Quantify the Protein-Protein Interaction Effects on Adsorption related Lubricating Behaviors of α -Amylase on a Glass Surface

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This supporting information contains (*i*) molecular weight determination, (*ii*) raw CD spectral data of native and adsorbed α - amylase responses under varying PPI conditions effect, (*iii*) friction test using pin-on-disk tribometer, and (*iv*) theoretical calculation of the monolayer adsorption capacity.

S.1 Determination of the molecular weight

The molecular weight determination of α - amylase using sodium dodecyl sulfatepolyacrylamide gel electrophoresis (SDS-PAGE) was performed according to a previous study[1]. The chemicals required for the analysis was prepared in an appropriate quantity. For the formation of separating gel, 1.5 M Tris was made and the pH value was adjusted to 8.8. For the stacking gel preparation, 1 M Tris was prepared, and the pH value was adjusted to 6.8. The concentration of the gel was determined according to the molecular weight of the protein sample. 1 mg/ml of α - amylase solution was mixed with 4 X SDS loading buffer (2% SDS; 10% glycerol; 0.5% bromophenol blue and 0.5M Tris-HCl, pH 6.8) and kept in boiling water (100°C) for 10 min. Then ten µl of the sample was loaded to each lane of 5 to 15% Bis-Tris gradient gels and separated at 50A and 120 V/gel using Mini Protean Tetra Cell units (Bio-Rad Laboratories, Inc., Richmond, CA). Following the separation, the gel was stained with Coomassie Brilliant Blue G-250 (Bioshop Canada Inc., Burlington, ON, Canada) for 20 minutes and then destained twice with acetic acid (PanReac AppliChem GmbH, Ottoweg, Darmstadt, Germany).

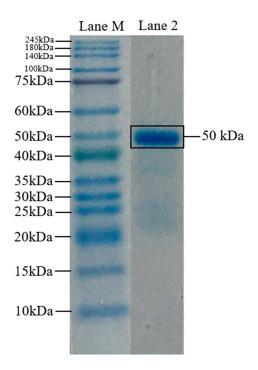


Figure S1. SDS-PAGE of α -amylase from barley malt used in this study. Lane M and Lane 2 denotes the marker and α -amylase loaded lane, respectively.

From the figure, it is more apparent that the molecular weight of the α -amylase used in this study is about 50 kDa. This molecular weight of α -amylase determined was used as the mass of each α -amylase protein for the theoretical calculation of the single-layer protein adsorption capacity described in detail in the section S.4.

S.2 Raw CD spectral data of native and adsorbed α - amylase responses under varying PPI conditions effect

Fig. S2. Illustrates the sample CD spectra for the native (i.e., α -amylase protein solution) and adsorbed proteins on the glass surface when adsorbed from 0.1 mg/ml, 0.5 mg/ml and 1 mg/ml bulk solution concentrations for an adsorption time of 2 hours and then equilibrated in the nano-pure water for another 2 hours. This obtained molar ellipticity (θ_{mol}) was uploaded on the DichroWeb, an online database to know the percentage of helical and sheet content [2, 3].

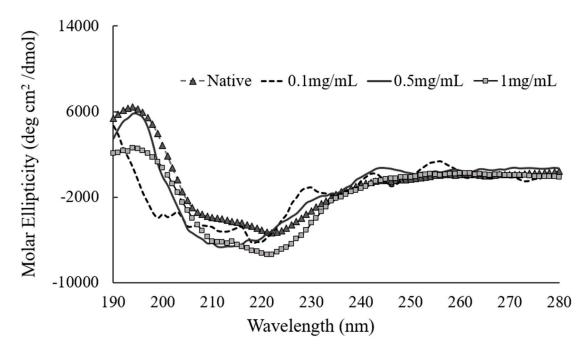


Figure S2. CD spectra for Native and adsorbed α -amylase on glass surface when adsorbed from 0.1 mg/ml, 0.5 mg/ml and 1 mg/ml bulk solution concentrations. (Average of 3 spectra). "Native" denotes the native α -amylase protein in solution.

