

Table S1. Colony site coordinates and date of collection for *V. vulgaris* samples used for total RNA transcriptomes from the native Belgian range and invasive New Zealand range.

Range	Colony	Site	Coordinates	Date
Invasive New Zealand	1	Six Mile Reserve	S -41.7685; E 172.9572	March 2014
	2	Tin Line Reserve	S -41.2809; E 173.5087	March 2014
	3	Braeburn Track	S -41.7954; E 172.5156	March 2014
Native Belgium	4	Wijgmaal, Leuven	N 50.5539; E 4.4143	September 2014
	5	Hasselt, Hasselt	N 50.5523; E 5.2017	September 2014
	6	Wezemaal, Leuven	N 50.5720; E 4.4444	September 2014

Table S2. Primers used in this study. *indicates Moku-like virus primers that were used for qPCR.

Name	Sequence (5'-3')	Product size (bp)	
<i>Virus primers</i>		<i>Sequencing</i>	<i>Screening</i>
Moku-like_88F	GTAAGTGGCTATCTCTGCTCG	1195	
Moku-like_1282R	GGTAACATCTCAATATACACACCC		
Moku-like_1119F	GTTGATACGATTGTTAATATTTGGG	1093	
Moku-like_2211R	TAATTGGTTCTCTAGTCTTAGTTCC		
Moku-like_2106F	GGAGATGTTAATTGTGATAATCCC	1391	106
Moku-like_3496R	GGAGTCACATACGTCTCTTCC		
Moku-like_3218F	TGTTGGATGTGCTATTAGTTGG	1128	279
Moku-like_4345R	TTGCCAAAGAGAGTATTCATTCC		
Moku-like_4265F	CATCCCTGTAGTGTACGC	1118	
Moku-like_5382R	GCCAGTATATCATACATTCTTGG		
Moku-like_5299F	GGAAGTCACATATAACTCAGC	1307	
Moku-like_6605R	GACTTTATCCAATGTATCACCAC		
Moku-like_6511F	CAATTCGATCTTATGTTTACTGG	1120	
Moku-like_7630R	GTTAAAGGTTTAGTAAGCATTTCC		
Moku-like_7517F	GAATGATGATGATGGAAATGCG	1290	
Moku-like_8806R*	CCAGGAATAGCACATTTTCGC		
Moku-like_8664F*	CGTCTTAAAGGCTTAGCACC	1255	143
Moku-like_9918R	CCGTGTTATAGCCACAACG		
Luteo-like_11F	GCGTTTCCGTTTCTATATTTTTGTG	1685	
Luteo-like_1695R	GACGATTTCTTCTGCTTCTTCG		
Luteo-like_1324F	GGACCAACTTATTCAGACTGC	1790	372
Luteo-like_3113R	CACCTTTGTTTCACCTCACAC		
Luteo-like_2364F	AAGAGTTGGCTGGTCCCCTT	483	
Luteo-like_2846R	CCCTTGAAGAGCATACCGGC		
<i>Reference gene primers for qPCR [1]</i>			
Ndufa8_Apis_F	GCACGATTACCAAGACCAA		78
Ndufa8_Apis_R	GGTGGAGCTACAGGCTCAGG		
Pros54_Apis_F	TCGAACCAAGATGGTACTGGAA		100
Pros54_Apis_R	TTGTTGTGCTTGCAGTCGTG		

Table S3. Total number of viruses present, and number of virus-like contigs assembled *de novo* from each transcriptome. The percentage of RNA-seq reads aligning to virus in each sample is indicated. (Colonies from Nelson, New Zealand (1-3) and Leuven, Belgium (4-6)).

	Larvae						Workers						Gynes					
	1	2	3	4	5	6	1	2	3	4	5	6	1	2	3	4	5	6
Total viruses present	7	11	4	7	21	31	9	5	4	7	5	4	1	2	1	1	3	1
# virus-like >200 bp	105	243	54	102	337	403	45	9	16	90	114	71	2	3	1	1	5	2
contigs >1000 bp	15	48	8	12	50	69	13	5	8	23	15	8	2	2	1	1	3	1
% of reads aligned to virus	0.27	0.15	0.08	0.6	0.75	0.76	0.21	0.43	0.05	0.99	0.49	0.27	0.04	0.22	0.05	0.13	0.15	0.17

Table S5: Average fold coverage and percentage of RNA reads aligning to *Vespula vulgaris* Luteo-like virus 1 and *Vespula vulgaris* Moku-like virus. (Colonies from Nelson, New Zealand (1-3) and Leuven, Belgium (4-6)).

