

Table S1. Primers list

	Forward (5'-3')	Reverse (3'-5')
<i>app-1</i>	CACGAAGGACCGATCGGAAT	TAAGCACTTGGCTGGCATGA
<i>asp-3</i>	TGCACAACAGATTTGATTGC	AGACAACATCGTTGTCAACG
<i>cdc-42</i>	GATCAAGTGCCTCGTTCGTTG	TGTGACGGCGTAATTGTCGA
<i>cpl-1</i>	GACGTCTCTTCGGTGA CTCC	TGGGTATCACGCCAGTCAAC
<i>cpr-5</i>	TGTGTCGACTCCTGCACTTC	TGGATTTGCTCGACCTTCTT
<i>ctsa-3.2</i>	TTGAAACACGGGCCATGGTA	CAAGTGGCACAAAGTGACCG
<i>djr-1.1</i>	GCACTCTTGAGCCATGGAGT	TGCCACTGACAACAACACGA
<i>djr-1.2</i>	AGAGCGTGTCACTTGTCACT	ACTCTGTCTGCTGATAACAACG
<i>glod-4</i>	GTGCCAAGACCATCGACTTT	TCTTGCTCCATCGTCCGTTAT
<i>gst-4</i>	TTTGATGCTCGTGCTCTTGC	AATGGGAAGCTGGCCAAATG
<i>hsp-16.2</i>	TGCAGAATCTCTCCATCTGAG T	TGGTTTAAACTGTGAGACGTTG A
<i>hsp-4</i>	AACCTACTCGTGCGTTGGAG	TCTCCGGAAAACGCAACGTA
<i>hsp-6</i>	TCGTGAACGTTTCAGCCAGA	CTCAGCGGCATTCTTTTCGG
<i>hsp-70</i>	CTACATGCAAAGCGATTGGA	GGCGTAGTCTTGTTCCTTC
<i>lap-1</i>	ATTGCTCGGACCTTCTGCAA	TCCTCGTTGGTGAGCTTGTC
<i>lec-1</i>	TCGTTCCACTTCAACCCACG	GAATGGGTTCTTTCCCTCGC
<i>lec-2</i>	TGCTGAAATGGCCCATGAGT	GGTGTGAAGGTTGACGGTGA
<i>lec-3</i>	CCTGGAAGTGTCTCGTCCGTA	TGGGTTCTTTCCCTCTCGTT
<i>lec-4</i>	CTCGACGCTGGACAGACATT	CACCACCTCCCTGGAGAAGA
<i>lec-5</i>	ATATGGCAGTTGCTGGTCTT	TTGACACGCTTTCCTTTTGG
<i>lec-6</i>	CGCCAATCCATTCCAGCAGA	ATGAGATGGACTCCGTGGGA
<i>lec-7</i>	CTCAGCTCCGACATAGCCAC	ACCGGGAACCGATGATCAA
<i>lec-8</i>	GCTCAAGCACCACGATCACT	TCAACTGGGAAACGGTGTGG
<i>lec-9</i>	CAGGAACCTTTGCTCTCCCC	ACCGGAGATCAGGTTGACGA
<i>lec-10</i>	TTCACCACTCGGAGCACTTC	AATCCGATGGCTTGAACCGA
<i>lec-11</i>	CGTTGCAATGACTGTTTCGAG	ATCGGAGTTGGAGCAACAGG
<i>lec-12</i>	AAAAACCGGATGGTTCAGCG	CGAATCGTTCTTCGTTGCC
<i>lgg-1</i>	TACCAGGACCATCACGAGGA	CGACCTCTCCTCCATACACAC
<i>lmp-2</i>	TCGCTGGACTCATTGTGCTC	ATGAGCTTCTGGGTTGACCA
<i>nas-11</i>	GACCCTGCCAACCAGTTTA	CGATCATTGCGAAGGTGCTG
<i>nas-28</i>	CAGCTGTCTGAGCTTCAAGG	AACGTATCACAGTTTGGCCC
<i>ostb-1</i>	AGGAGAAACTGCTGCTGTGA	ACGGTTAGCTCATGTCCTCG
<i>pept-1</i>	TGTCATCACGGCAGCAGAAA	AATCTCCAGCAGCAGTGGTC
<i>pmp-3</i>	AGTTCAGTTGCTCTCGTTGC	AGGCCAGCAAACATTCGAAG
<i>scav-1</i>	GTA CTGCAAGGACCCGA ACT	CCGACACGAATGCTGGTAGA
<i>scav-2</i>	AACCGCCTTTCTTGAAGTCG	TCCAAAATCGCTTCGCACAC
<i>scav-3</i>	TTACGATCCTGCTCTTCCGC	TGGCATCGTGACAAGTCGAT
<i>scav-4</i>	ACGGGTTCAAGGTGGAAGAC	ACATGGCCAGTTGGGGAAAT

