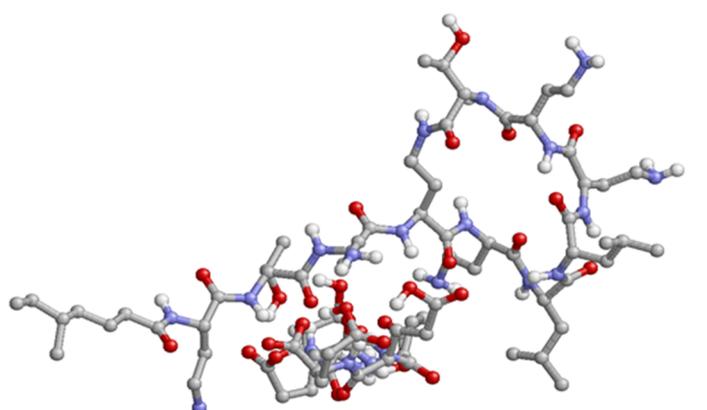
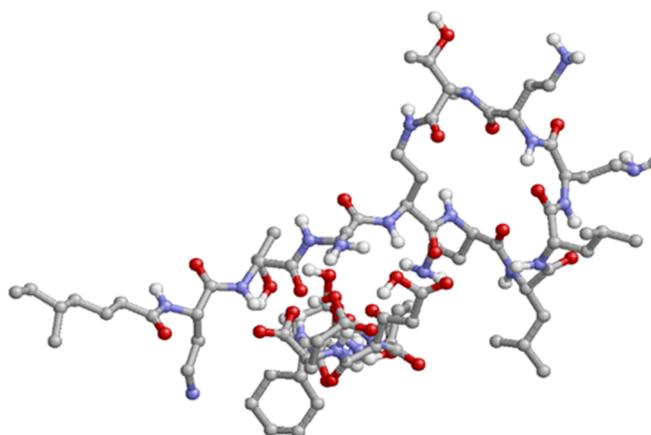


Supplementary Materials: Polypeptide self-assembled nanoparticles as delivery systems for polymyxins B and E

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(a)



(b)

Figure S1. Molecular docking of interaction between polymyxin E and pentapeptides: (a) Glu₅; (b) Glu₂PheGlu₂.

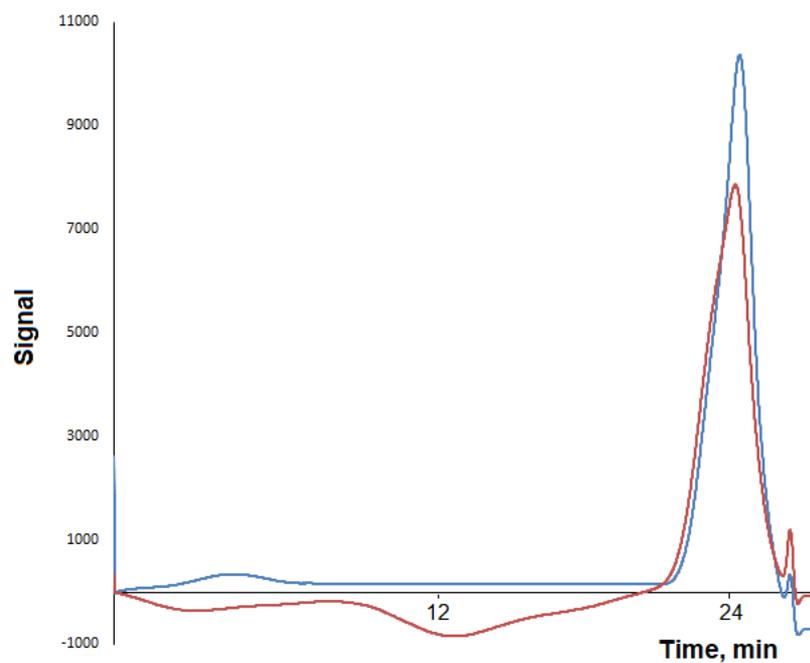


Figure S2. SEC traces of P(Glu-co-DPhe): blue line – sample 1; red line – sample 2.

