

## **Supplementary Materials: Association of Common Variants of TNFSF13 and TNFRSF13B Genes with CLL Risk and Clinical Picture, as well as Expression of Their Products – APRIL and TACI Molecules**

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**Supplementary Table S1.** Genotype distribution of the *APRIL* (*TNFSF13*; 17p13.1) polymorphisms in patients and controls.

<i>APRIL</i>		Patients (N=439)		Controls (N=477)		OR	CI95%	Patients vs. Controls
<i>TNFSF13</i>								
polymorphisms		N	%	N	%			
<b>rs11552708</b>	GG	385	87.70	417	87.42	1*		
	GA	53	12.10	57	11.95	1.01	0.68;1.50	$\chi^2_{df=2} = 0.85$
Gly67Arg	AA	1	0.20	3	0.63	0.46	0.07;3.16	p=0.782
<b>HWE</b>		p=0.833		p=0.448				
		f=0.03		f=0.031				
		CI95%=-0.08;0.06		CI95%=-0.06;0.14				
<b>rs4968210</b>	GG	166	37.80	157	32.90	1*		$\chi^2_{df=2} = 2.58$
	GA	198	45.10	237	49.70	0.79	0.59;1.05	p=0.275
Intron 5	AA	75	17.10	83	17.40	0.86	0.58;1.25	
<b>HWE</b>		p=0.232		p=0.778				
		f=0.057		f=0.020				
		CI95%=-0.04;0.15		CI95%=-0.11;0.07				
<b>rs6608</b>	CC	350	79.70	385	80.71	1*		
	CT	85	19.40	88	18.45	1.06	0.76;1.48	$\chi^2_{df=2} = 0.14$
3'UTR	TT	4	0.90	4	0.84	1.10	0.30;4.10	p=0.92
<b>HWE</b>		p=0.804		p=0.947				
		f=0.02		f=0.02				
		CI95%=-0.10;0.07		CI95%=-0.09;0.07				

Abbreviations : OR, odds ratio; CI, confidence intervals; HWE, Test for Hardy-Weinberg equilibrium; *f*, departure from HWE; \* the reference group;

**Supplementary Table S2.** Genotype distribution of the *TACI* (*TNFRSF13B*; 17p11.2) polymorphisms in patients and controls.

<i>TACI</i>		Patients (N=439)		Controls (N=477)		OR	CI95%	Patients vs. Controls		
<i>TNFRSF13B</i>										
polymorphisms		N	%	N	%					
<b>rs12051889</b>	CC	385	87.70	416	87.20	1*		$\chi^2_{df=2} = 1.35$ p=0.515		
	CT	51	11.60	60	12.60	0.92	0.62;1.37			
	TT	3	0.70	1	0.20	2.52	0.37;17.16			
<b>HWE</b>		p=0.414		p=0.912						
		f=0.04		f=-0.03						
		CI95%=-0.06;0.16		CI95%=-0.08;0.045						
<b>rs8072293<sup>a</sup></b>	TT	211	48.10	236	49.48	1*		$\chi^2_{df=2} = 0.25$ p=0.881		
	Thr27Thr	TC	195	44.40	204	42.77	1.07		0.82;1.40	
	CC	33	7.50	37	7.75	1.00	0.60;1.65			
<b>HWE</b>		p=0.209		p=0.506						
		f=-0.06		f=-0.036						
		CI95%=-0.15;0.03		CI95%=-0.12;0.053						
<b>rs11656106</b>	TT	206	46.90	238	49.90	1*		$\chi^2_{df=2} = 0.87$ p=0.649		
	TC	194	44.20	197	41.30	1.14	0.87;1.49			
	CC	39	8.90	42	8.80	1.07	0.67;1.72			
<b>HWE</b>		p=0.576		p=0.912						
		f=-0.03		f=0.006						
		CI95%=-0.12;0.06		CI95%=-0.08;0.09						
<b>rs11078355</b>	AA	157	35.80	158	33.10	158	1*	$\chi^2_{df=2} = 2.03$ p=0.362		
	Ser277Ser	AG	206	46.90	246	51.60	246		0.84	0.63;1.12
	GG	76	17.30	73	15.30	73	1.05		0.71;1.55	
<b>HWE</b>		p=0.554		p=0.185						
		f=0.028		f=-0.06						
		CI95%=-0.07;0.12		CI95%=-0.16;0.021						

Abbreviations : OR, odds ratio; CI, confidence intervals; HWE, Test for Hardy-Weinberg equilibrium; *f*, <sup>a</sup> T>C according to the frequency of alleles, \* the reference group;

**Supplementary Table S3.** Average values of APRIL MFI in CD19<sup>+</sup> CLL cells in relation to *APRIL* SNP genotypes.

<i>APRIL</i> SNP	Genotype			F-test	p-value	LSD*
MFI APRIL <sup>+</sup> CD19 <sup>+</sup> CLL cells	AA	GA	GG			
<b>rs3803800</b>	<b>AA</b>	<b>GA</b>	<b>GG</b>			
average	81.49	86.76	80.80	0.124	0.883	(AA.GA.GG)
n	8	19	45			
<b>rs11552708</b>	<b>AA</b>	<b>GA</b>	<b>GG</b>			
average	59.63	91.63	81.71	0.36	0.702	(AA.GA.GG)
n	1	8	63			
<b>rs4968210</b>	<b>AA</b>	<b>GA</b>	<b>GG</b>			
average	79.6	75.49	92.32	1.159	0.32	(AA.GA.GG)
n	12	32	28			
<b>rs6608</b>	<b>CC</b>	<b>CT</b>	<b>TT</b>			
average	78.11	104.34	59.63	2.031	0.139	(CC.CT.TT)
n	57	14	1			

\*The ANOVA test with different (homogeneous) groups identified based on Fisher's LSD *post hoc* test was applied.

**Supplementary Table S4.** Average percentage of APRIL<sup>+</sup> CD19<sup>+</sup> CLL cells according to APRIL SNP genotypes.

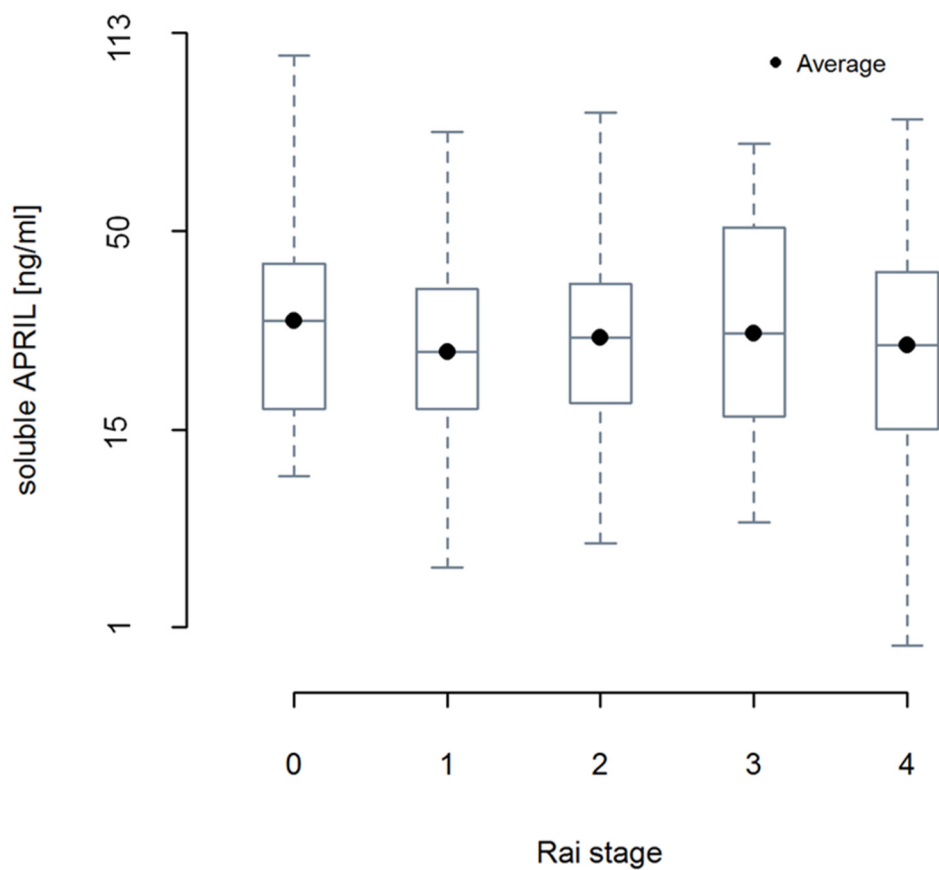
APRIL SNP % CD19 <sup>+</sup> APRIL <sup>+</sup> CLL cells	Genotype			F-test	p-value	LSD*
<b>rs3803800</b>	<b>AA</b>	<b>GA</b>	<b>GG</b>			
average	4.91	8.14	11.18	2.277	0.11	(AA.GA.GG)
n	8	19	45			
<b>rs11552708</b>	<b>AA</b>	<b>GA</b>	<b>GG</b>			
average	14.16	6.52	9.79	0.58	0.562	(AA.GA.GG)
n	1	8	63			
<b>rs4968210</b>	<b>AA</b>	<b>GA</b>	<b>GG</b>			
average	12.07	12.29	6.17	3.802	<b>0.0271</b>	<b>GG (AA.GA)</b>
n	12	32	28			
<b>rs6608</b>	<b>CC</b>	<b>CT</b>	<b>TT</b>			
average	10.08	6.9	14.16	0.774	0.465	(CC.CT.TT)
n	57	14	1			

\*The ANOVA test with different (homogeneous) groups identified based on Fisher's LSD *post hoc* test was applied.

