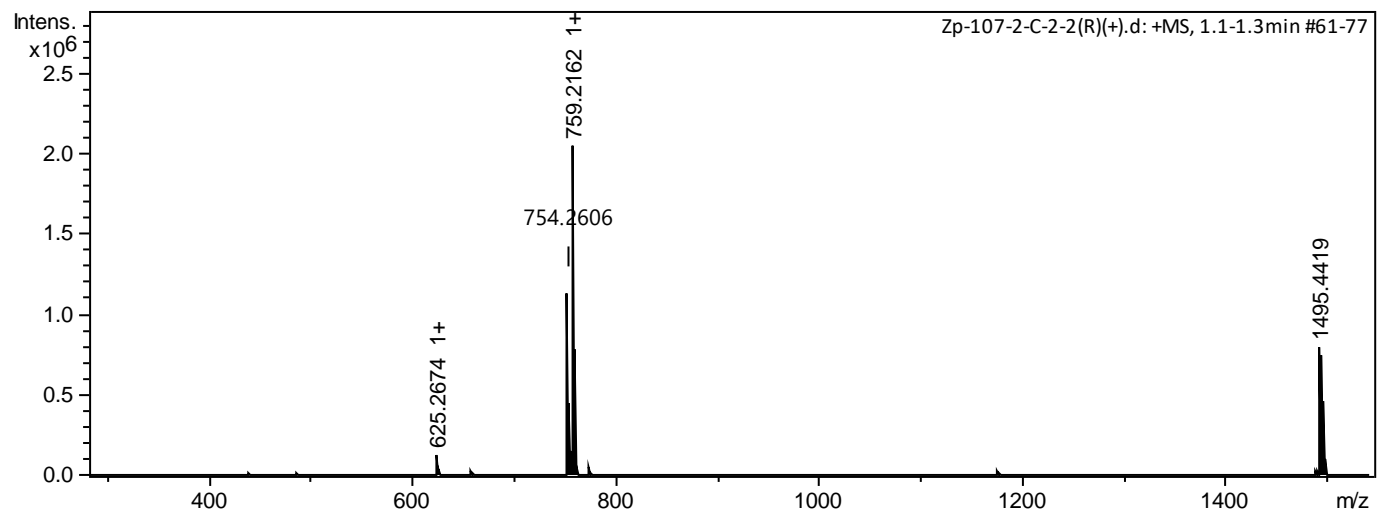
Figure S112. HRESIMS for 10b ((S)-MTPA ester)

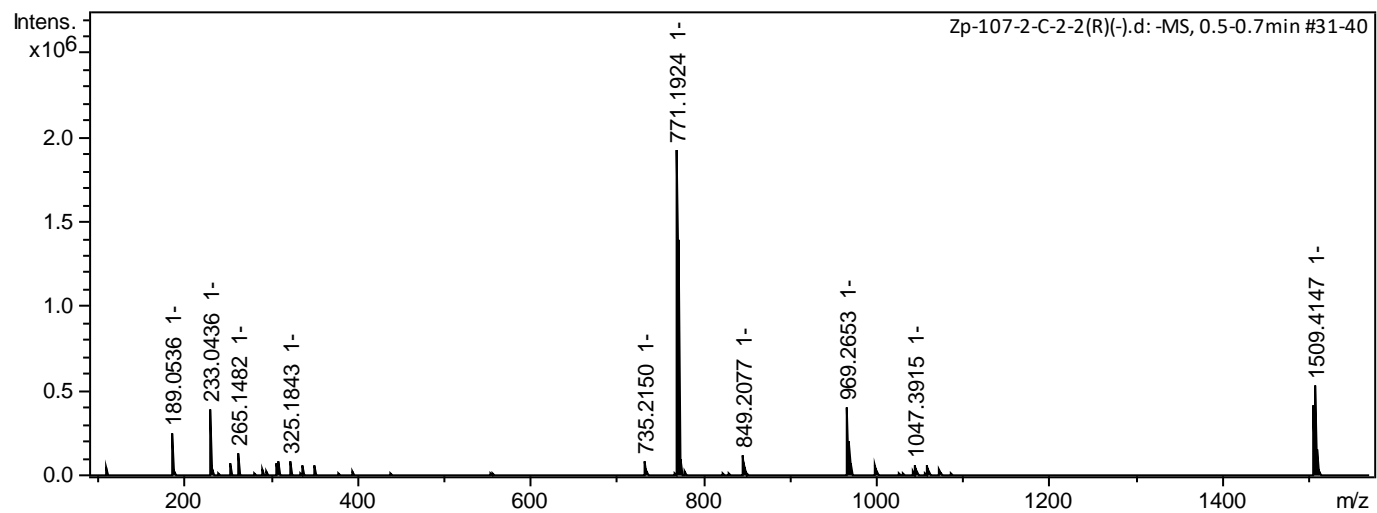
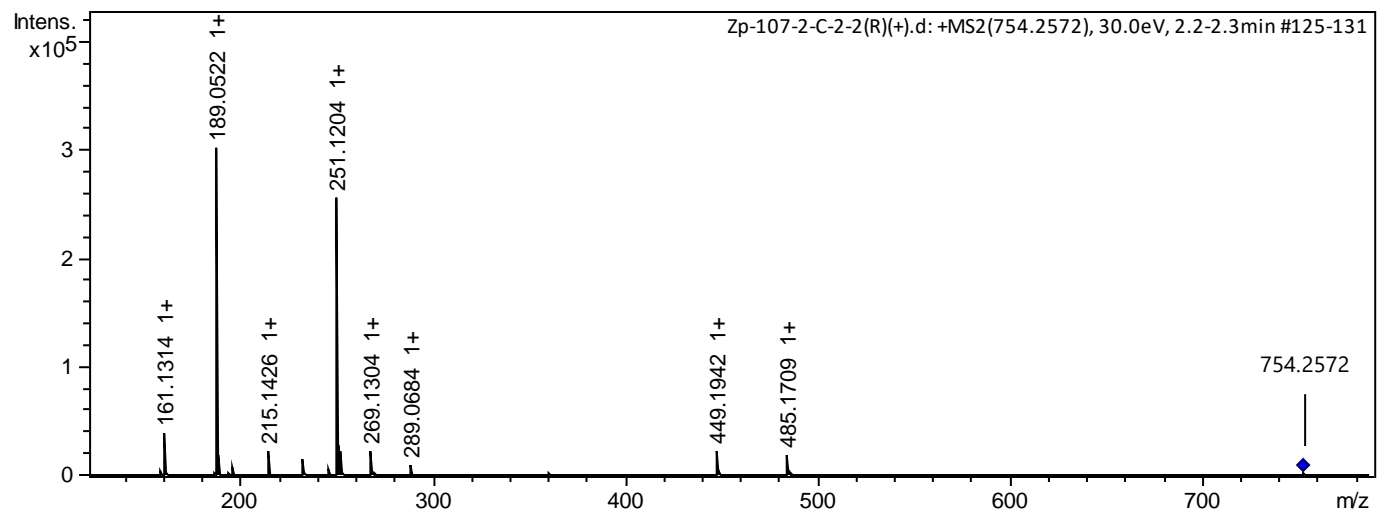