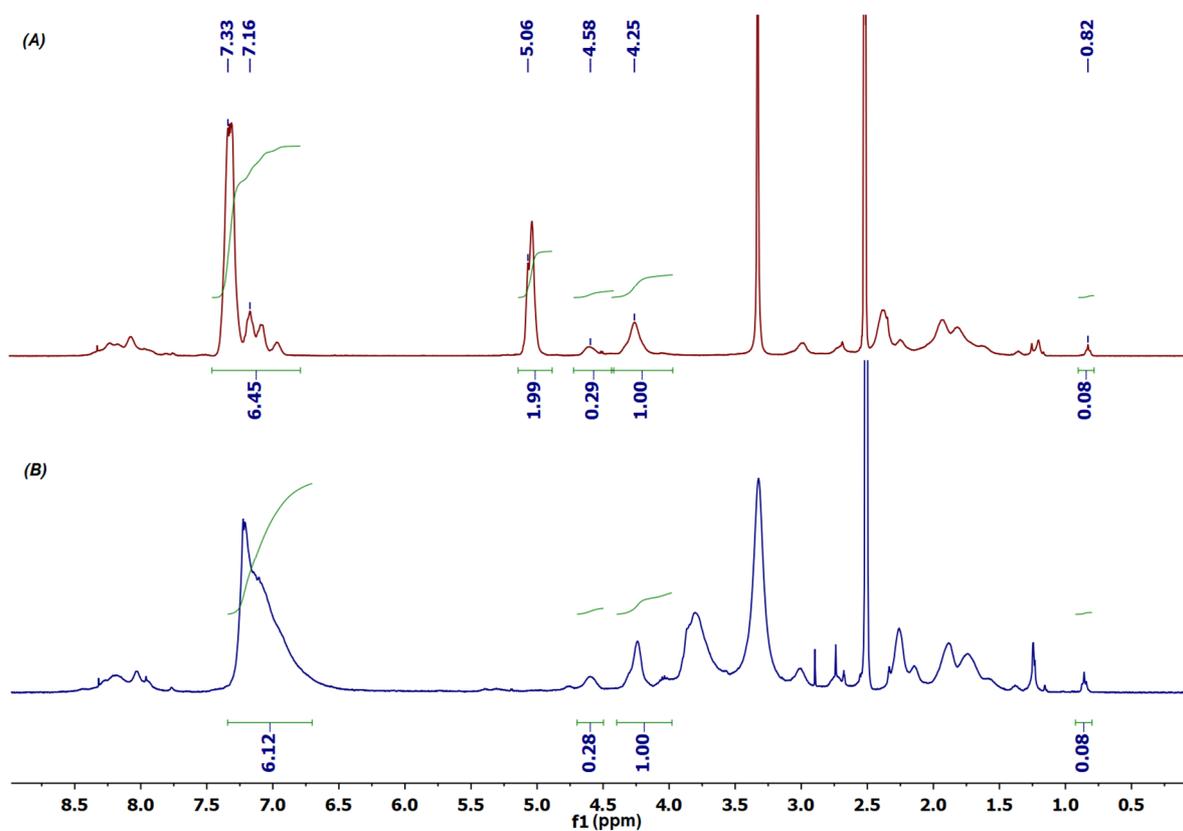
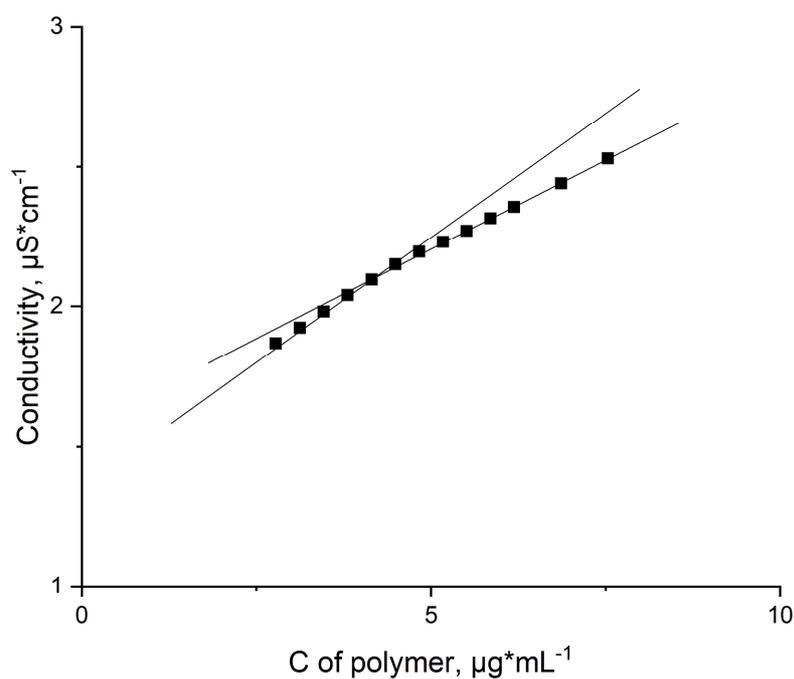