**Supplementary Table S5.** Average levels of plasma sAPRIL according to *APRIL* SNP genotypes.

<i>APRIL</i> SNP sAPRIL level [ng/ml]	Genotype			F-test	p-value	LSD
<b>rs3803800</b>	<b>AA</b>	<b>GA</b>	<b>GG</b>			
average	31.80	25.69	29.84	0.699	0.499	(AA.GA.GG)
n	13	43	66			
<b>rs11552708</b>	<b>AA</b>	<b>GA</b>	<b>GG</b>			
average	-	23.28	29.09	0.86	0.354	(GA.GG)
n	0	11	111			
<b>rs4968210</b>	<b>AA</b>	<b>GA</b>	<b>GG</b>			
average	32.47	29.5	25.27	0.921	0.401	(AA.GA.GG)
n	20	62	40			
<b>rs6608</b>	<b>CC</b>	<b>CT</b>	<b>TT</b>			
average	29.56	25.5	14.1	1.056	0.351	(CC.CT.TT)
n	99	21	2			

\*The ANOVA test with different (homogeneous) groups identified based on Fisher's LSD *post hoc* test was applied.



**Supplementary Figure S1.** Levels of sAPRIL in groups of CLL patients according to the Rai stage

Box-and-whiskers plots in standard manner with expected value as central points, 1st and 3rd quartiles and min-max values. Average levels of sAPRIL in groups of patients divided according to Rai stage (0, 1, 2, 3, 4) are 31.33, 25.84, 28.3, 29.06, 27.02 ng/ml, respectively. Figure shows average levels of sAPRIL as well as 1st and 3rd quartiles with min and max levels in each group of Rai stage. There is no relationship between Rai stage and sAPRIL level.

**Supplementary Table S6.** ANOVA table with linear contrast for IgA levels [g/l] in CLL patients according to rs3803800G>A genotype.

Box-Cox transformation	rs3803800G>A			
	$\lambda = -0.096$	AA	GA	GG
<i>n</i>	16	68	104	
Mean	0.616	0.304	0.130	
Standard deviation	0.417	0.72	0.803	
Variance homogeneity	$\chi^2_{df=3} = 6.087; p = 0.04767$ (Fligner-Killeen test)*			
ANOVA table				
Source of variability	Degrees of freedom	Sum of squares	Mean of squares	F-test p-value
Rai stage (linear contrast)	1	2.385	2.385	F=4.247
Error	186	104.45	0.562	p=0.0178*

\* F-distribution and p-value estimated numerically based on 10000 bootstrap samples.

Please note, that ANOVA was performed on data transformed with Box-Cox transformation ( $\lambda = -0.096$ ). There is minor heterogeneity of variances between groups and correlation between means and SD, so the distribution of the F statistic and p-value were estimated with bootstrap method.



**Supplementary Table S7.** ANOVA table with linear contrast for CD19<sup>+</sup>TACI<sup>+</sup> percentage according to Rai stage.

Box-Cox transformation	Rai stage			
	0	1	2	3-4
$\lambda = 0.146$				
<i>n</i>	50	43	40	13
Mean	2.54	2.73	2.88	3.06
Standard deviation	1.82	2.08	2.03	2.59
Variance homogeneity	$\chi^2_{df=3} = 4.384; p = 0.223$ (Fligner-Killeen test)			
ANOVA table				
Source of variability	Degrees of freedom	Sum of squares	Mean of squares	F-test p-value
Rai stage (linear contrast)	1	4.12	4.12	<b>F=1.0116</b>
Error	144	586.16	4.07	<b>p=0.3162</b>

Please note, that ANOVA was performed on data transformed with Box-Cox transformation ( $\lambda = 0.146$ ). The average levels of CD19<sup>+</sup>TACI<sup>+</sup> percentage of leukemic cells in groups of patients with Rai stage 0, 1, 2, and 3-4 are 8.7, 10.0, 11.1 and 12.5%, respectively. Pearson's correlation coefficient between these averages and Rai scores 0, 1, 2, 3 is  $r_{altering} = 0.998$ . It means that there is a strong linear relationship between Rai stage and expected percentage value of CD19<sup>+</sup>TACI<sup>+</sup> cells, but this coefficient ignores a huge variability of (%) CD19<sup>+</sup>TACI<sup>+</sup> cells within groups with particular Rai score. When this additional within-group variation is incorporated, the effect size correlation between Rai stage and (%) of CD19<sup>+</sup>TACI<sup>+</sup> cells is  $r_{effect\ size} = 0.084$ . This within-group variability is a reason why mean of squares for the linear contrast in ANOVA is practically the same as mean of squares for error, thus  $F_{1;144}=1.0116; p=0.3162$ .

**Supplementary Data 8.** Prediction of functional effects for: *TNFSF13* rs3803800G>A, rs4968210G>A and *TNFRSF13B* rs4985726C>G and rs11078355A>G.

Since elucidating the function of risk variants and variants associated with phenotypic features is an important step towards a better understanding of the biological processes involved in disease development and outcomes, we used publicly available sources, described below to examine a potential functional relevance of rs3803800G>A, rs4968210G>A, rs4985726C>G, and rs11078355A>G genetic variants.

rs3803800G>A, a missense variant of *TNFSF13* (Ser96Asn, exon 2) and *TNFSF12-TNFSF13* (Ser176Asn, exon 7) has been predicted to be a benign variant by PolyPhen2 (benign: 0.036) and as a tolerated substitution by SIFT (score:0.597, median: 2.93). According to HaploReg tool (v4.1) [1,2] this element is conserved and does not exist in LD with other SNPs (Supplementary Figure S2A).

Of note, *TNFSF13* is located in close proximity to the *TNFSF12* gene for another member of the *TNFSF*, namely the TNF-like weak inducer of apoptosis (TWEAK, *TNFSF12*). The adjacent localisation and the same transcriptional direction of these two genes enables the production of mRNA molecule (*TNFSF12-TNFSF13*) and protein (TWE-PRIL) consisted of the cytoplasmic/transmembrane and stalk domains of TWEAK and receptor binding domains of APRIL due to and intergenic splicing event.

Given the data provided by Bojarska-Junak et al. [3] showed a higher expression of APRIL mRNA and intracellular APRIL in peripheral blood (PB) CD19<sup>+</sup> leukemic cells than in PB CD19<sup>+</sup> cells isolated from the control group, we checked if rs3803800 localizes to any regulatory region by applying the ENCODE dataset [4-5]. One matching candidate is the Cis Regulatory Element (cCRE) accession number EH38E1844485 (hg38), which was shown for rs3803800G>A with a proximal enhancer like signature (Supplementary Figure S2B,C, respectively). This cCRE was associated with *TNFSF13* expression and according to ENCODE the IKZF1 transcription factor may bind within this cCRE, which was observed for GM12878 cell line. These data suggest that rs3803800 may potentially localize to regulatory region. The Supplementary Figure S2B,C presents rs3803800 surrounded by regulatory elements and TF binding sites (hg38). We have also performed an additional examination with application of GTEx [6] multi-tissue eQTLs analysis to see if this variant affect expression in other cells and tissues (Supplementary Figure S2D). The rs3803800 appeared to be significantly associated with *TNFSF13* expression in many tissues (Meta-Analysis RE2: p-value=5.2×10<sup>-55</sup>). This analysis showed that the minor allele A of rs3803800 (reference allele in GTEx) is in the majority of tissues associated with higher *TNFSF13* expression.

rs4968210G>A variant constitutes G to A substitution in intron 5 of *TNFSF12-TNFSF13* transcript as well as of *TNFSF12* transcript. As described earlier, we observed that A allele is associated with a higher average percentage of CD19<sup>+</sup>APRIL<sup>+</sup> CLL cells. Taking this into consideration we employed ENCODE [4-5] to check if this variant or any variant in LD with it (Supplementary Figure S3a) [1-2] localize to regulatory elements. According to ENCODE, rs4968210 variant does not overlap any cCRE, but is located 981 bp from cCRE EH38E1844474 (hg38) which is predicted to have a distal enhancer like signature inter alia in B cells and GM12878 (Supplementary Figure S3B and 3C) cell line and is associated with *TNFSF12-TNFSF13* mRNA expression. Additionally, another SNP in LD with rs4968210 (Supplementary Figure S3A) [1-2] namely rs12942590 is located within 221 bp distance of this cCRE (Supplementary Figure S3B and C). Accordingly, the transcription factors important for B cell biology such as EBF1, PAX5, SPI1, IKZF1, IKZF2, and BHLHE40 were shown to bind in this region (Supplementary Figure S3B and 3C).

The ENCODE data suggest that the impact of rs4968210 observed by us on phenotype of CD19<sup>+</sup> B cells in relation to APRIL may be associated with the fact that rs4968210 and rs12942590 are located near potential regulatory element associated with B cells biology and *TNFSF12-TNFSF13* expression. To further evaluate the association between rs4968210 and *TNFSF12-TNFSF13* expression we performed an additional analysis with application of GTEx multi-tissue eQTLs analysis to see if this variant affect expression in other cells and tissues (Supplementary Figure S3D). The rs4968210 turned out to be significantly associated with *TNFSF12* expression in many types of tissues (Meta -Analysis RE2: p-value=1.2x10<sup>-94</sup>). This analysis indicates that the rs4968210G allele (reference allele in GTEx) is associated with higher *TNFSF12* (*TWEAK*) expression. in the majority of tissues for that the data were available. However, in EBV-transformed lymphocytes an opposite effect can be observed, unfortunately this association was not significant. This is in line with our observation that patients with rs4968210GA and rs4968210AA genotypes had higher average percentage of CD19<sup>+</sup>APRIL<sup>+</sup> CLL cells. The rs4968210 was also associated with *TNFSF13* eQTL expression in multiple tissues (Meta -Analysis RE2: p-value=1.9x10<sup>-11</sup>). The result of this analysis suggests that rs4968210 may be associated with allele specific expression which seems to be tissue specific (Supplementary Figure S3E).

