

Review

Cancer Stem Cells in Tumor Microenvironment of Adenocarcinoma of the Stomach, Colon, and Rectum

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Simple Summary: Gastrointestinal cancers have a high mortality rate worldwide, and the progression of the disease is related to cancer stem cells. Until now, its relationship with the microenvironment has been poorly understood. We describe the molecules and different pathways activated during this interaction and the new targeting therapies for cancer cells and microenvironment modulation. This approach could impact the way gastrointestinal cancers are managed.



Citation: Islas, J.F.; Quiroz-Reyes, A.G.; Delgado-Gonzalez, P.; Franco-Villarreal, H.; Delgado-Gallegos, J.L.; Garza-Treviño, E.N.; Gonzalez-Villarreal, C.A. Cancer Stem Cells in Tumor Microenvironment of Adenocarcinoma of the Stomach, Colon, and Rectum. *Cancers* **2022**, *14*, 3948. <https://doi.org/10.3390/cancers14163948>

Academic Editor: Alexander Nikitin

Received: 30 June 2022

Accepted: 9 August 2022

Published: 16 August 2022

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Abstract: Gastrointestinal adenocarcinomas are one of the world's deadliest cancers. Cancer stem cells and the tissue microenvironment are highly regulated by cell and molecular mechanisms. Cancer stem cells are essential for maintenance and progression and are associated with resistance to conventional treatments. This article reviews the current knowledge of the role of the microenvironment during the primary establishment of gastrointestinal adenocarcinomas in the stomach, colon, and rectum and its relationship with cancer stem cells. We also describe novel developments in cancer therapeutics, such as targeted therapy, and discuss the advantages and disadvantages of different treatments for improving gastrointestinal cancer prognosis.

Keywords: cancer stem cells; gastrointestinal adenocarcinomas; colon cancer and gastric cancer

1. Introduction

Cancer is a multi-step process during which cells acquire an untamed ability to grow, proliferate, and (most times) differentiate, ultimately leading to either improper organ growth or the establishment of inadequate cells in locations where they contribute negatively to the body homeostasis, causing (amongst other things) high levels of inflammation [1–5]. Colorectal and gastric cancers are the most prevalent cancer in the digestive track. They represent the third and fifth leading causes of cancer-related deaths. With over 500,000–990,000 new cases worldwide, they typically have a five-year survival rate (mainly due to late diagnosis) [6–8].

Smoking, alcohol consumption, and obesity are some of the most likely risk factors associated with this disease. Those factors can alter the tumor microenvironment modulating the most relevant tumor-promoting functions: inflammation, angiogenesis, metabolism, and epithelial-mesenchymal transition (EMT) [7,9,10]. Cells presented in the tumor microenvironment secrete different growth and differentiation factors, leading to an imbalance between cancer cells and cancer stem cell (CSC) self-renewal and differentiation. In particular, CSCs are a small subpopulation inside the tumor heterogeneity with clonal and tumor initiation ability and also have been related to chemoresistance [11,12]. Together,

these components build a microenvironment that favors cancer cell proliferation and dissemination, providing them protection from apoptosis and host immune surveillance [13]. Thus, this study describes the interactions between CSCs and the tumor microenvironment (TME) which allow for cancer progression and cancer cell resistance. Gastric and colorectal cancers are related diseases that share risk factors, initial features, and pathophysiological mechanisms with multiple similarities that affect the TME and CSC (as shown in Figure 1).

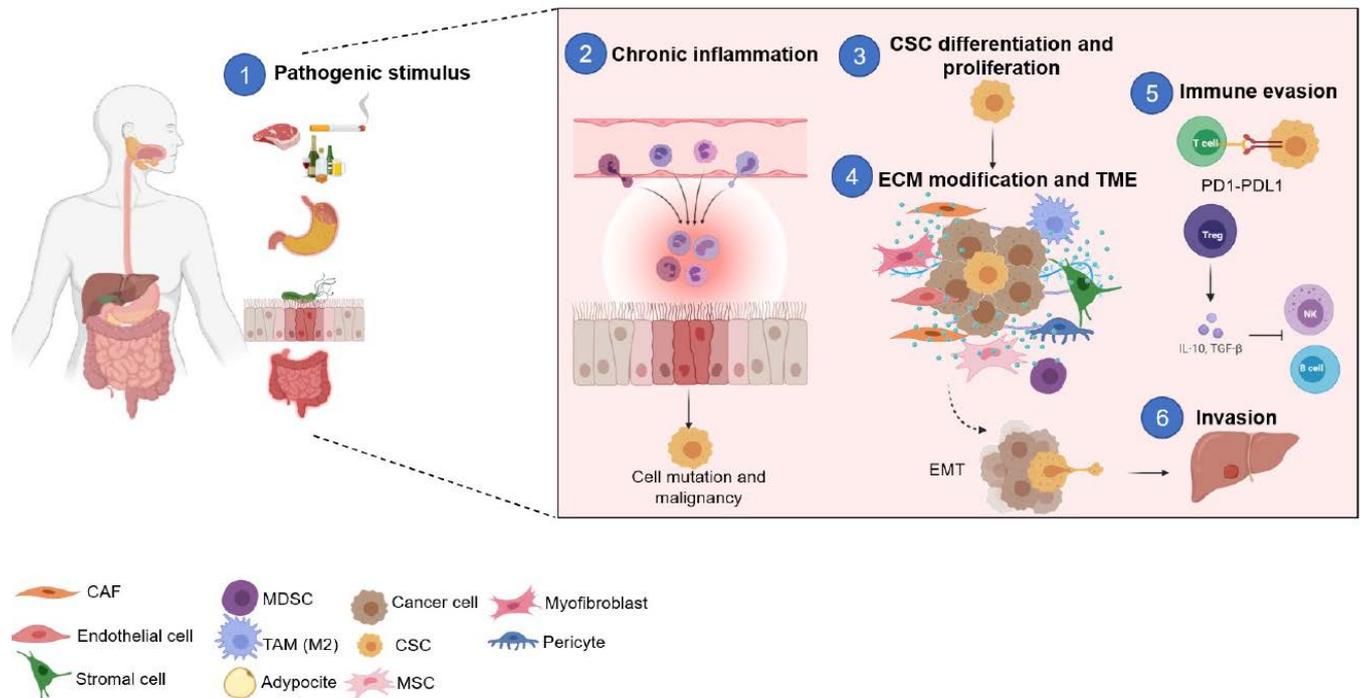


Figure 1. Impact of microenvironment during the development of gastric and colon cancer. Risk factors such as smoking, red meat, high alcohol intake, bacterial infections, and colitis could lead to chronic inflammation of the gastrointestinal tract, inducing cell malignization. All of these changes in the microenvironment induce cancer stem cell (CSC) differentiation and proliferation, with the contribution of cells that are attracted by inflammation and CSC. In addition, CSCs induce immune evasion and take advantage of immune regulatory mechanisms such as T regs, which allow epithelial-mesenchymal transition (EMT), invasion, and metastasis.

2. Generalities and Risk Factors by Gastrointestinal Cancers

Most colorectal tumors develop slowly through a series of morphological, histological, genetic, and epigenetic changes. They are generally asymptomatic until they reach a considerable size, thus increasing the risk for adenomas to develop into colorectal cancer (CRC) [14,15]. Several factors increase the risk of developing this disease. Among the most studied were a red and/or processed meat diet, high alcohol intake, tobacco use, and sedentarism [16]. Other risk factors associated with the development of CRC are non-modifiable, such as a family history of colorectal polyps, CRC, lynch syndrome, inflammatory bowel disease, type 2 diabetes, and racial and ethnic backgrounds [17]. CRC typically develops from focal changes within benign, precancerous polyps. These polyps are localized growths or aggregations of abnormal cells within the intestinal mucosa that protrude into the intestinal lumen [17].

Whether sporadic or hereditary, gene mutations also increase the risk for CRC. Some mutations in DNA mismatch repair (MMR) genes, such as MLH1, MSH2, PMS2, and adenomatous polyposis coli gene (APC), are uncommon and have a low prevalence in hereditary cancer [18]. The chromosomal instability pathway is observed in 65–70% of sporadic cancers. The first mutations develop within the APC gene cell division and the

next mutations develop in the KRAS oncogene, which affects cell differentiation, growth, motility, and survival. This causes a loss of function of the p53 gene, which acts as a transcription and apoptosis regulator that results in carcinogenesis [19]. Another epigenetic alteration seen in SSP-based CRC is aberrant gene promoter region hypermethylation, which inhibits gene transcription, regulating growth-promoting genes, bone morphogenic protein 3, and n-Myc downstream-regulated gene 4 (NDRG4) [17,19,20].

CRC can be classified among subtypes based on transcriptomic profiles. Moreover, every molecular subtype presents a different microenvironment [21,22]. These are summarized in Table 1.

Table 1. CRC subtypes.

| Subtypes | Frequency | Characteristics | Mutations | TME Associated | Clinical Relevance | Reference |
|--|-----------|--|---|---|---|-----------|
| CMS*1 Microsatellite instability immune | 14% | Diffuse immune infiltration ↓TGFβ inside TME | BRAF MSH6, RNF43, ATM, TGFB2, PTEN | ↓CAF ↑active adoptive immune response | Worse survival after relapse | |
| CMS2 Canonical | 37% | Epithelial features, CIN, activated WNT and MYC signaling pathways | APC, KRAS, TP53, PIK3CA | ↓CAF ↓immunogenic response | | |
| CMS3 Metabolic | 13% | Epithelial and dysregulation metabolic | APC, KRAS, TP53, PIK3CA | ↓CAF ↑active adoptive immune response, affected glutaminolysis, lipidogenesis, damage of mechanisms DNA repair | | [23–25] |
| CMS4 Mesenchymal | 23% | upregulation of EMT, TGF-β activation, angiogenesis, stromal infiltration activation mesenchymals and complement | APC, KRAS, TP53, PIK3CA | Many CAF, inflammation promote EMT | Worse relapse-free and overall survival | |
| Un-classified | 13% | Mixed phenotype of multiple CMSs or intratumoral heterogeneity | | | | |

* CMS, consensus molecular subtypes.

Common risk factors for gastric cancer (GC) include older age, male sex, tobacco smoking, radiation, and family history. Other factors are obesity, helicobacter pylori infection, gastroesophageal reflux disease, and diet (such as a high intake of salty foods or low vitamin A and C intake), nitrosamines, chemicals, smoked food, and high alcohol consumption [7]. In particular, high salt intake and tobacco are associated with increased incidence and mortality of GC [26]. The carcinomas are a consequence of a multistep process starting from chronic inflammatory through a sequence of atrophic gastritis, metaplasia, low-/high-grade dysplasia, and finally cancer. Gastritis is caused by chronic inflammation and is commonly regarded as a point of no return for carcinogenesis [27]. This results from persistent infection, which may evolve chronic atrophy gastritis, and subsequent changes in the gastric mucosa. *Helicobacter pylori* is the most prevalent bacterial pathogen currently classified as a group one carcinogen by the World Health Organization (definite carcinogen) [28].

Environmental and genetic factors can lead to cancer initiation by mutations that improve stemness and invasion. CSC are tumor cells that have the main properties of self-renewal, clonal tumor initiation capacity, and clonal long-term repopulation potential associated with the initiation, progression, and therapy resistance of cancer [29]. CSCs interaction with the microenvironment preserves their phenotypic plasticity, protects them from immune system elimination, and facilitates the growth and expansion of cancer cells [30].

