



Article A Novel 3D Tree-Modeling Method of Incorporating Small-Scale Spatial Structure Parameters in a Heterogeneous Forest Environment

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Abstract: Currently, 3D tree modeling in a highly heterogeneous forest environment remains a significant challenge for the modeler. Previous research has only focused on morphological characteristics and parameters, overlooking the impact of micro-environmental factors (e.g., spatial-structural diversification and habitat heterogeneity) and providing less structural information about the individual tree and decreasing the applicability and authenticity of 3D tree models in a virtual forest. In this paper, we chose a mixed-forest conversion of Chinese fir (Cunninghamia lanceolata) plantations in a subtropical region of China as our study subject and proposed a novel 3D tree-modeling method based on a structural unit (TMSU). Our approach modified traditional rule-based tree modeling (RTM) by introducing a nonlinear mixed-effect model (NLME) to study the coupling response between the spatial structures and morphological characteristics (e.g., tree height (H), height-to-crown base (H_{CB}), and crown width (CW)) of three dominant trees (e.g., Cunninghamia lanceolata (SM), Machilus pauhoi (BHN), and Schima superba (MH)) and develop a prediction model of the morphological characteristic by incorporating forest-based structural parameters. The results showed that: (1) The NLME model in TMSU was found to better fit the data and predict the morphological characteristics than the OLS model in RTM. As compared to the RTM morphological model, the prediction accuracy of the TMSU model of morphological features was improved by 10.4%, 3.02%, and 17.8%, for SM's H, H_{CB}, and CW, respectively; 6.5%, 7.6%, and 8.9% for BHN's H, H_{CB}, and CW, respectively; and 13.3%, 15.7%, and 13.4% for MH's H, H_{CB}, and CW, respectively. (2) The spatial-structural parameters of crowding (C_i), mingling (M_i), and dominance (U_i) had a significant impact on the morphological characteristics of SM, BHN, and MH in TMSU. The degree of crowding, for example, had a positive relationship with tree height, height-to-crown base, and crown width in SM, BHN, and MH; under the same crowding conditions, mingling was positively correlated with tree crown width in SM, and dominance was positively correlated with tree height but negatively correlated with height-to-crown base in BHN; under the same crowding and mingling, dominance was positively correlated with height-to-crown base in MH. (3) Using 25 scenes based on the value class of C_i , M_i for SM, 25 scenes based on the value class of C_i , U_i for BHN, and 125 scenes based on the value class of C_i , M_i , U_i for MH, we generated the model libraries for the three dominating species based on TMSU. As a result, our TSMU method outperformed the traditional 3D tree-modeling method RTM in a complex and highly heterogeneous spatial structure of a forest stand, and it provided more information concerning the spatial structure based on the neighborhood relationships than the simple morphological characteristics; a higher morphological prediction accuracy with fewer parameters; and the relationship between the spatial-structural parameters and the morphological characteristics of a reference tree.

Keywords: tree modeling; spatial structure; heterogeneous conversion forest; morphological characteristics model



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1. Introduction

The current 3D tree-modeling methods, such as those based on LIDAR point-cloud data and RGB-D images, have high precision but are costly and computationally intensive [1–3]. Furthermore, it is not appropriate to reconstruct 3D tree models in a dense and highly heterogeneous forest stand because there is insufficient space to obtain sufficient data. Therefore, 3D tree modeling, based on forestry rules, provides a set of generative rules to create plant organs iteratively, though it requires expertise in botany to manually adjust the many parameters in order to model a specific tree [4]. This method has proven to be an effective and popular method for virtual forest reconstruction [5,6]. For example, functional-structural plant models (FSPMs) require specific parameters to model a tree, such as tree height, diameter at breast height, crown width, and height-to-crown base [7]. Therefore, rule-based tree modeling can be easily controlled by modelers without a computer science background [8]. However, the applicability of rule-based modeling is limited because it is based on only a few parameters and overlooks the influence of micro-environmental factors (e.g., spatial-structure diversification and habitat heterogeneity) in a complex and changing forest ecosystem.

Exploring the coupling response between variables related to the spatial structure and morphological characteristics is vital in 3D tree modeling based on a structural unit in a highly heterogeneous forest environment. However, the current research has mostly focused on the coupling responses between unilateral or incomplete characteristics of the spatial structure and tree-crown attributes. For example, researchers suggested that when mixed with other species, the reference tree changed the typical structural attributes of the crown [9-12], such as branching patterns [10,13] and the general shape of the crown [11-14]. These changes could be attributed to the amount of light available, which is the most important resource for which trees compete in a forest canopy [12–15]. Researchers discovered that trees adjusted the shapes and sizes of their crowns in response to competition with their neighbors under different spatial-structure conditions [16], such as in crowded conditions [17,18]. With the rapid application of airborne laser-scanning techniques, more studies are being conducted at the individual tree level and in a more intelligent direction. For example, Pommerening et al. (2021) introduced the CanopyShotNoise model to analyze the response of the canopy pattern under different forest conditions [19]. Fischer et al. (2020) proposed a new simulation-based approach, the canopy constructor, to quantify the forest structure and infer the allometric-scaling relationship in a crowded environment [20].

Morphological models, such as height–diameter, crown width–diameter, and heightto-crown base height–diameter, are critical components of rule-based 3D tree modeling. The prediction accuracy of a model significantly influences the applicability and authenticity of 3D tree models in a virtual forest. Traditional morphological models have adopted the ordinary least squares (OLS) regression for the simple form and its variables. However, the morphological data from many sample plots have hierarchical structures and may be correlated, which has resulted in biased estimations of the variance of the morphological parameters and invalidated the hypothesis when the parameter estimates were analyzed [21,22]. Lately, the popularity and use of mixed-effect models have increased in forestry and morphological modeling practices [23–25]. Researchers have suggested that, as compared to OLS, the nonlinear mixed-effect model (NLME) was more reliable due to its assumption of random and independent observations, as well as the presence of autocorrelations [26–29].

Therefore, in order to model 3D trees in a spatially diverse and heterogeneous forest environment, we chose a mixed forest of Chinese fir (*Cunninghamia lanceolata*) as our subject, which was based on a typical mixed forest stand in a subtropical region of China. The following two aspects were explored in this study: (1) to develop the morphological characteristics (e.g., tree height, height-to-crown base, and crown width) of the model at the individual tree level and with highly heterogeneous spatial structures by introducing a NLME model and (2) to optimize the traditional rule-based 3D tree-modeling method by incorporating individual parameters of spatial structures based on a small-scale forest stand in order to develop a new dominant species' model library under heterogeneous forest conditions.

2. Materials and Methods

2.1. Study Areas

Our study site was located in the ShanXia Forest Farm in Jiangxi Province in Southern China $(27^{\circ}44'45'' \text{ N}, 114^{\circ}39'15'' \text{ E})$ (see Figure 1). The site has a mid-subtropical climate and a mean annual temperature of 19.15 °C, with the highest temperature of 40.3 °C occurring in June and July and the lowest temperature of $-2.1 \,^{\circ}$ C occurring in February. Furthermore, annual precipitation totaled 1722.3 mm, with the majority of the precipitation falling in April, May, and June. The annual photoperiod was 1657.4 h, with the longest photoperiod occurring in July, August, September, and October and the shortest occurring in January and February. The Chinese fir mixed plantations selected in this study were established in 1986 with an initial planting spacing of $1m \times 1m$. Thinning measures were taken when the plantations were 13 years and 20 years old. The first thinning measure was to reduce the intraspecific competition for resource by increasing the growth space and the second thinning measure was to fell the low-quality trees and replant broadleaved trees, such as Machilus pauhoi and Schima superba, which are high-value, fire-resistant trees. During the development of the mixed plantations, some Chinese firs died, allowing companion species or generations to develop in natural succession, such as Vernicia fordii, Liquidambar formosana Hance, Paulowinia fortune, Sapium discolorr, Castanopsis fargesii, etc.

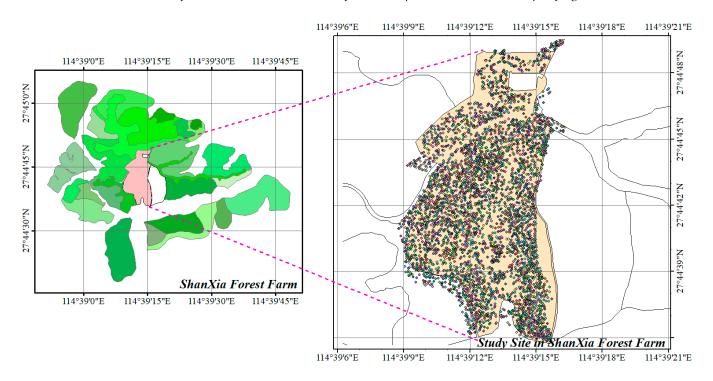


Figure 1. Location of our study site in a subtropical area of China. Notes: the pink area in the left map represents the study region at ShanXia Forest Farm; colorful points in the right map represent different tree species in the study site.

