

Article **Improvement of Thermal Behavior of Rattan by Lignosulphonate Impregnation Treatment**

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Abstract: Lignin derived from black liquor has a lot of potentials, particularly in its thermal stability, for making value-added chemicals. The purpose of this study was to determine the effect of washing frequency during hydrochloric acid lignin isolation on the properties of eucalyptus kraft lignin. To improve its thermal characteristics and enable its usage as an additive flame retardant, the isolated lignin was synthesized into lignosulphonate. The lignin produced by $3\times$ and $5\times$ washing treatments had a purity of 85.88 and 92.85%, respectively. An FTIR analysis indicated that lignosulphonate was successfully synthesized from isolated lignin after $3\times$ and $5\times$ washing treatments, as the S=O bond was detected at around 627 cm $^{-1}$. The lignosulphonate exhibited a purity of 71.89 and 67.21%, respectively. Thermal gravimetry and differential scanning calorimetry analysis revealed that the lignin and lignosulphonate after $3\times$ and $5\times$ washing treatments had a char residue of 44, 42, 32, and 48%, respectively. Glass transition temperatures (Tg) of 141, 147, 129, and 174 °C were observed. According to the findings, washing frequency increases lignin purity and Tg, thereby improving the thermal properties of lignosulphonate. Furthermore, the flammability of rattan impregnated with lignosulphonate was V-0 in the UL-94 vertical burning test.

Keywords: chemical-thermal properties; Eucalyptus lignin; lignosulphonate; flammability performance; washing treatment

1. Introduction

The pulp and paper industries in Indonesia play an important role in the nation's economy. These industries contributed 0.67% to Indonesia's Gross Domestic Product (GDP) in 2021 and brought in 7.5 billion US dollars in foreign exchange for the country. The pulp and paper industry in Indonesia ranks first in ASEAN and third in Asia for pulp production, fourth in Asia for paper production, and eighth and sixth in the world, respectively, for pulp and paper production [\[1\]](#page-19-0). In 2018, it was reported that the production of pulp and paper in Indonesia reached 11 and 16 million tons, respectively [\[2\]](#page-19-1). High pulp and paper production also generates a lot of waste, especially black liquor, in which the ratio of black liquor to the pulp is 7:1. Black liquor is composed of 15% solid residue, the majority of which is organic $(2/3)$ and inorganic $(1/3)$ chemicals [\[3\]](#page-19-2). Utilizing byproducts from the pulp and paper industry can reduce waste, improve resource efficiency, and improve environmental health [\[4\]](#page-19-3).

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Black liquor is primarily an organic substance with a high lignin concentration that has enormous potential as a raw material for a variety of valuable applications [\[5\]](#page-19-4). However, only 2%–5% of lignin has been used for high-value-added biomaterials, with the remaining lignin being used to heat boilers in the pulp industry [\[6,](#page-19-5)[7\]](#page-19-6). To maximize the economic benefits of lignin, several factors must be considered, including yield, heterogeneous lignin structure, purification, and reactivity [\[8\]](#page-19-7). When lignin is combined with other polymers, the size of the lignin as well as the method of lignin preparation have an effect on the final product [\[7,](#page-19-6)[9\]](#page-19-8). Lignin can be optimized by converting it into bioproducts [\[8\]](#page-19-7), one of which is its usage as a flame-retardant additive due to its excellent thermal properties. Lignin modification is a viable option to enhance the lignin reactivity before conversion into bioproducts [\[5\]](#page-19-4). The thermal properties of lignin are highly variable and can be influenced by a number of factors, including the source of the lignin, the extraction method, the low molecular weight of some species, moisture content, and chemical modifications [\[10](#page-19-9)[–12\]](#page-20-0).

The ability of lignin to produce char when heated and degraded is one of the most crucial aspects that determine its flame retardancy behavior. Char can slow down the rate at which polymeric materials burn, making it more difficult for oxygen to enter the combustion zone [\[13\]](#page-20-1). A flame retardant is a substance that, when applied to another substance, reduces the combustibility of the treated substance when exposed to heat or flame [\[14\]](#page-20-2).

Fire-retardant chemicals such as halogens, nitrogen, phosphorous, boric acid, inorganic metal compounds, and borax are currently used in consumer goods. Despite the fact that this practice is suspected of producing combustion byproducts or other undesirable side effects, these fire-retardant chemicals are still used [\[15](#page-20-3)[–17\]](#page-20-4). The use of halogen-free natural materials that also serve as fire retardants can benefit a variety of products, including electronics, furniture, and other household items. This provides a safer alternative [\[18\]](#page-20-5). Lignin has been employed as a refractory additive for polylactic acid in the investigation of Costes et al. [\[19\]](#page-20-6), polyamide [\[20\]](#page-20-7), and wood–plastic composites [\[21\]](#page-20-8). Lignin has been modified in various ways, such as by sulfonation into water-soluble lignosulphonate, in order to improve the flammability behavior of lignin. This has been accomplished by modifying the lignin. It is proposed that these properties will make the preparation easier when mixing with hydrophilic polymers such as woody and non-woody biomass. When compared to commercial flame retardants such as ammonium polyphosphate (APP), lignosulfonate, such as calcium lignosulfonate, is cheaper than APP and is more easily accessible and costeffective. Lignosulphonates have a significant molecular weight, aliphatic hydroxyl groups, and sulfonate groups. These are critical in determining their dispersing effectiveness and utility for various uses, including oil well dispersants, coal-water slurry dispersants, dye dispersion, polymer composites, adhesives, water-reducing agents, pesticides, ceramic colloidal processing, and carbon black [\[7,](#page-19-6)[22](#page-20-9)[–24\]](#page-20-10). However, up to now, investigation on the use of lignosulfonate as a flame retardant is very limited. Angelini et al. [\[25\]](#page-20-11) investigated the fire retardancy performance of particleboards containing lignosulphonate in the system. In the study, lignosulphonate was purified by dialysis against water as a carbohydrate-free dialyses lignin (LD) before being combined with urea-formaldehyde as a particleboard binder. This treatment causes the reduction of the peak of heat release rate by 15 wt.%, smoke production by 25%, and an increase in char formation of particleboard. For industrial purposes, flame retardants need to achieve a V-0 rating in UL-94 and a limiting oxygen index (LOI) of up to 28% [\[26\]](#page-20-12). Therefore, this is the first study to investigate the use of synthesized lignosulphonate from hardwood lignin for the impregnation of non-woody biomass. It is a novel approach to using lignosulphonate that could lead to a wide range of future applications.

This study aimed to investigate the lignin properties of the kraft black liquor of *Eucalyptus* sp., derived from pulp and paper industry by-products, for improving the flammability behavior of biobased materials such as rattan. The possibility of sulfonating Eucalyptus kraft lignin into lignosulphonate was also investigated, with the main focus on the effect of washing treatment in lignin isolation on the lignosulphonate properties. It is also critical to optimize lignosulphonate synthesis conditions, apply lignin and lignosulphonate-based

refractory additives to rattan, and assess the material's flammability. There are 600 rattan species in eight genera, approximately 300 of which are found in Indonesia. Indonesia produces 80%–90% of the world's rattan, making it the leader in rattan harvesting and household products, including further than $\frac{1}{2}$ exportation $[27]$. Rattan is a raw material that is used for a variety of different types of work and household products, including furniture. Rattan, on the other hand, has a number of disadvantages, including a low resistance to harmful organisms, such as powder post beetle attacks, and a high risk of ignition due to its organic nature [\[28\]](#page-20-14). It is anticipated that the increase in the quality of rattan as a raw material will lead to an increase in the functional value of items that are made from rattan.

2. Materials and Methods *2.1. Materials*

2.1. Materials The materials used in this study included kraft-black liquor from *Eucalyptus* sp with

The materials used in this study included kraft-black liquor from *Eucalyptus* sp. with a solid content of ±65% provided from PT. Toba Pulp Lestari in Sosor Ladang Village, Porsea District, Toba Samosir Regency, North Sumatra, Indonesia. HCl, NaOH, H₂SO₄, and $\text{Na}_2\text{S}_2\text{O}_5$ were used in lignosulphonate synthesis and lignin isolation. Lignin reference (catalog number: 370959) and lignosulfonate reference (catalog number: 471038) from Sigma Aldrich were used as a comparison to the characteristic of the isolated lignin and synthesized lignosulfonate. All analytical-grade chemicals in the process and analysis were used without purification.

2.2. Methods $\sum_{i=1}^{n}$

This research was carried out in several stages, with a flow chart presented in Figure [1.](#page-2-0)

Figure 1. Research flow chart. **Figure 1.** Research flow chart.