Figure S3. NMR spectra of P(Glu(OBzl)-co-DPhe) (a) and P(Glu-co-DPhe) (b) in DMSO-d₆.

Table S1. Characteristics used for calculations of M_{w0} for synthesized copolymers by static light scattering.

Sample	$dn/dc, \text{cm}^3/\text{g}$	$A_2, \text{cm}^3 \cdot \text{mol} \cdot \text{g}^{-2}$
1	0.1132	-8.56E-05
2	0.1171	-3.49E-04

**Figure S4.** Dependence of conductivity on polymer concentration in solution: determination of critical micelle concentration (CMC) by conductometry.**Table S2.** Characteristics of PLA and PLA-*b*-PEG nanoparticles used as control for comparison phagocytosis rate.

Nanoparticles	D_H, nm	ζ -potential, mV
PLA	92 ± 13	-8.1 ± 2.2
PLA- <i>b</i> -PEG	121 ± 16	-0.6 ± 1.1

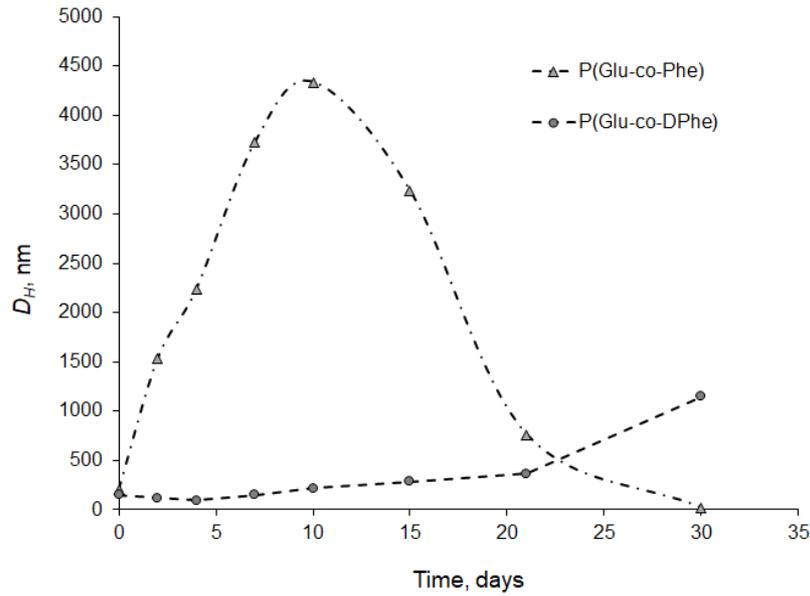


Figure S5. Stability of P(Glu-co-Phe) and P(Glu-co-DPhe) nanoparticles in the enzyme containing medium. *Conditions:* 0.01 M PBS containing papain; the concentration of nanoparticles was 1.0 mg/mL; the concentration of papain was 0.05 mg/mL; the suspension was incubated at 37 °C. In 15 days the medium was replaced with a fresh portion.