2.2. Data Collection

The data was acquired from 5 ha sample plots of a planted mixed forest dominated by *Cunninghamia lanceolata, Machilus pauhoi*, and *Schima superba* in March and April 2022 (Figure 2). To accurately locate the trees, the plot was divided into 10 subplots of 100 m \times 50 m each (Figure 2). Tree height (H), height-to-crown base (H_{CB}), diameter at breast height (DBH), and crown width (CW) were measured for all trees with a diameter at breast height of (DBH) \geq 5 cm, and their locations were recorded using a total station (TOPCON-

GTS-602AF). Crown width was computed as the arithmetic mean of two crown widths obtained from measurements of four crown radii, representing two azimuths [30]. The first azimuth was defined as the direction from the subject tree to the center of the measurement plot; the second azimuth was perpendicular to the first. In each quadrant, the crown radii were measured as the horizontal distance from the center of the tree bole to the greatest extent of the crown from the bole. The branch tip was located by vertical sighting with a clinometer [31]. Figure 2 depicts a distribution map of the tree species, and each tree population was assigned three layer classes, based on tree height, to select the dominant trees: upper layer trees were $H \ge 20$ m; middle layer trees were $10 \text{ m} \le H < 20$ m; and lower layer trees were $1.3 \text{ m} \le H < 10 \text{ m}$.

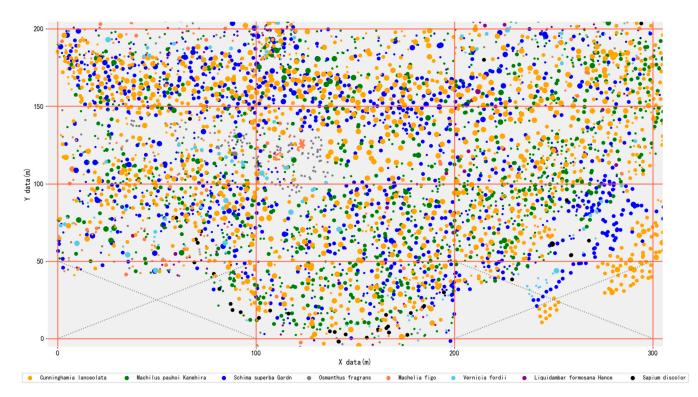


Figure 2. Spatial distribution of eight dominant tree populations at different layers in 5 ha (50 m \times 100 m) planted mixed forest in Shanxia Forest Farm. The upper layer tree was tree height (H) > 20 m; the middle layer tree was 10 m \leq H <20 m; and the lower layer tree was 1.3 m \leq H < 10 m. Different sizes of circles represent the layers in the forest, where the large-size circle is the upper layer and the middle-size circle is the middle layer, and the small-size circle is the lower layer in the mixed forest.

To eliminate edge effects and improve the accuracy of the spatial-structure parameters (SSP), we established a 5 m buffer zone around the plot. In the statistical analysis, only the trees in the reduced window (X, Y) = ($300 \text{ m} \times 200 \text{ m}$, except (0 m–100 m, 0–50), (200 m–300 m, 0 m–50 m)) were used as reference trees, and the individual trees in the buffer zone were only considered to be the nearest neighbors of the trees in the reduced window [32]. This edge correction enabled the individual evaluation of each tree to determine whether all *n*-nearest-neighbors were actually located within the plot.

2.3. Data Analysis

2.3.1. Identification of Dominant Populations for 3D Tree Modeling by the Important Value Index

The importance value (IV) of a species was defined as the average of its relative density (RD), relative frequency (RF), and relative dominance (Rd) and was calculated using the quadratic method [33] for each layer of the planted mixed forest. A total of 32 tree species belonging to 30 genera and 21 families were identified in the planted mixed

forest. The total stand density (DBH \geq 5 cm) was nearly 996.8 trees/ha, with a total basal area of 161.2 m²/ha. Table 1 shows the IVs of the forest's eight dominant tree populations. *Cunninghamia lanceolata, Machilus pauhoi*, and *Schima superba* accounted for 88.24% of the total stand density, and their IVs were clearly superior to those of the other tree species in different stand layers. As a result, they were identified as the dominant populations in the forest for 3D tree modeling (Table 1).

Table 1. IV and characteristics of eight dominant tree populations in the planted mixed forest.

	Density	IV (%)				Stand Characteristics				
Species	(Trees ha ⁻¹)	Upper Layer	Median Layer	Lower Layer	All	DBH (cm)	Height (m)	Basal Area (m ²)	Dominant Height (m)	
<i>Cunninghamia</i> <i>lanceolata</i> (Lamb.) Hook.	273.0	14.57 ± 7.14	15.13 ± 5.19	3.28 ± 1.86	17.18 ± 3.74	24.86 ± 8.91	16.50 ± 5.40	80.83	26.8	
<i>Machilus pauhoi</i> Kanehira	252.6	2.53 ± 1.68	10.38 ± 2.73	7.04 ± 3.13	11.13 ± 3.44	13.82 ± 8.25	10.63 ± 3.83	25.72	23.5	
<i>Schima superba</i> Gardn. et Champ.	259.0	1.21 ± 1.25	12.78 ± 4.21	4.32 ± 1.88	12.39 ± 4.39	15.32 ± 7.02	11.85 ± 3.55	27.67	23.4	
Osmanthus fragrans (Thunb.) Lour.	62.0	0.00 ± 0.00	0.00 ± 0.00	1.82 ± 3.56	1.65 ± 2.94	10.37 ± 3.31	5.74 ± 1.12	2.88	8.2	
Machelia figo (Lour.) Spreng.	48.0	0.07 ± 0.21	0.81 ± 0.90	1.12 ± 1.35	1.59 ± 1.60	13.69 ± 8.03	9.30 ± 3.76	4.36	18.8	
Vernicia fordii (Hemsl.) Airy Shaw	35.6	0.21 ± 0.55	1.34 ± 1.23	0.41 ± 0.74	1.52 ± 1.65	22.60 ± 9.47	14.21 ± 3.40	7.54	21.1	
<i>Liquidambar</i> <i>formosana</i> Hance	27.0	0.03 ± 0.09	0.63 ± 0.90	0.74 ± 0.86	1.24 ± 1.43	16.26 ± 7.63	12.19 ± 3.45	3.01	18.2	
Sapium discolor (Champ. ex Benth.) Muell. Arg.	21.8	0.00 ± 0.00	0.91 ± 1.76	0.13 ± 0.40	0.74 ± 1.21	22.22 ± 7.83	13.33 ± 1.92	4.21	15.4	

Notes: *IVs*, DBH, and Height are mean \pm SD; the upper layer was tree height (H) \geq 20 m; the middle layer was 10 m \leq tree height (H) < 20 m; and the lower layer was 1.3 m tree height (H) < 10 m.

2.3.2. Nonlinear Mixed-Effect Morphological Model of Three Predominant Trees

(1) Base morphological model selection.

In this paper, we chose 16 H~DBH models, 5 HCB~H and DBH models, and 11 CW~DBH models from the literature for appraisal as the base models [27,28,34,35]. These candidate base models were fitted to the 10 plot data (5 ha) using nonlinear least squares regression. The three statistical variables (e.g., coefficient of determination (R²), mean error (E), root-mean-squared error (RMSE)) were used to evaluate the performance of the base models (see Figure 3). The H, H_{CB}, and CW base models evaluated in this study are shown in Table 2.

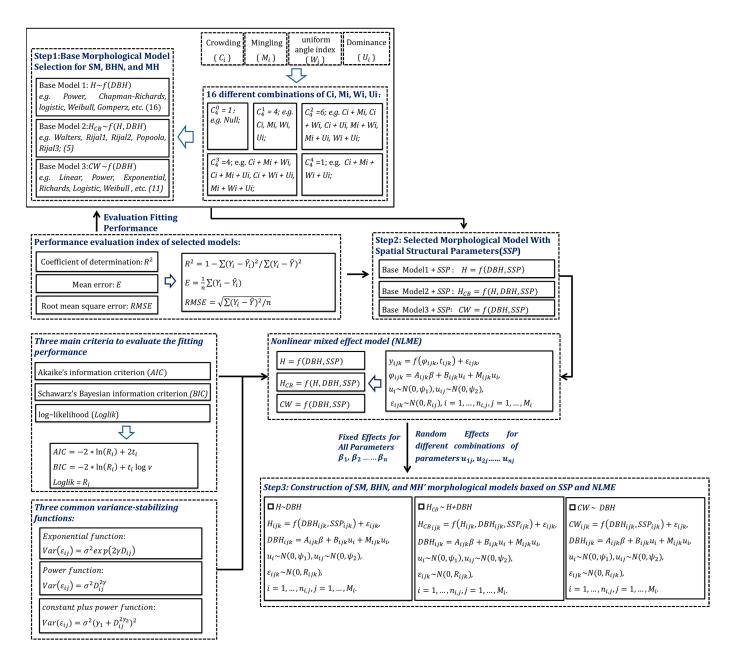


Figure 3. Schematic diagram showing the nonlinear mixed-effect morphological model with spatialstructure parameters (SSP). It includes three steps of morphological construction based on SSP and NLME, evaluation index, and variance-stabilizing functions. The first step is the selection of the base morphological models of SM, BHN, and MH; the second step is the selection of the morphological model with spatial-structure parameters (SSP); and the third step is the construction of the nonlinear mixed-effect of SM, BHN, and MH. SM, BHN, and MH represent *Cunninghamia lanceolata*, *Machilus pauhoi*, and *Schima superba*, respectively. **Table 2.** Base model selection: H is the total tree height (m), DBH is the diameter at breast height (cm), H_{CB} is the height-to-crown base (m), and CW is the crown width (m); a, b, and c are the parameters of the model. M1~M32 represents the formulae of the morphological characteristic models.