3. Cancer Stem Cells and Tumoral Microenvironment

CSC exhibit several remarkable abilities mentioned elsewhere. However, since they can derive from normal stem cells, they might also arise through dedifferentiation of mature somatic cells to reacquire stem cell characteristics. Studies have shown that CSCs are also responsible for therapy resistance by developing an increased expression of drug transporters, maintaining a slow dividing state, and efficient DNA damage repair mechanisms [31]. The identification of CSCs is based on markers found in normal stem cells, and these vary according to the type of cancer. CD44, CD24, and CD133 are usually common in gastrointestinal cancers. NANOG, OCT4, SOX2, and KLF4 transcription factors are considered critical of stem self-renewal and pluripotency regulators, mediating tumor proliferation and differentiation [27]. Also, overexpression of components from signaling pathways such as JAK/STAT, WNT, NOTCH, SHH, PI3K/phosphatase and Tensin homolog, and NF- κ B signaling pathways are involved in renewal, differentiation, and uncontrolled proliferation of CSC [32].

Inside the tumor, the CSC niche can be a product of numerous factors specific to the TME and considered important factors extrinsically influencing tumor heterogeneity. These CSC can affect the tumor's aggressiveness or invasion, modulating the normal growth and development of resident stem cells [30,33]. In turn, the accumulation of key mutations appears to begin specifically within the CSC compartment. Related EMT markers TGF- β 1, TWIST, SNAIL, SLUG, vimentin, and CD44 were upregulated, and CDH1 mRNA levels were decreased in the gastric mucosa of patients with dysplasia or early GC in comparison with controls. A variant of CD44, a hyaluronic acid receptor (CD44v8-10), was identified as the predominant CD44 variant expressed in GC cells and contributes to tumor progression, possibly by enhancing oxidative stress defense [30].

Another proposed origin of CSCs is derived from the transformation of bone marrow-derived mesenchymal stem cells (BM-MSCs) in the GC, which served as primary cell components contributing to tumor progress, migration, and angiogenesis [34]. It has been observed that after infection and inflammatory stimulation of BM-MSCs, they migrate to the gastric epithelium and participate in tissue repair of the gastric mucosa [35].

A tumor's diverse cell population tends to be erratic rather than regulated. However, CSC within the tumor is exposed to several biomolecules lead to an imbalance between CSC self-renewal and differentiation [35]. In CRC, MSC-derived exosomes are a double-edged sword in cancer development, metastasis, and invasion [36,37]. Tumor-associated stromal cells are key contributors to the tumor microenvironment, these arise from distinct sources, consisting of the basement membrane, non-malignant cells (fibroblasts, BM-MSC, adipocytes), immune cells (macrophages, lymphocytes), extracellular matrix, and vasculature (endothelial cells, pericytes) [38]. Thus, the CSC microenvironment produces factors that stimulate CSC self-renewal, induce angiogenesis, and recruit immune cells and other stromal cells that secrete additional factors to promote tumor cell invasion, as shown in Table 2.

Table 2. Tumor microenvironment factors that improve CSC activity.

| Cancer | Cell | Factor | Activity in CSC | Reference |
|---------|--------------------|---|---|------------|
| Gastric | CAFs | IL-6, IL-8, IL-1, IL-22, TGF- β , PGE-2, FGF, TNF- α , and IL-1 β , CXCL12 | Promotes EMT and tumor invasion | [13,39] |
| | MSC | WNT5a, Gremlin-1, miR-214, miR-221, and miR-222 | Tumor growth and metastasis | [13,40,41] |
| | TAMs (M2) | EGF, HGF, PDGF, FGF, VEGF, MGF-E8, MCP-1, COX-2/PGE-2, IFN- γ and ROS | Improves cell growth, drug resistance, upregulation of CD44 | [13,42,43] |
| | Endothelial cells | CXCR4 | Tumor invasion | [13] |
| | Myofibroblasts | R-spondin3 | Proliferation of Axin2+ Lgr5– stem cells | [13] |
| | BMDCs | IL-6 and HGF | Increase proliferation and stemness | [43] |
| Colon | TAMs (M2) | IL-10, PD-1 | Immune evasion | [44] |
| | Tregs | IL-10, TGF- β , PD-L1, PD-L2 and CTLA-4 | Immune evasion | [44,45] |
| | CAFs | TGF- β 1, IL- β 1, IL-6, IL-33, ROS, C-X-C chemokine receptor (CXC), MMPs, lysyl oxidase, miR-21, TNF- α , and alpha-smooth muscle actin (aSMA), HGF | ECM remodeling, stemness phenotype | [41,46] |
| | Granulocytes MDSCs | ROS | Induce hypoxic phenotype | [44] |
| | Endothelial cells | SNAIL, Jagged-1, AKT | Proliferation, stemness, and induce drug resistance | [41,46] |
| | MSCs | VEGF | Angiogenesis and liver metastasis | [41] |
| | BMDCs CD34+ CD31– | MMP9, MMP2 | Tumor invasion | [41] |

4. Crosstalk between Oncogenic Signaling and Metabolic Pathways of CSC: Role of Stroma-Derived Chemokines in the Local Invasion of Primary Tumor

Inflammation and tissue damage attract cells that cooperate with tumor TME as immune cells, stromal cells, adipocytes, and ECM components, whose main role is inhibiting apoptosis, protection of tumoral cells, promotion of proliferation, immune evasion, and invasion [10,47]. ECM is formed by a variety of substances, including collagen, elastin, laminin, fibronectin, and modulators such as metalloproteinase (MMP), which cleaves the ECM components and is crucial for tissue remodeling [48]. The persistent stimulus of inflammation in the tissue affects the homeostasis of cells, matrix, and cytokines. This finally leads to fibrosis and remodeling of ECM, which contributes to the establishment of tumors [49].

One example of this process is when the GC microenvironment becomes infiltrated by CAFs and TAMs, leading to excessive fibrosis [50]. The stimulus broadens the range of proinflammatory cytokines such as IL-6, TGF- β , FGF (fibroblast growth factor), TNF- α , and IL-1 β that promote EMT [51]. Also, these cells induce fibrosis in ECM, which is associated with a worse prognosis in GC. In colitis-associated cancer, Liang et al. demonstrated that sphingosine-1-phosphate (S1P) induces an amplifying loop of SIPR1 and NF- κ B/IL-6/STAT3 [52]. Interleukins such as IL-6 and gp130-related are the main activators of the JAK2/STAT3 pathway in CSCs. In addition, IL-6 promotes the survival of pre-malignant intestinal epithelial cells, which then transforms into tumoral cells [53,54]. CAFs enhance

GC cell migration and EMT through the secretion of IL-6 [50]. Once activated, the STAT-3 signaling pathway induces Tlr2 gene transcription in the gastric epithelium, which after overexpression, promotes proliferation and inhibits gastric epithelial cell apoptosis [55]. Tumor cells take advantage of STAT-3 by increasing immune evasion by inhibiting the maturation of dendritic cells (DC), which activate the adaptive immune response [56]. In recent years, WNT5a signaling in CAF has been implicated in tumor progression. In turn, overexpression of β -catenin and WNT5a indicates a poor prognosis since they promote cell growth, migration, invasion, and EMT of digestive tract tumors [39]. In addition, CSCs promote tumor infiltration of TAM through the CD44 receptor, which is upregulated by miR-328 suppression. Moreover, MSCs present an immunomodulatory role on lymphocytes B, T, dendritic cells, macrophages, and MDSCs. Therefore, MSC could affect CD4+ T cell migration and differentiation, T helper 17 homeostasis modulation, and response [51]. In addition, STAT3 mediated TWIST expression and EMT can be activated by EGF/EGFR [52].

Chemokines act by interacting with specific G protein-coupled receptors, and chemokines from TME can facilitate tumor progression or remodeling of the tumor niche by signal transduction [57]. Thus, these proteins may play a crucial role in the pathogenesis of CRC, and GC, as shown in Table 3. However, the high heterogeneity of the cell context limits transcription factors potential to induce gene expression on CSCs and their changes to improve tumor progression [58]. It is being proposed that transcription factors such as the Yes-Associated Protein (YAP)/Transcriptional Co-activator with PDZ-binding Motif (TAZ) pathway lead to epithelial phenotype repression and participate together with factors of the SNAIL family, such as TWIST and Zeb, which are the main regulators of EMT [59]. YAP overexpression has been reported in GC, and CRC; protein levels are associated with poor prognosis, tumor stage, and metastasis [60–62].

Inflammation and tissue damage are a potent chemokine source; both processes recruit cells that cooperate with tumor TME as immune cells, stromal cells, adipocytes, and ECM components. The main role of these cells is to inhibit apoptosis, protect tumoral cells, promote proliferation, and immune evasion and invasion [10,47].

Gastrointestinal epithelial cell infection by *H. pylori* and the expression of CXCL1 gastric cancer cells is necessary to stimulate the migration of bone marrow-derived stromal cells (BMDC) by CXCR2 signaling; moreover, the expression of CD271 by BMDC is related with invasion and worse prognosis [63]. Tumor-associated macrophages (TAM) are associated with tumor stage and metastasis. There are two types of macrophage differentiation: M1 macrophages that produce type I proinflammatory cytokines such as IL-1 β , IL-1 α , IL-12, TNF- α , and GFAP, and M2 macrophages that produce type II anti-inflammatory cytokines such as IL-4, IL-6, and IL-10, related to the pro-tumorigenic activity. The change from one phenotype to another depends on the TME, and the high infiltration of M2 is related to a worse prognosis [64]. Chronic gastritis induced by *H. pylori* infection is associated with 90% of gastric cancer cases. It has been reported that this infection causes atrophy of acid-secreting parietal cells (PC), which increases CD44+ stem cell proliferation in the gastric isthmus by pERK [26,65]. Some gastrointestinal hormones, such as gastrin or neurotransmitters and acetylcholine (Ach), may also play unique roles in the antral stem cell niche. Gastrin is secreted from G cells and is responsible for HCl secretion in parietal cells (PCs). These cells reside near the antral isthmus region [66]. Gastrin also induces the expression of EGFR ligands as heparin-binding EGF and trefoil family factor 2, which activate the cholecystikinin receptor (CCK-BR), and thus, motility, secretion, migration, and proliferation of gastric cells [67]. Moreover, MMPs and tissue inhibitors of metalloproteinases (TIMPs) decrease E-Cadherin and ECM degradation cell change interactions and paracrine signals, helping malignant transformation [68]. For example, *H. pylori* increase IL-21 expression in infected gastric mucosa and promote gelatinases, MMP-2, MMP-7, and MMP-9 synthesis through NF- κ B in activated B cells [68]. Some other proteins increase their expression during GC, such as phosphoglycerate kinase 1 (PGK1), CXCR4, CXCL12, and β -catenin [46], which promote EMT. In addition, CAFs secrete multiple proinflammatory cytokines such as TGF- β 1, IL- β 1, IL-6, IL-33, ROS, C-X-C chemokine receptor (CXC),

MMPs, lysyl oxidase, miR-21, TNF- α , and α SMA [46]. All of these factors contribute to tissue fibrosis, and later fibrogenesis activates ECM remodeling. In addition, fibrogenesis activates the chemoresistance-inducing factor SNAIL in epithelial cells, promoting proliferation and inducing drug resistance [69]. Table 4 includes the MMPs present in GC and CRC.