The Supplementary Figure S3A presents SNPs in LD with rs4968210 and Figure S3B,C present 4968210 and rs12942590 surrounded by regulatory elements and TF binding sites (hg38) in PB B cells as well as in GM12878 cell line, respectively.

rs4985726C>G variant is located in intron 1 of *TNFRSF13B* gene. According to HaploReg tool (v4.1) [1-2] this variant exists in LD with 10 intronic SNPs of *TNFRSF13B* and one missense rs34562254G>A (Pro251Leu) variant of *TNFRSF13B* (Supplementary Figure S4A). The rs34562254G>A (Pro251Leu) variant was predicted to be a possibly damaging variant by PolyPhen2 (Exome Variant server: 0.728; Ensembl: 0.476). Rs4985726C>G did not overlap with any cCRE both in PB B cells as well as in GM12878 cell line (Supplementary Figure S4B and S4C, respectively). Accordingly, we did not observe association between this variant and TACI expression in PB CLL cells. Similarly, analysis with GTEx portal [6] did not reveal significant eQTLs associated with rs4985726C>G. However, ENCODE [4-5] analysis for SNPs in LD with rs4985726 revealed data which may provide some potential explanation for the association of rs4985726 with the susceptibility to CLL risk observed by us. One of such SNPs, namely, rs57382045 did not overlap with any cCRE, however it is located 299 bp from EH38E1849614 cCRE (Supplementary Figure S4D,E). This cCRE was predicted to have distal enhancer-like signature in B cells with many TFs shown to bind in this region among others IKZF1 and IKZF2 (Supplementary Figure S4D). What is interesting, EH38E1849614 cCRE seems to be inactive (low DNase) in GM12878 cell line (Supplementary Figure S4E). The Supplementary Figure 4A presents SNPs in LD with rs4985726, Supplementary Figures S4B-E present rs4985726 and rs57382045, chromatin marks in B cells, and GM12878 cell line (hg38).

rs11078355A>G is a synonymous variant (exon 5, Ser277Ser) of the *TNFRSF13B* gene. According to HaploReg tool (v4.1) [1-2] this variant exists in LD with five SNPs (Supplementary Figure S6A). Analysis with application of Human Splicing Finder (HSF) [7] showed that this variant may potentially cause alternation of splicing, by affecting the binding site for 9G8 (also known as SRSF7) splicing factor (Supplementary Figure S5A). Additionally, our studies revealed an association between genotypes of rs11078355A>G and expression of TACI receptor on CD19<sup>+</sup>TACI<sup>+</sup> leukemic cells. Given that, we checked if rs11078355 and/or any variant in LD with it localize to any cCREs with possible impact on B cell biology. We did not find such cCREs (Supplementary Figure S6B,C). Application of GTEx portal [6] did not reveal significant eQTLs and sQTLs for rs11078355A>G

associated with *TNFRSF13B* mRNA levels. The Supplementary Figure S6 displays rs11078355 and the surrounding *TNFRSF13B* gene region as well as SNPs in LD with rs11078355.

A

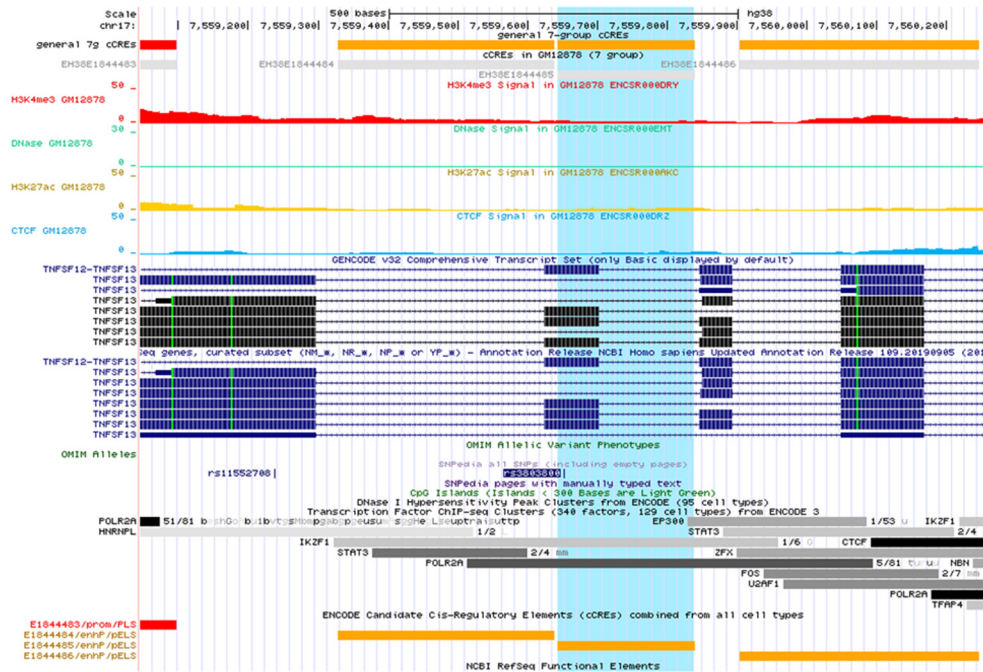
Query SNP: **rs3803800** and variants with  $r^2 \geq 0.8$

chr	pos (hg38)	LD (r <sup>2</sup> )	LD (D')	variant	Ref Allele	AFR freq	AMR freq	ASN freq	EUR freq	SIPhy cons	Promoter histone marks	Enhancer histone marks	DNAse	Proteins bound	Motifs changed	NHGRI/EBI GWAS hits	GRASP QTL hits	Selected eQTL hits	GENCODE genes	#SNP func annot
17	7559652	1	1	<b>rs3803800</b>	A	0.29	0.70	0.67	0.78		11 issues	17 issues	8 issues		ERalpha-MZF1:1-4	4 hits	10 hits	4 hits	TNFSF13	missense

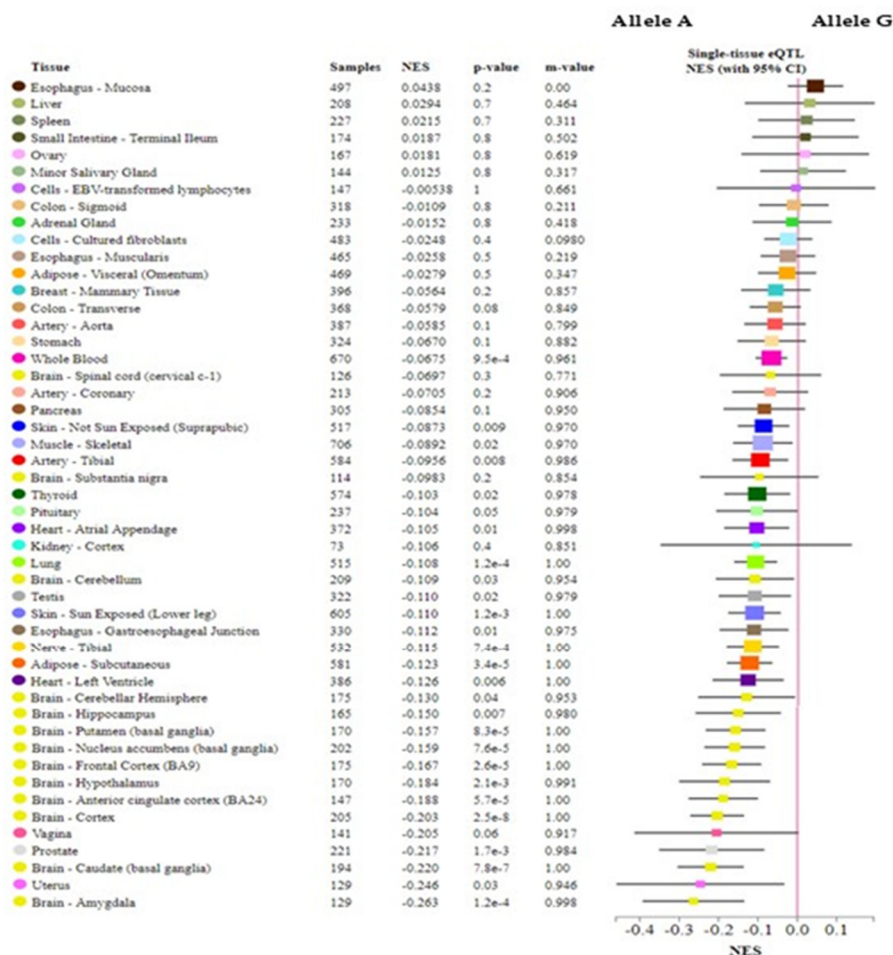
B



C



D



Supplementary Figure S2. *In silico* analysis - rs3803800 of *TNFSF13*.

