

Review

The Summary of Nitrification Process in Mainstream Wastewater Treatment

Hongjun Zhao¹, Yan Guo^{1,2,*}, Qunhui Wang^{1,2} , Ze Zhang¹, Chuanfu Wu^{1,2}, Ming Gao^{1,2,*} and Feng Liu³

¹ School of Energy and Environmental Engineering, University of Science and Technology Beijing, Beijing 100083, China

² Beijing Key Laboratory of Resource-Oriented Treatment of Industrial Pollutants, Beijing 100083, China

³ TUS-Environmental Science and Technology Development Corporation, Ltd., Beijing 100084, China

* Correspondence: ustbgy2015@163.com (Y.G.); gaoming402@gmail.com (M.G.)

Abstract: The application of the mainstream partial nitrification/anammox (PN/A) process is promising due to the huge cost reduction compared to traditional biological nitrogen removal. However, the nitrite production rate (NPR) of a biological nitrification pre-treatment process is relatively lower than the nitrite consumption rate in a pure anammox reactor with a high nitrogen loading rate (NLR). Thus, the NPR is the rate-limiting step for operating the PN/A process with a higher NLR. Various studies have attempted to improve mainstream NPR. A comprehensive review of these processes is needed for the actual application of the PN/A process. This study focuses on: (1) various nitrite production processes that have emerged in recent years; (2) the main microbial species and characteristics involved in biological nitrification; (3) the existing problems and the N₂O emission problem of these processes; and (4) a proposed novel and promising PN/A process facilitated with photocatalyst oxidation. This review is expected to provide references and a basis for the research on the nitrification step of the application of the mainstream PN/A process.



Citation: Zhao, H.; Guo, Y.; Wang, Q.; Zhang, Z.; Wu, C.; Gao, M.; Liu, F.

The Summary of Nitrification Process in Mainstream Wastewater Treatment. *Sustainability* **2022**, *14*, 16453.

<https://doi.org/10.3390/su142416453>

Academic Editors: Yu-You Li, Yu Qin and Jiayuan Ji

Received: 7 November 2022

Accepted: 7 December 2022

Published: 8 December 2022

Publisher's Note: MDPI stays neutral with regard to jurisdictional claims in published maps and institutional affiliations.



Copyright: © 2022 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (<https://creativecommons.org/licenses/by/4.0/>).

Keywords: nitrite production; nitrification; photocatalytic oxidation; mainstream wastewater; N₂O emission; anammox

1. Introduction

Since its discovery in the 1990s, the anammox process has been widely developed as a promising technology due to its obvious advantages of energy reduction and high efficiency. The anammox process has shed light on the revolution of wastewater treatment plants (WWTPs) towards the energy-neutral even output installations [1,2]. In an anammox reaction, the reactants of ammonium (NH₄⁺) and nitrite (NO₂⁻) can be mainly converted to nitrogen gas to achieve nitrogen removal. The reaction ratio of NO₂⁻-N/NH₄⁺-N is usually considered to be 1.146 for mainstream wastewater [3]. Given that the main nitrogen species in actual mainstream wastewater is NH₄⁺-N, the required NO₂⁻-N/NH₄⁺-N ratio for the anammox reaction can be achieved through the conversion of the part of influent NH₄⁺-N to nitrite. However, the nitrite consumption rates of pure anammox reaction in many studies are noticeably higher than the nitrite production rate (NPR) in nitrification research for mainstream wastewater [4–6]. Thus, the nitrification process is the rate-limiting step of the partial nitrification/anammox (PN/A) process for chasing higher nitrogen removal rate (NRR) and strategies and solutions to improve NPR are urgently needed.

As is indicated in Figure 1, there are two main strategies for achieving nitrite production: the biological nitrification process and the physicochemical photocatalytic oxidation process. In the physicochemical photocatalytic oxidation process, the reaction involves oxidizing ammonia to nitrite with a combination of light and a photocatalyst. The main drawback of this method is that the remaining oxidant may oxidize nitrite to nitric acid. Current research shows that changing the type of catalyst used can effectively inhibit the oxidation of nitrite. TiO₂ modified with Ag₂O showed excellent purification performance

in the study of photocatalytic oxidation treating 100 mgNH₄⁺-N/L synthesis wastewater. Among them, the nitrite production efficiency (NPE) reached 40.8% with 0.12 kg/m³/d NPR at a temperature of 21 °C, which is twice the catalytic effect of ordinary TiO₂ [7,8]. This method is promising because the temperature and ammonium concentration are close to mainstream wastewater treatment conditions. Additionally, photocatalytic oxidation will not cause N₂O emission like the biological nitrification process does and N₂O production can be ignored without aeration [9].

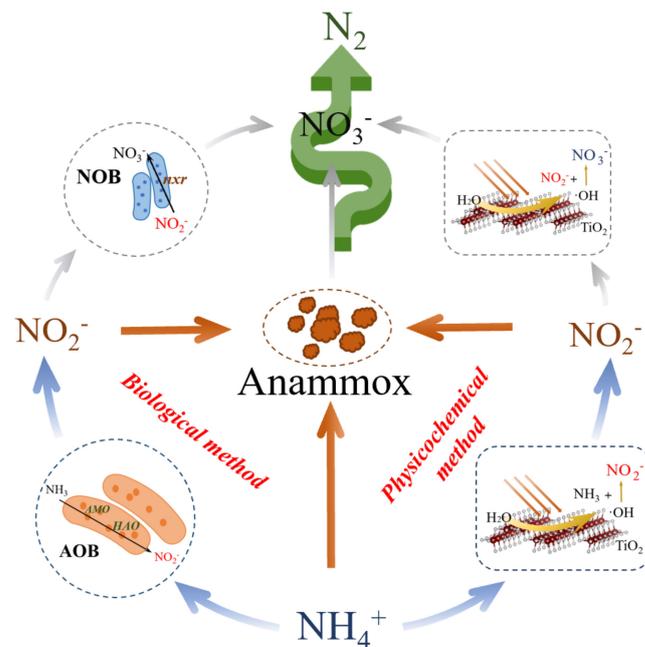


Figure 1. The nitrogen removal pathway of nitrite production and anammox process. Nitrite production process mainly includes a biological method and a physicochemical method.

The biological nitrification method is to oxidize ammonium to nitrite using ammonium-oxidizing bacteria (AOB). One of the major reasons for the difficulty in achieving a high NPR and NPE for biological nitrification is the presence of nitrite-oxidizing bacteria (NOB), which can further oxidize nitrite to nitrate. Generally, the control of operating conditions such as temperature, pH, free ammonium (FA), free nitrite acid (FNA), etc., can effectively suppress NOB [3,10]; however, the effect of these conditions cannot reach the requirements of anammox in mainstream wastewater [11,12]. Therefore, some studies have attempted to improve the NPR of nitrification through physical, chemical, and other assisted methods and have made promising progress. For example, in a previous study nitrification assisted by light for synthetic wastewater treatment was achieved with an NPR of 0.13 kg/m³/d at 25 °C and influent 30 mg NH₄⁺-N/L [13]. Another study obtained an NPR higher than 1.0 kg/m³/d by adjusting salinity to 10 g NaCl/L at 30 °C and 100 mgNH₄⁺-N/L [14]. However, NO₂⁻ may be reduced to Nitrous oxide (N₂O) by AOB at elevated nitrite concentrations or low oxygen conditions in the nitrification process, which is known as [15,16]. Higher NPR values increase the concentration of nitrite in the reactor, which may exacerbate nitrifier denitrification and higher NPR may therefore weaken the nitrification efficiency. However, the fluctuation of influent quality and quantity, low temperature, improper pH, and other harsh conditions will have unignorable effects in the application of biological nitrification and photocatalyst oxidation for actual mainstream treatment. Therefore, it is significant to conduct a comprehensive review of both these processes for selecting a properly effective method to achieve the efficient nitrite production and less N₂O in specific practical situations.

This paper summarizes the current methods of nitrite production, reviews the microbial main species and characteristics involved in biological nitrification process, outlines the

existing problems and N₂O emissions in various nitrite production processes, and puts forward the promising process of photocatalytic oxidation for nitrite production and anammox process for nitrogen removal.

2. Photocatalyst Oxidation

There are many physicochemical methods for wastewater ammonium removal including stripping [17], adsorption [18], photocatalytic oxidation [7], breakpoint chlorination [19] and more. These methods usually directly convert ammonium into harmless nitrogen or nitrate, except for photocatalytic oxidation, which can be used to generate nitrite and accumulate. Photocatalytic oxidation refers to a process in which light irradiates the surface of a photocatalyst, resulting in the electrons on its surface absorbing the photo energy and becoming photoexcited electrons. The photoexcited electrons convert to the empty conduction band from the filled valence band, leaving holes in the valence band. Subsequently, an electron–hole pair (e[−]-h⁺) is generated [20]. Holes combine with H₂O to form a variety of oxidants, mainly hydroxyl radicals, and pollutants are degraded by reacting with hydroxyl radicals in the wastewater, as is shown in Equations (1) and (2) [21]. The main pollutant, ammonia, will be oxidized by hydroxyl radicals to various products such as NO₃[−], NO₂[−], and N₂.



Titanium dioxide (TiO₂)-based materials are promising photocatalysts for NH₄⁺-N/NH₃ oxidation [7,9,22,23]. Nevertheless, the performance of the naked TiO₂ is unsatisfactory, due to the fact that recombination (in which the valence band holes and conduction band electrons simply recombine to liberate heat or light [24]) is easy to incur in most instances when light shines on TiO₂. However, TiO₂ modified with metals or their oxides can inhibit the recombination. Particles of metals or their oxides dispersed on the surface of TiO₂ can also produce electrons and holes. These electrons and holes change with electrons and holes produced by TiO₂ to inhibit the recombination [7,24]. Therefore, these photocatalysts have higher efficiency.

Modifying TiO₂ catalysts with different metals makes their surface adsorption energy intensity different. According to the difference of surface adsorption energy intensity, it has been suggested that the removal of ammonium can be divided into two ways [7,25], as is shown in Figure 2. In path I, the hydroxyl radical formed by Equation (1) reacts with ammonium to produce amino radicals and the two amino radicals combine to produce hydrazine. Hydrazine will be photochemically converted to diazene (N₂H₂). Finally, diazene can easily decompose into hydrogen and nitrogen under ambient temperature and pressure. The Pt and Pd surfaces have the best medium atomic nitrogen affinity for the production of dinitrogen, which makes the use of such catalysts more conducive to the production of nitrogen through path I [22]. In path II, ammonium molecules or amino radicals react with hydroxyl radicals to form hydroxylamine. Hydroxylamine is continuously oxidized by hydroxyl radicals to form nitrite ions and nitrate ions. The surface affinity of Ru and Rh is too strong to recombine and Au and Ag are too weak to produce active intermediates. Therefore, TiO₂ catalysts modified with Ru, Rh, Au and Ag are comprised to realize stable nitrite production. Some other catalysts can also be used to produce NO₂[−] and NO₃[−] in the photocatalytic oxidation reaction, such as atomic single layer graphitic-C₃N₄ (SL g-C₃N₄) [26].

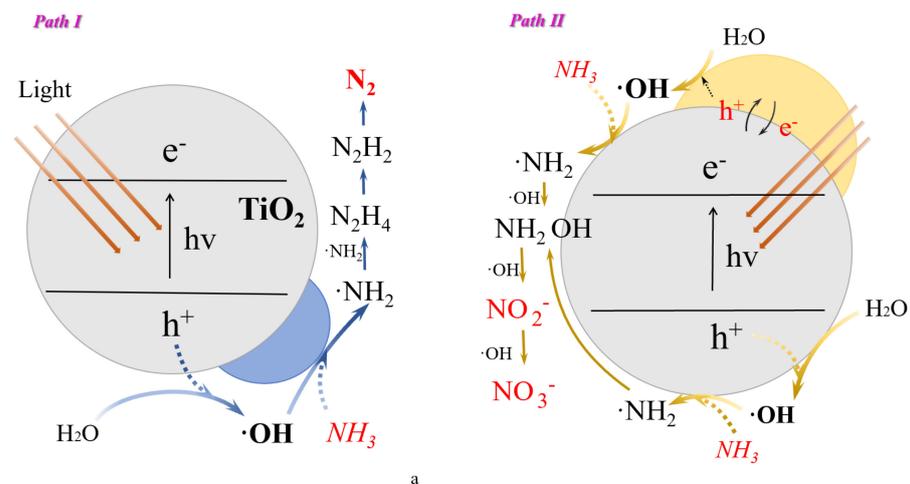


Figure 2. Possible reaction path of ammonium under photocatalytic oxidation: (a) the pathway to generate nitrogen; (b) the pathway to generate nitrite.