2.2.1. Lignin Isolation of Black Liquor 2.2.1. Lignin Isolation of Black Liquor

A total of 50 g of black liquor was put into a 500 mL beaker glass and diluted with A total of 50 g of black liquor was put into a 500 mL beaker glass and diluted with distilled water to 500 mL. The pH 2 was then attained by gradually adding the 2 M HCl distilled water to 500 mL. The pH 2 was then attained by gradually adding the 2 M HCl solution while stirring. The sample solution was kept at 25° C for 24 h to allow it to settle totally. After that, the solution was decanted employing a pickpocketed top (solution), then washed with distilled water as much as 350 mL per washing. The decantation stage was carried out for 3 and 5 washes. The precipitate and a small amount of solution were filtered, and the formed precipitate was put in pre-coated trays with heat-resistant plastic. The sample was oven dried at a temperature of 45 °C for 24 h. The dried lignin was crushed. The properties of the crushed dried lignin were then investigated, such as the yield of isolated lignin, calculated with Equation (1). The isolated lignin was also analyzed for purity, moisture content, ash content, phenolic hydroxyl groups, solubility in dioxane and NaOH, SEM-EDX, functional groups, lignin monomer, and thermal properties. \mathcal{L} is not thermal properties.

was carried out for 3 was held 5 was held 5 was held 3 was held and a small amount of solution were solution w

The yield of lignin (
$$
\%
$$
) = $\frac{\text{the dry weight of isolated lignin}}{\text{the dry weight of black liquor}} \times 100\%$ (1)

2.2.2. Lignosulphonate Synthesis 2.2.2. Lignosulphonate Synthesis

In a three-neck flask with a stirrer, 150 mL of water was used to dissolve 5 g of lignin and 2.5 g of $Na₂S₂O₅$. Afterward, 10% NaOH was added to the solution to achieve a pH of 7. The solution was synthesized for 4 h at 90 $^{\circ}$ C. A rotating vacuum evaporator Figure 2010 The solution was symmetric for 4 h at 90 °C. The solution's water content, and (Buchi R-300, Switzerland) was used to lower the obtained solution's water content, and a Buchner funnel was used to filter it. Methanol was added to the filtrate to precipitate $Na₂S₂O₅$. The obtained filtrate was oven dried at 60 °C for 24 h. \overline{a} is a \overline{b}

2.2.3. Impregnation of Rattan

Rattan samples were dried at 103 ± 2 °C until their weight remained constant. In the impregnation tube, the sample was submerged in the impregnation solution (the percentage of the lignosulphonate-distilled water solution was 40%). The tube was sucked at a vacuum of 50 cmHg for 60 min at a pressure of 0.8 MPa for 2 h. Figure 2 depicts the design of the impregnation part of the tube (Modification tube, Cibinong, Indonesia). After being wrapped in aluminum foil for 12 h, the sample was taken out and put back in the oven at 103 ± 2 °C until the weight was constant. Samples were soaked in water for 24 h before oven drying at 103 \pm 2 °C until the weight was constant to determine leachability (Equation (2)). The effectivity of impregnation was evaluated using weight percent gain (WPG) as determined in Equation (3). The effectivity of impregnation was evaluated using weight percent gains were gained with α

$$
Leachability (\%) = \frac{(W1 - W3)}{(W1 - W0)} \times 100\% \tag{2}
$$

$$
WPG\ (%) = \frac{(W1 - W0)}{W0} \times 100\% \tag{3}
$$

where W0 denotes dry-weight oven before treatment, W1 denotes dry-weight oven after where W0 denotes dry-weight oven before treatment, W1 denotes dry-weight oven after treatment, and W3 denotes dry-weight oven after soaking in water for 24 h. treatment, and W3 denotes dry-weight oven after soaking in water for 24 h.

Figure 2. Impregnation process. **Figure 2.** Impregnation process.

2.2.4. Lignin Properties Analysis: Moisture Content, Ash Content, Lignin Purity, and Solubility in NaOH and Dioxane

The TAPPI T264 cm-97 method [\[29\]](#page-20-15) was used to determine the moisture content of the lignin. Petri dishes were dried in a 105 °C oven for 4 h. Afterward, the sample was put into the desiccator for 30 min and weighed. Isolated lignin of about 0.1 g was weighed in a petri dish. The sample was put in the oven at 105 °C for 24 h, then put in a desiccator for 30 min and weighed. The moisture content in the lignin sample was calculated according to the standard mentioned (Equation (4)).

Moisture content (
$$
\%
$$
) = $\frac{B - (C - A)}{B} \times 100\%$ (4)

where A is the dry weight of the petri dish (g), B is the dry weight of the sample (g), and C is the dry weight of the petri dish and sample (g).

The TAPPI T211 om-02 method [\[30\]](#page-20-16) was used to determine the lignin ash content. Crucible porcelain was dried at 525 \pm 25 °C for 30–60 min in a furnace (Blue M, New Columbia, OH, USA), then cooled in a desiccator for 30–60 min before measuring the dry weight. An amount of 1 g of the isolated lignin was placed in a crucible porcelain and heated in a furnace for 6 h at a temperature of 525 ± 25 °C. After cooling for 30–60 min in a desiccator, the sample was weighed. The following formula (Equation (5)) was used to determine the amount of ash in the lignin sample:

$$
Ash content (%) = \frac{(C - A)}{B} \times 100\% \tag{5}
$$

where A is the dry weight of the crucible porcelain (g), B is the dry weight of the sample (g), and C is the dry weight of the porcelain cup and ash (g).

The NREL standard (NREL/TP-510-42618) [\[31\]](#page-20-17) was used to evaluate the lignin's purity. The first step in determining acid-insoluble lignin (AIL) was to dry an empty IG3 filter glass at 25 °C for 4 h, then put it in a desiccator for 30 min, and then measure the oven-dried weight. A 0.3 g sample of lignin was placed in a vial and added to 3 mL of 72% H2SO⁴ *w*/*v*, which was then agitated for 2 h at room temperature using a magnetic stirrer. The sample was transferred to a Duran bottle, and 84 mL of distilled water was added until the H₂SO₄ concentration was 4% *w*/*v*. The sample was autoclaved for 1 h at 121 °C and then filtered using an IG3 filter glass. For measuring the acid-soluble lignin (ASL), 10 mL of the filtrate was taken and stored before analysis. After being cleaned with 50 mL of hot water, the sample in the IG3 filter glass was oven dried for 24 h at 105 °C. The sample was placed in a desiccator for 30 min, after which the dry weight was measured. In order to determine the amount of ashed lignin, the residue from the IG3 filter glass was transferred to the crucible porcelain and heated in a furnace at 525 ◦C for 6 h. The AIL was determined by following Equation (6).

$$
AIL (%) = \frac{(A - B) - (C - D)}{E} \times 100\% \tag{6}
$$

where A = dry weight of IG3 and lignin (g), B = dry weight of IG3 (g), C = dry weight of crucible porcelain and ash (g), $D =$ dry weight of crucible porcelain (g), $E =$ dry weight of lignin (g).

A UV-Vis UV-1800 spectrophotometer (Shimadzu, Kyoto, Japan) was used to test the absorbance of the diluted filtrate to measure ASL at a wavelength of 240 nm. An H_2SO_4 4% *w*/*v* solution was used as a blank for auto-zero absorbance of UV-Vis spectrophotometers. The absorbance range for the measurement of acid-dissolved lignin was 0.7–0.9. ASL was obtained from the following Equation (7):

$$
\text{ASL} \left(\% \right) = \frac{A \times V \times fp}{\varepsilon \times BKO \times 1} \times 100\% \tag{7}
$$

where A denotes sample absorbance, V denotes filtrate volume (86.73 mL), fp is dilution factor, ε refers to molar absorptivity (25 M $^{-1}$ cm $^{-1}$), BKO means the dry weight of lignin (g), and l is cuvette length (1 cm).

The total phenolic hydroxyl (phOH) content of lignin was evaluated based on Cantador et al. [\[32\]](#page-20-18); the solution was filtered using microfiltration (0.45 µm) after 20 mg of lignin was dissolved in 10 mL of dioxane and 10 mL of 0.2 M NaOH. A total of 4 mL of the mixture solution was diluted with 50 mL of NaOH to a concentration of 0.08 g/L , and the absorbance was recorded using a UV-Vis Shimadzu UV-1800 spectrophotometer at a wavelength of 200–600 nm with lignin solution and pH 6 buffer as reference. The absorbance of maximum spectra at 300 and 350 nm was used to calculate total phOH by the following Equation (8).

Total phOH $\text{(mmol/g)} = (0.425 \times A_{300 \text{ nm}} \text{ (NaOH)} + (0.812 \times A_{350 \text{ nm}} \text{ (NaOH)} \times a)$ (8)

where *A* is absorbance, *a* is correction term is $1/(c.1) \cdot 10/17$, *c* is lignin solution concentration (g/L) .