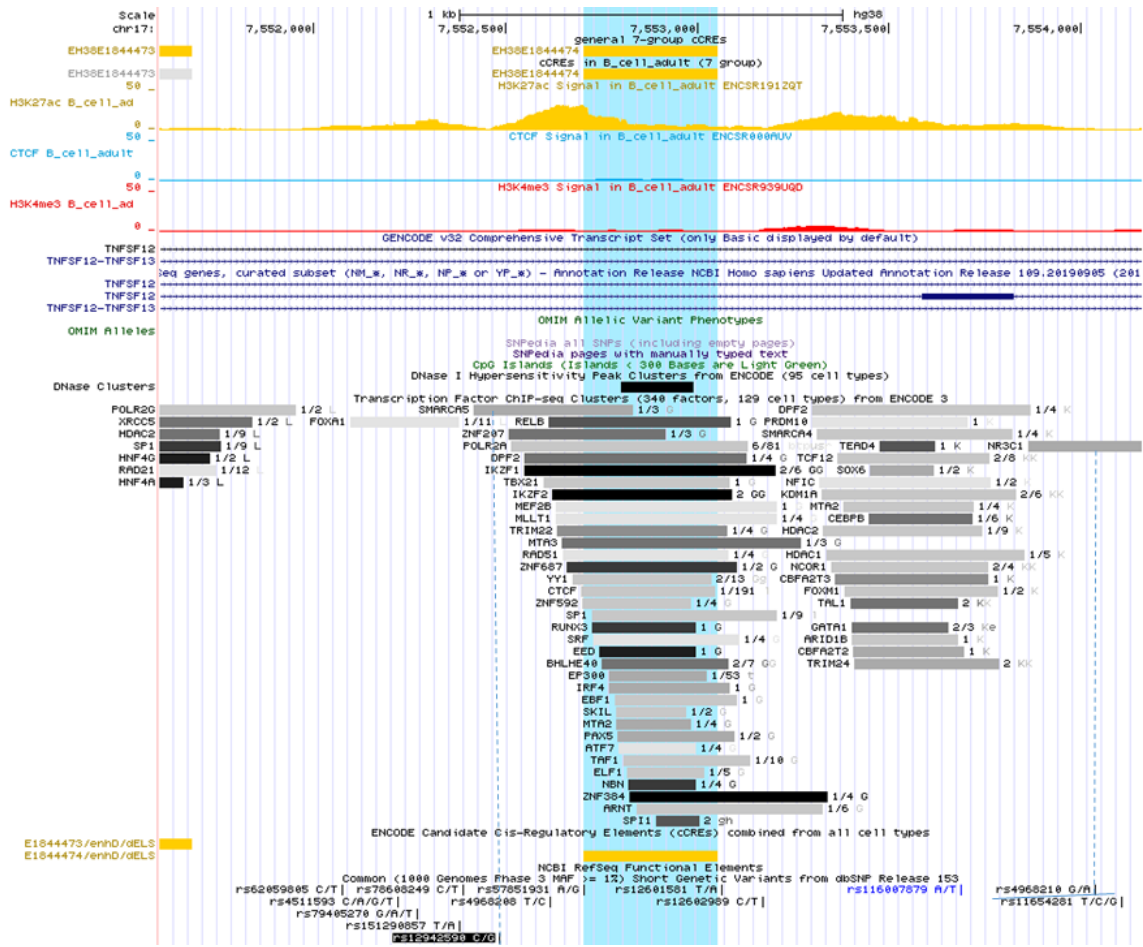
(A) The result of analysis performed with application of HaploReg v4.1 tool (<https://pubs.broadinstitute.org/mammals/haploreg/haploreg.php>) (screen shot) [1-2] (B) Location of rs3803800 (highlighted) of *TNFSF13* gene in relation to data from ENCODE project showing potential regulatory element-binding sites upstream and downstream of rs3803800 in B cells and GM12878 cell lines (C). Figures were downloaded from the UCSC genome browser (<http://genome.ucsc.edu>) [8-10]. A view of the region surrounding the rs3803800 includes also the representation of the ChIP-seq data from the ENCODE project, representing transcription factors (TFs) binding sites (black boxes denote the sites with stronger evidence for TFs binding in that region) (D) GTEx multi-tissue eQTLs analysis of association between rs3803800 and *TNFSF13* expression (Meta-Analysis RE2: p-value=5.2x10<sup>-55</sup>).

A

Query SNP: **rs4968210** and variants with  $r^2 \geq 0.8$

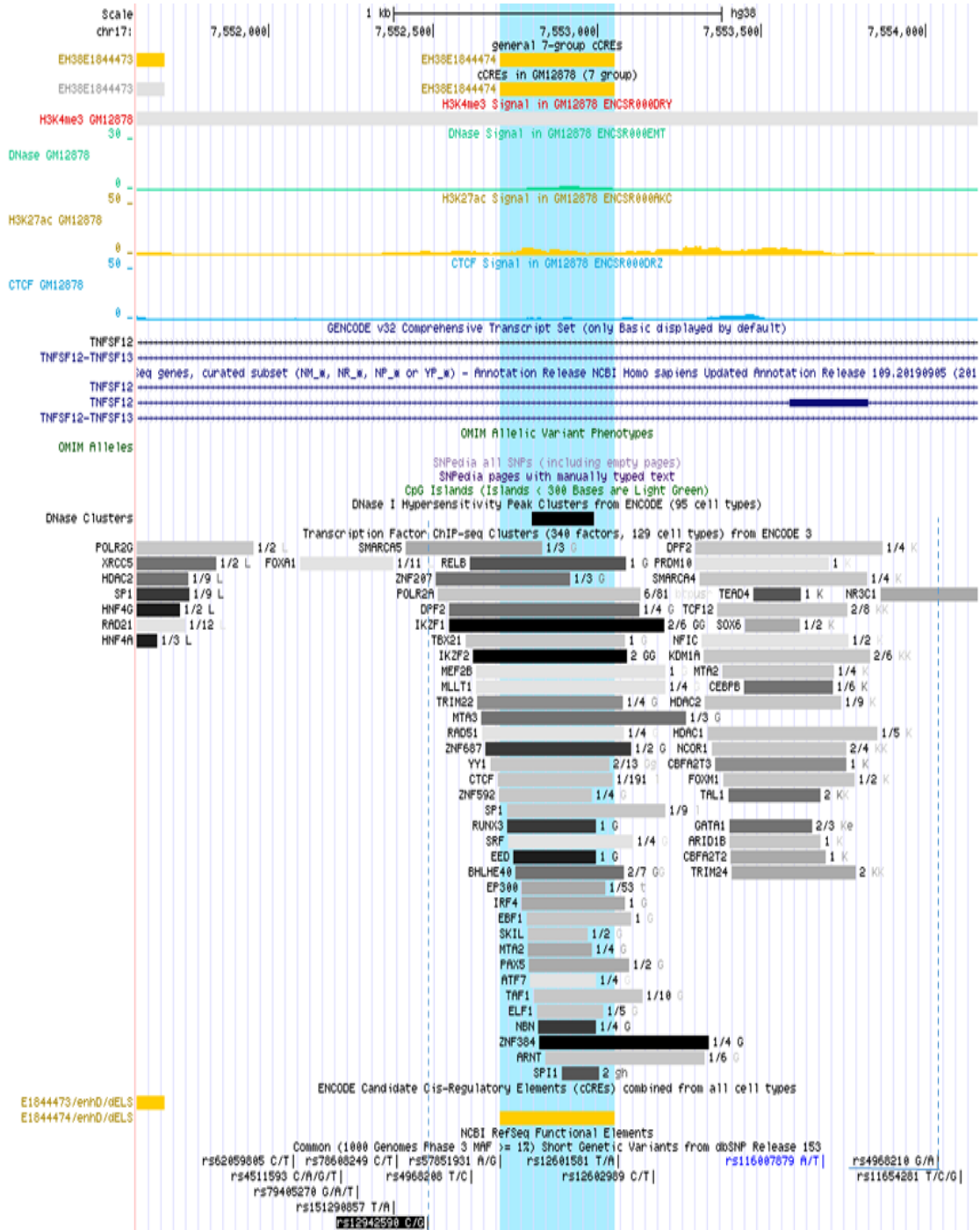
chr	pos (hg38)	LD (r <sup>2</sup> )	LD (D)	variant	Ref Alt	AFR freq	AMR freq	ASN freq	EUR freq	SPY freq	Promoter histone marks	Enhancer histone marks	DNase	Proteins bound	Motifs changed	NIH/IEB GWAS hits	GRASP QTL hits	Selected eQTL hits	GENCODE genes	dbSNP func annot
17	7538942	0.8	0.95	rs4968216	G A	0.02	0.33	0.27	0.40			5 Issues	BRN1, LING	CFOS	LBP-1, LBP-9			21 hits	1.7kb 5' of Y_RNA	
17	7541585	0.82	0.97	rs4968221	G A	0.12	0.33	0.27	0.40						9 altered motifs			19 hits	4.5kb 5' of Y_RNA	
17	7542075	0.87	0.95	rs148852655	A 20-mer	0.25	0.47	0.47	0.42						YY1			16 hits	4.9kb 5' of Y_RNA	
17	7552484	0.88	1	rs12842590	C G	0.02	0.32	0.28	0.40			5 Issues			4 altered motifs			22 hits	TNFSF12	intronic
17	7554035	1	1	rs4968210	G A	0.45	0.51	0.46	0.43		BLD	8 Issues	CRVX	GR	CCNT2, HP1-site-factor		5 hits	20 hits	TNFSF12	intronic
17	7554376	0.85	0.99	rs118271455	C T	0.14	0.33	0.28	0.39		BLD				E2F, Mat, SREBP			20 hits	TNFSF12	intronic
17	7555265	0.83	-0.99	rs12837543	A T	0.23	0.45	0.41	0.53		BLD, SKIN						3 hits	14 hits	TNFSF12	intronic
17	7557368	0.87	0.99	rs1128953	G A	0.02	0.32	0.28	0.40		5 Issues	17 Issues	8 Issues	RAD21	SREBP		1 hit	23 hits	TNFSF12	3'UTR