	meas	calc	Δ (ppm)
[M+Cl] ⁻	771,1933	771,1932	-0,1
[M+NH ₄] ⁺	754,2601	754,2552	-6,5
[M+Na] ⁺	759,2157	759,213	-3,6

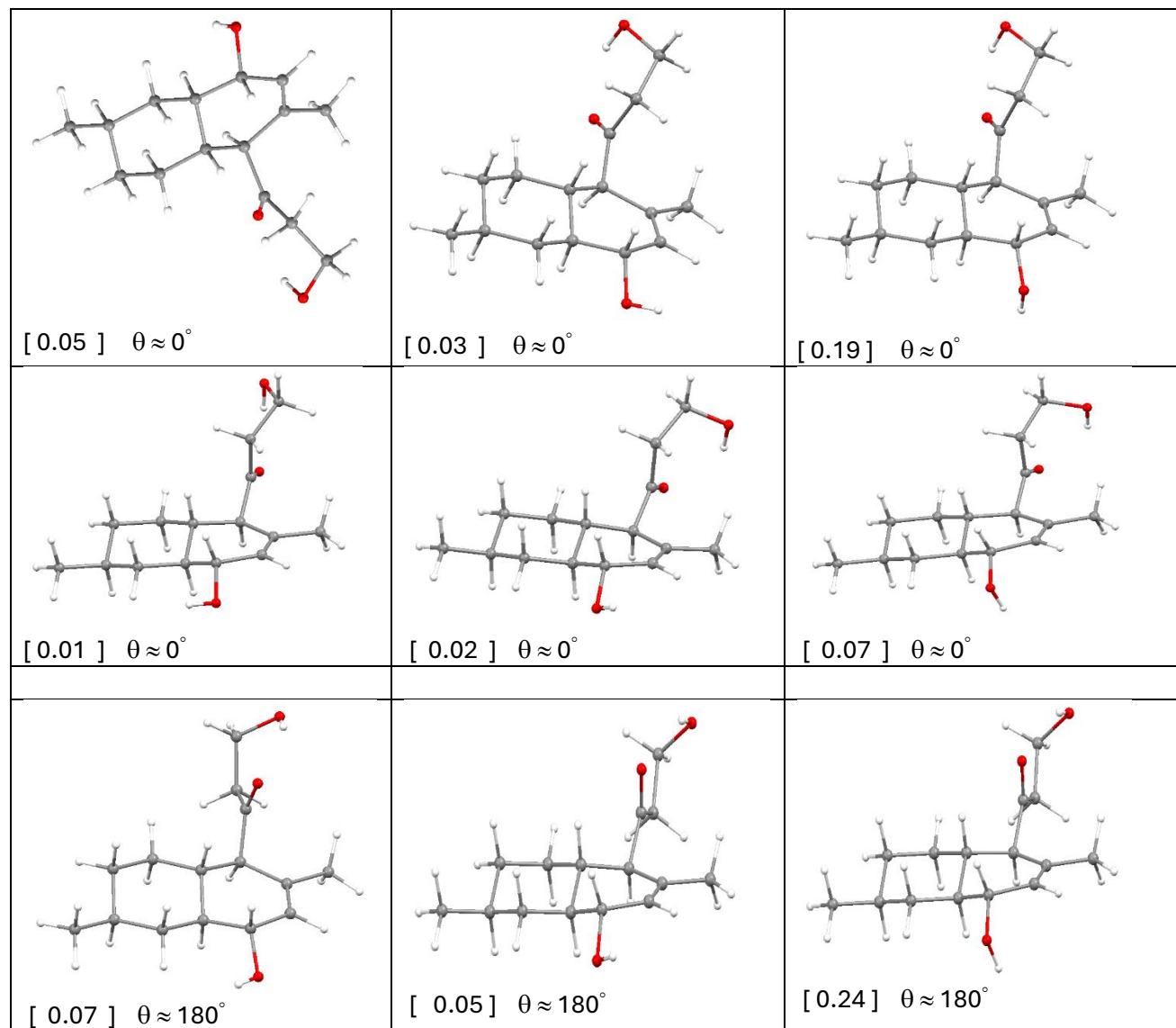
Figure S113. HRESIMS for 10a ((R)-MTPA ester)

