

Review

Dietary Advanced Glycation End Products: Digestion, Metabolism and Modulation of Gut Microbial Ecology

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Abstract: The formation of advanced glycation end products (AGEs) in foods is accelerated with heat treatment, particularly within foods that are cooked at high temperatures for long periods of time using dry heat. The modern processed diet is replete with AGEs, and excessive AGE consumption is thought to be associated with a number of negative health effects. Many dietary AGEs have high molecular weight and are not absorbed in the intestine, and instead pass through to the colon, where they are available for metabolism by the colonic bacteria. Recent studies have been conducted to explore the effects of AGEs on the composition of the gut microbiota as well as the production of beneficial microbial metabolites, in particular, short-chain fatty acids. However, there is conflicting evidence regarding the impact of dietary AGEs on gut microbiota reshaping, which may be due, in part, to the formation of alternate compounds during the thermal treatment of foods. This review summarises the current evidence regarding dietary sources of AGEs, their gastrointestinal absorption and role in gut microbiota reshaping, provides a brief overview of the health implications of dietary AGEs and highlights knowledge gaps and avenues for future study.

Keywords: advanced glycation end products; maillard reaction products; heat-treated diets; microbiota; short-chain fatty acids

1. Introduction

Advanced glycation end products (AGEs) is a term used to describe a heterogeneous group of compounds that are formed through a series of nonenzymatic reactions, together termed the Maillard reaction, involving an amino acid with a free amino group (such as lysine or arginine) and a free carbonyl group of a reducing sugar. This initial amine–carbonyl reaction results in an imine, often referred to as a Schiff base, which undergoes rearrangement to form an intermediate glycation product, the Amadori product, which is a more stable α -ketoamine. These Schiff bases and Amadori products are initial glycation products, and at this stage are reversible. Amadori products can also undergo degradation to form dicarbonyl compounds, such as methylglyoxal (MG), and then undergo further degradation, oxidation, reduction and condensation reactions, leading to irreversible AGE formation [1], as illustrated in Figure 1. Major AGEs present in vivo or from exogenous sources include methylglyoxal–hydroimidazolone (MG-H1), carboxymethyl–lysine (CML), carboxyethyl–lysine (CEL), pentosidine and pyrrolidine [2]. Given the heterogeneity of AGEs formed, it is often difficult to fully quantify their formation, and subsequently CML, CEL and pentosidine are commonly used as biomarkers for the glycation process [3]. Furthermore, the heterogeneity of AGEs formed during the heat treatment of foods is greater than that of those formed physiologically, and the diversity of dietary AGEs has hindered efforts to study their effects [4]. It should be noted that the

AGE pyrraline is formed exclusively within food products and thus measuring pyrraline may provide a better indicator of the effects of AGEs of exclusively dietary origin [5]. The objective of this review is to describe the food sources, absorption and health effects of dietary AGEs and provide a summary of in vitro and in vivo studies that have provided high AGE intervention and assessed the composition of the gut microbiome.

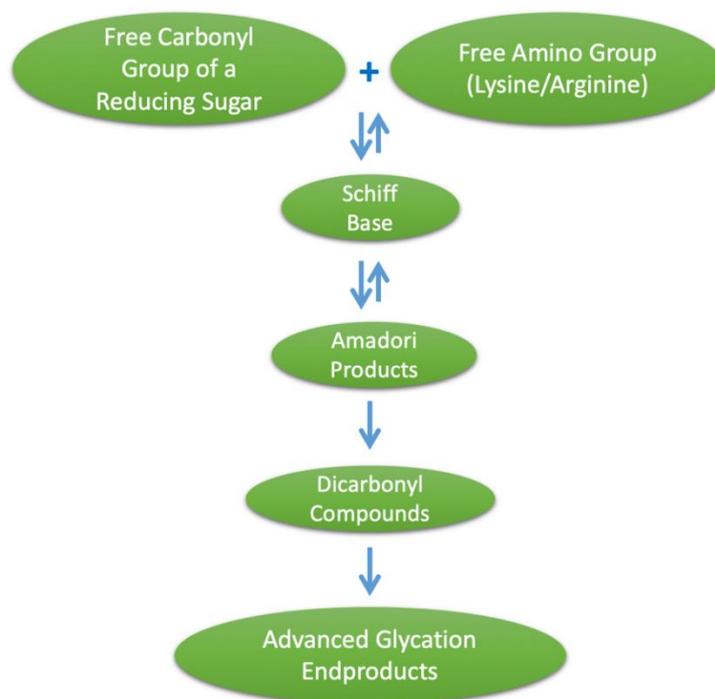


Figure 1. Formation of advanced glycation end products (AGEs).