Table 3. Chemokines in the tumor microenvironment.

| Chemokine | Receptor/Pathway | Gastrointestinal Cancer Improvement | Reference |
|---------------|--|---|-----------|
| IL-6 | IL-6R, activation of JAK2-STAT3 | Promotes proliferation and EMT | [53] |
| WNT5a | Frizzled receptor, activation of WNT/ β -catenin pathway | Promote cell growth, migration, invasion and EMT | [70] |
| PGK1 | Upregulates CXCR4 and β -catenin | Promotes EMT and metastasis | [46,71] |
| IL-21 | IL-21R, increase NF- κ B in activated B cells | promotes gelatinases, MMP-2, MMP-7, and MMP-9, and EMT | [68] |
| Gal-1 | Prch, activation of Hedgehog signaling | Promote tumor invasion and EMT | [72] |
| POSTN | ERK and p38 pathways | Proliferation, invasion, and migration of cancer cells | [73] |
| CXCL12 | CXCR4 | Improves TME and angiogenesis, lamellipodia and filopodia, cell adhesion to ECM | [74] |
| CXCL8 | CXCR1/CXCR2 regulates the expression of MMP-9, intracellular adhesion molecule (ICAM)-1, and E-cadherin. | Increased invasion, migration, and adhesion of cancer cells | [74] |
| CXCL1 | CXCR2, higher levels of MMP-2 and MMP-9 and upregulation of Ras and STAT3 | Tumor progression, increased migration, and invasive potential | [74] |
| CXCL5 | ERK/SNAIL pathway | Progression and metastasis of GC and activation of pro-tumor neutrophils | [75] |
| CXCL7 | CXCR2 | Promote tumor growth and activation of pro-tumor neutrophils | [76] |
| CXCL9, CXCL10 | CXCR3 | Promotes metastasis to lymph nodes | [77] |
| CXCL8 | CXCR2 | Increase proliferation and invasive capacity | [76] |
| CXCL11 | CXCR3 and CXCR7 | Promotes cell growth and EMT | [78] |
| CXCL16 | CXCR6 | Enhanced the recruitment of tumor-infiltrating lymphocytes | [74] |

Table 4. Matrix metalloproteinases in gastrointestinal cancers.

| Cancer | MMP | Role in Cancer | Reference |
|------------|-----------------------------|---|-----------|
| Gastric | 2, 1 and 9 | Promotes in tumor invasion, especially degradation of the basement membrane | [79] |
| | 13 and MT1-MMP and/or MMP-2 | Progression of GC | [80] |
| | 7 | Promotes metastasis | [80] |
| Colorectal | 1 | Correlates with tumor stage and poor prognosis, level of invasion, lymph node involvement, and metastasis | [79,81] |
| | 2 | Correlates with cancer invasion. | [80,81] |
| | 3 and 9 | Cancer progression and poor prognosis | [80] |
| | 9 | Contributes to inflammation and metastasis | [80,81] |
| | 7 | Relates to nodal or distant metastasis, and cell proliferation | [80,81] |
| | 12 | Expression reduces tumor growth and increases survival | [81] |
| | 13 | Related with advanced cancer stage and poor survival | [80,81] |

5. Participation of Extracellular Matrix Components in Cancer Progression

The role of the ECM has been demonstrated in all stages of GC and CRC, from onset to metastasis [82,83]. However, it has been shown that some components of the protein family are shared by gastrointestinal tumors and regulate a key aspect in the early stages of tumor biology, participating in the regulation of CSC (which can be favored under the stimulation of some signs or changes in the microenvironment by ECM proteins) [84]. Intestinal epithelial cells express integrins in healthy and pathological circumstances. Integrins are categorized based on the preferred ligands; they bind to collagens for $\alpha1/\alpha2$ coupled to $\beta1$, laminins for $\alpha3/\alpha6/\alpha7$ coupled to $\beta1$ or $\beta4$, tenascin for $\alpha9\beta1$, and RGD-containing ligands (fibronectin, osteopontin, and vitronectin) for $\alpha5/\alpha8$ coupled to $\beta1$ and αV coupled to $\beta1/\beta3/\beta5/\beta6/\beta8$. Recent works address the altered expression of certain integrins and their involvement in GC and CRC progression [85]. For example, integrin V6 can be expressed by CRC cells, which can then inactivate TGF β by integrin $\alpha v\beta6$ subsequently activating fibroblasts that promote tumor invasion [86]. CRC integrin $\alpha v\beta8$ expressed on tumor cells is reported as a crucial regulatory function during cell adhesion in the tumor microenvironment and supports the activation of TGF- $\beta1$ [87]. Several of these integrins, including integrin subunits $\beta1$, $\alpha6$, $\beta3$, and $\beta4$, concentrated in healthy adult stem and progenitor cells, are also signs of CSCs. Another characteristic of CRC is mainly overexpression of collagen types I, VI, VII, VIII, X, XI, and XVIII. For example, increased expression of type 1 collagen promotes EMT and stem cell marker expression by activation of PI3K/AKT/Snail signaling pathway conducted by integrin $\alpha2\beta1$ [88]. In addition, recent studies reported a higher expression of collagen XVII, which was significantly associated with the progression of cancer and by interaction with laminin-5 (Laminin-332), which is essential for epithelial cell migration and basement membrane attachment and, according to some studies, is a determinant for CCR initiation [89]. Figueiredo et al. reported that mutant E-cadherin conduces to an increase in $\beta1$ integrin expression associated with higher grade tumors and reduced overall survival of the GC patient [90]. Some of these proteins promote key CSC functions such as: (1) EMT; (2) immune surveillance modulation; (3) self-renewal/maintenance; and (4) metabolic reprogramming and matricellular proteins. These functions are also involved in the cellular mechanical response, such as mechano-sensor integrins, which are receptors for many proteins [91]. Table 5 summarizes the most important proteins associated with CSC and early stages and progression of colorectal and gastric cancer. In addition, these ECM proteins have been recently proposed as markers of early diagnosis and poor prognosis of gastrointestinal neoplasms associated with CSC signaling pathways. For example, Galectin-1 (Gal-1) is associated with *H. pylori* infection in GC, the increase in the expression of β -catenin, vimentin, and Hedgehog, and the decrease in E-cadherin expression [92,93]. In CRC, a higher expression in endothelial and tumor cells has been found in the stroma that promotes tumor invasion and progression. Although some reports have not found a coincidence with the tumor stage and progression in Gal-1 expression, more reports show that its expression is significant in the early and late stages of the disease. Therefore, it is an interesting target for gastrointestinal neoplasias [94]. The extracellular carbohydrate structure of Gal-1 produced by CAFs may interact with Integrin 1 to promote Gli1 expression, which may ultimately activate Wnt/-catenin signaling and result in the EMT process in GC cells [95,96]. To provide another example, a rigid, collagen-rich, or POSTN-rich ECM allows macrophage polarization to a pro-tumorigenic M2 phenotype. After recruitment, M2 macrophages activate several CSC survival signaling pathways, including Src, NF- κ B, STAT3/SOX2, and SHH. ECM can also affect the proliferation and activation of T cells, which are necessary to capture and kill CSCs. POSTN is overexpressed by CAFs and constitutes the primary tumor niche by supporting the proliferation of cancer cells through the ERK signaling pathway in GC [84].

Table 5. ECM deregulated components in colorectal and GC with associated stemness.

| ECM Component | Type of Cancer | Role in Cancer Stemness | Clinical Relevance | Ref |
|---|-----------------------|--|--|-----------|
| Tenascin | Gastric Colorectal | Upregulation of NOTCH ligand, Jagged 1 and other NOTHC components; enhance the expression of LGR5 and MSI1, the WNT and NOTCH signaling Stemness and tumorigenicity maintenance; increases expression of CD133 and Cmi1. Improve EMT and clonogenicity in CRC CSCs through $\alpha 2\beta 1$ integrin; | Increased expression in pre-malignant and malignant epithelia | [97,98] |
| Fibrous protein Collagen type I | Gastric Colorectal | enhance tumor potential and self-renewal of ALDH+ CSCs through $\beta 1$ integrin and FAK signaling | Overexpression correlated with overall survival | [99–101] |
| Fibronectin | Gastric Colorectal | FN is a marker of cancer stemness and induces EMT, promote resistance and poor prognosis | Activates intracellular signaling, mediated by integrins, TLRs, Wnt/ β catenin, and P13K, t | [102] |
| Secreted protein Acidic and Rich in Cysteine (SPARC) Gastric | Colorectal | Associated with EMT | Overexpression better prognosis Overexpression and chemosensitivity Survival prognosis and the clinical features of the tumor were significantly associated with survival, differentiation, and staging. | [103,104] |
| Periostin (POSTN) | Colorectal Gastric | POSTN promotes stemness and mesenchymal phenotype in human epithelial cells; plays an essential role in the crosstalk between CSCs and the niche leading to metastasis; recruits Wnt ligands, and increases signaling by promoting CSC maintenance and expansion | Correlation with tumor size, grade of cell differentiation, lymph node metastasis, serosal invasion, clinical stage, and 5-year survival rates. | [105–108] |
| Biglycan | Colorectal Gastric | Biglycan is highly expressed in colon CSCs and promotes chemoresistance of colon cancer cells by activating NF- κ B signaling | High levels of biglycan are associated with cancer aggressiveness, tumor stage, lymph node metastasis, and worse overall patient survival | [109,110] |
| Galectin | Colorectal Gastric | Regulated by HIF-1 and it plays vital pro-tumorigenic roles within the tumor microenvironment. | Pathogenesis of gastrointestinal malignancies, favoring tumor development, aggressiveness, metastasis, immunosuppression, and angiogenesis. | [111] |

Another important component in ECM are miRNAs. miR-206 downregulation in GC enhances GC stem cells (GCSCs) and carcinogenesis. At the same time, its overexpression suppresses GCSCs formation and associated tumorigenesis through ETS homologous factor (EHF) downregulation [112]. In GC, miR-17-92 members can help show metaplasia, but more importantly, they can be found at the early pre-niche stage serving a significant role as biomarkers [113]. In CRC, one of the most studied pathologies, circulating levels of miR-17 and miR-92a have been associated with early stages (primary tumor establishment)

formation [3,114]. Having members of this cluster circulating (if further analyzed and quantified) can give us an idea of cancer staging in the gastrointestinal tract. Moreover, miR-340 is a well-studied miRNA known for its role in the early establishment of CRC [115,116]. The primary miRNAs that participate in several pathways and lead to or inhibit gastric and colon cancers are shown in Figure 2.

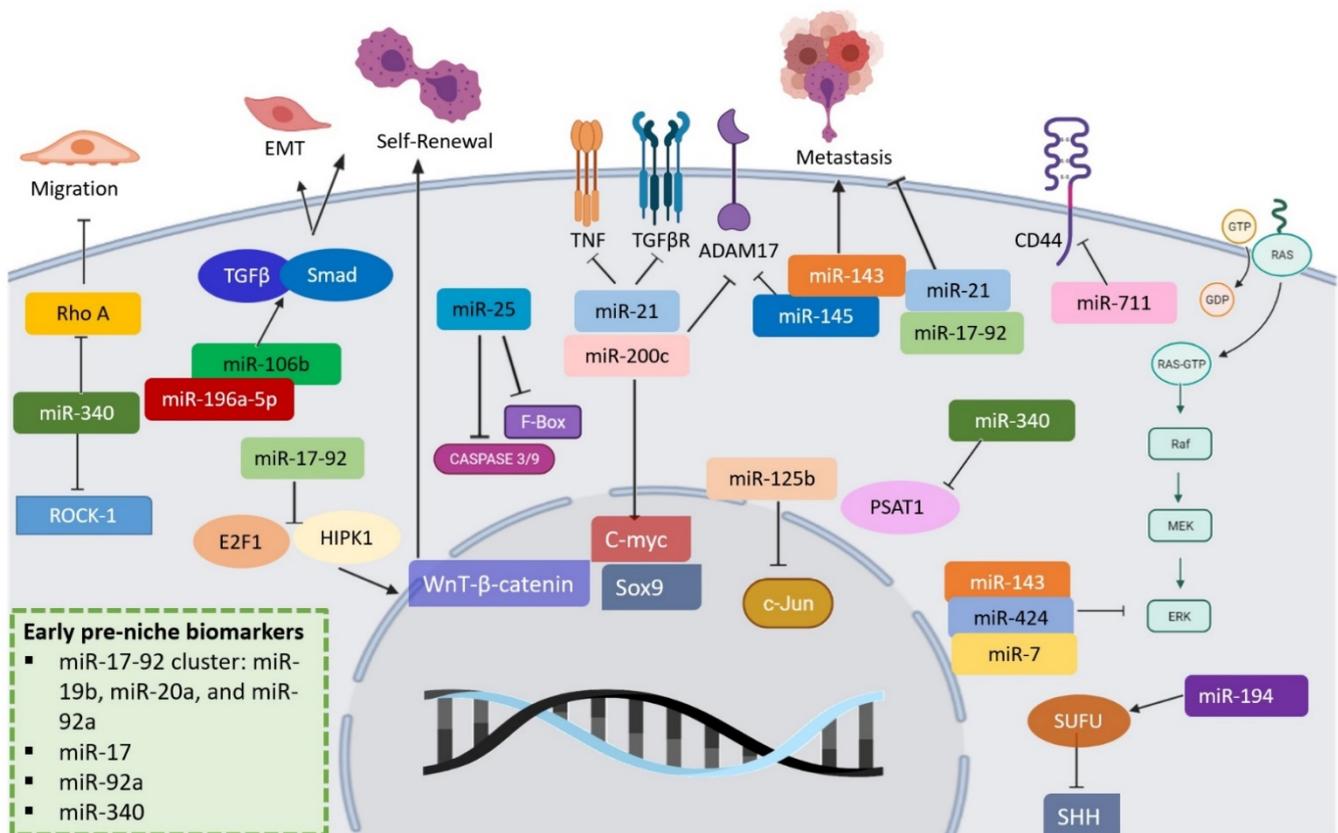


Figure 2. Regulatory roles of miRNAs in gastrointestinal cancers. miRNAs participate in the modification of several pathways that lead to or inhibit gastric and colon cancers. Some are present since cancer pre-niche and are related to EMT, migration, proliferation, invasion, and metastasis-development.