The solubility of lignin in base (NaOH) and organic solvent (dioxane) was analyzed based on a study by Hermiati et al. [\[33\]](#page-20-19). Each 5 mg of lignin sample was dissolved in 5 mL of dioxane/distilled water at a ratio of 9:1 or NaOH at pH 12 as the base solvent. The absorbance of both solutions was assessed using a UV-Vis spectrophotometer with a 200–400 nm wavelength range after being diluted 50 times.

2.2.5. Determination of Lignin Monomers

The Pyrolysis-gas chromatography-mass spectrometry (PyGCMS) (Shimadzu GC/MS system QP-2020 NX, Shimadzu, Japan) technique, based on the method of Nakagawa-Izumi et al. [\[34\]](#page-20-20), was used to analyze the lignin types of monomers such as guaiacyl, and syringyl units. The detail analysis condition was similar to that reported in *Acacia mangium* lignin, as reported in a study by Solihat et al. [\[35\]](#page-20-21). The ratio between the peak areas of the chromatograms for the pyrolysis products of the syringyl and guiacyl lignin units served as the basis for calculating the syringyl/guiacyl ratio. The pyrolysis products were analyzed by evaluating mass spectra and retention times with the data library in NIST LIBRARY 2017.

2.2.6. Thermal Analysis: Thermogravimetric Analysis (TGA) and Differential Scanning Calorimetry (DSC)

In a PerkinElmer Thermogravimetric Analyzer (TGA 4000, PerkinElmer, Waltham, MA, USA), Each ~4 mg of lignins and lignosulphonates was weighed into a standard ceramic crucible and subsequently heated at a rate of 10 $°C/min$ from a temperature of 25 \degree C to 750 \degree C. The testing was performed under an argon gas atmosphere with a 20 mL/min flow. The analysis results by TGA were used to determine the weight loss curve against temperature.

The Perkin Elmer DSC (DSC 4000, PerkinElmer, Waltham, MA, USA) was used to determine the glass transition temperature (Tg) value and curing characteristics of lignin that could be computed automatically by DSC 4000. Pyris 1 PerkinElmer software was used to conduct a DSC analysis of lignin and lignosulphonate samples. A total of about 4 mg of weighted and condensed samples were utilized in a standard aluminum pan. The samples were heated at a temperature of 30–100 °C with a heating rate of 10 °C/min and held at that temperature for 3 min. Next, it was cooled from 100 °C to -20 °C with a cooling rate of 10 $\rm{°C/min}$, then held at that temperature for 3 min. Afterward, it was heated to a temperature between 30–200 °C at a rate of 10 °C per minute and maintained for 1 min. Once more, cooling was done at a rate of 10 $\mathrm{^{\circ}C}$ /min from 200 $\mathrm{^{\circ}C}$ to 30 $\mathrm{^{\circ}C}$.

2.2.7. Morphological Characteristics and Elemental Analysis by FESEM-EDX and Ultimate Analyzer

The FESEM (Thermo Fisher SEm Quattro S, The Netherlands) equipped with an EDX Detector, heating stage, WetSTEM, and tensile stage, was used to conduct a topographic and morphological investigation of lignin and lignosulphonate. After placing the sample inside a carbon tube, micrographs were taken under a high vacuum at magnifications of $2500\times$, with a working distance of 11 mm and an acceleration voltage of 5.0 kV. In this magnification, the elemental content of samples was recorded.

Besides EDX analysis, the samples were also analyzed for their elemental content with an ultimate analyzer. The ultimate analyzer (CHNSO LECO CHN-2000, St. Joseph, MI, USA) was used to identify the lignin and lignosulfonate's elemental makeup, which

included carbon (C), hydrogen (H), nitrogen (N), and sulfur (S). The amount of oxygen was estimated by deducting 100 from the sum of the C, H, N, and S contents. Then, the empirical $(C_aH_bO_cS_d)$ n was calculated to determine the number of double bond equivalent (DBE) in lignin and lignosulfonate molecules, according to Equation (9). [\[36\]](#page-20-22).

$$
DBE = C - \frac{H}{2} + \frac{N}{2} + 1 \tag{9}
$$

C denotes carbon; *H* denotes hydrogen and *N* denotes nitrogen.

2.2.8. Lignosulphonate Properties: Yield and Purity

The lignosulphonate yield was determined according to Andriani and Nasrudin [\[37\]](#page-21-0). It was calculated based on the weight ratio between dry lignosulphonate and the total amount of liquid lignosulphonate used. Furthermore, the yield was presented as a weight percentage of lignosulphonate per volume (*w*/*v*) as presented in Equation (10).

The lignosulphonate yield
$$
(\% w/v) = \frac{(D \times A)}{(B \times C)} \times 100\%
$$
 (10)

where A is the initial volume (mL), B is sample volume (mL), C is the dry weight of the lignin (g), and D is the lignosulphonate weight (g).

The lignosulphonate's purity was examined based on Andriani and Nasrudin [\[37\]](#page-21-0). In total, 100 mL of distilled water was dissolved with 0.1 g of lignosulphonate, and 5 mL of the mixture was taken and then diluted to 200 mL. The pH of the solution was adjusted to pH 4 with the addition of 0.125 N NaOH or 0.2 N HCL. A UV-Vis spectrometer with a 232 nm wavelength was used to measure the absorbance after the solution was transferred to a 250 mL volumetric flask with previously its volume fixed using distilled water. The purity of lignosulfonate was determined with Equation (11)

$$
Purity of lignosulphonate (%) = \frac{A232 \times FP}{Factor \times g \times 10} \times 100\%
$$
 (11)

Description *A*232 is the absorbance at 232 nm, *FP* is dilution factor (50), Factor is NaLS factor (35).

Flammability testing was conducted according to UL-94 (Serpong, Indonesia) standard using a vertical burning test (Figure [3a](#page-7-0)). Rattan impregnated with lignosulphonate was tested for flammability. Rattan without any impregnation treatment was used as a control. The sample with a size of $125 \times 13 \times 4$ mm was clamped vertically. Then the sample was burned using methane gas for 10 s at a fixed distance. Afterward, the time (t) after burning until the fire was extinguished was calculated. There are 3 ratings for material in this testing, including V-0 (flame extinguishes within 10 s with no dripping), V1 (flame extinguishes within 30 s with no dripping), and V2 (flame extinguishes within 10 s with dripping).

The manual burning test of samples with the gas torch is according to SNI1740-2008 [\[38\]](#page-21-1) with modification. A sample that had been treated with lignosulfonate was burned manually for two minutes using a gas torch (Figure [3b](#page-7-0)). The sample was then oven dried for 24 h at 105 ◦C in an oven before being weighed and analyzed. When the fire had completely burned, the charred sample was re-weighed.

Figure 3. Flammability testing by UL-94 in vertical burning test (**a**) and gas torch (**b**). **Figure 3.** Flammability testing by UL-94 in vertical burning test (**a**) and gas torch (**b**).

3. Results and Discussion 3. Results and Discussion

3.1. Chemical Component and Solubility of Lignin 3.1. Chemical Component and Solubility of Lignin

Kraft pulping is an effective method to delignify wood as raw materials in the pulp Γ and paper industry for pulp production. Through this process, black liquor is generated, and paper industry for which contains a rich lignin content. Depending on the properties of black liquor in alkaline or acid conditions, lignin can be extracted from this source by precipitation or isolation
 techniques. During the lignin isolation process, acids, bases, or organic solvents have been
 known to precipitate lignin [\[7\]](#page-19-6). Lignin might begin to break down into low-molecular-
known to precipitate lignin [7]. Lignin might begin to break down into low-molecularweight fragments, altering its physico-chemical characteristics $[39]$. I recipitation occurs
when acidic substances are added to alkaline black liquor because of the rise in proton $[H^+]$ when acidic substances are added to alkaline black liquor because of the rise in proton [11] concentration and protonation of ionized phenolic groups in lignin macromolecules. In entermation and protonation of ionized phenone groups in lignin inderomolectiles. In alkaline environments, strong acids dissociate to create lignin with a very high hydroxyl environments, strong acids absociate to create lignin with a very rightly drong.
group concentration [\[40\]](#page-21-3). Also, strong acids such as HCl, which has a pKa of −7, lower the pH of alkaline liquor until it reaches the pKa of phenolic hydroxyl groups, which is around pH of alkaline liquor until it reaches the pKa of phenolic hydroxyl groups, which is around pKa−10. In this reaction, the photoionization of phenolic groups increases, causing ligning pKa-10. molecules to assume a neutral condition, reducing the electrostatic repulsive force between lignin molecules and precipitating part of the lignin molecules [\[41\]](#page-21-4). Hidayati et al. [\[42\]](#page-21-5) added that a decrease in this force creates less hydrophilic lignin molecules, while lignin added that a decrease in this force creates less hydrophilic lignin molecules, while lignin solubility decreases, and lignin deposits increase. weight fragments, altering its physico-chemical characteristics [\[39\]](#page-21-2). Precipitation occurs

As Solihat et al. [\[35\]](#page-20-21), Hermiati et al. [\[33\]](#page-20-19), and Salsabila et al. [\[43\]](#page-21-6) studied, the isolation method, including the frequency of washing treatment during lignin isolation of *Acacia* magium kraft black liquor, influences the lignin characteristics. Washing treatment is one method to increase the purity of lignin; therefore, in this study, an investigation of the washing treatment during acid isolation of black liquor has been observed on lignin characteristics and then lignosulphonate properties.