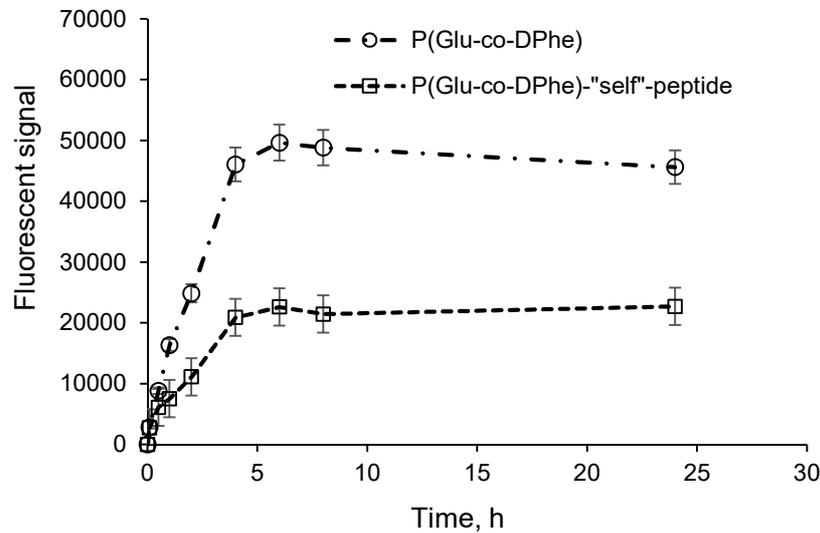
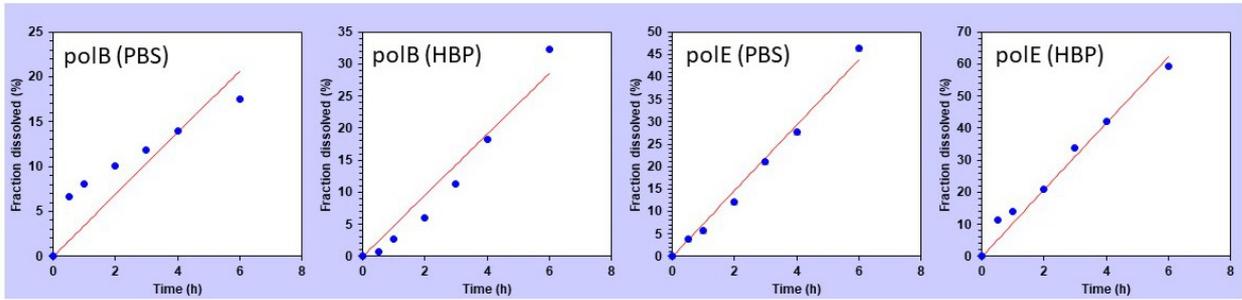
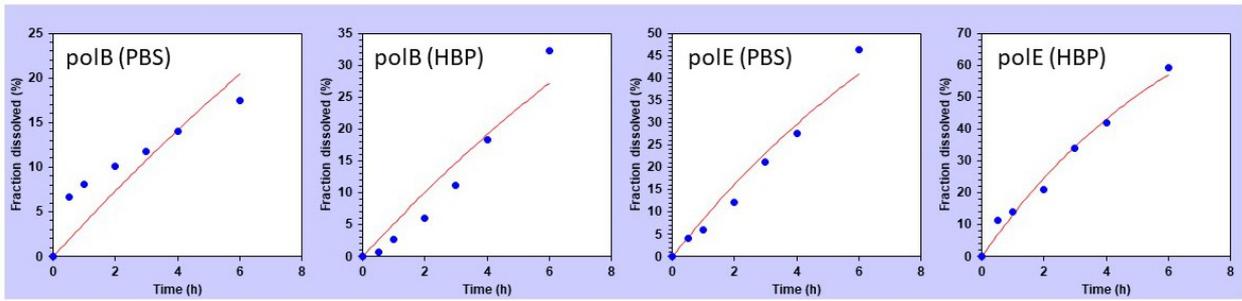


Figure S6. Uptake rate of fluorescently-labeled P(Glu-co-DPhe) NPs and these modified with "self"-peptide by macrophages. *Conditions:* J774A.1 cell line (mouse macrophages); mouse "self"-peptide: GNYTCEVTELTREGETIIEELK; flow cytometry; concentration of NPs was 50 μ g/mL; amount of bound "self-peptide" as 50 μ g/mg of NPs; dansylcadaverine was used as a fluorescent label.