Under ultraviolet light and extreme alkaline conditions (pH = 12.5), synthetic wastewater (130 mgNH₄⁺-N/L) treated by TiO₂ showed results including a 26% ammonium-removal efficiency (ARE), a 25.5% NPE and a 0.13 kg/m³/d NPR [8]. However, the reaction condition occurred at 30 °C and the high pH value is quite difficult to achieve under the actual treatment conditions of mainstream wastewater. Therefore, more suitable catalysts are needed. From above, using TiO₂ modified with Au, the synthetic mainstream wastewater (130 mgNH₄⁺-N/L) treatment was achieved with a 27% ARE at 30 °C and a pH value of 10, and the NPE and NPR could reach 21.6% and 0.11 kg/m³/d, respectively [27]. It is obvious that this catalyst yielded a higher ARE with a lower pH condition, however, the NPE and NPR were lower than that achieved by naked TiO₂. Au and Ag are similar in surface adsorption energy intensity characteristic. Therefore, TiO₂ modified with Ag or Ag₂O may also be promising. When the pH value was 11 and the temperature was 21 °C, ARE could reach 47.3% through TiO₂ modified with Ag₂O for synthetic wastewater (100 mgNH₄⁺-N/L) treatment, where the NPE and NPR were 40.8% and 0.12 kg/m³/d, respectively [7]. Compared to the above catalysts, the temperature of the wastewater treated by this catalyst was lower, which is closer to the mainstream conditions and seems to be a more suitable choice. Additionally, the NPE of this catalyst was higher, so more nitrite could be produced to participate in anammox. Other types of catalysts were also investigated. The activated carbon (AC-NiFe₂O₄) catalyst bonded on the iron–nickel oxide could oxidize 87% of ammonium and almost all of it was converted to nitrite with 0.15 kg/m³/d NPR at 25 °C and a pH of 10.5 in the presence of H₂O₂ [28]. This catalyst is also expected to be used in the nitrite production process of mainstream wastewater.

3. Biological Nitrification

The conversion of ammonium to nitrite achieved through biological metabolism is known as nitrification. A nitrification reaction is expressed in Equation (3) [29]. Nitrification-based nitrogen removal processes mainly include the nitrite shunt process and the PN/A process. The nitrite shunt process is composed of nitrification and denitrification, and the PN/A process consists of nitrification and anammox. The nitrite shunt process consists of two steps: nitrification and denitrification, which involve the oxidation of ammonium to nitrite by AOB followed by nitrite reduction to nitrogen using denitrifying bacteria [30]. Equations (4)–(7) express denitrification, the nitrite shunt process, anammox, and the PN/A reaction, respectively [29,31]. The main configuration of the nitrite shunt process is a one-stage mode [30,32,33], while the configuration of the PN/A process includes one-stage [34] and two-stage modes [35,36], as are shown in Figure 3. The two-stage mode can be further divided into two sub-modes. In the first sub-mode, the wastewater enters the anammox unit after meeting the requirement (NO₂⁻/NH₄⁺ ratio = 1.146) through the partial nitrification

unit. In the second sub-mode, according to the theoretical calculation of the equations and under ideal conditions, part (53.4%) of the wastewater enters the nitritation unit where ammonium is completely oxidized into nitrite. Subsequently, the effluent of the nitritation unit is mixed with the remaining wastewater (46.6%) to meet the requirement ($\text{NO}_2^-/\text{NH}_4^+$ ratio = 1.146) and the mixed wastewater enters the anammox unit. However, in practice, AOB cannot convert all ammonia into nitrite. Therefore, the actual amount used to generate nitrous acid should be significantly higher than 53.4%. Some studies show that a proportion of 57.8–65.9% of mainstream wastewater is used to produce nitrous acid and, finally, 93% of the ammonia removal rate is achieved [36]. Compared to the nitrite shunt process, the PN/A process uses less oxygen based on the theoretical calculation (a 0.436 mol O_2 /mol NH_4^+ -N reduction in oxygen consumption) [37]. The nitrite shunt process has a higher NRE than PN/A because all ammonium is oxidized to nitrite and no nitrate is produced, but its obvious disadvantage is that a carbon source is required in the denitritation step [38]. This paper summarizes the biological nitritation process by dividing this section into two parts: pure nitritation process and assisted nitritation process for a more detailed introduction.

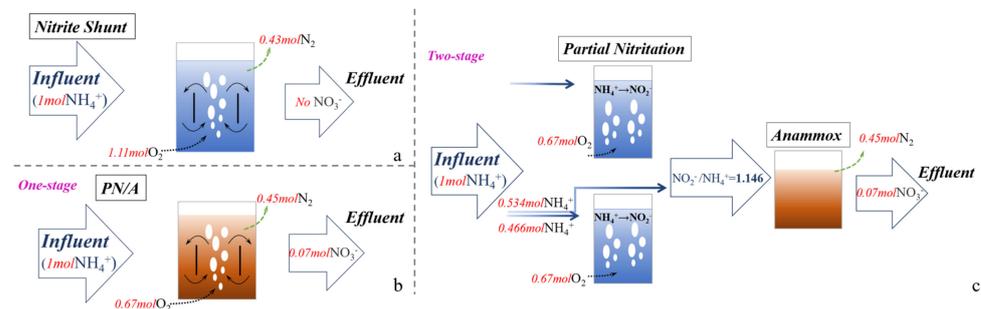
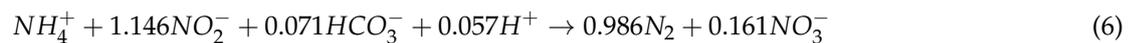
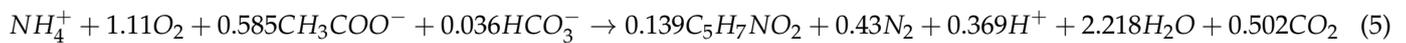
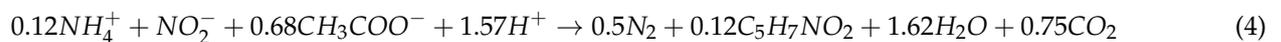
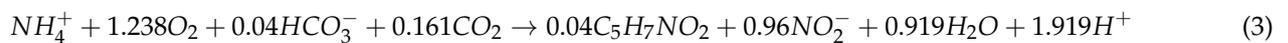


Figure 3. The process flow of nitritation-based nitrogen removal processes: (a) one-stage nitrite shunt process; (b) one-stage PN/A process; and (c) two-stage PN/A process.

3.1. Pure Nitritation

3.1.1. Research Progress

Pure nitritation refers to nitritation without an abiotic auxiliary method. NOB inhibition is an important issue in pure biological nitritation. Given that NOB can proliferate more rapidly than AOB in improper conditions, part of the nitrite oxidation will be in vain, which can finally decrease the NPE. Consequently, various measures in pure nitritation such as regulating and controlling temperature, DO, pH, sludge retention time (SRT) and hydraulic retention time (HRT) are adopted to inhibit NOB activity and growth. The main studies that achieved desirable nitritation through pure nitritation are shown in Table 1.

In Table 1, NPE, NPR and the nitrite accumulation ratio (NAR) are calculated as follows:

$$\text{NPE} = \frac{[\text{NO}_2^- - \text{N}]}{[\text{NH}_4^+ - \text{N}]_{inf}} \times 100\% \quad (8)$$

$$\text{NPR} = \text{NLR} \times \text{NPE} \quad (9)$$

$$\text{NAR} = \frac{[\text{NO}_2^- - \text{N}]}{[\text{NH}_4^+ - \text{N}]_{inf} - [\text{NH}_4^+ - \text{N}]_{eff}} \times 100\% \quad (10)$$

where $[\text{NO}_2^- - \text{N}]$ refers to nitrite generated, $[\text{NH}_4^+ - \text{N}]_{\text{inf}}$, $[\text{NH}_4^+ - \text{N}]_{\text{eff}}$ refer to the ammonium concentration of the influent and effluent, respectively, and NLR refers to the nitrogen load rate.

A previous study reported the highest NPR of $0.74 \text{ kg/m}^3/\text{d}$ with a 57.63% NPE for treating synthetic wastewater ($63 \text{ mgNH}_4^+ - \text{N/L}$) at 25°C [39]. A maximum NPE of 86.8% with 75 mg/L of the influent $\text{NH}_4^+ - \text{N}$ has also been reported, while the NPR merely reached a common level at $0.20 \text{ kg/m}^3/\text{d}$ [40]. Thus, it can be observed that a relatively high NPR value may not be conducive to obtaining a high NPE and vice versa. The reason for this may be the effect of HRT on the operating efficiency of the process; a shorter HRT will increase the NLR and contribute to a higher NPR, but a shorter HRT will result in incomplete oxidation of the ammonia and a significant reduction in the NPE [5,6,41]. Table 1 indicates that temperature can significantly affect NPE. Generally, the NPE increases as the temperature increases. Moreover, the influent ammonium concentration exerts a great impact on NPR and NPE. NPE and NPR can reach relatively high values even at a low temperature with high influent ammonium concentration [39,40,42]. The NPR in most nitrification processes is lower than $0.1 \text{ kg/m}^3/\text{d}$ due to the low ammonium concentration and low temperature of mainstream wastewater [43–45].

Table 1. Nitrite production ratio (NPR) and nitrite production efficiency (NPE) in recent pure nitrification process.

Wastewater	Temp.	pH	$\text{NH}_4^+ - \text{N}_{\text{inf}}$	DO	HRT	NAR	ARE	NPR	NPE	Refs.
	$^\circ\text{C}$									
Syn-wastewater	10	8	70	1.2	5.7	88.58	50	0.13	44.29	[46]
Syn-wastewater	20	8	47	0.42	-	-	-	0.04	40.10	[43]
Real wastewater	20.1	7.29–7.55	75	-	8	96.1	90.3	0.20	86.78	[40]
Syn-wastewater	25	-	37	8	20	70.38	96	0.03	67.57	[44]
Syn-wastewater	25	7.8–8.5	63	-	1	-	-	0.74	57.63	[39]
Real wastewater	25.5	7.1–7.4	70	-	3	93	60	0.31	55.8	[42]
Real wastewater	25.5	7.25	33.4	0.21	4	65.66	81.44	0.10	48.02	[41]
Syn-wastewater	30	8	100	1.5	8	55.43	98.5	0.16	54.6	[47]
Syn-wastewater	32	7.8–8	50	0.5–1	24	59.36	96.35	0.03	51.37	[4]
Syn-wastewater	33	7.5–8.1	50	-	8	48.59	87.4	0.06	38.14	[5]
Municipal wastewater	34	7.5–8.0	45	-	-	43.09	82.8	-	32.05	[5]
Syn-wastewater	35	8	50	-	2	31.98	93.8	0.18	30	[6]
Syn-wastewater	30–35	8–8.5	60	0.3	12	77.46	80	0.08	64.4	[48]

3.1.2. Microbial Information

Thus far, many kinds of AOB have been detected, as is shown in Table 2. Four genera, namely *Nitrosomonas*, *Nitrospira*, *Nitrosovibrio*, and *Nitrosolobus*, are common [49]. The dominant AOB is generally the *Nitrosomonas* genus. However, under some specific conditions, the main AOB genera of the nitrification process are altered. A study has reported that, in a nitrification process realised by polyvinyl alcohol gel beads, the *Ignavibacterium* genus is the major AOB [5]. Different from common AOB, *Ignavibacterium* is a chemoheterotroph with a versatile metabolism in the phylum *Chlorobi*. Other research has demonstrated that the *Nitrosomonadaceae Ellin6067* genus could play a pivotal role in inappropriate growth environments such as low temperature or light [13]. These recently detected genera are few. Hence, their main characteristics are not comprehensive and need to be further studied.

Table 2. The reported AOB species and their main features in pure nitrification.