In a bit more detail, in CD44 (+) cells, miR-106b enhances gastric stem cell (GCSC) traits such as EMT, self-renewal, and invasion through the modulation of the TGF- β /Smad signaling pathway [112]. miR-145 can further affect the stemness property of CD44 (+) cells, reducing levels of ADAM17 and SOX9, which regulate invasiveness. Loss of this regulation leads to tumor initiation via IL-6 inflammatory processes [117]. In addition, miR-196a-5p has been upregulated in CD44 + cells, and suppression led to less colony formation and invasion of GC stem cells, suggesting a significant role in EMT and invasion of this cell population [83]. miR-143, miR-145, miR-21, and miR-194 significantly distorted the epithelium of several cancers; these miRNAs relate to the target cell cycle (cdc25), PTEN signaling, FSCN1, ZEB2, and MDM2 [118–122].

Over 10 years, cluster miR-143 and miR-145 were shown to coregulate FSCN1 (early marker), taking part in metastasis [123,124]. Over time, this has led to identifying novel lymphatic directed biomarkers such as miR-135b-5p, miR-15b-5p, and miR-195-5p, correlated to pathological classifications. miR-145 seems to target c-Myc, inducing p53. Another interesting miRNA is miR-25, as its effects in GCs include viability, cell migration, and growth through an increase in cyclin D1. In colon cancer, miR-25 further decreases apoptosis by inhibiting caspases 3/9 [125]. In GC, miR-25 directly correlates with invasion by repression of the F-box and WD-40 domain protein 7 [126]. Tumor suppressor FBXW7

regulates degradation of the growth proteins, cyclin E, c-Myc, Mcl-1, mTOR, Jun, and Notch [126].

6. Treatment against Gastrointestinal Cancers

In usually incurable advanced tumors with metastases, patients get conventional therapy options such as surgery, radiotherapy, and chemotherapy [127]. However, chemoresistance has been reported as one of the main causes of treatment ineffectiveness, where CSCs overexpress P-gp, ABCG2, Bcl-2, and surviving overexpression [128]. Studies have reported that cell resistant to cisplatin, 5-FU or docetaxel showed hyperexpression of the stem cell marker proteins, CD44, CD133, ALDH1A1, NOTCH1, Oct4, and SOX2 showing increased migration and invasion, formation of spheroids, colonies, and tumorigenic [129]. This information, coupled with an increase in morbidity and mortality in gastrointestinal system cancers, forces us to continue researching new strategies to provide effective treatment that avoids cancer cell resistance.

New cancer therapies are directed at stimulating the immune system to eliminate CSC and revert immunotolerance. These immunotherapies have been studied in CRC, particularly in MSI/MMR status. Some immune components in the TME regulate tumor development. Thus, immune cells are a promising target for cancer immunotherapy [45]. Checkpoint inhibitors, such as the CTLA-4 and PD-1 inhibitors tremelimumab, and nivolumab, have presented limited activity. In addition, the Check-Mate-142 trial (Phase II) evaluated nivolumab and nivolumab plus ipimumab in patients with metastatic CRC with MSI-H/dMMR or MSS/pMMR status, showing improvement in patient survival [130]. Moreover, the FDA-approved nivolumab (Opdivo) combined with chemotherapy for initial treatment of patients with advanced or metastatic GC due to improvement in survival compared with placebo [131].

In the KEYNOTE-164 phase II trial, pembrolizumab monotherapy reached a one-year overall survival of 72% in patients pretreated with MSI-H mCRC [132]. In addition, the combination of pembrolizumab with cisplatin and 5-FU in a phase II clinical trial in subjects with recurrent metastatic GC (KEYNOTE-059/NCT02335411) increased survival, encouraging FDA approval of pembrolizumab in September 2017 as third-line therapy for recurrent or metastatic cancer [131]. Adjuvant therapy with FOLFOX and PD-1/PD-L1 inhibitors achieved an objective response rate (ORR) of up to 50% and increased survival. In MSI-H/dMMR CRC, checkpoint inhibitors should be used after therapy with first-line drugs such as fluoropyrimidine, oxaliplatin, and irinotecan [130].

CAR-T cell therapies present efficacy against hematological malignancies and solid tumors, and recently CAR-T cells have been applied in gastrointestinal tumors against antigens such as epidermal growth factor receptor 2 (HER2), carcinoembryonic antigen (CEA), mucin 1 (MUC1) and epithelial cell adhesion molecule (EpCAM) to delay tumor growth in murine models [133]. MUC1-specific CAR-T cells effectively target MUC1-positive tumor cells. EpCAM-targeted CAR-T cells are being evaluated in clinical trials (phase I/II trials NCT02617134 and NCT02725125) to assess their safety and efficacy [131]. G17DT is a vaccine that neutralizes gastrin-17, a hormone required for the growth of GC cells. The phase II/III clinical trial (NCT00042510) showed that G17DT induced specific affinity against gastrin antibody, which inhibits tumor proliferation and metastasis [134,135].

Table 6 includes the new target molecules and pathways against CSC in gastrointestinal cancer.

Table 6. Target molecules and pathways for gastrointestinal cancer stem cells.

| Therapeutic Agent | Inhibitory Mechanism | Mode of Action | Ref |
|--|---|---|-----------|
| Gemcitabine | EMT | Reduce the frequency of CTC | [136,137] |
| Apatinib napabucasin (BBI-608), pacritinib | EMT/Angiogenesis | Targeting Jak2/STAT3 block PI3K/AKT and VEGFR2/RAF/MEK/ERK signaling pathways | [138–140] |
| Artesunate | Cell oncosis | β -catenin | [141] |
| DKN-01 | Wnt/ β catenin signaling | DKK1 | [141,142] |
| Berberine | EMT | Smad-independent and growth factor- β signaling pathway | [143,144] |
| Metformin | EMT | Tyrosine kinase and topoisomerase II. SFRP2 silencer inhibitor | [141,145] |
| Genistein | CSCs characteristics by Gli1 signaling pathway. | PI3K and mTOR | [141,146] |
| DS-7423 | Apoptosis by p53 induction | IL-6/STAT3 signal pathway | [147] |
| Wogonin | EMT | N- and E-cadherin, STAT3 pathway, and cofilin pathway | [148] |
| Bigelovin | EMT | | [148] |
| Cordycepin | EMT. Upregulating cancer cell apoptosis and eliciting cell cycle arrest | Upregulation of CLEC2 via the PI3K/Akt signaling pathway | [149] |
| Dichloroacetate | Increased responsiveness to 5-FU | PDK-1 | [150] |
| CART-133 | Tumor cell apoptosis | CSC CD133+ | [151] |
| Sulfasalazine | CD44v-positive cancer cells | xCT | [152] |
| LGK974, Foxy-5, PRI-724 | Wnt/ β -catenin signaling | PORCN inhibitor, WNT5A mimic, β -catenin/CREBBP inhibitor | [153] |
| Ginsenoside Rg3 combined with cisplatin | TME | EMT | [154] |

7. Conclusions

Cancer is the final step from several cell changes due to genetic mutations and environmental epigenetic changes. As has been observed, gastrointestinal cancers share important characteristics from their beginning, mainly due to the development of a pre-niche that allows CSCs proliferation and establishment in tissue. Additionally, inflammation seems to be a central pillar in gastrointestinal cancer development, as cancer risk factors induce a state of chronic cytokine activation. Moreover, cancer cells use similar signaling pathways to survive. Signals from the pre-niche, such as ECM proteins, chemokines, and miRNAs derived from recruited cells, also enrich the tumor microenvironment and modulate the CSC response. Thus, the cooperation of all of these factors to develop tumorigenesis is necessary. It is not surprising that immunotherapy has been proposed as a strategy for managing gastrointestinal cancers, wherein a change in TME signals is introduced to eliminate CSC.

Author Contributions: J.F.I. wrote, analyzed, and corrected the manuscript. A.G.Q.-R. and P.D.-G. performed literature analysis and wrote, discussed the revised manuscript, and designed the images. H.F.-V. and J.L.D.-G. designed and revised information contained on tables and performed literature analysis and wrote treatment section. E.N.G.-T. and C.A.G.-V. made the literature analysis and wrote, discussed, and revised the manuscript of this review. E.N.G.-T. and C.A.G.-V. contributed equally to this work. All authors have read and agreed to the published version of the manuscript.

Funding: This research received no external funding.

Acknowledgments: Special thanks to Sergio Lozano for reviewing this manuscript.

Conflicts of Interest: All authors indicated no potential conflicts of interest in publishing this manuscript.