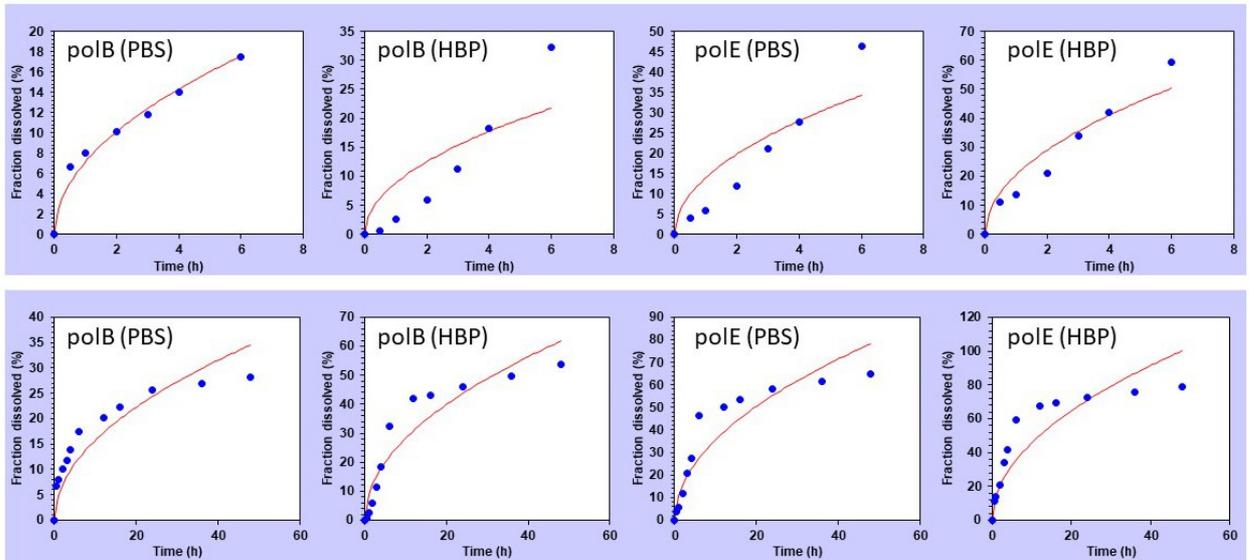
Zero-order $F=k_{zo} * t$



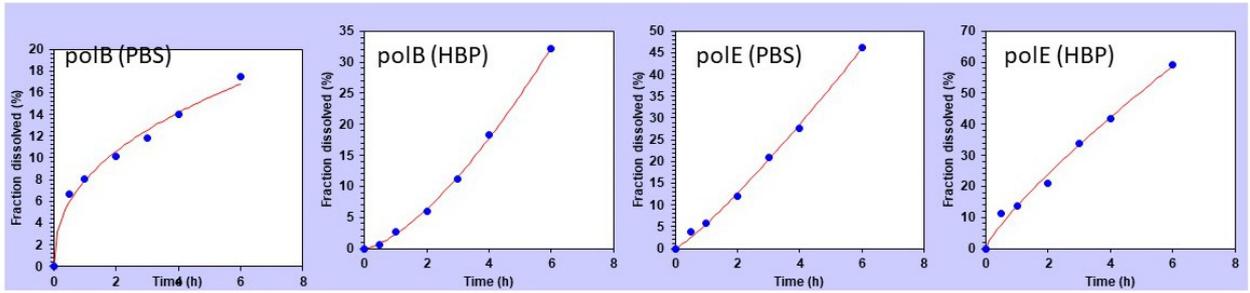
First-order $F=100*[1-Exp(-k_{fo} * t)]$