AOB		Limit of NaCl Tolerance	Limit of Ammonia Tolerance at pH 7.8	Optimum			Main Characteristics
Genus	Species	%	mg/L	Temp. (°C)	pH	Salinity (%)	
<i>Nitrosomonas</i>	<i>Nitrosomonas europaea</i>	2.9	6800	25–30	7.5–8.0	-	Common in WWTPs.
	<i>Nitrosomonas aestuarii</i>	4.1	5100	30	7.5–8.0	1.8	Salt requirement. Common in marine and estuarine waters.
	<i>Nitrosomonas communis</i>	1.8	3400	30	7.5–8.0	-	Common in soils.
	<i>Nitrosomonas eutropha</i>	2.9	8500	30	7.5–8.0	-	Tolerance of increasing ammonia concentrations. Common in municipal and industrial sewage disposal systems.
	<i>Nitrosomonas halophilus</i>	5.9	6800	30	7.5–8.0	1.5	Salt requirement. Common in brackish water.
	<i>Nitrosomonas marine</i>	4.7	3400	30	7.5–8.0	2.3	Salt requirement. Common in marine waters and salt lakes.
	<i>Nitrosomonas mobilis</i>	3.5	5100	30	7.5–8.0	0.6	Found from brackish water environments and sewage disposal plants.
	<i>Nitrosomonas nitrosa</i>	1.2	1700	30	7.5–8.0	-	Found in eutrophic environments.
	<i>Nitrosomonas oligotropha</i>	1.2	850	30	7.5–8.0	-	Tend to be found in low ammonium concentration or poor-ammonia environments.
	<i>Nitrosomonas ureae</i>	1.8	1700	30	7.5–8.0	-	Tend to be found in low ammonium concentration and soils.
	<i>Nitrosomonas cryotolerans</i>	3.5	6800	22–30	7.0–8.5	1.8	Can survive at a low ammonia concentration. Common in marine environments.
		<i>Nitrosomonas sp. NP1</i>	1.8	3400	30	7.5–8.0	-
	<i>Nitrosomonas sp. Nm143</i>	-	-	30	7.5–8.0	-	Common in intermediate brackish sites and estuarine sediments.
<i>Ignavibacterium</i>		-	-	33–35	7.5–8.6	-	Heterotrophic nitrifying bacteria. It can be observed as polyvinyl alcohol gel beads.
<i>Nitrosococcus</i>	<i>Nitrosococcus halophilus</i>	9.4	10,200	30	7.5–8.0	3.5–4.7	The habitat in salt lakes.
	<i>Nitrosococcus oceani</i>	-	17,000	30	7.5–8.0	2.3–2.9	Grows only in seawater.
<i>Nitrospira</i>	<i>Nitrospira briensis</i>	-	-	25–30	7.5	-	Low growth rate and low abundance. Common in grasslands, heath, forest soils, and mountainous areas.
<i>Nitrosovibrio</i>	<i>Nitrosovibrio tenuis</i>	-	-	25–30	7.7–7.8	-	Grows slowly. Common in oligotrophic soils or natural soils.
<i>Nitrosolobus</i>	<i>Nitrosolobus multififormis</i>	-	-	25–30	7.5	-	Common in agricultural amended soils and freshwater.

3.1.3. Influencing Factors

1. Dissolved oxygen:

Different concentrations of DO can cause different degrees of effects on AOB and NOB activities. Taking advantage of this difference, strict control of DO concentration in the reaction process can effectively inhibit NOB and produce a better nitrosation effect [43,50]. Studies have reported that, when DO concentration is below 0.51 mgO₂/L in the biofilm reactor, there is a complete and long-term NOB suppression resulting in a low relative nitrate production ratio (10%) [43]. Additionally, for traditional aeration, the oxygen mass transfer rate is limited by its solubility. Altering aeration can enhance oxygen transfer, thus promoting the growth of AOB and its dominance in the microbial community [51]. Oxygen transfer rate can be greatly improved (18 times) by using the micro–nano aeration method, allowing for a significant increase in DO concentration [44]. It can also considerably increase the abundance of hydroxylamine oxidoreductase (hao) and ammonium monooxygenase (amoA) to enhance the activity of AOB. Generally, high-DO concentrations have an adverse impact on nitrification. In a one-stage PN/A process, a high DO concentration inhibits the growth of anaerobic ammonium-oxidizing bacteria (AnAOB), weakens the inhibition of NOB, and leads to nitrification failure [44]. However, in the treatment of industrial wastewater, toxic pollutants contained in the wastewater selectively inhibit the NOB activity and growth—similar to inhibitors that assist in achieving the nitrification process—allowing nitrification at higher DO concentrations and reducing the problem of maintaining low-DO concentrations. However, in the low concentration of ammonia wastewater, selective inhibitors do not exist and efficient operation cannot be achieved by relying on DO concentration control alone.

2. Sludge retention time:

SRT directly affects nitrification efficiency. The minimum doubling time of AOB is 7–8 h shorter than that of NOB (i.e., 10–13 h) [52]. Therefore, SRT adjustment can change the structure of the microbial community. Researches have proven that with a short SRT (<2 d), NOB can be flushed successfully [53]. The production of nitrite is also promoted by a short SRT [54]. When selecting an SRT with a minimum reproduction time higher than that of NOB, conditions such as DO are required to suppress NOB [46]. However, when the SRT approaches infinity or sludge is not discharged, DO, temperature, and other factors may not effectively inhibit NOB and other methods may need to be introduced for assistance.

3. Temperature:

The temperature of the reactor is also an important influencing factor, and its influence is reflected in two main aspects. First, temperature affects microorganism activity. Studies have demonstrated that ARE is only 50% at 10 °C but can exceed 90% after warming to 27 °C [45,46]. The reason for this is mainly that NOB have lower growth rates than AOB when the temperature exceeds 24 °C, but the growth rates of NOB will be faster when the temperature is below 15 °C, making it dominant in the microbial community [55]. Secondly, temperature affects FA and FNA concentrations. The concentration of FA in the same solution at 25 °C is one-sixth of that at 35 °C, however, the concentration of FNA at 25 °C is five times higher than that at 35 °C [56]. Different FA and FNA concentrations will have different effects on AOB and NOB, so the effect of temperature changes on the nitrification process is multifaceted. In real wastewater treatment, nitrification operating conditions are generally below 25 °C and it is difficult to achieve stable operation only in terms of the effect of temperature on microbial activity. It is also necessary to combine some other conditions of control in addition to FA and FNA for efficient operation.

4. Potential of hydrogen:

The effect of pH is similar to that of temperature, and it involves the activity of AOB and NOB and the equilibrium of FA and FNA. The *Nitrosomonas* and *Nitrobacter* genera are the most common AOB and NOB, respectively, in the nitrification process. For the *Nitrosomonas* genus, the optimum pH varies from 7.9 to 8.2, whereas for the *Nitrobacter*

genus it ranges from 7.2 to 7.6 [57]. Research has suggested that nitrite accumulation can be achieved at a pH greater than 7.5 [58]. Therefore, the general nitrification process will set the pH at about 7.8. The relationship amongst FA, FNA and pH is shown in Equations (11) and (12) [52,59].

$$FA(\text{mg/L}) = \frac{17}{14} \times \frac{NH_3-N \times 10^{pH}}{10^{pH} + \exp\left(\frac{6344}{273+T}\right)} \quad (11)$$

$$FNA(\text{mg/L}) = \frac{46}{14} \times \frac{NO_2-N}{10^{pH} \times \exp\left(-\frac{2300}{273+T}\right)} \quad (12)$$

5. Free nitrite acid:

Inhibition of NOB activity by FNA in wastewater is a common method. It was shown that the concentration of FNA in the range of 0.42–1.72 mgHNO₂-N/L reduced AOB activity by 50%, however, it could inhibit NOB in the range of 0.011–0.07 mgHNO₂-N/L and, in the range of 0.026–0.22 mg, HNO₂-N/L could completely inhibit NOB [60]. FNA during the nitrification process can cause NOB to be inhibited by the accumulation of nitrite production. The genera *Nitrospira* and *Nitrotoga* in NOB are not susceptible to the effect of FNA [33,61]; therefore the effect of FNA is not enough to completely inhibit NOB [43]. Additionally, the inhibitory effect of FNA on NOB is weakened for biofilm reactors. Even with a long treatment time of 24 h the biofilm will still protect NOB from inhibition [43]. Therefore, relying on FNA inhibition of NOB alone may not achieve efficient nitrification.

6. Free ammonium:

FA prevents the oxidation of ammonium and nitrite ions during the nitrification process. The inhibition of FA in nitrite oxidation by NOB begins at a concentration of 0.1–1.0 mgNH₃-N/L, while ammonium oxidation by AOB is inhibited when the concentration reaches the range of 10–150 mgNH₃-N/L FA [62]. Therefore, FA concentrations in the range of 1–10 mgNH₃-N/L were chosen to be suitable for nitrification. Research has reported that FA concentrations in the 5–10 mgNH₃-N/L range can effectively inhibit NOB and have no effect on the activity of AOB [63]. However, in general practical applications, FA concentrations higher than 10 mgNH₃-N/L are adopted to ensure complete NOB inhibition [64,65].

3.2. Assisted Nitrification Process

Several studies have attempted to improve the NPR to meet the requirements of the anammox reaction in recent years. Nitrification in these studies can be achieved through a number of methods including light, ultrasound, magnetic field, metal ions, salinity, and others. The main principles are shown in Figure 4 and the relevant data are in Table 3. The maximum NPR can reach 1.05 kg/m³/d at 25 °C–26 °C and 200 mgNH₄⁺-N/L, assisted by CuO NPs, with an NPE of 80% [66]. This value is significantly higher than that of other metals, and the study can largely satisfy the nitrite ion consumption rate of the anaerobic ammonia oxidation reaction. According to Table 3, HRT will significantly affect the NPR and a too-long HRT is not conducive to achieving a high NPR. Temperature is also an important parameter. The NPR and NPE will increase with temperature. This paper will present the factors influencing the assisted nitrification process through these 6 aspects.

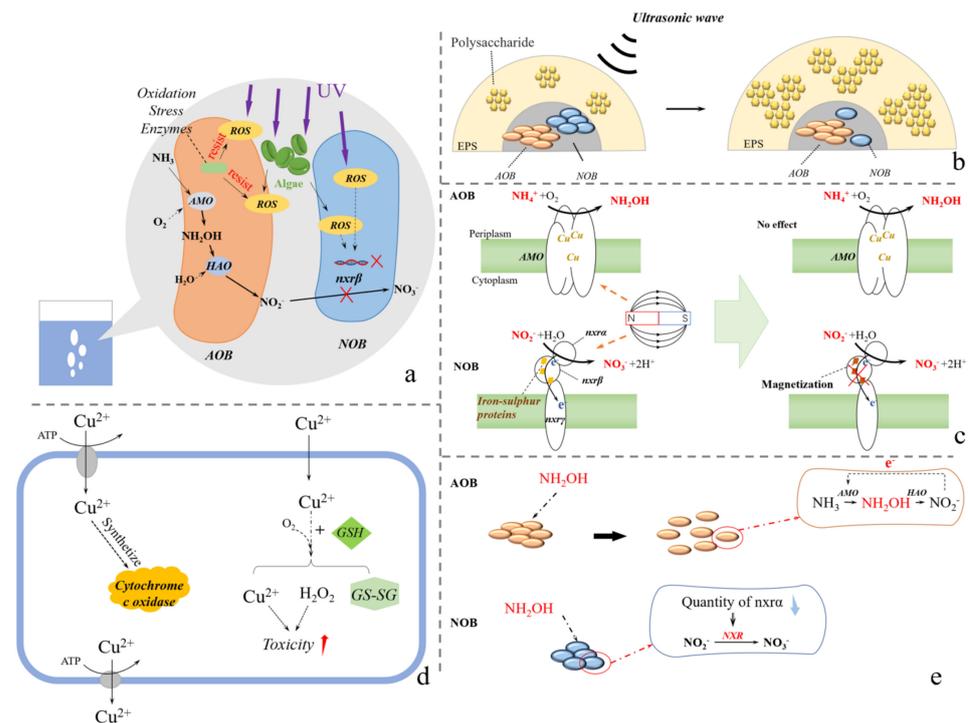


Figure 4. The principles of assisted nitrification process: (a) light-assisted nitrification process; (b) ultrasonic-assisted nitrification process; (c) magnetic field-assisted nitrification process; (d) copper ion-assisted nitrification process; and (e) hydroxylamine-assisted nitrification process.