As demonstrated in Table [1,](#page-8-0) the yield of lignin extracted from black liquor is affected by differences in the frequency of washing. It supports the preceding finding [\[33,](#page-20-19)[35,](#page-20-21)[43\]](#page-21-6), where the higher the frequency of washing with distilled water, the lower the yield, and vice versa. This treatment also affects lignin's purity, which is a total of AIL and ASL. Increasing washing frequency tends to increase the lignin purity. Compared to the purity of A. mangium isolated lignin [33,35], the isolated Eucalyptus kraft lignin has lower purity but

was still higher than commercial lignin. It is proved that the different sources of biomass and isolation methods affected this finding.

Chemical Components (%)	Lignin Reference		Lignin Eucalyptus 3×1 Lignin Eucalyptus $5 \times$
Water content	3.54 ± 0.07	1.58 ± 0.90	2.50 ± 1.83
Yield	$\overline{}$	39.28	36.24
Ash content	$3.67 + 0.28$	$4.20 + 0.20$	3.43 ± 1.05
AII.	86.67 ± 0.44	$76.04 + 0.29$	83.67 ± 1.09
ASL.	3.67 ± 0.11	$9.84 + 0.22$	9.18 ± 0.12
Total of ph-OH	7.09	6.74	5.13

Table 1. Chemical components of Eucalyptus lignin after 3 and 5 washing treatments.

AIL acid-insoluble lignin; ASL: acid-soluble lignin.

Lignin isolated by $3\times$ and $5\times$ washing treatments has a low water content of 1.58 and 2.5%. Suhartati et al. [\[44\]](#page-21-7) believe that the inability of lignin to readily bind to water is to blame for its relatively low water content (hydrophobic). Agustiany [\[45\]](#page-21-8) also stated that lignin has hydrophobic properties, which prevent moisture content changes and make it difficult to bind with water molecules. This content is nearly similar to that of Sameni et al. [\[46\]](#page-21-9), with the water content of isolated lignin from black liquor being 1.13%. However, this water content is lower than Solihat et al. [\[35\]](#page-20-21) and commercial lignin as references. The ash content of isolated lignin after $3\times$ and $5\times$ washing treatments was 4.20%–3.43%. This might be related to the lignin leached during the isolation procedure. The loss of impurities or inorganic materials during the isolation process may be the source of the low ash content in lignin [\[46\]](#page-21-9). Suhartati et al. [\[44\]](#page-21-7) claim that the black liquor's starting material impacts variations in lignin ash content. The ash content affects the lignin purity, whereas a high ash content causes low lignin purity [\[47\]](#page-21-10) Therefore, there was a lower lignin purity in the Eucalyptus isolated lignin than that of Solihat et al. [\[35\]](#page-20-21) and Acacia kraft lignin can also be affected by lower ash content. The high ash content in lignin is caused by dirty raw materials, and the type of acid used in the lignin isolation process influences the impurities produced [\[48\]](#page-21-11). The presence of impurities such as silica and inorganic components (sulfur, potassium, and sodium) is often difficult to remove, thus affecting the resulting yield [\[47\]](#page-21-10). Eucalyptus lignin after $3\times$ washing treatment contains higher sulfur, chlorine, natrium, and silica elements than Eucalyptus after $5\times$ washing treatments based on elemental analysis that will discussed later in Section [3.5](#page-14-0) (morphological and elemental analysis), these elements can be identified as ash in lignin. Therefore, washing treatment is necessary in the isolation process. Kraft pulping is included in the sulfur-bearing process [\[7](#page-19-6)[,26\]](#page-20-12), in which chemicals for delignification contain sulfur; therefore, this element is found in ash determination.

One important consideration while choosing an economically viable lignin isolation process was lignin recovery [\[35\]](#page-20-21). The lignin yield isolated from black liquor is up to 39.28%, which is in the expected yield range (20%–40%), as reported by Vishtal and Kraslawski [\[8\]](#page-19-7). This yield is higher than Acacia lignin isolated with single and fractionated steps [\[35\]](#page-20-21) and oil palm empty fruit bunch lignin, as reported by Hidayati et al. [\[42\]](#page-21-5). Hidayati et al. employed the lignin isolation with hydrochloric acid of 15.61% resulting in the highest lignin yield. The protonation of the phenolic groups in lignin results in a decrease in the electrostatic repulsion between lignin molecules generating lignin deposition in the isolation process [\[41\]](#page-21-4). As a result, the solubility of lignin decreases, and the lignin molecules become less hydrophilic.

The total phenolic hydroxyl content of isolated lignin after $3\times$ washing treatment is higher than that of $5\times$ washing treatment. These values are lower than phenolic hydroxyl content in Acacia kraft lignin and also in the lignin reference. The isolation method, source of biomass, and kraft pulping conditions contribute to this difference. Kraft pulping and precipitation of lignin with HCl contribute to creating a condensed structure in lignin and phenolic hydroxyl groups [\[49](#page-21-12)[,50\]](#page-21-13). Several chemical groups found in lignin, including phenolic hydroxyl groups [49,50]. Several chemical groups found in lignin, including hydroxyl (phenolic or aliphatic), hydrolyzed ether bonds, carboxyl, methoxy, carbonyl, hydroxyl (phenolic or aliphatic), hydrolyzed ether bonds, carboxyl, methoxy, carbonyl, benzyl alcohol, and condensing units in C2, C5, and C6, influence lignin's reactivity [\[51\]](#page-21-14). benzyl alcohol, and condensing units in C2, C5, and C6, influence lignin's reactivity [51]. Additionally, as a larger degree of condensation promotes the thermal degradation of the Additionally, as a larger degree of condensation promotes the thermal degradation of the polymer, the total phenolic hydroxyl content is connected with the Tg value [\[52\]](#page-21-15). polymer, the total phenolic hydroxyl content is connected with the Tg value [52].

This study used UV spectrophotometry dissolved in dioxane-distilled water and This study used UV spectrophotometry dissolved in dioxane-distilled water and NaOH to analyze lignin isolated by $3\times$ and $5\times$ washing as well as a lignin reference to determine the purity of the lignin with semi-quantitative results and molecular distribu-tion [\[35\]](#page-20-21). Figure [4b](#page-9-0) shows that the dioxane-distilled water solution has two prominent peaks for lignin after 3 \times and 5 \times washing treatment at around 240 nm and 280 nm, respectively. The difference in washing frequency treatment does not influence the UV spectra pattern that is similar to the lignin reference. Each kind of lignin emits light in the visible .
range, with the intensity varying just slightly. The absorption coefficients of isolated lignins are nearly identical even though the purity of the lignin $5\times$ washing treatment is higher than that of the lignin reference. Ibrahim et al. [\[53\]](#page-21-16) assert that this results in the bonding of the unconjugated phenolic group (aromatic ring). Ammar et al. [\[54\]](#page-21-17) also found that the presence of unconjugated phenolic groups in the lignin structure is indicated by the visible peak in the wavelength of 280 nm. Lignin was also soluble in NaOH, as presented in Figure 4a. Two peaks appeared in the spectra identified as unconjugated phenolic groups (excitation of π - π ^{*}) with absorption at 215–222 nm as the first peak. This band is the effect of hypsochromic NaOH [35]. In the wavelength of 296-303 nm, conjugated phenolic groups can correspond to the conjugate[d ph](#page-21-18)enolic group due to π - π ^{*} excitation [55]. Similar spectra were also found in Acacia lignin [35] when dissolving in NaOH and dioxane. The highest peak belongs to the lignin reference that correlates with the highest phenolic hydroxyl groups, as presented in Table 1 .

Isolated lignin from Eucalyptus sp was then subjected to sulfonation to produce Isolated lignin from Eucalyptus sp was then subjected to sulfonation to produce lignosulphonate. Lignosulphonate derived from lignin after 3× washing treatment lignosulphonate. Lignosulphonate derived from lignin after 3× washing treatment (ligno-sulphonate 3×) presents a higher yield than lignosulphonate [5×](#page-10-0) (Table 2). Huang et al. [\[56\]](#page-21-19) stated that the material, duration, and temperature utilized during the sulfonation process affect the yield of lignosulphonate. The yield increases as the material-to-lignin ratio used increases. The appropriate time for the sulfonation process is around 4 h; even if done longer, the yield does not increase significantly.