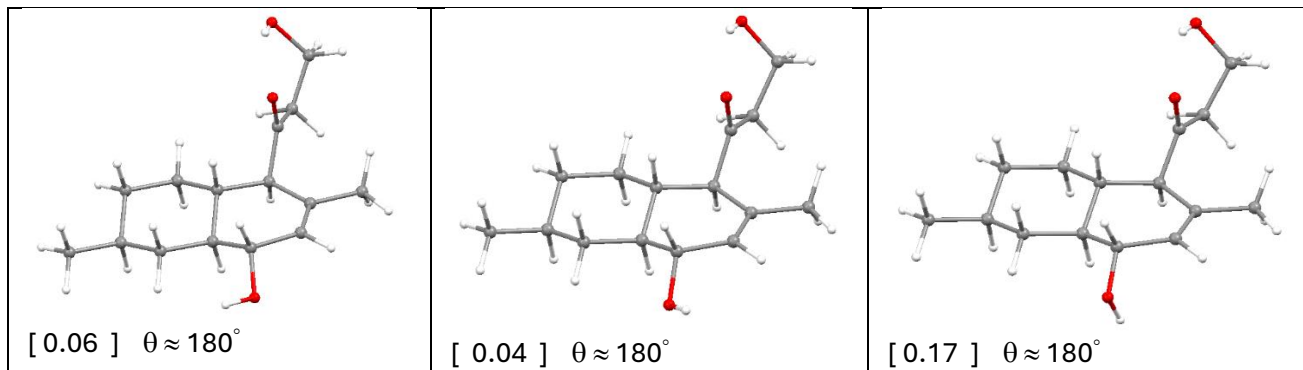




	meas	calc	Δ (ppm)
[M+Cl] ⁻	771,1924	771,1932	1,0
[M+NH ₄] ⁺	754,2606	754,2552	-7,2
[M+Na] ⁺	759,2162	759,213	-4,2

Figure S114. The main conformations of **1**





*the statistical weights are in square brackets

Figure S115. ECD spectra, calculated for different conformations of 1.

The ratio of intensities of bands at $\lambda \approx 210$ nm and $\lambda \approx 290$ nm in the total ECD spectrum is determined by the ratio, in which conformations $\theta \approx 0^\circ$ and $\theta \approx 180^\circ$ are presented in the sample. It is seen that theory based on PCM approach somewhat overestimates the percentage of $\theta \approx 180^\circ$ conformations. The same situation was obtained for compound **3** (see Figure S116)

We used the notation $\theta = \angle \text{O}-\text{C}(3)-\text{C}(4)-\text{H}(4)$.

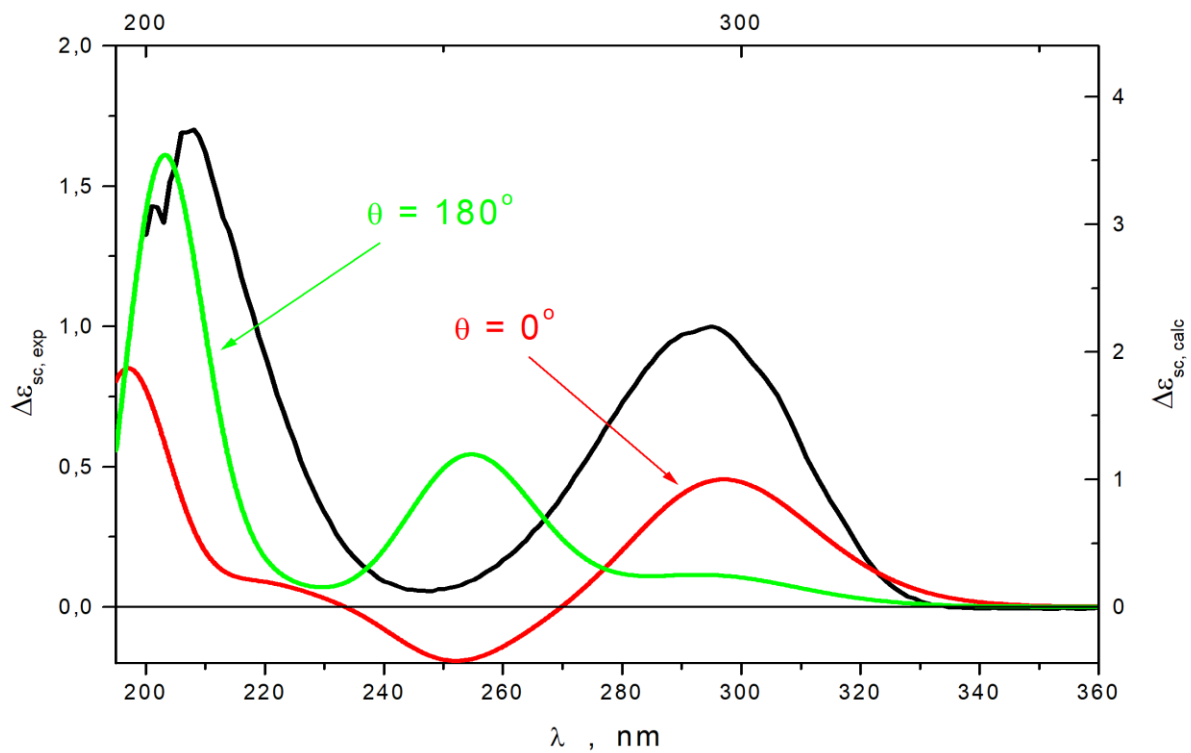


Figure S116. ECD spectra, calculated for different conformations of 3.