Table 3. NPR and NPE in assisted nitrification processes.

Method	Wastewater	Condition	Temp.	pH	$\text{NH}_4^+ - \text{N}_{\text{inf}}$	DO	HRT	NAR	ARE	NPR	NPE	Refs.
			°C		mg/L	mg/L	h	%	%	kg/m ³ /d	%	
Light	Syn-wastewater	1000 $\mu\text{mol}/(\text{m}^2\text{s})$	25	7.4	30	1	3.85	70	100	0.13	70	[13]
	Syn-wastewater	0.87 $\mu\text{E}/(\text{Ls})$	25–28	7.4–7.6	50	0.5	6	80	100	0.10	48	[67]
	Syn-wastewater	0.25 W/mL	18	7.5–8.5	60	-	16	90	100	0.05	50	[68]
Ultrasound	Syn-wastewater	0.25 W/mL	25	8.1	60	-	16	90	95	0.08	85.5	[69]
	Syn-wastewater	0.15 W/ml	-	8	60	-	16	85	95	0.07	80.75	[70]
Magnetic field	Syn-wastewater	15 mT	25	7.5–8.0	100	-	12	90.1	95	0.12	60	[71]
Metal ion	Syn-wastewater	Cu^{2+} 0.61 mg/L	18.1	8	125	8.7	5	-	90	0.23	38.4	[72]
	Syn-wastewater	Y^{3+} 5 mg/L	23	8.9	150	-	-	-	70	0.30	70	[73]
	Syn-wastewater	Zn^{2+} 10 mg/L	23–26	8.05	200	0.1–0.15	12	-	90	0.30	75	[74]
	Syn-wastewater	CuO NPs 5 mg/L	25–26	7.7–7.8	200	0.2	3.67	-	80	1.05	80	[66]
	Syn-wastewater	La^{3+} 5 mg/L	-	8.9	150	0.2	-	-	100	0.37	90	[75]
Salinity	Industrial wastewater	6.6 gNaCl/L	29	-	161	0.1–1.5	48.24	61.6	76.9	0.03	35.80	[76]
	Syn-wastewater	10 gNaCl/L	30	7.5–8.0	100	-	8	-	60	0.15	48.62	[14]
	Industrial wastewater	8.6 gNaCl/L	31	7.6	220	0.5–3.5	31.2	80	90	0.08	47	[77]
	Syn-wastewater	13.5 gNaCl/L	32	8	150	0.3	2.4	73.3	-	1.05	70	[78]
Others	Domestic sewage	NH_2OH 5 mg/L	19.5–28.2	7.9	70	3	3	-	100	0.28	50	[79]
	Syn-wastewater	PNCT 8 mg/L	20	7.50–7.80	70	-	6	94.94	-	0.23	81.1	[80]
	Syn-wastewater	formic acid 1380 mg/L	25	7.9	127.1	2	8.6	-	94.6	0.33	94.32	[81]
	Syn-wastewater	NH_2OH 10 mg/L	25	7.8–8.2	100	5	8	-	57	0.17	56.9	[82]
	Syn-wastewater	N_2H_4 2–5 mg/L	30	8	30	0.09–0.25	2	82	90	0.18	51.2	[83]
	Syn-wastewater	N_2H_4 2–5 mg/L	30	-	42	0.3	2.32	82.9	97.4	0.23	52.2	[84]
	Syn-wastewater	formic acid 1380 mg/L	-	7.9	120	-	4	-	95	0.62	86.45	[85]

3.2.1. Light

Light affects the growth and metabolism of some microorganisms in water. Previous studies have demonstrated that photosensitivity differences exist between AOB and NOB, with NOB being more sensitive to light than AOB [13,86]. This principle is shown in Figure 4a. Reactive oxygen species (ROS) can be generated in wastewater under light and can indirectly lead to a series of damages, mainly the oxidative damage of cell proteins, cell membranes, and nucleic acids [87,88]. However, AOB have a critical oxidation stress enzyme that can be used to protect themselves, while NOB do not [89,90]. Therefore, NOB will be affected by light, mainly in the form of NOB *nxrβ* gene un-expression and irreversible inhibition of NOB activity. Meanwhile, under light irradiation, a large amount of algae is produced in nitrification and forms a symbiotic system with AOB [67]. The algae also produce ROS, so this method can effectively inhibit NOB [91].

The effect of light intensity on NPE is very obvious. When the strength exceeds 300 $\mu\text{mol}/(\text{m}^2 \cdot \text{s})$, AOB were inhibited and nitrification fails [92]. Within a small range of light intensities, AOB activity initially increases and then decreases with an increase in light strength, whereas NOB activity directly decreases. Therefore, an optimum light intensity will be available to maximize the promotion of AOB and inhibit NOB, allowing nitrification to operate efficiently. Studies have observed a 1.31–1.43-fold increase in AOB activity at 25 °C under optimal conditions ($p = 0.03\text{--}0.08$ kJ/mgVSS) [13]. Moreover, the photoinhibition effect of AOB and NOB and the light time have a direct ratio [13]. Therefore, selecting light-assisted nitrification for long-term treatment can continuously inhibit the effect of NOB and make nitrite ions more likely to accumulate.

3.2.2. Ultrasonic

Research has found that low-frequency ultrasound can accelerate microbial growth by increasing the abundance of functional genes to promote signal transduction, cell movement, and membrane transport [68,93,94]. Differences in the effects of low-frequency ultrasound on AOB and NOB exist, and ultrasound at certain frequencies and intensities can promote the oxidation process of ammonia by AOB while inhibiting the oxidation of nitrite ions by NOB. During ultrasound-assisted nitrification process, an increase in extracellular polymer (EPS) content, especially a significant increase in polysaccharide (PS) content, was observed (shown in Figure 4b). EPSs have a positive effect on the efficiency of the mass transfer and the activity of AOB while inhibiting NOB activity [70]. Therefore, nitrification assisted by low-frequency ultrasound can be feasible.

The effect of low-frequency ultrasound intensity is similar to that of light. A suitable range also exists to increase the AOB activity and decrease the NOB activity [93]. Thus, this method is promising for use in mainstream wastewater treatment. Moreover, the temperature in the reactor can be considerably improved through ultrasound, weakening the inhibitory effect of low temperature on AOB. Generally, nitrification operations at 18 °C may fail according to the previous paper, but ultrasound-assisted nitrification can realise more than 90% of NAR [68]. Hence, the application of ultrasound can provide a good temperature environment for mainstream treatment [95].

3.2.3. Magnetic Field

Generally, microorganisms are magnetic to some extent. Thus, under an external magnetic field, enzyme activity and cell membrane permeability are affected to a certain extent [96,97]. Subsequently, the microbial metabolism also becomes affected. Nitrite oxidoreductase (*nxr*), which oxidizes nitrite to nitrate in NOB, can recruit iron–sulphur proteins to realise redox reaction [98], as is shown in Figure 4c. Iron is magnetised by the static magnetic field (SMF), but the active site of ammonium monooxygenase (AMO) in AOB is copper, which is not magnetised. Therefore, SMF only inhibits NOB [71]. The SMF makes the sludge gather closely and form an oxygen concentration gradient so that AOB are easily enriched while NOB are washed off [99]. The strength of the SMF also has an appropriate range to realise nitrification. Within this range, the DO utilisation rate is

improved, nitrification is achieved faster than through pure biological nitrification, and the DO concentration is maintained at 0.5–0.7 mg/L at 35 °C [100]. Therefore, the magnetic field-assisted method is recommended for actual applications.

3.2.4. Metal Ions

Metal ions are indispensable elements for microbial growth and metabolism. As shown in Figure 4d, copper ions play a considerable role in cytochrome c oxidase and related enzymes that participate in cell respiration. Generally, copper ions enter cells through active and passive transport [101]. However, copper ions enter cells rapidly and not specifically through passive transport at a high periplasm concentration. The copper ions' concentration in the cytoplasm then increases and the copper ions promote the reaction with glutathione (GSH) and oxygen to produce oxidized disglutathione (GS-SG), copper ions, and H₂O₂ [102]. Copper ions are not removed, and H₂O₂ increases toxicity. This may cause gene mutation and reduce the growth rate. Other metal ions have different effects on microorganisms; for example, Mn²⁺ can promote the respiration of AnAOB [103], and Zn²⁺ can be used for the synthesis of some zinc-containing enzymes [103].

Generally, metal ions are important co-factors of metalloproteinases, and some enzymes have a low metal ion concentration [101]. However, at a high concentration metal ions chemically combine with some enzymes and affect the structures and activity of these enzymes, finally inhibiting microbial activity [104]. For example, at 1 mmol/L concentration, Cu²⁺ effectively inhibits the activity of AOB and NOB at 25 °C [105]. Therefore, the promotion or inhibition of AOB and NOB depends on the exposure dose [73]. Most studies have reported that an increase in dosage initially increases the activity of AOB to the peak then gradually decreases it [66,73–75]. However, the activity of NOB decreases directly. Therefore, a suitable dose range exists to achieve nitrification.

3.2.5. Salinity

For mainstream wastewater, high salinity greatly increases the difficulty of denitrification. The main reason for this is that high salinity may inhibit microbial enzymes and metabolism and decrease cell activity [106,107]. Furthermore, high salinity affects the osmotic pressure of cells and even leads to plasmolysis [78]. Studies have indicated that AOB activity is inhibited under high salinity (25 gNaCl/L) at 35 °C [108]. However, at a low concentration (13.5 gNaCl/L), salt can promote the growth of AOB at 32 °C, such as in *Unclassified Nitrosomonadaceae* [78]. By adding salt (5.2, 7.6 and 10.2 gNaCl/L), the NAR can increase to above 95% at 22 °C and 36.2 mgNH₄⁺-N/L [109]. After high salinity acclimation, AOB and NOB show increased activity [110,111]. Thus, a low NPE may be obtained, but the NPR may increase [34,112]. Low NPE makes this method impossible to use alone.

3.2.6. Others

In addition to the categories provided above, other substances can inhibit NOB. These include hydroxylamine, hydrazine, formic acid, and some antibiotics. Hydroxylamine is the intermediate product of nitrification and a concentration of 10 mg/L can make nitrification highly stable at 25 °C and 100 mgNH₄⁺-N/L [82]. As shown in Figure 4e, hydroxylamine disperses cells to improve the mass transfer efficiency for AOB. Abundant hydroxylamine promotes the production of nitrite, which produces many electrons to circulate to the AMO enzyme and increase the consumption of ammonia [113]. However, it may inhibit the growth of NOB by inhibiting the induction of nxr and decreasing the nxr α enzyme [82,114]. Therefore, hydroxylamine can be selected to assist in nitrification and this method can obtain high efficiency [115].

Hydrazine, being an intermediate product of anammox, can also promote nitrification. Exogenous hydrazine can be partly converted into hydroxylamine to inhibit NOB, and another part causes hydroxylamine to be produced disproportionately during nitrification to reduce the effect on AOB [83]. Hydrazine can also improve AnAOB activity and can

be completely consumed without extra pollution, thereby making hydrazine a frequently selected inhibitor [84]. Studies have indicated that formic acid can effectively and selectively inhibit *nxrβ* gene transcription, leading to the inhibition of nitrite oxidation and promotion of nitrite accumulation [81,116]. Therefore, formic acid-assisted nitritation is also a suitable method. As a common antibiotic, PNCT can often be detected in wastewater [117]. PNCT can inhibit AOB and NOB through DNA and protein damage and destruction of the carbon fixation pathway [80]. However, the abundance of *Nitrosomonas* is always higher than that of *Nitrospira*, so AOB is always dominant in the reactor. Therefore, the PNCT-assisted method can also achieve stable nitritation.

