

Supplementary Table 1: Primers used in this study

Primer name	Sequence (5' – 3')	Function
<i>SmpilA</i> upF	GTAT <u>GTCGAC</u> GCCAATCGCCCCTATGCTGG	Anneals 1,047 to 1,028 bp upstream to the D1585 <i>pilA</i> start codon, <i>Sall</i> site underlined
<i>SmpilA</i> upR-OE	ATGGGATTAGCAGCCAGAGCACAGGATCGG TCTGG	The 20 nucleotides at the 3' end anneal to the seven codons following the D1585 <i>pilA</i> start codon and the first 15 nucleotides in bold overlap with the 5' end of the downstream fragment
<i>SmpilA</i> downF-OE	CCGATCCTGTGCTCTGGCTGCTAATCCCATC TGGA	The first 15 nucleotides in bold overlap with the 3' end of the upstream fragment, the last 20 nucleotides at the 3' end anneal to the last three codons in D1585 <i>pilA</i> and 11 bp following the stop codon
<i>SmpilA</i> downR	CCTC <u>AAGCTT</u> CCCCAACCACTTGTCTGC	Anneals 902 to 921 bp downstream to the D1585 <i>pilA</i> stop codon, <i>HindIII</i> site underlined
280 <i>pilA</i> upR	GCCCA <u>AAGCTT</u> CATGTTACGATCATCTGGG	Anneals 1,046 to 1,027 bp upstream to the 280 <i>pilA</i> start codon on the reverse strand. <i>HindIII</i> site underlined.
280 <i>pilA</i> upF-OE	GGCGTACTTCTTCAGCATTTTGGTACATCCC CAAG	The 20 nucleotides at the 3' end anneal to the 280 <i>pilA</i> start codon and 17 bp upstream, and the first 15 nucleotides in bold overlap with the 5' end of the downstream fragment.
280 <i>pilA</i> downR-OE	GGATGTACCAAAATGCTGAAGAAGTACGCC CCGAC	The first 15 nucleotides in bold overlap with the 3' end of the upstream fragment, and the last 20 nucleotides at the 3' end anneal to seven out of 12 codons upstream of the 280 <i>pilA</i> stop codon
280 <i>pilA</i> downF	GGCAG <u>TGACG</u> GAACTTGATCTCGTCCAGC	Anneals 1,082 to 1,063 bp downstream of the 280 <i>pilA</i> stop codon. <i>Sall</i> site underlined.
<i>PapilA</i> F	GCGT <u>GTCGAC</u> CCAGTTTCCTTGATCGTGCC	Anneals upstream of PA01 <i>pilA</i> gene. <i>Sall</i> site underlined.
<i>PapilA</i> R	GCCGA <u>AAGCTT</u> GAGGAACCCAATCACAACGG	Anneals downstream of PA01 <i>pilA</i> gene. <i>HindIII</i> site underlined.
<i>PapilE</i> F	CCGAGGATCCGATCGAGAAAGAACAGCCCC	Anneals upstream of the PA01 <i>pilE</i> gene. <i>BamHI</i> site underlined.
<i>PapilE</i> R	GCGGA <u>AAGCTT</u> GCGGGAGGAGAACATTACCT	Anneals downstream of the PA01 <i>pilE</i> gene. <i>HindIII</i> site underlined.
<i>SmpilA</i> F	CCA <u>AAGCTT</u> GACCCATCCGTGAAATAGCTGCC	Anneals upstream of D1585 <i>pilA</i> start codon. <i>Sall</i> site underlined.
<i>SmpilA</i> R	CGCC <u>AAGCTT</u> ACGAGCCGACAAAAGAAAGGC	Anneals downstream of D1585 <i>pilA</i> stop codon. <i>HindIII</i> site underlined.
<i>SmpilE</i> F	GTCTG <u>TGAC</u> CAGTAACCCAGTGCGAGGA	Anneals upstream of the D1585 <i>pilE</i> gene. <i>Sall</i> site underlined.
<i>SmpilE</i> R	GCCCA <u>AAGCTT</u> CTAACCGGCTGAGCTATTCG	Anneals downstream of the D1585 <i>pilE</i> gene. <i>HindIII</i> site underlined.
280 <i>pilA</i> F	GCAAG <u>TGAC</u> CAGACCGATCCTGTGCTCTG	Anneals upstream of the 280 <i>pilA</i> gene. <i>Sall</i> site underlined.
280 <i>pilA</i> R	GACCA <u>AAGCTT</u> CCCCTAGTTCGCTTCATGGC	Anneals downstream of the 280 <i>pilA</i> gene. <i>HindIII</i> site underlined.