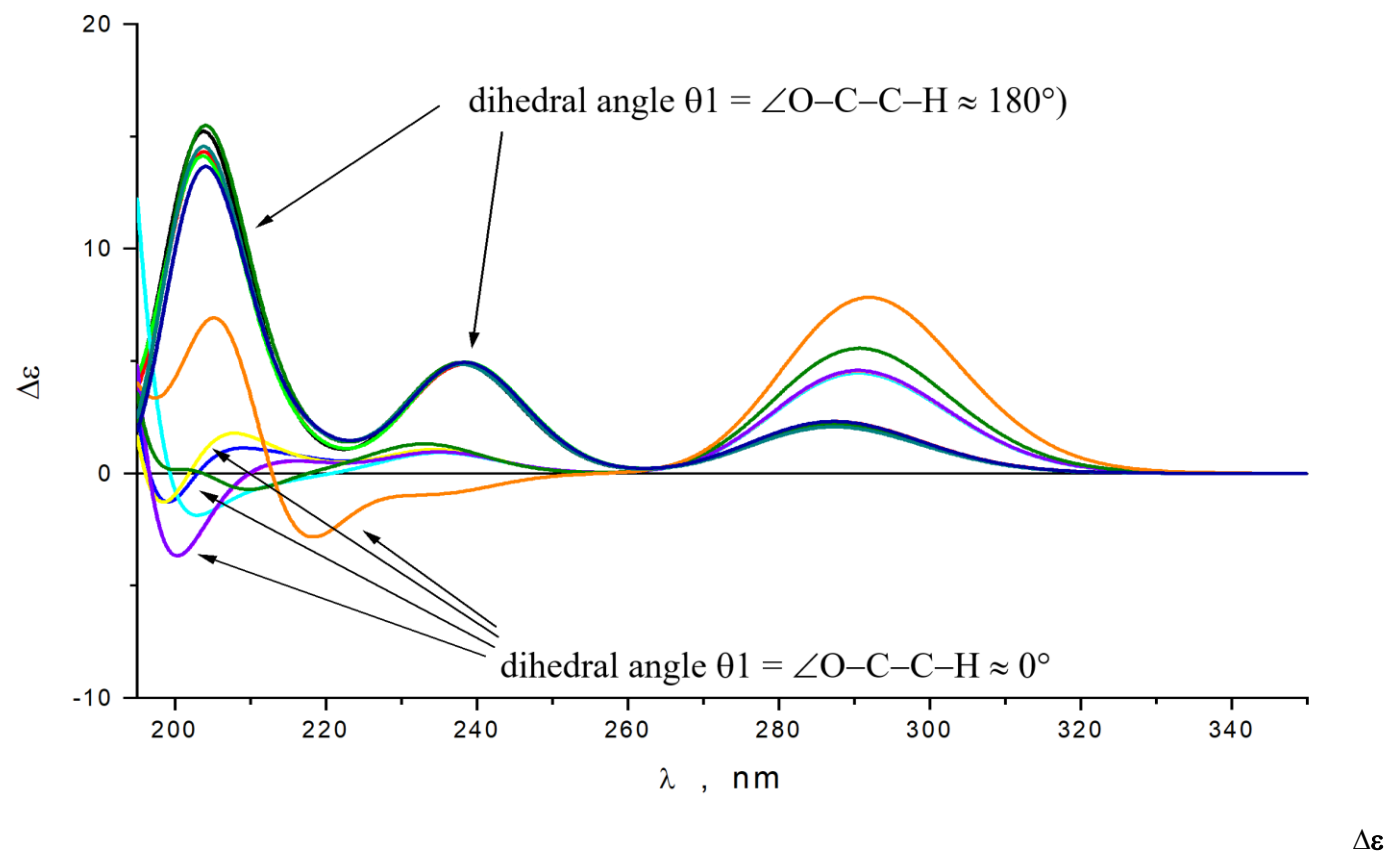


Figure S117. ECD spectra, simulated for different relative amounts of conformations with dihedral angle $\theta_1 = \angle\text{O}-\text{C}-\text{C}-\text{H} \approx 180^\circ$ and $\theta_1 = \angle\text{O}-\text{C}-\text{C}-\text{H} \approx 0^\circ$