Model	Formula	Model Form	Model	Formula	Model Form
M1	$H \sim 1.3 + DBH^2 / (a * DBH + b)^2$	Naslund	M17	$H_{CB} \sim H/(1 + exp(a + b*DBH))$	Walters
M2	H~1.3 + a*DBH/(1 + DBH) ^b	Curtis	M18	$H_{CB} \sim H/(1 + exp(a + b*DBH))^{1/2}$	Rijal 1
M3	H~1.3 + a*exp(-b/DBH)	Schumacher	M19	$H_{CB} \sim H/(1 + exp(a + b*DBH))^{1/6}$	Rijal 2
M4	H~1.3 + a*(1 - exp(-b*DBH))	Meyer	M20	$H_{CB} \sim H/(a + exp(-b*DBH))$	Popoola
M5	H~1.3 + a*DBH ^b	Power	M21	$H_{CB} \sim H/(1 - a * exp(-b * DBH))$	Rijal 3
M6	H~1.3 + a*DBH/(b + DBH)	Michaelis-Menten	M22	CW~a + b*DBH	Linear
M7	$H \sim 1.3 + \exp(a - b/(DBH + 1))$	Wykoff	M23	$CW \sim a + b * DBH + c * DBH^2$	Quadratic
M8	H~1.3 + a*((1 - exp(-b*DBH)))	Chapman–Richards	M24	CW~a*DBH ^b	Power
M9	$H \sim 1.3 + DBH^2 / (a * DBH^2 + b * DBH + c)$	Prodan	M25	CW~a*(1-exp(-b*DBH))	Monomolecular
M10	$H \sim 1.3 + a/(1 + b * exp(-c * DBH))$	logistic	M26	$CW \sim (DBH/(a + b * DBH))^2$	Hossfeld I
M11	$H \sim 1.3 + a*(1-exp(-b*DBH^{c}))$	Weibull	M27	CW~a*(b) ^{DBH}	Compound
M12	H~1.3 + a*exp(-b*exp(-c*DBH))	Gomperz	M28	CW~exp(a + b*DBH)	Growth
M13	$H\sim 1.3 + a*DBH^{(b*DBH-c)}$	Sibbesen	M29	CW~a*exp(b*DBH)	Exponential
M14	$H\sim 1.3 + a * exp(-b * DBH^{-c})$	Korf	M30	CW~a*(1 - exp(-b*DBH))	Richards
M15	$H \sim 1.3 + a \exp(-b/(DBH + c))$	Ratkowsky	M31	$CW \sim a/(1 + b \cdot exp(-c \cdot DBH))$	Logistic
M16	$H \sim 1.3 + a/(1 + 1/(b*DBH^{c}))$	Hossfeld IV	M32	CW~a*(1 - exp(-b*DBH))	Weibull

(2) Additional predictor variables.

A structural unit was defined as a neighborhood consisting of a focal tree and its four nearest neighbors [36]. In a mixed forest, tree size (DBH or crown), tree crowding, tree species, and tree distribution in space were used to describe any structural unit synchronously. These variables were easily expressed by a set of structural stand parameters: W_i , M_i , C_i , and U_i [37,38]. Therefore, there were 16 different combinations of W_i , M_i , C_i , and U_i for selecting the best performing model according to two rules: higher prediction accuracy and lower computational cost (see Figure 3).

$$W_{i} = \frac{1}{n} \sum_{j=1}^{n} Z_{ij}, Z_{ij} = \begin{cases} 1, & if \ \alpha - angle \ is \ smaller \ than \ \alpha_{0} \\ 0, & otherwise \end{cases}$$
(1)

where W_i is defined as the proportion of angles α smaller than the standard angle α_0 (72°) and reflects the degree of uniformity of tree distribution.

$$M_{i} = \frac{1}{n} \sum_{j=1}^{n} V_{ij}, V_{ij} = \begin{cases} 1, & \text{if neighbor } j \text{ is not the same species as reference tree } i \\ 0, & \text{otherwise} \end{cases}$$
(2)

where M_i is defined as the proportion of the four nearest neighbors that are of a different species than reference tree *i*, and it reflects the spatial segregation of different species in a multispecies forest.

$$U_{i} = \frac{1}{n} \sum_{j=1}^{n} K_{ij}, K_{ij} = \begin{cases} 0, & if \text{ neighbor } j \text{ is smaller than reference tree } i \\ 1, & otherwise \end{cases}$$
(3)

where U_i is defined as the degree of DBH differentiation in this paper and represents the relationship between the size of the reference tree *i* and its four nearest neighbors.

$$C_{i} = \frac{1}{n} \sum_{j=1}^{n} P_{ij}, P_{ij} = \begin{cases} 1, & \text{if average crowns of neighbor j and reference tree i} \\ & \text{is smaller than that of reference i} \\ 0, & \text{otherwise} \end{cases}$$
(4)

where C_i reflects the relationship between the canopy of the reference tree *i* and its four nearest neighbors and can reveal the degree of crowding of the reference tree *i*.

(3) Construction of mixed-effect morphological model.

We have incorporated the fixed effects for all parameters in the morphological characteristics models and compared different combinations of all parameters with random effects in the model to identify the parameters that needed to be included, according to the value of the Akaike information criteria (AIC) and Schwarz's Bayesian information criterion (BIC), which were the main criteria for evaluating the fitting performance. In order to solve the problem of heteroscedasticity in the morphological models, we used three typical variance-stabilizing functions (e.g., exponential function, power function, and constant-plus-power function) (4–6) (Figure 3) and selected the best-performing function based on the AIC and the log-likelihood (Loglik) values (see Figure 3).

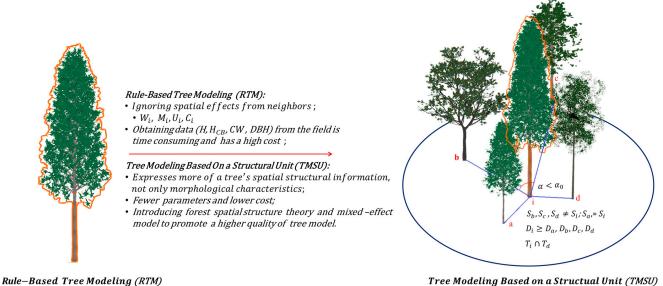
The graphic exploration and correlational analysis were used for evaluation of the contributions of the covariate predictors to the H, H_{CB} , and CW models. All analyses were conducted using R, version 4.1.2. The best-performing model was selected to include additional covariate predictors for formulations of the H, H_{CB} , and CW mixed-effect models.

(4) Assessment and analysis of morphological models

The main differences between rule-based tree modeling (RTM) and 3D tree modeling based on a structural unit (TMSU) were the morphological models' parameter-estimation model and whether the tree modeling considered the spatial structure. In general, the parameter estimation in a traditional morphological model in RTM was based on the ordinary least squares (OLS) model, whereas in TMSU, it was based on a nonlinear mixed-effect model (NLME). In order to validate the feasibility and superiority of the morphological models in TMSU for 3D tree modeling, statistics (R², E, RMSE) and a residual plot of three dominant tree-based morphological models were applied to assess and analyze the morphological models based on OLS and NLME.

2.3.2.1. 3D Tree Modeling Based on a Structural Unit

Rule-based tree modeling (RTM) employs morphological parameters, such as H, H_{CB}, DBH, and CW, to reconstruct a reference tree *i*. However, ignoring the influence of spatial structure on reference tree *i* at a small scale would result in a decrease in the quality of 3D tree modeling and the reality of a virtual forest in forest visualization applications. Our research proposed a new method of 3D tree modeling based on a structural unit (TMSU) that incorporated structural forest theory into 3D tree modeling and achieved 3D tree modeling at the individual tree level with few parameters (see Figure 4). A nonlinear mixed-effect model was introduced in this study to improve the prediction accuracy of the morphological characteristics and parameters (e.g., H, H_{CB}, CW) by avoiding the random effect from the plot and increasing the applicability of the method by reducing the time and computational demand. Based on the predicted morphological data, we reconstructed a 3D tree model with our Creating Tree System (CTS) [39–42], which was developed by the Institute of Forest Resource Information Techniques of the Chinese Academy of Forestry (CAF). The CTS used in this study was based on an improved IFS algorithm [43], and it provided a graphical user interface, 3D visualization tools, and a python scripting interface, as well as the ability to create 3D tree models by selecting the tree species and inputting the H, H_{CB}, DBH, and CW parameters.



DBH= 25.9 cm, H= 21.5 m, HCB= 6.7 m, CW= 5.3 m

Tree Modeling Based on a Structual Unit (TMSU) DBH = 25.9 cm, H = 23.1 m, HCB = 8.1 m, CW = 4.3 m Wi = 0.25, Mi = 0.75, Ui = 0.00, Ci = 0.25

Figure 4. Three-dimensional tree modeling based on RTM and TMSU.

3. Results

3.1. Three-Dimensional Tree Morphological Model Development at the Individual Tree Level

(1) Function selection.

Table 2 displays the fit statistics of the functions. Although the evaluation indices for all the functions were nearly identical, M6 (Chapman – Richard for SM and BHN's H – D), M5 (Power for MH's H – D), M17 (Walters for SM, BHN, and MH's H_{CB} – D), M31(Logistic for SM and BHN's CW – D), and M24 (Power for MH's CW – D) performed slightly better with the fewest parameters. As a result, these functions were chosen as the basic nonlinear model for building the morphological model.