4. N₂O Emission

As a major greenhouse gas in WWTPs, N₂O emissions weaken the advantage and efficiency of nitritation-based nitrogen removal processes due to nitrite consumption and environmental pollution. N₂O is produced in nitritation unit via two pathways: NH₂OH oxidation, and nitrifier denitrification by AOB [118,119]. The first case refers to the incomplete oxidation of NH₂OH produced in the nitritation N₂O. The other pathway describes how NO₂[−] is reduced by AOB, as shown in Figure 5. Nitrifier denitrification is generally considered the main pathway in nitritation [15]. N₂O emission is affected by DO and nitrite concentration in the nitritation unit. The denitrification by AOB is stimulated and the N₂O emission increases when DO is less than 0.5 mg/L at 27 °C with 600 mgNH₄⁺-N/L [120], whereas nitritation will be promoted and N₂O emission will be reduced with the increase of DO concentration [121]. The range (DO < 0.5 mg/L) is generally suitable to inhibit NOB and realise pure nitritation. Therefore, there is a great risk of N₂O emission at low-DO concentration in pure nitritation. The research demonstrated that nitrite concentration and N₂O emission are in a direct ratio [15]. In a one-stage process, nitrite is generated and consumed at the same time. In addition, the nitrogen removal rate of anammox in mainstream wastewater is greater than 1.2 kg/m³/d; that is, nitrite consumption rate is greater than 0.70 kg/m³/d [3]. The NPR is significantly lower than nitrite consumption rate according to Table 1. Therefore, nitrite cannot theoretically accumulate to produce N₂O in a one-stage process, and a one-stage process is more promising than a two-stage process. For photocatalytic oxidation, the NH₂OH produced by ammonia oxidation will be directly oxidized to nitrite under anaerobic conditions due to the strong oxidation of hydroxyl radicals [9]. The reduction of nitrite can be neglected under the condition of NH₂OH existence. Therefore, nitrite production through photocatalytic oxidation is also environmentally friendly.

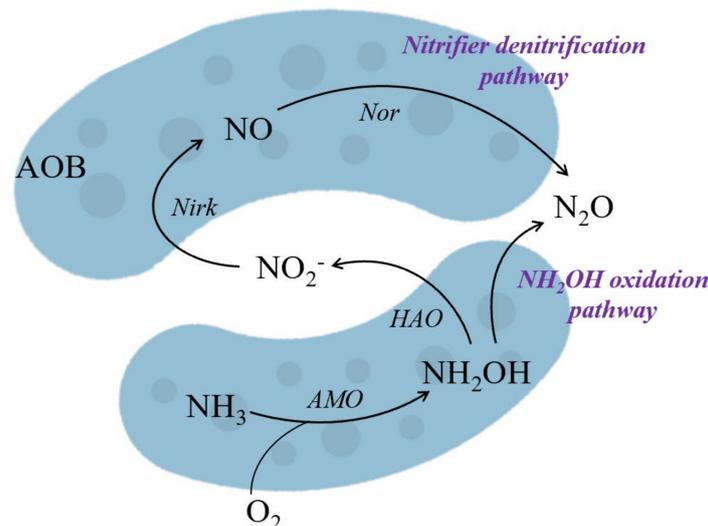


Figure 5. N₂O emission pathways in nitritation processes. N₂O can be emitted through nitrifier denitrification and the NH₂OH oxidation pathway.

5. Existing Problems

According to the above, photocatalytic oxidation is the only physicochemical method that can realize nitritation at present. However, this method is still in the lab-scale and has not been used in practice [25]. In its actual application, the chromaticity and turbidity of wastewater will significantly affect the efficiency. In addition, most of current studies tend to oxidize ammonium directly to nitrogen to reduce subsequent treatment [122]. This direction seems to be more environmentally friendly, but the treatment efficiency is poor. This case also results in less existing research and that the used catalysts are incomplete for nitrite production. Therefore, further research is needed in this direction.

There are some unresolved questions regarding these nitritation processes assisted by various methods. Light, ultrasonic, and magnetic field methods are more susceptible to fluctuations in water quality and quantity. For example, the density and morphology of sludge will have a significant impact on the shading for light-assisted nitritation [13]. Additionally, use of light, ultrasonic, and magnetic field methods will increase energy consumption, so it makes sense to use less energy to achieve better results in the practical application of these methods. Metal ions and salinity are not suitable to assist in nitritation alone given that long-term treatment of metal ions will increase the inhibition concentration and that there will still be a small amount of *Nitrospira* restoring activity when the salinity drops to a low concentration, even if the salt concentration is very high [77]. Cost of operation should also be considered. A higher concentration is required to more effectively inhibit NOB, but it will increase the cost. Organic compounds are frequently used for selective inhibitors. However, these inhibitors have some adverse other effects: hydroxylamine has unstable chemical properties, limited inhibition of Nitrobacter, and high chemical cost; hydrazine will inhibit AnAOB under mainstream conditions; formic acid is corrosive and difficult to store; and when antibiotics are incompletely removed they will aggravate the pollution of water quality [79,80,123]. Moreover, treatment of selective inhibitors will increase the operation cost and affect microorganisms other than NOB for the long term [124].

Given that photocatalytic oxidation has undeniable advantages, a two-stage PN/A process that combines photocatalytic oxidation for nitrite production and anammox for nitrogen removal can be designed for the follow-up treatment of the anaerobic digestion effluent of mainstream wastewater. The process flow is shown in Figure 6. First, wastewater will be treated in the COD removal unit before it enters the photocatalytic oxidation unit after passing through the setting tank and undergoing pH adjustment. The requirements of the anammox reaction will be met in this unit. Second, nitrogen will be removed in the anammox unit and water will be discharged after sedimentation. Unlike in the above-mentioned processes, the NPR of photocatalytic oxidation is higher and can meet the requirement of the anammox reaction at 21 °C and 100 mgNH₄⁺-N/L. This process does not require aeration and does not produce sludge in photocatalytic oxidation, so operating costs will be significantly reduced. As this process does not consume chemicals, the effects on the subsequent anammox process, drug storage and environmental pollution need not be considered.

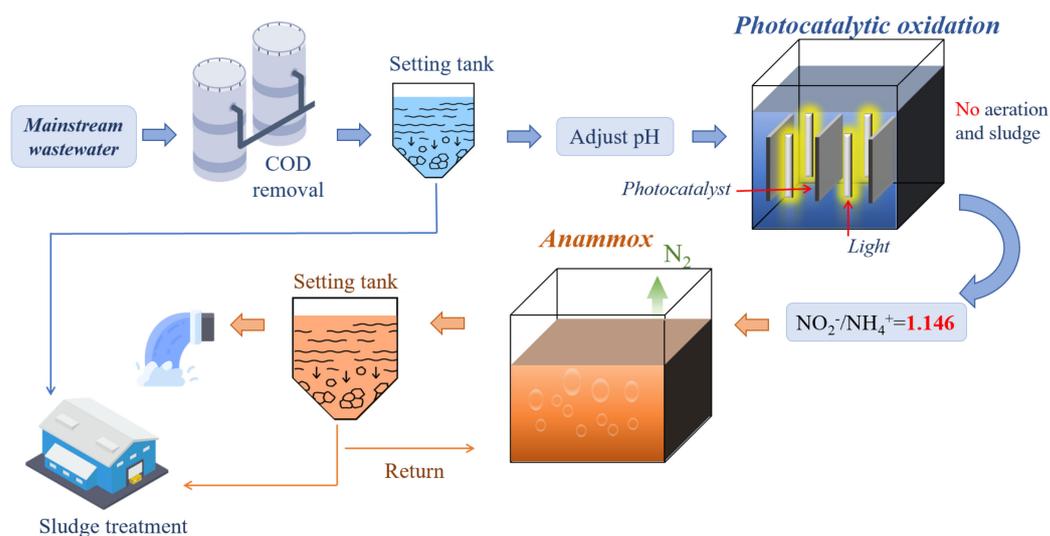


Figure 6. The novel two-stage nitrogen removal process.

6. Conclusions

This paper comprehensively summarizes nitrite production processes. The microbial main species and characteristics involved in biological nitrification processes are also reviewed. The principles of the various assisted nitrification processes and the influencing factors present are described in detail. A comparison of the studies reveals that the efficiency of nitrification is strongly influenced by HRT and temperature; the NPR increases as HRT shortens, while the NPE increases with increasing temperature. This provides a reference for the practical application of the PNA process. In addition, the N_2O emission in the nitrite production process is analysed, and a one-stage nitrification-based process and photocatalytic oxidation process are environmentally friendly. Finally, a two-stage process that combines photocatalytic oxidation and anammox is proposed. This process has a good prospect for development under mainstream conditions.

Author Contributions: H.Z. and Y.G. have reviewed the literature, summarized the information, prepared the tables and figures, and drafted the manuscript. F.L. and Z.Z. contributed to the designed figures in this work. C.W. and Q.W. contributed to the review and revision of the manuscript. M.G. planned and supervised the work. All authors have read and agreed to the published version of the manuscript.

Funding: This work was supported by the National Key R&D Program of China (2019YFC1906304; 2019YFC1906302), and the National Environmental and Energy Base for International Science & Technology Cooperation.

Institutional Review Board Statement: Not applicable.

Informed Consent Statement: Not applicable.

Data Availability Statement: The datasets generated or analysed during the current study are available in the Web of Science repository [<http://apps.webofknowledge.com/> accessed on 1 June 2022].

Conflicts of Interest: The authors declare no conflict of interest.

Abbreviations

WWPTs: wastewater treatment plants; NPR: nitrite production rate; NPE: nitrite production efficiency; PN/A: partial nitrification/anammox; NRR: nitrogen removal rate; AOB: ammonium-oxidizing bacteria; NOB: nitrite-oxidizing bacteria; DO: dissolved oxygen; FA: free ammonium; FNA: free nitrite acid; N_2O : Nitrous oxide; SRT: sludge retention time; HRT: hydraulic retention time; NAR: nitrite accumulation ratio; hao: hydroxylamine oxidoreductase; AMO: ammonium monooxygenase; amoA: monooxygenase; AnAOB: anaerobic ammonium-oxidizing bacteria; ROS: reactive oxygen species;

EPS: extracellular polymer; PS: polysaccharide; nxr: nitrite oxidoreductase; SMF: static magnetic field; GSH: glutathione; GS-SG: disglutathione; ARE: ammonium removal efficiency.