2. Endogenous Sources of AGEs

AGE-modified proteins accumulate within the body with aging and are thought to play a role in a number of age-related diseases including diabetes and neurodegenerative and cardiovascular diseases [6]. Endogenous AGE formation is accelerated during circumstances of high plasma glucose, such as is prevalent in diabetes [7]. Furthermore, there is evidence to indicate that excessive dietary intake of sugars, particularly fructose, contributes to endogenous AGE formation [8]. In situations of oxidative stress, oxidation of fatty acids and glucose can lead to the production of reactive carbonyls, such as MG, which contribute to AGE formation [9].

3. Exogenous Sources of AGEs

In addition to the endogenous formation of AGEs that can occur as described above, exogenous ingestion of AGEs can significantly contribute to their accumulation. Cigarette smoke contains glycation products that are highly reactive and lead to the formation of AGEs [10], which may contribute to the increased AGE accumulation in serum [11] and tissue [12] observed in cigarette smokers. AGEs are also formed in foods, and their formation is highly dependent on cooking methods. The heat treatment of food results in the generation of Maillard reaction products (MRPs), including AGEs, which improve the aroma and flavour of food products [2]. Food processing and cooking techniques that utilise dry heat (frying, roasting, baking, grilling, barbecuing) result in greater AGE formation compared with techniques that use lower temperatures for longer periods of time with higher water content, such as boiling or steaming [13,14]. Bread is a well-known source of MRPs, including AGEs, and reducing baking temperature from 250 °C to 200 °C resulted in a 20% lower accumulation of CML [15]. Higher pH levels can also increase the formation of AGEs, as the alkaline

conditions promote amino groups being in the basic deprotonated form, increasing reactivity [16]. Pretreating foods prior to cooking with acidic solutions, such as marinating with vinegar or lemon, to lower their pH has been demonstrated to reduce the formation of AGEs in food products [17].

The modern Western diet is replete with heat-treated foods that contribute to the intake of AGEs [18]. Foods that contribute large quantities of AGEs to the modern diet include processed cereal products such as biscuits, bakery products and extruded breakfast cereals [19]; powdered milk; and fish, chicken or meat cooked with dry heat [13]. It is estimated that modern dietary intake ranges from 25 to 75 mg/d for AGEs and 500 to 1200 mg/d for Amadori products, with milk and bakery products being the major contributors [20]. The highest concentrations of CML, CEL and MG-H1 were found in biscuits, cereal products, high-heat-processed meats, peanuts and peanut butter [13]. Dietary AGE intake has been shown to correlate with plasma-free AGEs, but not protein-bound AGEs in the plasma [21], suggesting that plasma protein-bound AGEs are more indicative of endogenous AGE formation. While exogenous and endogenous AGEs are considered to be two separate sources, recently the observation has been made that they may act synergistically and contribute to an overall glycotoxin burden, which may increase the detrimental health effects of AGEs [18].

4. Exogenous AGEs and Health

Numerous studies support the idea that dietary consumption of AGEs contributes to oxidative stress and inflammation in animal models; however, results have been less consistent in human trials [22]. In humans, Vlassara et al. observed that two weeks of a high-AGE diet was associated with increases in circulating C-reactive protein (CRP) and tumour necrosis factor alpha (TNF- α) protein and expression in peripheral blood mononuclear cells [23], whilst Semba et al. found no effect of six weeks of high-AGE feeding on CRP, interleukin 6 (IL-6) or TNF- α receptors [24]. Two-week consumption of a diet high in AGEs was associated with an increase in inflammatory markers and albuminuria in a cohort of overweight and obese but otherwise healthy people [25]. A recent meta-analysis of randomised controlled trials utilising a low-AGE diet was associated with significant reductions in TNF- α and 8-isoprostane levels [26]. It is interesting to note that even a single high-AGE meal can result in acute endothelial dysfunction [27].