Morphological models of the three predominant trees were the following:

$$SM: H_{ij} = 1.3 + a * (1 - ex p(-b * D_{ij})) + \varepsilon_{ij}$$

$$(5)$$

$$H_{CB_{ij}} = H_{ij}/(1 + exp(a + b * D_{ij})) + \varepsilon_{ij}$$
(6)

$$CW_{ij} = a/(1 + b * exp(-c * D_{ij})) + \varepsilon_{ij}$$

$$\tag{7}$$

BHN: $H_{ij} = 1.3 + a * (1 - exp(-b * D_{ij})) + \varepsilon_{ij}$

$$H_{CB_{ij}} = H_{ij}/(1 + exp(a + b * D_{ij})) + \varepsilon_{ij}$$
⁽⁹⁾

$$CW_{ij} = a/(1 + b * exp(-c * D_{ij})) + \varepsilon_{ij}$$
(10)

MH: $H_{ii} = 1.3 + a * D_{ii}^{b} + \varepsilon_{ii}$

$$H_{CB_{ij}} = H_{ij} / (1 + exp(a + (b * M_i + c * C_i + d * U_i) * D_{ij})) + \varepsilon_{ij}$$
(12)

$$CW_{ij} = a * D_{ij}^{\ b} + \varepsilon_{ij} \tag{13}$$

where H_{ij} , D_{ij} , H_{CBij} , CW_{ij} are the height (m), the diameter at breast height (cm), the height to-crown base (m), and the crown width (m) of tree *i* in the plot *j*; *a*, *b*, and *c* are the formal parameters; and ε_{ij} is an error term.

(2) Incorporating additional predictor variables.

We tested 16 different combinations of W_i , M_i , C_i , and U_i to select the variables that made a significant contribution to the variation of the morphological characteristic (H, H_{CB}, CW) in order to determine the influence of the spatial structure of individual trees. The statistical results showed that C_i , M_i , and U_i were strongly correlated with the morphological variation and, thus, were included in Equations (5)–(13). The following was the final base model used to build the morphological variation mixed-effect model:

(8)

(11)

$$H_{CB_{ij}} = H_{ij}/(1 + exp(a + (b + c * C_i) * D_{ij})) + \varepsilon_{ij}$$

$$\tag{15}$$

$$CW_{ij} = a/(1 + b * exp((-c + d * C_i + e * M_i) * D_{ij})) + \varepsilon_{ij}$$

$$(16)$$

BHN:
$$H_{ij} = 1.3 + a * (1 - exp((-b + c * C_i + d * U_i) * D_{ij})) + \varepsilon_{ij}$$
 (17)

$$H_{CB_{ij}} = H_{ij} / (1 + exp(a + (b * C_i + c * U_i) * D_{ij})) + \varepsilon_{ij}$$
(18)

$$CW_{ij} = a/(1 + b * exp((-c + d * C_i) * D_{ij})) + \varepsilon_{ij}$$
⁽¹⁹⁾

MH:
$$H_{ii} = 1.3 + (a + c * M_i + d * C_i) * D_{ii}{}^b + \varepsilon_{ii}$$
 (20)

$$H_{CB_{ij}} = H_{ij} / (1 + exp(a + (b * M_i + c * C_i + d * U_i) * D_{ij})) + \varepsilon_{ij}$$
⁽²¹⁾

$$CW_{ij} = (a + c * C_i + d * M_i) * D_{ij}^{\ b} + \varepsilon_{ij}$$

$$(22)$$

where H_{ij} , D_{ij} , H_{CBij} , CW_{ij} are the height (m), the diameter at breast height (cm), the heightto-crown base (m), and the crown width (m) of tree *i* in the plot *j*; *a*, *b*, and *c* are the formal parameters; C_i , M_i , and U_i are the degree of crowding, mingling, and dominance of the reference tree *i* base on structural unit; and ε_{ij} is an error term.

(3) Constructing mixed-effect model with parameters for the spatial structure.

After considering the model parameters (*a*, *b*, *c*, *d*, *e*) involved, there were seven different combinations of random effects at the sample plot level for Equations (14), (15) and (18), 15 different combinations for Equations (17), (19) and (20)–(22), and 31 different combinations for Equation (16). When fitted to the data, only model alternatives 5, 6, and 10 converged in Equations (14)–(16), respectively; whereas 12, 6, and 1 converged in Equations (17)–(19), respectively. Model alternatives 5, 9, and 6 converged in Equations (20)–(22), respectively. Among the models that converged, the NLME model with random parameters on a/b/c in Equation (14), *c* in Equation (15), a/b in Equation (16), b/c in Equation (17), b/c in Equation (18), a/b/d in Equation (19), *a* in Equation (20), *a* in Equation (21), and c/d in Equation (22) showed the smallest AIC and BIC. Three dominant trees' morphological models with mixed effects became the following:

SM:
$$H_{ij} = 1.3 + (\beta_1 + u_{1j}) * (1 - exp((-\beta_2 - u_{2j} + (\beta_3 + u_{3j}) * C_i) * D_{ij})) + \varepsilon_{ij}$$
 (23)

$$H_{CB_{ij}} = H_{ij} / (1 + exp(\beta_1 + (\beta_2 + (\beta_3 + u_{1j}) * C_i) * D_{ij})) + \varepsilon_{ij}$$
(24)

$$CW_{ij} = (\beta_1 + u_{1j})/(1 + (\beta_2 + u_{2j}) * exp((-\beta_3 + \beta_4 * C_i + \beta_5 * M_i) * D_{ij})) + \varepsilon_{ij}$$
(25)

BHN:
$$H_{ii} = 1.3 + \beta_1 * (1 - exp((-\beta_2 - u_{1i} + (\beta_3 + u_{2i}) * C_i + \beta_4 * U_i) * D_{ii})) + \varepsilon_{ii}$$
 (26)

$$H_{CB_{ij}} = H_{ij} / (1 + exp(\beta_1 + (\beta_2 + u_{1j}) * C_i + (\beta_3 + u_{2j}) * U_i) * D_{ij})) + \varepsilon_{ij}$$
(27)

$$CW_{ij} = (\beta_1 + u_{1j})/(1 + (\beta_2 + u_{2j}) * exp((-\beta_3 + (\beta_4 + u_{3j}) * C_i) * D_{ij})) + \varepsilon_{ij}$$
(28)

MH:
$$H_{ii} = 1.3 + (\beta_1 + u_{1i} + \beta_3 * M_i + \beta_4 * C_i) * D_{ii}^{\beta_2} + \varepsilon_{ii}$$

$$H_{CB_{ij}} = H_{ij}/(1 + exp(\beta_1 + u_{1j} + (\beta_2 * M_i + \beta_3 * C_i + \beta_4 * U_i) * D_{ij})) + \varepsilon_{ij}$$
(30)

$$CW_{ij} = (\beta_1 + (\beta_3 + u_{1j}) * C_i + (\beta_4 + u_{2j}) * M_i) * D_{ij}^{\beta_2} + \varepsilon_{ij}$$
(31)

where $\beta_1 - \beta_5$ are mixed-effect parameters; u_{1j} , u_{2j} , u_{3j} , and u_{4j} are random-effect parameters generated by the plot on β_1 , β_2 , β_3 , β_4 , respectively; and ε_{ij} is an error term.

(4) Within-plot variance–covariance (R) structure.

Because our data did not show spatial autocorrelations, we assumed the matrix of the within-plot error autocorrelation was an identity matrix. However, we found heteroscedasticity in our preliminary analysis even after the random effects were included in the morphological model with SSP. We then evaluated three variance functions for their heteroscedasticity-reducing performances. The results showed that Equations (24), (27)–(30) and (31) produced significantly different results than the models without variance functions, whereas Equations (23), (25) and (26) did not. According to the AIC and Loglik values, the power variance function and constant power variance function performed the best among the tested variance functions (Table 3). Therefore, the final modes were the following: Equation (23) + null, Equation (24) + power function, Equation (25) + power

(29)

function, Equation (26) + power function, Equation (27) + power function, Equation (28) + constant plus power function, Equation (29) + power function, Equation (30) + power function, and Equation (31) + constant plus power function. By including the variance function, the model's AIC decreased from 3438.982 to 3421.136 in Equation (24), from 1935.082 to 1934.881 in Equation (25), from 3825.354 to 1934.881 in Equation (26), from 3322.544 to 3231.132 in Equation (27), from 3160.599 to 2342.501 in Equation (28), from 4316.121 to 4310.291 in Equation (29), from 3798.104 to 3699.936 in Equation (30), and from 2462.686 to 2416.827 in Equation (31).

Table 3. Performance of mixed-effect model Equations (23)–(31) using morphological data with different variance functions. SM, BHN, and MH represent *Cunninghamia lanceolata, Machilus pauhoi,* and *Schima superba,* respectively.