References

- Lackner, S.; Gilbert, E.M.; Vlaeminck, S.E.; Joss, A.; Horn, H.; van Loosdrecht, M.C. Full-scale partial nitrification/anammox experiences—An application survey. *Water Res.* **2014**, *55*, 292–303. [[CrossRef](#)] [[PubMed](#)]
- Yang, E.; Chen, J.; Jiang, Z.; Deng, Z.; Tu, Z.; Wang, H.; Wu, S.; Kong, Z.; Hendrik Sanjaya, E.; Chen, H. Insights into rapidly recovering the autotrophic nitrogen removal performance of single-stage partial nitrification-anammox systems: Reconstructing granular sludge and its functional microbes synergy. *Bioresour. Technol.* **2022**, *361*, 127750. [[CrossRef](#)] [[PubMed](#)]
- Guo, Y.; Chen, Y.; Webeck, E.; Li, Y.Y. Towards more efficient nitrogen removal and phosphorus recovery from digestion effluent: Latest developments in the anammox-based process from the application perspective. *Bioresour. Technol.* **2020**, *299*, 122560. [[CrossRef](#)] [[PubMed](#)]
- Wang, X.; Wang, T.; Yuan, L.; Xing, F. One-step start-up and subsequent operation of CANON process in a fixed-bed reactor by inoculating mixture of partial nitrification and Anammox sludge. *Chemosphere* **2021**, *275*, 130075. [[CrossRef](#)] [[PubMed](#)]
- Wang, J.; Liang, J.; Sun, L.; Shen, J.; Wang, M. Achieving reliable partial nitrification and anammox process using polyvinyl alcohol gel beads to treat low-strength ammonia wastewater. *Bioresour. Technol.* **2021**, *324*, 124669. [[CrossRef](#)] [[PubMed](#)]
- Chen, H.; Wang, H.; Yu, G.; Xiong, Y.; Wu, H.; Yang, M.; Chen, R.; Yang, E.; Jiang, C.; Li, Y.Y. Key factors governing the performance and microbial community of one-stage partial nitrification and anammox system with bio-carriers and airlift circulation. *Bioresour. Technol.* **2021**, *324*, 124668. [[CrossRef](#)]
- Ren, H.-T.; Liang, Y.; Han, X.; Liu, Y.; Wu, S.-H.; Bai, H.; Jia, S.-Y. Photocatalytic oxidation of aqueous ammonia by Ag₂O/TiO₂ (P25): New insights into selectivity and contributions of different oxidative species. *Appl. Surf. Sci.* **2020**, *504*, 144433. [[CrossRef](#)]
- Altomare, M.; Chiarello, G.L.; Costa, A.; Guarino, M.; Selli, E. Photocatalytic abatement of ammonia in nitrogen-containing effluents. *Chem. Eng. J.* **2012**, *191*, 394–401. [[CrossRef](#)]
- Shibuya, S.; Aoki, S.; Sekine, Y.; Mikami, I. Influence of oxygen addition on photocatalytic oxidation of aqueous ammonia over platinum-loaded TiO₂. *Appl. Catal. B* **2013**, *138–139*, 294–298. [[CrossRef](#)]
- Van Hulle, S.W.; Vandeweyer, H.J.; Meesschaert, B.D. Engineering aspects and practical application of autotrophic nitrogen removal from nitrogen rich streams. *Chem. Eng. J.* **2010**, *162*, 1–20. [[CrossRef](#)]
- Law, Y.; Ye, L.; Wang, Q.; Hu, S.; Pijuan, M.; Yuan, Z. Producing free nitrous acid—A green and renewable biocidal agent—From anaerobic digester liquor. *Chem. Eng. J.* **2014**, *259*, 62–69. [[CrossRef](#)]
- Bartroli, A.; Perez, J.; Carrera, J. Applying Ratio Control in a Continuous Granular Reactor to Achieve Full Nitrification under Stable Operating Conditions. *Environ. Sci. Technol.* **2010**, *44*, 8930–8935. [[CrossRef](#)]
- Wang, L.; Qiu, S.; Guo, J.; Ge, S. Light Irradiation Enables Rapid Start-Up of Nitrification through Suppressing nxrB Gene Expression and Stimulating Ammonia-Oxidizing Bacteria. *Environ. Sci. Technol.* **2021**, *55*, 13297–13305. [[CrossRef](#)]
- Liu, C.; Yu, D.; Wang, Y.; Chen, G.; Tang, P.; Huang, S. A novel control strategy for the partial nitrification and anammox process (PN/A) of immobilized particles: Using salinity as a factor. *Bioresour. Technol.* **2020**, *302*, 122864. [[CrossRef](#)]
- Wunderlin, P.; Mohn, J.; Joss, A.; Emmenegger, L.; Siegrist, H. Mechanisms of N₂O production in biological wastewater treatment under nitrifying and denitrifying conditions. *Water Res.* **2012**, *46*, 1027–1037. [[CrossRef](#)]
- Wrage, N.; Velthof, G.L.; Van Beusichem, M.L.; Oenema, O. Role of nitrifier denitrification in the production of nitrous oxide. *Soil Biol. Biochem.* **2001**, *33*, 1723–1732. [[CrossRef](#)]
- Chen, T.-L.; Chen, L.-H.; Lin, Y.J.; Yu, C.-P.; Ma, H.-w.; Chiang, P.-C. Advanced ammonia nitrogen removal and recovery technology using electrokinetic and stripping process towards a sustainable nitrogen cycle: A review. *J. Clean. Prod.* **2021**, *309*, 127369. [[CrossRef](#)]
- Wang, H.; Gui, H.; Yang, W.; Li, D.; Tan, W.; Yang, M.; Barrow, C.J. Ammonia nitrogen removal from aqueous solution using functionalized zeolite columns. *Desalin. Water Treat.* **2013**, *52*, 753–758. [[CrossRef](#)]
- Pressley, T.A.; Bishop, D.F.; Roan, S.G. Ammonia-nitrogen removal by breakpoint chlorination. *Environ. Sci. Technol.* **1972**, *6*, 622–628. [[CrossRef](#)]
- Gaya, U.I.; Abdullah, A.H. Heterogeneous photocatalytic degradation of organic contaminants over titanium dioxide: A review of fundamentals, progress and problems. *J. Photochem. Photobiol. C* **2008**, *9*, 1–12. [[CrossRef](#)]
- Ikeda, K.; Sakai, H.; Baba, R.; Hashimoto, K.; Fujishima, A. Photocatalytic Reactions Involving Radical Chain Reactions Using Microelectrodes. *J. Phys. Chem. B* **1997**, *101*, 2617–2620. [[CrossRef](#)]
- Lee, J.; Park, H.; Choi, W. Selective photocatalytic oxidation of NH₃ to N₂ on platinumized TiO₂ in water. *Environ. Sci. Technol.* **2002**, *36*, 5462. [[CrossRef](#)] [[PubMed](#)]
- Zhu, X.; Castleberry, S.R.; Nanny, M.A.; Butler, E.C. Effects of pH and Catalyst Concentration on Photocatalytic Oxidation of Aqueous Ammonia and Nitrite in Titanium Dioxide Suspensions. *Environ. Sci. Technol.* **2005**, *39*, 3784–3791. [[CrossRef](#)] [[PubMed](#)]
- Zhou, W.; Liu, H.; Wang, J.; Liu, D.; Du, G.; Cui, J. Ag₂O/TiO₂ nanobelts heterostructure with enhanced ultraviolet and visible photocatalytic activity. *ACS Appl. Mater. Interfaces* **2010**, *2*, 2385–2392. [[CrossRef](#)] [[PubMed](#)]
- Zhang, G.; Ruan, J.; Du, T. Recent Advances on Photocatalytic and Electrochemical Oxidation for Ammonia Treatment from Water/Wastewater. *ACS ES&T Engg* **2020**, *1*, 310–325. [[CrossRef](#)]

26. Wang, H.; Su, Y.; Zhao, H.; Yu, H.; Chen, S.; Zhang, Y.; Quan, X. Photocatalytic oxidation of aqueous ammonia using atomic single layer graphitic-C₃N₄. *Environ. Sci. Technol.* **2014**, *48*, 11984–11990. [[CrossRef](#)]
27. Altomare, M.; Selli, E. Effects of metal nanoparticles deposition on the photocatalytic oxidation of ammonia in TiO₂ aqueous suspensions. *Catal. Today* **2013**, *209*, 127–133. [[CrossRef](#)]
28. Xiao, B.; Liu, S.-Q. Photocatalytic Oxidation of Ammonia via an Activated Carbon-Nickel Ferrite Hybrid Catalyst under Visible Light Irradiation. *Acta Phys.-Chim. Sin.* **2014**, *30*, 1697–1705. [[CrossRef](#)]
29. Ma, B.; Xu, X.; Ge, S.; Li, B.; Wei, Y.; Zhu, H.; Nan, X.; Peng, Y. Reducing carbon source consumption through a novel denitrification/anammox biofilter to remove nitrate from synthetic secondary effluent. *Bioresour. Technol.* **2020**, *309*, 123377. [[CrossRef](#)]
30. Moomen, S.; Ahmed, E. Development of partial nitrification as a first step of nitrite shunt process in a Sequential Batch Reactor (SBR) using Ammonium Oxidizing Bacteria (AOB) controlled by mixing regime. *Bioresour. Technol.* **2016**, *221*, 85–95. [[CrossRef](#)]
31. Guo, Y.; Luo, Z.; Shen, J.; Li, Y.Y. The main anammox-based processes, the involved microbes and the novel process concept from the application perspective. *Front. Environ. Sci. Eng.* **2022**, *16*, 84. [[CrossRef](#)]
32. Zaman, M.; Kim, M.; Nakhla, G. Simultaneous partial nitrification and denitrifying phosphorus removal (PNDPR) in a sequencing batch reactor process operated at low DO and high SRT for carbon and energy reduction. *Chem. Eng. J.* **2021**, *425*, 131881. [[CrossRef](#)]
33. Duan, H.; Ye, L.; Lu, X.; Yuan, Z. Overcoming nitrite oxidizing bacteria adaptation through alternating sludge treatment with free nitrous acid and free ammonia. *Environ. Sci. Technol.* **2019**, *53*, 1937–1946. [[CrossRef](#)]
34. Guo, Y.; Sugano, T.; Song, Y.; Xie, C.; Chen, Y.; Xue, Y.; Li, Y.Y. The performance of freshwater one-stage partial nitrification/anammox process with the increase of salinity up to 3.0. *Bioresour. Technol.* **2020**, *311*, 123489. [[CrossRef](#)]
35. Choi, M.; Chaudhary, R.; Lee, M.; Kim, J.; Cho, K.; Chung, Y.C.; Bae, H.; Park, J. Enhanced selective enrichment of partial nitrification and anammox bacteria in a novel two-stage continuous flow system using flat-type poly (vinylalcohol) cryogel films. *Bioresour. Technol.* **2020**, *300*, 122546. [[CrossRef](#)]
36. Jin, P.; Li, B.; Mu, D.; Li, X.; Peng, Y. High-efficient nitrogen removal from municipal wastewater via two-stage nitrification/anammox process: Long-term stability assessment and mechanism analysis. *Bioresour. Technol.* **2019**, *271*, 150–158. [[CrossRef](#)]
37. Chen, J.; Zeng, J.; He, Y.; Sun, S.; Wu, H.; Zhou, Y.; Chen, Z.; Wang, J.; Chen, H. Insights into a novel nitrogen removal process based on simultaneous anammox and denitrification (SAD) following nitrification with in-situ NOB elimination. *J. Environ. Sci.* **2023**, *125*, 160–170. [[CrossRef](#)]
38. Gao, Z.; Ma, Y.; Liu, Y.; Wang, Q. Waste cooking oil used as carbon source for microbial lipid production: Promoter or inhibitor. *Environ. Res.* **2022**, *203*, 111881. [[CrossRef](#)]
39. Guo, Y.; Xie, C.; Chen, Y.; Urasaki, K.; Qin, Y.; Kubota, K.; Li, Y.Y. Achieving superior nitrogen removal performance in low-strength ammonium wastewater treatment by cultivating concentrated, highly dispersive, and easily settleable granule sludge in a one-stage partial nitrification/anammox-HAP reactor. *Water Res.* **2021**, *200*, 117217. [[CrossRef](#)]
40. Li, S.; Li, J.; Yang, S.; Zhang, Q.; Li, X.; Zhang, L.; Peng, Y. Rapid achieving partial nitrification in domestic wastewater: Controlling aeration time to selectively enrich ammonium oxidizing bacteria (AOB) after simultaneously eliminating AOB and nitrite oxidizing bacteria (NOB). *Bioresour. Technol.* **2021**, *328*, 124810. [[CrossRef](#)]
41. Rong, C.; Luo, Z.; Wang, T.; Guo, Y.; Kong, Z.; Wu, J.; Ji, J.; Qin, Y.; Hanaoka, T.; Sakemi, S.; et al. Chemical oxygen demand and nitrogen transformation in a large pilot-scale plant with a combined submerged anaerobic membrane bioreactor and one-stage partial nitrification-anammox for treating mainstream wastewater at 25 degrees C. *Bioresour. Technol.* **2021**, *341*, 125840. [[CrossRef](#)] [[PubMed](#)]
42. Wang, Z.; Peng, Y.; Li, J.; Liu, J.; Zhang, Q.; Li, X.; Zhang, L. Rapid initiation and stable maintenance of municipal wastewater nitrification during the continuous flow anaerobic/oxic process with an ultra-low sludge retention time. *Water Res.* **2021**, *197*, 117091. [[CrossRef](#)] [[PubMed](#)]
43. Van Tendeloo, M.; Xie, Y.; Van Beeck, W.; Zhu, W.; Lebeer, S.; Vlaeminck, S.E. Oxygen control and stressor treatments for complete and long-term suppression of nitrite-oxidizing bacteria in biofilm-based partial nitrification/anammox. *Bioresour. Technol.* **2021**, *342*, 125996. [[CrossRef](#)] [[PubMed](#)]
44. Yao, G.J.; Ren, J.Q.; Zhou, F.; Liu, Y.D.; Li, W. Micro-nano aeration is a promising alternative for achieving high-rate partial nitrification. *Sci. Total Environ.* **2021**, *795*, 148899. [[CrossRef](#)]
45. Li, Z.; Wei, C.; Chen, Y.; Chen, B.; Qiu, G.; Wan, J.; Wu, H.; Zhu, S.; Zhao, H. Achieving nitrification in an aerobic fluidized reactor for coking wastewater treatment: Operation stability, mechanisms and model analysis. *Chem. Eng. J.* **2021**, *406*, 126816. [[CrossRef](#)]
46. Reino, C.; Carrera, J. Impact of the nitrifying community dynamics on the partial nitrification process performed by an AOB-enriched culture in a granular sludge airlift reactor. *J. Environ. Chem. Eng.* **2021**, *9*, 106691. [[CrossRef](#)]
47. Huang, T.; Zhao, J.; Wang, S.; Lei, L. Fast start-up and enhancement of partial nitrification and anammox process for treating synthetic wastewater in a sequencing batch biofilm reactor: Strategy and function of nitric oxide. *Bioresour. Technol.* **2021**, *335*, 125225. [[CrossRef](#)]
48. Huff Chester, A.L.; Eum, K.; Tsapatsis, M.; Hillmyer, M.A.; Novak, P.J. Enhanced Nitrogen Removal and Anammox Bacteria Retention with Zeolite-Coated Membrane in Simulated Mainstream Wastewater. *Environ. Sci. Technol. Lett.* **2021**, *8*, 468–473. [[CrossRef](#)]