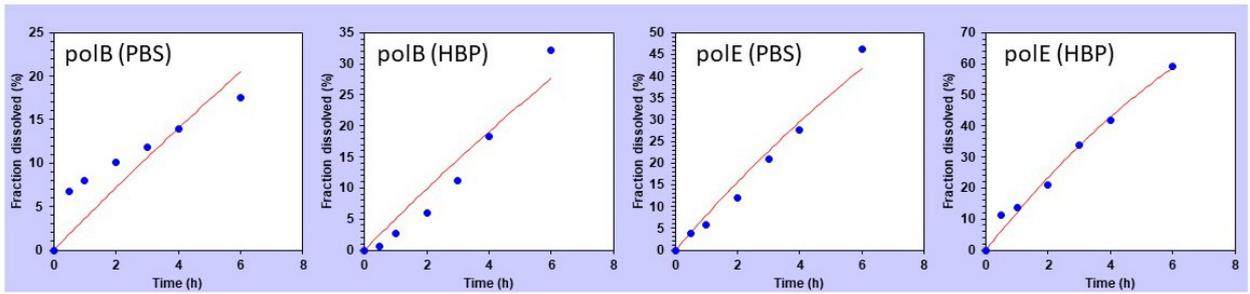
Higuchi $F=k_H * t^{0.5}$



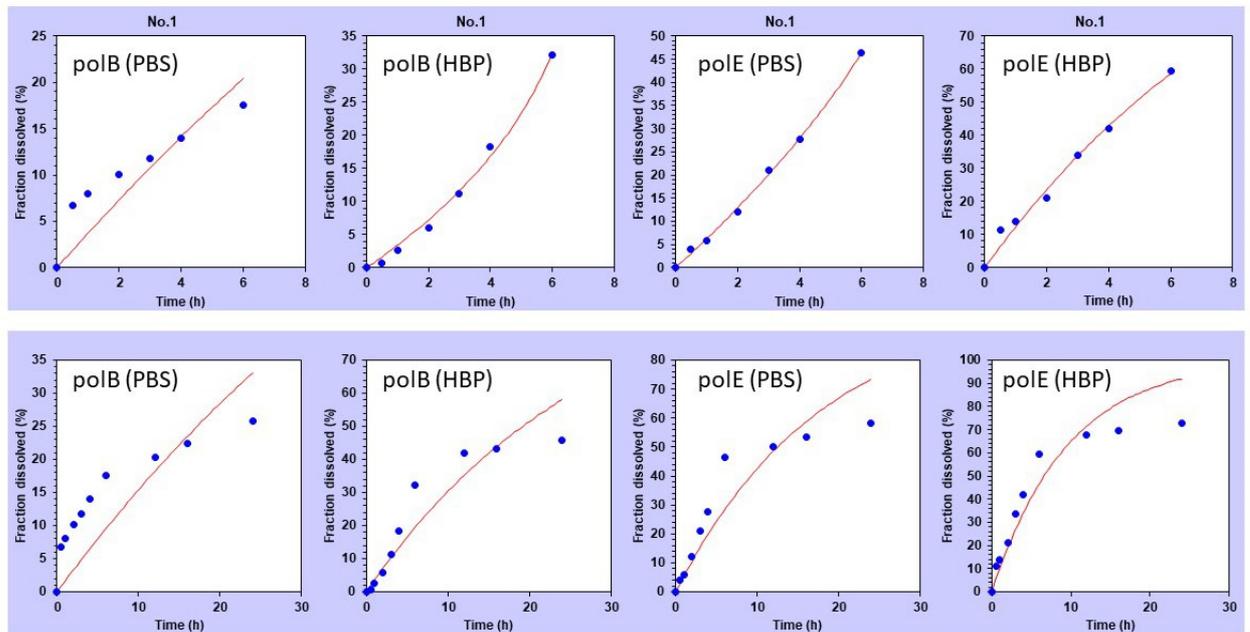
Korsmeyer-Peppas $F = k_{KP} * t^n$



Hixson-Crowell $F = 100 * [1 - (1 - k_{HC} * t)^3]$

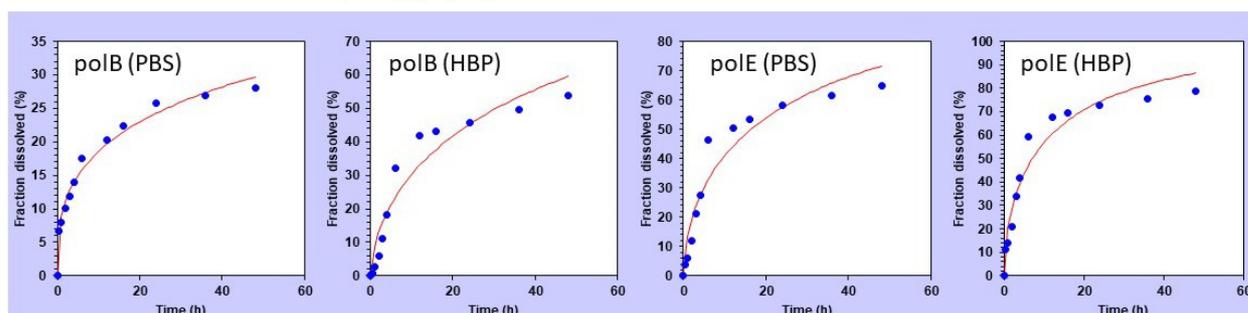


Hopfenberg $F = 100 * [1 - (1 - k_{HB} * t)^n]$

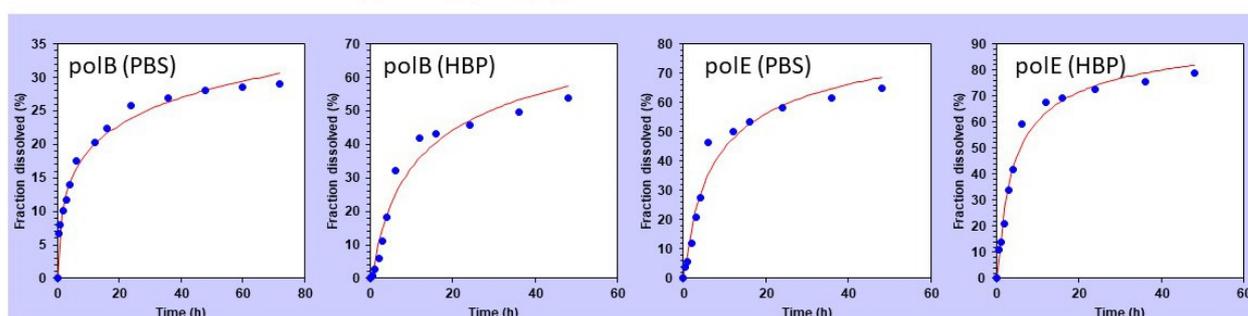


Weibull

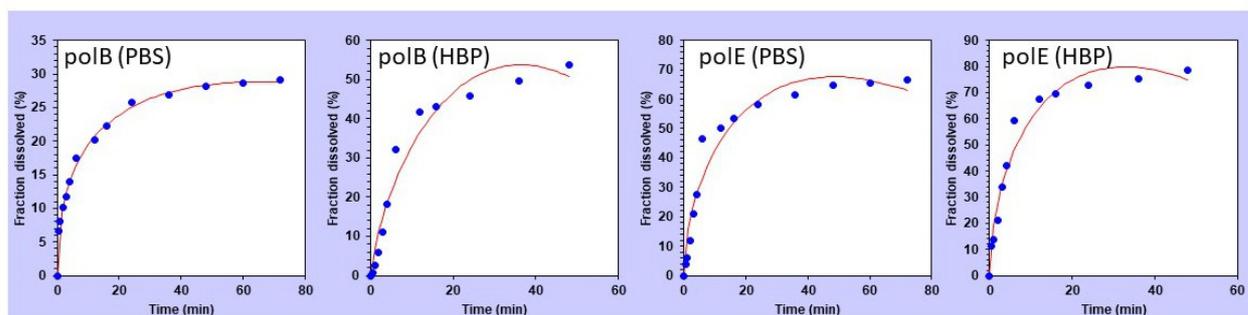
$$F=100*\{1-Exp[-(t^\beta)/\alpha]\}$$

**Gompertz**

$$F=100*Exp\{-\alpha*Exp[-\beta*log(t)]\}$$