Species		Evaluation	Residual Variance Function						
	Model	Indices	Null	Exponential Function	Power Function	Constant Plus Power Function			
	E	AIC	4336.224	4337.334	4337.458	4339.477			
	Equation (23)	Loglik	-2158.11	-2157.667	-2157.729	-2157.739			
		p Value		p > 0.05	p > 0.05	p > 0.05			
		AIC	3438.982	3421.818	3421.136	3423.136			
SM	Equation (24)	Loglik	-1712.491	-1702.909	-1702.568	-1702.568			
		p Value		p < 0.0001	p < 0.0001	p < 0.0001			
		AIC	1935.082	1935.461	1934.881	1936.881			
	Equation (25)	Loglik	-960.5411	-959.7304	-959.441	-959.4404			
		p Value		p > 0.05	p > 0.05	p > 0.05			
BHN	Equation (26)	AIC	3825.354	3827.209	3825.14	3827.14			
		Loglik	-1906.677	-1906.604	-1905.57	-1905.57			
		p Value		p > 0.05	p > 0.05	p > 0.05			
	Equation (27)	AIC	3322.544	3249.631	3231.132	3235.042			
		Loglik	-1906.677	-1616.815	-1607.57	-1608.521			
		p Value		p < 0.0001	p < 0.0001	p < 0.0001			
	Equation (28)	AIC	3160.599	2365.384	2449.84	2342.501			
		Loglik	-1574.299	-1175.692	-1217.92	-1163.251			
		p Value	p < 0.0001	p < 0.0001	p < 0.0001	p < 0.0001			
		AIC	4316.121	4313.26	4310.291	4312.291			
	Equation (29)	Loglik	-2150.061	-2147.63	-2146.145	-2146.145			
		p Value		p < 0.05	p < 0.05	p < 0.05			
	E	AIC	3798.104	3701.172	3699.936	3701.532			
MH	Equation (30)	Loglik	-1888.052	-1838.586	-1837.968	-1837.766			
		p Value		p < 0.0001	p < 0.0001	p < 0.0001			
		AIC	2462.686	2418.486	2427.477	2416.827			
	Equation (31)	Loglik	-1223.343	-1200.243	-1204.739	-1198.413			
		p Value		<i>p</i> < 0.0001	<i>p</i> < 0.0001	<i>p</i> < 0.0001			

(5) Estimation of parameters.

All parameter estimates of the OLS and NLME models at the sample plot level with the power variance function or constant plus power variance function were significantly different from zero (p < 0.05), and the parameters are listed in Table 4. When the fixed parameters were substituted into Equations (23)–(31), the morphological model for SM, BHN, and MH, respectively, became the following:

SM:
$$H_{ij} = 1.3 + (36.7543 + u_{1j}) * (1 - exp((-0.0236 - u_{2j} + (-0.0045 + u_{3j}) * C_i) * D_{ij})) + \varepsilon_{ij}$$
 (32)

$$H_{CB_{ij}} = H_{ij}/(1 + exp(0.3896 + (0.0073 + (-0.0096 + u_{1j}) * C_i) * D_{ij})) + \varepsilon_{ij}$$
(33)

$$CW_{ij} = (5.5212 + u_{1j})/(1 + (2.4130 + u_{2j}) * exp((-0.0520 + (-0.0310 * C_i) + 0.0162 * M_i) * D_{ij})) + \varepsilon_{ij}$$
(34)

BHN:
$$H_{ij} = 1.3 + 20.0990 * (1 - \exp((-0.0399 - u_{1j} + (-0.0133 + u_{2j}) * C_i + (-0.0078 * U_i)) * D_{ij})) + \varepsilon_{ij}$$
 (35)

$$H_{CB_{ij}} = H_{ij} / (1 + exp(0.0408 + (-0.0308 + u_{1j}) * C_i + (0.0133 + u_{2j}) * U_i) * D_{ij})) + \varepsilon_{ij}$$
(36)

$$CW_{ij} = (6.7421 + u_{1j})/(1 + (3.2953 + u_{2j}) * exp((-0.0721 + (-0.0455 + u_{3j}) * C_i) * D_{ij})) + \varepsilon_{ij}$$
(37)

MH:
$$H_{ij} = 1.3 + (2.3741 + u_{1j} - 0.2014 * M_i + 0.0622 * C_i) * D_{ij}^{0.5686} + \varepsilon_{ij}$$
 (38)

$$H_{CB_{ij}} = H_{ij} / (1 + exp(0.5479 + u_{1j} + (0.0277 * M_i + 0.0132 * C_i - 0.0026 * U_i) * D_{ij})) + \varepsilon_{ij}$$
(39)

$$CW_{ij} = (0.6514 + (0.2820 + u_{1j}) * C_i + (0.0519 + u_{2j}) * M_i) * D_{ij}^{0.5591} + \varepsilon_{ij}$$

$$\tag{40}$$

Table 4. Parameter estimates and evaluation indices of each model. SM, BHN, and MH represent *Cunninghamia lanceolata, Machilus pauhoi,* and *Schima superba,* respectively. If the letters a, b, and c, are the same, it represents no significant difference between models; otherwise, there are significant differences between models. ** and *** represent significant and extremely significant, respectively.

<u> </u>		Model		Fi	Evaluation Indexes					
Species	Formula		β_1	β_2	β_3	β_4	β_5	AIC	Loglik	p Value
SM	H-D	Equation (5) Equation (14) Equation (23)	33.4665 32.5791 36.7543	0.0266 0.0252 0.0236	-0.0037 -0.0045			4686.538 4350.543 4336.224	-2340.27 -2168.27 -2158.11	a b *** c ***
	H _{CB} -H	Equation (6) Equation (15) Equation (24)	0.4501 0.4230 0.3896	-0.0012 0.0061 0.0073	-0.0087 -0.0096			3488.336 3441.651 3421.136	-1741.17 -1716.83 -1702.57	a b *** c **
	CW-D	Equation (7) Equation (16) Equation (25)	5.9591 5.5826 5.5212	2.4952 2.3971 2.4130	0.0538 0.0461 0.0520	-0.0367 -0.0310	0.0153 0.0162	2224.718 2030.871 1934.881	$-1108.36 \\ -1009.44 \\ -959.441$	a b *** c ***
BHN	H-D	Equation (8) Equation (17) Equation (26)	19.3411 19.6529 20.0990	0.0540 0.0399 0.0399	-0.0123 -0.0133	-0.0072 -0.0078		3970.13 3935.849 3825.14	-1982.07 -1962.93 -1905.57	a b *** c ***
	H _{CB} -H	Equation (9) Equation (18) Equation (27)	0.2933 0.0384 0.0408	$\begin{array}{c} 0.0043 \\ -0.0310 \\ -0.0308 \end{array}$	0.0202 0.0133	 		3404.81 3335.732 3231.132	-1699.41 -1663.87 -1607.57	a b *** c ***
	CW-D	Equation (10) Equation (19) Equation (28)	8.3246 8.4723 6.7421	4.1547 3.7601 3.2953	0.0885 0.0501 0.0721	-0.0386 -0.0455		3218.003 3188.058 2342.501	$-1605 \\ -1589.03 \\ -1163.25$	a b *** c ***
	H-D	Equation (11) Equation (20) Equation (29)	2.5166 2.6616 2.3741	0.5378 0.5135 0.5686	-0.3857 -0.2014	0.3370 0.0622		4577.814 4526.287 4310.291	$-2285.91 \\ -2258.14 \\ -2146.15$	a b *** c ***
MH	H _{CB} -H	Equation (12) Equation (21) Equation (30)	0.8568 0.5636 0.5479	-0.0036 0.0253 0.0277	0.0185 0.0132	-0.0103 -0.0026		3980.823 3919.156 3699.936	-1987.41 -1954.58 -1837.97	a b *** c ***
	CW-D	Equation (13) Equation (22) Equation (31)	0.8478 0.5918 0.6514	0.5819 0.5606 0.5591	0.3178 0.2820	0.1041 0.0519		2749.965 2554.176 2416.827	$-1371.98 \\ -1272.09 \\ -1198.41$	a b *** c ***

In order to validate the effect of the plot level and SSP on the model parameters, we recorded the performance statistics and parameters of the final NLME and OLS models. According to the AIC, Loglik, and *p*-value in Table 4, As shown, Equations (23)–(31) had the smallest AIC and the largest Loglik values, and there was a significant difference between Equations (5) and (14); Equations (14) and (23); Equations (6) and (15); Equations (15) and (24); Equations (7) and (16); Equations (16) and (25); Equations (8) and (17); Equations (17) and (26); Equations (9) and (18); Equations (18) and (27); Equations (10) and (19); Equations (19) and (28); Equations (11) and (20); Equations (20) and (29); Equations (12) and (21); Equations (21) and (30); Equations (13) and (22); and, finally, between Equations (22) and (31), which indicated that the plot and SSP had significant effects on the morphological models of SM, BHN, and MH.

(6) Assessment and analysis of morphological models.

Most models of the morphological characteristics in TMSU differed significantly from the models in RTM, according to Table 5. For example, as compared to the morphological models of Equations (5)–(13) and (23)–(31), which were based on NLME and incorporated variables of the spatial structurers, we were able to significantly increase the prediction accuracy of the morphological characteristics by 10.4%, 3.02%, and 17.8% for SM's H, HCB, and CW, respectively; 6.5%, 7.6%, and 8.9% for BHN's H, HCB, and CW, respectively; and 13.3%, 15.7%, and 13.4% for MH's H, HCB, and CW, respectively.