Supplementary Table 2: Characteristics of *P. aeruginosa* PA01 transposon mutants

Mutant ID/ Strain Name	Gene affected	Transposon	Genome Insertion Position	DLP1 lysis	Source
PW8621	<i>pilA</i>	<i>lacZ-hah</i>	5069310	-	[21]
PW8622	<i>pilA</i>	<i>phoA-hah</i>	5069368	-	[21]
PA01_lux_18_G2	<i>pilB</i>	<i>mini-Tn5-luxCDABE</i>	5069913	-	[20]
PA01_lux_50_H10	<i>pilB</i>	<i>mini-Tn5-luxCDABE</i>	5071244	-	[20]
PA01_lux_67_D1	<i>pilB</i>	<i>mini-Tn5-luxCDABE</i>	5070860	-	[20]
PA01_lux_97_B10	<i>pilB</i>	<i>mini-Tn5-luxCDABE</i>	5070077	-	[20]
PA01_lux_38_F5	<i>pilE</i>	<i>mini-Tn5-luxCDABE</i>	5104853	-	[20]
PA01_lux_41_C7	<i>pilE</i>	<i>mini-Tn5-luxCDABE</i>	5104839	-	[20]
PA01_lux_50_D5	<i>pilE</i>	<i>mini-Tn5-luxCDABE</i>	5104869	-	[20]
PW7438	<i>pilF</i>	<i>phoA-hah</i>	4264656	-	[21]
PA01_lux_44_E9	<i>pilJ</i>	<i>mini-Tn5-luxCDABE</i>	452821	-	[20]
PW9471	<i>pilN</i>	<i>phoA-hah</i>	5679378	-	[21]
PW9465	<i>pilQ</i>	<i>phoA-hah</i>	5676840	-	[21]
PW9466	<i>pilQ</i>	<i>phoA-hah</i>	5676900	-	[21]
PA01_lux_80_E5	<i>pilR</i>	<i>mini-Tn5-luxCDABE</i>	5096068	-	[20]
PA01_lux_18_G4	<i>pilS</i>	<i>mini-Tn5-luxCDABE</i>	5094527	-	[20]
PA01_lux_42_D11	<i>pilS</i>	<i>mini-Tn5-luxCDABE</i>	5094527	-	[20]
PA01_lux_53_B6	<i>pilS</i>	<i>mini-Tn5-luxCDABE</i>	5094527	-	[20]
PA01_lux_80_C7	<i>pilS</i>	<i>mini-Tn5-luxCDABE</i>	5094527	-	[20]
PA01_lux_32_G12	<i>pilT</i>	<i>mini-Tn5-luxCDABE</i>	436863	-	[20]
PA01_lux_46_D4	<i>pilT</i>	<i>mini-Tn5-luxCDABE</i>	436504	-	[20]
PW1730	<i>pilU</i>	<i>lacZ-hah</i>	438793	+	[21]
PA01_lux_19_D2	<i>pilV</i>	<i>mini-Tn5-luxCDABE</i>	5098940	-	[20]
PA01_lux_73_C10	<i>pilV</i>	<i>mini-Tn5-luxCDABE</i>	5099241	-	[20]
PA01_lux_20_D4	<i>pilYI</i>	<i>mini-Tn5-luxCDABE</i>	5101178	-	[20]
PA01_lux_51_H7	<i>pilYI</i>	<i>mini-Tn5-luxCDABE</i>	5102516	-	[20]
PA01_lux_82_C12	<i>pilYI</i>	<i>mini-Tn5-luxCDABE</i>	5102516	-	[20]
PA01_lux_97_G2	<i>pilYI</i>	<i>mini-Tn5-luxCDABE</i>	5100755	-	[20]
PA01_lux_67_E3	<i>fimV</i>	<i>mini-Tn5-luxCDABE</i>	3496099	+/-	[20]
PA01_lux_20_D1	<i>fimV</i>	<i>mini-Tn5-luxCDABE</i>	3497859	+/-	[20]
PA01_lux_21_F1	<i>fimV</i>	<i>mini-Tn5-luxCDABE</i>	3496114	+/-	[20]
PA01_lux_97_D2	<i>algR</i>	<i>mini-Tn5-luxCDABE</i>	5923165	+/-	[20]
PA01_lux_50_H9	<i>PA2806</i>	<i>mini-Tn5-luxCDABE</i>	3160973	-	[20]