Excessive AGE consumption in mice has been implicated in the development of hepatic inflammation in the absence of steatosis [28], and in a rat model of nonalcoholic fatty liver disease, high dietary AGEs exacerbated liver injury, inflammation and liver fibrosis [29]. Chronic AGE intake has been postulated to lead to cognitive decline and Alzheimer's disease [30]. Several studies in animal models using both healthy animals and a 5/6 nephrectomy model of chronic kidney disease (CKD) illustrated that six weeks of high-AGE feeding increased proteinuria [31,32]. In a diabetic mouse model with db/db mice, four months of high-AGE feeding was associated with an increase in albuminuria [33]. Low-AGE diets have been shown to improve markers of inflammation and oxidative stress in haemodialysis patients [34] and those with stage 3 CKD [35]; however, neither of these studies reported on kidney function. There is mounting evidence that excessive consumption of dietary AGEs contributes to inflammation and oxidative stress, which has implications for a number of chronic disease states.

5. Mechanisms of AGE-Induced Damage Exogenous AGEs and Health

AGEs can exert deleterious health effects and contribute to the progression of disease states through a number of mechanisms. Crosslinking of extracellular matrix proteins by AGEs contributes to thickening of the basement membrane and glomerulosclerosis within the kidney as well as arterial stiffness, which contributes to atherosclerosis [36]. AGEs have also been observed in vitro to induce basement membrane hypertrophy of the blood-brain barrier [37], and AGE modification of matrix proteins affected cellular migration and adhesion properties of T cells in vitro [38]. Guanine and adenine nucleotides contain amino groups that are susceptible to glycation, and AGE modification of DNA can lead to single strand breaks and mutations [39], which have been suggested as potential

contributors to the development of renal disease [40]. There is also evidence that AGEs can act as catalytic sites for free radical formation, contributing to further damage [41].

AGEs interact with a number of cell surface receptors, of which the receptor for AGEs (RAGE) has been the most widely studied [42]. RAGE is expressed by numerous cells, including neurons, immune cells, endothelial cells and myocytes [43]. AGE–RAGE binding leads to activation of intracellular nuclear factor of κ light-chain-enhancer in activated B cells and subsequently increases the expression of a number of proinflammatory cytokines, including TNF- α , IL-6 and interleukin 1 [44]. The ligation of RAGE by AGEs also increases nicotinamide adenine dinucleotide phosphate oxidase and the production of mitochondrial reactive oxygen species [45]. AGE ligation with RAGE is implicated in the progression of a number of disease states, including cancer, atherosclerosis, stroke, metabolic syndrome and kidney disease [46,47]. There are a number of mechanisms by which AGEs, of either endogenous or exogenous origin, may impart deleterious effects.

6. AGE Absorption

Early studies indicated that upon oral administration, only 10–30% of AGEs are absorbed into the systemic circulation [48,49]. AGEs can exist in either free form as a single amino acid or a free low-molecular-weight (LMW, <5 kDa) peptide, or bound to proteins, forming high-molecular-weight (HMW) compounds [18]. Free CML can be absorbed by simple diffusion [50], whilst intestinal absorption of pyrroline is considered to happen mostly as a dipeptide rather than a free amino acid [51], with the dipeptide form of pyrroline absorbed across the intestinal epithelium using peptide transporter 1 (PEPT1) [52]. Further research has identified that PEPT1 is also utilised for absorption of the dipeptide versions of CML, CEL and MG-H1 [53]. Absorption of the AGE pentosidine was greater when it was provided in its free form (in brewed coffee) than protein-bound (in bakery products) [54], and diets containing protein-bound CML were associated with significantly higher faecal excretion of CML compared with diets containing the LMW fraction [55]. Modification of proteins with AGEs reduces the digestibility of the proteins [56], limiting their ability to undergo proteolytic digestion [18]. As HMW proteins require proteolytic digestion prior to absorption [2], the LMW fractions of AGEs are more likely to be absorbed [57], with the nonabsorbed AGEs progressing through the gastrointestinal tract. Tessier et al. used ^{13}C -labelled CML incorporated into ^{13}C -CML-bovine serum albumin (BSA) to identify CML of exclusively dietary origin in the diets of rats for 30 days. Organ accumulation, primarily in the kidneys, and faecal excretion of ^{13}C -labelled CML was observed, indicating that intestinal CML absorption from dietary protein-bound CML does occur; however, a proportion is not absorbed, instead passing through the gastrointestinal tract [58]. There is a strong correlation between dietary intake of CML and faecal excretion [55,59,60], suggesting an increased opportunity for interaction between AGEs and the colonic microbiome. Between 20% and 50% of ingested CML appears to be excreted in the faeces [59,60], suggesting that there is a proportion of ingested AGEs that are not absorbed and not defecated and may be metabolised intraluminally by the microbiome. It is recognised that knowledge of the specific absorption mechanisms of dietary AGEs is limited, and further studies are required to elucidate them [61].