Sample	Water Content (%) Yield (%)		Purity $(\%)$	Solubility
Lignosulphonate reference	5.37	-	16.19	Water soluble
Lignosulphonate Eucalyptus $3\times$	1.58	93.39	71.89	Water soluble
Lignosulphonate Eucalyptus $5\times$	2.50	91.95	67.21	Water soluble

Table 2. Chemical components of lignosulphonate after 3× and 5× washing treatments.

Furthermore, the sulfonation process can only be carried out at a maximum temperature of 110 ◦C; higher temperatures would result in lower yields. Lignin becomes water-soluble when it is converted into lignosulphonate, losing its hydrophobic properties. Lignosulphonate includes water-soluble polyelectrolytes because they have anions in the form HSO_3^- and $\text{SO}_3{}^{2-}$ and have a broad molecular weight distribution [\[10\]](#page-19-9).

Lignosulphonate after $3\times$ washing has the highest purity of 71.89% compared to lignosulphonate with $5\times$ washing and commercial lignosulphonate. It may be affected by the higher ASL content of lignin after $3\times$ washing treatment. Nawawi et al. [\[57\]](#page-21-20) suggested that a significant proportion of ASL has a high β-O-4 structure, making it more reactive. The purification procedure and the lignin origin as raw materials influence the lignosulphonate purity. According to Muharom et al. [\[58\]](#page-21-21), a small number of contaminants, the majority of which are sodium bisulfite, are responsible for the high purity of lignosulphonates. Andriani and Nasrudin [\[37\]](#page-21-0) reported that the high purity of lignosulphonates results from employing lignin in isolation, including other compound components, which can lower the purity.

The yield of the lignosulphonate after $3\times$ washing treatment was higher than the yield of the lignosulphonate with $5\times$ washing treatment, at 93.39 and 91.95%, respectively. Huang et al. [\[56\]](#page-21-19) reported that the material, duration, and temperature utilized have an impact on the yield of lignosulphonate produced during the sulfonation process. The yield increases with the utilization of the material-to-lignin ratio. The sulfonation procedure should be completed within a maximum of 4 h. The yield does not greatly increase even when it is performed for up to 4 h. Furthermore, a temperature above the sulfonation process's maximum operating temperature of 110 ◦C will simply decrease the yield that is obtained. When lignosulphonate is generated from lignin, it transforms the substance so that it is soluble in water. Because they contain HSO_3^- , and $SO_3^2^-$ anions, and have a broad molecular weight range, lignosulphonates are polyelectrolytes that are able to dissolve in water [\[59\]](#page-21-22).

3.2. Functional Group of Lignin and Lignosulphonate

FTIR analysis is used to characterize functional groups of obtained lignin, as well as to evaluate the structure of isolated lignin in general and compare it to lignin reference structures. Figure 5 shows the structure of isolated lignin after $3\times$ and $5\times$ washing treatment, and commercial lignin as a reference. The spectrum at wavenumbers around 3300 cm $^{-1}$ (no 1) corresponds to hydroxyl group (O–H) vibrations in phenolic and aliphatic structures. Jeong et al. [\[60\]](#page-21-23) reported that the hydroxyl group occurs at 3362 cm $^{-1}$. Muharom et al. [\[58\]](#page-21-21) report that a large band associated with the OH group was found at wavenumbers around 3200–3400 $\rm cm^{-1}$, while Solihat et al. [\[35\]](#page-20-21) identified it at 3500–3400 $\rm cm^{-1}$. The presence of a wide band at the peak (no 1) indicates the presence of a higher phenolic O-H content due to the breaking of the C–O bond during the lignin extraction process resulting in the formation of phenolic rings. The larger molar mass is most likely the cause of the lower O−H spectrum intensity, which was linked to an increase in lignin concen-tration [\[61\]](#page-21-24). The presence of a small peak in the spectrum at 2928 cm⁻¹ indicates C-H stretching in methylene from the side chain and aromatic methoxy group [\[62](#page-21-25)[,63\]](#page-21-26) or aliphatic and aromatic C−H groups [\[58\]](#page-21-21). In the lignin structure, wave numbers 1688, 1588, and 1503 cm⁻¹ show the presence of a conjugated carbonyl with a benzene ring and a benzene ring vibration [\[64\]](#page-22-0). It is in agreement with a previous study in which vibration of the aromatic skeleton in lignin in the wave number of 1460–1590 cm⁻¹ [\[65\]](#page-22-1) also appeared in Acacia kraft lignin [\[35\]](#page-20-21). The aromatic skeletal vibration with deformation in C−H

asymmetric bonds on methyl groups [\[61](#page-21-24)[,66\]](#page-22-2), C−O bonds on syringyl monomers [\[67\]](#page-22-3), and C−C, C−O, and C=O bonds on guaiacyl monomers [\[68\]](#page-22-4) were displayed in the spectra of C−C, C−O, and C=O bonds on guaiacyl monomers [68] were displayed in the spectra of wavenumbers 1453, 1368, and 1211 cm $^{-1}$, respectively. The spectra 1125 and 1089 cm $^{-1}$ indicate the deformation of the C–H bond in the syringyl monomer [68] and the aromatic indicate the deformation of the C–H bond in the syringyl monome[r \[6](#page-22-4)8] and the aromatic deformation of C–H, the presence of strains of the C–O and C–C bonds [67]. deformation of C–H, the presence of strains of the C–O and C–C bond[s \[6](#page-22-3)7].

Forests **2022**, *13*, x FOR PEER REVIEW 12 of 24

Figure 5. FTIR spectra of lignin and lignosulphonate at: (**a**) wavenumber range of 4000–400 cm[−]1, **Figure 5.** FTIR spectra of lignin and lignosulphonate at: (**a**) wavenumber range of 4000–400 cm−¹ , and (**b**) wavenumber range of 1800–400 cm[−]1. and (**b**) wavenumber range of 1800–400 cm−¹ .

The lignosulphonate synthesized from the Eucalyptus lignin is very similar to that of The lignosulphonate synthesized from the Eucalyptus lignin is very similar to that of the lignosulphonate reference (Figure 4). The O-H bonds are also found in spectra with the lignosulphonate reference (Figure [4\)](#page-9-0). The O-H bonds are also found in spectra with the wide band that is visible in wave number 3312 cm⁻¹. The small peak in the wavenumber of 2921 cm^{−1} is associated with C-H stretching vibration on the methyl and methylene groups. The peaks at the wavenumber of 1581 and 1420 cm^{−1} revealed the backbone vibration of the $C = \overline{C}$ bond on the aromatic ring and the symmetrical and asymmetrical bending vibrations of the C-H bond on the methyl group. Sharp peaks at 1111 and 1040 cm⁻¹ corresponded to the presence of the C-O-C bond strain and the C-O and S=O bond strain, respectively. The transformation of lignin into lignosulphonates can be identified as successful because of the presence of a new spectrum peak at wave number 627 cm⁻¹, which denotes an S-O \bm{b} bond [69,70].

3.3. Lignin Composition 3.3. Lignin Composition

PyGCMS is a versatile method for determining lignin degradation products and PyGCMS is a versatile method for determining lignin degradation products and other components such as carbohydrates or additives [\[71\]](#page-22-7). The elemental composition, amount of produced products, nature of lignins, and isolation technique can all be clarified using the PyGCMS approach [\[72\]](#page-22-8). The Eucalyptus lignin employed in this research is derived from hardwood. Due to the different raw materials used in the isolated lignin, the syringyl (S)/guiacyl (G) ratio of isolated lignin could not be compared to the reference lignin. The abundance of lignin monomers may vary depending on the raw material, such as hardwood, softwood, or grass. Hardwood lignin contains S and G units in specific ratios. Like Acacia lignin [\[35\]](#page-20-21), which is included in hardwood lignin, the S unit dominates the pyrolysis composition. Meanwhile, the lignin reference contains more G units which are typical for softwood lignin, with fewer p-hydroxyl phenyl (H) units [\[73\]](#page-22-9). In addition,
example: Solihat et al. [\[35\]](#page-20-21) reported that reference lignin is rich in G-lignin, and proposed that it is a
 kind of softwood lignin. Compared to Acacia lignin, the S/G ratio of Eucalyptus lignin is
. higher. It is demonstrated that the difference in lignin sources, as well as a method for lignin in the difference in lignin sources, as well as a method for lignin isolation, contributes to this finding. Because of the presence of the C-5 position, which
 $\frac{1}{2}$ binds to C-C, lignin with a high percentage of G units is less reactive [\[51\]](#page-21-14). However, the two isolated lignins have more S units than G units, making them more reactive due to the

methoxy groups found in the structure of S-type lignin [\[74\]](#page-22-10). The S/G ratio of Eucalyptus lignin after $3\times$ washing treatment is 4.49, which is higher than Eucalyptus lignin after $5\times$ washing treatment (Table [3\)](#page-12-0) suggesting that lignin after $3\times$ washing treatment is more reactive than lignin with $5\times$ washing treatment. This finding is related to the fact that the amount of ASL present is directly proportional to the high S ratio [\[75\]](#page-22-11). Because the pulping process with a high S ratio reduces the condensation reaction, lignin reactivity and condensation levels are inversely related. As C-C linkages are more difficult to break down into aromatic monomers than ether bonds, condensed structures can reduce yield [\[76\]](#page-22-12).