More information on strains is available at <http://pseudomutant.pseudomonas.com> for *mini-Tn5-luxCDABE* mutants and <http://www.gs.washington.edu/labs/manoil/libraryindex.htm> for *lacZ-hah* and *phoA-hah* mutants.

Supplementary Table 3: Comparison of the presence of Phage-tail_3 domain and tail fiber domains of experimentally confirmed pili-receptor phages.

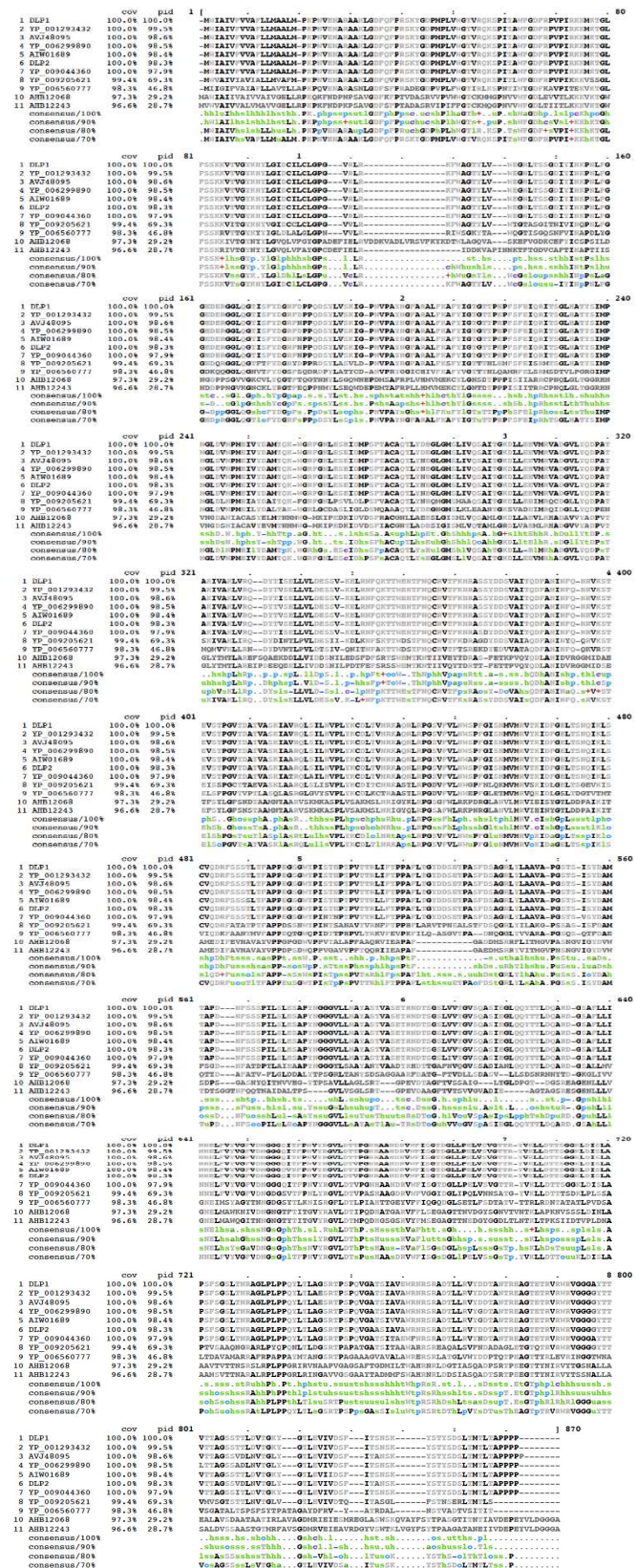
Bacteriophage	Accession	Morphology	Phage-tail_3 domain	Tail fiber domains
<i>Stenotrophomonas</i> phage DLP1	KR537872.1	<i>Siphoviridae</i>	Yes	-
<i>Stenotrophomonas</i> phage DLP2	KR537871.1	<i>Siphoviridae</i>	Yes	-
<i>Stenotrophomonas</i> phage DLP4	MG018224.1	<i>Siphoviridae</i>	Yes	Yes (1x)
<i>Pseudomonas</i> phage D3112	NC_005178	<i>Siphoviridae</i>	Yes	-
<i>Pseudomonas</i> phage MP22	NC_009818	<i>Siphoviridae</i>	Yes	-
<i>Pseudomonas</i> phage B3	NC_006548	<i>Siphoviridae</i>	Yes	-
<i>Pseudomonas</i> phage JBD26	JN811560	<i>Siphoviridae</i>	Yes	-
<i>Pseudomonas</i> phage JBD68	KY707339	<i>Siphoviridae</i>	-	-
<i>Pseudomonas</i> phage MP29	EU272036.1	<i>Siphoviridae</i>	Yes	-
<i>Pseudomonas</i> phage MP42	JQ762257.1	<i>Siphoviridae</i>	Yes	-
<i>Pseudomonas</i> phage JBD69	KU199708.1	<i>Siphoviridae</i>	Yes	-
<i>Pseudomonas</i> phage JBD93	NC_030918.1	<i>Siphoviridae</i>	Yes	-
<i>Pseudomonas</i> phage JBD88a	NC_020200.1	<i>Siphoviridae</i>	-	-
<i>Pseudomonas</i> phage JBD5	NC_020202.1	<i>Siphoviridae</i>	Yes	-
<i>Pseudomonas</i> phage JBD30	NC_020198.1	<i>Siphoviridae</i>	Yes	-
<i>Xylella</i> phage Paz	KF626666	<i>Siphoviridae</i>	-	Yes (1x) ^a
<i>Xylella</i> phage Prado	KF626667	<i>Siphoviridae</i>	-	Yes (1x) ^a