49. Chen, H.; Liu, K.; Yang, E.; Chen, J.; Gu, Y.; Wu, S.; Yang, M.; Wang, H.; Wang, D.; Li, H. A critical review on microbial ecology in the novel biological nitrogen removal process: Dynamic balance of complex functional microbes for nitrogen removal. *Sci. Total Environ.* **2023**, *857*, 159462. [[CrossRef](#)]
50. Wang, H.; Yu, G.; He, W.; Du, C.; Deng, Z.; Wang, D.; Yang, M.; Yang, E.; Zhou, Y.; Sanjaya, E.H.; et al. Enhancing autotrophic nitrogen removal with a novel dissolved oxygen-differentiated airlift internal circulation reactor: Long-term operational performance and microbial characteristics. *J. Environ. Manag.* **2021**, *296*, 113271. [[CrossRef](#)]
51. Cao, Y.; Loosdrecht, M.; Daigger, G.T. Mainstream partial nitrification–anammox in municipal wastewater treatment: Status, bottlenecks, and further studies. *Appl. Microbiol. Biotechnol.* **2017**, *101*, 1365–1383. [[CrossRef](#)]
52. Soliman, M.; Eldyasti, A. Ammonia-Oxidizing Bacteria (AOB): Opportunities and applications—A review. *Rev. Environ. Sci. Bio/Technol.* **2018**, *17*, 285–321. [[CrossRef](#)]
53. Hellinga, C.; Schellen, A.; Mulder, J.W.; Van, L.M.C.M.; Heijnen, J.J. The SHARON process: An innovative method for nitrogen removal from ammonium-rich waste water. *Water Sci. Technol.* **1998**, *37*, 135–142. [[CrossRef](#)]
54. Yamamoto, T.; Takaki, K.; Koyama, T.; Furukawa, K. Long-term stability of partial nitrification of swine wastewater digester liquor and its subsequent treatment by Anammox. *Bioresour. Technol.* **2008**, *99*, 6419–6425. [[CrossRef](#)]
55. Rodriguez-Sanchez, A.; Gonzalez-Martinez, A.; Martinez-Toledo, M.V.; Garcia-Ruiz, M.J.; Osorio, F.; Gonzalez-Lopez, J. The Effect of Influent Characteristics and Operational Conditions over the Performance and Microbial Community Structure of Partial Nitrification Reactors. *Water* **2014**, *6*, 1905–1924. [[CrossRef](#)]
56. Gabarro, J.; Ganigue, R.; Gich, F.; Ruscalleda, M.; Balaguer, M.D.; Colprim, J. Effect of temperature on AOB activity of a partial nitrification SBR treating landfill leachate with extremely high nitrogen concentration. *Bioresour. Technol.* **2012**, *126*, 283–289. [[CrossRef](#)]
57. Alleman, J.E. Elevated Nitrite Occurrence in Biological Wastewater Treatment Systems. *Water Sci. Technol.* **1985**, *17*, 409–419. [[CrossRef](#)]
58. Villaverde, S.; García-Encina, P.; Fdz-Polanco, F. Influence of pH over nitrifying biofilm activity in submerged biofilters. *Water Res.* **1997**, *31*, 1180–1186. [[CrossRef](#)]
59. Chen, H.; Tu, Z.; Wu, S.; Yu, G.; Du, C.; Wang, H.; Yang, E.; Zhou, L.; Deng, B.; Wang, D.; et al. Recent advances in partial denitrification-anaerobic ammonium oxidation process for mainstream municipal wastewater treatment. *Chemosphere* **2021**, *278*, 130436. [[CrossRef](#)]
60. Zhou, Y.; Oehmen, A.; Lim, M.; Vadivelu, V.; Ng, W.J. The role of nitrite and free nitrous acid (FNA) in wastewater treatment plants. *Water Res.* **2011**, *45*, 4672–4682. [[CrossRef](#)]
61. Ma, B.; Lan, Y.; Wang, Q.; Yuan, Z.; Peng, Y. Inactivation and adaptation of ammonia-oxidizing bacteria and nitrite-oxidizing bacteria when exposed to free nitrous acid. *Bioresour. Technol.* **2017**, *245*, 1266–1270. [[CrossRef](#)] [[PubMed](#)]
62. Liu, Y.; Ngo, H.H.; Guo, W.; Peng, L.; Wang, D.; Ni, B. The roles of free ammonia (FA) in biological wastewater treatment processes: A review. *Environ. Int.* **2019**, *123*, 10–19. [[CrossRef](#)] [[PubMed](#)]
63. Chung, J.; Shim, H.; Park, S.J.; Kim, S.J.; Bae, W. Optimization of free ammonia concentration for nitrite accumulation in shortcut biological nitrogen removal process. *Bioprocess Biosyst. Eng.* **2006**, *28*, 275–282. [[CrossRef](#)] [[PubMed](#)]
64. Chung, J.; Shim, H.; Lee, Y.W.; Bae, W. Comparison of influence of free ammonia and dissolved oxygen on nitrite accumulation between suspended and attached cells. *Environ. Technol.* **2005**, *26*, 21–33. [[CrossRef](#)] [[PubMed](#)]
65. Sui, Q.; Liu, C.; Zhang, J.; Dong, H.; Zhu, Z.; Wang, Y. Response of nitrite accumulation and microbial community to free ammonia and dissolved oxygen treatment of high ammonium wastewater. *Appl. Microbiol. Biotechnol.* **2016**, *100*, 4177–4187. [[CrossRef](#)]
66. Zhang, X.; Zhou, Y.; Yu, B.; Zhang, N.; Wang, L.; Fu, H.; Zhang, J. Effect of copper oxide nanoparticles on the ammonia removal and microbial community of partial nitrification process. *Chem. Eng. J.* **2017**, *328*, 152–158. [[CrossRef](#)]
67. Chu, Z.; Huang, X.; Su, Y.; Yu, H.; Rong, H.; Wang, R.; Zhang, L. Low-dose Ultraviolet-A irradiation selectively eliminates nitrite oxidizing bacteria for mainstream nitrification. *Chemosphere* **2020**, *261*, 128172. [[CrossRef](#)]
68. Huang, S.; Zhu, Y.; Lian, J.; Liu, Z.; Zhang, L.; Tian, S. Enhancement in the partial nitrification of wastewater sludge via low-intensity ultrasound: Effects on rapid start-up and temperature resilience. *Bioresour. Technol.* **2019**, *294*, 122196. [[CrossRef](#)]
69. Huang, S.; Zhu, Y.; Zhang, G.; Lian, J.; Liu, Z.; Zhang, L.; Tian, S. Effects of low-intensity ultrasound on nitrite accumulation and microbial characteristics during partial nitrification. *Sci. Total Environ.* **2020**, *705*, 135985. [[CrossRef](#)]
70. Tian, S.; Huang, S.; Zhu, Y.; Zhang, G.; Lian, J.; Liu, Z.; Zhang, L.; Qin, X. Effect of low-intensity ultrasound on partial nitrification: Performance, sludge characteristics, and properties of extracellular polymeric substances. *Ultrason. Sonochem.* **2021**, *73*, 105527. [[CrossRef](#)]
71. Jia, W.; Zhang, J.; Lu, Y.; Li, G.; Yang, W.; Wang, Q. Response of nitrite accumulation and microbial characteristics to low-intensity static magnetic field during partial nitrification. *Bioresour. Technol.* **2018**, *259*, 214–220. [[CrossRef](#)]
72. Schopf, A.; Delatolla, R.; Mathew, R.; Tsitouras, A.; Kirkwood, K.M. Investigation of copper inhibition of nitrifying moving bed biofilm (MBBR) reactors during long term operations. *Bioprocess Biosyst. Eng.* **2018**, *41*, 1485–1495. [[CrossRef](#)]
73. Su, H.; Zhang, D.; Antwi, P.; Xiao, L.; Liu, Z.; Deng, X.; Asumadu-Sakyi, A.B.; Li, J. Effects of heavy rare earth element (yttrium) on partial-nitrification process, bacterial activity and structure of responsible microbial communities. *Sci. Total Environ.* **2020**, *705*, 135797. [[CrossRef](#)]
74. Zhang, X.; Zhou, Y.; Zhang, N.; Zheng, K.; Wang, L.; Han, G.; Zhang, H. Short-term and long-term effects of Zn (II) on the microbial activity and sludge property of partial nitrification process. *Bioresour. Technol.* **2017**, *228*, 315–321. [[CrossRef](#)]