Sample	Proportion $S(\%)$	Proportion $G(\%)$	S/G Ratio
Lignin reference	0.59	45.85	0.01
Lignin Eucalyptus $3\times$	55.06	12.25	4.49
Lignin Eucalyptus $5\times$	50.61	14.67	3.45

Table 3. The proportion of types of monomers in Eucalyptus lignin with PyGCMS.

3.4. Thermal Behavior of Lignin and Lignosulphonate

As a fire retardant, the investigation of thermal characteristics such as Tg and mass loss is the most critical assessment that must be performed. Besides the burning test, TGA and DSC have been reported as methods for assessing the thermal and flammability of natural fiber-reinforced composites [\[16\]](#page-20-23). The ability of lignin with a phenolic structure to suppress fire depends on the origin of the lignin and the extraction technique. This ability as a flame retardant is influenced by its thermal properties, which include glass transition temperature (Tg), thermal degradation, molecular weight, lignin purity, and phenolic concentration [\[26\]](#page-20-12). The varied Tg values were due to the heterogeneous structures and the broad molecular weight of isolated lignin samples [\[76\]](#page-22-12). These factors also affect interchain hydrogen bonding, cross-linking density, and rigid phenyl groups [\[77\]](#page-22-13). Although several studies reported an increase in char residue with higher Tg values, the results may be inconsistent due to the plant sources and extraction conditions [\[46\]](#page-21-9).

As can be seen from the study in Figure [6,](#page-13-0) there are three distinct steps to the degradation of the mass loss process in the lignin and lignosulphonate samples. There was a steady weight loss in the first stage when the temperature ranged from $40-130$ °C. This indicates that the water content of the lignin sample had been evaporated [\[78\]](#page-22-14). Li et al. [\[79\]](#page-22-15) reported that water evaporation begins at a temperature of 55 ◦C while Kazak et al. [\[80\]](#page-22-16) argue that chemically bonded water can be released at temperatures as low as 400 ◦C. It is suggested that water molecules are bound tightly in the lignin structure of the lignin reference, and it is possible that the lignin reference loses its free water at a temperature of 60–70 °C. This mass loss occurred in the lignin samples at 60–70 °C, with weight loss percentages of 4, 11, and 10% for the reference lignin, Eucalyptus after $3\times$ and $5\times$ washing treatment, respectively. At 50–60 ℃, the weight loss of the lignosulphonate reference, and eucalyptus after $3\times$, and $5\times$ washing treatment are 4, 8, and 7%, respectively. The second stage involves the carbohydrate breakdown attached to the lignin structure, which begins between 150–250 \degree C [\[81\]](#page-22-17). In this stage, the decomposition of lignin into degradation products, such as aromatic decomposition as a phenolic compound, including ether linkages among the C9 units, might occur [\[82\]](#page-22-18). The mass loss of lignin and lignosulphonate peaks in both figures at a temperature of 300–400 °C, brought on by the release of CO and CO₂ from the lignin structure [\[83\]](#page-22-19). Compared to isolated lignin after $3\times$ and $5\times$ washing treatment, which both have a mass loss of 34%, the lignin reference had a mass loss at the rate of 43%. Lignosulphonate reference, Eucalyptus lignin after $3\times$ and $5\times$ washing treatment results in less mass loss than lignin, which loses mass at rates of 15, 28, and 35%.

Figure 6. TGA and DTG of lignin (**a**) and lignosulphonate (**b**). **Figure 6.** TGA and DTG of lignin (**a**) and lignosulphonate (**b**).

The beginning degradation temperatures for the third stages of the lignin and The beginning degradation temperatures for the third stages of the lignin and lignosulphonate samples were 500 °C and 400 °C, respectively. However, these steps demon-strated the degradation of the lignin's aromatic ring [\[84\]](#page-22-20). The proportion of residues produced by the three distinct lignins at temperatures over 700 °C was nearly the same, with residues of the reference lignin, Eucalyptus lignin after $3\times$ and $5\times$ washing treatment being 44, 44, and 42%, respectively. The three lignosulphonate samples had varying amounts of residue. The lignosulphonate reference, Eucalyptus lignin after $3\times$ and 5× washing treatments were 68, 32, and 48%, respectively. The proportion of isolated 5× washing treatments were 68, 32, and 48%, respectively. The proportion of isolated lignin residue is similar to the research by Handika [et a](#page-22-17)l. $[81]$, which demonstrated that when isolated lignin is combined with hydrochloric acid, it yields 46% residue, allowing it to be used at high temperatures. According to Gordobil et al. [\[52\]](#page-21-15), The presence of aromatic phenyl groups and hydroxyl groups increases the durability of the aromatic structure in lignin and protects it from temperature damage. As a result, lignin has high thermal stability. Based on the results of the TGA thermal analysis, the synthesized lignin into lignosulphonates can improve thermal stability and be used as a refractory additive.

DSC analysis of lignin thermal properties can provide the Tg value of lignin and DSC analysis of lignin thermal properties can provide the Tg value of lignin and lignosulphonate, which benefits future strategy utilization [52]. Eucalyptus lignin's Tg lignosulphonate, which benefits future strategy utilization [\[52\]](#page-21-15). Eucalyptus lignin's Tg values after 3 \times and 5 \times washing treatments are higher than the Tg of the reference lignin at 141, 147, and 141 °C (Table [4\)](#page-13-1), respectively. These Tg are lower compared to the Tg of Acacia lignin [35,43] and the Tg of *Eucalyptus grandis* lignin at about 161 °C [85]. However, Acacia lignin [\[35,](#page-20-21)[43\]](#page-21-6) and the Tg of *Eucalyptus grandis* lignin at about 161 ◦C [\[85\]](#page-22-21). However, Tg obtained in this study is higher than the Tg value of kraft lignin from Eucalyptus, Tg obtained in this study is higher than the Tg value of kraft lignin from Eucalyptus, which is 133 °C [\[86\]](#page-22-22). The difference in Tg is influenced by molecular weight, thermal history, lignin isolation method, adsorbed water $[87]$, and ash content. Solihat et al. $[35]$ discovered that single-step lignin with lower ash content has a higher Tg than lignin fractioned step. This Tg value influences the stability of lignin at high temperatures; thus, single-step lignin is more stable and suitable for high-temperature process conditions [\[35\]](#page-20-21).

Table 4. Tg value of lignin and lignosulphonate. **Table 4.** Tg value of lignin and lignosulphonate.

The Eucalyptus lignosulphonate with $5\times$ washing treatment shows a higher Tg value (174 °C) than the reference lignosulphonate (158 °C). It is proposed that the char residue of lignosulphonate with $5\times$ washing treatment be increased. Different value ranges represent variations in the lignin structure's flexibility or stiffness, affecting elements such as molecular mass, cross-link density, and hydrogen connections between chains [\[46\]](#page-21-9). Solihat et al. [\[88\]](#page-22-24) claimed that the Tg value shows that the endothermic process changed the lignin structure from a glass-like state to a rubbery (plastic) state. The Tg value of lignosulphonate with $5\times$ washing treatment compared to that of Eucalyptus lignin with the same water treatment frequency, indicating improvement in thermal properties, char residue, and flammability behavior.