References

1. Hwang, W.L.; Lan, H.Y.; Cheng, W.C.; Huang, S.C.; Yang, M.H. Tumor stem-like cell-derived exosomal RNAs prime neutrophils for facilitating tumorigenesis of colon cancer. *J. Hematol. Oncol.* **2019**, *12*, 10. [[CrossRef](#)] [[PubMed](#)]
2. Intlekofer, A.M.; Finley, L.W.S. Metabolic signatures of cancer cells and stem cells. *Nat. Metab.* **2019**, *2*, 177–188. [[CrossRef](#)] [[PubMed](#)]
3. Gonzalez-Villarreal, C.A.; Quiroz-Reyes, A.G.; Islas, J.F.; Garza-Treviño, E.N. Colorectal Cancer Stem Cells in the Progression to Liver Metastasis. *Front. Oncol.* **2020**, *10*, 1511. [[CrossRef](#)] [[PubMed](#)]
4. Pandurangan, A.K.; Divya, T.; Kumar, K.; Dineshbabu, V.; Velavan, B.; Sudhandiran, G.; Pandurangan, A.; Divya, T.; Kumar, K.; Dineshbabu, V.; et al. Colorectal carcinogenesis: Insights into the cell death and signal transduction pathways: A review. *World J. Gastrointest. Oncol.* **2018**, *10*, 244–259. [[CrossRef](#)] [[PubMed](#)]
5. Pretzsch, E.; Bösch, F.; Neumann, J.; Ganschow, P.; Bazhin, A.; Guba, M.; Werner, J.; Angele, M. Mechanisms of Metastasis in Colorectal Cancer and Metastatic Organotropism: Hematogenous versus Peritoneal Spread. *J. Oncol.* **2019**, *2019*, 7407190. [[CrossRef](#)]
6. Alsop, B.R.; Sharma, P. Esophageal Cancer. *Gastroenterol. Clin. North Am.* **2016**, *45*, 399–412. [[CrossRef](#)]
7. Karimi, P.; Islami, F.; Anandasabapathy, S.; Freedman, N.D.; Kamangar, F. Gastric Cancer: Descriptive Epidemiology, Risk Factors, Screening, and Prevention. *Cancer Epidemiol. Biomark. Prev.* **2014**, *23*, 700–713. [[CrossRef](#)]
8. Weitz, J.; Koch, M.; Debus, J.; Höhler, T.; Galle, P.R.; Büchler, M.W. Colorectal cancer. *Lancet* **2005**, *365*, 153–165. [[CrossRef](#)]
9. Labianca, R.; Beretta, G.D.; Kildani, B.; Milesi, L.; Merlin, F.; Mosconi, S.; Pessi, M.A.; Prochilo, T.; Quadri, A.; Gatta, G.; et al. Colon cancer. *Crit. Rev. Oncol. Hematol.* **2010**, *74*, 106–133. [[CrossRef](#)]
10. Kwilas, A.J. The Tumor Microenvironment in Esophageal Cancer. *Oncogene* **2016**, *35*, 5337–5349. [[CrossRef](#)]
11. Zhao, Y.; Bao, Q.; Renner, A.; Camaj, P.; Eichhorn, M.; Ischenko, I.; Angele, M.; Kleespies, A.; Jauch, K.W.; Bruns, C. Cancer stem cells and angiogenesis. *Int. J. Dev. Biol.* **2011**, *55*, 477–482. [[CrossRef](#)] [[PubMed](#)]
12. Plaks, V.; Kong, N.; Werb, Z. The Cancer Stem Cell Niche: How Essential is the Niche in Regulating Stemness of Tumor Cells? *Cell Stem Cell* **2015**, *16*, 225–238. [[CrossRef](#)] [[PubMed](#)]
13. Oya, Y.; Hayakawa, Y.; Koike, K. Tumor microenvironment in gastric cancers. *Cancer Sci.* **2020**, *111*, 2696–2707. [[CrossRef](#)]
14. Lochhead, P.; Chan, A.T.; Giovannucci, E.; Fuchs, C.S.; Wu, K.; Nishihara, R.; O'Brien, M.; Ogino, S. Progress and Opportunities in Molecular Pathological Epidemiology of Colorectal Premalignant Lesions. *Am. J. Gastroenterol.* **2014**, *109*, 1205. [[CrossRef](#)] [[PubMed](#)]
15. Conteduca, V.; Sansonno, D.; Russi, S.; Dammacco, F. Precancerous colorectal lesions (Review). *Int. J. Oncol.* **2013**, *43*, 973–984. [[CrossRef](#)] [[PubMed](#)]
16. Rawla, P.; Sunkara, T.; Barsouk, A. Epidemiology of colorectal cancer: Incidence, mortality, survival, and risk factors. *Prz. Gastroenterol.* **2019**, *14*, 89–103. [[CrossRef](#)]
17. Simon, K. Colorectal cancer development and advances in screening. *Clin. Interv. Aging* **2016**, *11*, 967–976. [[CrossRef](#)]
18. De'angelis, G.L.; Bottarelli, L.; Azzoni, C.; De'angelis, N.; Leandro, G.; Di Mario, F.; Gaiani, F.; Negri, F. Microsatellite instability in colorectal cancer. *Gastroenterology* **2010**, *138*, 2073–2087. [[CrossRef](#)]
19. Pino, M.S.; Chung, D.C. The Chromosomal Instability Pathway in Colon Cancer. *Gastroenterology* **2010**, *138*, 2059–2072. [[CrossRef](#)]
20. O'Brien, M.J.; Yang, S.; Mack, C.; Xu, H.; Huang, C.S.; Mulcahy, E.; Amorosino, M.; Farraye, F.A. Comparison of microsatellite instability, CpG island methylation phenotype, BRAF and KRAS status in serrated polyps and traditional adenomas indicates separate pathways to distinct colorectal carcinoma end points. *Am. J. Surg. Pathol.* **2006**, *30*, 1491–1501. [[CrossRef](#)]
21. Wang, M.; Busuttill, R.A.; Pattison, S.; Neeson, P.J.; Boussioutas, A. Immunological battlefield in gastric cancer and role of immunotherapies. *World J. Gastroenterol.* **2016**, *22*, 6373–6384. [[CrossRef](#)] [[PubMed](#)]
22. Balkwill, F.; Mantovani, A. Inflammation and cancer: Back to Virchow? *Lancet* **2001**, *357*, 539–545. [[CrossRef](#)]
23. Guinney, J.; Dienstmann, R.; Wang, X.; de Reyniès, A.; Schlicker, A.; Soneson, C.; Marisa, L.; Roepman, P.; Fessler, E.; Sousa, F.; et al. The Consensus Molecular Subtypes of Colorectal Cancer. *Nat. Med.* **2015**, *21*, 1350–1356. [[CrossRef](#)] [[PubMed](#)]
24. Thanki, K.; Nicholls, M.E.; Gajjar, A.; Senagore, A.J.; Qiu, S.; Szabo, C.; Hellmich, M.R.; Chao, C. Consensus Molecular Subtypes of Colorectal Cancer and their Clinical Implications. *Int. Biol. Biomed* **2017**, *3*, 105–111.
25. Wang, W.; Kandimalla, R.; Huang, H.; Zhu, L.; Li, Y.; Gao, F.; Wang, X.; Kong, H.; Sammons, C.A.; Street, W.; et al. Molecular subtyping of colorectal cancer: Recent progress, new challenges and emerging opportunities. *Semin. Cancer Biol.* **2019**, *55*, 37–52. [[CrossRef](#)]
26. Guggenheim, D.E.; Shah, M.A. Gastric cancer epidemiology and risk factors. *J. Surg. Oncol.* **2013**, *107*, 230–236. [[CrossRef](#)]
27. Xiao, S.; Zhou, L. Gastric Stem Cells: Physiological and Pathological Perspectives. *Front. Cell Dev. Biol.* **2020**, *8*, 571536. [[CrossRef](#)]
28. Ahn, H.J.; Lee, D.S. *Helicobacter pylori* in gastric carcinogenesis. *World J. Gastrointest. Oncol.* **2015**, *7*, 455–465. [[CrossRef](#)]
29. Codony-Servat, J.; Rosell, R. Cancer stem cells and immunoresistance: Clinical implications and solutions. *Transl. Lung Cancer Res.* **2015**, *4*, 689–703. [[CrossRef](#)]
30. Prager, B.C.; Xie, Q.; Bao, S.; Rich, J.N. Cancer Stem Cells: The Architects of the Tumor Ecosystem. *Cell Stem Cell* **2019**, *24*, 41–53. [[CrossRef](#)]
31. Dzobo, K.; Senthebane, D.A.; Ganz, C.; Thomford, N.E.; Wonkam, A.; Dandara, C. Advances in Therapeutic Targeting of Cancer Stem Cells within the Tumor Microenvironment: An Updated Review. *Cells* **2020**, *9*, 1896. [[CrossRef](#)] [[PubMed](#)]
32. Matsui, W.H. Cancer stem cell signaling pathways. *Medicine* **2016**, *95*, S8–S19. [[CrossRef](#)] [[PubMed](#)]

33. Zhao, Y.; Dong, Q.; Li, J.; Zhang, K.; Qin, J.; Zhao, J.; Sun, Q.; Wang, Z.; Wartmann, T.; Jauch, K.W.; et al. Targeting cancer stem cells and their niche: Perspectives for future therapeutic targets and strategies. *Semin. Cancer Biol.* **2018**, *53*, 139–155. [[CrossRef](#)] [[PubMed](#)]
34. Becerril-Rico, J.; Alvarado-Ortiz, E.; Toledo-Guzmán, M.E.; Pelayo, R.; Ortiz-Sánchez, E. The cross talk between gastric cancer stem cells and the immune microenvironment: A tumor-promoting factor. *Stem Cell Res. Ther.* **2021**, *12*, 498. [[CrossRef](#)]
35. Fu, Y.; Li, H.; Hao, X. The self-renewal signaling pathways utilized by gastric cancer stem cells. *Tumor Biol.* **2017**, *39*, 1010428317697577. [[CrossRef](#)]
36. Zhou, J.; Tan, X.; Tan, Y.; Li, Q.; Ma, J.; Wang, G. Mesenchymal stem cell derived exosomes in cancer progression, metastasis and drug delivery: A comprehensive review. *J. Cancer* **2018**, *9*, 3129–3137. [[CrossRef](#)]
37. Muralikumar, M. Current understanding of the mesenchymal stem cell-derived exosomes in cancer and aging. *Biotechnol. Rep.* **2021**, *31*, e00658. [[CrossRef](#)]
38. Bussard, K.M.; Mutkus, L.; Stumpf, K.; Gomez-Manzano, C.; Marini, F.C. Tumor-associated stromal cells as key contributors to the tumor microenvironment. *Breast Cancer Res.* **2016**, *18*, 84. [[CrossRef](#)]
39. Qin, Y.; Wang, F.; Ni, H.; Liu, Y.; Yin, Y.; Zhou, X.; Gao, G.; Li, Q.; Qi, X.; Li, J. Cancer-associated fibroblasts in gastric cancer affect malignant progression via the CXCL12-CXCR4 axis. *J. Cancer* **2021**, *12*, 3011–3023. [[CrossRef](#)]
40. Haddad, R.; Saldanha-Araujo, F. Mechanisms of T-cell immunosuppression by mesenchymal stromal cells: What do we know so far? *Biomed Res. Int.* **2014**, *2014*. [[CrossRef](#)]
41. Batista, S.; Gregório, A.C.; Otake, A.H.; Couto, N.; Costa-Silva, B. The Gastrointestinal Tumor Microenvironment: An Updated Biological and Clinical Perspective. *J. Oncol.* **2019**, *2019*. [[CrossRef](#)] [[PubMed](#)]
42. Fu, L.; Bu, L.; Yasuda, T.; Koiwa, M.; Akiyama, T.; Uchihara, T.; Baba, H.; Ishimoto, T. Gastric cancer stem cells: Current insights into the immune microenvironment and therapeutic targets. *Biomedicines* **2020**, *8*, 7. [[CrossRef](#)] [[PubMed](#)]
43. Sawayama, H.; Ishimoto, T.; Baba, H. Microenvironment in the pathogenesis of gastric cancer metastasis. *J. Cancer Metastasis Treat.* **2018**, *4*, 10. [[CrossRef](#)]
44. Therapy, C. The Tumor Microenvironment in Colorectal Cancer Therapy. *Cancers* **2019**, *11*, 1172.
45. Koi, M.; Carethers, J.M. The colorectal cancer immune microenvironment and approach to immunotherapies. *Futur. Oncol.* **2017**, *13*, 1633–1647. [[CrossRef](#)]
46. Brücher, B.L.D.M.; Jamall, I.S. Precancerous niche (PCN), a product of fibrosis with remodeling by incessant chronic inflammation. *4 Open* **2019**, *2*, 11.
47. Han, P.; Cao, P.; Hu, S.; Kong, K.; Deng, Y.; Zhao, B.; Li, F. Esophageal microenvironment: From precursor microenvironment to premetastatic niche. *Cancer Manag. Res.* **2020**, *12*, 5857–5879. [[CrossRef](#)]
48. Stamenkovic, I. Extracellular matrix remodelling: The role of matrix metalloproteinases. *J. Pathol.* **2003**, *200*, 448–464. [[CrossRef](#)]
49. Cox, T.R.; Erler, J.T. Remodeling and homeostasis of the extracellular matrix: Implications for fibrotic diseases and cancer. *DMM Dis. Model. Mech.* **2011**, *4*, 165–178. [[CrossRef](#)]
50. Ham, I.H.; Lee, D.; Hur, H. Role of cancer-associated fibroblast in gastric cancer progression and resistance to treatments. *J. Oncol.* **2019**, *2019*. [[CrossRef](#)]
51. Dominguez, C.; David, J.M.; Palena, C. Epithelial-mesenchymal transition and inflammation at the site of the primary tumor. *Semin. Cancer Biol.* **2017**, *47*, 177–184. [[CrossRef](#)] [[PubMed](#)]
52. Li, B.; Huang, C. Regulation of EMT by STAT3 in gastrointestinal cancer (Review). *Int. J. Oncol.* **2017**, *50*, 753–767. [[CrossRef](#)]
53. Grivennikov, S.; Karin, E.; Terzic, J.; Mucida, D.; Yu, G.Y.; Vallabhapurapu, S.; Scheller, J.; Rose-John, S.; Cheroutre, H.; Eckmann, L.; et al. IL-6 and STAT3 are Required for Survival of Intestinal Epithelial Cells and Development of Colitis-Associated Cancer. *Cancer Cell* **2009**, *15*, 103–113. [[CrossRef](#)]
54. Jin, W. Role of JAK/STAT3 Signaling in the Regulation of Metastasis, the Transition of Cancer Stem Cells, and Chemoresistance of Cancer by Epithelial-Mesenchymal Transition. *Cells* **2020**, *9*, 217. [[CrossRef](#)] [[PubMed](#)]
55. Tye, H.; Kennedy, C.L.; Najdovska, M.; McLeod, L.; McCormack, W.; Hughes, N.; Dev, A.; Sievert, W.; Ooi, C.H.; Ishikawa, T.O.; et al. STAT3-Driven Upregulation of TLR2 Promotes Gastric Tumorigenesis Independent of Tumor Inflammation. *Cancer Cell* **2012**, *22*, 466–478. [[CrossRef](#)] [[PubMed](#)]
56. Su, Y.L.; Banerjee, S.; White, S.V.; Kortylewski, M. STAT3 in tumor-associated myeloid cells: Multitasking to disrupt immunity. *Int. J. Mol. Sci.* **2018**, *19*, 1803. [[CrossRef](#)]
57. Pawluczuk, E.; Łukaszewicz-Zajac, M.; Mroczko, B. The role of chemokines in the development of gastric cancer—diagnostic and therapeutic implications. *Int. J. Mol. Sci.* **2020**, *21*, 8456. [[CrossRef](#)]
58. Bocci, F.; Gearhart-Serna, L.; Boareto, M.; Ribeiro, M.; Ben-Jacob, E.; Devi, G.R.; Levine, H.; Onuchic, J.N.; Jolly, M.K. Toward understanding cancer stem cell heterogeneity in the tumor microenvironment. *Proc. Natl. Acad. Sci. USA* **2019**, *116*, 148–157. [[CrossRef](#)]
59. Kim, H.; Lee, S.; Shin, E.; Seong, K.M.; Jin, Y.W.; Youn, H.S.; Youn, B.H. The Emerging Roles of Exosomes as EMT Regulators in Cancer. *Cells* **2020**, *9*, 861. [[CrossRef](#)]
60. Yin, F.; Dong, J.; Kang, L.; Liu, X. Hippo-YAP signaling in digestive system tumors. *Am. J. Cancer Res.* **2021**, *11*, 2495–2507.
61. Qu, Y.; Zhang, L.; Wang, J.; Chen, P.; Jia, Y.; Wang, C.; Yang, W.; Wen, Z.; Song, Q.; Tan, B.; et al. Yes—Associated protein (YAP) predicts poor prognosis and regulates progression of esophageal squamous cell cancer through epithelial—Mesenchymal transition. *Exp. Ther. Med.* **2019**, *18*, 2993–3001. [[CrossRef](#)] [[PubMed](#)]

