

Supplementary material

Table S1. Primers employed for molecular characterization of the *H. pylori* isolates from the Antiguo Hospital Civil de Guadalajara.

Primer	Sequence	Amplicon (pb)	Source
<i>cagA</i> -F <i>cagA</i> -R	ACC CTA GTC GGT AAT GGG TTA GGC TGT TAG TAG CGT AAT TGT C	643	Occhialini <i>et al.</i> , 2001 [42]
<i>vacAm1</i> -F <i>vacAm1</i> -R	GGT CAA AAT GCG GTC ATG G CCA TTG GTA CCT GTA GAA AC	290	García <i>et al.</i> , 2006 [43]
<i>vacAm2</i> -F <i>vacAm2</i> -R	GGA GCC CCA GGA AAC ATT G CAT AAC TAG CGC CTT GCA C	352	García <i>et al.</i> , 2006 [43]
<i>vacAs1a</i> -F <i>vacAs1a</i> -R	GTC AGC ATC ACA CCG CAA C CTG CTT GAA TGC GCC AAA C	190	García <i>et al.</i> , 2006 [43]
<i>vacAs1b</i> -F <i>vacAs1b</i> -R	AGC GCC ATA CCG CAA GAG CTG CTT GAA TGC GCC AAA C	187	García <i>et al.</i> , 2006 [43]
<i>vacAs2</i> -F <i>vacAs2</i> -R	GCT AAC ACG CCA AAT GAT CC CTG CTT GAA TGC GCC AAA C	199	García <i>et al.</i> , 2006 [43]
<i>tonB</i> -F <i>tonB</i> -R	GGA TAC CTC AAA ACG CGC AT TCA ATT GGA CTC CGC CTT CT	197	Designed in primer3
<i>ureA</i> -F <i>ureA</i> -R	GCC AAT GGT AAA TTA GTT CTC CTT AAT TGT TTT TAC	410	Russo <i>et al.</i> , 1999 [44]
16S-fD1 16S-rD1	CCGAATTCGTCGACAACAGAGTTTGAT CCTGGCTCAG CCCGGGATCCAAGCTTAAGGAGGTGAT CCAGCC	1500	Weisburg <i>et al.</i> , 1991 [19]
<i>ureC</i> -F <i>ureC</i> -R	TAT AAT CAT GAA AAT TTT TGG GAC T TTA GCT GCA GTT AGC ACA AAT GCC CTT C	1344	De Reuse <i>et al.</i> , 1997 [7]

Table S2. Thermal profiles for amplification of different genes present in the *H. pylori* strain genome through end-point PCR.

Gene*	Denaturalization (initial denaturalization)	Alignment	Extension (final extension)
<i>cagA</i>	94°C 30 s (3 min)	54°C, 30 s	72°C, 1 min (5min)
<i>vacAm1</i>	95°C 1 min (1min)	52°C, 1 min	72°C, 1 min (7 min)
<i>vacAm2</i>	94 °C 30 s (3 min)	54°C, 30 s	72°C, 1 min (5 min)
<i>vacAs1a</i>	95 °C 1 min (5 min)	52°C, 1 min	72°C, 1 min (7 min)
<i>vacAs1b</i>	95 °C 1 min (5 min)	52°C, 1 min	72°C, 1 min (7 min)
<i>vacAs2</i>	95 °C 1 min (5 min)	52°C, 1 min	72°C, 1 min (7 min)
<i>tonB</i>	94°C 30 s (3 min)	54°C, 30 s	72°C, 1 min (5 min)
<i>ureA</i>	94°C 30 s (3 min)	42°C, 30 s	72°C, 1 min (5 min)
16S	94°C 1 min (1min)	54°C, 1 min	72°C, 1 min (5 min)
<i>ureC</i>	95°C 1 min (1min)	55°C, 2 min	72°C, 2 min (7 min)

*34 amplification cycles. The polymerase Dream Taq DNA enzyme (Thermo Fisher Scientific, USA) was used.

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