Table 5. Parameter estimates and evaluation indices of each model. SM, BHN, and MH represent *Cunninghamia lanceolata, Machilus pauhoi*, and *Schima superba*, respectively; u_{1j} , u_{2j} , u_{3j} are the random effect parameters caused by the jth sample plot on β_1 , β_2 , β_3 , respectively.

Species	Formula	Model ⁻	Variance Components							Fit Statistics		
			$\sigma^2_{u_{1j}}$	$\sigma^2_{u_{2j}}$	$\sigma^2_{u_{3j}}$	$\sigma_{u_{1j}}\sigma_{u_{2j}}$	$\sigma_{u_{1j}}\sigma_{u_{3j}}$	$\sigma_{u_{2j}}\sigma_{u_{3j}}$	R ²	E	RMSE	
	H-D	Equation (5) Equation (14) Equation (23)	${4.99 \times 10^0}$	 7.51 × 10 ⁻⁵	2.90×10^{-5}	-9.86×10^{-1}	8.83×10^{-1}	-9.47×10^{-1}	0.7155 0.7369 0.8203	$\begin{array}{r} -0.0201 \\ -0.0199 \\ -0.0190 \end{array}$	2.7644 2.6584 2.1971	
SM -	H _{CB} -H	Equation (6) Equation (15) Equation (24)	3.78×10^{-6}				 		0.6121 0.6313 0.6406	0.0635 0.0739 0.0670	$1.4820 \\ 1.4450 \\ 1.4265$	
	CW-D	Equation (7) Equation (16) Equation (25)	 1.24 × 10 ⁻¹	2.95×10^{-2}		-1.81×10^{-1}	 		0.4276 0.5341 0.5965	$\begin{array}{r} -0.0004 \\ -0.0004 \\ -0.0013 \end{array}$	0.7671 0.6921 0.6441	
	H-D	Equation (8) Equation (17) Equation (26)	3.17×10^{-6}			-8.83×10^{-1}			0.6752 0.6887 0.7376	0.0250 0.0201 0.0205	2.1741 2.1285 1.9542	
BHN	H _{CB} -H	Equation (9) Equation (18) Equation (27)	2.49×10^{-5}	 1.61 × 10 ⁻⁴		-4.30×10^{-1}			$\begin{array}{c} 0.4095 \\ 0.4542 \\ 0.4860 \end{array}$	0.0482 0.0383 0.0467	1.5897 1.5284 1.4832	
-	CW-D	Equation (10) Equation (19) Equation (28)	3.59×10^{0}		3.47×10^{-13}	 1.00×10^{0}	-3.20×10^{-2}	-3.20×10^{-2}	0.4594 0.4782 0.5271	$\begin{array}{c} -0.0012 \\ -0.0015 \\ 0.0032 \end{array}$	1.4319 1.4068 1.3393	
	H-D	Equation (11) Equation (20) Equation (29)	 7.43 × 10 ⁻²						0.5161 0.5426 0.6473	$\begin{array}{c} 0.0008 \\ -0.0021 \\ -0.0029 \end{array}$	2.4594 2.3912 2.0999	
MH -	H _{CB} -H	Equation (12) Equation (21) Equation (30)	3.77×10^{-2}				 		$\begin{array}{c} 0.3018 \\ 0.3468 \\ 0.4282 \end{array}$	$\begin{array}{c} -0.0148 \\ -0.0177 \\ 0.0078 \end{array}$	1.8170 1.7575 1.6443	
	CW-D	Equation (13) Equation (22) Equation (31)	1.10×10^{-2}	2.76×10^{-3}		-9.95×10^{-1}	 		0.5322 0.6180 0.6658	0.0020 0.0054 0.0077	0.9734 0.8796 0.8228	

A box plot of the residuals of the dependent variable classes of three dominant trees was examined to further assess the feasibility of the fitted models based on OLS and NLME (see Figure 5). In each plot, nearly all absolute values of the standard residuals based on OLS and NLME models were less than two. Furthermore, the residuals from the NLME model showed no significant trends, whereas some residual plots from the OLS model did, such as the HCB model of *Cunninghamia lanceolata, Machilus pauhoi*, and *Schima superba*, as well as the CW model of *Machilus pauhoi* and *Schima superba*. Only a few larger deviations were observed for trees that were unusually large or small. The residual from the NLME model exhibited no variance in heteroskedasticity. As a result, the NLME models Equations (32)–(40) had superior statistical performance and could be used for SM, BHN, and MH's 3D tree modeling in TMSU, respectively.

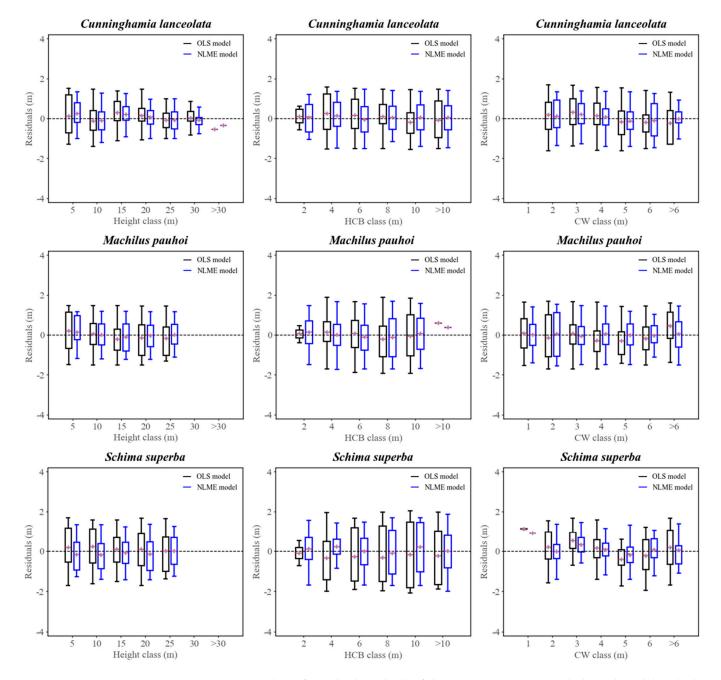


Figure 5. Box plots of standard residuals of three dominant tree morphological models. The box length, whisker length, horizontal lines in red, and plus signs in a box represent the interquartile range (IQR); the class minimum and maximum values in the IQR; and the class median and mean values, respectively. *Cunninghamia lanceolata, Machilus pauhoi,* and *Schima superba* are represented by SM, BHN, and MH, respectively.

3.2. Analysis of 3D Tree Modeling Based on a Structural Unit

We discovered that the reference tree's C_i , M_i , and U_i had a substantial influence on the morphological features of morphological characteristic of SM, BHN, and MH, according to the analytical results of the nonlinear mixed-effect morphological models of three dominant species. For example, (i) crowding and mingling had a significant impact on SM's morphological characteristics: the higher the degree of crowding, the higher the tree height and the height-to-crown base, the smaller the crown width, while under the same crowding, the higher the mingling, the bigger the crown width. (ii) Crowding and dominance had an impact on BHN's morphological characteristics: the bigger the crowding, the higher the tree height and the height-to-crown base, the smaller the crown width, while under the same crowding, the higher the dominance, the bigger the tree height and the lower the height-to-crown base. (iii) Crowding, dominance, and mingling all had an impact on the morphological characteristics of MH: the bigger the crowding, the higher the tree height and the smaller the crown width, while under the same crowding and mingling, the higher the dominance, the higher the height-to-crown base. (see Figure 6).

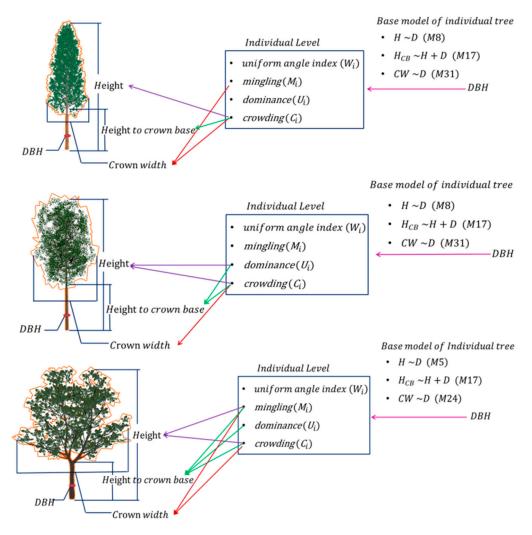


Figure 6. Three-dimensional tree modeling of three predominant trees based on a structural unit.

In this study, we have selected three very different scenes for SM, BHN, and MH as examples to reconstruct 3D tree modeling with a structural unit (see Figure 7). Based on the value classes of C_i , M_i , and U_i , we constructed the SM and BHN model libraries with 25 scenes each (see Appendix A Figures A1 and A2) and an MH model library with 125.