62. Jiang, L.; Zhang, J.; Xu, Q.; Wang, B.; Yao, Y.; Sun, L.; Wang, X.; Zhou, D.; Gao, L.; Song, S.; et al. YAP promotes the proliferation and migration of colorectal cancer cells through the Glut3/AMPK signaling pathway. *Oncol. Lett.* **2021**, *21*, 312. [[CrossRef](#)] [[PubMed](#)]
63. Kasashima, H.; Yashiro, M.; Nakamae, H.; Masuda, G.; Kinoshita, H.; Morisaki, T.; Fukuoka, T.; Hasegawa, T.; Sakurai, K.; Toyokawa, T.; et al. Bone marrow-derived stromal cells are associated with gastric cancer progression. *Br. J. Cancer* **2015**, *113*, 443–452. [[CrossRef](#)] [[PubMed](#)]
64. Wang, X.L.; Jiang, J.T.; Wu, C.P. Prognostic significance of tumor-associated macrophage infiltration in gastric cancer: A meta-analysis. *Genet. Mol. Res.* **2016**, *15*, gmr15049040. [[CrossRef](#)]
65. Khurana, S.S.; Riehl, T.E.; Moore, B.D.; Fassan, M.; Ruggie, M.; Romero-Gallo, J.; Noto, J.; Peek, R.M.; Stenson, W.F.; Mills, J.C. The hyaluronic acid receptor CD44 coordinates normal and metaplastic gastric epithelial progenitor cell proliferation. *J. Biol. Chem.* **2013**, *288*, 16085–16097. [[CrossRef](#)]
66. Craven, C.J. A hypothesis of couplet molecules and couplet cells in gastric function and an association with *Helicobacter pylori*. *BMC Gastroenterol.* **2016**, *16*, 16. [[CrossRef](#)]
67. Smith, J.P.; Nadella, S.; Osborne, N. Gastrin and Gastric Cancer. *Cell. Mol. Gastroenterol. Hepatol.* **2017**, *4*, 75–83. [[CrossRef](#)]
68. Brücher, B.L.D.M.; Jamall, I.S. Transition from normal to cancerous cell by precancerous niche (PCN) induced chronic cell-matrix stress. *4Open* **2019**, *2*, 14. [[CrossRef](#)]
69. Wang, Y.; Shi, J.; Chai, K.; Ying, X.; Zhou, B. The Role of Snail in EMT and Tumorigenesis. *Curr. Cancer Drug Targets* **2013**, *13*, 963–972. [[CrossRef](#)]
70. Astudillo, P. Wnt5a Signaling in Gastric Cancer. *Front. Cell Dev. Biol.* **2020**, *8*, 110. [[CrossRef](#)]
71. Zieker, D.; Königsrainer, I.; Tritschler, I.; Löffler, M.; Beckert, S.; Traub, F.; Nieselt, K.; Bühler, S.; Weller, M.; Gaedcke, J.; et al. Phosphoglycerate kinase 1 a promoting enzyme for peritoneal dissemination in gastric cancer. *Int. J. Cancer* **2010**, *126*, 1513–1520. [[CrossRef](#)] [[PubMed](#)]
72. Chong, Y.; Tang, D.; Gao, J.; Jiang, X.; Xu, C.; Xiong, Q.; Huang, Y.; Wang, J.; Zhou, H.; Shi, Y.; et al. Galectin-1 induces invasion and the epithelial-mesenchymal transition in human gastric cancer cells via non-canonical activation of the hedgehog signaling pathway. *Oncotarget* **2016**, *7*, 83611–83626. [[CrossRef](#)] [[PubMed](#)]
73. Zhong, H.; Li, X.; Zhang, J.; Wu, X. Overexpression of periostin is positively associated with gastric cancer metastasis through promoting tumor metastasis and invasion. *J. Cell. Biochem.* **2019**, *120*, 9927–9935. [[CrossRef](#)] [[PubMed](#)]
74. Lee, H.J.; Song, I.C.; Yun, H.J.; Jo, D.Y.; Kim, S. CXC chemokines and chemokine receptors in gastric cancer: From basic findings towards therapeutic targeting. *World J. Gastroenterol.* **2014**, *20*, 1681–1693. [[CrossRef](#)]
75. Mao, Z.; Zhang, J.; Shi, Y.; Li, W.; Shi, H.; Ji, R.; Mao, F.; Qian, H.; Xu, W.; Zhang, X. CXCL5 promotes gastric cancer metastasis by inducing epithelial-mesenchymal transition and activating neutrophils. *Oncogenesis* **2020**, *9*, 63. [[CrossRef](#)]
76. Desurmont, T.; Skrypek, N.; Duhamel, A.; Jonckheere, N.; Millet, G.; Leteurtre, E.; Gosset, P.; Duchene, B.; Ramdane, N.; Hebbat, M.; et al. Overexpression of chemokine receptor CXCR2 and ligand CXCL7 in liver metastases from colon cancer is correlated to shorter disease-free and overall survival. *Cancer Sci.* **2015**, *106*, 262–269. [[CrossRef](#)]
77. Pączek, S.; Łukaszewicz-Zajac, M.; Mroczko, B. Chemokines—What Is Their Role in Colorectal Cancer? *Cancer Control* **2020**, *27*, 1073274820903384. [[CrossRef](#)]
78. Gao, Y.J.; Liu, D.L.; Li, S.; Yuan, G.F.; Li, L.; Zhu, H.Y.; Cao, G.Y. Down-regulation of CXCL11 inhibits colorectal cancer cell growth and epithelial-mesenchymal transition. *Onco. Targets. Ther.* **2018**, *11*, 7333–7343. [[CrossRef](#)]
79. Shimada, W. Matrix metalloproteinases and their inhibitors in arthropathy. *J. Jpn. Orthop. Assoc.* **1995**, *69*, 791–797.
80. Verma, S. Matrix metalloproteinases and gastrointestinal cancers: Impacts of dietary antioxidants. *World J. Biol. Chem.* **2014**, *5*, 355. [[CrossRef](#)]
81. Said, A.H.; Raufman, J.P.; Xie, G. The role of matrix metalloproteinases in colorectal cancer. *Cancers* **2014**, *6*, 366–375. [[CrossRef](#)] [[PubMed](#)]
82. Fernandes, E.; Sores, J.; Cotton, S.; Peixoto, A.; Ferreira, D.; Freitas, R.; Reis, C.A.; Santos, L.L.; Ferreira, J.A. Esophageal, gastric and colorectal cancers: Looking beyond classical serological biomarkers towards glycoproteomics-assisted precision oncology. *Theranostics* **2020**, *10*, 4903–4928. [[CrossRef](#)]
83. Andreuzzi, E.; Capuano, A.; Poletto, E.; Pivetta, E.; Fejza, A.; Favero, A.; Doliana, R.; Cannizzaro, R.; Spessotto, P.; Mongiat, M. Role of extracellular matrix in gastrointestinal cancer-associated angiogenesis. *Int. J. Mol. Sci.* **2020**, *21*, 3686. [[CrossRef](#)] [[PubMed](#)]
84. Nallanthighal, S.; Heiserman, J.P.; Cheon, D.-J. The Role of the Extracellular Matrix in Cancer Stemness. *Front. Cell Dev. Biol.* **2019**, *7*, 86. [[CrossRef](#)]
85. Hou, S.; Wang, J.; Li, W.; Hao, X.; Hang, Q. Roles of Integrins in Gastrointestinal Cancer Metastasis. *Front. Mol. Biosci.* **2021**, *8*, 708779. [[CrossRef](#)] [[PubMed](#)]
86. Zhang, Z.; Vuori, K.; Reed, J.C.; Ruoslahti, E. The $\alpha 5 \beta 1$ integrin supports survival of cells on fibronectin and up-regulates Bcl-2 expression. *Proc. Natl. Acad. Sci. USA* **1995**, *92*, 6161–6165. [[CrossRef](#)]
87. Zhou, M.; Niu, J.; Wang, J.; Gao, H.; Shahbaz, M.; Niu, Z.; Li, Z.; Zou, X.; Liang, B. Integrin $\alpha v \beta 8$ serves as a Novel Marker of Poor Prognosis in Colon Carcinoma and Regulates Cell Invasiveness through the Activation of TGF- $\beta 1$. *J. Cancer* **2020**, *11*, 3803–3814. [[CrossRef](#)]
88. Wu, X.; Cai, J.; Zuo, Z.; Li, J. Collagen facilitates the colorectal cancer stemness and metastasis through an integrin/PI3K/AKT/Snail signaling pathway. *Biomed. Pharmacother.* **2019**, *114*, 108708. [[CrossRef](#)]