75. Su, H.; Zhang, D.; Antwi, P.; Xiao, L.; Luo, W.; Deng, X.; Lai, C.; Liu, Z.; Shi, M.; Manefield, M.J. Unraveling the effects of light rare-earth element (Lanthanum (III)) on the efficacy of partial-nitrification process and its responsible functional genera. *Chem. Eng. J.* **2021**, *408*, 127311. [[CrossRef](#)]
76. Giustinianovich, E.A.; Campos, J.L.; Roeckel, M.D.; Estrada, A.J.; Mosquera-Corral, A.; Val Del Rio, A. Influence of biomass acclimation on the performance of a partial nitrification-anammox reactor treating industrial saline effluents. *Chemosphere* **2018**, *194*, 131–138. [[CrossRef](#)]
77. Val Del Rio, A.; Pichel, A.; Fernandez-Gonzalez, N.; Pedrouso, A.; Fra-Vazquez, A.; Morales, N.; Mendez, R.; Campos, J.L.; Mosquera-Corral, A. Performance and microbial features of the partial nitrification-anammox process treating fish canning wastewater with variable salt concentrations. *J. Environ. Manag.* **2018**, *208*, 112–121. [[CrossRef](#)]
78. Li, X.; Yuan, Y.; Yuan, Y.; Bi, Z.; Liu, X.; Huang, Y.; Liu, H.; Chen, C.; Xu, S. Effects of salinity on the denitrification efficiency and community structure of a combined partial nitrification- anaerobic ammonium oxidation process. *Bioresour. Technol.* **2018**, *249*, 550–556. [[CrossRef](#)]
79. Li, J.; Zhang, Q.; Li, X.; Peng, Y. Rapid start-up and stable maintenance of domestic wastewater nitrification through short-term hydroxylamine addition. *Bioresour. Technol.* **2019**, *278*, 468–472. [[CrossRef](#)]
80. Wu, Z.; Gao, J.; Cui, Y.; Li, D.; Dai, H.; Guo, Y.; Li, Z.; Zhang, H.; Zhao, M. Metagenomics insights into the selective inhibition of NOB and comammox by phenacetin: Transcriptional activity, nitrogen metabolism and mechanistic understanding. *Sci. Total Environ.* **2022**, *803*, 150068. [[CrossRef](#)]
81. Wang, J.; Liu, Y.; Li, W. Model-based assessment of nitrification using formic acid as a selective inhibitor. *J. Clean. Prod.* **2020**, *276*, 124290. [[CrossRef](#)]
82. Xu, G.; Xu, X.; Yang, F.; Liu, S.; Gao, Y. Partial nitrification adjusted by hydroxylamine in aerobic granules under high DO and ambient temperature and subsequent Anammox for low C/N wastewater treatment. *Chem. Eng. J.* **2012**, *213*, 338–345. [[CrossRef](#)]
83. Xiang, T.; Gao, D. Comparing two hydrazine addition strategies to stabilize mainstream deammonification: Performance and microbial community analysis. *Bioresour. Technol.* **2019**, *289*, 121710. [[CrossRef](#)] [[PubMed](#)]
84. Xiang, T.; Gao, D.; Wang, X. Performance and microbial community analysis of two sludge type reactors in achieving mainstream deammonification with hydrazine addition. *Sci. Total Environ.* **2020**, *715*, 136377. [[CrossRef](#)] [[PubMed](#)]
85. Wang, J.P.; Liu, Y.D.; Meng, F.G.; Li, W. The short- and long-term effects of formic acid on rapid nitrification start-up. *Environ. Int.* **2020**, *135*, 105350. [[CrossRef](#)]
86. Akizuki, S.; Natori, N.; Cuevas-Rodríguez, G.; Toda, T. Application of nitrifying granular sludge for stable ammonium oxidation under intensive light. *Biochem. Eng. J.* **2020**, *160*, 107631. [[CrossRef](#)]
87. Santos, A.L.; Moreirinha, C.; Lopes, D.; Esteves, A.C.; Henriques, I.; Almeida, A.; Domingues, M.R.; Delgadillo, I.; Correia, A.; Cunha, A. Effects of UV radiation on the lipids and proteins of bacteria studied by mid-infrared spectroscopy. *Environ. Sci. Technol.* **2013**, *47*, 6306–6315. [[CrossRef](#)]
88. Sharma, A.K.; Singh, H.; Chakrapani, H. Photocontrolled endogenous reactive oxygen species (ROS) generation. *Chem. Commun.* **2019**, *55*, 5259–5262. [[CrossRef](#)]
89. Lucker, S.; Wagner, M.; Maixner, F.; Pelletier, E.; Koch, H.; Vacherie, B.; Rattei, T.; Damste, J.S.; Spieck, E.; Le Paslier, D.; et al. A *Nitrospira* metagenome illuminates the physiology and evolution of globally important nitrite-oxidizing bacteria. *Proc. Natl. Acad. Sci. USA* **2010**, *107*, 13479–13484. [[CrossRef](#)]
90. Laloo, A.E.; Wei, J.; Wang, D.; Narayanasamy, S.; Vanwonterghem, I.; Waite, D.; Steen, J.; Kaysen, A.; Heintz-Buschart, A.; Wang, Q.; et al. Mechanisms of Persistence of the Ammonia-Oxidizing Bacteria *Nitrosomonas* to the Biocide Free Nitrous Acid. *Environ. Sci. Technol.* **2018**, *52*, 5386–5397. [[CrossRef](#)]
91. Rezayian, M.; Niknam, V.; Ebrahimzadeh, H. Oxidative damage and antioxidative system in algae. *Toxicol. Rep.* **2019**, *6*, 1309–1313. [[CrossRef](#)]
92. Lipschultz, F.; Wofsy, S.C.; Fox, L.E. The effects of light and nutrients on rates of ammonium transformation in a eutrophic river. *Mar. Chem.* **1985**, *16*, 329–341. [[CrossRef](#)]
93. Zheng, M.; Liu, Y.C.; Xin, J.; Zuo, H.; Wang, C.W.; Wu, W.M. Ultrasonic Treatment Enhanced Ammonia-Oxidizing Bacterial (AOB) Activity for Nitrification Process. *Environ. Sci. Technol.* **2016**, *50*, 864–871. [[CrossRef](#)]
94. Pitt, W.G.; Ross, S.A. Ultrasound increases the rate of bacterial cell growth. *Biotechnol. Prog.* **2010**, *19*, 1038–1044. [[CrossRef](#)]
95. Zheng, M.; Liu, Y.C.; Xu, K.N.; Wang, C.W.; He, H.; Zhu, W.; Dong, Q. Use of low frequency and density ultrasound to stimulate partial nitrification and simultaneous nitrification and denitrification. *Bioresour. Technol.* **2013**, *146*, 537–542. [[CrossRef](#)]
96. Moore, R.L. Biological effects of magnetic fields: Studies with microorganisms. *Can. J. Microbiol.* **1979**, *25*, 1145–1151. [[CrossRef](#)]
97. Filipic, J.; Kraigher, B.; Tepus, B.; Kokol, V.; Mandic-Mulec, I. Effects of low-density static magnetic fields on the growth and activities of wastewater bacteria *Escherichia coli* and *Pseudomonas putida*. *Bioresour. Technol.* **2012**, *120*, 225–232. [[CrossRef](#)]
98. Richardson, D.J. Bacterial respiration: A flexible process for a changing environment. *Microbiology* **2000**, *146* (Pt 3) Pt 3, 551–571. [[CrossRef](#)]
99. Niu, C.; Liang, W.; Ren, H.; Geng, J.; Ding, L.; Xu, K. Enhancement of activated sludge activity by 10-50 mT static magnetic field intensity at low temperature. *Bioresour. Technol.* **2014**, *159*, 48–54. [[CrossRef](#)]
100. Wang, Z.; Liu, X.; Ni, S.Q.; Zhang, J.; Zhang, X.; Ahmad, H.A.; Gao, B. Weak magnetic field: A powerful strategy to enhance partial nitrification. *Water Res.* **2017**, *120*, 190–198. [[CrossRef](#)]
101. Nies, D. Microbial heavy metal resistance. *Appl. Microbiol. Biotechnol.* **1999**, *51*, 730–750. [[CrossRef](#)] [[PubMed](#)]

102. Noctor, G.; Queval, G.; Mhamdi, A.; Chaouch, S.; Foyer, C.H. Glutathione. *Arab. Book* **2011**, *9*, e0142. [[CrossRef](#)] [[PubMed](#)]
103. Strous, M.; Pelletier, E.; Mangenot, S.; Rattei, T.; Lehner, A.; Taylor, M.W.; Horn, M.; Daims, H.; Bartol-Mavel, D.; Wincker, P. Deciphering the evolution and metabolism of an anammox bacterium from a community genome. *Nature* **2006**, *440*, 790–794. [[CrossRef](#)] [[PubMed](#)]
104. Hu, Z.; Chandran, K.; Grasso, D.; Smets, B.F. Comparison of nitrification inhibition by metals in batch and continuous flow reactors. *Water Res.* **2004**, *38*, 3949–3959. [[CrossRef](#)] [[PubMed](#)]
105. Hu, Z.; Chandran, K.; Grasso, D.; Smets, B.F. Impact of metal sorption and internalization on nitrification inhibition. *Environ. Sci. Technol.* **2003**, *37*, 728. [[CrossRef](#)] [[PubMed](#)]
106. Zhao, Y.; Park, H.-D.; Park, J.-H.; Zhang, F.; Chen, C.; Li, X.; Zhao, D.; Zhao, F. Effect of different salinity adaptation on the performance and microbial community in a sequencing batch reactor. *Bioresour. Technol.* **2016**, *216*, 808–816. [[CrossRef](#)]
107. Yang, G.F.; Yu, J.J.; Ping, Z. The inhibition of the Anammox process: A review. *Chem. Eng. J.* **2012**, *197*, 67–79. [[CrossRef](#)]
108. García-Ruiz, M.; Castellano-Hinojosa, A.; González-López, J.; Osorio, F. Effects of salinity on the nitrogen removal efficiency and bacterial community structure in fixed-bed biofilm CANON bioreactors. *Chem. Eng. J.* **2018**, *347*, 156–164. [[CrossRef](#)]
109. Liu, Y.; Peng, C.Y.; Tang, B.; Wang, S.Y.; Zhao, K.F.; Peng, Y.Z. Determination effect of influent salinity and inhibition time on partial nitrification in a sequencing batch reactor treating saline sewage. *Desalination* **2009**, *246*, 556–566.
110. Campos, J.L.; Mosquera-Corral, A.; Sánchez, M.; Méndez, R.; Lema, J.M. Nitrification in saline wastewater with high ammonia concentration in an activated sludge unit. *Water Res.* **2002**, *36*, 2555–2560. [[CrossRef](#)]
111. Panswad, T.; Anan, C. Specific oxygen, ammonia, and nitrate uptake rates of a biological nutrient removal process treating elevated salinity wastewater. *Bioresour. Technol.* **1999**, *70*, 237–243. [[CrossRef](#)]
112. Huang, X.; Mi, W.; Ito, H.; Kawagoshi, Y. Probing the dynamics of three freshwater Anammox genera at different salinity levels in a partial nitritation and Anammox sequencing batch reactor treating landfill leachate. *Bioresour. Technol.* **2021**, *319*, 124112. [[CrossRef](#)]
113. Soler-Jofra, A.; Perez, J.; van Loosdrecht, M.C.M. Hydroxylamine and the nitrogen cycle: A review. *Water Res.* **2021**, *190*, 116723. [[CrossRef](#)]
114. Wang, Y.; Wang, H.; Zhang, J.; Yao, L.; Wei, Y. Deciphering the evolution of the functional genes and microbial community of the combined partial nitritation-anammox process with nitrate build-up and its in situ restoration. *RSC Adv.* **2016**, *6*, 111702–111712. [[CrossRef](#)]
115. Sui, Q.; Wang, Y.; Wang, H.; Yue, W.; Chen, Y.; Yu, D.; Chen, M.; Wei, Y. Roles of hydroxylamine and hydrazine in the in-situ recovery of one-stage partial nitritation-anammox process: Characteristics and mechanisms. *Sci. Total Environ.* **2020**, *707*, 135648. [[CrossRef](#)]
116. Li, H.; Yao, H.; Zhang, D.; Zuo, L.; Ren, J.; Ma, J.; Pei, J.; Xu, Y.; Yang, C. Short- and long-term effects of manganese, zinc and copper ions on nitrogen removal in nitritation-anammox process. *Chemosphere* **2018**, *193*, 479–488. [[CrossRef](#)]
117. Albaiges, J.; Casado, F.; Ventura, F. Organic indicators of groundwater pollution by a sanitary landfill. *Water Res.* **1986**, *20*, 1153–1159. [[CrossRef](#)]
118. Chen, H.; Zeng, L.; Wang, D.; Zhou, Y.; Yang, X. Recent advances in nitrous oxide production and mitigation in wastewater treatment. *Water Res.* **2020**, *184*, 116168. [[CrossRef](#)]
119. Massara, T.M.; Malamis, S.; Guisasola, A.; Baeza, J.A.; Noutsopoulos, C.; Katsou, E. A review on nitrous oxide (N₂O) emissions during biological nutrient removal from municipal wastewater and sludge reject water. *Sci. Total Environ.* **2017**, *596–597*, 106–123. [[CrossRef](#)]
120. Lv, Y.; Ju, K.; Sun, T.; Wang, L.; Miao, R.; Liu, T.; Wang, X. Effect of the dissolved oxygen concentration on the N₂O emission from an autotrophic partial nitritation reactor treating high-ammonium wastewater. *Int. Biodeterior. Biodegrad.* **2016**, *114*, 209–215. [[CrossRef](#)]
121. Peng, L.; Carvajal-Arroyo, J.M.; Seuntjens, D.; Prat, D.; Colica, G.; Pintucci, C.; Vlaeminck, S.E. Smart operation of nitritation/denitritation virtually abolishes nitrous oxide emission during treatment of co-digested pig slurry centrate. *Water Res.* **2017**, *127*, 1–10. [[CrossRef](#)] [[PubMed](#)]
122. Liu, S.-Q.; Zhou, Y.; Meng, Z.-D.; Wu, Z.-Y.; Zhu, X.-L. Smart photocatalytic removal of ammonia through molecular recognition of zinc ferrite/reduced graphene oxide hybrid catalyst under visible-light irradiation. *Catal. Sci. Technol.* **2017**, *7*, 3210–3219. [[CrossRef](#)]
123. Ma, H.; Niu, Q.; Zhang, Y.; He, S.; Li, Y.Y. Substrate inhibition and concentration control in an UASB-Anammox process. *Bioresour. Technol.* **2017**, *238*, 263–272. [[CrossRef](#)] [[PubMed](#)]
124. Gao, S.H.; Fan, L.; Peng, L.; Guo, J.; Agulló-Barceló, M.; Yuan, Z.; Bond, P.L. Determining multiple responses of *Pseudomonas aeruginosa* PAO1 to an antimicrobial agent, free nitrous acid. *Environ. Sci. Technol.* **2016**, *50*, 5305–5312. [[CrossRef](#)]