B

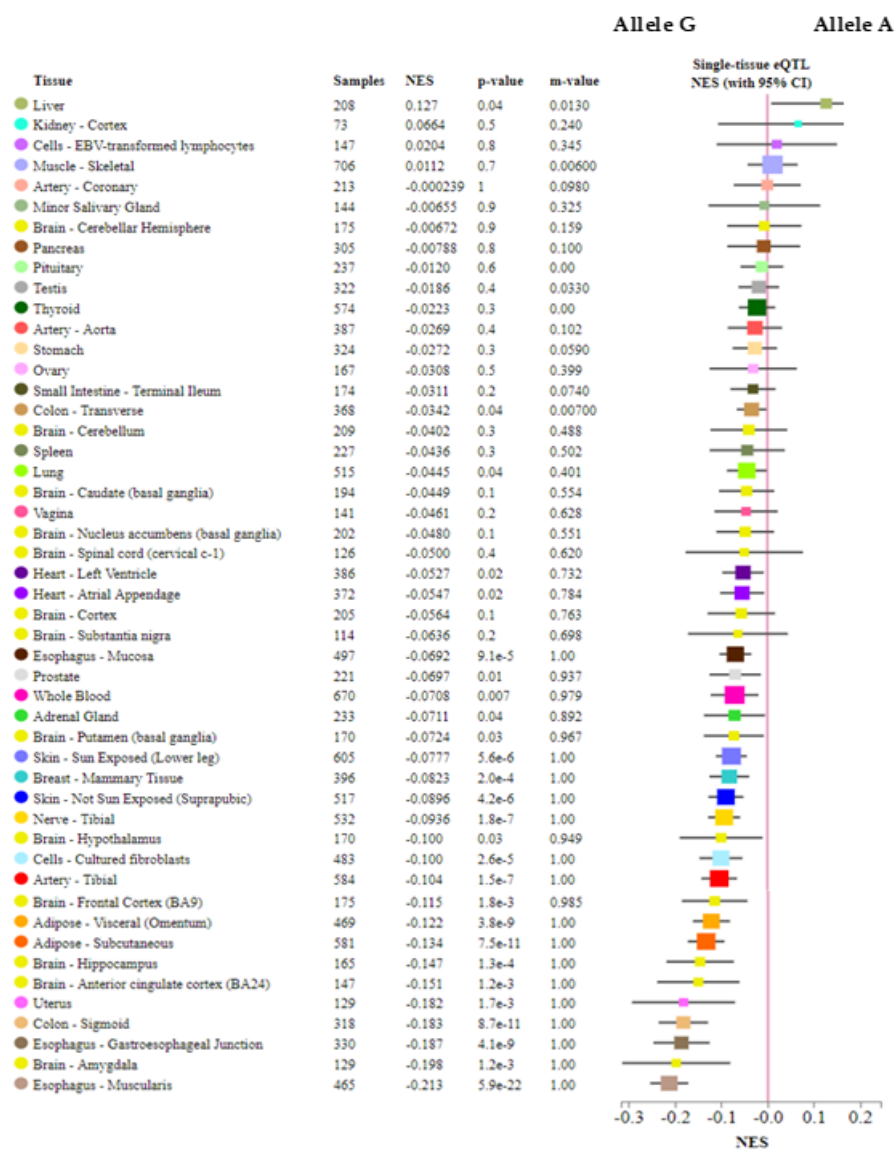




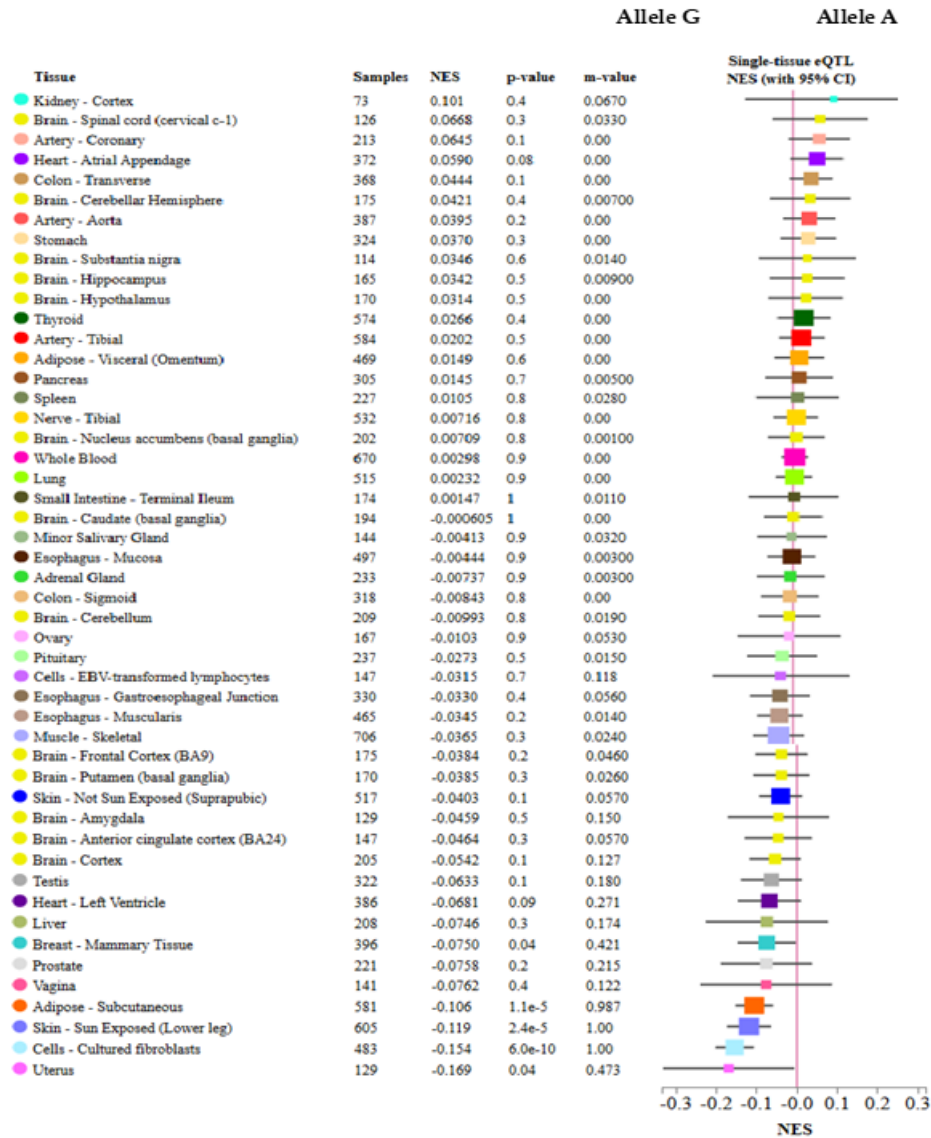
C



D



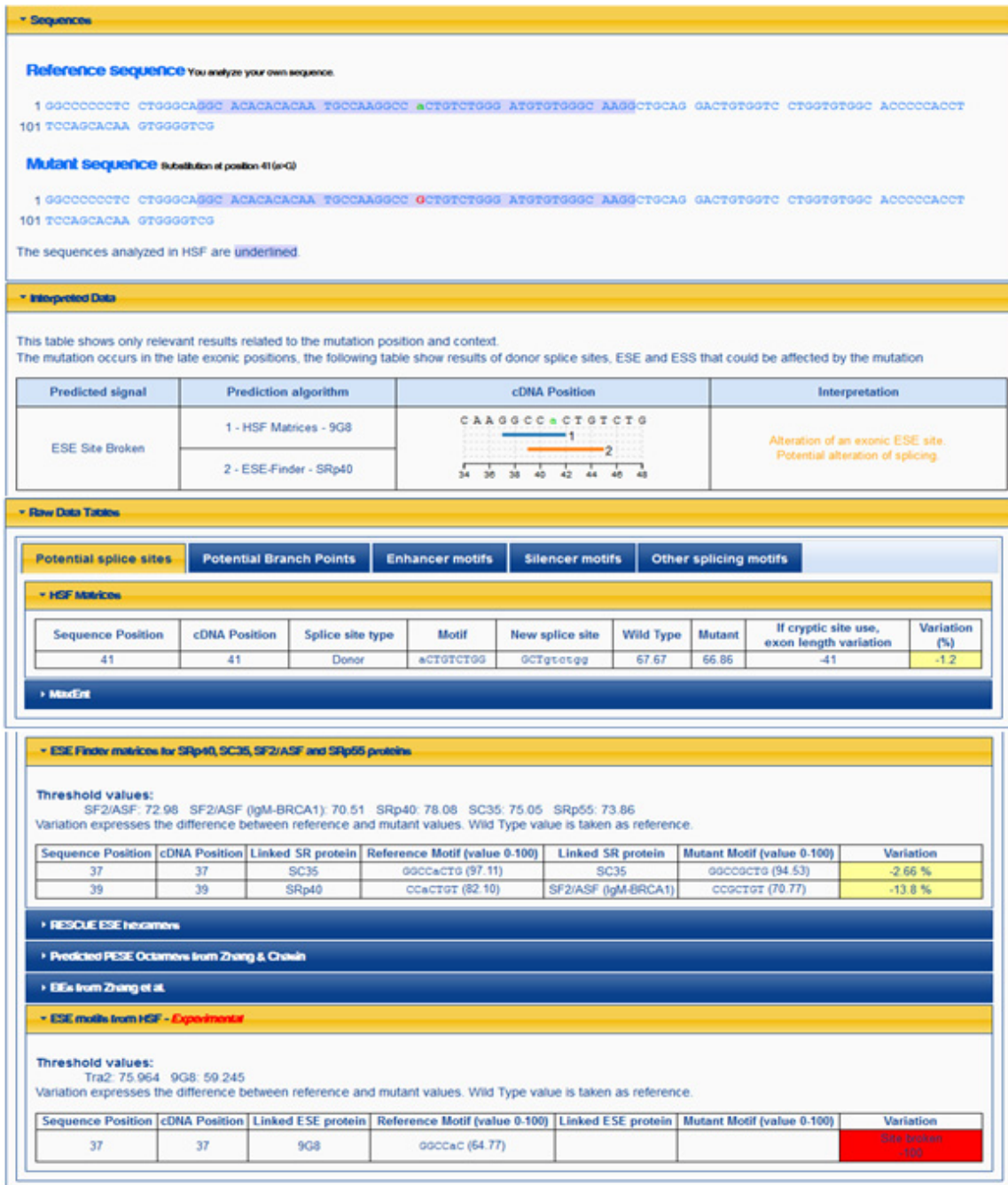
E



Supplementary Figure S3. In silico analysis - rs4968210 of TNFSF13.

(A) The result of analysis performed with application of HaploReg v4.1 tool (screen shot) [1-2]. (B) Location of rs4968210 (underlined) and rs12942590 (highlighted) (LD with rs4968210) of TNFSF13 gene in relation to data from ENCODE project showing potential regulatory element-binding in close proximity to rs4968210 in B cells and (C) in GM12878 cell line. A view of the region surrounding the rs4968210 and rs12942590 includes also the representation of the ChIP-seq data from the ENCODE project, representing transcription factors (TFs) binding sites (black boxes denote the sites with stronger evidence for TFs binding in that region). Figures were downloaded from the UCSC genome browser (<http://genome.ucsc.edu>) [8-10] (D) GTEx multi-tissue eQTLs analysis of association between rs4968210 and TNFSF12 expression (Meta-Analysis RE2: p-value=1.2x10<sup>-94</sup>) (E)

GTEx multi-tissue eQTLs analysis of association between rs4968210 and TNFSF13 expression (**Meta -Analysis RE2: p-value=1.9x10<sup>-11</sup>**).