89. Zeisberg, M.; Neilson, E.G. Biomarkers for epithelial-mesenchymal transitions. *J. Clin. Investig.* **2009**, *119*, 1429–1437. [[CrossRef](#)]
90. Figueiredo, J.; Ferreira, R.M.; Xu, H.; Gonçalves, M.; Barros—Carvalho, A.; Cravo, J.; Maia, A.F.; Carneiro, P.; Figueiredo, C.; Smith, M.L.; et al. Integrin β 1 orchestrates the abnormal cell—Matrix attachment and invasive behaviour of E-cadherin dysfunctional cells. *Gastric Cancer* **2022**, *25*, 124–137. [[CrossRef](#)]
91. Scott, L.E.; Weinberg, S.H.; Lemmon, C.A. Mechanochemical Signaling of the Extracellular Matrix in Epithelial-Mesenchymal Transition. *Front. Cell Dev. Biol.* **2019**, *7*, 135. [[CrossRef](#)] [[PubMed](#)]
92. Rossi, A.F.T.; Duarte, M.C.; Poltronieri, A.B.; Valsechi, M.C.; Jorge, Y.C.; De-Santi Neto, D.; Rahal, P.; Oliani, S.M.; Silva, A.E. Deregulation of annexin-A1 and galectin-1 expression in precancerous gastric lesions: Intestinal metaplasia and gastric ulcer. *Mediators Inflamm.* **2014**, *2014*, 478138. [[CrossRef](#)] [[PubMed](#)]
93. Yu, H.; Zeng, J.; Liang, X.; Wang, W.; Zhou, Y.; Sun, Y.; Liu, S.; Li, W.; Chen, C.; Jia, J. *Helicobacter pylori* promotes epithelial-mesenchymal transition in gastric cancer by Downregulating Programmed Cell Death Protein 4 (PDCD4). *PLoS ONE* **2014**, *9*, e105306. [[CrossRef](#)] [[PubMed](#)]
94. Bacigalupo, M.L.; Carabias, P.; Troncoso, M.F. Contribution of galectin-1, a glycan-binding protein, to gastrointestinal tumor progression. *World J. Gastroenterol.* **2017**, *23*, 5266–5281. [[CrossRef](#)]
95. Seropian, I.M.; González, G.E.; Maller, S.M.; Berrocal, D.H.; Abbate, A.; Rabinovich, G.A. Galectin-1 as an Emerging Mediator of Cardiovascular Inflammation: Mechanisms and Therapeutic Opportunities. *Mediators Inflamm.* **2018**, *2018*, 8696543. [[CrossRef](#)]
96. Zhang, X.; Zhang, C.; Ren, Z.; Zhang, F.; Xu, J.; Zhang, X.; Zheng, H. Curcumin affects gastric cancer cell migration, invasion and cytoskeletal remodeling through gli- β -catenin. *Cancer Manag. Res.* **2020**, *12*, 3795–3806. [[CrossRef](#)]
97. Tiitta, O.; Virtanen, I.; Sipponen, P.; Gould, V. Tenascin expression in inflammatory, dysplastic and neoplastic lesions of the human stomach. *Virchows Arch.* **1994**, *425*, 369–374. [[CrossRef](#)]
98. Murakami, T.; Kikuchi, H.; Ishimatsu, H.; Iino, I.; Hirotsu, A.; Matsumoto, T.; Ozaki, Y.; Kawabata, T.; Hiramatsu, Y.; Ohta, M.; et al. Tenascin C in colorectal cancer stroma is a predictive marker for liver metastasis and is a potent target of miR-198 as identified by microRNA analysis. *Br. J. Cancer* **2017**, *117*, 1360–1370. [[CrossRef](#)]
99. Zhang, Q.N.; Zhu, H.L.; Xia, M.T.; Liao, J.; Huang, X.T.; Xiao, J.W.; Yuan, C. A panel of collagen genes are associated with prognosis of patients with gastric cancer and regulated by microRNA-29c-3p: An integrated bioinformatics analysis and experimental validation. *Cancer Manag. Res.* **2019**, *11*, 4757–4772. [[CrossRef](#)]
100. Kirkland, S.C. Type I collagen inhibits differentiation and promotes a stem cell-like phenotype in human colorectal carcinoma cells. *Br. J. Cancer* **2009**, *101*, 320–326. [[CrossRef](#)]
101. Fang, S.; Dai, Y.; Mei, Y.; Yang, M.; Hu, L.; Yang, H.; Guan, X.; Li, J. Clinical significance and biological role of cancer-derived Type I collagen in lung and esophageal cancers. *Thorac. Cancer* **2019**, *10*, 277–288. [[CrossRef](#)] [[PubMed](#)]
102. Takigawa, H.; Kitadai, Y.; Shinagawa, K.; Yuge, R.; Higashi, Y.; Tanaka, S.; Yasui, W.; Chayama, K. Mesenchymal Stem Cells Induce Epithelial to Mesenchymal Transition in Colon Cancer Cells through Direct Cell-to-Cell Contact. *Neoplasia* **2017**, *19*, 429–438. [[CrossRef](#)] [[PubMed](#)]
103. Nakajima, M.; Yoshino, S.; Kanekiyo, S.; Maeda, N.; Sakamoto, K.; Tsunedomi, R.; Suzuki, N.; Takeda, S.; Yamamoto, S.; Hazama, S.; et al. High secreted protein acidic and rich in cysteine expression in peritumoral fibroblasts predicts better prognosis in patients with resectable gastric cancer. *Oncol. Lett.* **2018**, *15*, 803–812. [[CrossRef](#)] [[PubMed](#)]
104. Wu, J.; Zhang, J.R.; Jiang, X.Q.; Cao, X.G. Correlation between secreted protein acidic and rich in cysteine protein expression and the prognosis of postoperative patients exhibiting esophageal squamous cell carcinoma. *Mol. Med. Rep.* **2017**, *16*, 3401–3406. [[CrossRef](#)]
105. González-González, L.; Alonso, J. Periostin: A matricellular protein with multiple functions in cancer development and progression. *Front. Oncol.* **2018**, *8*, 225. [[CrossRef](#)]
106. Li, Z.; Zhang, X.; Yang, Y.; Yang, S.; Dong, Z.; Du, L.; Wang, L.; Wang, C. Periostin expression and its prognostic value for colorectal cancer. *Int. J. Mol. Sci.* **2015**, *16*, 12108–12118. [[CrossRef](#)]
107. Kikuchi, Y.; Kunita, A.; Iwata, C.; Komura, D.; Nishiyama, T.; Shimazu, K.; Takeshita, K.; Shibahara, J.; Kii, I.; Morishita, Y.; et al. The niche component periostin is produced by cancer-associated fibroblasts, supporting growth of gastric cancer through ERK activation. *Am. J. Pathol.* **2014**, *184*, 859–870. [[CrossRef](#)]
108. Lv, Y.J.; Wang, W.; Ji, C.S.; Jia, W.; Xie, M.R.; Hu, B. Association between periostin and epithelial-mesenchymal transition in esophageal squamous cell carcinoma and its clinical significance. *Oncol. Lett.* **2017**, *14*, 376–382. [[CrossRef](#)]
109. Pinto, F.; Santos-Ferreira, L.; Pinto, M.T.; Gomes, C.; Reis, C.A. The extracellular small leucine-rich proteoglycan biglycan is a key player in gastric cancer aggressiveness. *Cancers* **2021**, *13*, 1330. [[CrossRef](#)]
110. Zhu, Y.H.; Yang, F.; Zhang, S.S.; Zeng, T.T.; Xie, X.; Guan, X.Y. High expression of biglycan is associated with poor prognosis in patients with esophageal squamous cell carcinoma. *Int. J. Clin. Exp. Pathol.* **2013**, *6*, 2497–2505.
111. Ito, K.; Stannard, K.; Gabutero, E.; Clark, A.M.; Neo, S.Y.; Onturk, S.; Blanchard, H.; Ralph, S.J. Galectin-1 as a potent target for cancer therapy: Role in the tumor microenvironment. *Cancer Metastasis Rev.* **2012**, *31*, 763–778. [[CrossRef](#)]
112. Khan, A.Q.; Ahmed, E.I.; Elareer, N.R.; Junejo, K.; Steinhoff, M.; Uddin, S. Role of miRNA-Regulated Cancer Stem Cells in the Pathogenesis of Human Malignancies. *Cells* **2019**, *8*, 840. [[CrossRef](#)]
113. Li, H.; Wu, Q.; Li, T.; Liu, C.; Xue, L.; Ding, J.; Shi, Y.; Fan, D. The miR-17-92 cluster as a potential biomarker for the early diagnosis of gastric cancer: Evidence and literature review. *Oncotarget* **2017**, *8*, 45060–45071. [[CrossRef](#)]