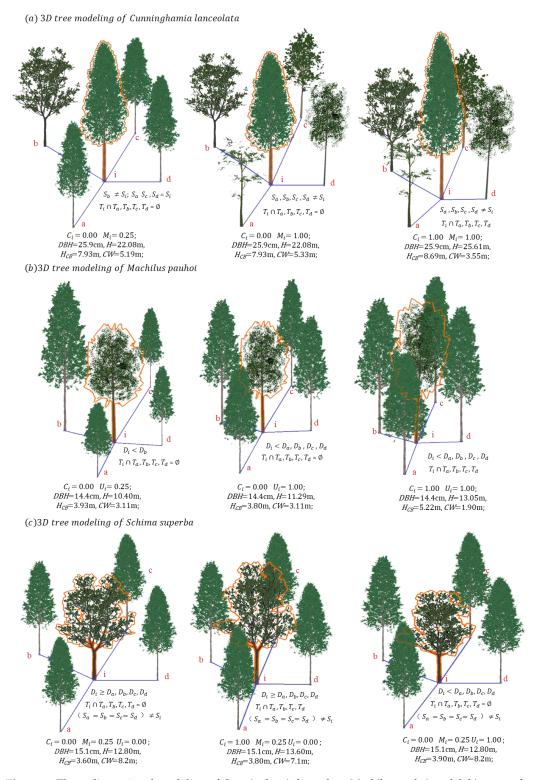


Figure 7. Three-dimensional modeling of *Cunninghamia lanceolata, Machilus pauhoi*, and *Schima superba* under different C_i , U_i , and M_i . The values of C_i , U_i , and M_i were 0.00, 0.25, 0.50, 0.75, and 1.00, respectively. *Cunninghamia lanceolata, Machilus pauhoi*, and *Schima superba* are represented by SM, BHN, and MH, respectively. Since *Cunninghamia lanceolata* (DBH = 25.9 cm), *Machilus pauhoi* (DBH = 14.4 cm), and *Schima superba* (DBH = 15.1 cm) occurred the most frequently in the study region, we chose them as our examples.

4. Discussion

4.1. The Development of Morphological Characteristic Models Involving Forest Spatial Structure in TMSU

A model of morphological characteristics involving forest spatial structure is an essential component of 3D tree modeling based on a structural unit. By acquiring several morphological parameters for 3D tree reconstruction, such as tree height, height-to-crown base, diameter at breast height, and crown width, RTM used fewer micro-characteristics of the individual trees [4–7]. However, forest environments are not homogeneous even at very small scales. The spatial structures must be incorporated into the morphological characteristics in order to accurately model 3D trees in an environment with a high degree of structural diversity and heterogeneity [12–14]. In our study, TMSU used four spatialstructure variables (e.g., W_i , M_i , C_i , and U_i) to describe the spatial structures and their effects on morphological traits. According to Table 4, we discovered that these spatial structures could increase the precision with which morphological characteristics were predicted due to the spatial heterogeneity of a reference tree. For example, as compared to the RTM model Equations (5)–(13), the TMSU model Equations (14)–(22) incorporated variables of the spatial structures and then improved its prediction accuracy of the morphological characteristics by 2.1%, 1.9%, and 10.7% for SM's H, H_{CB} , and CW, respectively; 1.4%, 4.5%, and 1.9% for BHN's H, H_{CB}, and CW, respectively; and finally, 2.7%, 4.5%, and 8.9% for MH's H, H_{CB}, and CW, respectively. However, when developing morphological characteristic models, researchers have suggested that adding excessive parameters affected model convergence and lengthened the time to compute parameter estimates, especially for the nonlinear mixed-effect modeling strategy used in this study [27,28,44]. Furthermore, adding a number of tree or spatial-structure variables to the inventory increased the cost and time required to obtain the measurements for those variables [29,44]. Therefore, for the final morphological prediction model of SM in TMSU, we selected one tree variable (D) and two spatial-structure variables (C_i , M_i); one tree variable (D) and two spatial-structure variables (C_i, U_i) for the final morphological prediction model of BHN; and one tree variable (D) and three spatial-structure variables (C_i , U_i , M_i) for the final morphological prediction model of MH.

Nonlinear mixed-effect models are appealing for the analysis of hierarchically structured data that are auto-correlated, as compared to the OLS model because of their flexibility in accounting for the covariate structure that is overlooked in conventional regression procedures. Therefore, in this study, we selected the Chapman-Richards, Walters, logistic, and power models as the base morphological models based on the statistical variables R^2 , E, and RMSE, and then we integrated them with a nonlinear mixed-effect model approach at the plot level to construct morphological models of three dominant trees involving spatialstructure parameters. As shown in Table 5, a random effect model in TMSU Equations (23)–(31) outperformed a model without a random effect in RTM Equations (14)–(22) in terms of prediction accuracy for the full dataset. The result was consistent with most research conclusions [24–28]. As a result, morphological models of three dominant trees in TMSU were chosen for 3D tree modeling due to their higher precision and spatial capabilities. In addition, the morphological models in TMSU were limited in their capacity to handle changes in spatial structures (e.g., those caused by human activity or natural succession) [45] and complete data acquisition that could reflect different spatial-structure conditions. Generally, as the spatial structure of the planted mixed forest would not be significantly altered in a short time, the models in TMSU could be utilized for producing a short-term forecast (less than 5 years). Therefore, in order to improve the applicability of the morphological model in TMSU, we will consider climate variables, in addition to natural succession and human activities, and devise more plots with different spatial-structure conditions in our next project [46,47].

4.2. Three-Dimensional Tree Modeling in a Highly Heterogeneous Forest Environment

In forest visualization research, accurate 3D tree modeling with heterogeneous spatial forest structures is required (e.g., forest quality assessment, health supervision, and forest management planning and assessment) [48]. Given the complexity of forest structures and heterogeneous habitats, describing the relationship between any individual tree and its neighbors in a 3D tree model is a significant issue. In this study, we addressed the challenges of 3D tree modeling in a complex and highly heterogeneous forest environment in order to determine the spatial-structure variables, explore the relationships between spatial structures and morphological characteristics, and achieve 3D tree modeling based on a structural unit.

Most researchers have developed various qualitative and quantitative indices to describe the reference tree and its neighbor trees in a forest stand, such as the index of Clark and Evans [49], the diffusion index [50], Ripley's K-function [51], the K-function [52], the Oring statistic [53], the Gini coefficient [54], Pielou's isolation index [55], the mean directional index [56], Gadow's species mingling [57], the mixed ratio [58], and the uniform angle index [59,60]. However, some of these indices have been predominantly used in forestry and ecology, and they do not meet the prerequisites for visually and intuitively portraying 3D tree modeling based on a structural unit [19,20]. In addition, certain approaches for analyzing spatial structures consider the neighboring tree species to some extent, although they can only describe the overall or unilateral structure of the reference tree population. For example, the size differentiation, mixed conditions, and the degree of crowding of the reference tree at different distance scales remained unknown when using W_i to analyze the distribution patterns. Therefore, these methods solely considered the distribution pattern and overlooked factors such as the tree size, the degree of crowding, and the degree of species aggregation. Furthermore, an entirely different structure could yield the same outcome. Obviously, it has been challenging to accurately depict the spatial structure of a tree population. The forest is a complex and dynamic ecosystem, particularly when it contains many different species and has a highly heterogeneous spatial structure. The four most significant characteristics of the population structure for any species in a mixed forest may be interspecific and intraspecific variances in tree sizes, species mingling, crowding, and distribution patterns [60]. The spatial structural parameters W_i , M_i , C_i , and U_i were used in TMSU to describe the relationships between any reference tree and its four adjacent neighboring trees. It uncovered reference tree population characteristics and modeled a 3D tree in a more accurate and thorough manner by simultaneously employing two or more pieces of structural information.

According to TMSU-based morphological models, we found that C_i , M_i , and U_i had a strong influence on the morphological characteristic of SM, BHN, and MH. For example, in SM, BHN, and MH, there was an identical phenomenon where the degree of crowding had a positive relationship with tree height, height-to-crown base, and crown width, which could be related to a growth response strategy of trees in a crowded and highly competitive environment to increase the tree's height to occupy the main layer of the forest and obtain more resources, such as light, moisture, and nutrition [17,60]. This could also promote the self-thinning of the branches, which increases the height-to-crown base for the limited availability of the light resource [10], as well as retaining a narrow crown width due to competition with conspecific and nonspecific species [16]. Furthermore, under the same degree of crowding, mingling was positively correlated with tree crown width in SM, and dominance was positively correlated with tree height but negatively correlated with the height-to-crown base in BHN; under the same crowding and mingling conditions, the dominance was positively correlated with the height-to-crown base in MH. The results could be linked to the competitive response mechanism of various tree species, such as light transmittance, shade tolerance, and biological characteristics. Therefore, considering the unique coupling responses between the spatial-structure parameters and the morphological characteristics, we created model libraries for the three dominant species with 25 scenes based on the value classes of C_i and M_i for SM; 25 scenes based on the value classes of C_i and U_i for BHN; and 125 scenes based on the value classes of C_i , M_i , and U_i for MH, respectively.

5. Conclusions

This study provided a new approach for 3D tree modeling TMSU. The differences between RTM and TMSU were focused on: (i) Four spatial structural parameters (W_i , M_i , C_i , and U_i) were incorporated into the morphological characteristic models of the reference trees and revealed the relationship between the spatial-structure variables and the morphological features. (ii) The nonlinear mixed-effect model was introduced into the parameter estimation of the TMSU-based morphological model for higher precision and applicability. By comparing the two approaches, RTM and TMSU, we found that (i) the prediction accuracy of the TMSU-based morphological model increased by 10.4%, 3.02%, and 17.8% for SM's H, HCB, and CW, respectively; 6.5%, 7.6%, and 8.9% for BHN's H, HCB, and CW, respectively; as compared to the RTM morphological models. (ii) TMSU provided more spatial-structure information with fewer parameters in the morphological models and revealed the relationships between the spatial structures and the morphological traits of the reference tree.