		Larvae						Workers						Gynes					
		1	2	3	4	5	6	1	2	3	4	5	6	1	2	3	4	5	6
<i>Luteo-like virus</i>	Fold coverage	1630	286	841	5030	46	7378	969	974	827	2964	2391	4006	480	519	550	1923	1026	2446
	% reads	0.12	0.02	0.08	0.34	0.01	0.43	0.06	0.07	0.04	0.15	0.22	0.23	0.04	0.04	0.05	0.02	0.06	0.17
<i>Moku-like virus</i>	Fold coverage					770	30					119						537	
	% reads					0.09	0.01					0.04						0.10	

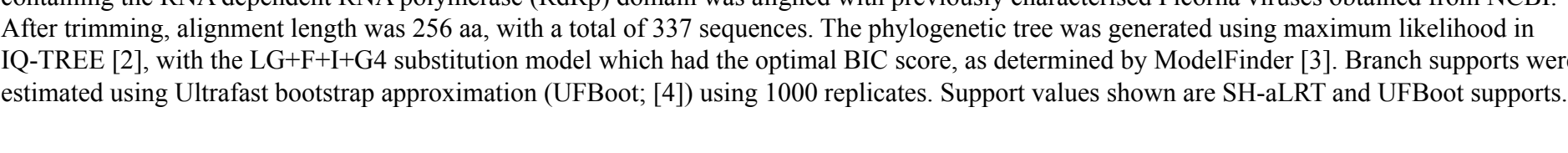




Figure S3. Detailed Partiti phylogenetic tree from Figure 1. Phylogenetic tree of the novel Partiti viruses identified in *Vesputula vulgaris*. Novel viruses are shown in red. The open reading frame containing the RNA dependent RNA polymerase (RdRp) domain was aligned with previously characterised Partiti viruses obtained from NCBI. After trimming, alignment length was 405 aa, with a total of 116 sequences. The phylogenetic tree was generated using maximum likelihood in IQ-TREE [2], with the LG+F+I+G4 substitution model which had the optimal BIC score, as determined by ModelFinder [3]. Branch supports were estimated using Ultrafast bootstrap approximation (UFBoot; [4]) using 1000 replicates. Support values shown are SH-aLRT and UFBoot supports.



Figure S4. Detailed Tombus phylogenetic tree from Figure 1. Phylogenetic tree of the novel Tombus viruses identified in *Vespula vulgaris*. Novel viruses are shown in red. The open reading frame containing the RNA dependent RNA polymerase (RdRp) domain was aligned with previously characterised Partiti viruses obtained from NCBI. After trimming, alignment length was 344 aa, with a total of 136 sequences. The phylogenetic tree was generated using maximum likelihood in IQ-TREE [2], with the rtREV+F+I+G4 substitution model which had the optimal BIC score, as determined by ModelFinder [3]. Branch supports were estimated using Ultrafast bootstrap approximation (UFBoot; [4]) using 1000 replicates. Support values shown are SH-aLRT and UFBoot supports.

Supplementary references

1. Cameron, R.C.; Duncan, E.J.; Dearden, P.K. Stable reference genes for the measurement of transcript abundance during larval caste development in the honeybee. *Apidologie* **2013**, *44*, 357-366.
2. Nguyen, L.-T.; Schmidt, H.A.; von Haeseler, A.; Minh, B.Q. Iq-tree: A fast and effective stochastic algorithm for estimating maximum-likelihood phylogenies. *Mol Biol Evol* **2014**, *32*, 268-274.
3. Kalyaanamoorthy, S.; Minh, B.Q.; Wong, T.K.F.; von Haeseler, A.; Jermiin, L.S. Modelfinder: Fast model selection for accurate phylogenetic estimates. *Nat Methods* **2017**, *14*.
4. Hoang, D.T.; Chernomor, O.; von Haeseler, A.; Minh, B.Q.; Vinh, L.S. Ufboot2: Improving the ultrafast bootstrap approximation. *Mol Biol Evol* **2017**, *35*, 518-522.