114. Fu, F.; Jiang, W.; Zhou, L. Circulating Exosomal miR-17-5p and miR-92a-3p Predict Pathologic Stage and Grade of Colorectal Cancer. *Transl. Oncol.* **2018**, *11*, 221–232. [[CrossRef](#)]
115. Algaber, A.; Al-Haidari, A.; Madhi, R.; Rahman, M.; Syk, I.; Thorlacius, H. MicroRNA-340-5p inhibits colon cancer cell migration via targeting of RhoA. *Sci. Rep.* **2020**, *10*, 16934. [[CrossRef](#)]
116. Sun, Y.A.N.; Zhao, X.; Zhou, Y.; Hu, Y.U. miR-124, miR-137 and miR-340 regulate colorectal cancer growth via inhibition of the Warburg effect. *Oncol. Rep.* **2012**, *28*, 1346–1352. [[CrossRef](#)] [[PubMed](#)]
117. Yu, C.-C.; Tsai, L.-L.; Wang, M.-L.; Yu, C.-H.; Lo, W.-L.; Chang, Y.-C.; Chiou, G.-Y.; Chou, M.-Y.; Chiou, S.-H. miR145 Targets the SOX9/ADAM17 Axis to Inhibit Tumor-Initiating Cells and IL-6–Mediated Paracrine Effects in Head and Neck Cancer. *Cancer Res.* **2013**, *73*, 3425–3440. [[CrossRef](#)]
118. Wu, X.; Ajani, J.A.; Gu, J.; Chang, D.W.; Tan, W.; Hildebrandt, M.A.T.; Huang, M.; Wang, K.Y.; Hawk, E. MicroRNA expression signatures during malignant progression from Barrett’s esophagus to esophageal adenocarcinoma. *Cancer Prev. Res.* **2013**, *6*, 196–205. [[CrossRef](#)] [[PubMed](#)]
119. Pack, S.D.; Karkera, J.D.; Zhuang, Z.; Pak, E.D.; Balan, K.V.; Hwu, P.; Park, W.S.; Pham, T.; Ault, D.O.; Glaser, M.; et al. Molecular cytogenetic fingerprinting of esophageal squamous cell carcinoma by comparative genomic hybridization reveals a consistent pattern of chromosomal alterations. *Genes Chromosom. Cancer* **1999**, *25*, 160–168. [[CrossRef](#)]
120. Shibuya, H.; Iinuma, H.; Shimada, R.; Horiuchi, A.; Watanabe, T. Clinicopathological and prognostic value of microRNA-21 and microRNA-155 in colorectal cancer. *Oncology* **2011**, *79*, 313–320. [[CrossRef](#)] [[PubMed](#)]
121. Wijnhoven, B.P.L.; Hussey, D.J.; Watson, D.I.; Tsykin, A.; Smith, C.M.; Michael, M.Z. MicroRNA profiling of Barrett’s oesophagus and oesophageal adenocarcinoma. *Br. J. Surg.* **2010**, *97*, 853–861. [[CrossRef](#)]
122. He, B.; Yin, B.; Wang, B.; Xia, Z.; Chen, C.; Tang, J. microRNAs in esophageal cancer (Review). *Mol. Med. Rep.* **2012**, *6*, 459–465. [[CrossRef](#)] [[PubMed](#)]
123. Liu, R.; Liao, J.; Yang, M.; Sheng, J.; Yang, H.; Wang, Y.; Pan, E.; Guo, W.; Pu, Y.; Kim, S.J.; et al. The Cluster of miR-143 and miR-145 Affects the Risk for Esophageal Squamous Cell Carcinoma through Co-Regulating Fascin Homolog 1. *PLoS ONE* **2012**, *7*, e33987. [[CrossRef](#)] [[PubMed](#)]
124. Faccini, J.; Ruidavets, J.B.; Cordelier, P.; Martins, F.; Maoret, J.J.; Bongard, V.; Ferrières, J.; Roncalli, J.; Elbaz, M.; Vindis, C. Circulating MIR-155, MIR-145 and let-7c as diagnostic biomarkers of the coronary artery disease. *Sci. Rep.* **2017**, *7*, 42916. [[CrossRef](#)] [[PubMed](#)]
125. Li, D.; Zhang, T.; Lai, J.; Zhang, J.; Wang, T.; Ling, Y.; He, S.; Hu, Z. MicroRNA—25/ATXN3 interaction regulates human colon cancer cell growth and migration. *Mol. Med. Rep.* **2019**, *19*, 4213–4221. [[CrossRef](#)]
126. Gong, J.; Cui, Z.; Li, L.; Ma, Q.; Wang, Q.; Gao, Y.; Sun, H. MicroRNA-25 promotes gastric cancer proliferation, invasion, and migration by directly targeting F-box and WD-40 Domain Protein 7, FBXW7. *Tumor Biol.* **2015**, *36*, 7831–7840. [[CrossRef](#)]
127. Glynne-Jones, R.; Grainger, J.; Harrison, M.; Ostler, P.; Makris, A. Neoadjuvant chemotherapy prior to preoperative chemoradiation or radiation in rectal cancer: Should we be more cautious? *Br. J. Cancer* **2006**, *94*, 363–371. [[CrossRef](#)]
128. Vaidya, F.U.; Sufiyah Chhipa, A.; Mishra, V.; Gupta, V.K.; Rawat, S.G.; Kumar, A.; Pathak, C. Molecular and cellular paradigms of multidrug resistance in cancer. *Cancer Rep.* **2020**, *2020*, e1291. [[CrossRef](#)]
129. Gunduz, M.; Gunduz, E.; Tamagawa, S.; Enomoto, K.; Hotomi, M. Cancer stem cells in Oropharyngeal Cancer. *Cancers* **2021**, *13*, 3878. [[CrossRef](#)]
130. Wang, D.-K.; Zuo, Q.; He, Q.-Y.; Li, B. Targeted Immunotherapies in Gastrointestinal Cancer: From Molecular Mechanisms to Implications. *Front. Immunol.* **2021**, *12*, 705999. [[CrossRef](#)]
131. Yang, L.; Wang, Y.; Wang, H. Use of immunotherapy in the treatment of gastric cancer (Review). *Oncol. Lett.* **2019**, *18*, 5681–5690. [[CrossRef](#)] [[PubMed](#)]
132. Le, D.T.; Kim, T.W.; van Cutsem, E.; Geva, R.; Jäger, D.; Hara, H.; Burge, M.; O’Neil, B.; Kavan, P.; Yoshino, T.; et al. Phase II open-label study of pembrolizumab in treatment-refractory, microsatellite instability–high/mismatch repair–deficient metastatic colorectal cancer: KEYNOTE-164. *J. Clin. Oncol.* **2019**, *38*, 11–19. [[CrossRef](#)] [[PubMed](#)]
133. Dominika, B. CAR-T Cell Therapy—An Overview of Targets in Gastric Cancer. *J. Clin. Med.* **2020**, *9*, 1894.
134. Data, A.; Bv, I. Gastrin 17 vaccine—Aphton: Anti-gastrin 17 immunogen, G17DT. *BioDrugs* **2003**, *17*, 223–225.
135. Gilliam, A.D.; Watson, S.A.; Henwood, M.; McKenzie, A.J.; Humphreys, J.E.; Elder, J.; Iftikhar, S.Y.; Welch, N.; Fielding, J.; Broome, P.; et al. A phase II study of G17DT in gastric carcinoma. *Eur. J. Surg. Oncol.* **2004**, *30*, 536–543. [[CrossRef](#)]
136. Zheng, X.; Carstens, J.L.; Kim, J.; Scheible, M.; Kaye, J.; Sugimoto, H.; Wu, C.; Lebleu, V.S.; Kalluri, R.; Biology, C. EMT Program is Dispensable for Metastasis but Induces Chemoresistance in Pancreatic Cancer. *Nature* **2015**, *527*, 525–530. [[CrossRef](#)] [[PubMed](#)]
137. Liao, Z.J.; Guo, Y.H.; Zhao, Z.; Yao, J.T.; Xu, R.; Nan, K.J. Gemcitabine inhibits the micrometastasis of non-small cell lung cancer by targeting the EpCAM-positive circulating tumor cells via the HGF/cMET pathway. *Int. J. Oncol.* **2014**, *45*, 651–658. [[CrossRef](#)]
138. Geng, R.; Song, L.; Li, J.; Zhao, L. The safety of apatinib for the treatment of gastric cancer. *Expert Opin. Drug Saf.* **2015**, *17*, 1145–1150. [[CrossRef](#)]
139. Nagaraju, G.P.; Farran, B.; Farren, M.; Chalikonda, G.; Wu, C.; Lesinski, G.B.; El-Rayes, B.F. Napabucasin (BBI 608), a potent chemoradiosensitizer in rectal cancer. *Cancer* **2020**, *126*, 3360–3371. [[CrossRef](#)]
140. Qureshy, Z.; Johnson, D.E.; Grandis, J.R. Targeting the JAK/STAT pathway in solid tumors. *J. Cancer Metastasis Treat.* **2020**, *6*, 27. [[CrossRef](#)]

141. Pádua, D.; Figueira, P.; Ribeiro, I.; Almeida, R.; Mesquita, P. The Relevance of Transcription Factors in Gastric and Colorectal Cancer Stem Cells Identification and Eradication. *Front. Cell Dev. Biol.* **2020**, *8*, 442. [[CrossRef](#)]
142. Wall, J.A.; Klempner, S.J.; Arend, R.C. The anti-DKK1 antibody DKN-01 as an immunomodulatory combination partner for the treatment of cancer. *Expert Opin. Investig. Drugs* **2020**, *29*, 639–644. [[CrossRef](#)]
143. Zou, J.; Li, C.; Jiang, S.; Luo, L.; Yan, X.; Huang, D.; Luo, Z. AMPK inhibits Smad3-mediated autoinduction of TGF- β 1 in gastric cancer cells. *J. Cell. Mol. Med.* **2021**, *25*, 2806–2815. [[CrossRef](#)] [[PubMed](#)]
144. Hu, Q.; Li, L.; Zou, X.; Xu, L.; Yi, P. Berberine attenuated proliferation, invasion and migration by targeting the AMPK/HNF4 α /WNT5a pathway in gastric carcinoma. *Front. Pharmacol.* **2018**, *9*, 1150. [[CrossRef](#)] [[PubMed](#)]
145. Yu, D.; Shin, H.S.; Lee, Y.S.; Lee, D.; Kim, S.; Lee, Y.C. Genistein attenuates cancer stem cell characteristics in gastric cancer through the downregulation of Gli1. *Oncol. Rep.* **2014**, *31*, 673–678. [[CrossRef](#)] [[PubMed](#)]
146. Kashiyama, T.; Oda, K.; Ikeda, Y.; Shiose, Y.; Hirota, Y.; Inaba, K.; Makii, C.; Kurikawa, R.; Miyasaka, A.; Koso, T.; et al. Antitumor activity and induction of TP53-dependent apoptosis toward ovarian clear cell adenocarcinoma by the dual PI3K/mTOR inhibitor DS-7423. *PLoS ONE* **2014**, *9*, e87220. [[CrossRef](#)] [[PubMed](#)]
147. Liu, Q.; Yu, S.; Li, A.; Xu, H.; Han, X.; Wu, K. Targeting interleukin-6 to relieve immunosuppression in tumor microenvironment. *Tumor Biol.* **2017**, *39*, 1010428317712445. [[CrossRef](#)] [[PubMed](#)]
148. Li, M.; Yue, G.G.L.; Song, L.H.; Huang, M.B.; Lee, J.K.M.; Tsui, S.K.W.; Fung, K.P.; Tan, N.H.; Lau, C.B.S. Natural small molecule bigelovin suppresses orthotopic colorectal tumor growth and inhibits colorectal cancer metastasis via IL6/STAT3 pathway. *Biochem. Pharmacol.* **2018**, *150*, 191–201. [[CrossRef](#)]
149. Wang, Y.; Lv, Y.; Liu, T.S.; Yan, W.D.; Chen, L.Y.; Li, Z.H.; Piao, Y.S.; An, R.B.; Lin, Z.H.; Ren, X.S. Cordycepin suppresses cell proliferation and migration by targeting CLEC2 in human gastric cancer cells via Akt signaling pathway. *Life Sci.* **2019**, *223*, 110–119. [[CrossRef](#)]
150. Hur, H.; Xuan, Y.; Kim, Y.B.; Lee, G.; Shim, W.; Yun, J.; Ham, I.H.; Han, S.U. Expression of pyruvate dehydrogenase kinase-1 in gastric cancer as a potential therapeutic target. *Int. J. Oncol.* **2013**, *42*, 44–54. [[CrossRef](#)]
151. Wang, Y.; Chen, M.; Wu, Z.; Tong, C.; Dai, H.; Guo, Y.; Liu, Y.; Huang, J.; Lv, H.; Luo, C.; et al. CD133-directed CAR T cells for advanced metastasis malignancies: A phase I trial. *Oncoimmunology* **2018**, *7*, e1440169. [[CrossRef](#)]
152. Shitara, K.; Doi, T.; Nagano, O.; Imamura, C.K.; Ozeki, T.; Ishii, Y.; Tsuchihashi, K.; Takahashi, S.; Nakajima, T.E.; Hironaka, S.; et al. Dose-escalation study for the targeting of CD44v+ cancer stem cells by sulfasalazine in patients with advanced gastric cancer (EPOC1205). *Gastric Cancer* **2017**, *20*, 341–349. [[CrossRef](#)] [[PubMed](#)]
153. Kim, M.J.; Huang, Y.; Park, J. II Targeting wnt signaling for gastrointestinal cancer therapy: Present and evolving views. *Cancers* **2020**, *12*, 3638. [[CrossRef](#)] [[PubMed](#)]
154. Pan, L.; Zhang, T.; Sun, H.; Liu, G. Ginsenoside Rg3 (Shenyi Capsule) Combined with Chemotherapy for Digestive System Cancer in China: A Meta-Analysis and Systematic Review. Evidence-based Complement. *Altern. Med.* **2019**, *2019*, 2417418. [[CrossRef](#)] [[PubMed](#)]