7. Dietary AGEs and Gut Microbial Composition

7.1. In Vitro Studies

In vitro work utilising human faecal samples has established that some AGEs become available to be selectively metabolised by certain members of the gut microbiota. Early work showed that AGEs reduce the growth rate of *Bacillus stearothermophilus* in an in vitro assay [62], and it has been established that some bacteria possess deglycation enzymes, which indicates that bacterial metabolism of glycated proteins is a possibility [63]. When faecal samples were taken from patients with ulcerative colitis (UC) and incubated with glycated BSA, there was an increase in the abundance of sulphate-reducing bacteria (SRB) and a decrease in *Bifidobacteria* and *Eubacterium rectale* compared with faeces that were incubated

with nonglycated BSA [64]. A reduction in jejunal alanine absorption has been observed in animal models of experimental colitis [65,66]; however, it is unknown whether UC affects the gastrointestinal absorption of AGE-modified amino acids. In healthy controls, incubation with glycated BSA also increased the abundance of SRB, but there was no change in other bacterial groups, indicating that the initial composition of the microbiota has an impact on how it interacts with glycated proteins [64]. Twenty-four-hour incubation of a faecal sample from one healthy human volunteer with glycated pea protein was associated with an increased abundance of *Bifidobacteria* and *Lactobacilli*, as assessed by fluorescent in situ hybridization (FISH) with genus-specific 16S rRNA-targeted oligonucleotide probes, when compared with nonglycated pea protein [67]. A similar study, which utilised the same FISH probes, found that glycated gluten was associated with an expansion of *Bifidobacteria* but a contraction of *Lactobacilli* compared with heated nonglycated gluten [68]. Consistent with the bifidogenic effect that was observed with glycated gluten and pea protein, a study that incubated faecal samples from healthy volunteers with HMW bread crust fraction showed increased *Bifidobacteria*, as assessed by plating on Beerens agar, a selective growth medium for *Bifidobacteria* [69]. A recent study analysing the effect of bread crust on the microbiome observed that there were large interindividual differences with regard to *Bifidobacteria*, but noted that there was a consistent reduction in *Enterobacteria* across subjects [70]. At the phylum level, fermentation of glycated fish protein was associated with an expansion of Firmicutes and a contraction of Bacteroidetes [71]. A study using isolated AGEs identified that CML and fructoselysine are metabolised by human colonic microbiota, whilst pyrraline is not [72]. These findings indicate that whilst glycated substrates do alter microbial composition in vitro, there is a lack of agreement between studies on the specific microbial changes, which may be due to the different glycated substrates used. A summary of in vitro fermentation studies that have shown the effects of AGEs on the human microbiome is presented in Table 1.

Table 1. Summary of in vitro studies.

Study	Stool Donors	High AGE Intervention	Control	<i>Bifidobacteria</i>	<i>Lactobacilli</i>	<i>Bacteroides</i>	<i>E. rectale</i>	<i>Escherichia/Shigella</i>	Other Microbiota	Total SCFAs	Acetic Acid	Propionic Acid	Butyric Acid
[64]	UC patients	Glycated BSA	Heated BSA	↓	-	↑	↓	-	↑ SRB, ↑ Clostridia	-	↔	↔	↔
	Healthy donors	Glycated BSA	Heated BSA	↔	-	↔	↔	-	↑ SRB, ↔ Clostridia	-	-	-	-
[68]	Healthy donors	Glycated gluten	Heated gluten	↑	↓	↔	-	-		-	↓	↓	↓
[71]	Healthy donors	Glycated fish protein	Heated fish protein	-	-	↓	-	↔	↓ Bact, ↑ Firm	↑	↑	↔	↔
[70] ¹	Healthy donors	Bread crust model	Prior to fermentation	↔	-	-	-	-	↓ Enterobacteria	-	-	-	-
[69]	Healthy donors	Bread	Prior to fermentation	↑	-	↓	-	-		-	-	-	-
[67]	Healthy donors	Glycated pea protein	Nonglycated pea protein	↑	↑	↑	-	-	↑ Enterobacteriaceae	-	↔	↔	↔