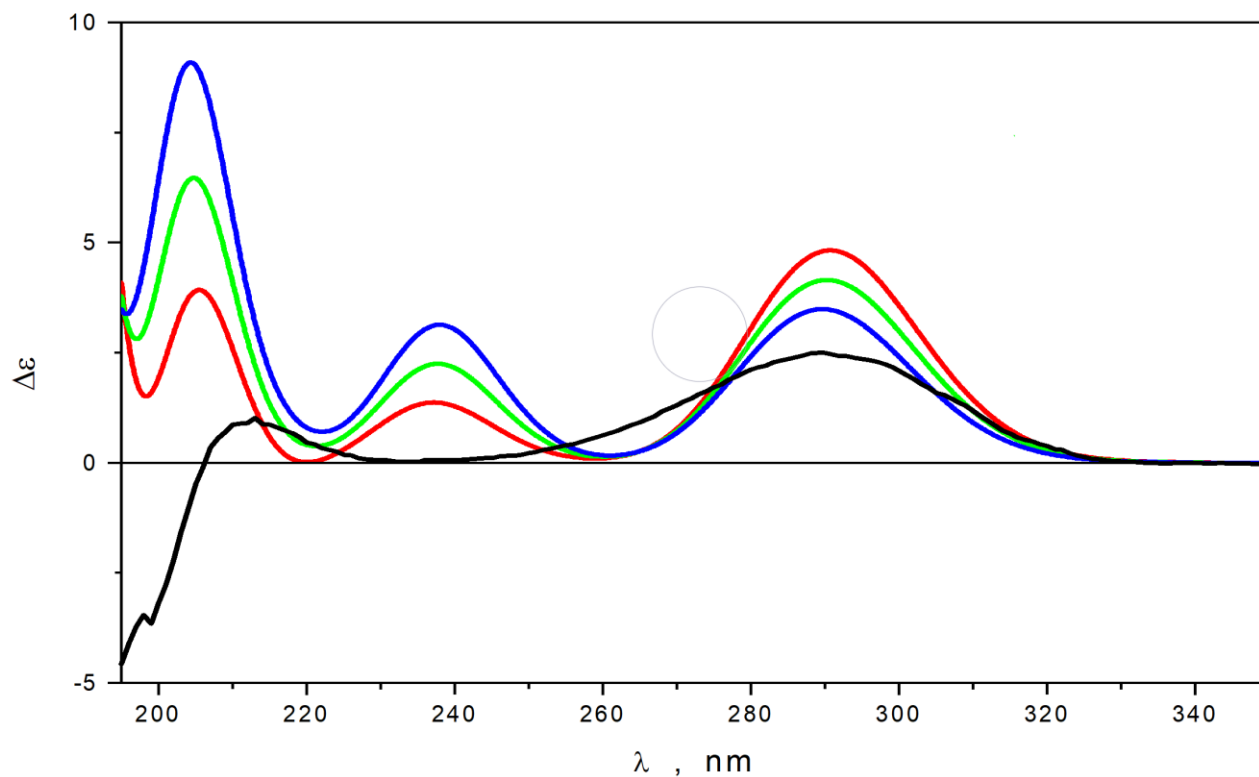


Figure S118. HPLC-MS/MS Analysis

The following mobile phases were used for the high pressure gradient: A, methanol+0.1% formic acid; B, water+0.1% formic acid. Eluent was pumped at a constant flow 0.2 mL/min with gradient (A:B, % by vol.): 80:20. Analysis time was 5 min. Injection volume was 5 μL .

The MS parameters were as follows: temperature of the heat block, and desolvation line was 200, and 250 $^\circ\text{C}$, respectively. Flow rate of nebulizing and dry gases (N_2) was 1.5 and 10 L/min, respectively. The capillary voltages were 4.5/-3.5 kV (for positive / negative ionization mode). The MS1 scan range was set at m/z 200-800. The MS2 scan range was set at m/z 100-800, CID energy was 50%

Figure S119. Key HMBC (blue arrows) and ¹H-¹H COSY (bold lines) correlations of 1–10.