3.5. Morphological and Elemental Analysis of Lignin

FESEM EDX was used to determine the morphological characteristics and the elemental constituents of isolated lignin after $3\times$ and $5\times$ washing treatments (Figure [7\)](#page-15-0). In order to assess and identify the materials and impurities present in a sample, EDS analysis may detect elemental content with a minimum concentration of 0.1% [\[89\]](#page-22-25). The types of lignin that influence the elemental analysis of kraft lignin can be seen in Table [5.](#page-15-1) After $3\times$ washing treatment, lignin is still composed of Na and S elemental content (Table [5\)](#page-15-1). This is affected by the kraft pulping process, which uses chemicals such as sodium hydroxide and, sodium sulfide as white liquor [\[7\]](#page-19-6). The Cl elemental content was found in isolated lignin caused by the utilization of hydrochloric acid in the lignin isolation process. It cannot be removed by washing treatment. Due to its abundance of functional groups and benzene rings, lignin is a prime candidate as a precursor for carbon-based products (Tables 6 and 7) and has a Is a prime candidate as a precursor for carbon-based products (tables 6 and 7) and has a
high carbon value [90]. Double bond equivalents (DBE) in lignin and lignosulfonate can be calculated using elemental analysis. DBE denotes a double bond in a structure that can be edictanced asing elemental analysis. DEE denotes a double bond in a stracture that can be
used to predict the possibility of an unconjugated bond, which is more reactive to other substances. A C=C bond can be found in the aromatic ring of lignin, which may be less than a single bond in an aliphatic structure. An additional reaction is easily promoted by the reactive site in the unsaturated bond. DBE also indicates the degree of lignin condensation researce of aromatic ring structures in lignin molecules [47]. $\frac{d}{dt}$ different structures are formed after the form lignin is subsequently to form light $\frac{d}{dt}$. number 627 cm α , which is the annumber of the state distribution of α and α lower α . In the state decay is a lower than α 3). T_{total} for μ is the empirical formula of light μ in T_{total} and μ and μ α different structures are formed after the form in α and α in the form light structure in the form light structure in α ed to prodict the possibility of an unconjugated bond which is more reactive to other syring than guiacyl as calculated in the S/G ratio $\frac{1}{2}$ ratio from PyGCMS and PyGCMS 3). no candidate as a precursor for carbon based products (fubres 0 and 7) and has a s you content than guiacyl as calculated in the S/G ratio from PyGCMS and $\frac{1}{2}$ 3). If as the presence of aromatic ring structures in lignin molecules $\left[47\right]$ sulfide as white liquor [7]. The CI elemental content was found in isolated lignin caused by
the utilization [o](#page-16-0)f hydrochloric acid in the lig[nin](#page-16-1) isolation process. It cannot be removed by
washing treatment. Due to its abund here. A C=C bond can be found in the aromatic ring of lignin, which may be less than e bond in an aliphatic structure. An additional reaction is easily promoted by the e site in the unsaturated bond. DBE also indicates th nances is a communication of the measure of a second community of the manufacture of the same of the same second in an alinhatic structure. An additional reaction is easily promoted by the s_g l as the presence of aromatic ring structures in lignin molecules K 71 3). as the presence of aromatic ring structures in lignin polecules $[47]$

Table 5. FESEM-EDX analysis on lignin and lignosulphonate. $\begin{bmatrix} 1 & 0 \\ 0 & 1 \end{bmatrix}$ **Table 5.** FESEM-EDX analysis on lignin and lignosulphonate.

Content (%) $\bf{(B)}$ **Reference E 3× E 5× Reference E 3× E 5×** $C_{\rm 3.3}$ $C_{\rm 3.3}$ $D_{\rm 3.3}$ $D_{\rm 3.3}$ $\mathcal{L}(\mathcal{L}(\mathcal{L}))=\mathcal{L}(\mathcal{L}(\mathcal{L}))$ $S \subset \mathbb{R}$, $S \subset \mathbb{R}$ \mathbb{C} . \mathbb{C} , \mathbb{C} $N_{\rm e}$ 1 $N_{\rm e}$ 1 $N_{\rm e}$ 1 $N_{\rm e}$ Si - 4.7 - 4.7 - 4.7 - 4.7 - 4.7 - 4.7 - 4.7 - 4.7 - 4.7 - 4.7 - 4.7 - 4.7 - 4.7 - 4.7 - 4.7 - 4.7 - 4.7 - 4.7 Al 0.6 - - - - - (D) **Content (%)** $\bf{(B)}$ **Reference E 3× E 5× Reference E 3× E 5×** $\mathcal{L} = \mathcal{L} \cup \mathcal{L}$ O 24.6 21.8 26.4 33.3 26.9 28.6 $\mathcal{S} \subset \mathcal{S} \subset \mathcal{S}$, and the set of $\mathcal{S} \subset \mathcal{S}$, and $\mathcal{S} \subset \mathcal{S}$ Cl - 2.4 - 2.5 1.3 - $\mathcal{A} = \mathcal{A} - 1 - 1 - 1$ Si - 4.7 - 5.8 - 4.7 - 5.8 - 5.8 - 5.8 - 5.8 - 5.8 - 5.8 - 5.8 - 5.8 - 5.8 - 5.8 - 5.8 - 5.8 - 5.8 - 5.8 - 5.8 Al 0.6 - - - - - (D) **Content (%)** $\bf{(B)}$ **Reference E 3× E 5× Reference E 3× E 5×** $\mathcal{L} = \mathcal{L} \cup \mathcal{L}$ \mathcal{L} and \mathcal{L} $\mathcal{L} = \{ \mathcal{L} \mid \mathcal{L} \in \mathcal{L} \}$ Cl - 2.4 - 2.4 - 2.4 - 2.4 - 2.4 - 2.4 - 2.4 - 2.4 - 2.4 - 2.4 - 2.4 - 2.4 - 2.4 - 2.4 - 2.4 - 2.4 - 2.4 - 2.4 \mathbb{R}^n and \mathbb{R}^n Si - 4.7 - 4.7 - 4.7 - 4.7 - 4.7 - 4.7 - 4.7 - 4.7 - 4.7 - 4.7 - 4.7 - 4.7 - 4.7 - 4.7 - 4.7 - 4.7 - 4.7 - 4.7 Al 0.6 - - - - - (D) **Content (%)** $\bf{(B)}$ **Reference E 3× E 5× Reference E 3× E 5×** C 73.3 57.0 70.4 35.9 49.4 49.1 $\mathcal{L}(\mathcal{A})=\mathcal{L}(\mathcal{A})$ and $\mathcal{L}(\mathcal{A})=\mathcal{L}(\mathcal{A})$. 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The set of \mathbb{R}^n $\mathcal{P} = \{ \mathcal{P} \mid \mathcal{P} \in \mathcal{P} \}$ Na 0.8 10.0 3.2 17.8 11.6 11.5 \mathbb{S}^n - \mathbb{S}^n , \mathbb{S}^n , \mathbb{S}^n , \mathbb{S}^n , \mathbb{S}^n , \mathbb{S}^n , \mathbb{S}^n Al 0.6 - - - - - (ν) **Content (%)** $\left(\frac{2}{\pi} \right)$ **Reference E 3× E 5× Reference E 3× E 5×** C 73.3 57.0 70.4 35.9 49.4 49.1 $\mathcal{L} \rightarrow \mathcal{L}$ \mathbb{C} . The set of \mathbb{C} and \mathbb{C} and \mathbb{C} . The set of \mathbb{C} Cl - 2.4 - 2.5 1.3 - Na 0.8 10.0 3.2 17.8 11.6 11.5 $\mathbb{S} \times \mathbb{S}$, $\mathbb{S} \times \mathbb{S}$ Al 0.6 - - - - - $(\boldsymbol{\nu})$ (**A**) (**B**) *Forests* **2022**, *13*, x FOR PEER REVIEW 16 of 24 (**C**) (**D**) (**E**) (**F**)

Figure 7. FESEM analysis on lignin reference (A), Eucalyptus $3 \times$ (B) and $5 \times$ (C) washing treatment; lignosulphonates reference (**D**), Eucalyptus 3× (**E**) and 5× (**F**) washing treatment. lignosulphonates reference (**D**), Eucalyptus 3× (**E**) and 5× (**F**) washing treatment.

Table 5. *Cont.*

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Table 6. Chemical constituent elements with FESEM EDX.

Table 7. Analysis C, H, O, N, and S of lignin and lignosulphonate.

Sample	C(%)	H $(\%)$	N(%)	O(%)	DBE	Empirical Formula
Lignin reference	61.07	5.94	0.66	32.33	59.43	$C_9H_{10.50}O_{3.57}$
Lignin eucalyptus $3\times$	57.26	5.53	0.12	37.09	55.56	$C_9H_{10.43}O_{4.37}$
Lignin eucalyptus $5\times$	56.86	5.66	0.16	37.32	55.11	$C_9H_{1075}O_{443}$
Lignosulphonate reference	16.59	2.92	12.08	68.41	22.17	$C_9H_{5,54}O_{8,12}$
Lignosulphonate eucalyptus $3\times$	28.24	3.52	12.09	56.15	33.53	$C_9H_{13,46}O_{13,42}$
Lignosulphonate eucalyptus $5\times$	24.86	3.72	12.74	58.68	30.37	$C_9H_{16,15}O_{15,93}$

DBE: double bond equivalent.

The DBE numbers of the Eucalyptus lignin after $3\times$ and $5\times$ washing treatments were also comparable with a value of 55.56 and 55.11, respectively. The DBE values of isolated lignin are similar to that of the reference lignin. The DBE values for the lignosulphonate Eucalyptus with $3\times$ and $5\times$ washing treatment were 33.53 and 30.37, respectively. Compared to the DBE of lignin, the DBE of lignosulfonate is lower. It is suggested that some OH groups in the aromatic structure of lignin are replaced by $SO₃H$ in electrophilic aromatic substitution. This significant difference in DBE also shows structural changes in molecules by sulfonation of lignin into lignosulfonate. However, washing did not change the structure of the lignin, as evidenced by similar DBE between lignin washed $3\times$ and $5\times$. A similar statement has been reported by Hermiati et al. [\[33\]](#page-20-19).