Bact, Bacteroidetes; BSA, bovine serum albumin; Firm, Firmicutes; SCFA, short-chain fatty acid, SRB, sulphate-reducing bacteria; UC, ulcerative colitis. ↑ = increased, ↓ = decreased, ↔ = no change, - = not reported. ¹ Reported SCFAs as ratios rather than absolute values and was not included.

7.2. Human In Vivo Studies

Whilst in vitro work has provided some insight into the effects of AGEs on the human microbiota, there is a dearth of studies modulating the AGE content of diets in humans and assessing the composition of the microbiome in vivo. A 2014 study assessed the effects of two weeks of a diet high in AGEs on the microbiome in adolescent males using targeted qPCR to measure the changes in seven bacterial groups [73]. This study observed that a high-AGE diet was associated with a contraction in *Lactobacilli*, which was negatively correlated with CML intake, and an expansion in Enterobacteria [73]. No significant differences were observed in *Bifidobacteria*, *Bacteroides*, *E. rectale* or *Clostridium leptum*. More recently, a one-month low-AGE intervention relative to the patient's regular diet, which was used as the high-AGE comparator, was undertaken in peritoneal dialysis patients [74]. This study assessed the composition of the microbiome by sequencing the V3-4 region of the 16S rRNA gene and reported that dietary AGE restriction was associated with a contraction of *Prevotella copri* and *Bifidobacterium animalis* and an expansion of *Alistipes indistinctus*, *Clostridium hathewayi*, *Clostridium citroniae* and *Ruminococcus gnavreaii*, whilst no difference was observed in α diversity [74]. The study authors noted that the relative abundance of *B. animalis*, a species generally considered to confer health benefits, that they found in peritoneal dialysis patients was eightfold lower than what was reported in healthy controls [75]. Interestingly, randomised controlled trials have shown that sevelamer, a phosphate binder commonly prescribed to patients with CKD, is associated with reductions in serum AGEs in haemodialysis patients [76] and diabetic patients with stage 2–4 CKD [77]. In vitro work has illustrated that sevelamer binds AGE-BSA [78], indicating that it may lower serum AGEs by sequestering intestinal AGEs and limiting gastrointestinal absorption; however, whether sevelamer-bound AGEs are available for microbial metabolism in the colon is unknown. Further work is required in different human cohorts to assess the impact of high-AGE diets on the composition of the microbiome, and to link these changes to health outcomes.

7.3. Animal In Vivo Studies

Whilst there is a paucity of human studies, a number of studies have been undertaken using animals and assessed either the caecal or faecal microbiome following a high-AGE intervention. A contraction in *Lactobacilli* observed in humans following a high-AGE diet [73] and in vitro with glycated gluten [68] was confirmed by a rat study showing that diets supplemented with either the LMW fraction, the HMW fraction or total bread crust were associated with reductions in *Lactobacilli* compared with a control diet [79].

Animals (mice and catfish) that were fed thermally treated (steamed) fish had subsequent decreases in microbial α diversity compared with animals that were fed nonthermally treated fish [80]. Several animal studies have shown that heat-treated diets are associated with phylum-level changes in the composition of the gut microbiome, including a decrease in Bacteroidetes and an increase in Firmicutes [80,81], although these findings were not observed in rats consuming glycated fish protein [82]. The effects on *Bifidobacteria* have been inconsistent between studies, with a contraction of *Bifidobacteria* observed in rats following either a heat-treated diet [73] or supplementation with bread crust [79], although another study showed an expansion of *Bifidobacteria* following breast crust supplementation [83]. Oral administration of CML in mice with dextran sodium sulphate (DSS)-induced colitis was associated with a contraction in both Proteobacteria and *Enterobacteriaceae* [84]. Similar to the in vitro findings, there are discrepancies between studies, which limits our ability to draw broad conclusions. In general, high-AGE diets were associated with a contraction in Bacteroidetes, *Bifidobacteria*, *Lactobacilli* and α diversity; however, the findings of some studies contradict this. A summary of in vivo studies that showed the effects of AGEs on the microbiome is presented in Table 2.