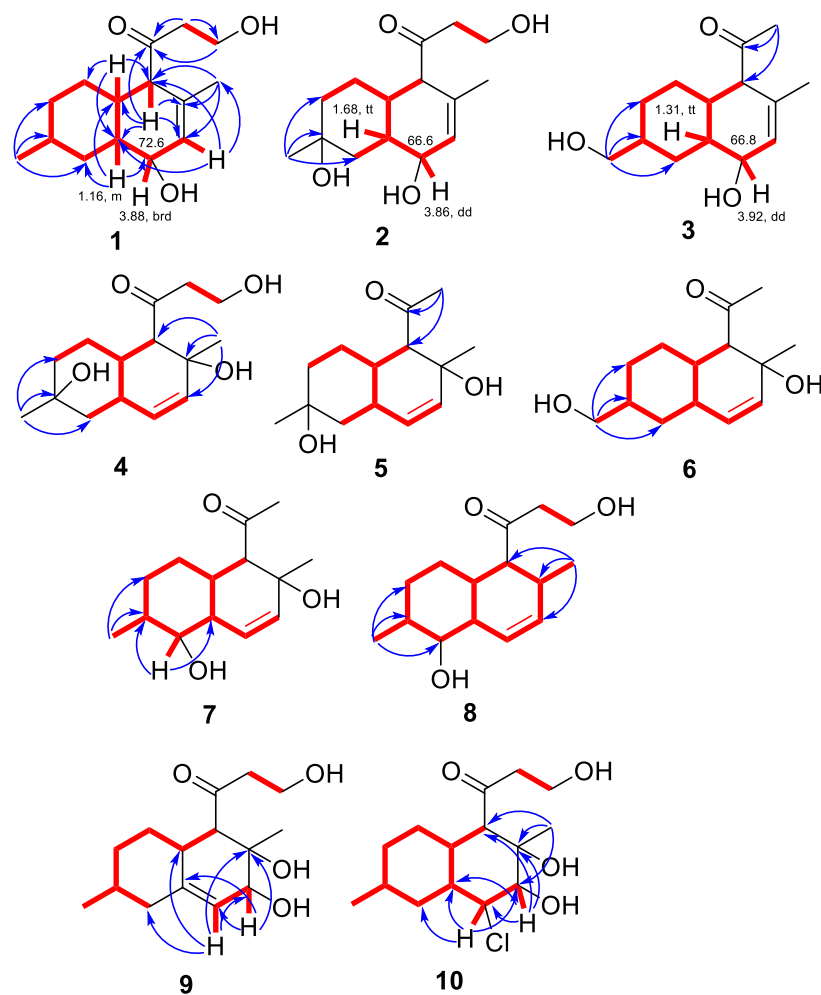
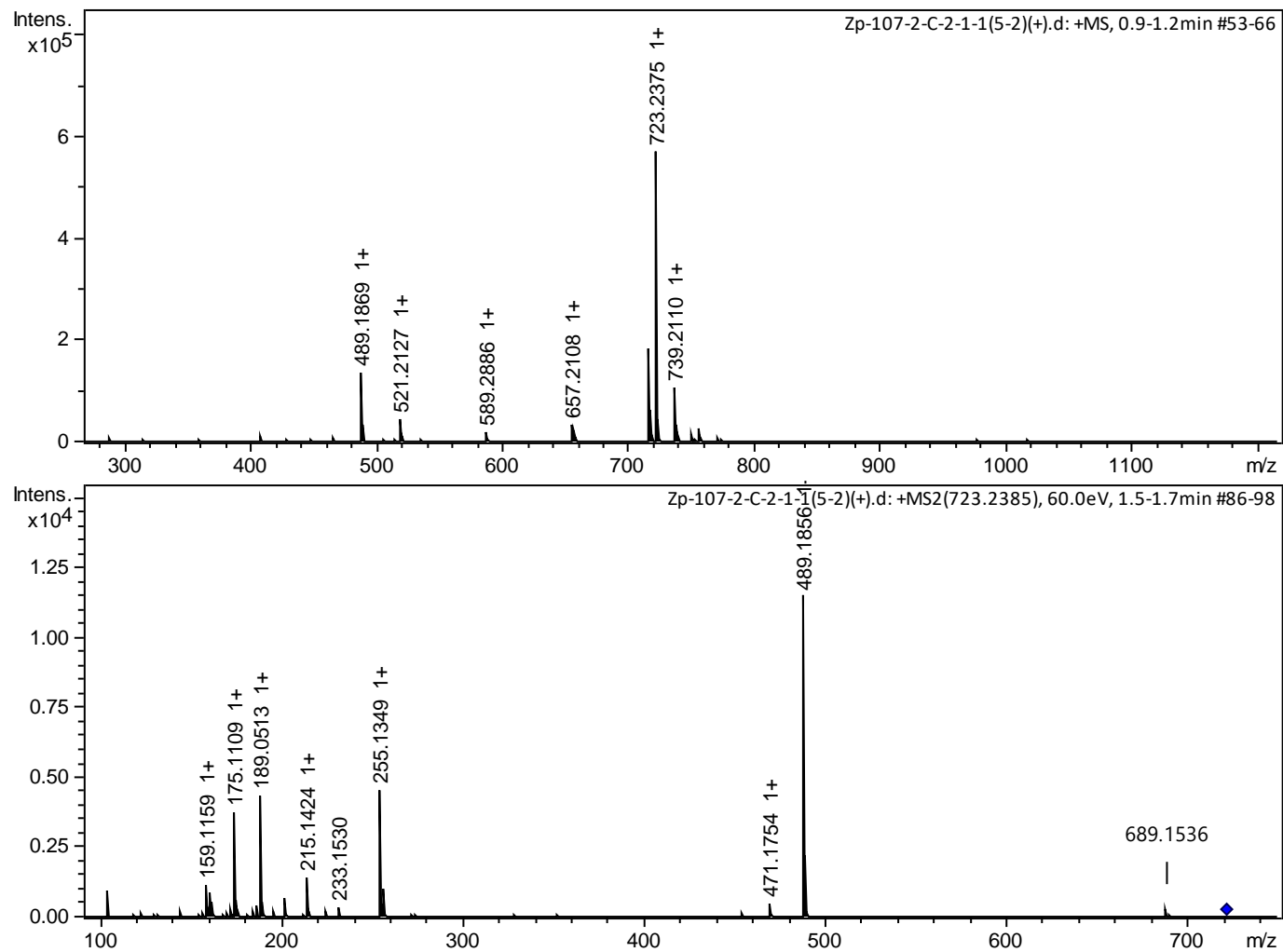


Figure S120. HRESIMS for 9b ((S)-MTPA ester)