<i>scav-5</i>	CTCAATAGCCGAGGGAGCAG	AGGATTTTTGCCGTGGGACT
<i>scav-6</i>	ATGCTTTACTGTCCGCTGCT	TTCGGCATAGCTGGAATCGG
<i>skr-1</i>	GCTGGGACGTCGAGTTTCTT	AGCAACAGTCTTGCAGGTAA
<i>skr-12</i>	TCCAATTGTCGATGCCCCAG	TCGATGGTGTAGCCGAGATT
<i>sod-3</i>	AAGCATCATGCCACCTACGT	CGCTGGTTGGAGAGCAATTG
<i>trx-1</i>	GAGACGCAATTGAGGCACTG	GAGCAGATACGTGCTCCAAC
<i>vha-6</i>	AGAATGCGTACACGAGAAAA	ATGCTCATCGTAGTCTGGAA
<i>vha-11</i>	GGCAGATTTGGTGAAGGCTG	CGACGAGAGTGTCGCGTATT
Y16B4A.2	TGCACAAATGCTTCCACCCA	ATGCAGCTTGGTTTTCTGTAG
ZK1307.8	GGAATGGAGTGGCCCAGAAG	ATTTTGCTGGCTCCGTGACT

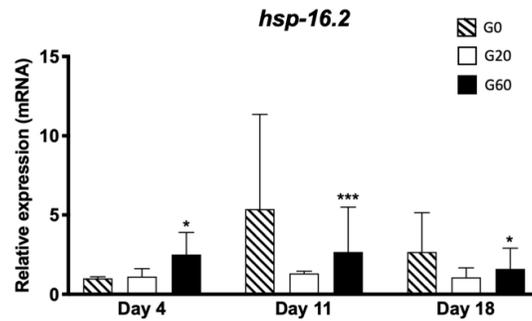


Figure S1: Relative expression of heat shock protein 16.2 (HSP-16.2) encoding gene. Young adult worms were fed for 4, 11 and 18 days with bacteria, which were pre-treated with 0 mM (G0), 20 mM (G20) or 60 mM (G60) of glyoxylic acid. Relative expression of genes coding for antioxidant, carbonyl stress and unfolded protein responses were analyzed by comparing all the conditions to the Ctrl at day 4. Data were expressed as means \pm standard deviation of 3 biological replicates. The *cdc-42* and *pmp-3* genes were used to normalize levels of gene expression. * $p \leq 0.05$, *** $p \leq 0.001$ G20 or G60 vs G0 for the same incubation time (Kruskal-Wallis, Dunn's multiple comparisons test).

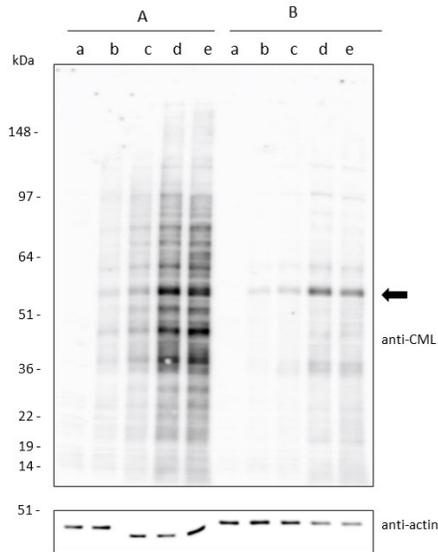


Figure S2: Digestion of bacterial glycosylated proteins by *C. elegans*. Young adult worms were fed with bacteria, which were incubated with 0 mM (G0) and 60 mM (G60) of glyoxylic acid. Some worms were only incubated with either G0 (a) or G60 (e) bacteria, while others were incubated with a mixed of the 2 bacteria types with a ratio G0:G60 of 7:1 (b), 3:1 (c) and 1:1 (d). After 4 days of culture, half of the worms were lysed (A) while the other half was incubated in the bacteria-free medium for 1 day before being washed and lysed (B). Western blots of worms' lysates were stained with anti-CML and anti-actin antibodies. The arrow shows a protein band poorly digested by the worms.

All results are representative of 3 independent experiments.