Author Contributions: L.W. and H.Z. (Huaiqing Zhang) proposed the novel 3D tree modeling method by incorporating spatial structural parameters under highly heterogeneous forest environment. L.W. and H.Z. (Huacong Zhang), T.Y., J.Z. and Y.L. collected and processed the morphological and spatial structural data of study site. L.W. and H.Z. (Huaiqing Zhang) contributed to writing the paper. All authors have read and agreed to the published version of the manuscript.

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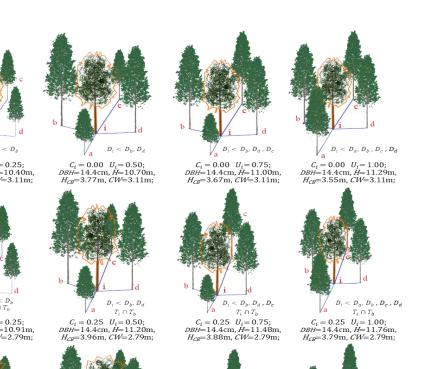
Data Availability Statement: Not applicable.

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Conflicts of Interest: The authors declare that there are no conflict of interest.

Appendix A d d d d $S_a, S_b, S_c, S_d \neq S_i$ $S_b, S_c, S_d = S_i$ $S_b, S_c, S_d \neq S_i; S_a$ = S $S_b, S_d \neq S_i; S_a, S_c = S_i$ $S_b \neq S_i; S_a S_c, S_d = S_i$ $C_i = 0.00 \quad M_i = 0.00;$ DBH=25.9cm, H=22.08m, $C_i = 0.00 M_i = 0.75;$ DBH=25.9cm, H=22.08m, $C_i = 0.00 \ M_i = 0.50;$ DBH=25.9cm, H=22.08m, $C_i = 0.00 M_i = 1.00;$ DBH=25.9cm, H=22.08m, $C_i = 0.00 \ M_i = 0.25;$ DBH=25.9cm, H=22.08m, *H_{CB}*=7.93m, *CW*=5.14m; H_{CB} =7.93m, CW=5.36m; *H_{CB}*=7.93m, *CW*=5.29m; H_{CB}=7.93m, CW=5.44m; H_{CB}=7.93m, CW=5.22m; h d d d d S_b, S_c, S_d $\neq S_i; S_a$ = S S_c , $S_d = S_i$ $S_h, S_c, S_d \neq S_i; S_{a} = S_i$ $S_h, S_d \neq S_i$; $S_a, S_c = S_i$ $S_a, S_b, S_c, S_d \neq S_i$ $T_i \cap T_b$ Va a Va $C_i = 0.25$ $M_i = 0.25$; DBH = 25.9 cm, H = 23.10 m, $H_{CB} = 8.18$ m, CW = 4.53 m; $C_i = 0.25 \quad M_i = 0.50;$ DBH=25.9 cm, H=23.10 m, $H_{CB}=8.18 \text{ m}, CW=4.62 \text{ m};$ $C_i = 0.25 \quad M_i = 0.75;$ DBH = 25.9 cm, H = 23.10 m, $H_{CB} = 8.18 \text{ m}, CW = 4.70 \text{ m};$ $C_i = 0.25 \quad M_i = 1.00;$ DBH=25.9 cm, H=23.10 m, $H_{CB}=8.18 \text{ m}, CW=4.77 \text{ m};$ $C_i = 0.25 \quad M_i = 0.00;$ DBH=25.9cm, H=23.10m, H_{CB}=8.18m, CW=4.44m; d d d d $\overset{\mathbf{d}}{=} S_i; \ S_a \ S_c, S_d = S_i$ S_b $S_a, S_b, S_c, S_d \neq S_i$ $S_a, S_b, S_c, S_d = S_i$ $S_b, S_d \neq S_i$; $S_a, S_c = S_i$ $T_i \cap T_b, T_d$ $S_b, S_c, S_d \neq S_i; S_a,=S_i$ $T_i \cap T_b$, T_d $T_i \cap T_b, T_d$ $T_i \cap T_b, T_d$ $T_i \cap T_b$, T_d ľa $C_i = 0.50 \quad M_i = 0.25;$ DBH = 25.9 cm, H = 24.14 m, $H_{CB} = 8.43 \text{ m}, CW = 4.05 \text{m};$ $C_i = 0.50 M_i = 0.50;$ DBH=25.9 cm, H=24.14 m, $H_{CB}=8.43 \text{ m}, CW=4.11 \text{ m};$ $C_i = 0.50 \quad M_i = 1.00;$ DBH=25.9cm, H=24.14m, $H_{CB}=8.43$ m, CW=4.22m; $C_i = 0.50 \quad M_i = 0.00;$ $C_i = 0.50 \quad M_i = 0.75$ $DBH=25.9 \text{ cm}, H=24.14 \text{ m}, H_{CB}=8.43 \text{ m}, CW=4.16 \text{ m};$ DBH=25.9cm, H=24.14m, H_{CB}=8.43m, CW=4.01m; d d 1 d d $\neq S_i; S_a S_c, S_d = S_i$ S_a , S_b , S_c , $S_d = S_i$ S_b $S_a, S_b, S_c, S_d \neq S_i$ $S_b, S_d \neq S_i; S_a, S_c = S_i$ $S_h, S_c, S_d \neq S_i; S_a, = S_i$ $V_{\mathbf{a}}^{T_i} \cap T_b, T_c, T_d$ $T_i \cap T_b, T_c, T_d$ $\bigvee_{a} T_{i} \cap T_{b}, T_{c}, T_{d}$ $T_i \cap T_b, T_c, T_d$ $T_i \cap T_b, T_c, T_d$ a $C_i = 0.75 M_i = 0.50;$ DBH=25.9cm, H=24.87m, $C_i = 0.75 \quad M_i = 1.00;$ DBH=25.9cm, H=24.87m, $C_i = 0.75 M_i = 0.00;$ $C_i = 0.75 \quad M_i = 0.25;$ DBH=25.9cm, H=24.87m, $C_i = 0.75$ $M_i = 0.75$; DBH=25.9cm, H=24.87m, DBH=25.9cm, H=24.87m, H_{CB} =8.56m, CW=4.00m; H_{CB}=8.56m, CW=3.71m; H_{CB}=8.56m, CW=3.79m; H_{CB}=8.56m, CW=3.86m; H_{CB}=8.56m, CW=3.93m; li i $\int_{S_a, S_b, S_c, S_d}^{1} \frac{d}{\neq S_i}$ d d d $S_a, S_b, S_c, S_d = S_i$ $S_a S_c, S_d = S_i$ $S_b, S_d \neq S_i; S_a, S_c = S_i$ $S_b, S_c, S_d \neq S_i; S_a = S_i$ S; **a** $T_i \cap T_a, T_b, T_c, T_d$ a $T_i \cap T_a, T_b, T_c, T_d$ $C_i = 1.00 \quad M_i = 0.50;$ DBH=25.9 cm, H=25.61 m, $H_{CB}=8.69 \text{ m}, CW=3.55 \text{ m};$ $C_i = 1.00 M_i = 0.75;$ DBH=25.9 cm, H=25.61 m, $H_{CB}=8.69 \text{ m}, CW=3.63 \text{ m};$ $C_i = 1.00 \quad M_i = 1.00;$ DBH=25.9 cm, H=25.61 m, $H_{CB}=8.69 \text{ m}, CW=3.70 \text{ m};$ $C_i = 1.0 \quad M_i = 0.00;$ $C_i = 1.00 \quad M_i = 0.25;$ DBH=25.9 cm, H=25.61 m, $H_{CB}=8.69 \text{ m}, CW=3.48 \text{ m};$ DBH=25.9cm, H=25.61m, H_{CB} =8.69m, CW=3.40m;

Figure A1. Three-dimensional modeling of SM under different C_i and M_i . The values of C_i and M_i were 0.00, 0.25, 0.50, 0.75, and 1.00. Since *Cunninghamia lanceolata* (SM) (DBH = 25.9 cm) occurred most frequently in the study region, we chose it as an example.



 $D_i < D_a, D_b, D_c, D_d$ $T_i \cap T_b, T_d$

d



 $\begin{array}{c} D_i < D_a, D_b, D_c, D_d \\ \mathbf{a} \quad T_i \cap T_b, T_c, T_d \end{array}$ Va

 $C_t = 0.75$ $U_t = 1.00;$ DBH=14.4 cm, H=12.64 m, $H_{CB}=4.71$ m, CW=2.17 m;

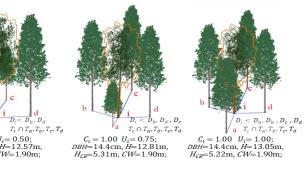


Figure A2. Three-dimensional modeling of BHN under different C_i and U_i . Notes: $D_i < C_i$ D_a , D_b , D_c , D_d indicated that the diameter of tree *i* was smaller than tree *a*, *b*, *c*, and *d*; $T_i \cap T_a$, T_b , T_c , T_d indicated that the canopy of tree *i* and tree *a*, *b*, *c*, and *d* overlapped; C_i and U_i had values of 0.00, 0.25, 0.50, 0.75, and 1.00. Since Machilus pauhoi (BHN) (DBH = 14.4 cm) occurred most frequently in the study region, we chose it as an example.



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