	meas	calc	Δ (ppm)
[M+Cl] ⁻	735,2157	735,2165	1,1
[M+Na] ⁺	723,2375	723,2363	-1,7

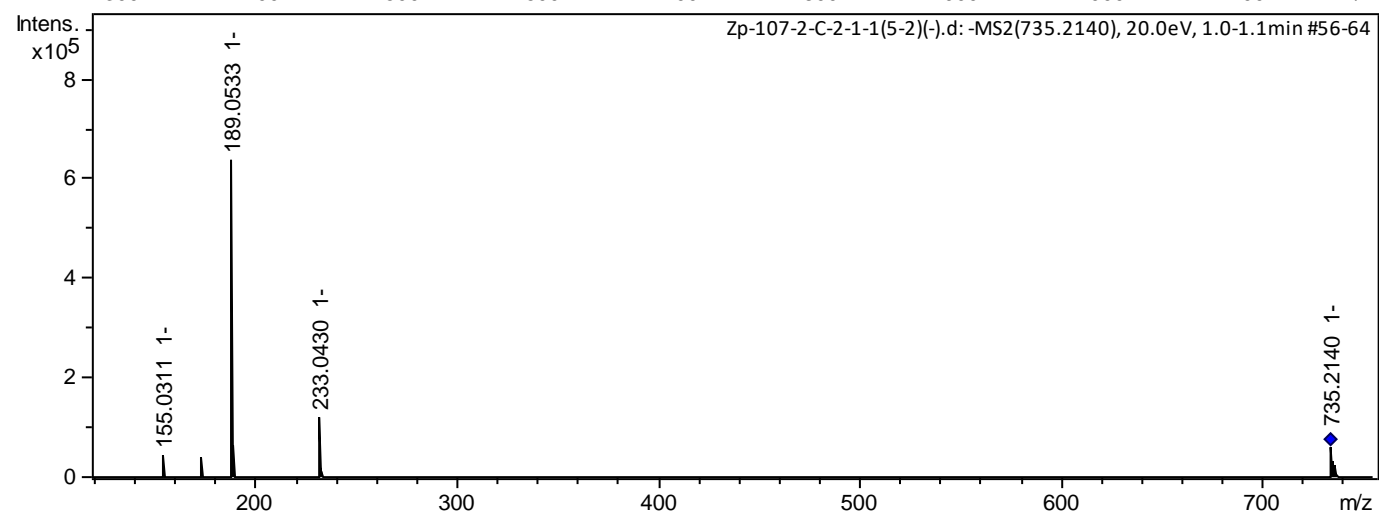
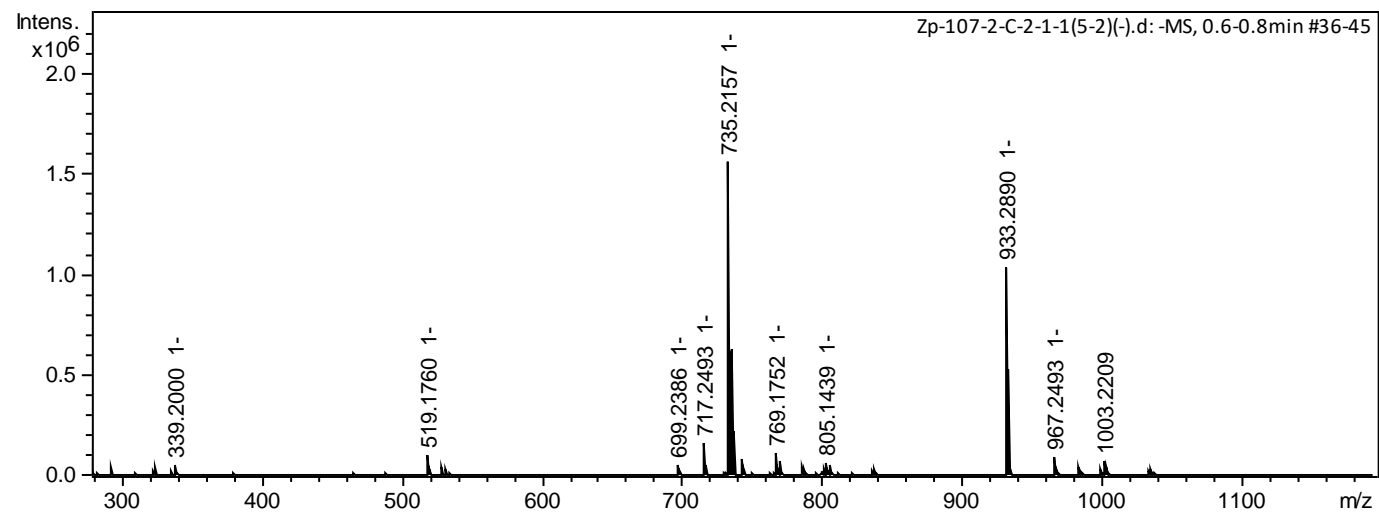
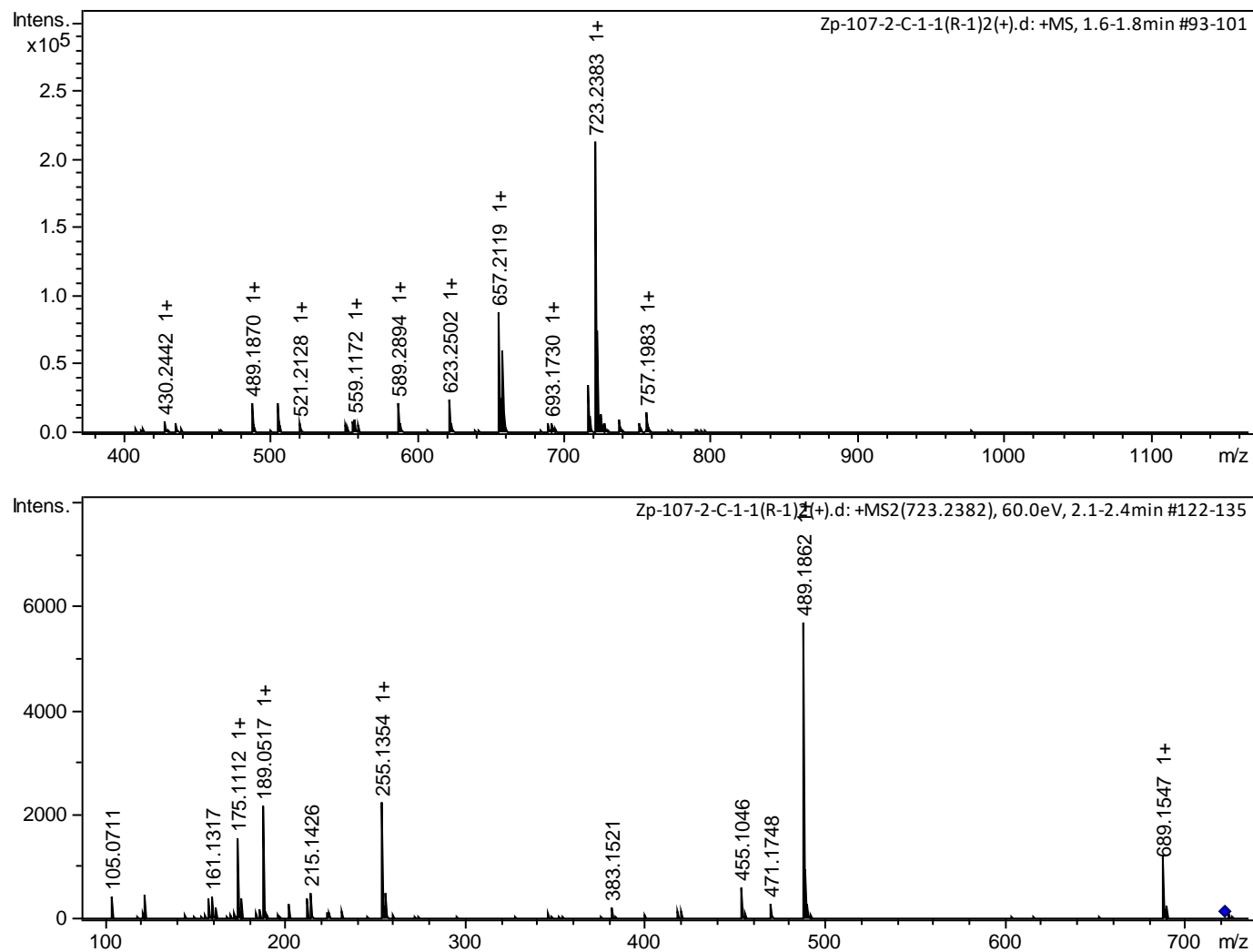