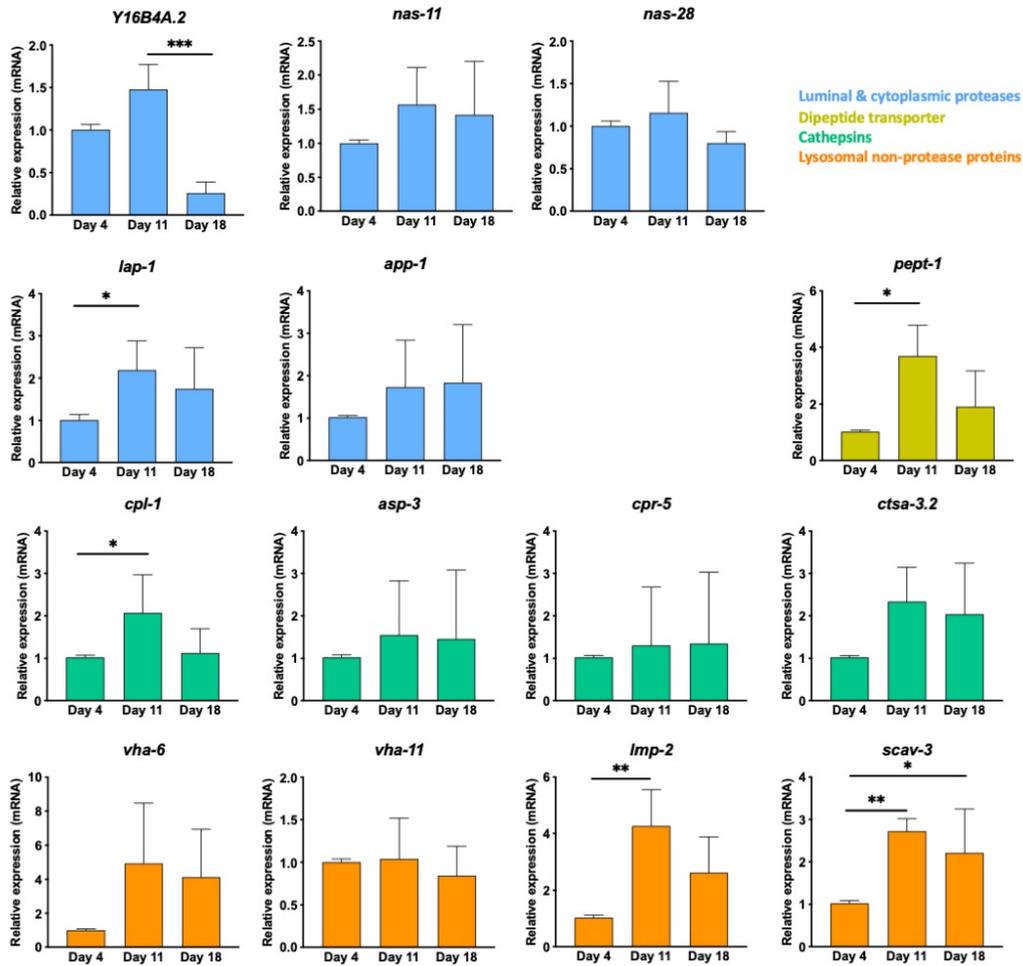


Figure S3: Impacts of dietary *Nε*-carboxymethyllysine (dCML) exposure on transcriptional regulation of some digestive enzymes and lysosomal proteins. Young adult worms were fed with bacteria, which were incubated with 0 mM of glyoxylic acid. After 4, 11, and 18 days of culture, relative expression of genes coding for luminal and/or cytoplasmic proteases, lysosomal cathepsins and lysosomal non-protease proteins were analyzed by comparing all the conditions to the day 4. Data were expressed as means \pm standard deviation of 3 biological replicates. The *cdc-42* and *pmp-3* genes were used to normalize levels of gene expression.

* $p \leq 0.05$, ** $p \leq 0.01$, *** $p \leq 0.001$ (Kruskal–Wallis, Dunn’s multiple comparisons test).

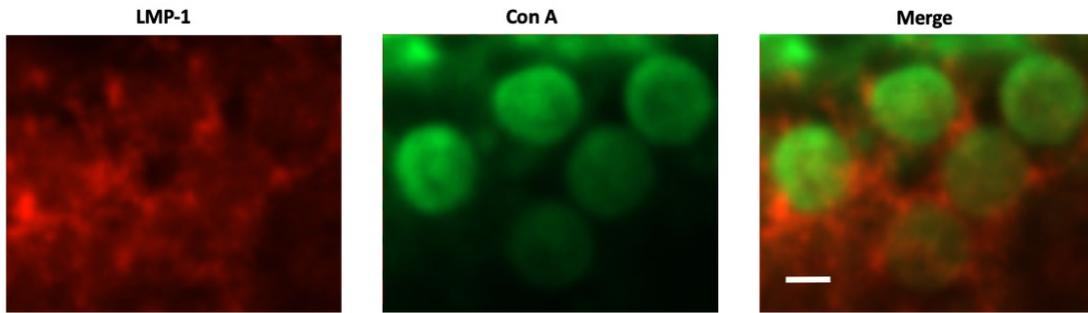


Figure S4: Mapping of lysosomal epitope and concanavalin A (Con A) staining on intestinal endosomes. Immunohistochemical assays were performed on worms fed for 4 days with bacteria, which were pretreated with 60 mM of glyoxylic acid, using Con A and anti-lysosome-associated membrane protein homolog 1 (LMP-1) antibody. Fluorescence micrographs with the mentioned lectin or antibody are shown. Scale bars = 2.5 μm .

All results are representative of 3 independent experiments.