Table 2. Summary of in vivo studies.

Study	Model	High-AGE Intervention	Control	<i>Bifidobacteria</i>	<i>Lactobacilli</i>	<i>Bacteroides</i>	<i>C. Leptum</i>	<i>E. rectale</i>	<i>Escherichia/Shigella</i>	Other Microbiota	Total SCFAs	Acetic Acid	Propionic Acid	Butyric Acid
[73]	Healthy males	High MRP diet	Low MRP diet	↓	☐	☐	☐	☐	↓	↓ Enterobacteria	-	-	-	-
	Wistar rats, male	Heated AIN93G (150 °C for 90 m)	AIN93G	↓	☐	☐	☐	☐	☐	☐ Enterobacteria	-	-	-	-
[79]	Wistar rats, male	Bread crust (10% diet)	AIN93G	↓	↓	☐	☐	☐	↑	☐ Enterobacteriaceae	↑	↑	↑	☐
		HMW BC (10% diet)	AIN93G	☐	↓	↑	☐	☐	☐	☐ Enterobacteriaceae	↑	☐	↑	↑
		LMW BC (10% diet)	AIN93G	☐	↓	☐	☐	☐	☐	☐ Enterobacteriaceae	↑	☐	↑	☐
[80]	C57BL/6 mice, male	Steamed fish (15m)	Fish	-	-	↓	-	-	-	↓ α diversity, ↓ Bact, ↑ FB	-	-	-	-
	Catfish, male	Steamed fish (15m)	Fish	-	-	↑	-	-	-	↓ α diversity, ↓ Bact, ↓ Firm	-	-	-	-
[81]	ApoE ^{-/-} mice, male	Heated high-fat diet (200 °C for 10 min)	High fat diet (40% E sat fat)	-	-	-	-	-	-	↓ Bact ↑ Firm, ↑ Clostridiales	-	-	-	-
[82]	SD rats, male	Glycated fish protein	Heated fish protein	-	-	☐	-	-	☐	↓ Firm, ↑ Actinobacteria	↑	☐	☐	↑
[85]	SD rats, male	Heated AIN93G (125 °C for 180 min)	AIN93G	-	-	↑	-	-	-	↓ α diversity, ↓ Bact, ↑ Proteobacteria	-	↓	☐	☐
[86]	C57BL/6 mice, male	Heated AIN93G (175 °C for 45 min)	AIN93G	-	-	↓	-	-	-	↓ Firm, ↑ Actinobacteria, ↓ Clostridia	-	↓	☐	↓
[83]	SD rats, male	Bread crust (13% diet)	A04	↑	-	-	-	-	-	☐ Bact, ☐ Firm	-	-	-	-
[84]	Balb/c mice, male, DSS-induced colitis	CML (1.6 mg/kg/d)	Saline	-	-	-	-	-	-	↓ Proteobacteria, ↓ Enterobacteriaceae	-	-	-	-
	Balb/c mice, male, healthy	CML (1.6 mg/kg/d)	Saline	-	-	-	-	-	-	↓ Lachnospiraceae, ↑ Desulfovibrionaceae	-	-	-	-
[74]	Peritoneal dialysis patients	Usual diet (high AGE)	Dietary AGE restriction	-	-	-	-	-	-	☐ α diversity, ↑ <i>Prevotella copri</i> , ↑ <i>B. animalis</i>	-	-	-	-

Bact, Bacteroidetes; FB, Firmicutes; SCFA, short-chain fatty acid; SD, Sprague-Dawley; MRP, Maillard reaction product; CML carboxymethyl-lysine. ↑ = increased, ↓ = decreased, ☐ = no change, - = not reported.

8. Dietary AGEs and Microbial Production of Short-Chain Fatty Acids

8.1. In Vitro Studies

In addition to measuring changes in the composition of the microbiome following exposure to glycated food components, several in vitro studies have also assessed the effects of these diets on the production of short-chain fatty acids (SCFAs). When faecal samples from patients with ulcerative colitis were fermented with glycated BSA, there was no change in SCFA production relative to the control heated BSA [64], which was consistent with the results seen when faecal samples from healthy volunteers were incubated with glycated pea protein [67]. A study that utilised glycated gluten observed reductions in the three main SCFAs, acetate, propionate and butyrate, following fermentation [68], whilst fermentation with glycated fish protein was associated with an increase in acetate, but not propionate or butyrate [71]. There is considerable variation in the effects of high-AGE interventions on the production of SCFAs from human faecal samples in vitro.