Ultimate analysis (Table [7\)](#page-16-1) shows the presence of N element, a protein-lignin complex that might be formed during the pulping process [\[91\]](#page-22-27) and its existence in the lignin indicated that isolation and washing treatment cannot remove the N element. The amount of N in the lignosulfonate increased dramatically, indicating that N was tightly bound to the lignin molecules as a protein-lignin complex and could not be lost during sulfonation. The lignin washing treatment has the potential to reduce contaminants. Sodium metabisulfite converts lignin into lignosulphonates, raising the elemental content of S for both lignosulphonates. In contrast, sodium hydroxide elevates the solution's pH to 7 to produce the elemental content of Na.

Ultimate analysis (Table [7\)](#page-16-1) presents the empirical formula of lignin and lignosulfonate that is deducted from the elemental component in the samples. Eucalyptus lignin after $3\times$ and $5\times$ washing treatment has a relatively similar formula because of the non-significant difference in the content of C, H, N, and O in lignins. This formula is also similar to the reference lignin indicating that the isolation process successfully produced lignin with a similar structure. The computation was carried out similarly for H and O, producing a chemical formula for lignin $(C_9H_xO_y)$ [\[92\]](#page-23-0).

The empirical formula of lignosulfonate differs from that of lignin, indicating that different structures are formed after lignin is sulfonated to form lignosulfonate. It is strengthened by the presence of a new peak in lignosulfonate FTIR spectra at wave number 627 cm−¹ , which is known as an S-O bond. As stated by Sameni et al. [\[93\]](#page-23-1), a lower weight percentage of carbon and a greater weight percentage of oxygen are present in lignins with

a greater amount of methoxyl groups. The Eucalyptus lignin has higher syringyl content than guiacyl as calculated in the S/G ratio from PyGCMS analysis (Table [3\)](#page-12-0).

3.6. Flammability Behaviour

Fire retardants can reduce the danger of fire in products while extending their useful lives and safeguarding the environment by reducing local pollutants and long-term environmental consequences by avoiding ignition, slowing fire propagation, and reducing flashover time [\[14\]](#page-20-2). Time to ignition, heat release rate, spontaneous ignition and flash point temperature, thermal stability index, smoke toxicity, mass loss, extinction flammability index, limiting oxygen index (LOI), fire resistance, and flame propagation on the surface are the most crucial factors.

Previously, Solihat et al. [\[26\]](#page-20-12) noted that even though lignin can reach better thermal properties of polymeric material, the complex structure, source difference, and isolation method can influence the resulting performance as a bio-based flame retardant. As a result, the functionalization of lignin is a method of improving performance. The sulfonation of kraft lignin into lignosulphonate was used in this study. Angelini et al. [\[25\]](#page-20-11) tried to use unmodified lignosulphonate to improve the fire retardancy of particleboard. The peak heat release rate (HRR) and total smoke production were successfully decreased, while char formation increased by using 15% wt.% of lignosulphonates.

Oven-dried rattan was impregnated with a 40% (*w*/*v*) lignosulphonate-distilled water solution. The impregnation procedure yielded a WPG that varied between 20%–23% as can be seen in Table [8.](#page-17-0) There was no significant difference between rattan impregnated with the lignosulfonate reference and lignosulfonate synthesized from Eucalyptus lignin. The leachability test was performed after the rattan had been impregnated by submerging the specimen in water for 24 h. The proportion of leachability ranges from 94 to 98% (Table [8\)](#page-17-0). The easily soluble in water nature of lignosulphonates [\[94\]](#page-23-2) contributes to the high percentage of leachability in impregnated rattan by causing significant loss of lignosulphonates from rattan during water submersion. Because impregnated rattan is not waterproof, it will have an impact on its future use for indoor purposes. In the future, lignosulfonate can be used as a fire retardant in exterior applications such as coating the material after impregnation or combining it with adhesive while preparing boards. Incorporating lignosulfonate with commercial hydrophobic polymers in the preparation of composites can improve both water and fire resistance. According to Lubis et al. [\[95\]](#page-23-3), impregnating ramie fibre with lignin-based polyurethane can increase heat stability by 6%.

Table 8. WPG and leachability of impregnated rattan.

Flammability behavior on impregnated rattan was initially assessed using UL-94 with the vertical burning mode (Figure [3a](#page-7-0)) and manual burning observations by gas torch (Figure [3b](#page-7-0)). The untreated rattan did not receive a UL-94 rating and burned out in 2 min when tested with a gas torch. One of the three impregnated rattans failed to meet the UL-94 V-0 rating. The received ratings of all samples are displayed in Table [9.](#page-18-0) After a burning test with a gas torch, the samples lost the weight in which the lignosulfonate impregnation has been successfully prevented in high weight loss of rattan (Table [10\)](#page-18-1). The time after burning for impregnated rattan by lignosulphonate after $3\times$ and $5\times$ washing treatments was less than 6 s, classifying it as V-0. The untreated rattan shows robust burning without molten droplets after the sample was burned for 10 s. In comparison, under the same circumstances, lignosulphonate-impregnated rattan may put out a fire in less than 6 s without dripping. The untreated rattan was totally burned until all samples were lost (Figure [8a](#page-18-2)) or burned out in the manual burning test, while impregnated rattan had the

ability to reduce the burning rate. This can be proved by the fact that the impregnated rattan was still partially intact after burning (Figure [8b](#page-18-2)–d). The impregnation of rattan with lignosulfonate resulted in a higher UL-94 rating classification than the lignosulfonate reference-treated rattan. Consequently, this result has demonstrated that the impregnation of lignosulphonates into rattan successfully improves the fire resistance of rattan. This treatment helps to improve flammability performance and meet industrial requirements, in which achieving a V-0 rating [\[26\]](#page-20-12).

Table 9. UL-94 classification of rattan with no treatment, impregnation of lignosulphonate reference, eucalyptus after $3\times$, and $5\times$ of washing treatments.

Sample	t after Flame (s)	t after Extinguished (s)	Dripping	Rating
Rattan	47.33	378.02	No	-
Impregnated rattan by lignosulphonate reference	41.67		No	$V-1$
Impregnated rattan by lignosulphonate Eucalyptus $3\times$	5.67		No	$V-0$
Impregnated rattan by lignosulphonate Eucalyptus $5\times$	2.07	0.67	No	V-0

Table 10. Flammability test of rattan with torch gas for 2 min.

Figure 8. Rattan with no treatment (a), impregnation of lignosulphonate reference (b), Eucalyptus 3× (**c**), and 5× (**d**) washing treatments after flammability testing. 3× (**c**), and 5× (**d**) washing treatments after flammability testing.

Table 9. UL-94 conclusions with no treatment, impregnation of lignosulphonate \mathbf{r} and \mathbf{r}

Lignin produced by 3 \times and 5 \times washing treatments had a yield of 39.28 and 36.24%, a water content of 1.38 and 2.30%, an ash content of 4.20 and 3.43%, a fighth purity of 83.88
and 92.85%, and a total of ph-OH of 6.74 and 5.13%, respectively. The washing treatment water content of 1.58 and 2.50%, an ash content of 4.20 and 3.43%, a lignin purity of 85.88

frequency increased the lignin purity. The TGA and DSC analysis revealed that the lignin had a char residue of 44.14 and 41.66%, with a glass transition temperature (Tg) of 140.86 and 146.72 °C, respectively. The FTIR analysis indicated that lignosulphonate had been successfully synthesized from isolated lignin which could be detected at 627 cm^{-1} for a S=O bond. Meanwhile, lignosulphonate after $3\times$ and $5\times$ washing treatments exhibited a low moisture content of 1.58 and 2.50%, a yield of 93.39 and 91.95%, with a purity of 71.89 and 67.21%. Mass residue by thermal gravimetric of lignosulphonate after $3\times$ and 5× washing treatment of 44 and 42%, and Tg of 128.95 and 173.51 ◦C, respectively. The V-0 rating classification allows lignosulphonate-impregnated rattan to extinguish a fire in less than 6 s without dripping. The incorporation of lignosulphonates into rattan improves the material's fire resistance and meets the needs of the industrial sector.

5. Patents

Indonesian Patent Officer has registered this invention with the patent registration numbers P00202205548 and P00202206965.

Author Contributions: Conceptualization, W.F. and D.S.N.; methodology, W.F. and E.W.M.; software, E.W.M.; validation, W.F. and D.S.N.; formal analysis, E.W.M. and S.H.L.; investigation, W.F.; resources, W.F. and A.H.I.; data curation, E.W.M. and W.F.; writing—original draft preparation, E.W.M., W.F., D.S.N., A.H.I. and S.H.L.; writing—review and editing, W.F., S.H.L. and D.S.N.; visualization, E.W.M.; supervision, W.F. and D.S.N.; project administration, W.F. and A.H.I.; funding acquisition, A.H.I. All authors have read and agreed to the published version of the manuscript.

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