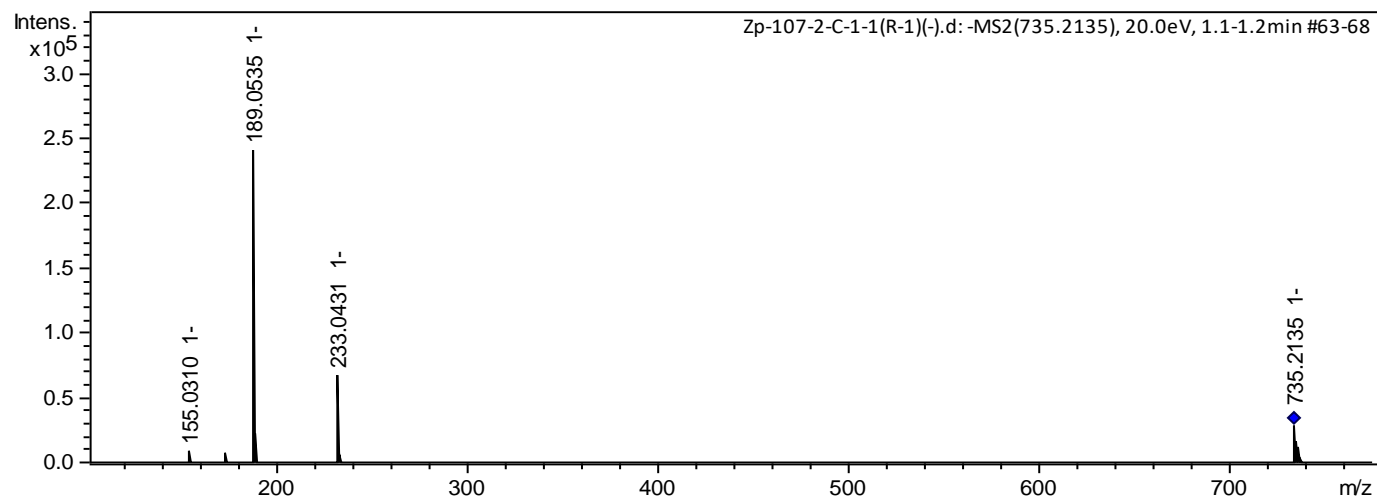
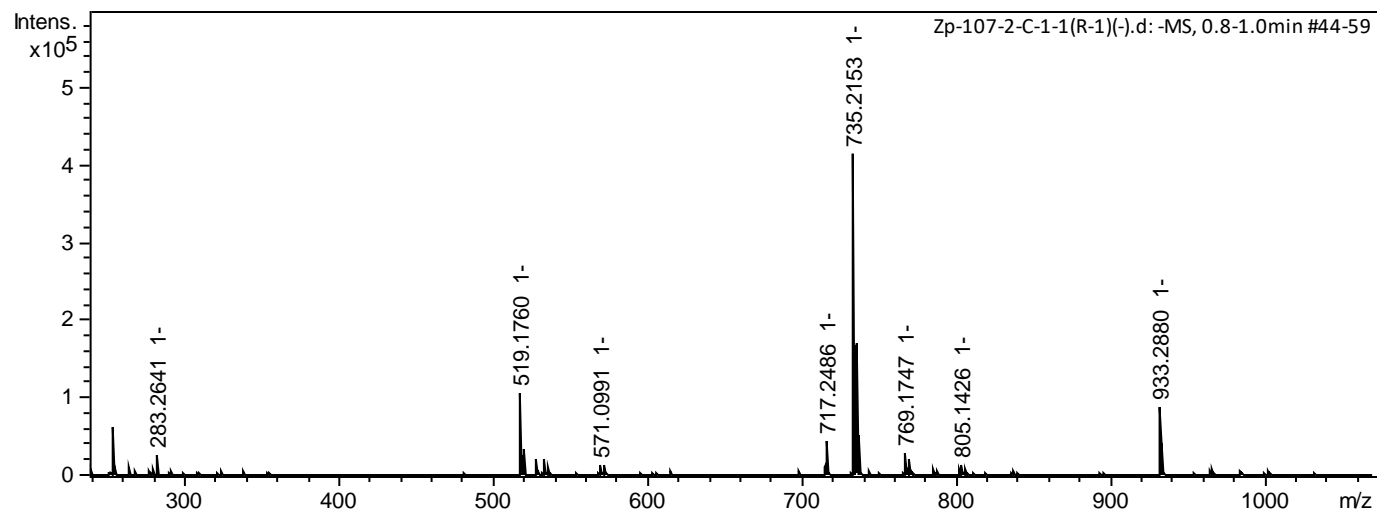


Figure S121. HRESIMS for 9a ((R)-MTPA ester)





	meas	calc	Δ (ppm)
[M+Cl] ⁻	735,2153	735,2165	1,6
[M+Na] ⁺	723,2383	723,2363	-2,8