S.3 Friction test using a pin-on-disk tribometer

A schematic of the experimental pin-on-disk tribometer was shown in Fig. S3. The upper stage with a pin was made of PDMS, a soft oral-mimetic surface[4]. The pin was rubbed against the glass with adsorbed α -amylase locked in the lower stage with a force of 0.5 N, which is the critical surface tension for saliva-coated tooth surfaces as reported in the previous studies[5]. The coefficient of friction was calculated by the ratio of the sliding friction force (F_x) to the forward loading force (F_z), as mention in equation (S1).

$$Coefficient of friction, COF = \frac{F_X}{F_Z}$$
(S1)

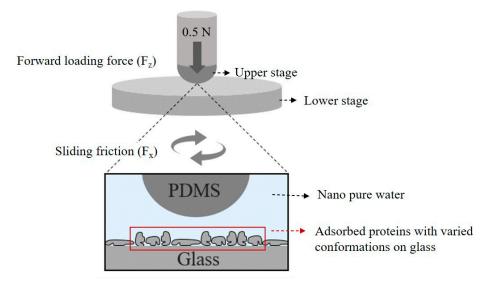


Figure S3. Schematic diagram of the friction testing machine.

α-Amylase protein concentration (mg/ml)	Obtained COF	Averaged COF
Native	0.009546	
Native	0.002883	0.0055
Native	0.004164	
0.1	0.13407	
0.1	0.127179	0.1393
0.1	0.156576	
0.5	0.098456	
0.5	0.107971	0.121
0.5	0.156606	
1	0.034066	
1	0.028057	0.0288
1	0.024344	

Table S1. The averaged values of coefficient of friction from the obtained value in the range of 800-900s

S.4 Theoretical calculation of the single-layer adsorption capacity

The monolayer adsorption capacity can be theoretically calculated by assuming that the structure of the α -amylase adsorbed on the glass surface is a spherical footprint. Previous studies have proved that the average density of the proteins with a molecular weight of around 40-50 kDa was about 1.41g/cm³[6]. Spherical volume (v) and surface area (A) of the α -amylase, as shown in equation (S2) and (S3) respectively, was used as per our assumption, where radius (r) can be calculated using (Equation S2).

$$v = \frac{4}{3}\pi r^3 \tag{S2}$$

$$A = \pi r^2 \tag{S3}$$

$$M = \frac{1}{\pi r^2} X \frac{1}{6.02 \times 10^{-20}} X \ 50000 \tag{S4}$$

$$V = 50000 X \frac{1}{6.02 * 10^{20}} X \frac{1}{1.41}$$
(S5)

Theoretical protein mass per unit area (M) (i.e., single-layer adsorption capacity) and the volume of each protein (V) (i.e., mass/density) adsorbed was calculated as 0.0004 mg/cm^3 and $5.97 \times 10^{-20} \text{cm}^3$ using the equation (S4) & (S5) respectively which was derived from the previous paper[7].

References:

[1] U.K. Laemmli, Cleavage of structural proteins during the assembly of the head of bacteriophage T4, nature 227(5259) (1970) 680-685.

[2] L. Whitmore, B.A. Wallace, DICHROWEB, an online server for protein secondary structure analyses from circular dichroism spectroscopic data, Nucleic acids research 32(Web Server issue) (2004) W668-73.

[3] L. Whitmore, B.A. Wallace, Protein secondary structure analyses from circular dichroism spectroscopy: Methods and reference databases, Biopolymers 89(5) (2008) 392-400.

[4] A. Sarkar, E. Andablo-Reyes, M. Bryant, D. Dowson, A. Neville, Lubrication of soft oral surfaces, Current Opinion in Colloid & Interface Science 39 (2019) 61-75.
[5] C.E. CHRISTERSSON, R.G. DUNFORD, P.O.J. GLANTZ, R.E. BAIER, Effect of critical surface tension on retention of oral microorganisms, European journal of oral sciences 97(3) (1989) 247-256.

[6] H. Fischer, I. Polikarpov, A.F. Craievich, Average protein density is a molecularweight-dependent function, Protein Science 13(10) (2004) 2825-2828.

[7] M. Rabe, D. Verdes, S. Seeger, Understanding protein adsorption phenomena at solid surfaces, Advances in colloid and interface science 162(1-2) (2011) 87-106.