<i>Xylella</i> phage Salvo	KF626668	<i>Siphoviridae</i>	Yes	- ^b
<i>Xylella</i> phage Sano	KF626665	<i>Siphoviridae</i>	Yes	- ^b
<i>Caulobacter</i> phage phiCbK	JX100813	<i>Siphoviridae</i>	Yes	-
<i>Pseudomonas</i> phage phiKMV	AJ505558	<i>Podoviridae</i>	-	Yes (1x) ^c
<i>Pseudomonas</i> phage MPK7	JX501340	<i>Podoviridae</i>	-	Yes (2x)
<i>Pseudomonas</i> phage LUZ19	NC_010326	<i>Podoviridae</i>	-	Yes (3x)
<i>Pseudomonas</i> phage F116	NC_006552	<i>Podoviridae</i>	-	-
<i>Pseudomonas</i> phage Pf-10	NC_027292	<i>Podoviridae</i>	-	Yes (1x)
<i>Pseudomonas</i> phage JBD23	KM389462.1	N/A ^d	Yes	-

a. Three tail fiber genes annotated, but only one tail fiber hit on CD-search; therefore, only counted as 1x tail fiber domains.

b. One tail fiber gene annotated, but no tail fiber hits on CD-search, therefore not counting as tail fiber domains.

c. Two tail fiber proteins annotated, but only one hit on CD-search; therefore, only counting as 1x tail fiber domains.

d. Worked with phage experimentally in a lab (Bondy-Delomy *et al.*, 2016. Prophages mediate defense against phage infection through diverse mechanisms. ISME Journal (10); 2854–2866) and sequenced, but no annotations or TEM images.



Supplementary Figure 1. An amino acid alignment of the DLP1 and DLP2 baseplate or central tail hub proteins with their ten nearest neighbours. Black lettered amino acids are identical; lighter shaded amino acids have less homology. Consensus cutoff levels and amino

acids are shown below the grouping. The alignment was performed using EMBL-EBI Multiple Sequence Alignment Tool and visually presented using M-view