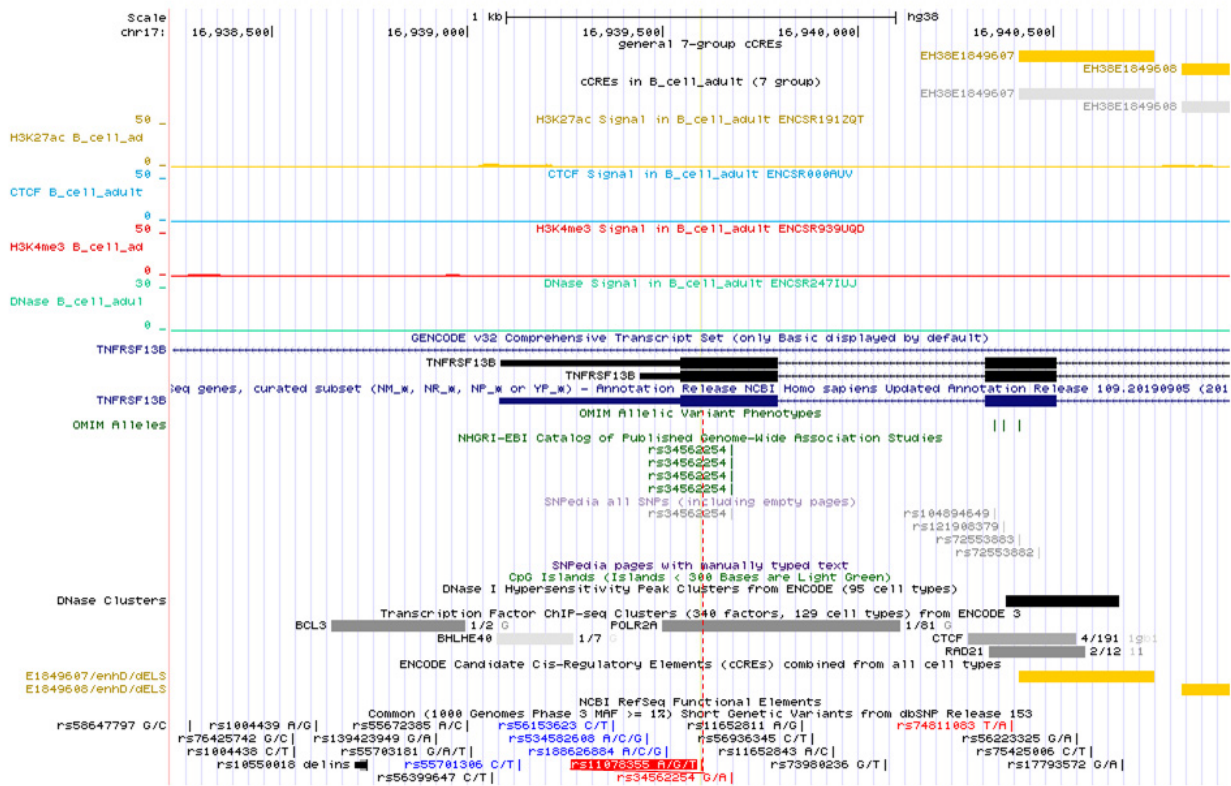


**Supplementary Figure S4.** Analysis of potential effects of rs11078355 A>G on splicing of *TNFRSF13B* gene. A Human Splicing Finder (HSF version 3.1) [7] prediction showing that allele G of rs11078355 may potentially cause alternation of an exonic ESE site (screen shot).

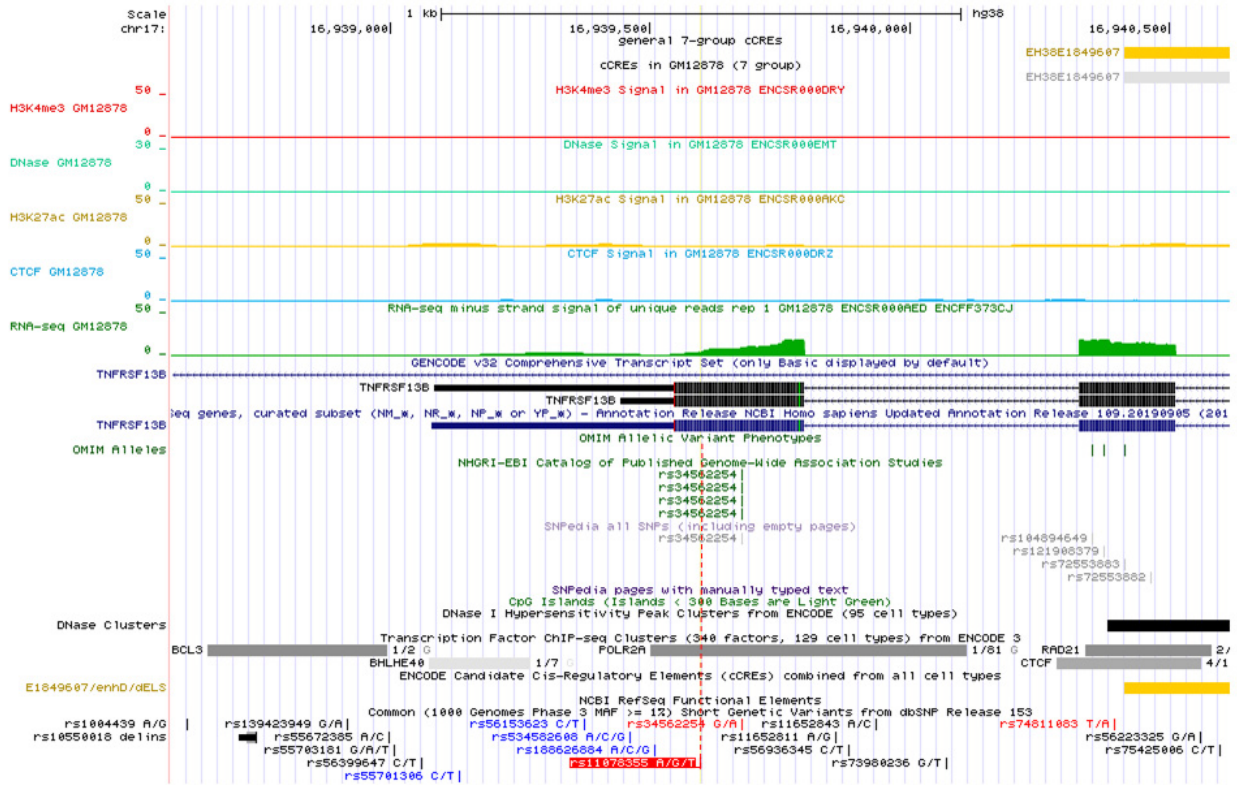
**A** Query SNP: **rs11078355** and variants with  $r^2 \geq 0.8$

chr	pos (hg38)	LD (r <sup>2</sup> )	LD (D')	variant	Ref Alt	AFR freq	AMR freq	ASN freq	EUR freq	SiPhy cons	Promoter histone marks	Enhancer histone marks	DNase	Proteins bound	Motifs changed	NHGRI/EBI GWAS hits	GRASP QTL hits	Selected eQTL hits	GENCODE genes	dbSNP func annot
17	16935416	0.97	0.99	rs55828532	A G	0.63	0.53	0.79	0.36			ADRL, MUS	4 tissues	FOSL2	8 altered motifs			4 hits	TNFRSF13B	
17	16939598	1	1	<b>rs11078355</b>	A G	0.60	0.55	0.79	0.37			5 tissues	BLD		5 altered motifs			3 hits	TNFRSF13B	synonymous
17	16942841	0.8	0.99	rs12938061	T C	0.56	0.56	0.79	0.41			4 tissues	10 tissues		9 altered motifs			4 hits	TNFRSF13B	intronic
17	16942862	0.8	0.99	rs12938073	T A	0.56	0.56	0.79	0.41			4 tissues	9 tissues		4 altered motifs			4 hits	TNFRSF13B	intronic
17	16942984	0.82	0.99	rs11078357	C G	0.67	0.56	0.79	0.41			5 tissues	BLD, MUS	POL2	Nix2			3 hits	TNFRSF13B	intronic
17	16943483	0.82	0.99	rs10852841	C G	0.59	0.55	0.79	0.41						5 altered motifs			3 hits	TNFRSF13B	intronic