8.2. In Vivo Studies

To date, no in vivo human studies that have utilised diets high in AGEs have reported concentrations of faecal SCFAs, thus the knowledge of the effects of high-AGE diets on these microbial metabolites comes from the results of animal studies. Glycated fish protein was associated with an increase in caecal and faecal butyrate concentrations [82], and heat-treated diets were associated with reductions in caecal SCFA concentrations [85,86], suggesting that high-AGE diet-induced alterations in the microbial composition led to a subsequent change in the production of microbial metabolites. Furthermore, high-AGE feeding in rats is associated with a decrease in mRNA and protein concentrations of occludin and zonula occludens-1, two tight junction proteins, indicating that high-AGE feeding is associated with increased intestinal permeability [85]. Whilst a thermally treated diet was associated with a reduction in SCFAs, a previous study using bread crust as a model of a high-MRP diet observed increased SCFA concentrations [79]. SCFA production was increased with supplementation with glycated fish protein, consistent with in vitro findings, or bread crust, but decreased when animals consumed baked chow.

9. Contradictory Findings

There remains some debate as to whether thermally treated diets are detrimental to the composition of the gut microbiome and, subsequently, health. This is due, in part, to the fact that not just AGEs are formed when foods are heat treated, but also other high-molecular-weight compounds, such as melanoidins, which can act as prebiotics in a bifidogenic fashion [87]. Results from in vitro studies have illustrated that a bifidogenic effect is observed with thermal treatment of bread crust [69], glycated gluten [68] and glycated pea protein [67]. However, in vivo it was observed that rats receiving bread crust-supplemented diets had a contraction of *Bifidobacteria* compared with control diets [79], and it was recently shown in an elegant experiment model utilising diets supplemented with either high-melanoidin bread crust, low-melanoidin bread crumb or a fibre-free melanoidin-free bread model that each diet had similarly bifidogenic effects compared with a control diet, indicating that the melanoidin content of bread is not responsible for the observed expansion of *Bifidobacteria* [83]. A previous report analysing the effect of bread crust melanoidins on the microbiome noted that there were large interindividual differences with regard to *Bifidobacteria*, but a consistent reduction in *Enterobacteria* across subjects [70].

Heat-treated diets have been found to be protective against inflammation and weight loss in a DSS-induced mouse model of colitis [88], which develops a dysbiotic microbiome [89]. Oral CML administration in the DSS-induced colitis mouse protected against weight loss and modulated the gut microbiome towards the control state, with a significant contraction of Proteobacteria [84], indicating that CML is in fact protective. Furthermore, the increased abundance of *Enterobacteriaceae* observed in DSS-induced colitis [89] was significantly reduced with CML administration [84]. Peritoneal dialysis

patients often have decreased *B. animalis*, a beneficial bacterium frequently included in probiotic supplementation formulas, compared with healthy controls. Dietary AGE restriction in a population of peritoneal dialysis patients resulted in decreased *B. animalis*, suggesting that dietary AGEs may promote the growth of this beneficial bacteria [74]. The contradictory effects on the microbiome and health outcomes observed with heat-treated processed foods have been ascribed to the heterogeneity of compounds that are formed during thermal treatment [90] and requires further elucidation.

10. Conclusions

AGEs are formed exogenously in foods, particularly foods that are cooked at high temperatures for prolonged periods of time using dry heat. Dietary intake of AGEs has increased considerably with the adoption of the modern processed diet, and excessive AGE intake is associated with inflammation, oxidative stress and the progression of CKD, at least in animal models. The absorption of dietary AGEs is limited, with evidence indicating that the majority of protein-bound AGEs pass through the gastrointestinal tract to the colon, where they can act as substrates for the gut microbiota. Recent research has highlighted the detrimental role the microbiome can have in a number of pathological conditions, and there are conflicting reports on whether dietary AGEs negatively or positively influence the composition of the microbiome. Furthermore, there have been mixed reports on the effects of high-AGE diets on the production of SCFAs. Alteration of the microbiome by dietary AGEs is of particular interest, given the prevalence of these compounds in the modern processed diet.

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