**Peppas-Sahlin**

$$F=K_1*t^m+K_2*t^{(2*m)}$$

**Baker-Lonsdale**

$$3/2*[1-(1-F/100)^{(2/3)}]-F/100=k_{BL}*t$$

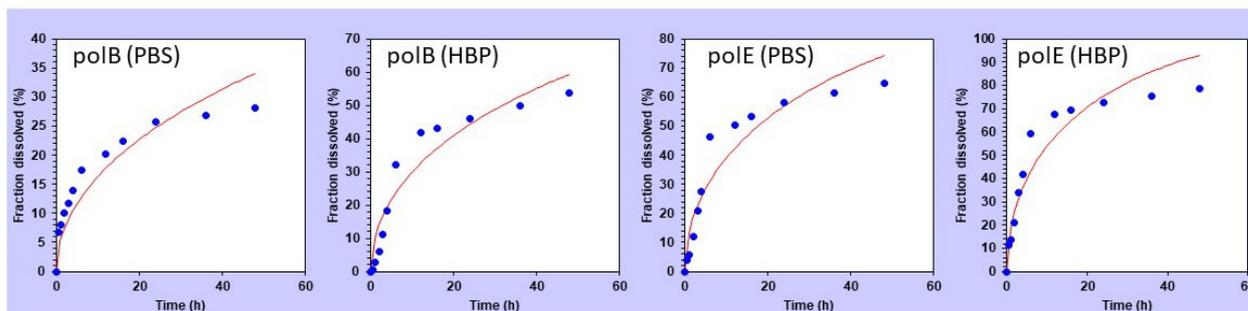


Figure S7. Drug dissolution kinetics models and approximation curves.

Abbreviations: PolE: polymyxin E; PolB: polymyxin B; PBS: 0.01 M sodium phosphate buffer, containing 0.15 mol/L NaCl; HBS: human blood plasma.

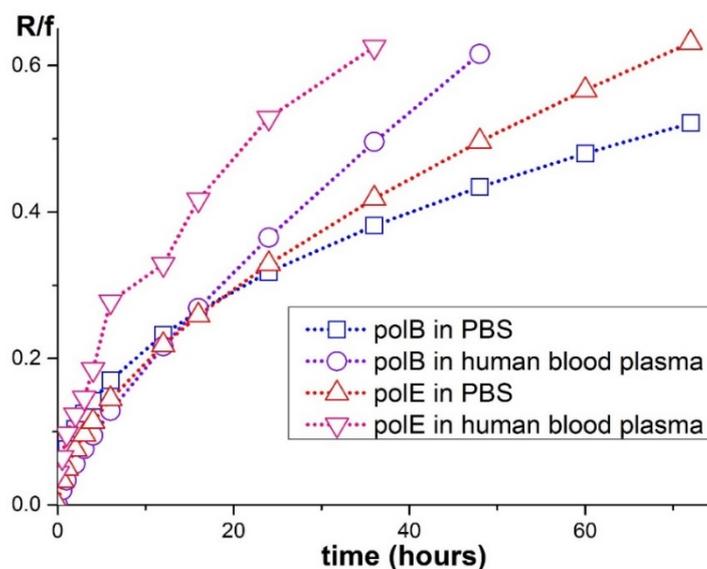


Figure S8. The dependence of drug dissolution mechanism with time. R/f is the ratio of polymer relaxation (R) to Fick's diffusion (f) as the driving mechanisms of drug release for the system. At definite time R/f was calculated as follows: $\frac{R}{f} = \frac{K_2 \times t^m}{K_1}$, where K_1 , K_2 and m are parameters determined by approximation of release data with Peppas-Sahlin model and t is time. *Abbreviations:* PolE: polymyxin E; PolB: polymyxin B; PBS: 0.01 M sodium phosphate buffer, containing 0.15 mol/L NaCl; HBS: human blood plasma.