**B**



C



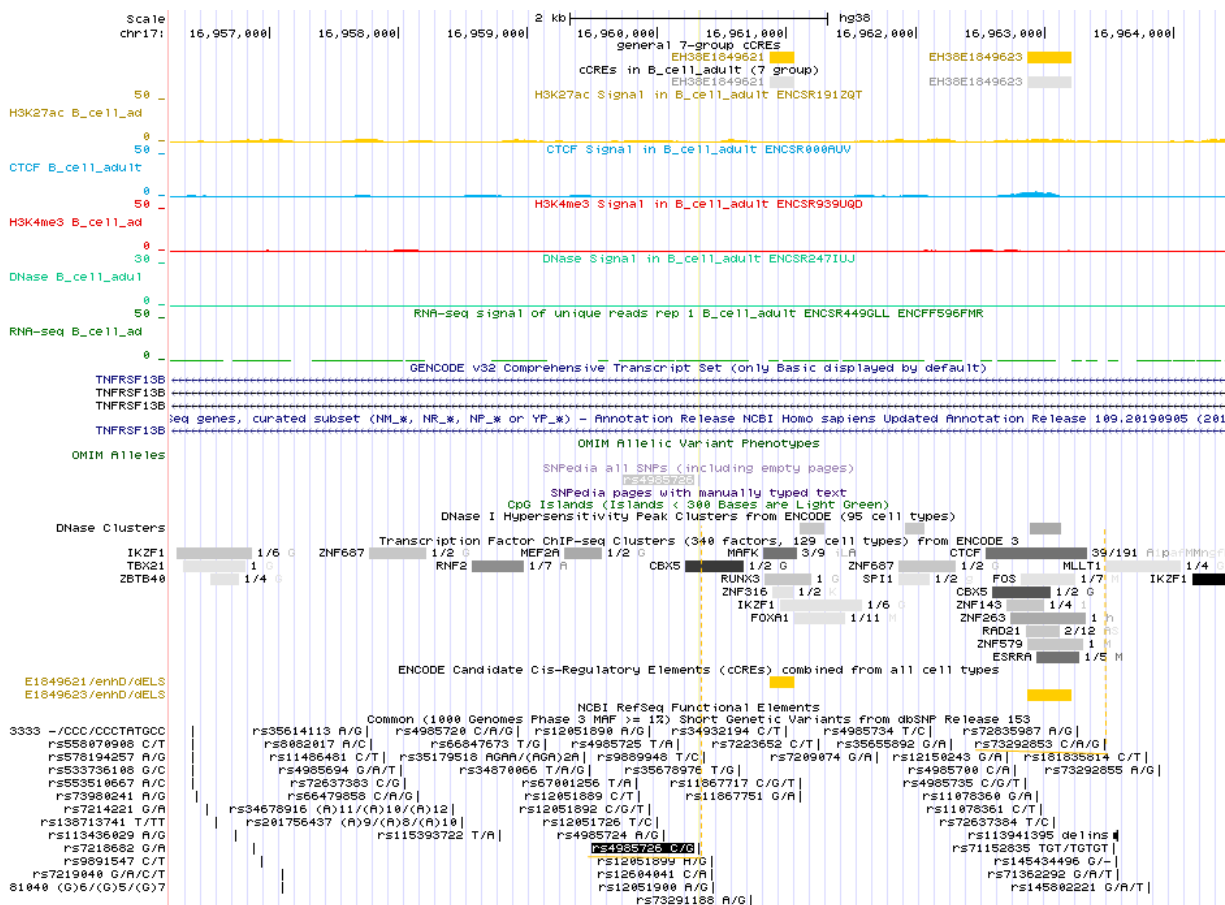
**Supplementary Figure S5.** *In silico* analysis - rs11078355 of *TNFRSF13B* (A) The result of analysis performed with application of HaploReg v4.1 tool (screen shot) [1-2]. Location of rs11078355 of *TNFRSF13B* gene (highlighted) in relation to data from ENCODE project for B cells (B) and GM12878 cell line (D) showing lack of cCRE in close distance to rs11078355. A view of the region surrounding the rs4968210 and rs12942590 includes also the representation of the ChIP-seq data from the ENCODE project, representing transcription factors (TFs) binding sites (black boxes denote the sites with stronger evidence for TFs binding in that region). Figures were downloaded from the UCSC genome browser (<http://genome.ucsc.edu>) [ 8-10].

A

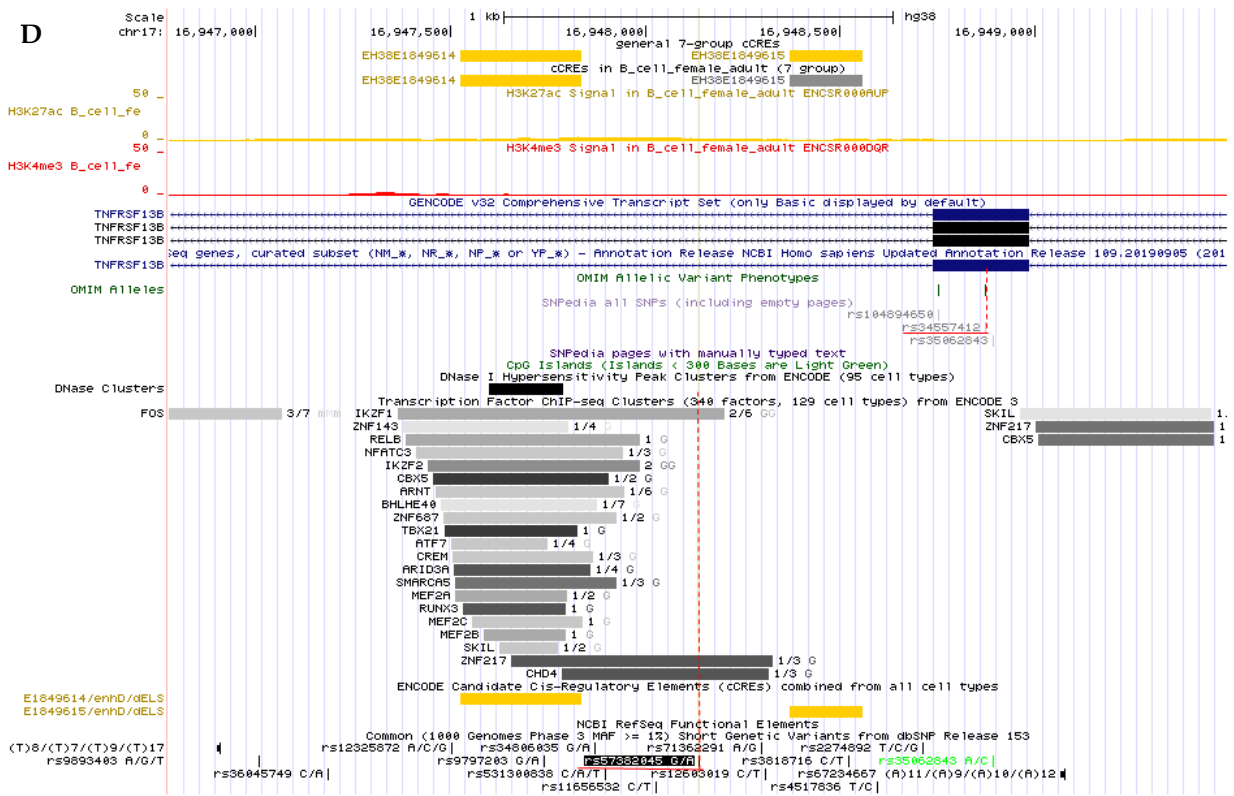
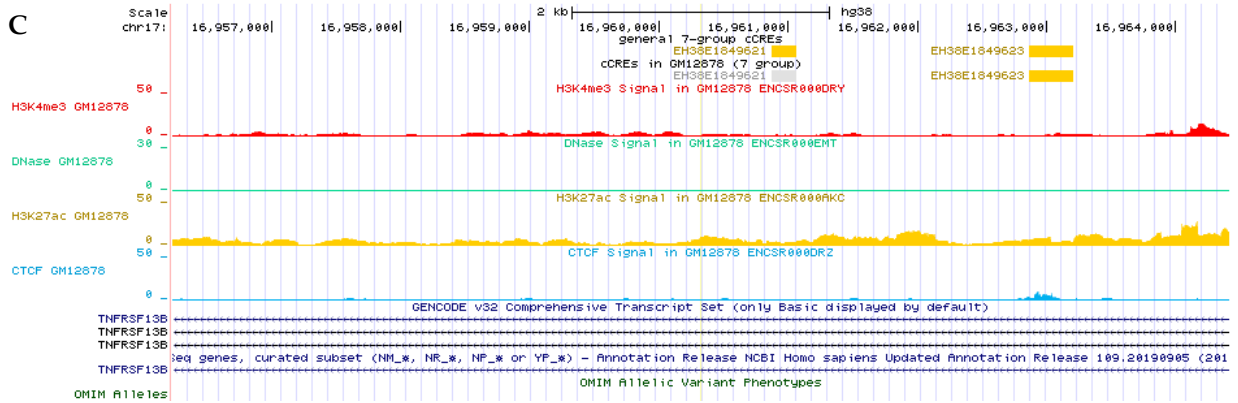
Query SNP: **rs4985726** and variants with  $r^2 \geq 0.8$

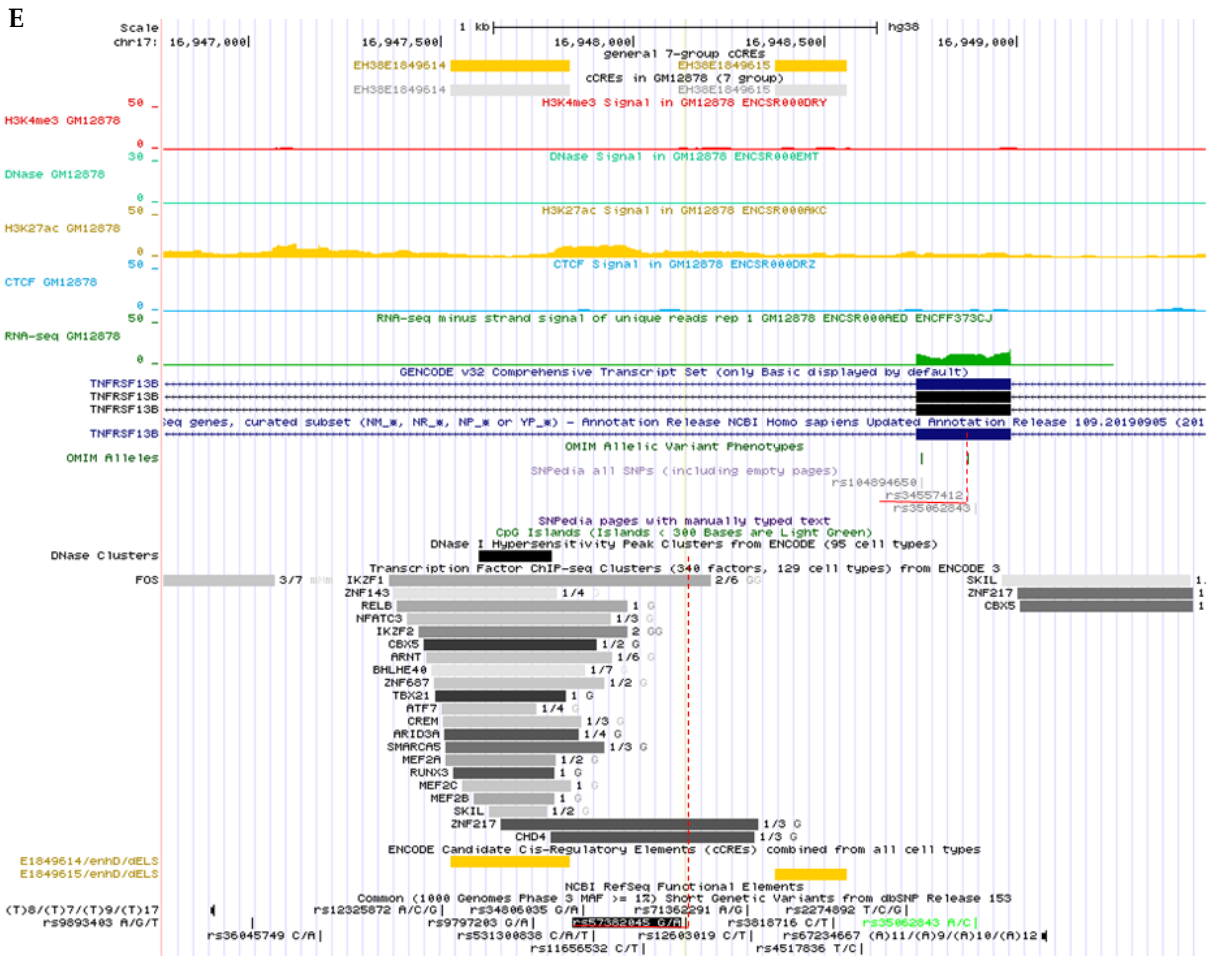
chr	pos (hg38)	LD (r <sup>2</sup> )	LD (D')	variant	Ref	Alt	AFR freq	AMR freq	ASN freq	EUR freq	SiPhy cons	Promoter histone marks	Enhancer histone marks	DNase	Proteins bound	Motifs changed	NHGRI/EBI GWAS hits	GRASP QTL hits	Selected eQTL hits	GENCODE genes	dbSNP func annot		
17	16938999	0.8	0.94	<b>rs55672385</b>	A	C	0.11	0.09	0.38	0.11		BLD	6 tissues	THYM, LNG		4 altered motifs				TNFRSF13B			
17	16939677	0.84	0.92	<b>rs34562254</b>	G	A	0.12	0.10	0.39	0.10			5 tissues		POL2	7 altered motifs				TNFRSF13B	missense		
17	16941853	0.89	0.97	<b>rs4792800</b>	A	G	0.12	0.10	0.39	0.11			4 tissues	11 tissues		12 altered motifs	1 hit			TNFRSF13B	intronic		
17	16945251	0.89	0.97	<b>rs4500785</b>	C	G	0.18	0.11	0.38	0.11			7 tissues	IPSC	POL2	AP-2, NRSF					TNFRSF13B	intronic	
17	16945436	0.93	0.97	<b>rs4561508</b>	C	T	0.13	0.10	0.38	0.11			7 tissues			GLI	4 hits				TNFRSF13B	intronic	
17	16945825	0.93	0.97	<b>rs4273077</b>	A	G	0.14	0.15	0.47	0.11			8 tissues	20 tissues	CTCF, RAD21	CTCF, Myf, Nkx2	2 hits				TNFRSF13B	intronic	
17	16948136	0.93	0.97	<b>rs57382045</b>	G	A	0.13	0.10	0.38	0.11		BLD, BRN	BLD		Brachyury, Msx-1						TNFRSF13B	intronic	
17	16951166	0.94	0.97	<b>rs74892229</b>	G	A	0.13	0.10	0.38	0.10			BLD	BLD		4 altered motifs					TNFRSF13B	intronic	
17	16953548	0.97	0.99	<b>rs57293260</b>	A	T	0.13	0.10	0.38	0.10		BLD, THYM	5 tissues	CTCF		Arid5a						TNFRSF13B	intronic
17	16953863	0.97	0.99	<b>rs57166795</b>	G	A	0.14	0.10	0.38	0.10		BLD, BRN, THYM				7 altered motifs						TNFRSF13B	intronic
17	16960324	1	1	<b>rs4985726</b>	C	G	0.08	0.09	0.38	0.10				BLD			2 hits	1 hit			TNFRSF13B	intronic	
17	16963463	0.94	0.97	<b>rs73292853</b>	C	G	0.32	0.17	0.47	0.10		BLD, PANC	IPSC			4 altered motifs						TNFRSF13B	intronic

B









**Supplementary Figure S6.** *In silico* analysis - rs4985726 of *TNFRSF13B* (A) The result of analysis performed with application of HaploReg v4.1 tool (screen shot) [1-2] (B, D) Location of rs4985726 (highlighted) and rs57382045 (highlighted) (LD with rs4985726) and of *TNFRSF13B* gene in relation to data from ENCODE project for B cells and (C, E) GM12878 cell line. The dotted lines indicated SNPs in LD with rs4985726. A view of the region surrounding the rs4968210 and rs12942590 includes also the representation of the ChIP-seq data from the ENCODE project, representing transcription factors (TFs) binding sites (black boxes denote the sites with stronger evidence for TFs binding in that region). Figures were downloaded from the UCSC genome browser (<http://genome.ucsc.edu>) [8-10].

**Supplementary Table S8.** Genotype distribution of the *BCMA* (*TNFRSF17*; 16p13.13) polymorphisms in patients and controls.

<i>BCMA</i> <i>TNFRSF17</i> polymorphisms		Patients (N=439)		Controls (N=477)		OR	CI95%	Patients vs. Controls
		N	%	N	%			
rs11570136 2KB Upstream Variant	TT	205	46.70	213	44.70	1*		
	TA	185	42.10	204	42.80	0.72	0.68 ;1.24	$\chi^2_{df=2} = 0.61$
	AA	49	11.20	60	12.60	0.85	0.56 ;1.30	p=0.735
<b>HWE</b>		p=0.587 f=0.035 CI95%=-0.06;0.13		p=0.3032 f=0.046 CI95%=-0.04;0.14				
rs2017662 Thr159Thr	GG	386	87.90	421	88.30	1*		$\chi^2_{df=2} = 2.18$
	GA	51	11.60	56	11.70	0.99	0.66; 1.49	p=0.336
	AA	2	0.50	0	0.00	5.45	0.26; 113.94	
<b>HWE</b>		p=0.8217 f=0.011 CI95%=0.07;0.12		p=0.1732 f=-0.062 CI95%=-0.08;-0.05				
rs373496 Ser81Asn	CC	424	96.60	453	95.00	1*		
	CT	14	3.20	24	5.00	0.63	0.33 ;1.23	$\chi^2_{df=2} = 3.14$
	TT	1	0.20	0	0.00	3.20	0.13 ;78.89	p=0.2210
<b>HWE</b>		p=0.027 f=-0.11 CI95%=-0.02;0.35		p=0.5730 f=-0.02 CI95%=-0.04;0.02				

rs2017662 GA+AA vs. GG (OR=1.03; CI95%=0.69-1.54; p-value=0.8765)

rs373496 CT+TT vs. CC (OR=0.68; CI95%=0.35-1.29; p-value=0.2269)

**Supplementary Table S9.** Basic statistics of the main variables considered in the paper.

<b>Rai</b>	<b>0</b>	<b>1</b>	<b>2</b>	<b>3</b>	<b>4</b>	<b>Sum</b>
<b>n</b>	73	67	74	12	20	246
<b>%</b>	29.7	27.2	30.1	4.9	8.1	100%
	<b>Min</b>	<b>Q1</b>	<b>Median</b>	<b>Sn</b>	<b>Q3</b>	<b>Max</b>
<b>IgA [g/l]</b>	0.13	0.8	1.375	0.82	2.192	127
<b>sAPRIL [ng/ml]</b>	0.02	16.8	28.1	15.8	42.75	162.4
<b>APRIL<sup>+</sup> CD19<sup>+</sup> [%]</b>	0.79	4.298	8.89	8	19.08	73.74
<b>APRIL<sup>+</sup> CD19<sup>+</sup> [MFI]</b>	30.12	53.47	82.82	38	121.2	226.6
<b>TACI<sup>+</sup> CD19<sup>+</sup> [%]</b>	0.18	3.043	10.32	10.1	33.09	92.31
<b>TACI<sup>+</sup> CD19<sup>+</sup> [MFI]</b>	18.88	28.45	38.35	16.1	58.05	152.8

Q1, Q3 – 1<sup>st</sup> and 3<sup>rd</sup> quartile

Sn – variability measure

MFI – Mean Fluorescence Intensity

**Supplementary Table S10.** Primer sequences, annealing temperatures and restriction enzymes used for PCR-RFLP genotyping.

SNP	Forward primer (5'-3')	Reverse primer (5'-3')	T <sub>a</sub>	Restriction enzyme
rs4985726	TTCTATTTCTTTTCTTGTCTAAGTGC	TGGTAGTTCACACTAAAGGAATG	52°C	<i>DpnII</i>
rs8072293 <sup>a</sup>	CATCAGGGACAAGAGGCC	CCTGACTGTGGGGCCAGA	55°C	<i>BslI</i>
rs11078355	AGCCCTGTGAGCACATCC	TCTTTCTCTCTCCCCTCCTCT	52°C	<i>AccI</i>

T<sub>a</sub> – annealing temperatures

<sup>a</sup> OneTaq Hot Start 2X Master Mix with GC Buffer (New England BioLabs) was used for PCR

**Supplementary Table S11.** TaqMan SNP Genotyping Assays used for genotyping with applying the allelic discrimination method.

<b>SNP</b>	<b>ID of TaqMan SNP Genotyping Assays</b>
rs11552708	C_25630192_20
rs3803800	C_12115389_10
rs4968210	C_26603822_10
rs6608	C_247220_20
rs12051889	C_31002822_20
rs11656106	C_12121510_10

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